

Reporting Summary

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Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
- A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- The statistical test(s) used AND whether they are one- or two-sided
Only common tests should be described solely by name; describe more complex techniques in the Methods section.
- A description of all covariates tested
- A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
Give P values as exact values whenever suitable.
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection

The sequencing reads were undertaken with PacBio Sequel 2 and Illumina NovaSeq platforms.

Data analysis

Khaper algorithm is freely available in GitHub (<https://github.com/lardo/khaper>) and calc_switchErr can be found in GitHub (https://github.com/tangerzhang/calc_switchErr). The codes (Khaper and calc_switchErr) also have been archived to Zenodo with the DOIs (10.5281/zenodo.4780792 and 10.5281/zenodo.4780666) and cited in refs.15 and 19.

Other programs used in this study are listed as follows:
CANU (version 1.9), BWA (version 0.7.15-r1140), Pilon (version 1.22), ALLHiC (version 0.1 <https://github.com/tangerzhang/ALLHiC>), Purge_haplotigs (https://bitbucket.org/mroachawri/purge_haplotigs/src/master/), Pseudohaploid (<https://github.com/schatzlab/pseudohaploid>), 3D-DNA (version 180922), GATK (version 3.8), SAMtools (version 1.9), VCFtools (version 0.1.16), BLASTN (version 2.7.1), BLASTP (version 2.7.1), LAST (version 959), R (version 3.5.1), BUSCO (version 3.0.2), LTR_retriever (version 2.8), RECON (version 1.08), RepeatScout (version 1.0.5), TEclass (version 2.1.3), TRF (version 4.07), LTR-FINDER (version 1.0.5), RepeatMasker (version open-4.0.7), LTRharvest (version 1.5.10), GeneWise (version 2.4.1), Trinity (version 2.3.2), RSEM (version 1.2.31), PASA (version 2.2.0), Augustus (version 2.4), SNAP(<https://github.com/KorfLab/SNAP>), BLAT (version 350), MAKER (version 2.31.10), OrthoMCL (version 1.1.4), MUSCLE (version 3.8.31), DensiTree (version 2.2.5), MCScanX (<http://chibba.pgml.uga.edu/mcscan2/>), MCscan ([https://github.com/tanghaibao/jcvi/wiki/MCscan-\(Python-version\)](https://github.com/tanghaibao/jcvi/wiki/MCscan-(Python-version))), iTOL (<https://itol.embl.de/>), proc10xG (<https://github.com/ucdavis-bioinformatics/proc10xG>), NUCMER (version 4.0.0), GMAP (version 2013-10-28), MAFFT (version 7.299b), Trimmomatic (version 0.33), IQ-Tree (version 1.6.12), RAXML (version 8.2.12), PLINK (version 1.9), ANGSD (version 0.932), PopLDdecay (version 3.31), ADMIXTURE (version 1.3.0), Selscan (version 1.3.0).

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

The raw sequencing reads of PacBio, Illumina, 10× Genomics, Hi-C, RNA-seq and ISO-seq have been deposited in the National Center for Biotechnology Information (NCBI) database with the accession number PRJNA665594 and/or in GSA database (<https://bigd.big.ac.cn/gsa/>) under the accession number PRJCA003090. The assembly and annotation have been archived in NCBI with the accession number JAFLEL000000000 and GWH (<https://bigd.big.ac.cn/gwh/>) with the accession numbers GWHASIV000000000 for the monoploid and GWHASIX000000000 for the haplotype-resolved genomes. The VCF that contains all of the clean SNPs were uploaded to Mendeley database (<https://data.mendeley.com/datasets/7hb33vd7sf/1>). In addition, three datasets that were used to assess the switch errors in the haplotype-resolved TGY genome assembly were deposited to Mendeley database (<http://dx.doi.org/10.17632/xpccyg5w2x.1>).

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

- Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

- Sample size
- Data exclusions
- Replication
- Randomization
- Blinding

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

- | n/a | Involved in the study |
|-------------------------------------|--|
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| <input checked="" type="checkbox"/> | <input type="checkbox"/> Eukaryotic cell lines |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Palaeontology |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Animals and other organisms |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Human research participants |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Clinical data |

Methods

- | n/a | Involved in the study |
|-------------------------------------|---|
| <input checked="" type="checkbox"/> | <input type="checkbox"/> ChIP-seq |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Flow cytometry |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> MRI-based neuroimaging |