

Expanded View Figures

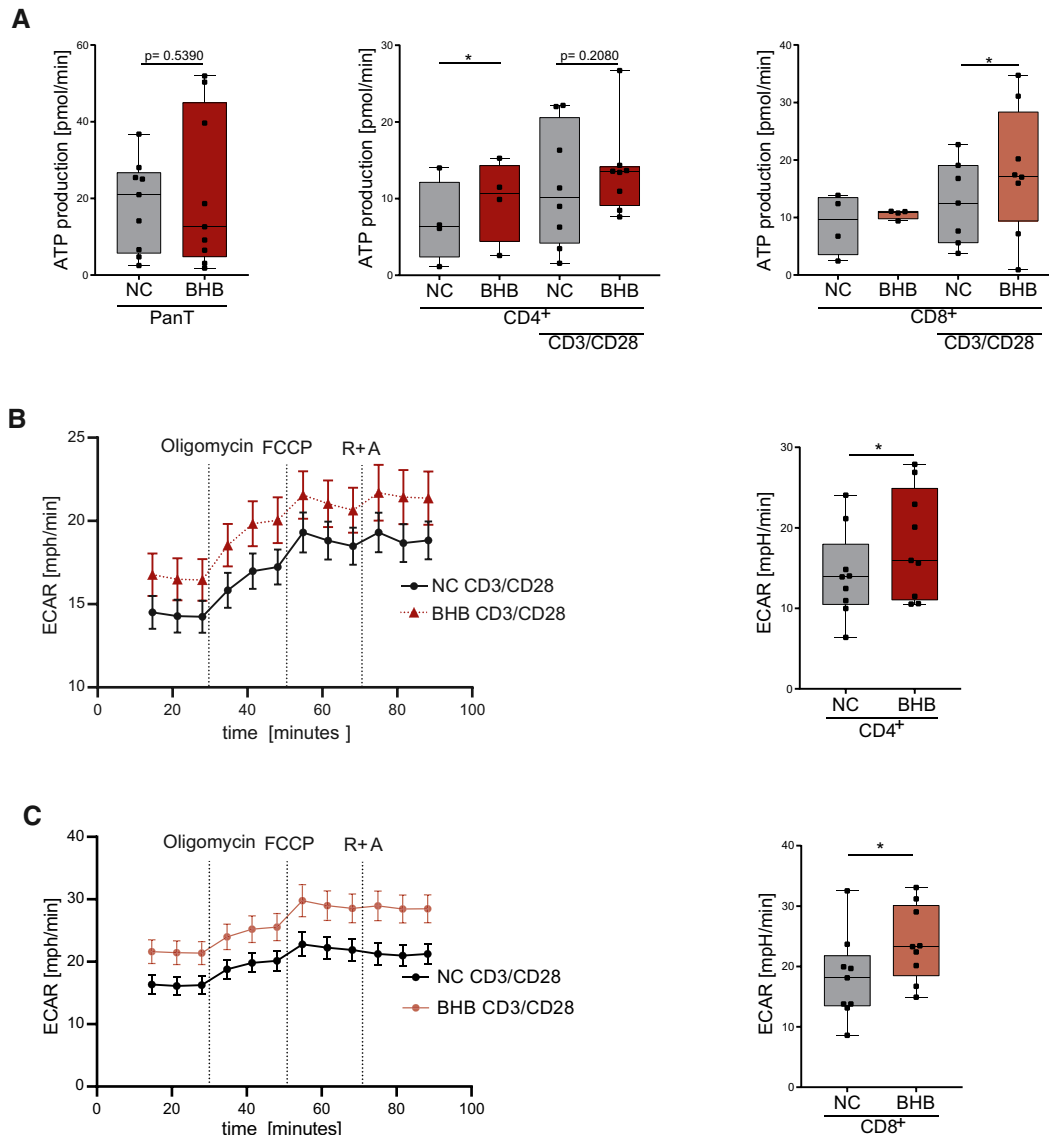


Figure EV1. Ketone bodies shift T-cell metabolism to oxidative phosphorylation.

