

Supporting Information

**Pseudo Natural Products—Chemical Evolution of  
Natural Product Structure**

*George Karageorgis, Daniel J. Foley, Luca Laraia, Susanne Brakmann, and  
Herbert Waldmann\**

anie\_202016575\_sm\_miscellaneous\_information.pdf

### **Author Contributions**

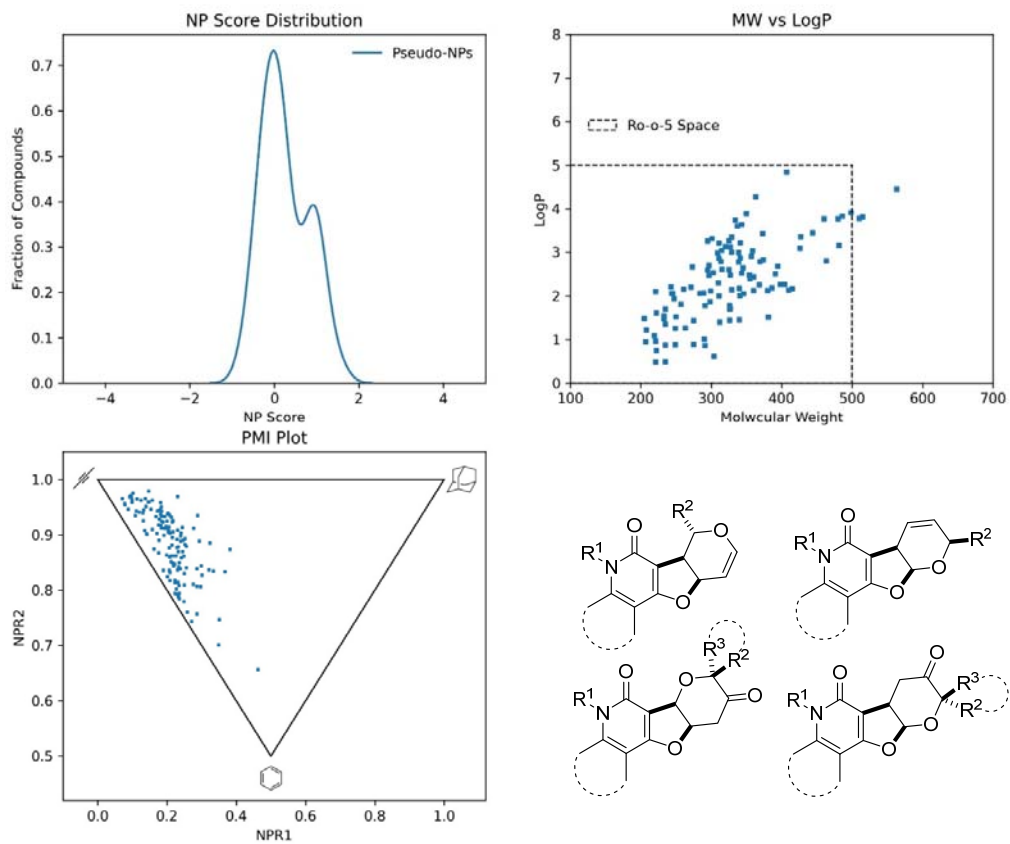
G.K. performed the cheminformatic analyses. All authors wrote and commented on the manuscript.

Molecular properties were calculated using RDKit (<http://www.rdkit.org>) running in Python 3.7.

