

Indoloxotriazines as Binding Molecules for the JAK2JH2 Pseudokinase Domain and Its V617F Variant

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Supporting Information

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1. Experimental Details

1.1 Database

The drug-like subset from ZINC12 database [1] containing more than 10 million compounds (10,637,968 – 2012-08-22) was used for the virtual screening. The compounds were processed with LigPrep (v 2.5) to generate three-dimensional structures with multiple conformers [2].

1.2 Virtual Screening

The database was employed in the docking calculations using the Glide 5.7.109 standard precision (SP) mode [3] to the crystal structure of JAK2 JH2 mutant V617F (PDBID:4FVR) [4]. Three hydrogen bond constraints with GLN626, GLU627 and VAL629 were added to increase binding affinity to the hinge area. The best ranked ca. 6400 were redocked with Glide XP [5], and 1K best complexes were visualized. The selection was based on the docking scores, on the favourable interactions with the receptor and also on the predictions of physical properties from QikProp [6]. Molecules containing unwanted structural features were removed (readily hydrolysable and/ or containing highly electrophilic functional groups).

1.3 Purchased Compounds

The identities of all purchased compounds were confirmed by ¹H NMR and high-resolution mass spectrometry; purity was normally > 95% as shown by high-performance liquid chromatography.

NMR spectra were recorded on Agilent DD2 600 (600 MHz), and DD2 400 (400 MHz) instruments. Mass determinations were performed using electrospray ionization on water Micromass ZQ (LC- MS) and on an Agilent Technologies 6890N (GC-MS). HRMS (ESI-TOF) analyses were performed on Waters Xevo QTOF equipped with Z-spray electrospray ionization source. The purity (≥ 95%) of all final synthesized compounds was determined by reverse phase HPLC, using a Waters 2487 dual λ absorbance detector with a Waters 1525 binary pump and a Phenomenex Luna 5μ C18(2) 250 x 4.6 mm column. Samples were run at 1 mL/min using gradient mixtures of 5-100% of water with 0.1% trifluoroacetic acid (TFA) (A) and 10:1 acetonitrile:water with 0.1% TFA (B) for 22 min followed by 3 min at 100% B.

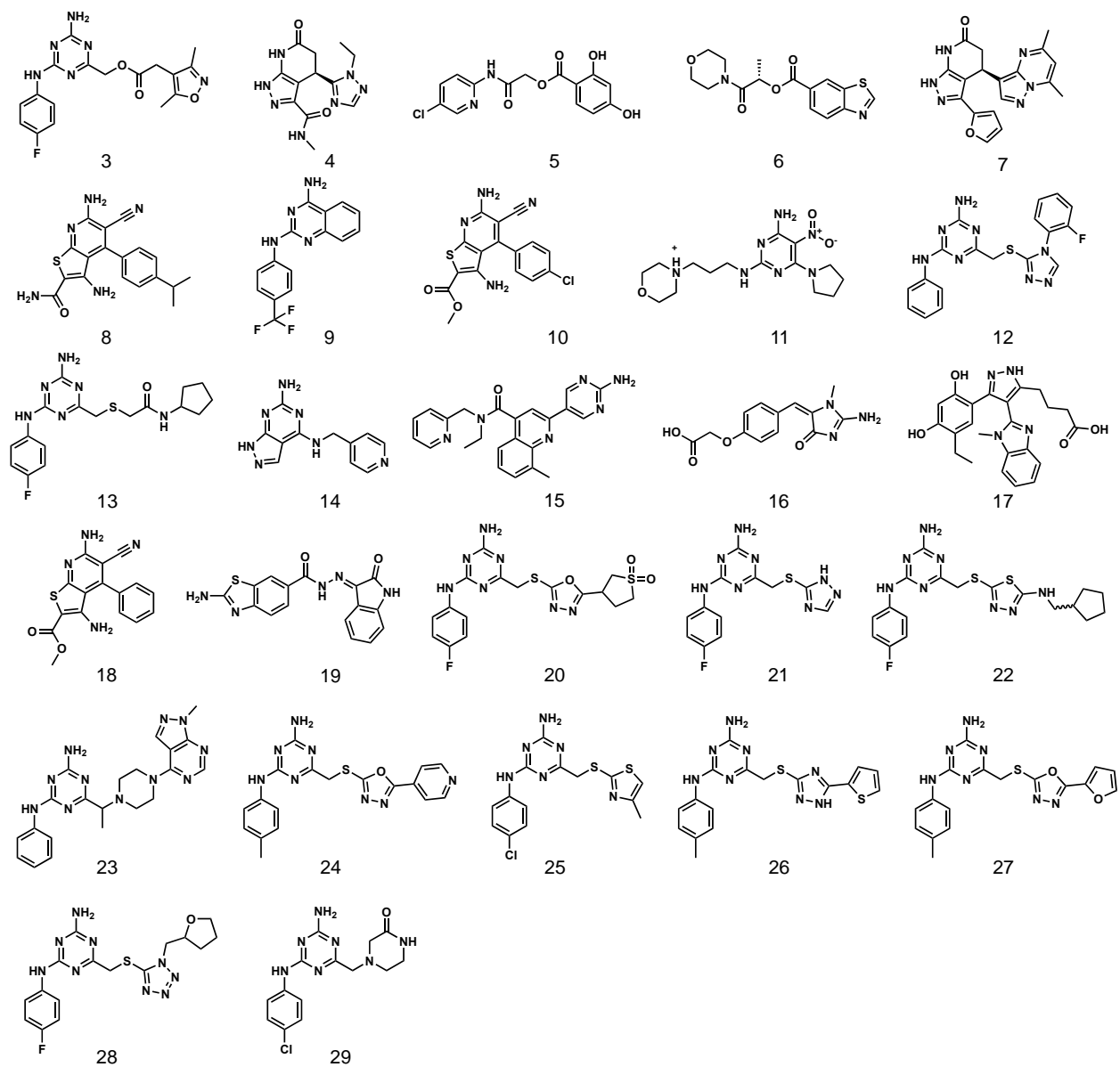


Figure ES1. Structures of the purchased (3-19) obtained from the virtual screening. Compounds 20-29 are analogues of 4, 12 and 15, found through SciFinder substructure-search, were also purchased and tested.

Table ES1. Experimentally binding affinities (K_d) using fluorescence polarization (FP) [7] against JAK2 JH1 WT, JAK2 JH2 WT and JAK2 JH2 VF, docking score and ZINC ID for the active compounds obtained from the virtual screening.

| Compound | K_d (μM) | | | Glide SP (kcal/mol) | ZINC ID |
|----------|-------------------------|------------|-----------|------------------------|----------|
| | JAK2 - WT | | JAK2-VF | | |
| | JH1 | JH2 | JH2 | | |
| 3 | NA | NA | 250 | -10.02 | 16675328 |
| 4 | NA | 300 | NA | -9.96 | 65372817 |
| 5 | 115 | NA | NA | -9.98 | 06080826 |
| 6 | NA | NA | 300 | -9.81 | 46804407 |
| 9 | 200 | 150 | 130 | -9.59 | 32617730 |
| 11 | NA | NA | 250 | -9.41 | 05111325 |
| 12 | NA | 92 | 40 | -9.76 | 05702514 |
| 13 | NA | 300 | 300 | -9.69 | 15495946 |
| 14 | NA | NA | 300 | -9.15 | 32511757 |
| 15 | 250 | 119 | 55 | -9.71 | 15495946 |
| 19 | NA | NA | 300 | -9.47 | 05706308 |
| 20 | NA | 292 | 75 | | |
| 21 | NA | 108 | 50 | | |
| 22 | NA | 149 | 65 | | |
| 23 | NA | 300 | 200 | | |
| 25 | 150 | 41 | 35 | | |
| 26 | 215 | 147 | 45 | | |
| 27 | 300 | 200 | 95 | | |
| 28 | NA | 85 | 45 | | |
| 29 | NA | 150 | 80 | | |

1.4 Fluorescence Polarization Assay

1.4.1 Determination of Tracer Affinity with JAK2

In a flat black bottom 96 well plate (Corning), the buffer (20 mM Tris-HCl pH 8.0, 150 mM NaCl, 20% Glycerol, 0.5 mM TCEP, 0.01% Tween 20) is added - 200 μL to column 1 (blank), 295 μL to column 2, 150 μL to columns 3-12. 5 μL of protein (179.0 μM JAK2-JH2-WT, 154.7 μM JAK2-JH2-V617F, 126.3 μM JAK2-JH1) were added to column 2. 150 μL was transferred, using a multichannel pipette, from column 2 to 3, 3 to 4, 4 to 5, until reaching the last column to make a serial dilution (1:2). 50 μl of 24.0 nM tracer were added from columns 2-12 and fluorescence polarization was measured at $\lambda_{\text{exc}} = 485 \pm 20$ nm, $\lambda_{\text{em}} = 535 \pm 25$ nm using an Infinite F500 plate reader until no FP variation was observed. From the lowest and highest FP values (tracer free and tracer fully bound to JAK) fraction of ligand bound to the protein to ligand total (L_b/L_t) was calculated for each concentration of the JAK2-JH2-WT, JAK2-JH2-V617F, and JAK2-JH1 (Figure S2B). Experiments were carried out by quadruplicates in three independent experiments. The data provided a typical saturation-binding curve and K_d was calculated fitting the results to the Hill equation using Prism 7.

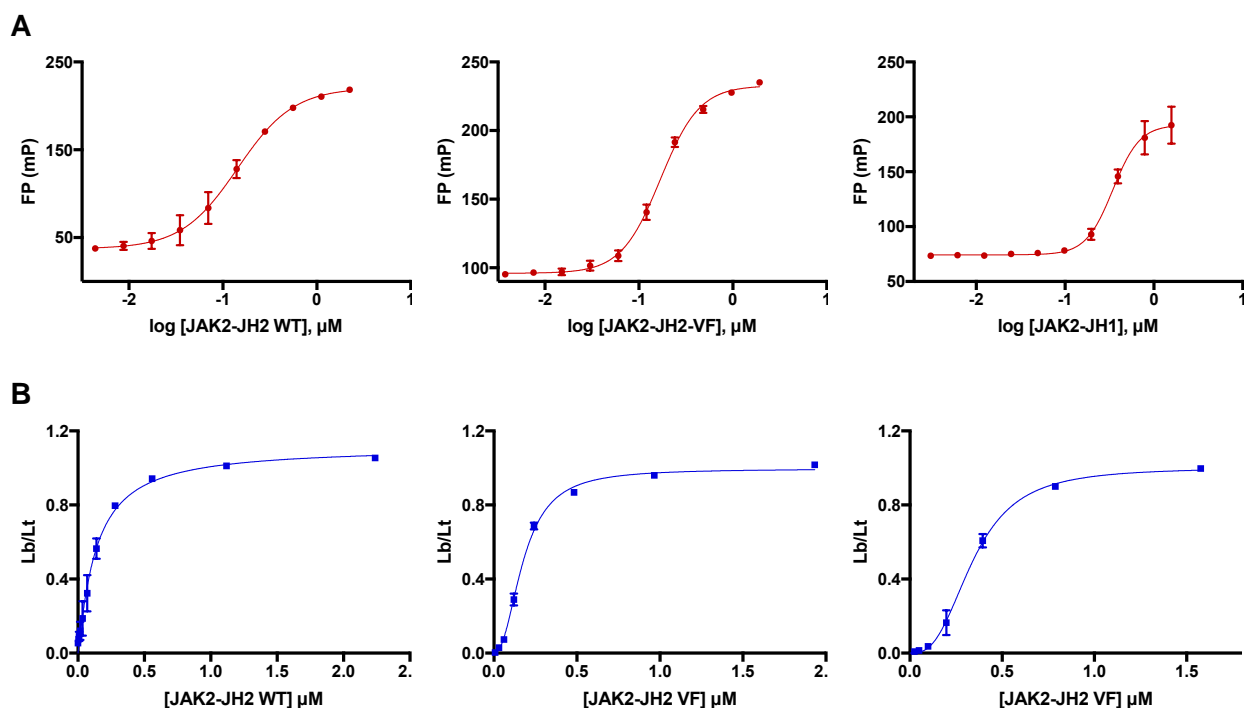


Figure ES2. Determination of binding affinities for tracer (6 nM) through saturation experiments. (A) Variation of FP values as a function of JAK2-JH2-WT, JAK2-JH2-V617F, and JAK2-JH1 concentration. (B) K_d determination for JAK2-JH2-WT, JAK2-JH2-VF, and JAK2-JH1. Lb/Lt = ratio of ligand bound to the total. Data from quadruplicate experiments in three independent assays. Mean \pm SEM plotted for all data.

1.4.2 Competitive FP Assay Protocol [8]

In a flat black bottom 96 well plate (Corning), 200 μ L of FP buffer (20 mM Tris-HCl pH 8.0, 150 mM NaCl, 20% Glycerol, 0.5 mM TCEP, 0.01% Tween 20) were added to column 1 (blank), 150 μ L to column 2, and 140 μ L to columns 3-12. 10 μ L of 2.96 μ M of JAK2-JH2 WT (3.52 μ M for JAK2-JH2-VF, and 6.93 μ M for JAK2-JH1), were added to columns 3-12, followed by the addition of 2 μ L of DMSO to columns 1-3. 2 μ L of inhibitor in DMSO at different concentrations were added from column 4 to 12. 50 μ L of 24 nM of tracer were added to columns 2-12. Fluorescence polarization was measured at $\lambda_{exc} = 485 \pm 20$ nm, $\lambda_{em} = 535 \pm 25$ nm for 1 hour. Experiments were carried out in quadruplicate with three independent experiments. Data were analyzed by a least-squares non-linear fit, generated using Prism 7 in order to determine the compound's IC_{50} . K_d values for each inhibitor were calculated using the following equation based on the IC_{50} , K_d of the tracer (K_d^t), total (L_t) and bound (L_b) tracer, as well as total protein concentration (P_t) [9].

$$K_d^I = \frac{L_b IC_{50} K_d^t}{P_t L_t + L_b (P_t - L_t + L_b - K_d^t)}$$

2. General Procedure for Preparation of Compound 33 Analogues

2.1 General Information

Reagents and solvents were obtained from commercial supplies and used without further purification. Reactions were monitored by thin-layer chromatography (TLC) using Merck pre-coated silica gel plates (analytical, SiO₂-60, F₂₅₄). TLC plates were visualized under U.V. light (254 nm). Organic solutions were concentrated under reduced pressure on a Büchi rotary evaporator. Microwave reactions were performed using a Biotage® Initiator⁺ microwave synthesizer using the standard settings. Flash column chromatography was performed on a Combiflash® Rf+ (Teledyne Isco, Lincoln, NE) with RediSep RF GOLD® (silica gel, particle size 20-40 µm) prepared cartridges. Preparative reverse phase HPLC was utilized for the purification of **33i-j**, **33l**, and **33n-o** using an Agilent 1260 Infinity II system equipped with G7161A Preparative Binary Pump., G7115A Diode Array Detector WR., G7157A Preparative Autosampler, G7159B Agilent Preparative Open-Bed Fraction Collector and an Agilent 5 Prep-C18 21× 100 mm column with a gradient of 0.1 % formic acid in water and 0.1 % formic acid in acetonitrile as the mobile phase. Purity assessment for **33i-j**, **33l**, and **33n-o** was conducted with the same system, using an Agilent Prep-C18 4.6 × 100 mm, 5µM particle size, scalar column with a gradient of 0.1 % formic acid in water and 0.1 % formic acid in acetonitrile as the mobile phase.

Nuclear magnetic resonance (NMR) spectra were recorded either on an Agilent DD₂ 400 (¹H NMR, ¹³C NMR recorded at 400, and 101 MHz, respectively), an Agilent DD₂ 500 (¹H NMR, ¹³C NMR recorded at 500, and 126 MHz, respectively), or an Agilent DD₂ 600 (¹H NMR, ¹³C NMR recorded at 600, and 151 MHz, respectively). All spectra were recorded at room temperature, 62 °C or 80 °C as noted. Chemical shifts are reported in ppm relative to deuterated solvent as an internal standard (δ_H DMSO-*d*₆ 2.50 ppm, δ_C DMSO-*d*₆ 39.52 ppm; δ_H Methanol-*d*₆ 3.31 ppm, δ_C Methanol-*d*₆ 49.00 ppm δ_H Acetone-*d*₆ 2.05 ppm, δ_C Acetone-*d*₆ 29.84 ppm, 206.26 ppm; δ_H Chloroform-*d* 7.26 ppm, δ_C Chloroform-*d* 77.16 ppm) with the following convention for describing multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, quint = quintet, m = multiplet, br = broad signal, dd = doublet of doublets, etc). High resolution mass spectroscopy (HRMS) measurements of assayed compounds were recorded using a Waters Acquity UPLC® coupled to a Waters Xevo® QTOF mass spectrometer equipped with a Waters ZSpray™ electrospray ionization source.