Human PBMCs were cultivated for 48 h in RPMI containing 80 mg/dl glucose (NC) and supplemented with 10 mM beta-hydroxybutyrate (BHB). T-cell stimulation was performed through CD3/CD28 Dynabeads at a bead:cell ratio of 1:8. Pan T cells and CD4⁺ and CD8⁺ T cells were isolated through magnetic cell separation. Mitochondrial metabolism was analyzed for each subpopulation using a Seahorse XF96 Analyzer.

A Mitochondrial ATP production was measured in pan T cells, CD4⁺ T cells, and CD8⁺ T cells, $n = 9$ (pan T) and $n = 4/8$ (unstimulated/stimulated CD4 and CD8) individual experiments.

B, C Extracellular acidification rate, measured in (B) CD4⁺ T cells and (C) CD8⁺ T cells, $n = 9$ biological replicates.

Data information: Data depicted as mean \pm SEM (ECAR) and box plots with median, 25th and 75th percentiles and range (all other). Dots indicating individual values.

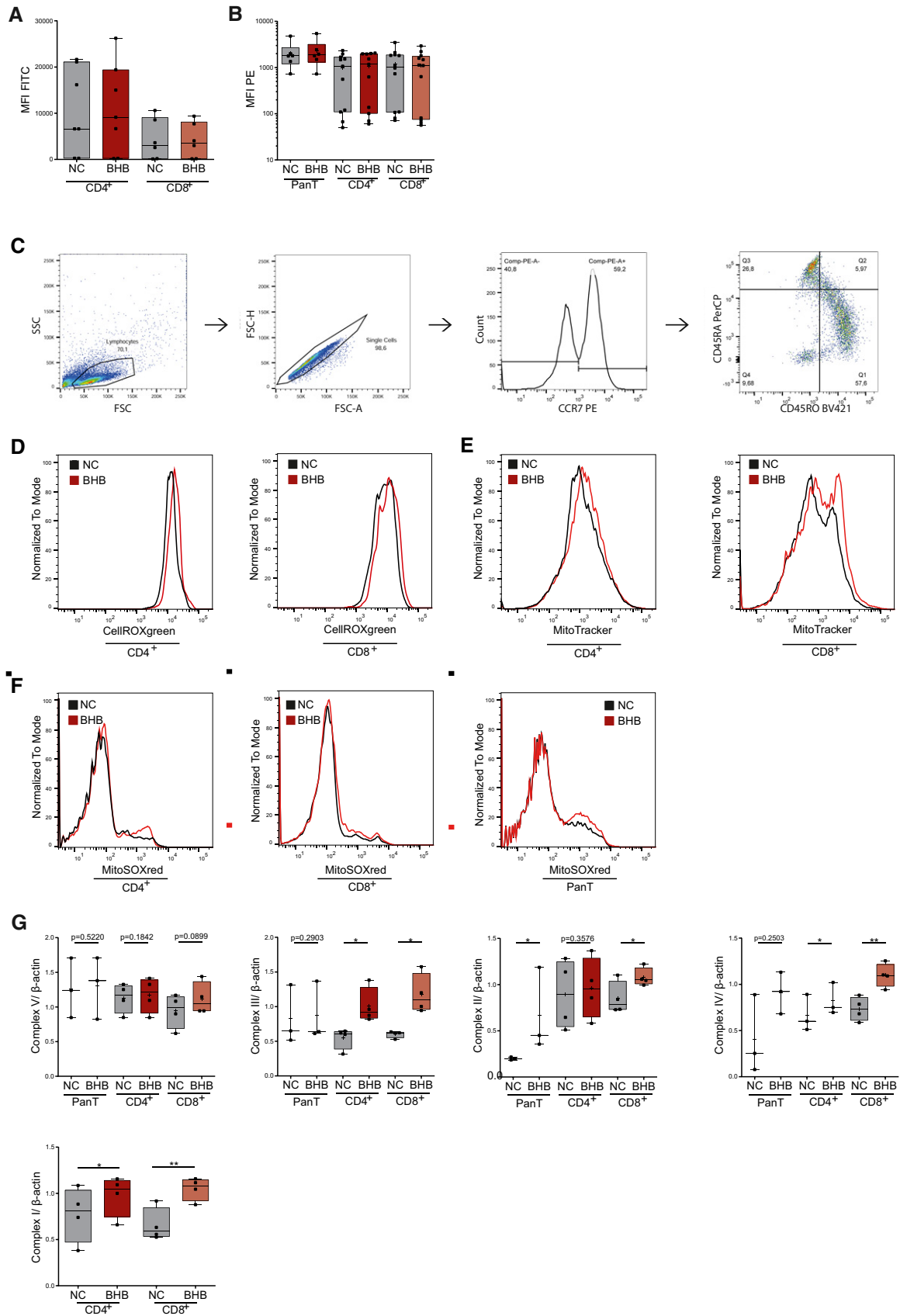
* $P < 0.05$ paired t-test/Wilcoxon matched-pairs signed rank test, as appropriate.

