

1 **Supplemental Material for**

2  
3 Topoisomerase I Essentiality, DnaA-independent Chromosomal Replication,  
4 and Transcription-Replication Conflict in *Escherichia coli*

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6 J Krishna Leela<sup>a</sup>, Nalini Raghunathan<sup>a</sup>, and J Gowrishankar<sup>a,b#</sup>

7  
8 <sup>a</sup>Laboratory of Bacterial Genetics, Centre for DNA Fingerprinting and  
9 Diagnostics, Hyderabad, India

10 <sup>b</sup>Indian Institute of Science Education and Research Mohali, SAS Nagar, India

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12 Running Head: Topoisomerase I essentiality in *E. coli*

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14 #Address correspondence to Dr J Gowrishankar (Orcid ID: 0000-0003-2483-  
15 9209), Tel: +91-172-2240266; Fax: +91-172-2240124; Email:  
16 [shankar@iisermohali.ac.in](mailto:shankar@iisermohali.ac.in)

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## Supplementary Methods

21 **Construction of shelter plasmids pHYD2382 and pHYD2390.** The pair of  
22 primers **5'-CCAAACTGCAGTCGTGCTATAGCGCCTGT-3'** and 5'-  
23 TTTGTTAAGCTTAACCTGACAGAATTAAAGG -3' (*PstI* and *HindIII* sites in them,  
24 respectively, italicized) were used with MG1655 genomic DNA to amplify the  
25 *topA*<sup>+</sup> gene by PCR, and the product was digested with *PstI* and *HindIII* before  
26 being cloned into the corresponding sites of plasmid vector pMU575, to  
27 generate the *topA*<sup>+</sup> shelter plasmid pHYD2382. The two primers are  
28 designated, respectively, as b1 and b2 in Supplementary Figure S7.

29 For constructing plasmid pHYD2390 (*topA*<sup>+</sup> *dnaA*<sup>+</sup> shelter plasmid), the  
30 DNA fragment present in plasmid pHYD2388 (1) carrying the *Salmonella*  
31 *enterica dnaA*<sup>+</sup> gene was sub-cloned *via* an intermediate plasmid vector into  
32 the *XbaI-KpnI* sites of plasmid pHYD2382.

33 **Generation of *topA* mutations by recombineering.** The Datsenko and  
34 Wanner method (2), with plasmids pKD13 and pKD46, was used for  
35 generating chromosomal *topA* mutations by recombineering in derivatives  
36 carrying the shelter plasmids pHYD2382 or pHYD2390. The primer pairs used  
37 for PCR with pKD13 template were: **for**  $\Delta topA::Kan$ , 5'-  
38 TCAACGTGCGACGCATTCCTGGAAGAATCAAATTAGGTAAGGTGAATATGATT  
39 CCGGGGATCCGTCGACC-3' and 5'-  
40 TGTTTATAAAAACCTGACAGAATTAAAGGTTATTTTTTTCCTTCAACCCATGTAG  
41 GCTGGAGCTGCTTCG-3' (designated primer Y); **for** *topA*-Ins480::Kan, 5'-  
42 CCAGCACTTTACCAAGCCGCCAGCCCGTTTCAGTGAAGCAATTCCGGGGATC  
43 CGTCGACC-3' (designated primer X) and 5'-  
44 GACGACCGATACCGCGTTTTTCCAGCTCTTTAACCAGCGATGTAGGCTGGAG  
45 CTGCTTCG-3'; and **for** *topA*-480 $\Delta$ , primers X and Y.

46 Given that the recombineering in these derivatives could have occurred  
47 on either the chromosome or the shelter plasmid, we employed simple genetic  
48 tests on several independent clones to classify them into the two categories.  
49 The three *topA*::FRT alleles on the chromosome were generated with the aid  
50 of plasmid pCP20, as described (2).