Mass spectrometric measurements for compounds **35-40**, **33i-j**, **33l**, and **33n-o** were performed with a Shimadzu Scientific Instruments QToF 9030 LC-MS system, equipped with a Nexera LC-40D xs UHPLC, consisting of a CBM-40 Lite system controller, a DGU-405 Degasser Unit, two LC-40D XS UHPLC pumps, a SIL-40C XS autosampler and a Column Oven CTO-40S. UV data was collected with a Shimadzu Nexera HPLC/UHPLC Photodiode Array Detector SPD M-40 in the range of 190 - 800nm. Mass spectra were subsequently recorded with the quadrupole time-of-flight (QToF) 9030 mass spectrometer. The samples were held at 4 deg C in the autosampler compartment. 0.3uL of each spiked solution were injected into a sample loop and separated on a

Shim-pack Scepter C18-120, 1.9um, 2.1x100mm Column, equilibrated at 40 deg C in a column oven. A binary gradient was used:

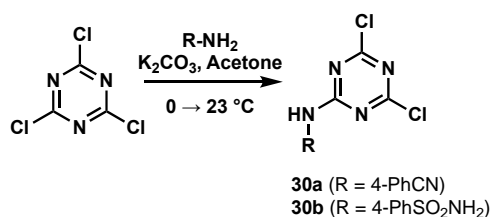
Solvent A: Water, HPLC grade Chromasolv, with 0.1% Formic Acid

Solvent B: Acetonitrile, HPLC grade Chromasolv, with 0.1% Formic Acid

The ionization source was run in "ESI" mode, with the electrospray needle held at +4.5kV. Nebulizer Gas was at 2 L/min, Heating Gas Flow at 10 L/min and the Interface at 300 deg C. Dry Gas was at 10 L/min, the Desolvation Line at 250 deg C and the heating block at 400 deg C. Mass spectra were recorded in the range of 50 to 2000 m/z in positive ion mode. Measurements and data post-processing were performed with LabSolutions 5.97 Realtime Analysis and PostRun.

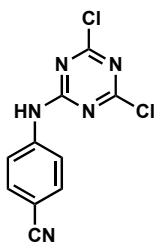
2.2 Synthesis of *Triazine* derivatives

Scheme ES1. Synthesis of Intermediates **30a-b**.



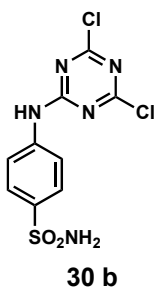
General Procedure A: Synthesis of Intermediates **30a-b.**

To a solution of cyanuric chloride (2.710 mmol, 1.0 eq.) in acetone (10 ml) at 0 °C was added potassium carbonate (5.420 mmol, 2.0 eq.) and the solution was stirred 20 minutes. The amino derivative (2.710 mmol, 1.0 eq.) was added portion wise and the reaction was allowed to warm at room temperature and stir overnight. The solvent was removed under reduced pressure. The product was collected by filtration, washed with H₂O and dried under vacuum.



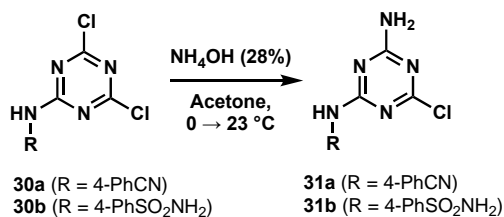
30 a

4-((4,6-dichloro-1,3,5-triazin-2-yl)amino)benzotrile (30a**)** (0.578 g, 81% yield) ¹H NMR (400 MHz, Methanol-*d*₄) δ 7.87 (d, *J* = 8.8 Hz, 2H), 7.75 (d, *J* = 8.9 Hz, 2H). HRMS (ESI) calcd for [M+H]⁺ C₁₀H₆Cl₂N₅ 266.0001, found 266.0006.



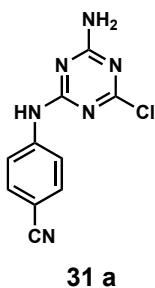
4-((4,6-dichloro-1,3,5-triazin-2-yl)amino)benzenesulfonamide (30b) (2.720 g, 31% yield)
¹H NMR (500 MHz, DMSO-*d*₆) δ 11.42 (s, 1H), 7.86 – 7.82 (m, 2H), 7.79 – 7.75 (m, 2H), 7.33 (s, 2H). HRMS (ESI) calcd for [M+H]⁺ C₉H₈Cl₂N₅O₂S 319.9776, found 319.9771.

Scheme ES2. Synthesis of Intermediates **31a-b**.

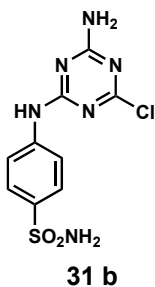


General Procedure B: Synthesis of Intermediates **31a-b**.

To a solution of **30** (14.5 mmol) in acetone (100 ml) at 0°C was added NH₄OH 28% (1 ml). The reaction was allowed to warm at room temperature and stir overnight. The precipitated product was collected by filtration and washed with H₂O.

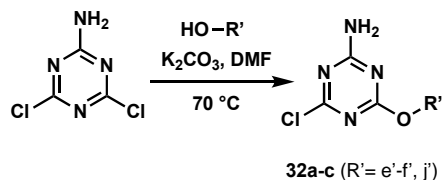


4-((4-amino-6-chloro-1,3,5-triazin-2-yl)amino)benzonitrile (31a) (3.034 g, 85% yield)
¹H NMR (400 MHz, Acetone-*d*₆) δ 9.25 (s, 1H), 8.06 (d, *J* = 8.6 Hz, 2H), 7.71 (d, *J* = 8.7 Hz, 2H), 7.15 (s, 1H), 7.00 (s, 1H). HRMS (ESI) calcd for [M+H]⁺ C₁₀H₈ClN₆ 247.0501, found 247.0504.



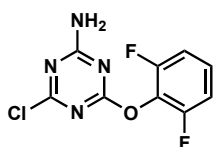
4-((4-amino-6-chloro-1,3,5-triazin-2-yl)amino)benzenesulfonamide (31b) (0.131 g, 70% yield)
¹H NMR (400 MHz, DMSO-*d*₆) δ 10.30 (s, 1H), 7.89 (d, *J* = 8.5 Hz, 2H), 7.80 – 7.67 (m, 4H), 7.25 (s, 2H). HRMS (ESI) calcd for [M+H]⁺ C₉H₁₀ClN₆O₂S 301.0274, found 301.0269.

Scheme ES3. Synthesis of Aromatic Ether Intermediates **32a-c**.



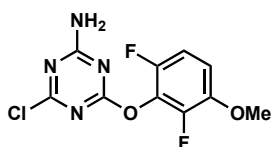
General Procedure C: Synthesis of Aromatic Ether Intermediates 32a-c.

A solution of 4,6-dichloro-1,3,5-triazin-2-amine (1.212 mmol, 1.0 eq.), functionalized phenol with (1.212 mmol, 1.0 eq.) and K_2CO_3 (3.636 mmol, 3.0 eq.) in DMF (2 mL) was stirred at 70°C for 18 hours. The reaction was cooled to room temperature, poured into H_2O and extracted with EtOAc. The organic layer was dried over Na_2SO_4 and the solvent was removed under reduced pressure. The crude mixture was loaded on silica to give the desired aromatic ether derivative after column chromatography.



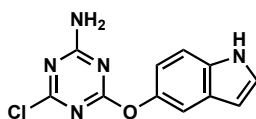
32 a

4-chloro-6-(2,6-difluorophenoxy)-1,3,5-triazin-2-amine (32a) (0.213 g, 68% yield) 1H NMR (400 MHz, $DMSO-d_6$) δ 8.37 (bs, 1H), 8.30 (bs, 1H), 7.46 – 7.22 (m, 3H). HRMS (ESI) calcd for $[M+H]^+$ $C_9H_6ClF_2N_4O$ 259.0198, found 259.0193.



32 b

4-chloro-6-(2,6-difluoro-3-methoxyphenoxy)-1,3,5-triazin-2-amine (32b) (0.164 g, 47% yield) 1H NMR (400 MHz, $DMSO-d_6$) δ 8.38 (bs, 1H), 8.32 (bs, 1H), 7.25 (td, $J = 9.7, 2.1$ Hz, 1H), 7.14 (td, $J = 9.3, 4.9$ Hz, 1H), 3.87 (s, 3H). HRMS (ESI) calcd for $[M+H]^+$ $C_{10}H_8ClF_2N_4O_2$ 289.0304, found 289.0299.

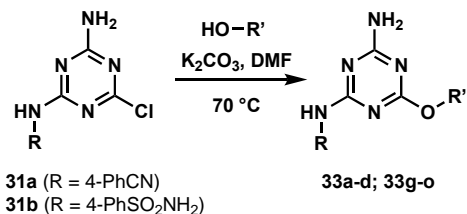


32 c

4-((1H-indol-5-yl)oxy)-6-chloro-1,3,5-triazin-2-amine (32c) (0.214 g, 68%) 1H NMR (400 MHz, $DMSO-d_6$) δ 11.19 (s, 1H), 8.01 (bs, 1H), 7.99 (bs, 1H), 7.46 – 7.37 (m, 2H), 7.33 (d, $J = 2.3$ Hz, 1H), 6.91 (dd, $J = 8.7, 2.3$ Hz, 1H), 6.43 (t, $J = 2.4$ Hz, 1H). HRMS (ESI) calcd for $[M+H]^+$ $C_{11}H_9ClN_5O$ 262.0496, found 262.0498.

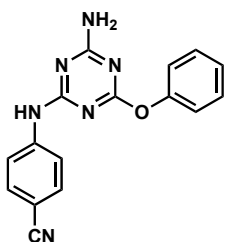
2.3 General Method for the Synthesis of the Aromatic Ether Analogs

Scheme ES4. Synthesis of Aromatic Ether Analogs **33a-d** and **33g-o**.



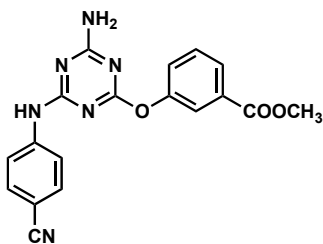
General Procedure C: Synthesis of Aromatic Ether Analogs **32a-d** and **33g-o**.

A solution of 4-((4-amino-6-chloro-1,3,5-triazin-2-yl)amino) derivative (0.20 mmol), functionalized phenol (0.30 mmol) and K₂CO₃ (0.60 mmol) in DMF (2 mL) was stirred at 70 °C for 18 hours. The reaction was cooled to room temperature, poured into H₂O and extracted with EtOAc. The organic layer was dried over Na₂SO₄ and the solvent removed under reduced pressure. The crude mixture was loaded on silica to give the desired aromatic ether derivative after column chromatography.



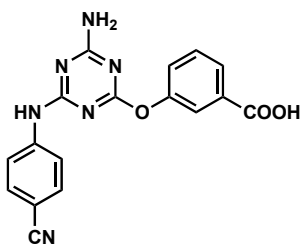
33 a

4-((4-amino-6-phenoxy-1,3,5-triazin-2-yl)amino)benzonitrile (33a) (62 % yield) ¹H NMR (400 MHz, Acetone-*d*₆) δ 9.01 (s, 1H), 7.94 (d, *J* = 7.2 Hz, 2H), 7.58 (d, *J* = 8.3 Hz, 2H), 7.47 (t, *J* = 7.5 Hz, 2H), 7.31 (t, *J* = 7.4 Hz, 1H), 7.20 (d, *J* = 7.9 Hz, 2H), 6.67 (d, *J* = 16.3 Hz, 2H). ¹³C NMR (101 MHz, Acetone-*d*₆) δ 170.02, 166.83, 153.66, 145.04, 133.49, 130.21, 126.09, 122.98, 120.43, 119.73, 105.43. HRMS (ESI) calcd. For [M+H]⁺ C₁₆H₁₃N₆O 305.1151, found 305.1155.



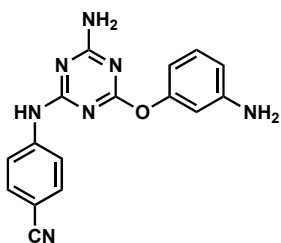
33 b'

Methyl 3-((4-amino-6-((4-cyanophenyl)amino)-1,3,5-triazin-2-yl)oxy)benzoate (33b') (59 % yield) ¹H NMR (600 MHz, Acetone-*d*₆) δ 9.04 (s, 1H), 7.95 (d, *J* = 7.6 Hz, 3H), 7.79 (s, 1H), 7.62 (t, *J* = 7.9 Hz, 1H), 7.58 (d, *J* = 7.6 Hz, 2H), 7.49 (d, *J* = 7.7 Hz, 1H), 6.74 (d, *J* = 50.9 Hz, 1H), 3.90 (s, 3H). ¹³C NMR (151 MHz, Acetone-*d*₆) δ 169.94, 166.58, 153.67, 133.51, 132.56, 130.58, 127.84, 127.03, 123.85, 120.53, 120.46, 119.70, 105.61, 52.60. HRMS (ESI) calcd. For [M+H]⁺ C₁₈H₁₅N₆O₃ 363.1206, found 363.1203.



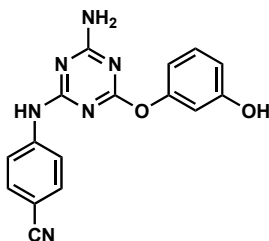
33 b

3-((4-amino-6-((4-cyanophenyl)amino)-1,3,5-triazin-2-yl)oxy)benzoic acid (33b) This compound was obtained by hydrolysis of **33b'** (45 % yield) ¹H NMR (600 MHz, DMSO-*d*₆) δ 13.19 (s, 1H), 10.02 (s, 1H), 7.84 (d, *J* = 7.7 Hz, 3H), 7.68 (s, 1H), 7.62 (d, *J* = 7.0 Hz, 2H), 7.57 (t, *J* = 7.8 Hz, 1H), 7.49 (d, *J* = 7.4 Hz, 1H), 7.38 (s, 2H). ¹³C NMR (151 MHz, DMSO-*d*₆) δ 170.67, 168.21, 166.66, 165.32, 152.17, 144.20, 132.70, 129.88, 126.19, 122.71, 119.45, 119.35, 103.33. HRMS (ESI) calcd for [M+H]⁺ C₁₇H₁₃N₆O₃ 349.1049, found 349.1043.



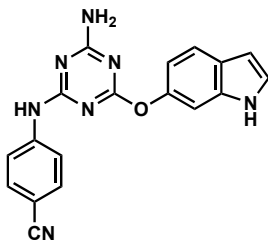
33 c

4-((4-amino-6-(3-aminophenoxy)-1,3,5-triazin-2-yl)amino)benzonitrile (33c) (26 % yield) ¹H NMR (600 MHz, Acetone-*d*₆) δ 9.00 (s, 1H), 7.97 (s, 2H), 7.59 (d, *J* = 8.1 Hz, 2H), 7.10 (t, *J* = 8.0 Hz, 1H), 6.60 (d, *J* = 7.8 Hz, 3H), 6.48 (s, 1H), 6.37 (d, *J* = 7.8 Hz, 1H), 4.84 (s, 2H). ¹³C NMR (151 MHz, Acetone-*d*₆) δ 170.09, 166.85, 154.63, 150.73, 145.15, 133.51, 130.37, 120.44, 119.80, 112.15, 110.75, 108.76, 105.33. HRMS (ESI) calcd for [M+H]⁺ C₁₆H₁₄N₇O 320.1260, found 320.1256.



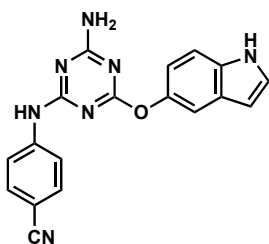
33 d

4-((4-amino-6-(3-hydroxyphenoxy)-1,3,5-triazin-2-yl)amino)benzonitrile (33d) (23 % yield). ¹H NMR (600 MHz, Acetone-*d*₆) δ 9.03 (s, 1H), 8.68 (s, 1H), 7.97 (s, 2H), 7.59 (d, *J* = 7.8 Hz, 2H), 7.26 (t, *J* = 7.9 Hz, 1H), 6.78 (d, *J* = 7.8 Hz, 1H), 6.66 (d, *J* = 8.9 Hz, 3H). ¹³C NMR (151 MHz, Acetone-*d*₆) δ 172.39, 170.04, 166.84, 159.23, 154.60, 145.08, 133.50, 130.62, 120.45, 119.76, 113.91, 110.34, 105.41. HRMS (ESI) calcd. For [M+H]⁺ C₁₆H₁₃N₆O₂ 321.1100, found 321.1095.



33 g

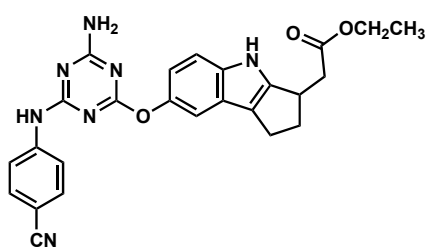
4-((4-((1H-indol-6-yl)oxy)-6-amino-1,3,5-triazin-2-yl)amino)benzonitrile (33g) (54 % yield) ¹H NMR (400 MHz, Acetone-*d*₆) δ 10.36 (s, 1H), 8.99 (s, 1H), 7.94 (s, 1H), 7.60 (d, *J* = 8.5 Hz, 1H), 7.49 (d, *J* = 7.4 Hz, 2H), 7.40 (s, 1H), 7.23 (s, 1H), 6.86 (d, *J* = 8.5 Hz, 1H), 6.63 (s, 2H), 6.55 (s, 1H). ¹³C NMR (151 MHz, Acetone-*d*₆) δ 170.09, 166.86, 148.96, 145.16, 137.17, 133.44, 126.73, 126.32, 121.15, 120.38, 119.76, 115.15, 105.51, 105.27, 102.40. HRMS (ESI) calcd for [M+H]⁺ C₁₈H₁₄N₇O 344.1260, found 344.1265.