Figure EV2. BHB does not increase ROS in unstimulated human T cells.

Human peripheral blood mononuclear cells (PBMCs) were cultivated for 48 h in RPMI containing 80 mg/dl glucose (NC) and supplemented with 10 mM beta-hydroxybutyrate (BHB). Native T cells were analyzed via flow cytometry using the indicated dyes and antibodies identifying T cells and CD4⁺/CD8⁺ T cell subsets.

- A Quantification of cellular ROS using CellROX, indicated by MFI FITC in human CD4⁺/CD8⁺ T cells, $n = 7/6$ biological replicates, $P = 0.4917/0.6324$.
- B Mitochondrial superoxide production quantified through MitoSOX staining, depicted as MFI PE in human pan/CD4⁺/CD8⁺ T cells, $n = 6/11/12$ biological replicates, $P = 0.0938/0.3209/0.1401$.
- C Representative gating strategy for memory cell quantification using flow cytometry.
- D–F Representative histogram plots for quantification of (D) cellular ROS using CellROXgreen, (E) mitochondrial mass using MitoTracker green and (F) mitochondrial ROS using MitoSOXred.
- G Densitometric analysis of Western blots of mitochondrial oxidative phosphorylation proteins in pan/CD4⁺/CD8⁺ T cells as indicated, $n = 3/4/4$ (CD4 complex IV, $n = 3$) biological replicates.

Data information: Data depicted as box plots with median, 25th and 75th percentiles and range, crosses indicating mean, dots indicating individual values. * $P < 0.05$, ** $P < 0.01$, paired t-test/Wilcoxon matched-pairs signed rank test, as appropriate.



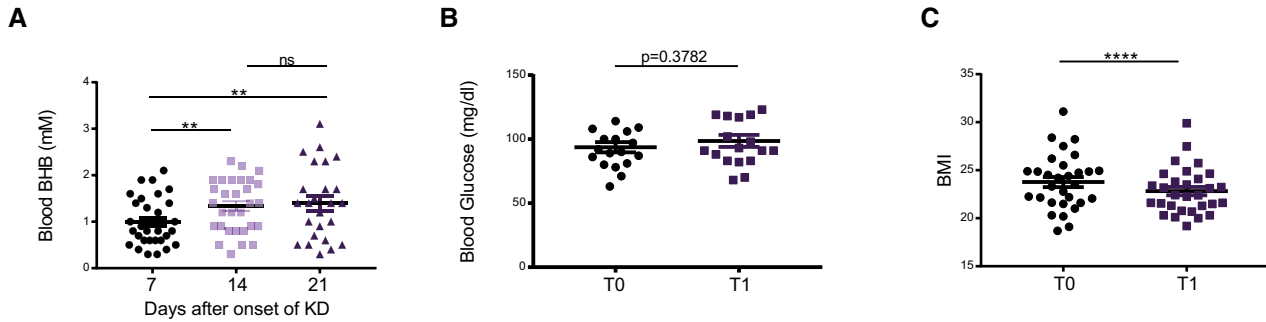


Figure EV3. Ketogenic diet *in vivo*.