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78 *coli* K-12 in-frame, single-gene knockout mutants: the Keio collection.  
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81 **Table S1.** List of *E. coli* strains

Strain <sup>a</sup>	Genotype <sup>b</sup>
MG1655	<i>E. coli</i> K-12 wild-type
MDS42	MG1655 with 42 engineered deletions in genome
GJ12134	MDS42 $\hat{e}$ ( <i>argF-lac</i> ) <i>U169</i>
GJ13519	MG1655 $\hat{e}$ ( <i>argF-lac</i> ) <i>U169</i> [ 80 lysogen]
GJ13531 <sup>c</sup>	GJ13519 <i>rho</i> ::Kan <i>att</i> ::P <sub>tac</sub> $\phi$ UvsW (Amp <sup>R</sup> )
GJ15603 <sup>d</sup>	GJ13519 <i>topA</i> ::Kan
GJ15604 <sup>d</sup>	GJ13519 <i>topA</i> -Ins480::Kan
GJ15630 <sup>d</sup>	GJ13519 <i>dnaA177 dgoT</i> ::FRT <i>rpoB</i> *35 <i>btuB</i> ::Tn10 <i>topA</i> -Ins480::FRT <i>tus</i> ::FRT
GJ15688 <sup>d</sup>	GJ13519 <i>topA</i> -480 ::Kan
GJ15697 <sup>d</sup>	GJ15603 <i>att</i> ::P <sub>tac</sub> $\phi$ UvsW (Amp <sup>R</sup> )
GJ16475 <sup>e</sup>	GJ13519 <i>rpoB</i> *35 <i>btuB</i> ::Tn10 <i>dnaA</i> ::FRT <i>tus</i> ::FRT <i>rhA</i> ::FRT
GJ16703	GJ13519 <i>rpoB</i> *35 <i>btuB</i> ::Tn10
GJ16813 <sup>d</sup>	GJ16703 <i>topA</i> ::Kan
GJ16814 <sup>d</sup>	GJ16703 <i>topA</i> -Ins480::Kan
GJ16815 <sup>d</sup>	GJ16703 <i>topA</i> -480 ::Kan
GJ16816 <sup>d</sup>	GJ12134 <i>topA</i> ::Kan
GJ16817 <sup>d</sup>	GJ12134 <i>topA</i> -Ins480::Kan
GJ16818 <sup>d</sup>	GJ12134 <i>topA</i> -480 ::Kan
GJ16819	GJ12134 <i>rpoB</i> *35 <i>btuB</i> ::Tn10
GJ16820 <sup>d</sup>	GJ16819 <i>topA</i> ::Kan
GJ16821 <sup>d</sup>	GJ16819 <i>topA</i> -Ins480::Kan
GJ16822 <sup>d</sup>	GJ16819 <i>topA</i> -480 ::Kan
GJ16854 <sup>d</sup>	GJ16703 <i>topA</i> -Ins480::FRT
GJ16880 <sup>d</sup>	GJ16854 <i>uvrD</i> ::Kan
GJ16902 <sup>d</sup>	GJ12134 <i>rpoB</i> *35 <i>btuB</i> ::Tn10 <i>dnaA177 dgoT</i> ::FRT <i>tus</i> ::FRT <i>topA</i> ::FRT
GJ16903 <sup>d</sup>	GJ12134 <i>rpoB</i> *35 <i>btuB</i> ::Tn10 <i>dnaA177 dgoT</i> ::FRT <i>tus</i> ::FRT <i>topA</i> -Ins480::FRT
GJ16904 <sup>d</sup>	GJ12134 <i>rpoB</i> *35 <i>btuB</i> ::Tn10 <i>dnaA177 dgoT</i> ::FRT <i>tus</i> ::FRT <i>topA</i> -480 ::FRT
GJ16921 <sup>d</sup>	GJ13519 <i>topA</i> -Ins480::FRT
GJ17776 <sup>d</sup>	GJ16819 <i>topA</i> ::FRT
GJ17777 <sup>d</sup>	GJ16819 <i>topA</i> -Ins480::FRT
GJ17778 <sup>d</sup>	GJ16819 <i>topA</i> -480 ::FRT

