

## **SUPPLEMENTARY FIGURE LEGENDS**

### **Suppl. Fig. 1. Syncrip transcript isoform quantification**

Transcript specific reanalysis of a published RNA sequencing dataset in rat cortical neurons (DIV7) (26). Violine plots show that transcript ENSRNOT00000074515 (Ensembl\_ID - corresponding to isoform X3 in the NCBI annotation (XM\_006243515)) is the highest expressed among Ensembl annotated Syncrip isoforms

### **Suppl. Fig. 2. Pie chart representing the distribution of iCLIP peaks (Piranha bin 20 – at least 2 replicates) in the defined regions of the complete rat genome (Rnor\_6.0).**

Different regions of NCBI RefSeq genes are indicated as 5' UTR, coding exons, introns and 3'UTR. Peaks mapping outside NCBI RefSeq genes are divided in two regions – intergenic and 1000 bp downstream of 3'UTR end.

### **Suppl. Fig. 3. Gene Ontology analysis of SYNCRIP targets**

Gene ontology (GO) analysis of molecular function (MF, upper panel) and cellular component (CC, lower panel) terms enriched in SYNCRIP iCLIP target genes. Displayed are the top 15 MF and CC terms based on statistical enrichment (P-values). The number of SYNCRIP iCLIP target genes in each term is highlighted on the right.

**Suppl. Fig. 4. A snapshot from NCBI genomic data viewer showing detailed mapping profile of iCLIP unique reads and peaks within selected SYNCRIP iCLIP target genes (DCX, Cd24 and Syt4).**

Total unique reads from only one replicate of SYNCRIP iCLIP were used to visualize mapping profile of iCLIP unique reads to rat genome (Rnor\_6.0), as an example (unique reads panel). Positions of significant peaks (Piranha peaks, bin 20 – at least 2 replicates) and a number of unique reads identified within each peak are provided as a separate panel.

**Suppl. Fig. 5. Schematic representation of luciferase reporter genes containing 2x miR-9 and 2x Let7 binding sites in the 3' UTR.**

Sequence of each miRNA binding site (black) together with its complementary mature miRNA (green) are provided. Mutations disrupting the binding of mature miRNA to the respective 3'UTR are indicated as a red box.

**Suppl. Fig. 6. Depletion of SYNCRIP leads to upregulation of luciferase reporter lacking poly-A tail.**

Luciferase reporter gene assays in rat cortical neurons (DIV 6-9) transfected with the indicated 2xmiR binding site (BS) reporters, containing histone stem loop (HSL) and hammerhead ribozyme (HhR), and shRNA constructs. Relative luciferase activity (firefly/renilla activity) is presented as mean  $\pm$  SD (n=4). For each reporter gene, results from co-transfection of a vector alone (without shRNA) were set to one. Statistical test is performed using two-way ANOVA and corrected for multiple

comparison with Tukey's post-hoc test (2x miR-9 BS-HSL-HhR:  $P < 0.05$ , 2x miR-9 BS-mut-HSL-HhR:  $P = \text{n.s.}$ ; BS:  $F(1, 12) = 3.039$ ,  $P = 0.107$ ; shRNA:  $F(1, 12) = 10.81$ ,  $P = 0.007$ ; BS x shRNA:  $F(1, 12) = 2.183$ ,  $P = 0.165$ ).

**Suppl. Fig. 7. Regulation of miR-9 and Let7 BS luciferase reporter expression by miRNAs occurs mainly at the level of mRNA translation.**

Rat cortical neurons (DIV 6) were transfected with the indicated 2xmiR binding site (BS) reporters and lysed at DIV 9 either for RNA extraction or luciferase activity measurements. RNA levels of luciferase reporters were determined using real-time PCR. Relative luciferase activity (firefly/renilla activity) and RNA levels are presented as mean  $\pm$  SD ( $n=3$ ). Statistical test is performed using one-sample Student's t-test (p value, \*  $< 0.05$ , \*\*  $< 0.01$ ).

**Suppl. Fig. 8. miRNA binding site enrichment analysis**

Enrichment plots, highlighting the results of miRNA binding site depletion (left panel) and enrichment (right panel) analyses on SYNCRIP iCLIP target genes against a specific gene expression background. Circle sizes indicate the overlap of predicted miRNA and SYNCRIP target genes. Plotted are all miRNAs with an overlap of at least five and a p-value equal or below 0.05. y-axis –  $-\log_{10}$  transformed p-value (hypergeometric test on miRNA binding sites), x-axis – fold enrichment of miRNA binding sites against the background. miRNA expression levels in DIV7 cortical neuronal cultures, as reported in (21), are color-coded.

## **SUPPLEMENTARY TABLES:**

**Suppl. Table 1.** General analysis pipeline of iCLIP sequencing

**Suppl. Table 2.** The list of all significant peaks ( $< 0.05$  FDR, present in at least 2 out of 3 replicates) identified using `-bin_size=1` parameter of Piranha software.

**Suppl. Table 3.** The list of all significant peaks ( $< 0.05$  FDR, present in at least 2 out of 3 replicates) identified using `-bin_size=20` parameter of Piranha software.