Healthy volunteers conducted a 3-week ketogenic diet (KD) with limited carbohydrate consumption of < 30 g/day.

- A Blood BHB concentration of healthy volunteers at the indicated time points as measured by point of care instruments.
 B Blood glucose concentration of healthy volunteers was analyzed prior to starting the diet (T₀) and again after 3 weeks of strict adherence to the diet (T₁).
 C Participants' BMI measured at the start and at the end of KD.

Data information: Data depicted as mean ± SEM, dots indicating individual values. ^{ns}P > 0.05, **P < 0.01, ****P < 0.0001, paired t-test/Wilcoxon matched-pairs signed rank test, as appropriate.

Figure EV4. Ketogenic diet primes human T cells to mitochondrial metabolism and memory cell development *in vivo*.

Forty-four healthy volunteers conducted a 3-week KD with a limited carbohydrate consumption of < 30 g/day. Blood was taken and analyzed prior to starting the diet (T₀) and again after 3 weeks of strict adherence to the diet (T₁). PBMCs were isolated. If applicable, T-cell stimulation was performed through CD3/CD28 Dynabeads at a bead:cell ratio of 1:8. Pan/CD4⁺/CD8⁺ T cells were separated via magnetic cell labeling. Mitochondrial metabolism was analyzed for each subpopulation using a Seahorse XF96 Analyzer.

- A Mitochondrial ATP production was measured in stimulated CD4⁺ and CD8⁺ T cells, n = 14 individual human subjects.
 B–E Extracellular acidification rate, measured in (B) unstimulated and (C) stimulated CD4⁺ T cells and (D) unstimulated and (E) stimulated CD8⁺ T cells, n = 6 individual human subjects.
 F, G Glycolytic proton efflux rate (glycoPER) and compensatory (maximum) glycoPER measured in stimulated (F) CD4⁺ T cells and (G) CD8⁺ T cells, depicted as mean ± 95% CI/mean ± SEM, n = 7/8 individual human subjects.
 H Quantification of mitochondrial membrane potential using JC1, indicated by MFI PE/FITC in human lymphocytes *in vivo*. n = 4/5 (unstimulated/stimulated) individual human subjects.
 I, J Quantification of (I) cellular and (J) mitochondrial ROS using CellROX/MitoSOX, indicated by MFI FITC and MFI PE in native human Pan/CD4⁺/CD8⁺ T cells, n = 8 (CellROX), 10/9/10 (MitoSOX) individual human subjects.
 K Quantification of mitochondrial mass using MitoTracker green, indicated by MFI FITC in native human pan/CD4⁺/CD8⁺ T cells, n = 10/10/11 individual human subjects.
 L TruCulture Interleukin (IL)1α/β protein quantification of LPS-stimulated whole blood samples, n = 11/12 individual human subjects.
 M TruCulture IL8 and IL12 subunit p40 protein quantification of unstimulated whole blood samples, n = 5 (IL8: T0 n = 4) individual human subjects. TruCulture IFNγ, IL4, IL6, IL23, and TNFα protein quantification of unstimulated whole blood samples were below the lower limit of quantification.

Data information: Data depicted as mean ± SEM (ECAR/glycoPER) and box plots (all other) with median, 25th and 75th percentiles and range, dots indicating individual values. *P < 0.05, **P < 0.01, paired t-test/Wilcoxon matched-pairs signed rank test, as appropriate.