GJ17783 <sup>d</sup>	GJ13519 <i>tus::FRT rpoB*35 btuB::Tn10 topA::FRT</i>
GJ17784 <sup>d</sup>	GJ13519 <i>tus::FRT rpoB*35 btuB::Tn10 topA-Ins480::FRT</i>
GJ17786 <sup>d</sup>	GJ17783 <i>dnaA::Kan</i>
GJ17787 <sup>d</sup>	GJ17784 <i>dnaA::Kan</i>
GJ17788 <sup>d</sup>	GJ13519 <i>tus::FRT rpoB*35 btuB::Tn10 topA-480 ::FRT dnaA::Kan</i>
GJ17789 <sup>d</sup>	GJ13519 <i>tus::FRT rpoB*35 btuB::Tn10 dnaA::FRT</i>
GJ17790 <sup>d</sup>	GJ17789 <i>topA::Kan</i>
GJ17791 <sup>d</sup>	GJ17789 <i>topA-Ins480::Kan</i>
GJ17792 <sup>d</sup>	GJ17789 <i>topA-480 ::Kan</i>
GJ18601	GJ13519 80 free
GJ18904 <sup>d</sup>	GJ17776 <i>dnaA::FRT</i>
GJ18905 <sup>d</sup>	GJ17777 <i>dnaA::FRT</i>
GJ18906 <sup>d</sup>	GJ17778 <i>dnaA::FRT</i>
GJ18907 <sup>d</sup>	GJ18904 <i>tus::Kan</i>
GJ18908 <sup>d</sup>	GJ18905 <i>tus::Kan</i>
GJ18909 <sup>d</sup>	GJ18906 <i>tus::Kan</i>
GJ18910	GJ18601 <i>rpoB*35 btuB::Tn10</i>
GJ18918 <sup>d</sup>	GJ18910 <i>topA-Ins480::FRT</i>
GJ18921 <sup>d</sup>	GJ18918 <i>dinG::Kan</i>
GJ18940 <sup>d</sup>	GJ17783 <i>dnaA::FRT</i>
GJ18941 <sup>d</sup>	GJ17784 <i>dnaA::FRT</i>
GJ18942 <sup>d</sup>	GJ13519 <i>tus::FRT rpoB*35 btuB::Tn10 topA-480 ::FRT dnaA::FRT</i>
GJ18945 <sup>d</sup>	GJ18918 <i>tus::Kan</i>
GJ18951 <sup>d</sup>	GJ18941 <i>rnhA::Kan</i>
GJ18952 <sup>d</sup>	GJ18941 <i>dinG::Kan</i>
GJ18976 <sup>d</sup>	GJ12134 <i>topA::FRT</i>
GJ18977 <sup>d</sup>	GJ12134 <i>topA-Ins480::FRT</i>
GJ18983 <sup>d</sup>	GJ18940 <i>rnhA::Kan</i>
GJ18997 <sup>d</sup>	GJ13519 <i>dnaA177 dgoT::FRT rpoB*35 btuB::Tn10 tus::FRT topA::FRT</i>
GJ18999 <sup>d</sup>	GJ13519 <i>dnaA177 dgoT::FRT rpoB*35 btuB::Tn10 tus::FRT topA-480 ::FRT</i>
GJ19608 <sup>d</sup>	GJ17783 <i>rnhA::Kan</i>
GJ19609 <sup>d</sup>	GJ17784 <i>rnhA::Kan</i>