33 h

4-((4-((1H-indol-5-yl)oxy)-6-amino-1,3,5-triazin-2-

yl)amino)benzonitrile (33h) (36 % yield) ¹H NMR (400 MHz, Acetone-*d*₆) δ 10.40 (s, 1H), 8.96 (s, 1H), 8.02 – 7.83 (m, 2H), 7.47 (d, *J* = 8.5 Hz, 3H), 7.43 (s, 1H), 7.34 (d, *J* = 2.2 Hz, 1H), 6.92 (dd, *J* = 8.7, 2.2 Hz, 1H), 6.60 (s, 2H), 6.51 (t, *J* = 2.5 Hz, 1H). ¹³C NMR (101 MHz, Acetone-*d*₆) δ 170.09, 166.86, 146.95, 145.18, 134.88, 133.41, 129.26, 127.00, 120.39, 119.77, 117.05, 113.40, 112.33, 105.21, 102.64. HRMS (ESI) calcd. for [M+H]⁺ C₁₈H₁₄N₇O 344.1260, found 344.1258.

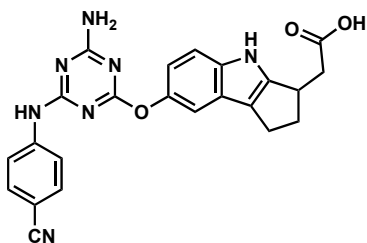


33 i'

Ethyl 2-(7-((4-amino-6-((4-cyanophenyl)amino)-1,3,5-triazin-2-yl)oxy)-1,2,3,4-tetrahydrocyclopenta[b]indol-3-yl)acetate (33i')

(52 % yield) To a flame dried flask equipped with a stir bar was added 51.9 mg of 4-((4-amino-6-chloro-1,3,5-triazin-2-yl)amino)benzonitrile (**31a**) (0.210 mmol), 55.9 mg of ethyl 2-(7-hydroxy-1,2,3,4-tetrahydrocyclopenta[b]indol-3-yl) acetate (0.216 mmol, 1.02 eq., commercially available) and 88.4 mg of potassium

carbonate (0.640 mmol, 3.04 eq.). The flask was flushed with nitrogen, at which point 3 mL of DMF were added and the solution was heated to 70 °C overnight for 15 hours. The mixture was then concentrated in vacuo and purified via column chromatography (0-100% EtOAc in Hexanes), yielding 50.4 mg of ethyl 2-(7-((4-amino-6-((4-cyanophenyl)amino)-1,3,5-triazin-2-yl)oxy)-1,2,3,4-tetrahydrocyclopenta[b]indol-3-yl)acetate (**33i'**) (0.107 mmol, 51.6% yield) as a colorless oil. ¹H NMR (500 MHz, Methanol-*d*₄) δ 7.66 (br s, 2H), 7.40 – 7.25 (m, 3H), 7.10 (d, *J* = 2.3 Hz, 1H), 6.81 (dd, *J* = 8.7, 2.3 Hz, 1H), 4.18 (q, *J* = 7.1 Hz, 2H), 3.60 (p, *J* = 6.9 Hz, 1H), 2.84 – 2.67 (m, 4H), 2.56 (dd, *J* = 15.9, 7.6 Hz, 1H), 2.22 – 2.12 (m, 1H), 1.25 (t, *J* = 7.1 Hz, 3H). Note: H of the COOH moiety was not observed. ¹³C NMR (126 MHz, Methanol-*d*₄) δ 174.34, 173.42, 169.98, 166.88, 148.49, 146.79, 145.32, 140.48, 133.65, 125.84, 120.80, 120.17, 119.67, 115.63, 112.93, 111.92, 105.48, 61.66, 40.53, 36.91, 36.77, 24.13, 14.56. MS (ESI) calcd for [M+H]⁺ C₂₅H₂₄N₇O₃ 470.2, found 470.1; [M+Na]⁺ 492.2, found 492.1; [M-H]⁺ 468.2, found 468.2.

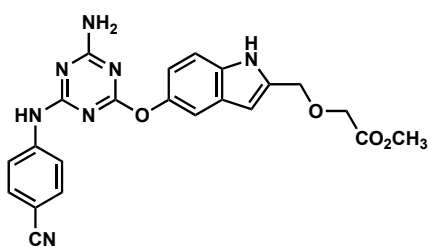


33 i

2-(7-((4-amino-6-((4-cyanophenyl)amino)-1,3,5-triazin-2-yl)oxy)-1,2,3,4-tetrahydrocyclopenta[b]indol-3-yl)acetic acid (33i)

(55 % yield) To a flask equipped with a stir bar was added 35.9 mg of **33i'** (0.0765 mmol), 3 mL of THF, and 3 mL of EtOH. 270 μL of 2M NaOH (aq.) (0.540 mmol, 7.06 eq.) was then added and the solution was allowed to stir under ambient atmosphere and

temperatures overnight for 15.5 hours. The solution was quenched with 2 M HCl (aq.) to pH 4 and concentrated *in vacuo*. The residue was then purified by reverse-phase HPLC Retention Time=13.396 min (5 % B- 100 % B over 20 minutes), yielding 18.6 mg of compound **33i** (0.0421 mmol, 55.1 % yield) as a white solid. ¹H NMR (500 MHz, DMSO-*d*₆) δ 10.71 (s, 1H), 9.89 (s, 1H), 7.87 (d, *J* = 8.4 Hz, 2H), 7.55 (d, *J* = 8.4 Hz, 2H), 7.30 (d, *J* = 8.7 Hz, 1H), 7.22 (br s, 1H), 7.19 (br s, 1H), 7.06 (d, *J* = 2.2 Hz, 1H), 6.77 (dd, *J* = 8.7, 2.3 Hz, 1H), 3.50 (p, *J* = 7.0, 6.5 Hz, 1H), 2.77 – 2.58 (m, 4H), 2.39 (dd, *J* = 16.0, 8.7 Hz, 1H), 2.14 – 2.03 (m, 1H). ¹³C NMR (126 MHz, DMSO-*d*₆) δ ¹³C NMR (126 MHz, dms) δ 173.49, 171.65, 168.31, 165.44, 147.64, 145.15, 144.43, 138.47, 132.65, 124.11, 119.39, 117.37, 114.43, 112.12, 110.58, 103.07, 48.60, 35.60, 35.09, 23.11. HRMS (ESI) calcd for [M+H]⁺ C₂₃H₂₀N₇O₃ 442.1628, found 442.1643.

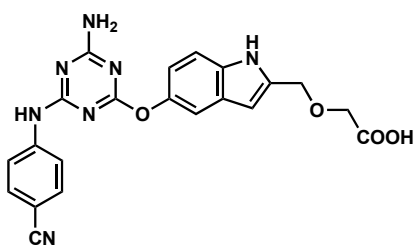


33j'

Methyl 2-((5-((4-amino-6-((4-cyanophenyl)amino)-1,3,5-triazin-2-yl)oxy)-1H-indol-2-yl)methoxy)acetate (33j'**)** (60 % yield)

Methyl 2-((5-hydroxy-1H-indol-2-yl)methoxy)acetate (**34**) (37.2 mg, 0.158 mmol, 1 eq.) was combined with 0.8 mL anhydrous dimethylformamide and potassium carbonate (69.7 mg, 0.5 mmol, 3.2 eq.) in a flame-dried pressure vial equipped with a rubber septum.

The mixture was stirred at rt for 5-10min, then was heated at 60 °C and stirred for another 5-10 min. 4-((4-amino-6-chloro-1,3,5-triazin-2-yl)amino)benzotrile (**31a**) (41.4 mg, 0.168 mmol 1.1eq.) was added, and the reaction stirred at 60° C for 11h. Afterwards, solvent was evaporated *in vacuo* and the residue was purified with normal-phase column chromatography (Dichloromethane/Methanol). Methyl 2-((5-((4-amino-6-((4-cyanophenyl)amino)-1,3,5-triazin-2-yl)oxy)-1H-indol-2-yl)methoxy)acetate (**33j'**) was isolated as a white solid. Yield: 42.5 mg, 60%. The corresponding carboxylic acid **33j** was also identified as one of the reaction products, and was isolated as a pale white solid. Yield: 15 mg, 22%. ¹H NMR (600 MHz, DMSO-*d*₆) δ 11.27 (s, 1H), 9.92 (s, 1H), 7.87 (brs, 2H), 7.55 (brs, 2H), 7.36 (d, *J* = 8.7 Hz, 1H), 7.28 (d, *J* = 2.2 Hz, 1H), 7.23 (brs, 2H), 6.90 (dd, *J* = 8.7, 2.3 Hz, 1H), 6.41 (d, 1H), 4.69 (s, 2H), 4.17 (s, 2H), 3.66 (s, 3H). ¹³C NMR (151 MHz, DMSO-*d*₆) δ 171.63, 170.44, 168.33, 165.44, 145.39, 144.42, 136.21, 134.27, 133.01, 132.67, 127.79, 119.39, 116.26, 112.35, 111.68, 103.09, 101.75, 66.30, 65.39, 51.50. [M+H]: 446.1571, found 446.1586, and 468.1405 [M+Na⁺].

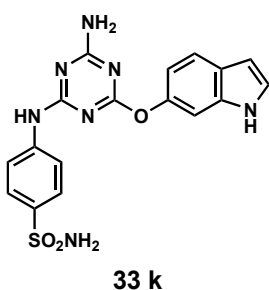


33j

2-((5-((4-amino-6-((4-cyanophenyl)amino)-1,3,5-triazin-2-yl)oxy)-1H-indol-2-yl)methoxy)acetic acid (33j**)** (27 % yield)

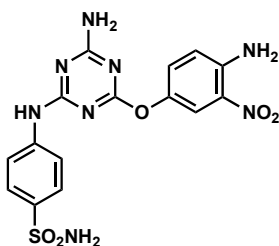
Methyl 2-((5-((4-amino-6-((4-cyanophenyl)amino)-1,3,5-triazin-2-yl)oxy)-1H-indol-2-yl)methoxy)acetate (**33j'**) (20 mg, 0.045 mmol, 1.0 eq.) was suspended in 4.5 mL dioxane. Sodium hydroxide (0.58 mL of a 2M aqueous solution, 1.16 mmol, 25.8

eq.) was added dropwise, and the reaction was allowed to stir at rt for 1h, when TLC indicated completion. Subsequently the reaction was concentrated to dryness, and 1.5mL water was added. The pH was adjusted to ~4, the precipitate was collected with centrifugation and removal of the supernatant water, followed by drying under a nitrogen stream. Then the residue was purified using a preparatory HPLC column with a gradient of acetonitrile with 0.1% formic acid /water with 0.1% formic acid, to afford **33j** as a pale-white solid. Yield 5.3 mg, 27%. Purity 99%. ¹H NMR (600 MHz, DMSO-*d*₆) δ 11.77 (s, 1H), 9.92 (s, 1H), 7.90 (d, *J* = 5.2 Hz, 2H), 7.57 (d, *J* = 6.9 Hz, 2H), 7.37 (d, *J* = 8.6 Hz, 1H), 7.30 – 7.15 (m, 3H), 6.86 (dd, *J* = 8.6, 2.3 Hz, 1H), 6.33 (s, 1H), 4.67 (s, 2H), 3.84 (s, 2H). Note: H of the COOH moiety was not observed. ¹³C NMR (151 MHz, DMSO-*d*₆) δ 172.58, 171.63, 168.32, 165.47, 145.21, 144.45, 137.90, 134.14, 132.70, 127.95, 119.43, 119.40, 115.76, 112.12, 111.62, 103.08, 100.49, 68.93, 65.33. [M+H]: 432.1415, found 432.1429, and 454.1247 [M+Na⁺].



4-((4-((1H-indol-6-yl)oxy)-6-amino-1,3,5-triazin-2-

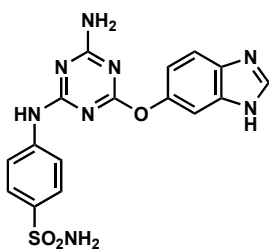
yl)amino)benzenesulfonamide (33k) (38 % yield) ¹H NMR (400 MHz, Acetone-*d*₆) δ 10.35 (s, 1H), 8.89 (s, 1H), 7.91 (s, 2H), 7.67 (d, *J* = 7.0 Hz, 2H), 7.59 (d, *J* = 8.4 Hz, 1H), 7.38 (s, 1H), 7.22 (s, 1H), 6.86 (d, *J* = 8.3 Hz, 1H), 6.58 (s, 2H), 6.54 (s, 1H), 6.41 (s, 2H). ¹³C NMR (101 MHz, Acetone-*d*₆) δ 170.05, 166.96, 148.96, 144.16, 138.00, 137.15, 127.61, 126.67, 126.28, 121.08, 119.85, 115.16, 105.44, 102.40. HRMS (ESI) calcd for [M+H]⁺ C₁₇H₁₇N₇O₃S 398.1035, found 398.1040.



4-((4-amino-6-(4-amino-3-nitrophenoxy)-1,3,5-triazin-2-

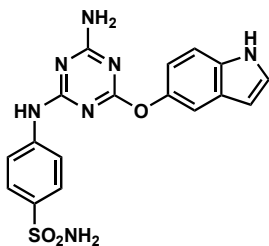
yl)amino)benzenesulfonamide (33l') (63 % yield) To a flame dried flask equipped with a stir bar 4-((4-amino-6-chloro-1,3,5-triazin-2-yl)amino)benzenesulfonamide (**31b**) was added (301.9 mg, 1.00 mmol), 4-amino-3-nitrophenol (154.4 mg, 1.00 mmol, 1.00 eq.) and potassium carbonate (416.7 mg, 3.02 mmol, 3.00 eq.). The flask was flushed with nitrogen, at which point 4 mL of DMF were added and the solution was heated to 70 °C overnight

for 16 hours. The mixture was then concentrated in vacuo, suspended in acetone, filtered, and washed with acetone. The supernatant was then collected, concentrated in vacuo, and suspended in dichloromethane. This mixture was then filtered, washed with dichloromethane, and the precipitate was isolated, yielding 263.1 mg (0.629 mmol) of (**33l'**) as a burnt yellow solid. ¹H NMR (500 MHz, Acetone-*d*₆) δ 9.00 (s, 1H), 7.91 (br s, 2H), 7.84 (s, 1H), 7.74 (d, *J* = 8.5 Hz, 2H), 7.35 (d, *J* = 8.5 Hz, 1H), 7.16 (d, *J* = 9.1 Hz, 1H), 7.14 (s, 2H), 6.72 (br s, 2H), 6.48 (br s, 2H). ¹³C NMR (126 MHz, Acetone-*d*₆) δ 172.42, 169.87, 166.78, 144.77, 143.89, 142.52, 138.20, 132.24, 131.10, 127.60, 120.51, 120.02, 118.34. MS (ESI) calcd for [M+H]⁺ C₁₅H₁₅N₈O₅S 419.1, found 419.1.



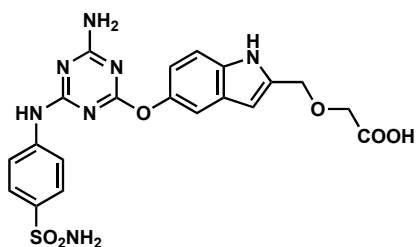
33 l

added. The vessel was then microwaved for 15 minutes at 130 °C. The contents of the flask were then concentrated in vacuo and purified on reverse phase HPLC, yielding 7.1 mg of 4-((4-((1H-benzo[d]imidazol-5-yl)oxy)-6-amino-1,3,5-triazin-2-yl)amino)benzenesulfonamide (0.0178 mmol, 28.8 % yield) as a white solid. ¹H NMR (600 MHz, DMSO-*d*₆) δ 9.81 (s, 1H), 8.31 (s, 1H), 7.80 (s, 2H), 7.62 (d, *J* = 8.6 Hz, 1H), 7.57 (s, 2H), 7.41 (d, *J* = 2.3 Hz, 1H), 7.24 (s, 1H), 7.17 (s, 3H), 7.06 (dd, *J* = 8.6, 2.3 Hz, 1H). ¹³C NMR (151 MHz, DMSO-*d*₆) δ 171.36, 168.26, 165.43, 163.01, 147.61, 142.91, 142.81, 136.80, 126.16, 118.91, 116.96. HRMS (ESI) calcd for [M+H]⁺ C₁₆H₁₅N₈O₃S 399.0988, found 399.0981.



33 m

4-((4-((1H-indol-5-yl)oxy)-6-amino-1,3,5-triazin-2-yl)amino)benzenesulfonamide (33m) (44 % yield) ¹H NMR (600 MHz, Acetone-*d*₆) δ 10.39 (s, 1H), 8.86 (s, 1H), 7.90 (s, 2H), 7.66 (s, 2H), 7.46 (d, *J* = 8.5 Hz, 1H), 7.42 (s, 1H), 7.34 (s, 1H), 6.93 (d, *J* = 8.0 Hz, 1H), 6.54 (s, 2H), 6.51 (s, 1H), 6.40 (s, 2H). ¹³C NMR (101 MHz, Acetone-*d*₆) δ 170.22, 167.12, 147.11, 144.37, 138.10, 135.01, 129.41, 127.78, 127.15, 120.01, 117.24, 113.50, 112.46, 102.80. HRMS (ESI) calcd for [M+H]⁺ C₁₇H₁₇N₇O₃S 398.1035, found 398.1029.