**Suppl. Table 4.** The number of unique reads and peaks from Suppl. T2 mapping to different genomic positions.

**Suppl. Table 5.** The number of unique reads and peaks from Suppl. T3 mapping to different genomic positions.

**Suppl. Table 6.** The list of SYNCRIP iCLIP target genes based on data from Suppl. Table T3 that maps to 3'UTR.

## SUPPLEMENTARY MATERIAL:

### DNA oligonucleotides

#### *Cloning*

##### Into pEGFP-N1

fw-N1\_X3\_Sall: CTAGTCGACCGCCACCATGGCTACAGAACATGTTAATGGA

rev-N1\_X3\_AgeI: GATCACCGGTTGTAACAGGTCAGGACCGG

fw-N1\_X7\_Sall: CTAGTCGACCGCCACCATGAAGACTTACAGGCAGAGAGAGA

rev-N1\_X7\_AgeI: same as rev-N1\_X3\_AgeI

##### Into pEGFP-C1

fw-C1\_X6\_XhoI: CTAGCTCGAGCAATGAAGACTTACAGGCAGAGAGAGA

rev-C1\_X6\_XbaI: CTAGTCTAGACTACTTCCACTGTTGCCCAAAG

##### into pIS1:

fw-2xmiR-9 BS:

CATTAACAGCTGATCAACCAAAGACTCCGTTAACAGCTGATCAACCAAAGA

rev-2xmiR-9 BS:

CTAGTCTTTGGTTGATCAGCTGTTAACGGAGTCTTTGGTTGATCAGCTGTTAATG  
AGCT

fw-2xmiR-9 BS-mut:

CATTAACAGCTGATCAACCGTTCCTCCGTTAACAGCTGATCAACCGTTCA

rev-2xmiR-9 BS-mut:

CTAGTGAACGGTTGATCAGCTGTTAACGGAGTGAACGGTTGATCAGCTGTTAAT  
GAGCT

fw-2xLet7 BS (Hmga2):

CCAATCAAACACACTACTACCTCTTAAGTCCCAGTATACCTCATT

rev-2xLet7 BS (Hmga2):

CTAGAAATGAGGTATACTGGGACTTAAGAGGTAGTAGTGTGTTTTGATTGGAGC  
T

fw-2xLet7 BS-mut:

CCAATCAAACACACTACTAACGCTTAAGTCCCAGTATAACGCATT

rev-2xLet7 BS-mut:

CTAGAAATGCGTTATACTGGGACTTAAGCGTTAGTAGTGTGTTTTGATTGGAGC  
T

fw-HSL-HhR: CTAGACTAGTATCTCGAGGCGGCCGC

Rev-HSL-HhR: CTAGATCGATAAGCTTGATATCGAATTCCC

into pSuper:

fw1-SYNCRIP-shRNA-1: GATCCCCTGAGAAAGCTGGACCTA

fw2-SYNCRIP-shRNA-1: TATTCAAGAGATATAGGTCCAGCTTTCTCATTTTTA

rev1- SYNCRIP-shRNA-1: AGCTTAAAAATGAGAAAGCTGGACCTA

rev1- SYNCRIP-shRNA-1: TATCTCTTGAATATAGGTCCAGCTTTCTCAGGG

fw1-SYNCRIP-shRNA-2: GATCCCCGATCAGAAGAGGAAAGA

fw2-SYNCRIP-shRNA-2: AATTCAAGAGATTTCTTTCCTCTTCTGATCTTTTTA

rev1-SYNCRIP-shRNA-2: AGCTTAAAAAGATCAGAAGAGGAAAGA

rev2-SYNCRIP-shRNA-2: AATCTCTTGAATTTCTTTCCTCTTCTGATCGGG

fw1-Cyt-SYNCRIP-shRNA - GATCCCCGATCAGAGCTTATACCT

fw2-Cyt-SYNCRIP-shRNA - AATTCAAGAGATTAGGTATAAGCTCTGATCTTTTTA

rev1-Cyt-SYNCRIP-shRNA - AGCTTAAAAAGATCAGAGCTTATACCT

rev2-Cyt-SYNCRIP-shRNA - AATCTCTTGAATTAGGTATAAGCTCTGATCGGG

### *Real-time PCR*

fw-U6: CTCGCTTCGGCAGCACA

rev-U6: AACGCTTCACGAATTTGCGT

fw\_12sRNA: ATACCGCCATCTTCAGCAAAC

rev-12sRNA: TTCTTTCCGCTTCATTGGCTA

fw-GAPDH: GCCTTCTCTTGTGACAAAGTGGA

rev-GAPDH: CCGTGGGTAGAGTCATACTGGAA

fw-HIPK2: AACACAAGCAGCGTGCAGAT

rev-HIPK2: TGTTTTTGGAGGTGGCAGTG

fw-Dcl1: AAATTTGCTGTTGTCAAGGAA

rev-Dcl1: GACCTCGTTTTGGATCATGTGT

fw-Cd24: TCCTACCCACGCAGATTTATTG

rev-Cd24: GATTTGGGGCAGCAGAGATACT

fw-DCX: GGAGTGCGCTACATTTACT

rev-DCX: GTCTGAGGAACAGACATAGCTT

fw-Syt4: CGGGGTGAACTTCTGGTCTC  
rev-Syt4: TGCCGCGCTTTTAAGACAAC  
fw-Firefly: GCTCAGCAAGGAGGTAGGTG  
rev-Firefly: TCTTACCGGTGTCCAAGTCC  
fw-Renilla: TTATTGAATCGGACCCAGGA  
re-Renilla: TTGAGAACTCGCTCAACGAA