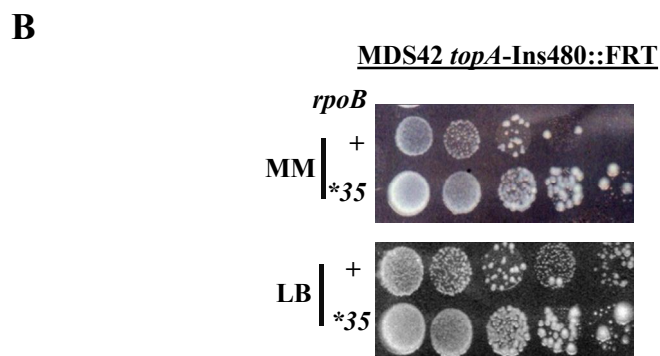
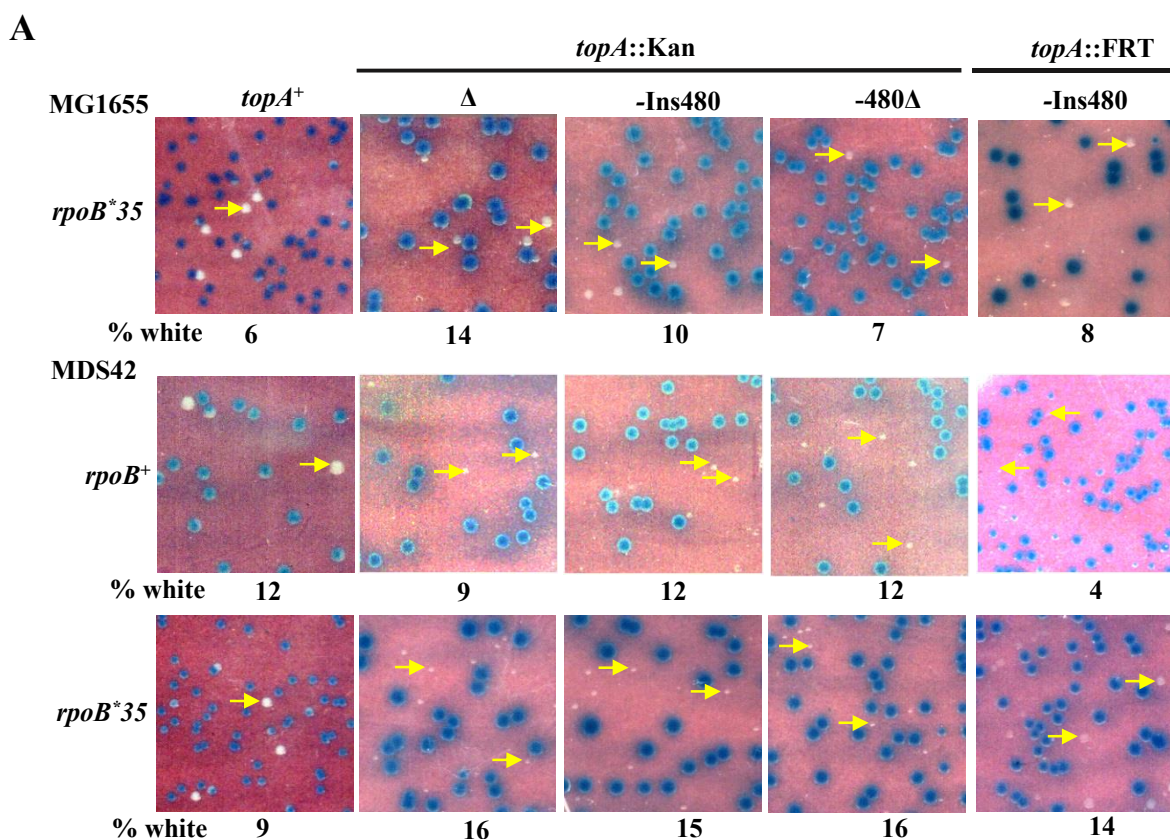
83 <sup>a</sup> Strain MG1655 was from our laboratory collection. Strains MDS42 (3),  
84 GJ12134 (4), GJ13519 (1), GJ13531 (5), and GJ16475 (1) have been  
85 described earlier; all other strains were constructed in this study.

86 <sup>b</sup> The following alleles and constructs have been described earlier:  $\Delta(\text{argF-}$   
87  $\text{lac})U169$  (6);  $\text{dnaA177}$  (7); Keio deletions of  $\text{rnhA}$ ,  $\text{tus}$ ,  $\text{dinG}$ ,  $\text{dgoT}$ ,  $\text{uvrD}$ , and  
88  $\text{tus}$  (8);  $\text{rpoB}^*35$  and  $\text{btuB}::\text{Tn10}$  (1); and  $\text{att } \lambda::\text{P}_{\text{tac}}\text{-UvsW(Amp}^{\text{r}})$  (4).

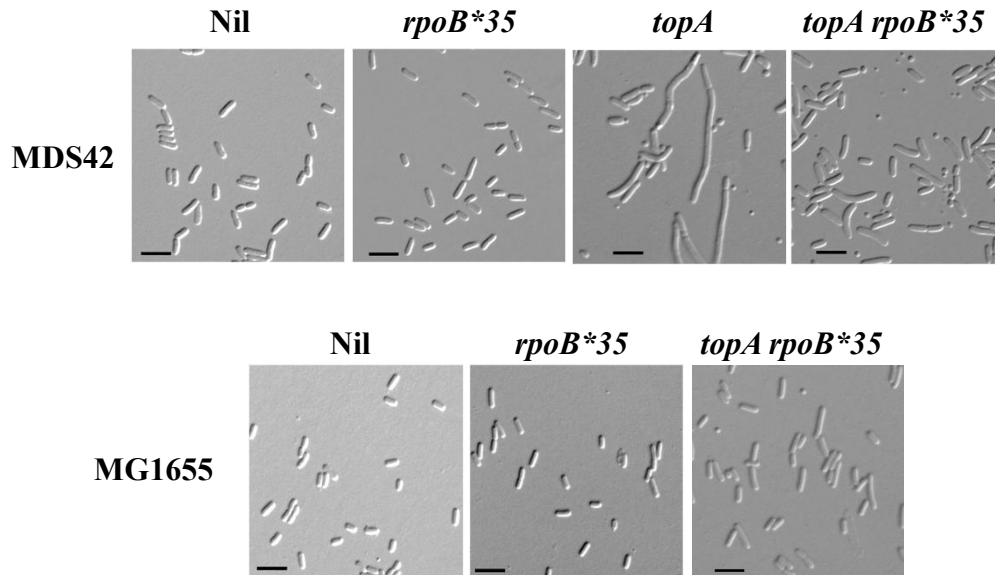
89 <sup>c</sup> Strain GJ13531 was routinely maintained as derivative with the shelter  
90 plasmid pHYD2411.