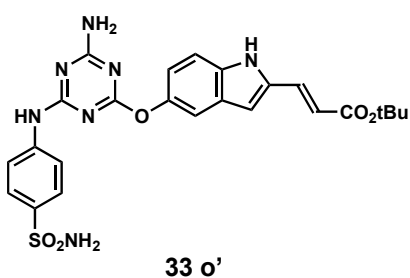


33 n

2-((5-((4-amino-6-((4-sulfamoylphenyl)amino)-1,3,5-triazin-2-yl)oxy)-1H-indol-2-yl)methoxy)acetic (34) (12.5 % yield) Methyl 2-((5-hydroxy-1H-indol-2-yl)methoxy)acetate (34 mg, 0.14 mmol, 1.0 eq.) was combined with 0.7 mL anhydrous dimethylformamide and potassium carbonate (58 mg, 0.42 mmol, 3.0 eq.) in a flame-dried pressure vial equipped with a rubber septum. The mixture was stirred

at rt for 5-10min, then was heated at 60°C and stirred for another 5-10min. 4-((4-amino-6-chloro-1,3,5-triazin-2-yl)amino)benzenesulfonamide (**31b**) (42.1 mg, 0.14 mmol 1.0 eq.) was added, and the reaction stirred at 60 °C for 37h. The mixture was concentrated and was subjected to normal-phase column chromatography (Dichloromethane/Methanol). The fractions containing the title product were collected and dried. The residue

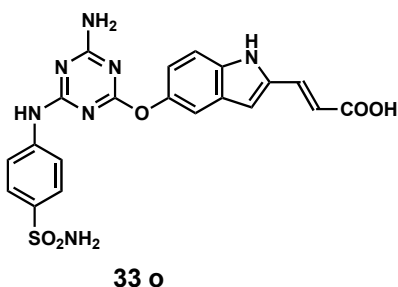
was further processed: 1 mL water was added and the pH was adjusted to ~4. The precipitate was collected with centrifugation and removal of the supernatant water, followed by drying under a nitrogen stream. Then the residue was purified using a preparatory HPLC column with a gradient of acetonitrile with 0.1% formic acid /water with 0.1% formic acid, to afford the title compound as a beige solid. ¹H NMR (600 MHz, DMSO-*d*₆) δ 11.28 (s, 1H), 9.78 (s, 1H), 7.83 (s, 2H), 7.58 (d, 2H), 7.36 (d, *J* = 8.6 Hz, 1H), 7.27 (d, *J* = 2.2 Hz, 1H), 7.24 – 7.07 (m, 4H), 6.89 (dd, *J* = 8.6, 2.1 Hz, 1H), 6.39 (s, 1H), 4.68 (s, 2H), 4.05 (s, 2H). Note: H of the COOH moiety was not observed. ¹³C NMR (151 MHz, DMSO-*d*₆) δ 171.61, 171.60, 168.33, 165.52, 145.38, 143.05, 136.76, 136.60, 134.20, 127.84, 126.23, 118.94, 116.15, 112.25, 111.61, 101.42, 66.55, 65.29. HRMS (ESI) calcd for [M+H]⁺ C₂₀H₂₀N₇O₆S 486.1196, found 486.1218, and 508.1037 [M+Na⁺], 95.5 % pure.



(E)-3-(5-((4-amino-6-((2-(3-(tert-butoxy)-3-oxoprop-1-en-1-yl)-1H-indol-5-yl)oxy)-1,3,5-triazin-2-yl)(4-sulfamoylphenyl)amino)benzenesulfonamide)acrylate (33o')

(62 % yield) To a flask dried in a vacuum desiccator equipped with a stir bar and containing 17.2 mg (0.0663 mmol) of tert-butyl (E)-3-(5-hydroxy-1H-indol-2-yl)acrylate (**35**) was added 21.1 mg (0.070 mmol, 1.06 eq.) of 4-((4-amino-6-chloro)-1,3,5-triazin-2-yl)amino)benzenesulfonamide (**31b**) and 28.9 mg (0.209 mmol, 3.15

eq.) of potassium carbonate. The flask was purged with nitrogen, at which point 3 mL of DMF was added. The solution was then heated to 70 °C and stirred overnight. The solution was then concentrated in vacuo, partitioned between EtOAc and water, and extracted with EtOAc. The organic layer was dried with Na₂SO₄, filtered, and concentrated in vacuo. The residue was purified via column chromatography (10:1 CH₂Cl₂:MeOH) yielding 21.4 mg (0.0410 mmol, 61.7 % yield) of (**33o'**) as a white solid. ¹H NMR (400 MHz, Methanol-*d*₄) δ 7.73 (d, *J* = 5.6 Hz, 2H), 7.64 (d, *J* = 5.6 Hz, 2H), 7.57 (d, *J* = 16.0 Hz, 1H), 7.40 (d, *J* = 8.8 Hz, 1H), 7.34 (d, *J* = 2.2 Hz, 1H), 7.02 (dd, *J* = 8.8, 2.3 Hz, 1H), 6.79 (s, 1H), 6.36 (d, *J* = 16.0 Hz, 1H), 1.55 (s, 9H). ¹³C NMR (101 MHz, Methanol-*d*₄) δ 173.21, 169.99, 168.26, 166.99, 147.46, 144.44, 137.86, 137.58, 136.56, 135.15, 129.97, 127.84, 120.31, 119.98, 118.72, 114.25, 112.77, 109.06, 81.71, 28.47. MS (ESI) calcd. For [M-H]⁻ C₂₄H₂₄N₇O₅S 522.2, found 522.2.

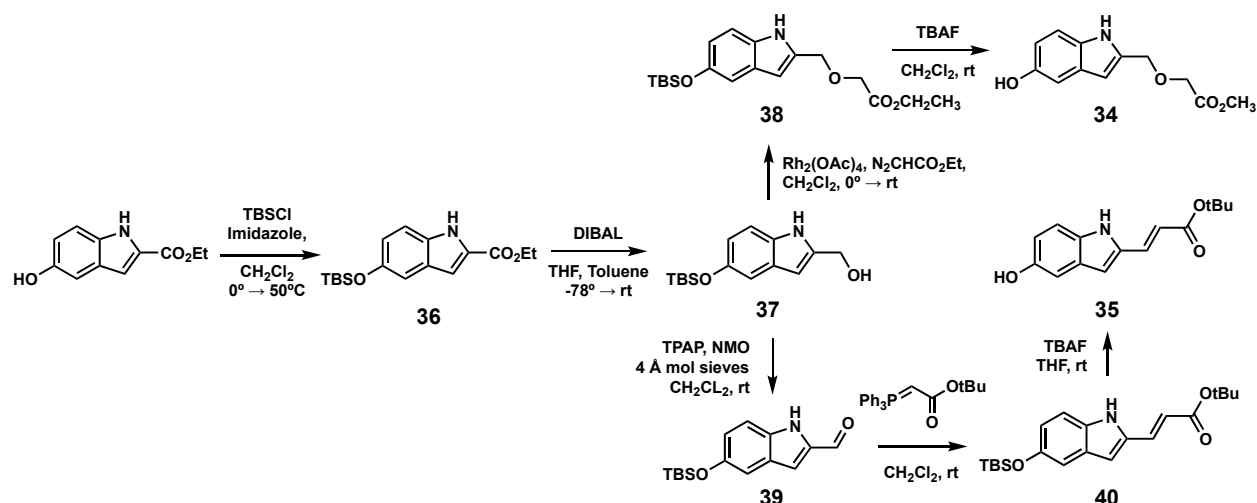


(E)-3-(5-((4-amino-6-((4-sulfamoylphenyl)amino)-1,3,5-triazin-2-yl)oxy)-1H-indol-2-yl)acrylic acid (33o)

(246 % yield) To a flask equipped with a stir bar and 18.0 mg (0.0344 mmol) of (E)-3-(5-((4-amino-6-((2-(3-(tert-butoxy)-3-oxoprop-1-en-1-yl)-1H-indol-5-yl)oxy)-1,3,5-triazin-2-yl)(4-sulfamoylphenyl)amino)benzenesulfonamide)acrylate (**33o'**) was added 2 mL of CH₂Cl₂ and 2 mL of TFA. The solution was stirred at room temperature

for 1 hour, at which point the mixture was concentrated in vacuo. The residue was then purified by reverse phase HPLC to yield the title compound (**33o**) as a white solid. ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 11.66 (s, 1H), 9.82 (s, 1H), 7.84 (d, $J = 5.4$ Hz, 2H), 7.59 (d, $J = 8.5$ Hz, 2H), 7.50 (d, $J = 15.9$ Hz, 1H), 7.38 (d, $J = 8.8$ Hz, 1H), 7.34 (d, $J = 2.3$ Hz, 1H), 7.24 (br s, 1H), 7.19 (br s, 3H), 7.00 (dd, $J = 8.7, 2.3$ Hz, 1H), 6.81 (s, 1H), 6.48 (d, $J = 16.1$ Hz, 1H). Note: H of the COOH moiety was not observed. ^{13}C NMR (101 MHz, $\text{DMSO-}d_6$) δ 171.49, 168.29, 165.50, 145.76, 143.02, 136.79, 135.56, 135.43, 128.17, 126.24, 118.94, 118.69, 112.89, 111.82, 107.15. HRMS (ESI) calcd. For $[\text{M}+\text{H}]^+$ $\text{C}_{20}\text{H}_{18}\text{N}_7\text{O}_5\text{S}$ 468.1085, found 468.1075. Retention Time=10.530 min (5 % B-100 % B over 20 minutes), 96.4 % pure.

Scheme ES5. Synthesis of Indole Intermediates **34** and **35**.



Ethyl 5-((*tert*-butyldimethylsilyl)oxy)-1*H*-indole-2-carboxylate (36**)** (0.733 g, 94 % yield) To a flame-dried flask, imidazole (299 mg, 4.39 mmol, 1.8 eq) was combined with anhydrous tetrahydrofuran (6 mL). Upon complete dissolution, ethyl 5-hydroxy-1*H*-indole-2-carboxylate (500 mg, 2.44 mmol, 1.0 eq) was added, the mixture stirred for 10 min, and was cooled to 0° C. Subsequently *tert*-butyldimethylsilyl chloride (551 mg, 3.66 mmol, 1.5 eq) was added portion-wise. The reaction was warmed to rt and then heated at 50 ° C for 12.5 h. Afterwards solvent was evaporated, dichloromethane (60mL) was added to the residue, and the organic layer was washed 3 times with 30mL water, 2 times with 30mL BRINE, and dried over sodium sulfate. After filtering and solvent evaporation, the title product was isolated as a pale-white solid. ^1H NMR (500 MHz, $\text{DMSO-}d_6$) δ 11.71 (s, 1H), 7.32 (d, $J = 8.8$ Hz, 1H), 7.03 (dd, $J = 11.2, 1.8$ Hz, 2H), 6.83 (dd, $J = 8.8, 2.3$ Hz, 1H), 4.32 (q, $J = 7.1$ Hz, 2H), 1.33 (t, $J = 7.1$ Hz, 3H), 0.96 (s, 9H), 0.17 (s, 6H). ^{13}C NMR (126 MHz, $\text{DMSO-}d_6$) δ 161.21, 148.88, 133.21, 127.83, 127.29, 119.50, 113.22, 110.08, 107.12, 60.29, 25.62, 17.91, 14.29, -4.53. HRMS (ESI) calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{17}\text{H}_{26}\text{NO}_3\text{Si}$ 320.1682, found 320.1685.

(5-((*tert*-butyldimethylsilyloxy)-1*H*-indol-2-yl)methanol (37) (1.60 g, 74 % yield) In a flame-dried flask, ethyl 5-((*tert*-butyldimethylsilyloxy)-1*H*-indole-2-carboxylate (**36**) (2.5 g, 7.83 mmol, 1 eq.) was dissolved in anhydrous tetrahydrofuran (50 mL). The mixture was brought to -78° C, followed by dropwise, slow addition of diisobutylaluminum(III) hydride (26.1 mL of 1.2M solution in toluene, 31.32 mmol, 4eq.) for over 1h. At the end of the addition, the reaction was warmed to rt, and was stopped at 1.5 h, when TLC indicated consumption of the starting material. Subsequently, the reaction was quenched with excess methanol until bubbling was ceased, and the solvents were removed under reduced pressure. The residue was dissolved in ethyl acetate, was washed 2 times with sodium bicarbonate, 2 times with BRINE, and was dried over sodium sulfate. After filtering and solvent evaporation, the residue was further purified with normal-phase column chromatography (Hexanes /Ethyl Acetate) to afford the title product as a yellow oil. ¹H NMR (600 MHz, DMSO-*d*₆) δ 10.82 (s, 1H), 7.17 (d, *J* = 8.6 Hz, 1H), 6.87 (d, *J* = 1.8 Hz, 1H), 6.58 (dd, *J* = 8.6, 2.2 Hz, 1H), 6.15 (s, 1H), 5.18 (t, *J* = 5.5 Hz, 1H), 4.56 (d, *J* = 5.5 Hz, 2H), 0.96 (s, 9H), 0.15 (s, 6H). ¹³C NMR (151 MHz, DMSO-*d*₆) δ 147.95, 140.96, 131.86, 128.51, 114.40, 111.41, 108.92, 98.25, 56.92, 17.97, 14.10, -4.44. HRMS (ESI) calcd for [M+H]⁺ C₁₅H₂₄NO₂Si 278.1576, found 278.1584.

Ethyl 2-((5-((*tert*-butyldimethylsilyloxy)-1*H*-indol-2-yl)methoxy)acetate (38) (0.249 g, 31 % yield) (5-((*tert*-butyldimethylsilyloxy)-1*H*-indol-2-yl)methanol (**37**) (0.605 g, 2.18 mmol, 1 eq.), was combined with anhydrous dichloromethane (3mL) in a flame-dried pressure vial equipped with a rubber septum. The mixture was cooled to 0° C and rhodium acetate dimer was added (0.034 g, 0.077 mmol, 0.035 eq.) The reaction was purged with nitrogen and was allowed to stir for 5-10 min at 0° C. Subsequently ethyl diazoacetate (395 μL, 87% wt solution in dichloromethane, 3.27 mmol, 1.5 eq.) was added slowly. After the addition was complete, the reaction was stirred for 10 min at 0° C, then warmed to rt and was allowed to run for 21h. The crude mixture was filtered through celite and washed with copious amounts of dichloromethane. The organic phase was washed 7 times with water, until discoloration, one time with BRINE, and was dried over sodium sulfate. Salts were filtered and solvent was evaporated under reduced pressure. The residue was further purified with normal-phase column chromatography (Hexanes /Ethyl Acetate) to afford the title product as a bright yellow waxy solid. ¹H NMR (600 MHz, DMSO-*d*₆) δ 11.01 (s, 1H), 7.19 (d, *J* = 8.6 Hz, 1H), 6.90 (d, *J* = 2.3 Hz, 1H), 6.62 (dd, *J* = 8.6, 2.3 Hz, 1H), 6.27 (d, *J* = 1.6 Hz, 1H), 4.63 (s, 2H), 4.13 – 4.10 (m, 4H, overlapping terminal CH₂ of ether, and CH₂ of ethyl ester), 1.21 – 1.18 (m, 3H), 0.96 (s, 9H), 0.15 (s, 6H). ¹³C NMR (151 MHz, DMSO-*d*₆) δ 169.94, 148.12, 135.56, 132.23, 128.13, 115.30, 111.68, 109.12, 101.22, 66.41, 65.42, 60.20, 25.70, 17.96, 14.06, -4.46. HRMS (ESI) calcd for [M+H]⁺ C₁₉H₃₀NO₄Si 364.1944, found 364.1943.

Methyl 2-((5-hydroxy-1*H*-indol-2-yl)methoxy)acetate (34) (0.075 g, 48 % yield) Ethyl 2-((5-((*tert*-butyldimethylsilyloxy)-1*H*-indol-2-yl)methoxy)acetate (**38**) (0.242 g, 0.67 mmol, 1.0 eq.) was dissolved in dichloromethane (8 mL) and tetrabutylammonium fluoride (0.8 mL, 1M solution in THF, 1.2 eq.) was added

dropwise. After the addition was complete, the reaction stirred at rt for 3 min and subsequently dried in vacuo. The residue was purified with normal-phase column chromatography (Dichloromethane/Acetonitrile/Methanol). Transesterification occurred during the process, converting the ethyl to methyl ester, thus affording the title product as a white solid. ¹H NMR (600 MHz, DMSO-*d*₆) δ 9.39 (d, *J* = 17.0 Hz, 1H), 8.00 (d, *J* = 8.7 Hz, 1H), 6.92 (s, 1H), 6.76 (dd, *J* = 8.7, 2.4 Hz, 1H), 6.44 (s, 1H), 4.89 (s, 2H), 4.47 (s, 2H), 3.38 (s, 3H). ¹³C NMR (151 MHz, DMSO-*d*₆) δ 165.75, 154.52, 135.45, 130.51, 127.26, 115.81, 113.02, 105.88, 103.03, 67.83, 67.43, 62.14.