Plots were generated using Matplotlib.<sup>[1]</sup>

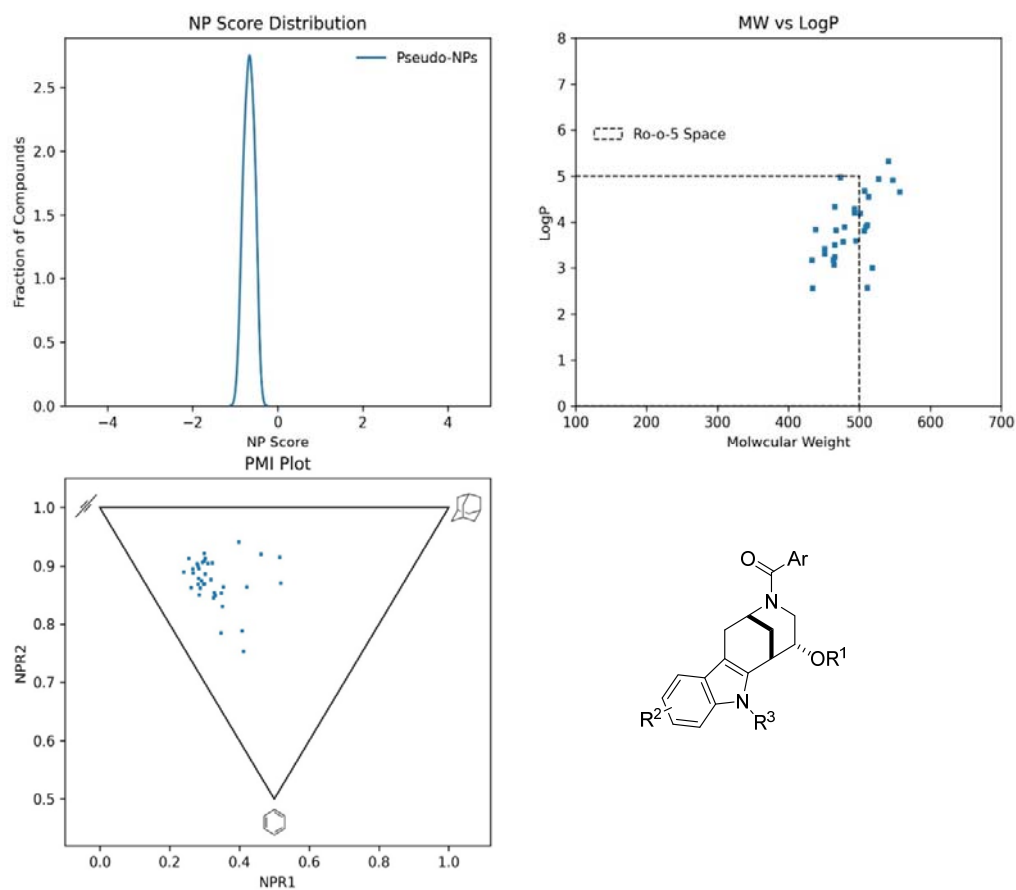
## Molecular properties of individual pseudo-NP classes used for the cheminformatic analyses:

### Pyranofuranopyridones<sup>[2]</sup>



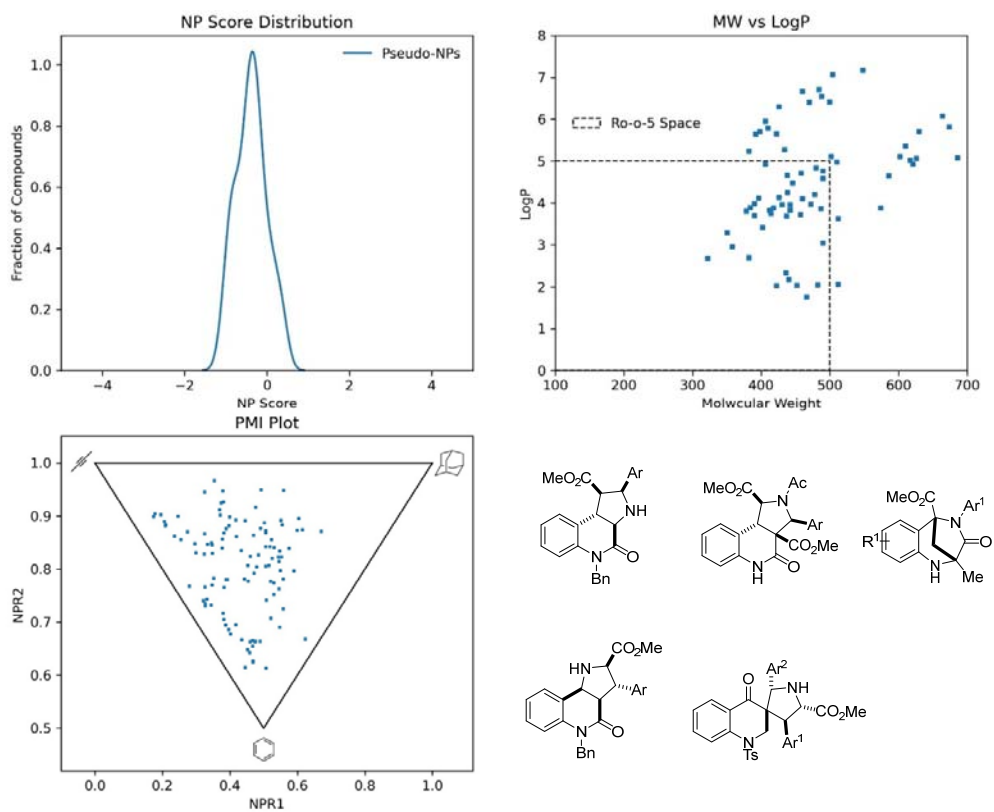
**Figure S1:** Molecular properties of pyranofuranopyridone pseudo-NPs. Top left: NP Score distribution. Top right: Molecular Weight against LogP. Dashed box denotes "Rule-of-five" space. Bottom left: PMI plot<sup>[3]</sup> of pyranofuranopyridone scaffold derivatives. Bottom right: Pyranofuranopyridone scaffolds structures.