*RNA-pull down*

Templates for in vitro transcription

fw-TAG-pre: AATTTAATACGACTCACTATAG  
rev-TAG-pre: AGTGAGTCGTATTAATT  
fw1-TAG-2xmiR-124 BS: GGAGAATAGATAGTCTGTTTTATATCTTAAGAGTGC  
fw2-TAG-2xmiR-124 BS: CTTATTATGTTGCTGCCACATTGTGCCTTTA  
rev1-TAG-2xmiR-124 BS:  
TAAAGGCACAATGTGGGCAGCAACATAATAAGGCACTCTT  
rev2-TAG-2xmiR-124 BS: AAGATATAAAACAGACTATCTATTCTCCCTAT  
fw1-TAG-2xmiR-124 BS-mut: same as fw1-TAG-2xmiR-124 BS  
fw2-TAG-2xmiR-124 BS-mut: GCCATTATGTTGCTGCCACATTGTTCGGTTTA  
rev1-TAG-2xmiR-124 BS-mut:  
TAAACCGACAATGTGGGCAGCAACATAATGGCGCACTCTT  
rev2-TAG-2xmiR-124 BS-mut: same as rev2-TAG-2xmiR-124 BS  
fw1-TAG-2xmiR-9 BS: GGAGAATAGATAGTATTAACAGCTGATCAACC  
fw2-TAG-2xmiR-9 BS: AAAGACTCCGTTAACAGCTGATCAACCAAAGA  
rev1-TAG-2xmiR-9 BS:  
TCTTTGGTTGATCAGCTGTTAACGGAGTCTTTGGTTGATC  
rev2-TAG-2xmiR-9 BS: AGCTGTTAATACTATCTATTCTCCCTAT  
fw1-TAG-2xmiR-9 BS-mut: same as fw1-TAG-2xmiR-9 BS  
fw2-TAG-2xmiR-9 BS-mut: GTTCACTCCGTTAACAGCTGATCAACCGTTCA  
rev1-TAG-2xmiR-9 BS-mut:  
TGAACGGTTGATCAGCTGTTAACGGAGTGAACGGTTGATC  
rev2-TAG-2xmiR-9 BS-mut: same as rev2-TAG-2xmiR-9 BS

DNA adapter

5' ACTATCTATTCTCCC 3' biotin

*iCLIP (from Sutandy et al., 2016)*

L3 linker: /5rApp/AGATCGGAAGAGCGGTTCAG/3ddC/

RT-CLIP1: PHO/NNAACCNNAAGATCGGAAGAGCGTCGTGGATCCTGAACCGC

RT-CLIP2: PHO/NNACAANNAAGATCGGAAGAGCGTCGTGGATCCTGAACCGC

RT-CLIP3: PHO/NNATTGNNAAGATCGGAAGAGCGTCGTGGATCCTGAACCGC

RT-CLIP4: PHO/NNCGCCNNAAGATCGGAAGAGCGTCGTGGATCCTGAACCGC

RT-CLIP5: PHO/NNGCCANNAAGATCGGAAGAGCGTCGTGGATCCTGAACCGC

RT-CLIP6: PHO/NNGACTNNAAGATCGGAAGAGCGTCGTGGATCCTGAACCGC

Cut\_oligo: GTTCAGGATCCACGACGCTCTTCAAAA

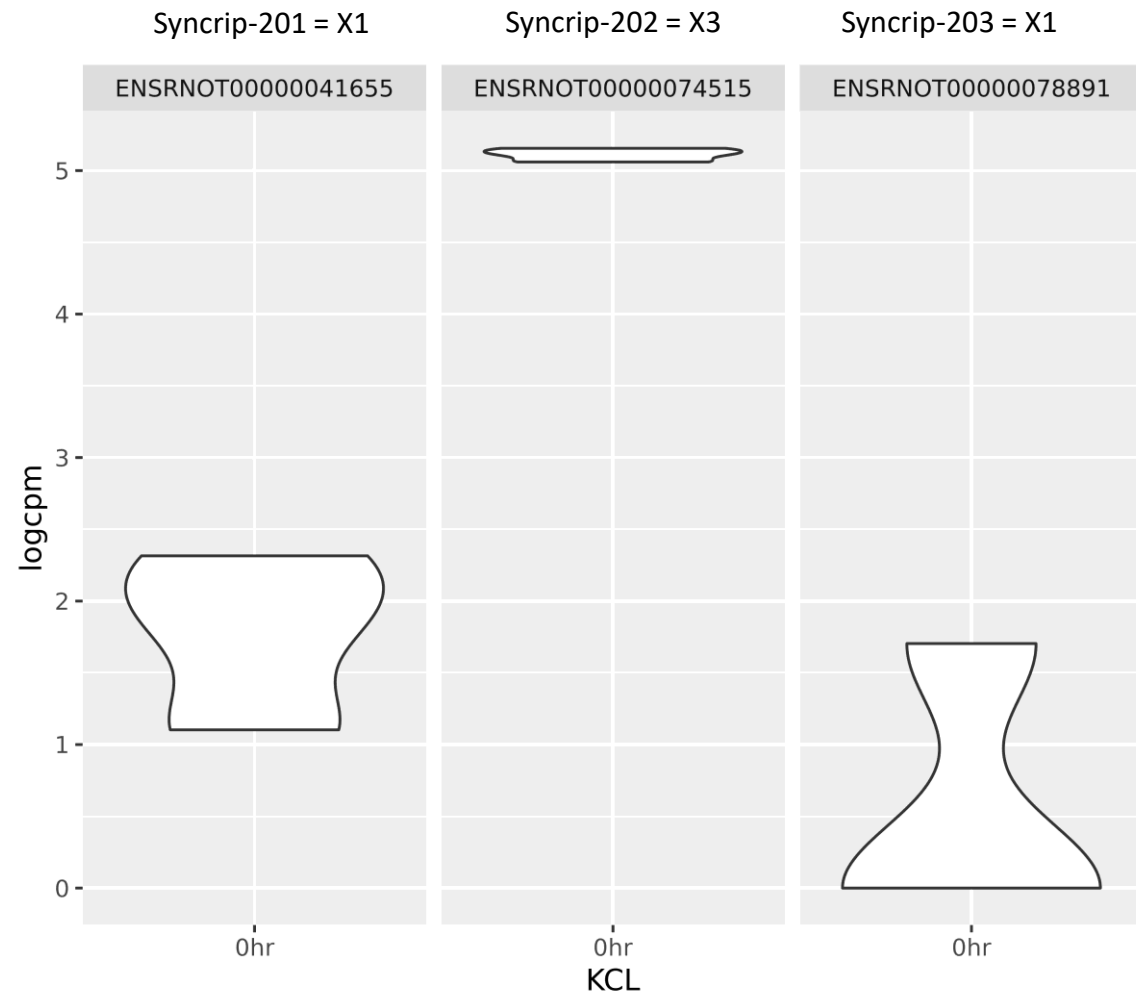
P5solexa:

AATGATACGGCGACCACCGAGATCTACACTCTTTCCCTACACGACGCTCTTCCG  
ATCT

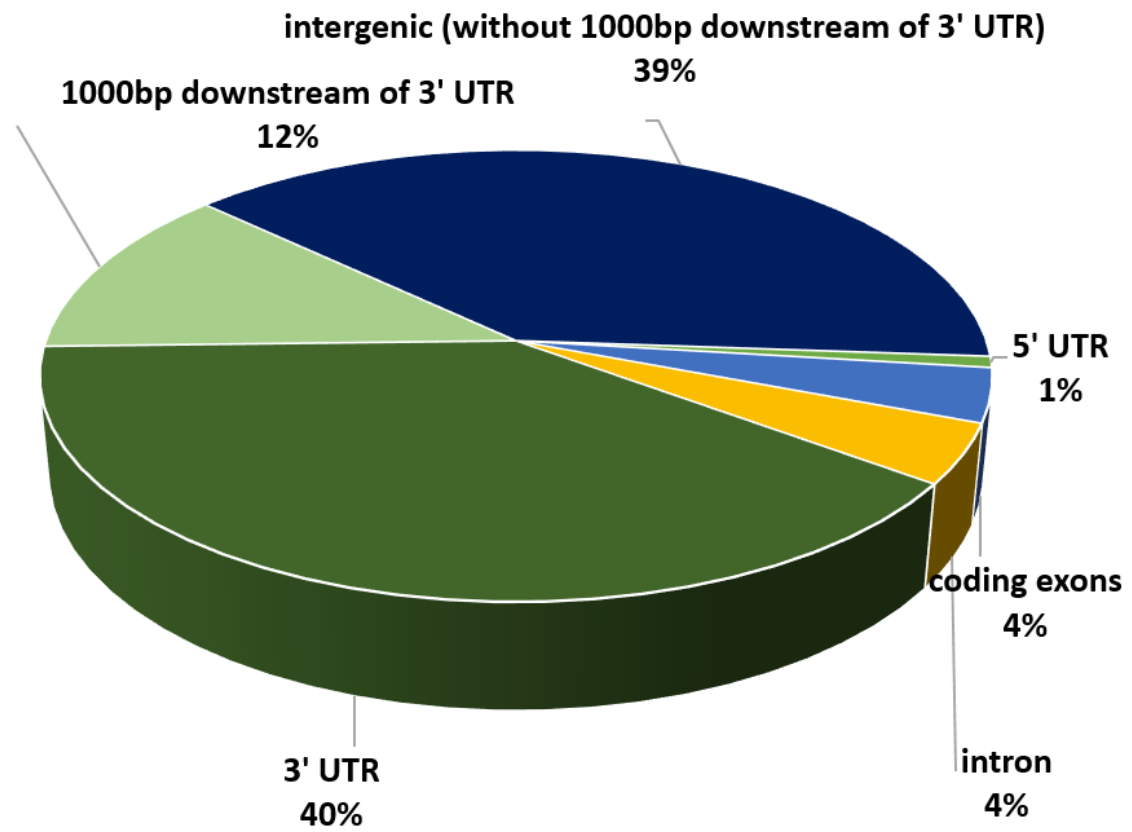
P3solexa:

