The BTB-domain transcription factor ZBTB2 recruits chromatin remodelers and a histone chaperone during the exit from pluripotency

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SUPPLEMENTARY INFORMATION

Supplementary Figures S1-S5 Supplementary Table S1: Screen results Supplementary Table S2: CRISPR KO, KI, and genotyping strategies; cell lines Supplementary Table S3: Gene expression tables with clusters and post- and pre-implantation epiblast genes Supplementary Table S4: AP-MS bait sequences and Y2H constructs Supplementary Table S5: AP-MS results tables Supplementary Table S6: BTB domain sequences Figure S1



Supplementary Figure S1: Related to Figure 1.

(A) Chromatograms of $Zfp42^{-/-}$ (cl.8, 15, 24) and WT (cl.9, 23) sibling clones under the respective gRNAs used (see also Table S2).

(**B,C**) Genotyping PCR of *Nmt1* and *Zbtb2* targeted clones, showing knockout (red) and sibling control clones (green) used in this study (see also Table S2).

(D,F) Average and standard deviation (SD) of *Nanog>GFP*-high cells of biological triplicates quantified as in Fig.1B, C, D (**D**). and Fig.1F (**F**). ** indicates p-values<0.001 and NS p-values>0.1 compared to corresponding *WT* controls.

(E) Anti-ERK and anti-phospho-ERK western-blot of lysates from *Nmt1^{-/-}* and *WT* clones.

(G) Scatterplot of log2FCs in gene expression of indicated contrasts. Green labels postimplantation (postEPI) and red pre-implantation epiblast (preEPI) specific genes (55).



Supplementary Figure S2: Identification of the ZBTB2-interacting subunits and domains. Related to Figure 2.

(A) Scheme of the ZBTB2-3xFLAG knock-in strategy (see also Table S2) and chromatogram of final homozygous cell line.

(B) *Nanog>GFP* intensities after switching from 2iLIF to N2B27 +CHIR +bFGF for 3 days in the presence of DOX of indicated genotypes.

(C) *Zbtb25* genotyping PCR of *Zbtb2^{-/-}*; *Zbtb25^{-/-}* clones; red labels clones used in this study.

(D) qPCR of *Zbtb2* relative to *GAPDH* in *WT* and ZBTB2-AVI overexpressing cells. Average and SD of technical triplicates, ** indicates p-value<0.001.

(E,G,H,I,K) Colony growth on QDOXA plates of strains expressing indicated proteins. Bait constructs are vertical and prey constructs are horizontal.

(F,J) Diagram of *Ubn2* (F) and *Gatad2b* (J) constructs used for Y2H analysis; + and – indicates positive and negative interactions as in (G,I and K).

(L) Genotyping PCR of *Znf639*^{-/-} clones; red labels clones used in this study.



Supplementary Figure S3: Lack of genetic interaction between ZBTB2 and ZBTB25. Related to Figure 3.

(A) qPCR of *Ubn1* and *Ubn2* relative to *GAPDH* in *WT* and UBN1-AVI- or UBN2-AVIoverexpressing cells. Average and SD of technical triplicates, * indicates p-value<0.01 and ** p-value<0.001.

(B) Volcano plot of protein enrichments in AP-MS of mESCs overexpressing UBN1-AVI compared to control BirA-only-expressing mESCs. ZBTB2 is indicated in blue and HiRA subunits in green.

(C,D) Colony growth on QDOXA plates of control matings for experiments presented in Fig.3B,C,F (C) and in Fig.3D (D) using ZBTB2's link region as control bait construct.

(E) *Zbtb25* genotyping PCR of *Zbtb25^{-/-}* clones; red labels clones used in this study.

(F,G) *Nanog>GFP* intensities after switching from 2iLIF to Serum-LIF after 60h of indicated genotypes (F). Dashed line indicates the threshold for quantification of GFP-high cells as in (G). (G) average and SD of biological triplicates; ** indicates p-values<0.001 and NS p-values>0.1 compared to the *WT* control.

Figure S4 Α gRNA#2 ccagagggtctacccgtgcttcttgagaaad ACCTACACTTCCAGAGG. TCTACCCGTGCTTCTTGAGAAACGCATT Ubn2 cl.10 gRNA#3 gcccttggggatgacatcccggacttaggtctg ACAACAGCCCTTGGGG-----GACTTAGGTCTGAACAG *Ubn2* cl.2 \mathbb{N} $\mathcal{M}\mathcal{M}\mathcal{M}\mathcal{M}$ $M \sim$ GTGTCACAACAGCCCTTGGGGATGACATCCCGGACTTAGGTCTGA Mannaman Ubn2 cl.15 Ŵ В 4.5 Zbtb2-HAF in *Ubn2^{-/-}* Zbtb2-HAF in W7 Srsf7 Hira Ubn2 4



Supplementary Figure S4: The Ep400 interaction is not mediated by the HiRA complex. Related to Figure 4.

(A) Chromatograms of $Ubn2^{-/-}$ (cl.10,2) and WT (cl.15) sibling clones under the respective gRNAs used (see also Table S2).

(B) Volcano plot of protein enrichments in AP-MS of overexpressed ZBTB2-HAF in *WT* compared to *Ubn2^{-/-}* mESCs. ZBTB2 and partner TFs are indicated in blue. The dashed line marks enrichment of the ZBTB2-HAF bait protein which is similar to that of NuRD subunits in red and Ep400 subunits in orange, while interaction with HiRA subunits in green is comparatively reduced in *Ubn2^{-/-}* mESCs.



Supplementary Figure S5: ZBTB2 binds and represses its own promoter. Related to Figure 5.

(A) Boxplot of log2 fold differential expression after 48h in Serum-LIF of pre-implantation (preEPI, red) and post-implantation (postEPI, green) epiblast specific genes (55) in indicated genotypes compared to *WT* cells.

(B,C) ZBTB2 (53), KAT5 (60) and CHD4 (25) (left scale), and DUX (17) (right scale) ChIP-seq rpkm centered on TSSs of cluster 1 (**B**) and cluster 2-9 genes (**C**), according to Fig. 5D.

(D) Heatmap of ZBTB2 (53), H3K4me3 (79), CHD4, MBD3 (25), KAT5, EP400 (60) and DUX (17) log2 fold ChIP-seq enrichment over respective controls at accessible (ATACseq, not shown) TSSs.

(E) Diagram of the *Zbtb2* locus, showing the ZBTB2 peak at the TSS, and the deletions in the *Zbtb2^{-/-}* clones 58I and 58M that give rise to transcripts that are detectable by the qPCR probe (TaqMan Probe) against endogenous *Zbtb2*.

(F) qPCR of endogenous *Zbtb2* relative to *GAPDH* in *WT* cells, *WT* cells over-expressing a *Zbtb2* construct that cannot be detected by the qPCR probe (TAQMANmut) and the *Zbtb2*^{-/-} clones depicted in Fig. S5E. Average and SD of technical triplicates; * indicates p-values<0.01 compared to *WT*.

(G) qPCR of *Kat5* and *Zbtb2* relative to *GAPDH* upon *Kat5* or control siRNA transfection. Average and SD of technical triplicates; ** indicates p-value<0.001 compared to control siRNA.