## Indomorphans<sup>[4]</sup>



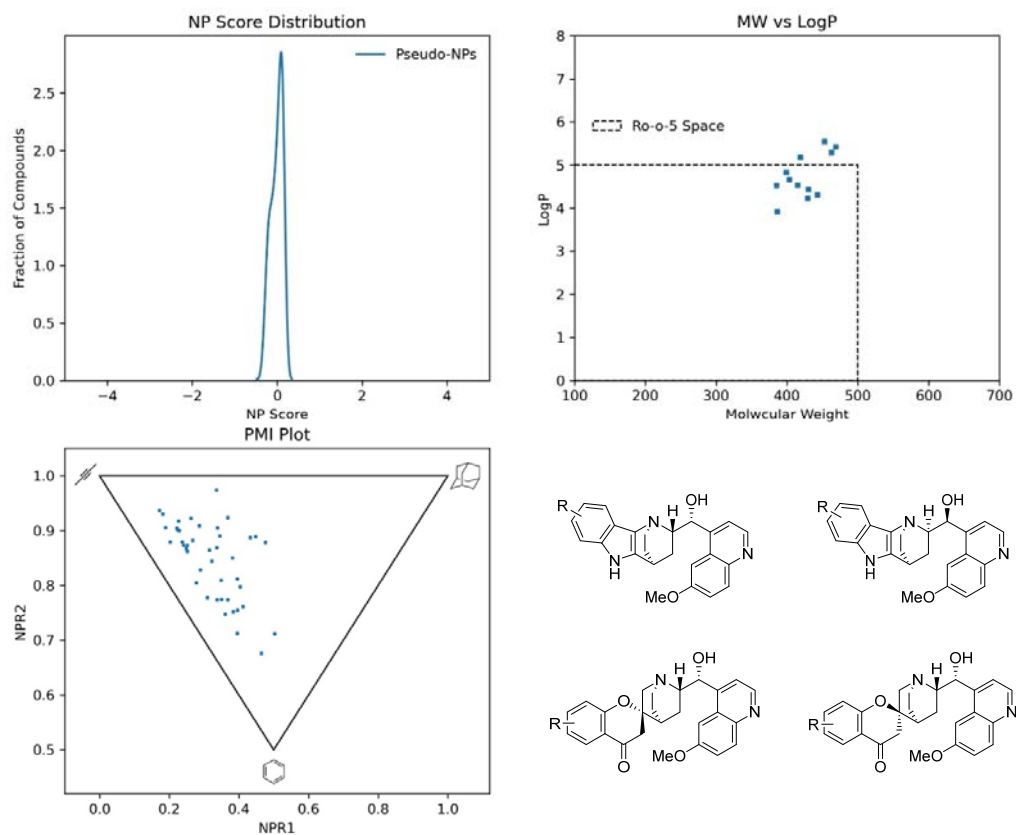
**Figure S2:** Molecular properties of indomorphan pseudo-NPs. Top left: NP Score distribution. Top right: Molecular Weight against LogP. Dashed box denotes “Rule-of-five” space. Bottom left: PMI plot<sup>[3]</sup> of indomorphan scaffold derivatives. Bottom right: Indomorphan scaffold structures.