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TCCGATCT

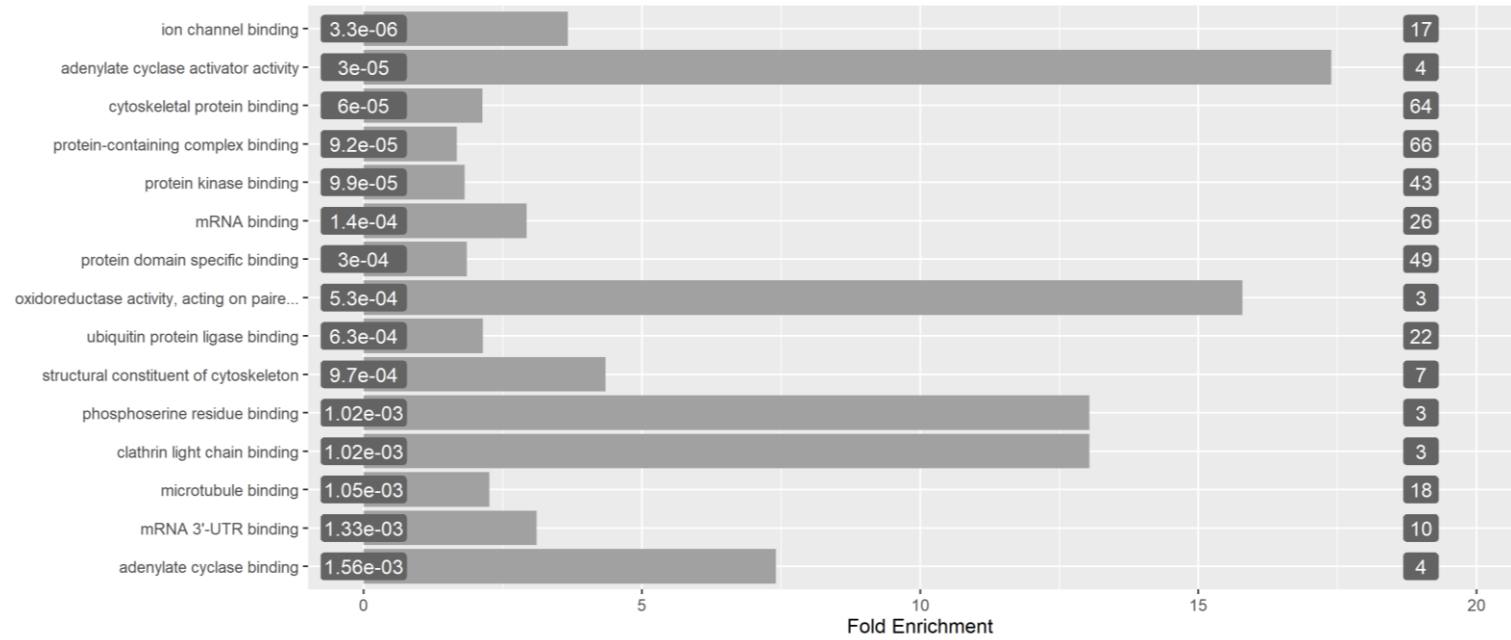




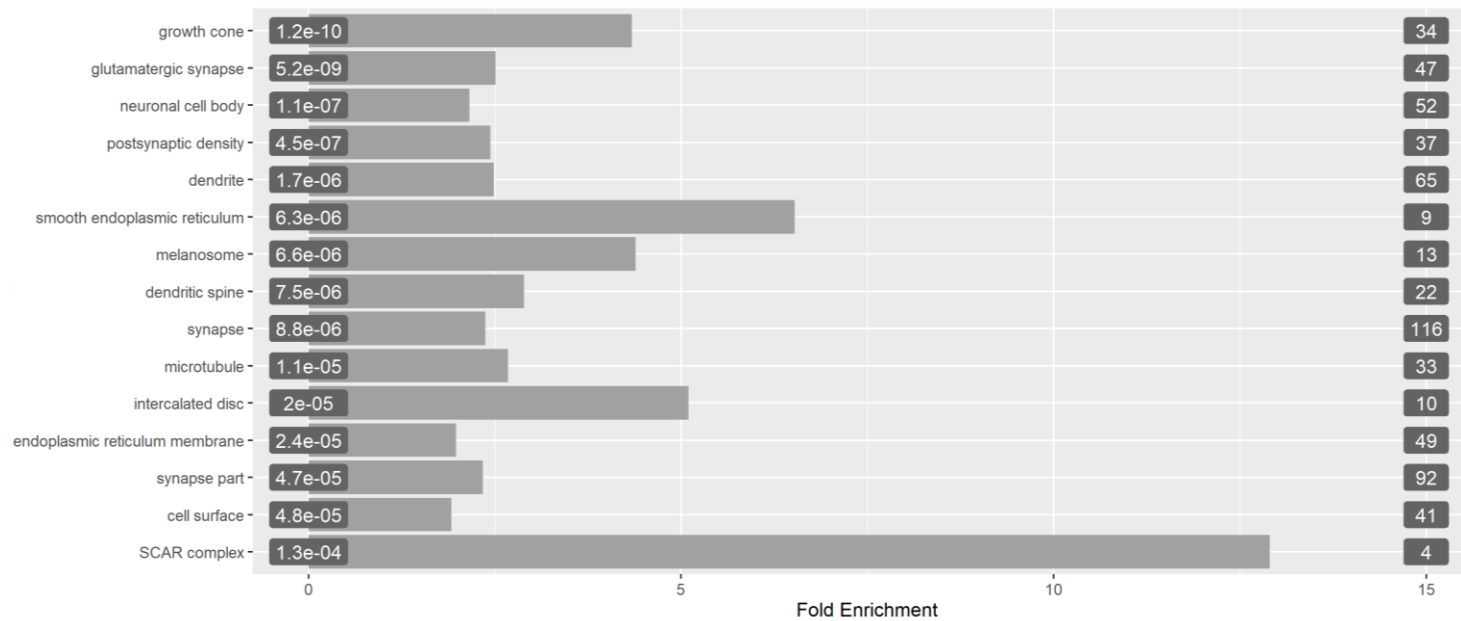
Suppl. Fig. 1



MF

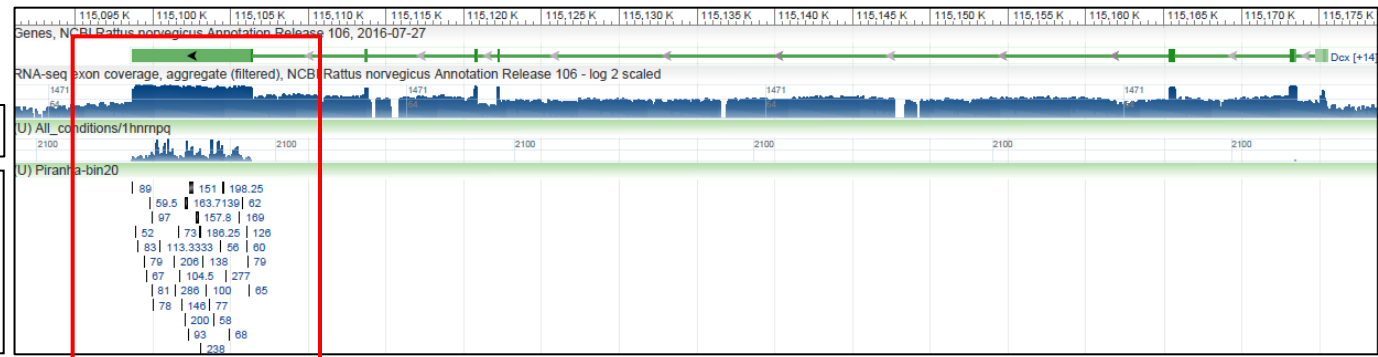


CC



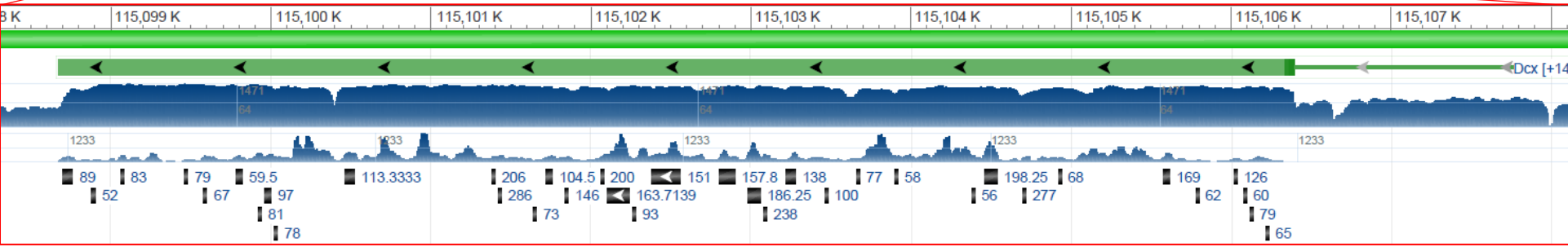
Suppl. Fig. 3