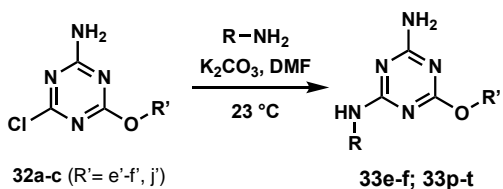
5-[tert-butyl(dimethyl)silyloxy-1*H*-indole-2-carbaldehyde (39) (0.67 g, 69 % yield) To a flask dried in a vacuum desiccator equipped with a stir bar and containing 0.098 mg, 0.354 mmol of [5-[tert-butyl(dimethyl)silyloxy-1*H*-indol-2-yl]methanol (**37**) was added 8.5 mg (0.024 mmol, 0.068 eq) of TPAP, 55.8 mg (0.484 mmol, 1.37 eq) of NMO, and 182.3 mg (500 mg/mmol) of 4 Å molecular sieves. The flask was purged under nitrogen, at which point 3 mL of CH₂Cl₂ was added and the solution was stirred for 23 hours at room temperature. The solution was concentrated in vacuo, and purified by column chromatography (0 to 66 % EtOAc in hexanes). ¹H NMR (500 MHz, Chloroform-*d*) δ 9.81 (s, 1H), 9.06 (s, 1H), 7.31 (d, *J* = 8.9 Hz, 1H), 7.16 (d, *J* = 1.1 Hz, 1H), 7.13 (d, *J* = 2.2 Hz, 1H), 6.98 (dd, *J* = 8.9, 2.3 Hz, 1H), 1.01 (s, 9H), 0.21 (s, 6H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 182.07, 150.37, 136.61, 133.99, 128.11, 122.88, 114.31, 113.09, 111.64, 25.88, 18.37, -4.28. MS (ESI) calcd. For [M+H]⁺ C₁₅H₂₂NO₂Si 276.1, found 276.2.

tert-butyl (E)-3-(5-((tert-butyl)dimethylsilyloxy)-1*H*-indol-2-yl)acrylate (40) (0.066 mg, 50 % yield) To a flask dried in a vacuum desiccator equipped with a stir bar and containing 97.7 mg (0.355 mmol) of 5-[tert-butyl(dimethyl)silyloxy-1*H*-indole-2-carbaldehyde (**39**) was added 133.5 mg (0.355 mmol, 1.00 eq) of tert-butyl 2-(triphenyl-λ⁵-phosphaneylidene)acetate. The flask was purged with nitrogen, and 3 mL CH₂Cl₂ was added. The solution was stirred overnight at room temperature, at which point the solution was concentrated in vacuo. The residue was purified using column chromatography (10 % EtOAc in hexanes). ¹H NMR (400 MHz, Chloroform-*d*) δ 8.75 (s, 1H), 7.56 (d, *J* = 16.0 Hz, 1H), 7.21 (d, *J* = 8.7 Hz, 1H), 7.03 (d, *J* = 2.3 Hz, 1H), 6.83 (dd, *J* = 8.7, 2.3 Hz, 1H), 6.67 (s, 1H), 6.24 (d, *J* = 16.0 Hz, 1H), 1.57 (s, 9H), 1.02 (s, 9H), 0.21 (s, 6H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 166.72, 149.81, 134.45, 133.81, 133.69, 129.24, 119.19, 117.39, 111.67, 110.47, 108.01, 80.86, 28.40, 25.91, 18.36, -4.29. MS (ESI) calcd. For [M+H]⁺ C₂₁H₃₂NO₃Si 374.2, found 374.2.

tert-butyl (E)-3-(5-hydroxy-1*H*-indol-2-yl)acrylate (35) (0.036 g, 85 % yield) To a flask dried in a vacuum desiccator equipped with a stir bar, purged with nitrogen, and 61.8 mg (0.165 mmol) of tert-butyl (E)-3-(5-((tert-butyl)dimethylsilyloxy)-1*H*-indol-2-yl)acrylate (**40**) was added 3 mL of THF and 0.35 mL (0.35 mmol, 0.212 eq) of 1M TBAF in THF. The solution was stirred for 2 hours at room temperature, at which point the solution was concentrated in vacuo. The residue was purified via column chromatography (0 to 100 % EtOAc in hexanes). ¹H NMR (400 MHz, Methanol-*d*₄) δ 7.51 (d, *J* = 15.9 Hz, 1H), 7.18 (d, *J* = 8.7 Hz, 1H), 6.90 (d, *J* = 2.3 Hz, 1H), 6.76 (dd, *J* = 8.7, 2.4 Hz, 1H), 6.59 (s, 1H), 6.27 (d, *J* = 15.9 Hz, 1H), 1.52 (s, 9H). ¹³C NMR (101 MHz,

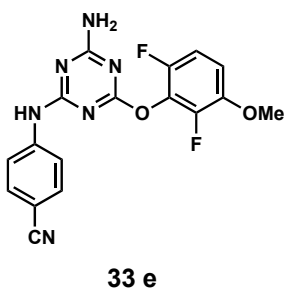
Methanol-*d*₄) δ 168.55, 152.04, 135.57, 134.93, 130.45, 117.39, 115.73, 112.79, 108.43, 105.49, 81.51, 28.47.
 MS (ESI) calcd. For $[M+Na]^+$ C₁₅H₁₇NNaO₃ 282.1, found 282.2.

Scheme ES6. Synthesis of Aniline Analogs **33e-f** and **33p-t**.

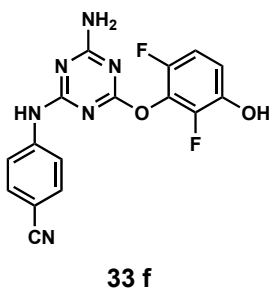


General Procedure E: Synthesis of Aniline Analogs 33e-f and 33p-t.

A solution of 4-((4-amino-6-chloro-1,3,5-triazin-2-yl)amino) derivative (0.20 mmol), functionalized aniline (0.30 mmol) and K₂CO₃ (0.60 mmol) in DMF (2 mL) was stirred at 70 °C for 18 hours. The reaction was cooled to room temperature, poured into H₂O and extracted with EtOAc. The organic layer was dried over Na₂SO₄ and the solvent removed under reduced pressure. The crude mixture was loaded on silica to give the desired aromatic ether derivative after column chromatography.



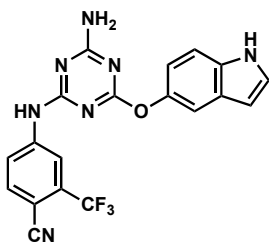
4-((4-amino-6-(2,6-difluoro-3-methoxyphenoxy)-1,3,5-triazin-2-yl)amino) benzonitrile (33e) (86%) ¹H NMR (400 MHz, Acetone-*d*₆) δ 9.13 (s, 1H), 8.03 – 7.78 (m, 2H), 7.58 (d, *J* = 8.4 Hz, 2H), 7.10 (d, *J* = 7.5 Hz, 2H), 6.85 (d, *J* = 30.5 Hz, 2H), 3.92 (s, 3H). ¹³C NMR (151 MHz, Acetone-*d*₆) δ 171.27, 170.05, 166.88, 151.10 (d, *J* = 2.4 Hz), 149.50 (d, *J* = 2.5 Hz), 147.23 (d, *J* = 4.2 Hz), 146.07 (dd, *J* = 9.1, 2.8 Hz), 145.58 (d, *J* = 4.3 Hz), 144.72, 120.65, 119.66, 113.48 – 109.28 (m), 105.86, 57.19. HRMS (ESI) calcd. For $[M+H]^+$ C₁₇H₁₃F₂N₆O₂ 371.1068, found 371.1066.



4-((4-amino-6-(2,6-difluoro-3-hydroxyphenoxy)-1,3,5-triazin-2-yl)amino)benzonitrile (33f) obtained by demethylation of **33e** (54%) To a solution of 4-((4-amino-6-(2,6-difluoro-3-methoxyphenoxy)-1,3,5-triazin-2-yl)amino)benzonitrile (60 mg, 0.16 mmol) in CH₂Cl₂ (0.5 mL) at 0°C was added phenylmethanethiol (0.37 mL) and AlCl₃ (32 mg, 0.24 mmol). The reaction was stirred for 30 minutes and a second portion of AlCl₃ was added. The reaction was

allowed to warm at room temperature and stirred for other 90 minutes. The reaction was quenched with a slow addition of H₂O and a saturated solution of NaHCO₃. The aqueous phase was extracted with EtOAc and the

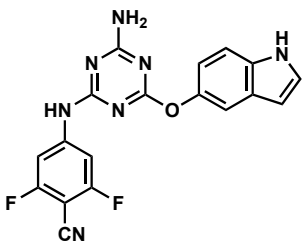
organic layer washed with brine and dried over Na₂SO₄. The solvent was removed under reduced pressure and the crude mixture loaded on silica to give the free alcohol after column chromatography. ¹H NMR (600 MHz, Acetone-*d*₆) δ 9.14 (s, 1H), 8.92 (s, 1H), 7.93 (s, 2H), 7.61 (d, *J* = 8.3 Hz, 2H), 7.06 – 6.94 (m, 2H), 6.85 (d, *J* = 42.9 Hz, 2H). ¹³C NMR (151 MHz, Acetone-*d*₆) δ 171.35, 170.02, 166.89, 150.18, 148.59, 146.54 (d, *J* = 4.2 Hz), 144.92 (d, *J* = 4.3 Hz), 144.73 (d, *J* = 14.6 Hz), 143.34 (d, *J* = 11.0 Hz), 133.53, 120.64, 119.68, 111.24 (dd, *J* = 19.9, 3.9 Hz), 105.81. HRMS (ESI) calcd for [M+H]⁺ C₁₆H₁₁F₂N₆O₂ 357.0912, found 357.0907.



33 p

4-((4-((1H-indol-5-yl)oxy)-6-amino-1,3,5-triazin-2-yl)amino)-2-

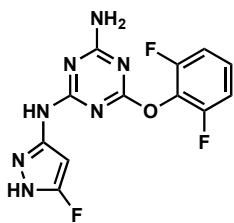
(trifluoromethyl)benzonitrile (33p) (0.032 g, 14% yield) ¹H NMR (600 MHz, DMSO-*d*₆) δ 11.16 (s, 1H), 10.26 (s, 1H), 8.24 (s, 2H), 7.90 – 7.77 (m, 1H), 7.45 – 7.36 (m, 3H), 7.32 (d, *J* = 2.3 Hz, 2H), 6.90 (dd, *J* = 8.7, 2.3 Hz, 1H), 6.42 (t, *J* = 2.5 Hz, 1H). ¹³C NMR (151 MHz, dms) δ 171.62, 168.26, 165.45, 145.20, 144.86, 135.87, 133.64, 131.46 (d, *J* = 31.6 Hz), 127.89, 126.62, 123.40, 121.88, 116.99 – 116.43 (m), 116.20, 115.71, 112.16, 111.78, 101.27, 99.40. HRMS (ESI) calcd for [M+H]⁺ C₁₉H₁₃F₃N₇O 412.1134, found 412.1127.



33 q

4-((4-((1H-indol-5-yl)oxy)-6-amino-1,3,5-triazin-2-yl)amino)-2,6-

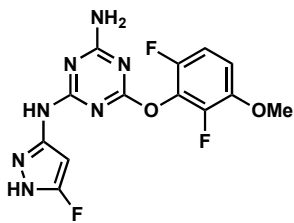
difluorobenzonitrile (33q) (0.021 g, 10% yield) ¹H NMR (600 MHz, DMSO-*d*₆) δ 11.16 (s, 1H), 10.32 (s, 1H), 7.64 (bs, 2H), 7.44 (bs, 2H), 7.41 (s, 1H), 7.40 (s, 1H), 7.31 (d, *J* = 2.3 Hz, 1H), 6.90 (dd, *J* = 8.8, 2.3 Hz, 1H), 6.42 (t, *J* = 2.5 Hz, 1H). ¹³C NMR (151 MHz, dms) δ 171.64, 168.29, 165.27, 163.41 (d, *J* = 7.2 Hz), 161.73 (d, *J* = 7.4 Hz), 145.16, 133.68, 129.68, 127.90, 126.66, 115.66, 112.10, 111.84, 110.33, 102.10, 101.94, 101.28. HRMS (ESI) calcd for [M+H]⁺ C₁₈H₁₂F₂N₇O 380.1071, found 380.1065.



33 r

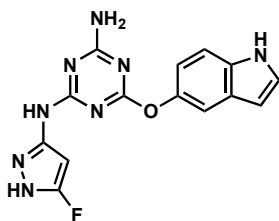
6-(2,6-difluorophenoxy)-N²-(5-fluoro-1H-pyrazol-3-yl)-1,3,5-triazine-

2,4-diamine (33r) (0.057 g, 23%) ¹H NMR (500 MHz, DMSO-*d*₆) δ 12.08 (s, 1H), 10.56 (s, 1H), 7.60 (d, *J* = 41.1 Hz, 2H), 7.40 – 7.22 (m, 3H), 5.49 (s, 1H). ¹³C NMR (126 MHz, DMSO-*d*₆) δ 169.73, 168.12, 164.10, 163.23, 161.36, 155.88 (d, *J* = 4.0 Hz), 153.91 (d, *J* = 4.2 Hz), 140.03, 128.15, 126.74, 112.63 (dd, *J* = 17.6, 4.2 Hz). HRMS (ESI) calcd for [M+H]⁺ C₁₂H₉F₃N₇O 324.0821, found 324.0816.



33 s

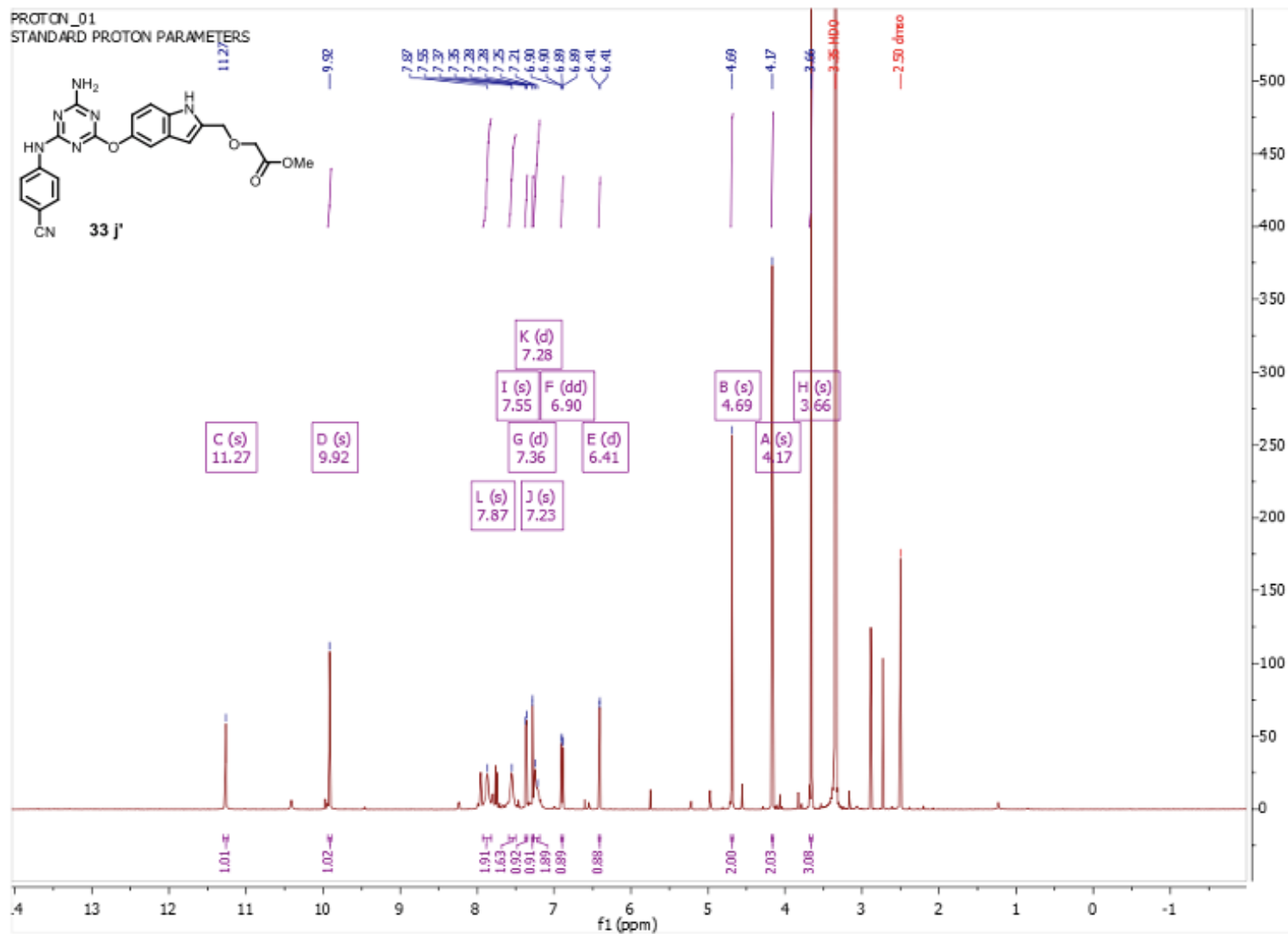
6-(2,6-difluoro-3-methoxyphenoxy)-*N*²-(5-fluoro-1*H*-pyrazol-3-yl)-1,3,5-triazine-2,4-diamine (33s) (0.060 g, 31%) ¹H NMR (600 MHz, DMSO-*d*₆) δ 12.09 (bs, 1H), 10.58 (bs, 1H), 7.66 (bs, 1H), 7.56 (bs, 1H), 7.21 (td, *J* = 9.6, 1.9 Hz, 1H), 7.09 (td, *J* = 9.1, 4.6 Hz, 1H), 5.49 (bs, 1H), 3.86 (s, 3H). ¹³C NMR (151 MHz, DMSO-*d*₆) δ 169.71, 168.15, 163.10, 161.55, 149.43, 147.84, 146.21 – 143.27 (m), 140.62, 128.94 – 128.75 (m), 110.64 – 110.49 (m), 109.87, 103.50 (d, *J* = 17.8 Hz), 56.61. HRMS (ESI) calcd for [M+H]⁺ C₁₃H₁₁F₃N₇O₂ 354.0926, found 354.0932.