91 <sup>d</sup> The indicated strains were routinely maintained as derivatives with the  
92 shelter plasmid pHYD2390.

93 <sup>e</sup> Strain GJ16475 was routinely maintained as derivative with the shelter  
94 plasmid pHYD2388.



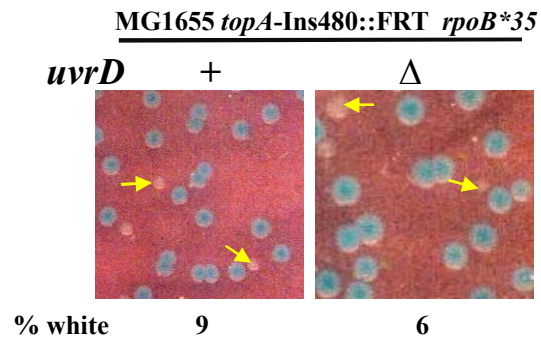
**Supplementary Figure S1. (A)** Blue-white screening assay on glucose-minimal A medium, of MG1655 or MDS42 strain derivatives with the *topA*<sup>+</sup> shelter plasmid pHYD2390 and the different *topA* alleles as indicated on the top of each column; the *rpoB* allele status is indicated at left of each row. Examples of white colonies are marked by the yellow arrows. From left to right, strains used for the panels were pHYD2390 derivatives of: row 1, GJ16703, GJ16813, GJ16814, GJ16815 and GJ16854; row 2, GJ12134, GJ16816, GJ16817, GJ16818 and GJ 18977; and row 3, GJ16819, GJ16820, GJ16821, GJ16822 and GJ17777. **(B)** Serial dilution-spotting of *rpoB*<sup>+</sup> and *rpoB*<sup>\*35</sup> derivatives of MDS42 *topA*-Ins480::FRT (GJ18977 and GJ17777, respectively) on LB and MM.



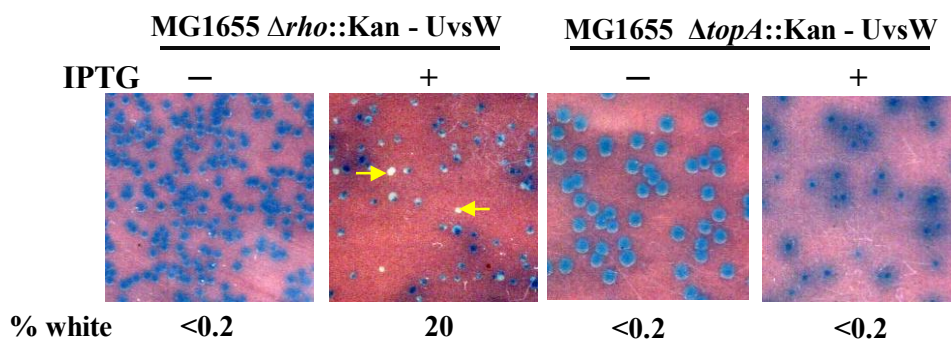
**Supplementary Figure S2.** Cell morphology in derivatives of MDS42 and MG1655 (top and bottom rows, respectively), visualized by differential interference contrast microscopy. Scale bar (at bottom left of each panel), 5  $\mu\text{m}$ . Relevant mutations in the strains are indicated on top of the corresponding panels. The *topA* allele in MDS42 derivatives was  $\Delta\textit{topA}::\text{Kan}$  and that in the MG1655 derivative was *topA-Ins480::Kan*. Strains used were (from left to right): top row, GJ12134, GJ16819, GJ16816, and GJ16820; and bottom row, GJ13519, GJ16703, and GJ16814.