DCX



Unique reads

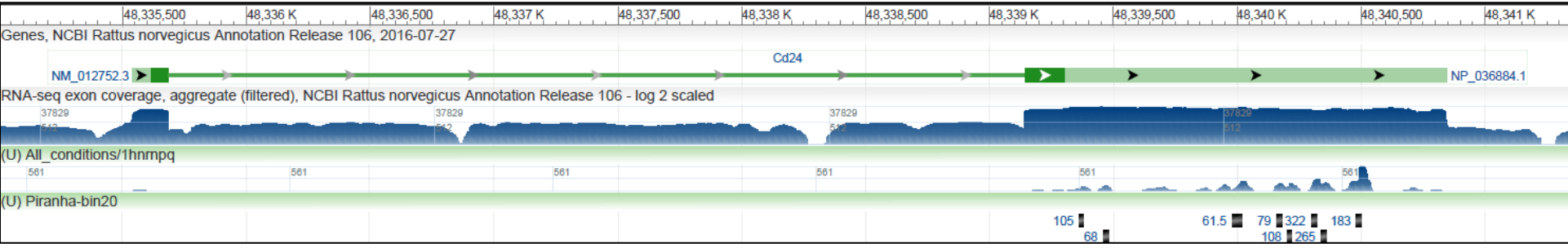
Piranha bin20 peaks  
(#unique reads/peak)



Unique reads

Piranha bin20 peaks  
(#unique reads/peak)

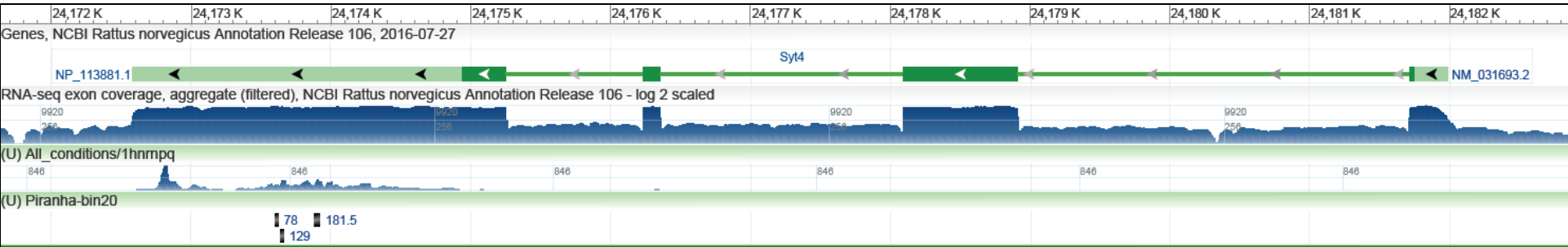
Cd24



Unique reads

Piranha bin20 peaks  
(#unique reads/peak)