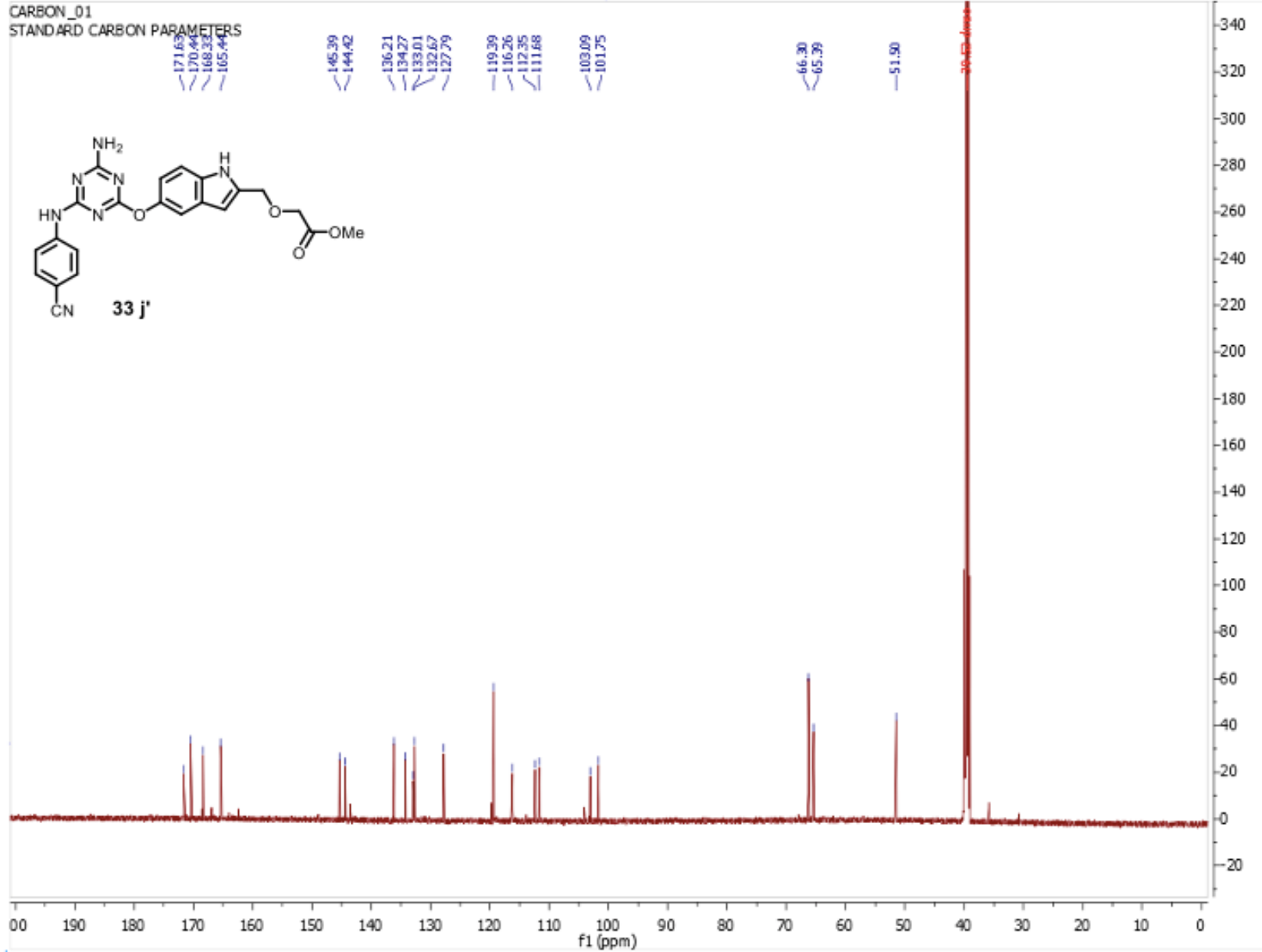
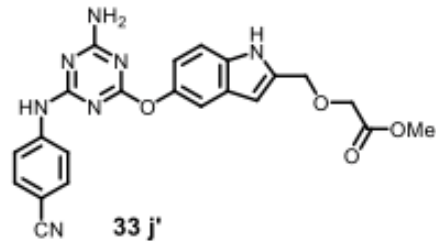
33 t

6-((1*H*-indol-5-yl)oxy)-*N*²-(5-fluoro-1*H*-pyrazol-3-yl)-1,3,5-triazine-2,4-diamine (33t) (0.017 g, 10%) ¹H NMR (500 MHz, DMSO-*d*₆) δ 12.07 (s, 1H), 10.38 (s, 1H), 7.96 (d, *J* = 7.1 Hz, 1H), 7.43 – 7.25 (m, 5H), 6.95 – 6.75 (m, 2H), 6.45 – 6.39 (m, 1H). ¹³C NMR (126 MHz, DMSO-*d*₆) δ 171.66, 170.08, 168.18, 164.29, 163.27, 161.39, 145.18, 133.56, 127.87, 126.59, 112.15, 111.74, 101.23. HRMS (ESI) calcd for [M+H]⁺ C₁₄H₁₂FN₈O 327.1118, found 327.1123.

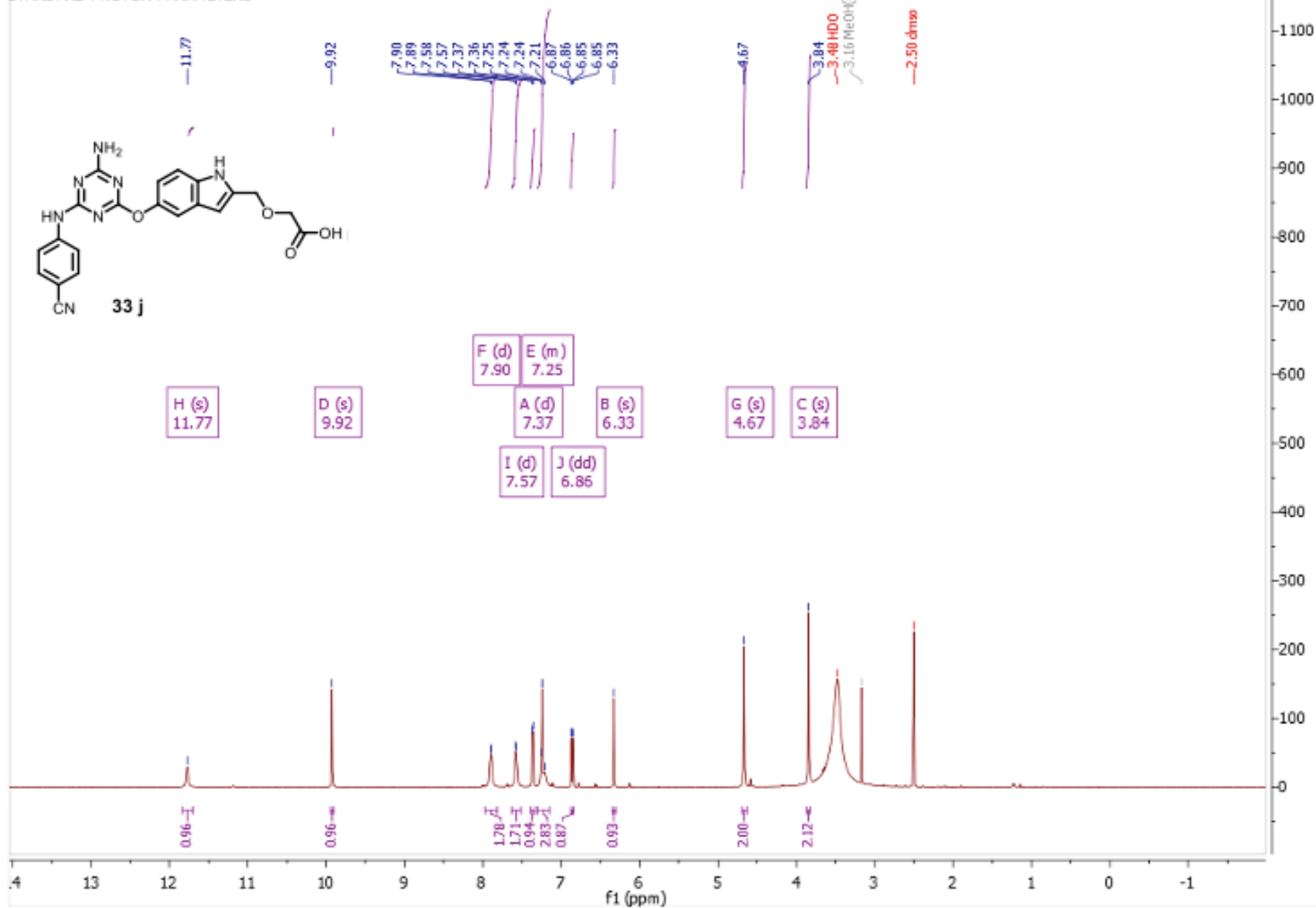
3. ¹H and ¹³C NMR Spectra



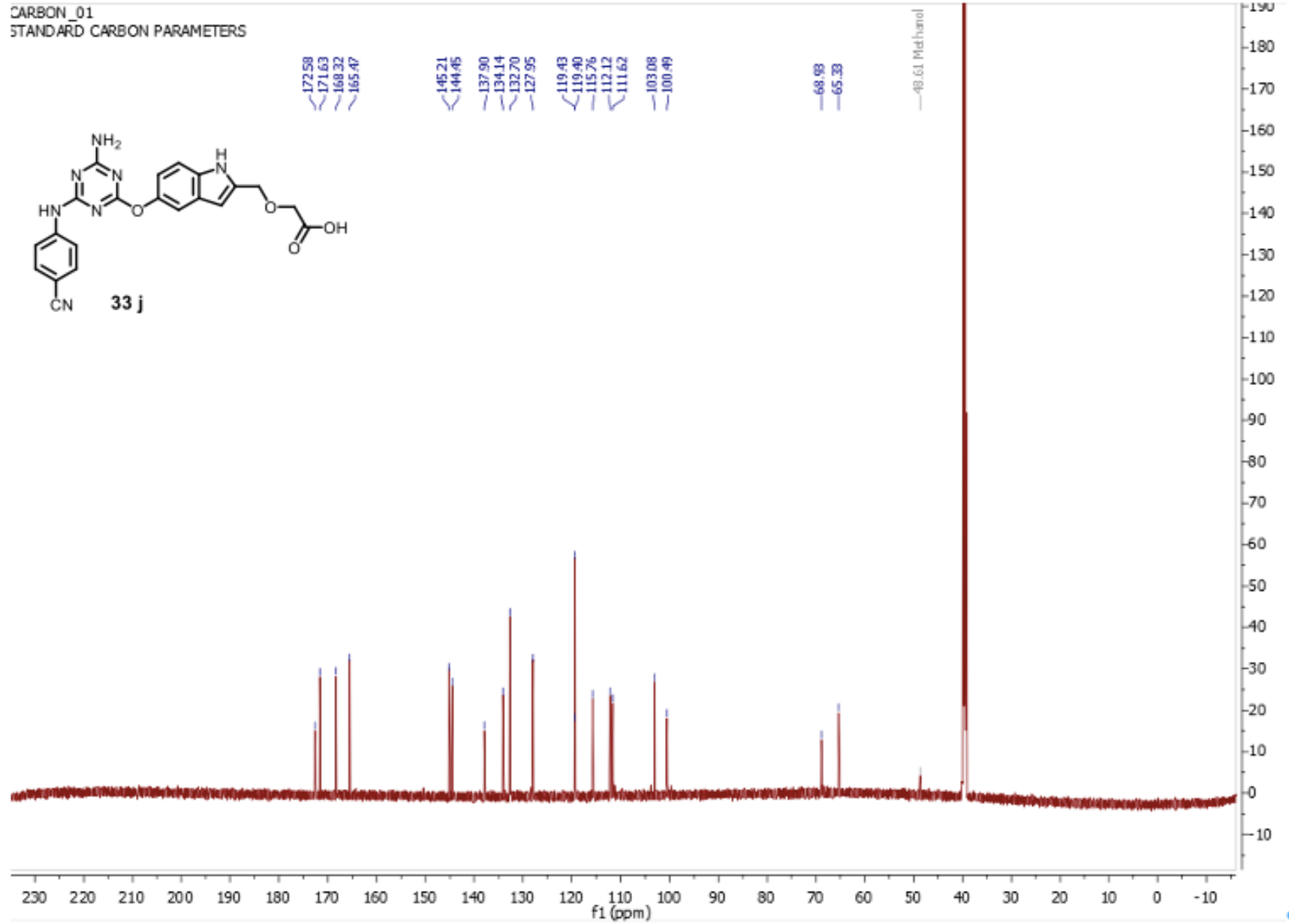
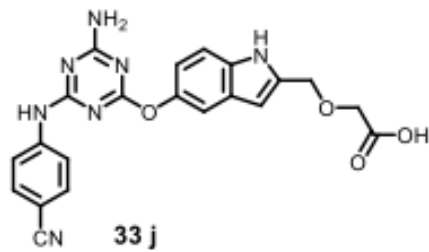
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STANDARD CARBON PARAMETERS