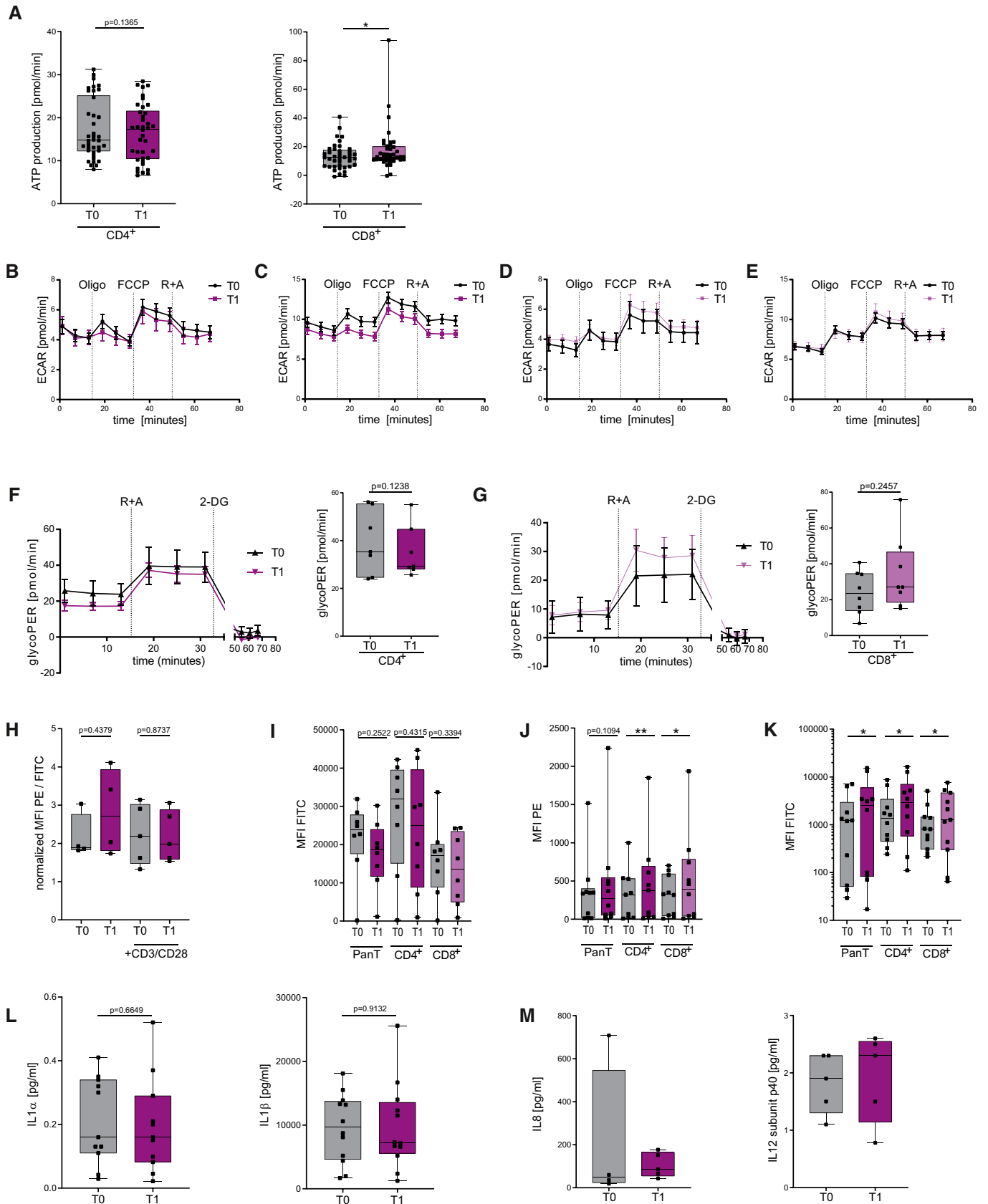


Figure EV4.