Syt4



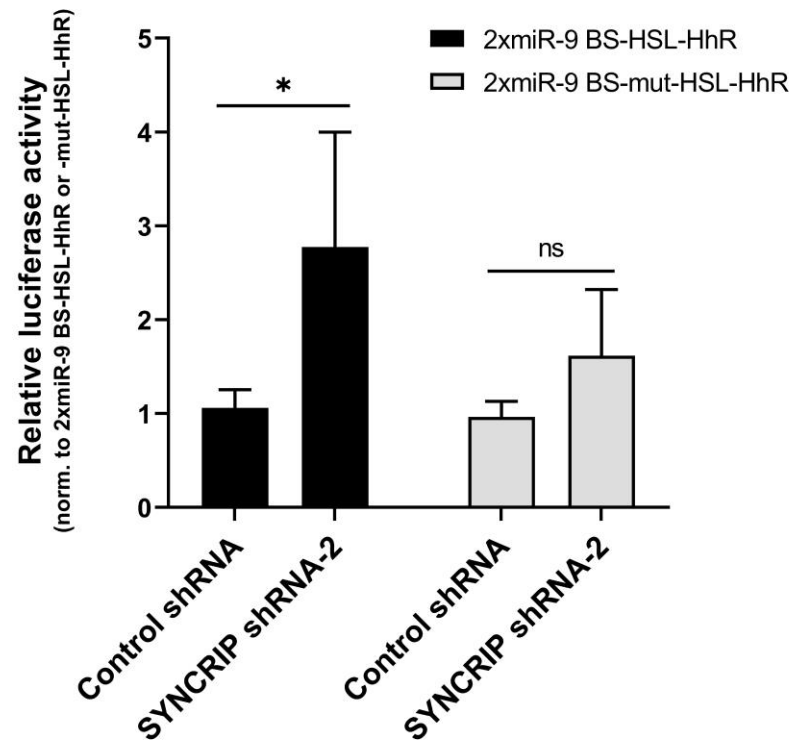
Unique reads

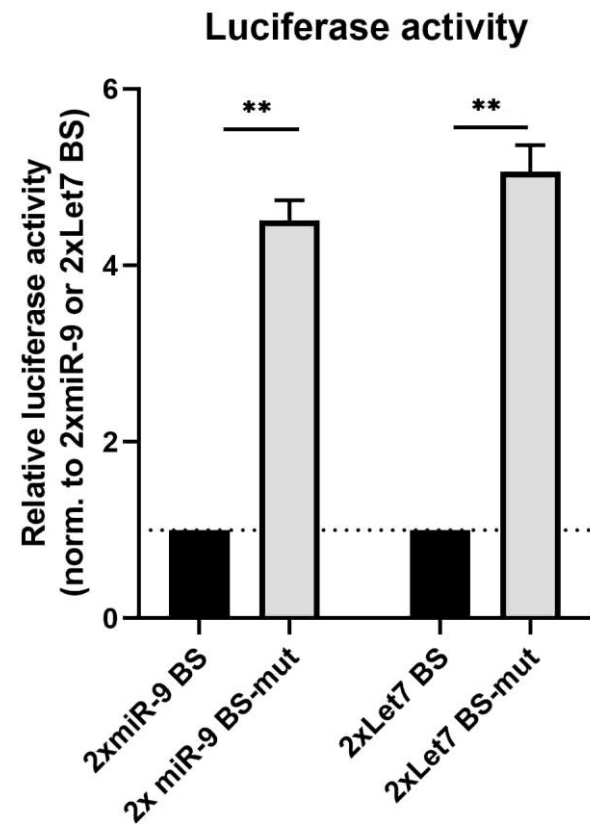
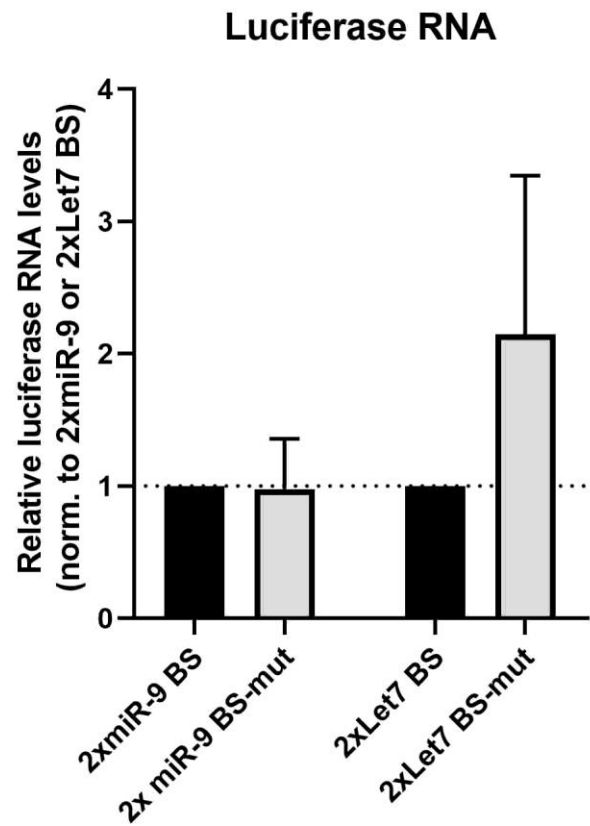
Piranha bin20 peaks  
(#unique reads/peak)