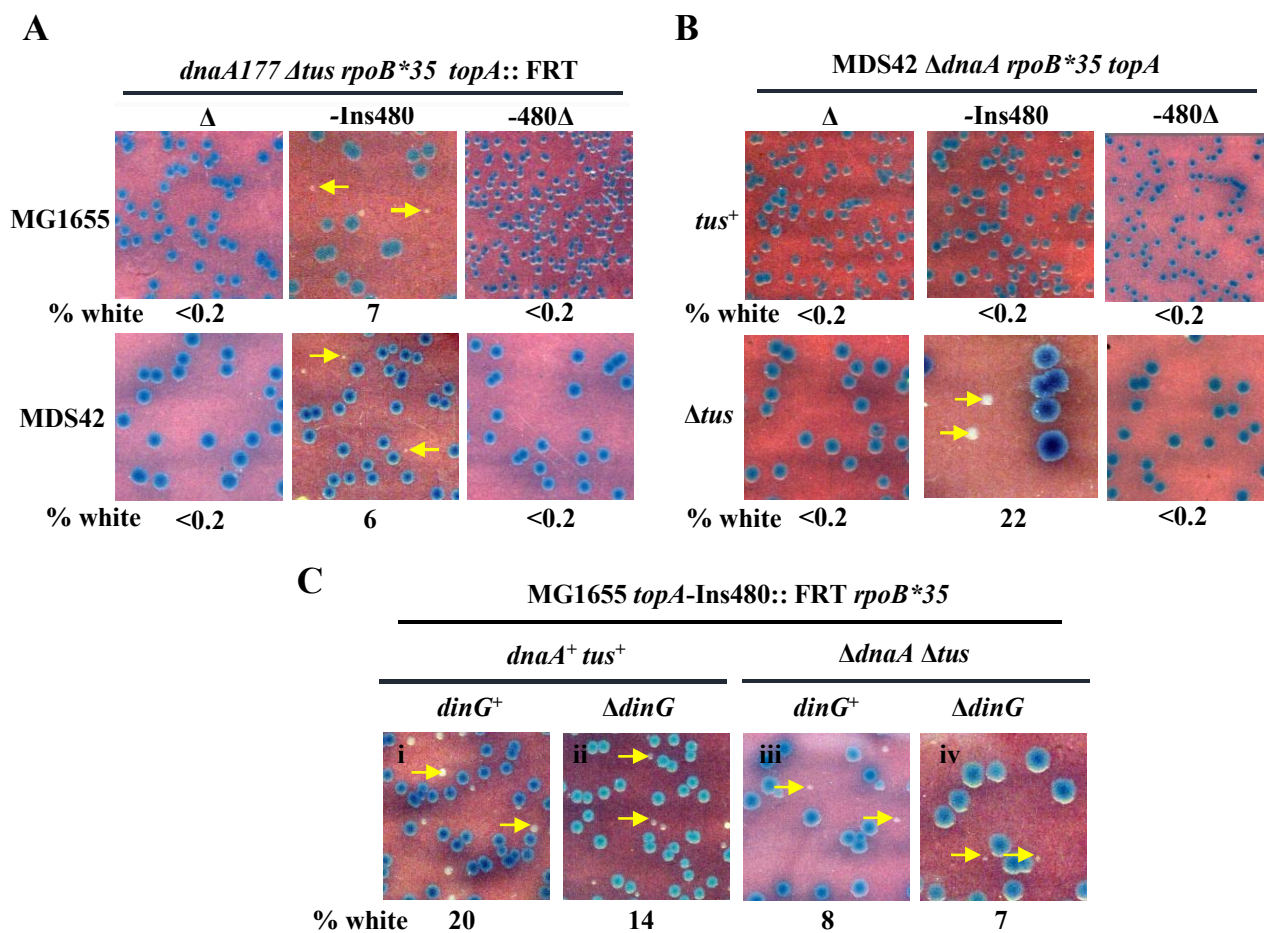
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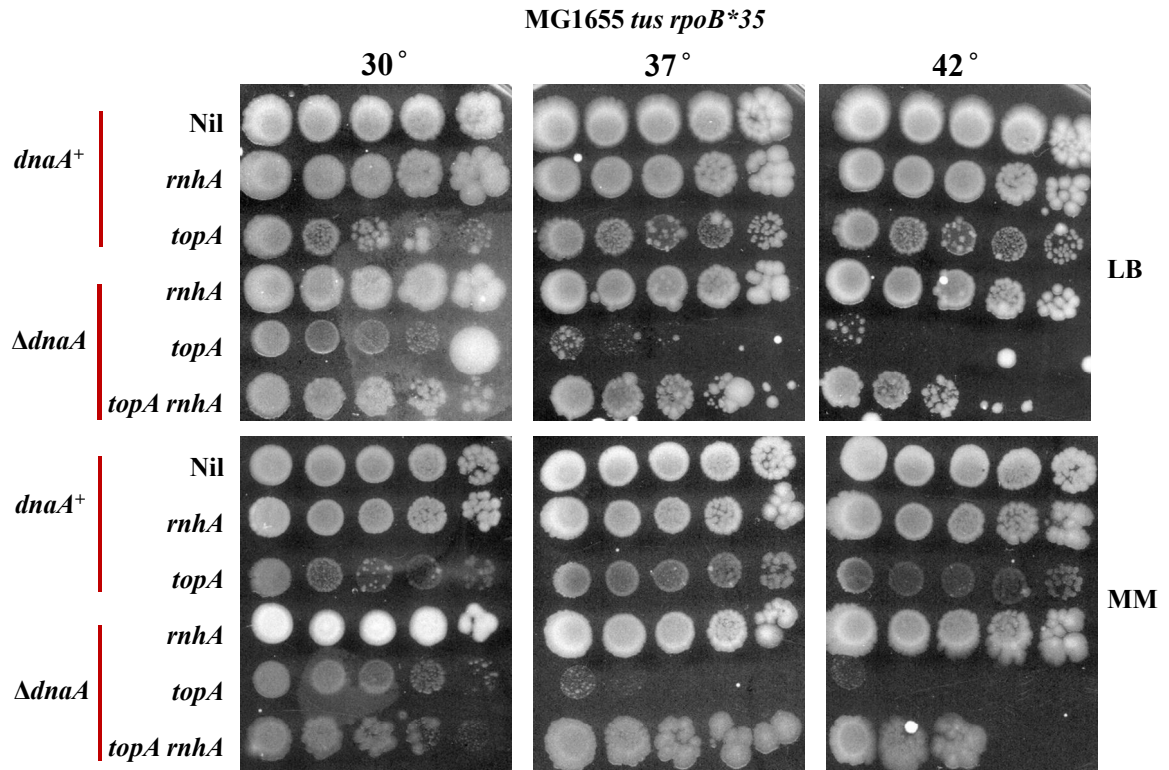
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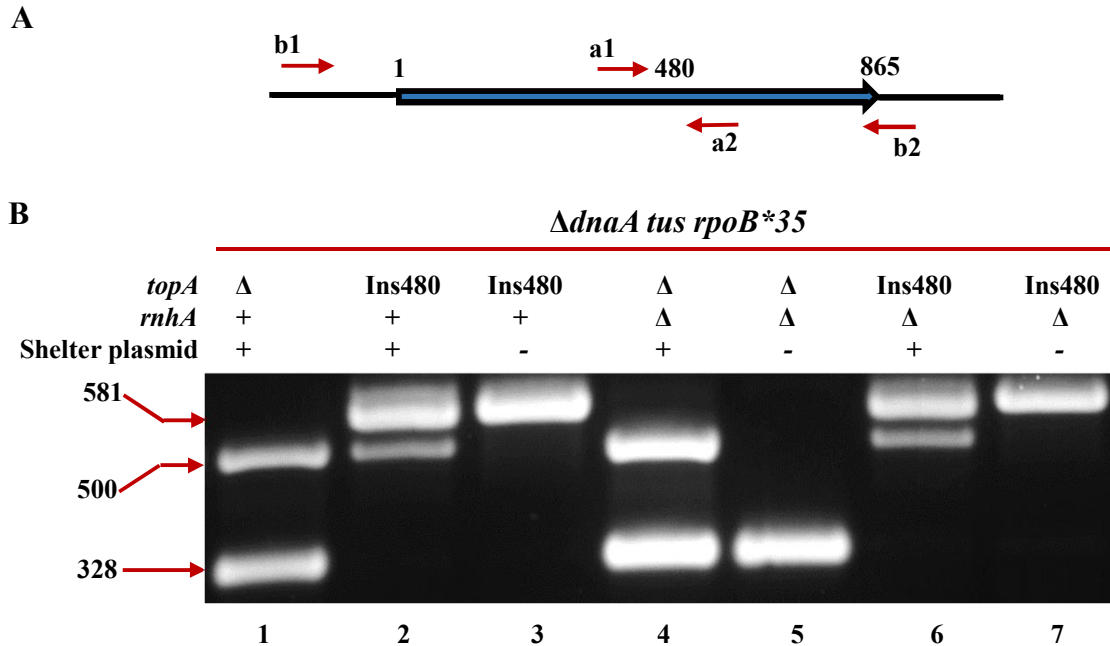
**Supplementary Figure S3.** Blue-white screening assays with MG1655 derivatives (carrying cognate shelter plasmids) on glucose-minimal A medium to demonstrate that *topA* lethality rescue by *rpoB*\*35 is UvrD-independent (**A**), and that UvsW expression rescues Δ*rho* lethality but not Δ*topA* lethality (**B**). Relevant genotypes are indicated on top of each of the panels. Examples of white colonies are marked by the yellow arrows. UvsW expression from a chromosomal  $P_{tac}$ -UvsW construct was regulated by the omission (-) or addition (+) of IPTG (latter at 100 μM and 40 μM, respectively, for the *rho* and *topA* mutants). Strains used were: (**A**) pHYD2390 derivatives of GJ16854 (*uvrD*<sup>+</sup>) and GJ16880 (Δ*uvrD*); and (**B**) pHYD2411 derivative of GJ13531 (Δ*rho*) and pHYD2390 derivative of GJ15697 (Δ*topA*).