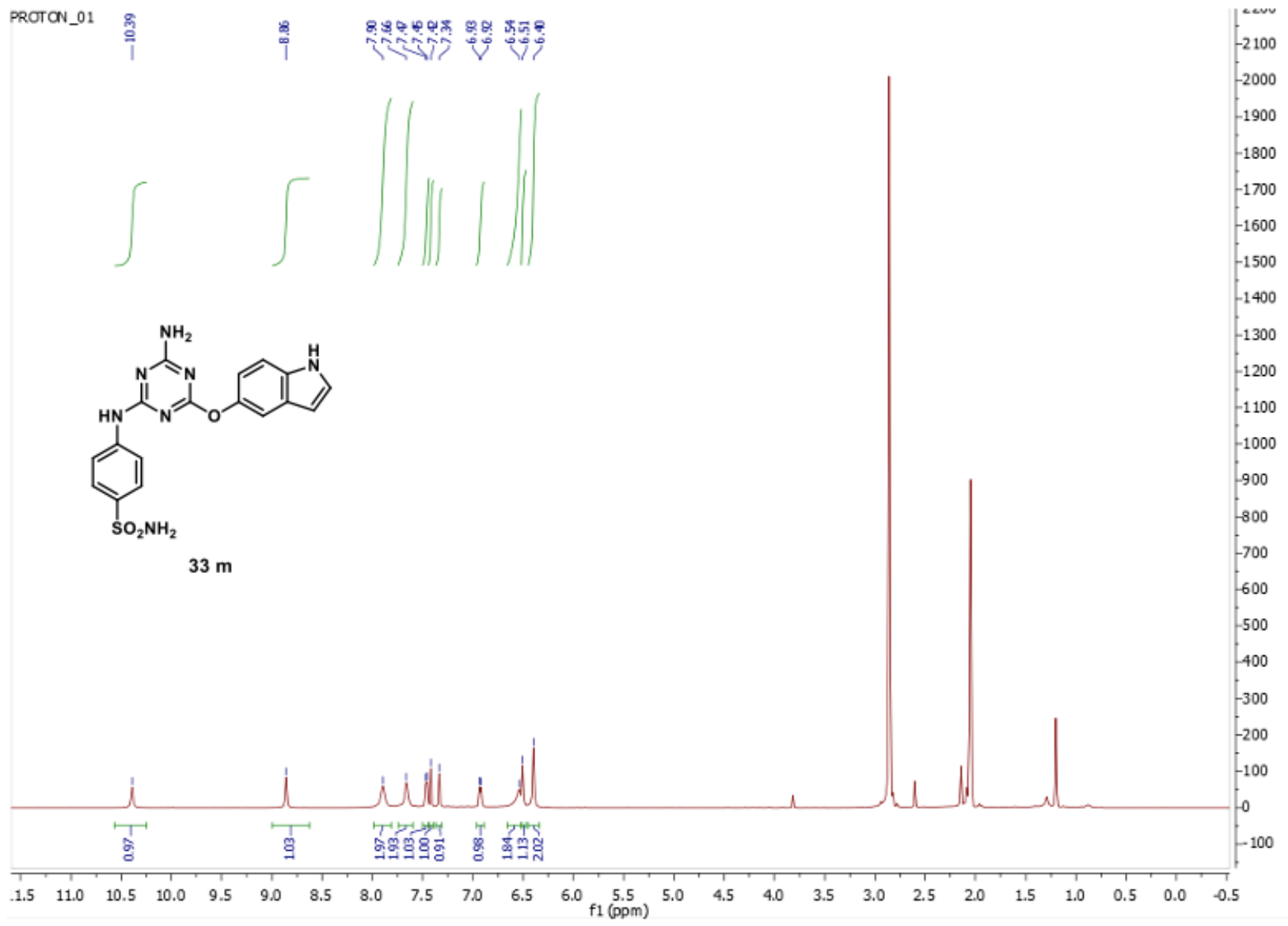


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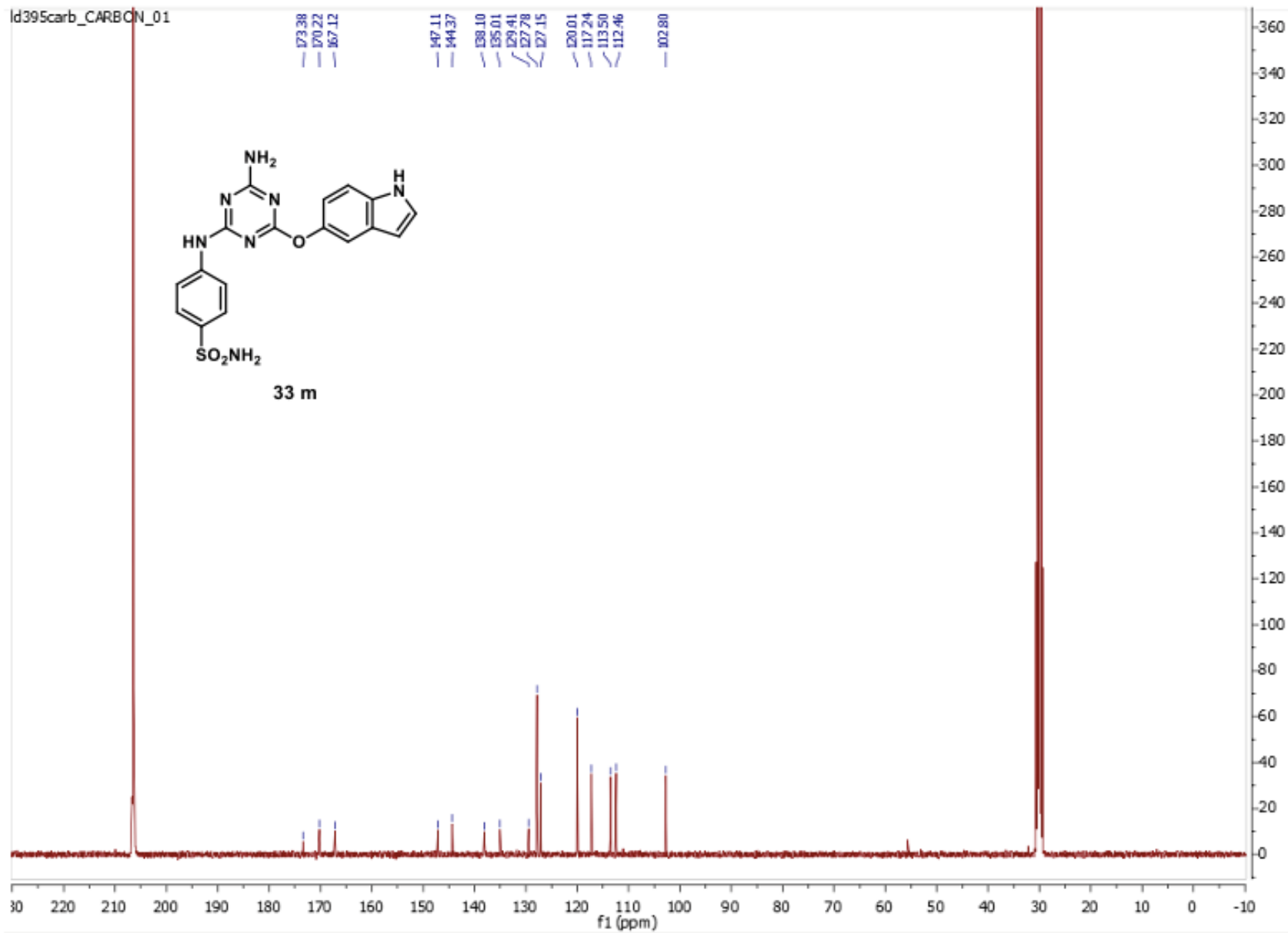


CARBON_01
STANDARD CARBON PARAMETERS

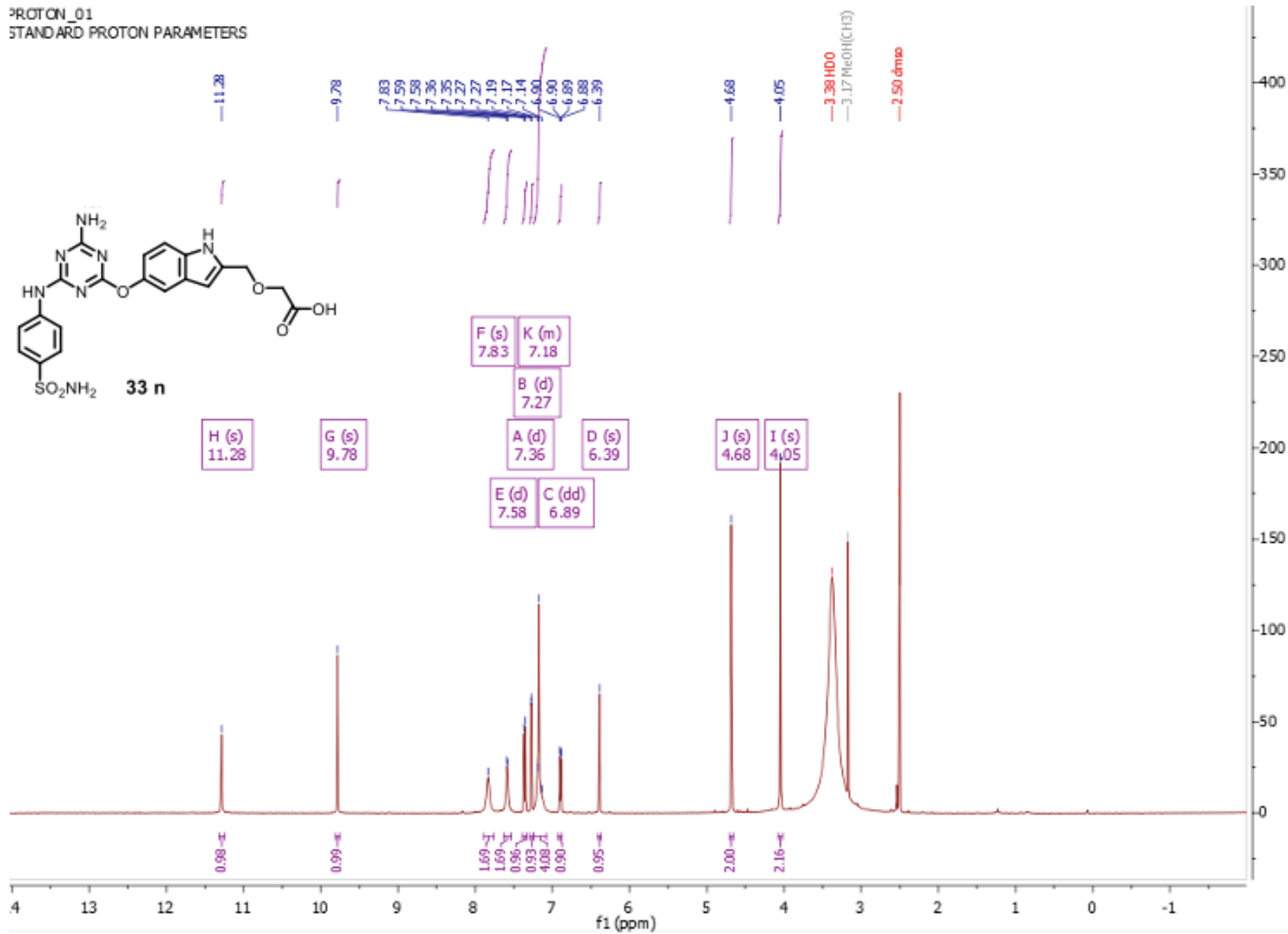
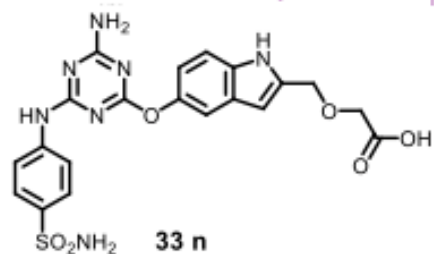