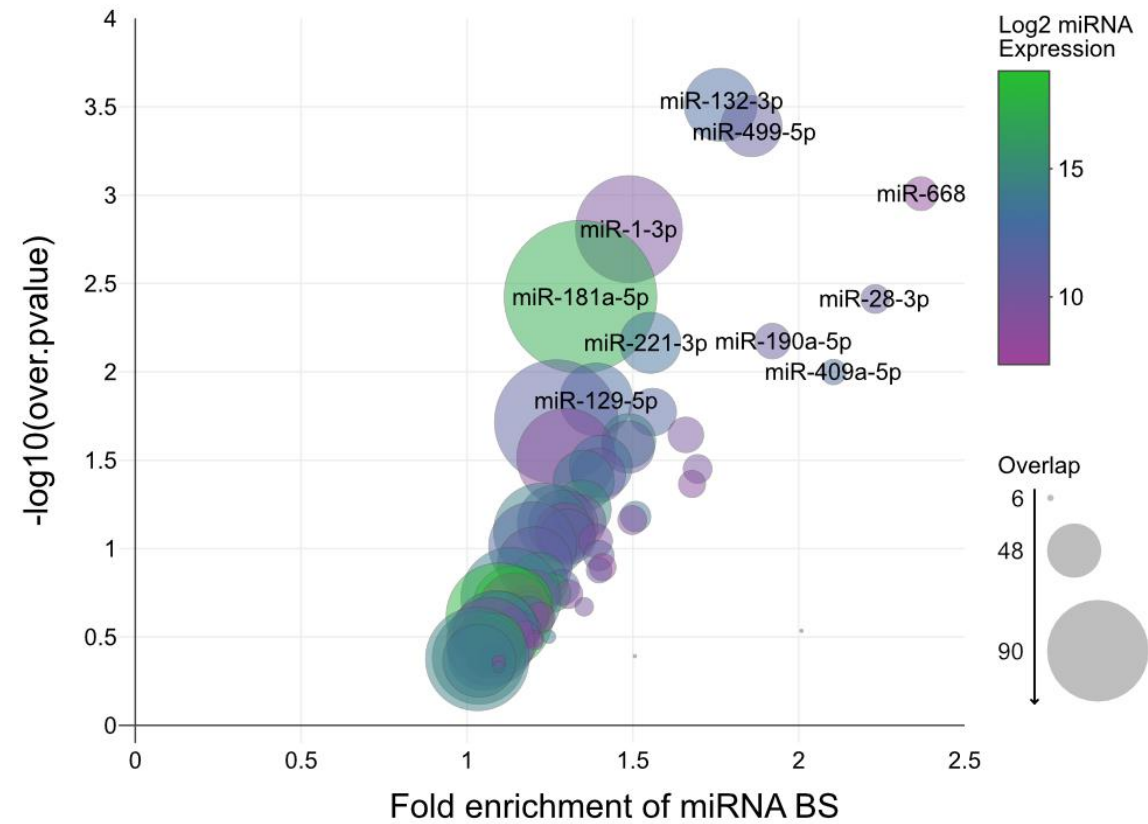
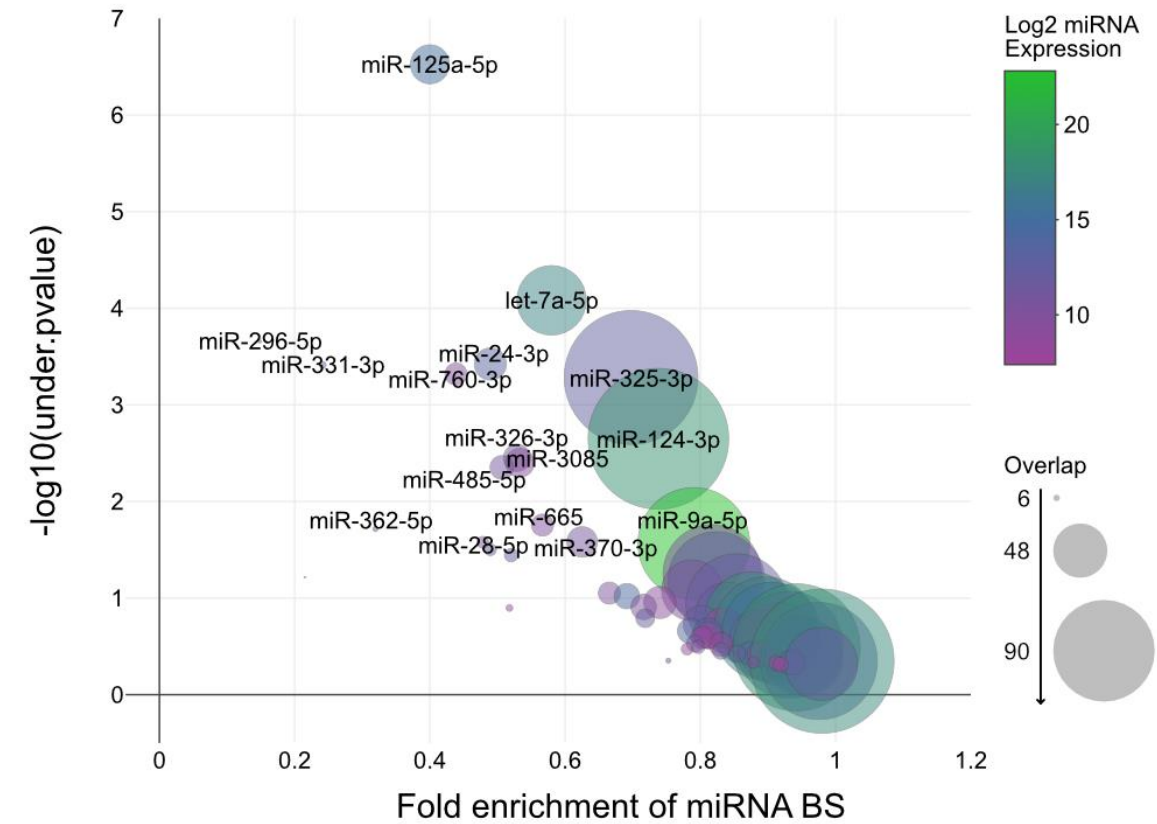
Suppl.  
Fig. 4



Suppl. Fig. 5







Suppl. Fig. 8