**Supplementary Figure S4.** Features of *dnaA* lethality rescue by *topA* alleles. **(A-C)** Blue-white screening assays were performed at 30° on glucose-minimal A with *topA*<sup>+</sup> *dnaA*<sup>+</sup> shelter plasmid pHYD2390 in strain derivatives of MG1655 or MDS42 whose relevant genotypes are indicated on top of each of the panels. Examples of white colonies are marked by the yellow arrows. From left to right, strains employed were pHYD2390 derivatives of: **A** top row, GJ18997, GJ15630 and GJ18999; **A** bottom row, GJ16902, GJ16903 and GJ16904; **B** top row, GJ18904, GJ18905 and GJ18906; **B** bottom row, GJ18907, GJ18908 and GJ18909; and **C**, GJ18918, GJ18921, GJ18941 and GJ18952.



**Supplementary Figure S5.** Serial dilution-spotting on LB and glucose-minimal A (MM) at indicated temperatures of derivatives of MG1655  $\Delta$ *tus rpoB\*35* with additional relevant alleles as indicated at left. The  $\Delta$ *dnaA*, *topA* and *rnhA* alleles were, respectively,  $\Delta$ *dnaA*::FRT, *topA*-Ins480::FRT, and  $\Delta$ *rnhA*::Kan. The six strains used were (from top): GJ17784/pHYD2390; GJ16475/pHYD2388; GJ17784; GJ16475; GJ18941; and GJ18951.



**Supplementary Figure S6.** PCR validation of chromosomal *topA* genotype in *ΔdnaA* derivatives without or with *rnhA* mutation. **(A)** Schematic depiction of *topA* ORF (865 codons); positions of PCR primer pairs a1-a2 (5'-TGTACCAGTTAATCTGGCGTCAG-3' and 5'-TCGTGATTTGCCACCTGGTCGAG-3', respectively) flanking the codon 480 region, and b1-b2 (sequences given in Supplementary Methods above) for the entire ORF, are marked. The former primer pair is expected to yield amplicons of size 500 bp and 581 bp from *topA*<sup>+</sup> and *topA*-Ins480::FRT, respectively, and no amplicon from *ΔtopA*::FRT. The latter primer pair is expected to yield amplicons of size 2.8 kb and 328 bp from *topA*<sup>+</sup> and *ΔtopA*::FRT, respectively; however, the conditions used for PCR amplification were such as not to yield the large-size amplicons such as of 2.8 kb length. **(B)** Amplicons detected by gel electrophoresis after PCR with both primer pairs together, on DNA preparations from strains of genotype indicated on top of each lane. The *topA*<sup>+</sup> *dnaA*<sup>+</sup> shelter plasmid was pHYD2390. Allele designations used for *topA*: Δ, *ΔtopA*::FRT and 480, *topA*-Ins480::FRT; and for *rnhA*: Δ, *ΔrnhA*::Kan. Migration positions of fragments of length 328, 500 and 581 bp are marked. Strains employed for the different lanes were: 1, GJ18940/pHYD2390; 2, GJ18941/pHYD2390; 3, GJ18941; 4, GJ18983/pHYD2390; 5, GJ18983; 6, GJ18951/pHYD2390; and 7, GJ18951.