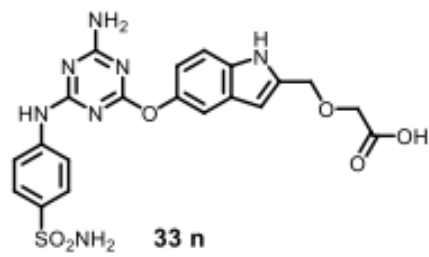
Id395carb_CARBON_01



PROTON_01
STANDARD PROTON PARAMETERS



CARBON_01
STANDARD CARBON PARAMETERS



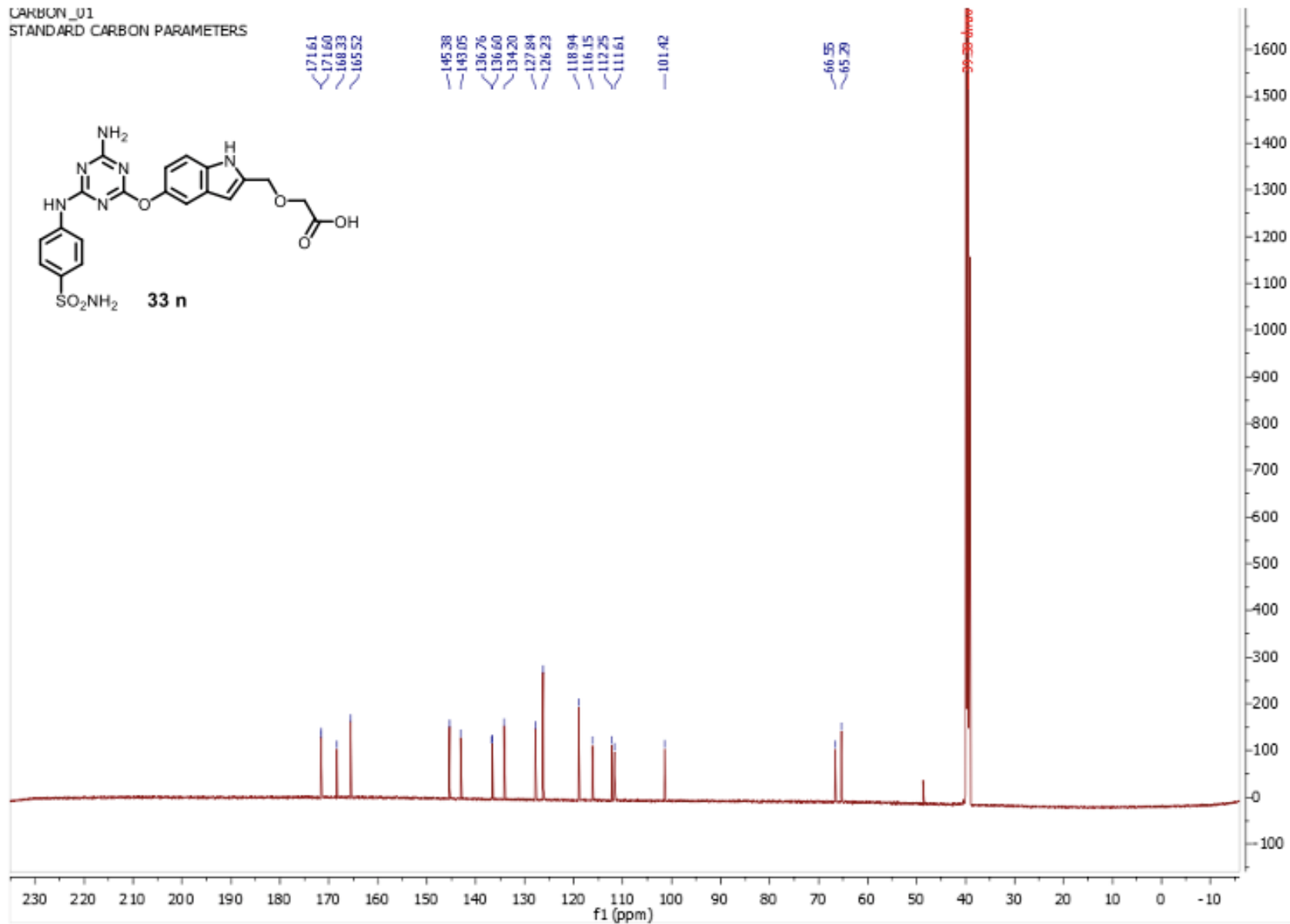
171.61
171.60
168.33
165.52

145.38
143.05
136.76
136.60
134.20
127.84
126.23

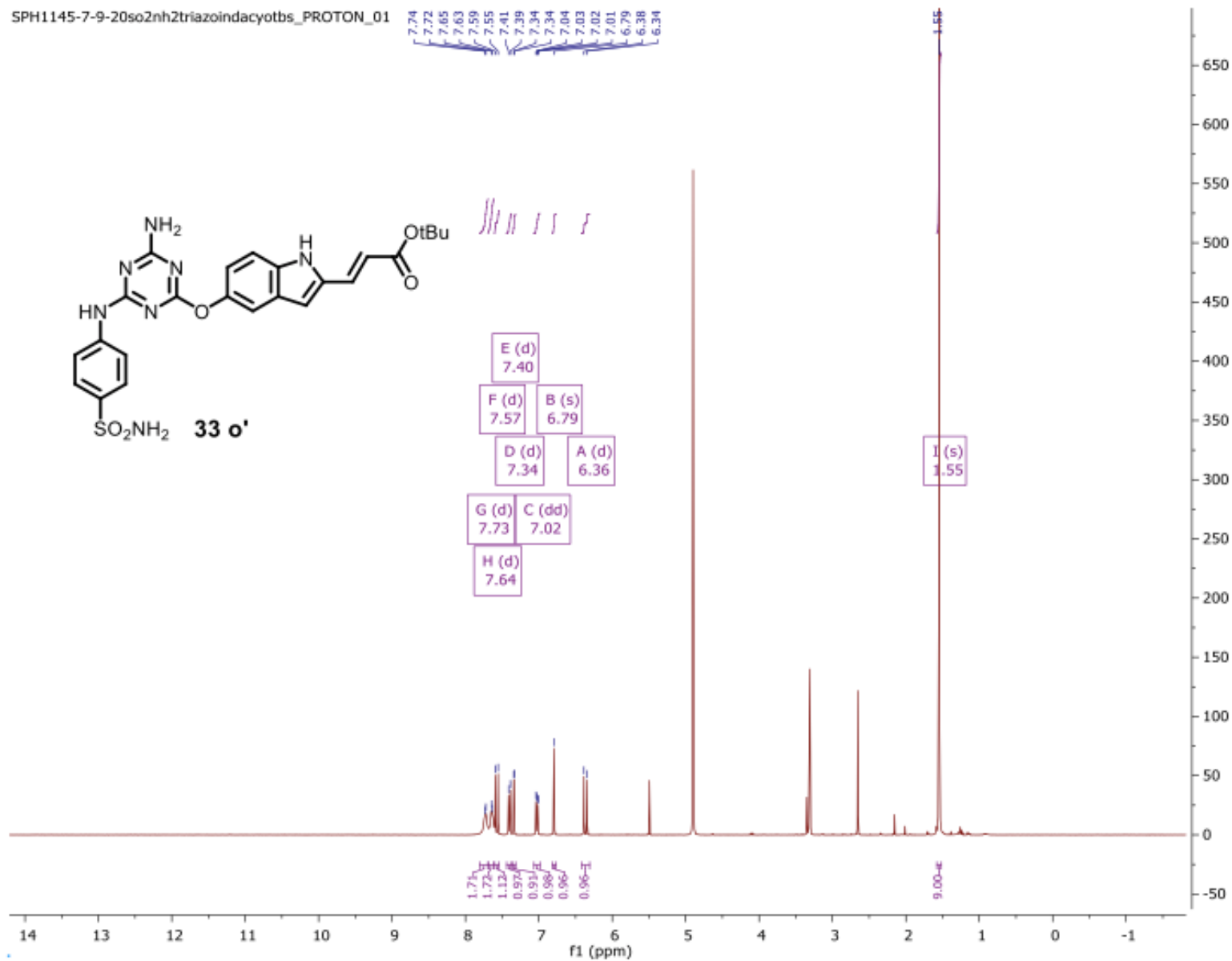
118.94
116.15
112.25
111.61
101.42

66.55
65.29

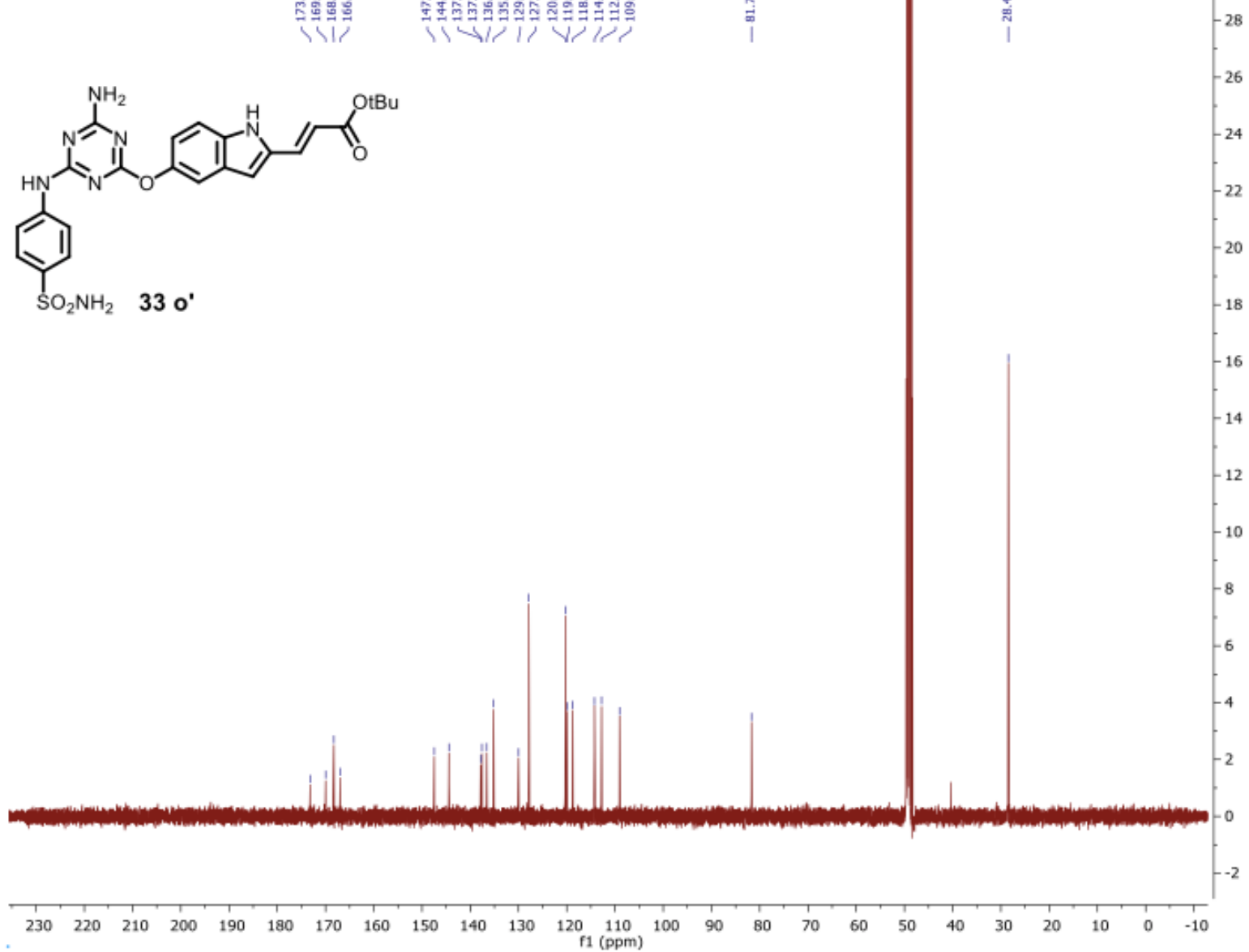
39.33 (DMSO-d₆)



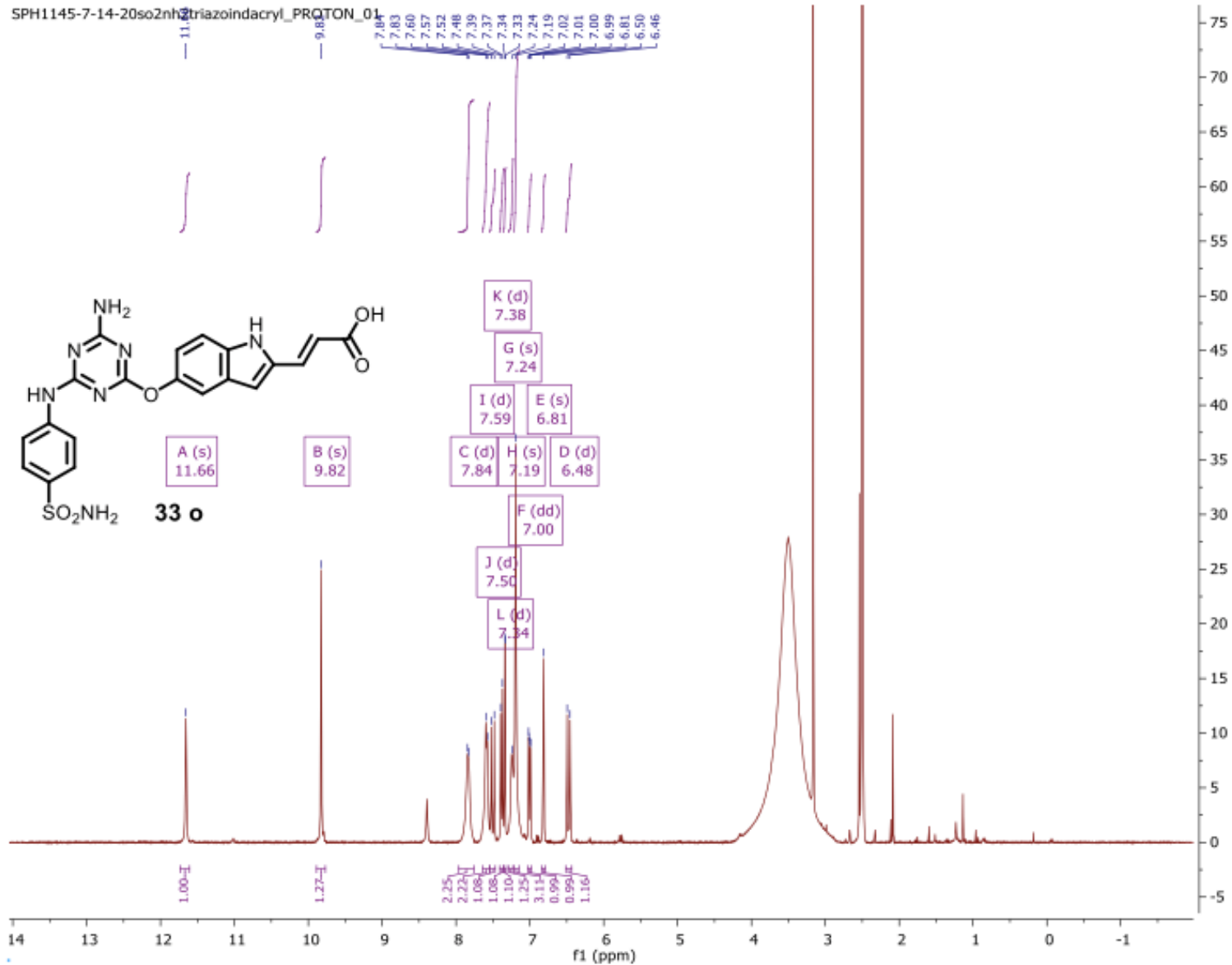
SPH1145-7-9-20so2nh2triazoindacotybs_PROTON_01



SPH1145-7-9-20so2nh2triazoindecryotbu CARBON 13C

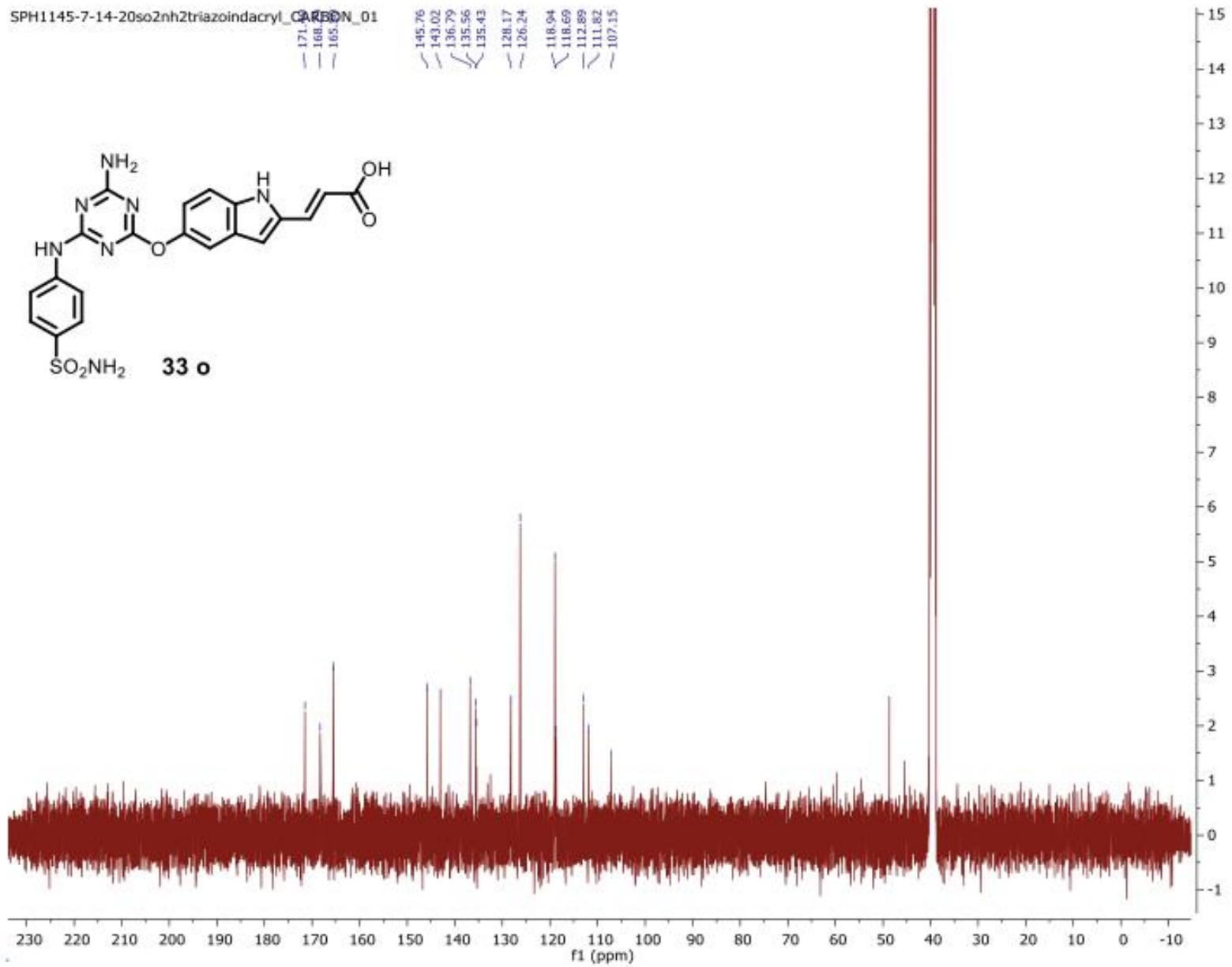
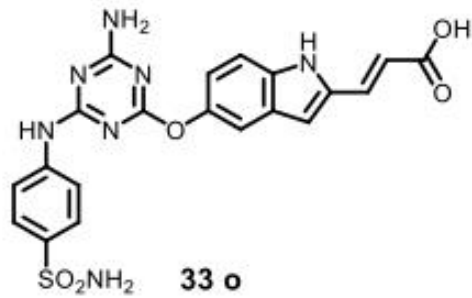


SPH1145-7-14-20so2nh2triazolindacryl_PROTON_01



SPH1145-7-14-20so2nh2triazolindacryl_CARBON_01

171.168165
145.76
143.02
136.79
135.56
135.43
128.17
126.24
118.94
118.69
112.89
111.82
107.15



4. Structure Determination by X-ray Crystallography

Protein expression and purification was performed identical to a previously described protocol [11]. JAK2 JH2 protein at a concentration of 6.0 mg/mL in a buffer consisting of 20 mM Tris pH 8.0, 100 mM NaCl, 10% glycerol, and 1 mM TCEP was crystallized by hanging drop vapor diffusion (room temperature) using reservoir solution consisting of 0.1 M Tris pH 8.0, 0.2 M sodium acetate, 12–24% PEG4000, and 1 mM TCEP. Crystal growth was initiated by streak seeding. Protein-ligand complexes were prepared by transferring empty JAK2 JH2 crystals into reservoir solution containing 4 mM small-molecule ligand and 8% DMSO for 24 h. Prior to freezing, crystals were cryo-protected by briefly exposing them to reservoir solution containing 20% glycerol.

Diffraction data were collected in-house on Rigaku MicroMax-007HF X-ray generator ($\lambda = 1.54 \text{ \AA}$) equipped with a Dectris Pilatus 200K detector. Datasets were indexed, integrated, and scaled using HKL2000 [12]. Phases were calculated by molecular replacement (search model: 4FVQ) using Phaser [13]. Structure refinement was performed using Phenix version 1.11.1-2575 [14]. Cartesian simulated annealing was performed as initial step of refinement, followed by refinement of XYZ coordinates, occupancies, and individual B factors. Manual model building was performed using Coot [15]. Small-molecule ligands' restraints were prepared using Phenix eLBOW [16]. Crystallographic data collection and refinements statistics are shown in Table ES2.

Table ES2. Crystallographic data collection and refinement statistics

| | small-molecule ligand | | |
|---|---------------------------|---------------------------|---------------------------|
| | 22 | 33b | 33m |
| PDB ID | 7JYQ | 7JYO | 6XJK |
| space group | $P2_1$ | $P2_1$ | $P2_1$ |
| unit cell parameters: | | | |
| <i>a</i> , <i>b</i> , <i>c</i> (Å) | | 44.6, 57.4, | 45.0, 57.6, |
| <i>α</i> , <i>β</i> , <i>γ</i> (°) | 44.4, 57.3, | 61.1, | 61.2, |
| | 61.0, | 90.0, 111.0, | 90.0, 111.5, |
| | 90.0, 110.6 | 90.0 | 90.0 |
| Matthews coefficient (Å ³ /Da) ^a | 47.0 | 47.0 | 47.0 |
| solvent content (%) ^a | 2.3 | 2.3 | 2.3 |
| | Diffraction data | | |
| resolution range (Å) | 50.00–1.86 (1.93–1.86) | 50.00–2.16 (2.24–2.16) | 50.00–2.02 (2.05–2.02) |
| unique reflections | 24049 (2297) | 15316 (1383) | 18548 (922) |
| <i>R</i> _{sym} (%) | 5.8 (22.6) | 5.7 (18.3) | 11.5 (54.5) |
| <i>CC</i> 1/2 | 0.970 (0.920) | 0.979 (0.930) | 0.930 (0.734) |
| $\langle I/\sigma(I) \rangle$ | 19.7 (3.3) | 15.2 (3.0) | 8.7 (2.1) |

| | | | |
|--|-------------|-------------|-------------|
| completeness (%) | 99.3 (96.5) | 98.5 (89.5) | 96.9 (96.8) |
| redundancy | 3.0 (2.6) | 3.0 (2.1) | 2.9 (2.6) |
| Wilson <i>B</i> factor (Å ²) | 21.2 | 24.5 | 23.8 |

Refinement

| | | | |
|---|------------|------------|------------|
| resolution range (Å) | 29.11–1.86 | 40.44–2.16 | 40.50–2.02 |
| reflections used (work/free) | 22852/1182 | 14548/751 | 17419/923 |
| <i>R</i> _{work} / <i>R</i> _{free} (%) | 17.9/22.1 | 18.5/24.6 | 22.7/27.3 |
| protein residues | 272 | 270 | 265 |
| inhibitor atoms | 23 | 26 | 29 |
| water molecules | 155 | 72 | 62 |
| RMSD from ideality: | | | |
| bond lengths (Å) | 0.006 | 0.007 | 0.007 |
| bond angles (°) | 0.8 | 0.8 | 0.8 |
| Ramachandran plot: ^b | | | |
| Ramachandran favored (%) | 98.9 | 96.6 | 97.7 |
| Ramachandran outliers (%) | 0.0 | 0.0 | 0.0 |
| Mean B factors (Å ²): | | | |
| protein | 22.4 | 29.8 | 28.5 |
| inhibitor | 31.4 | 25.3 | 29.0 |
| water molecules | 28.7 | 29.1 | 28.3 |

Values in parenthesis describe the highest resolution shell. ^aComputed with *CCP4 Matthews_coef* [17]. ^bComputed with *MolProbity* [18].

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