## Pyrroquinolines<sup>[5]</sup>



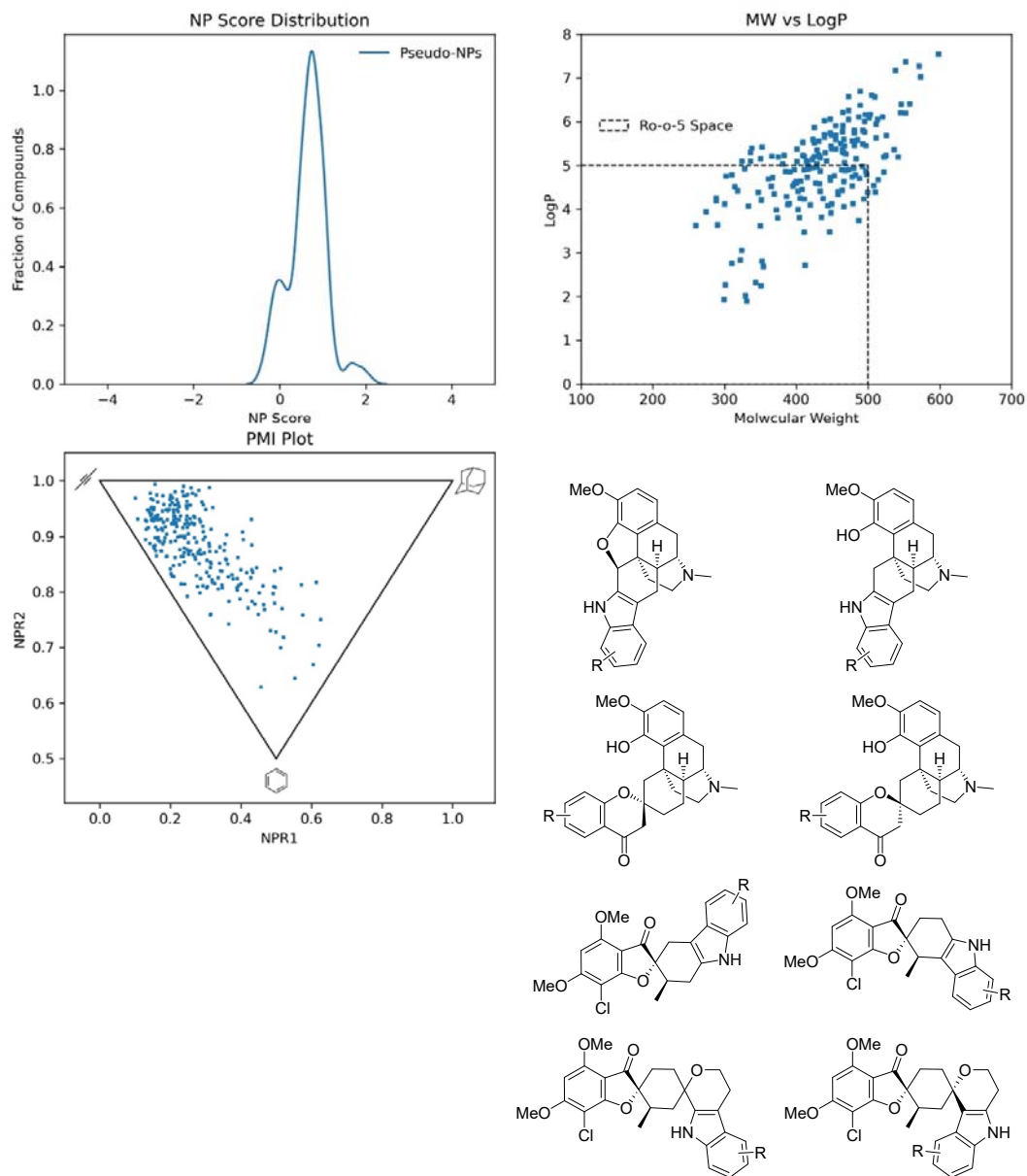
**Figure S3:** Molecular properties of pyrroquinoline pseudo-NPs. Top left: NP Score distribution. Top right: Molecular Weight against LogP. Dashed box denotes “Rule-of-five” space. Bottom left: PMI plot<sup>[3]</sup> of pyrroquinoline scaffold derivatives. Bottom right: pyrroquinoline scaffold structures.

Pseudo-NPs based on cinchona alkaloids<sup>[6,7]</sup>



**Figure S4:** Molecular properties of cinchona pseudo-NPs. Top left: NP Score distribution. Top right: Molecular Weight against LogP. Dashed box denotes “Rule-of-five” space. Bottom left: PMI plot<sup>[3]</sup> of cinchona pseudo-NP derivatives. Bottom right: cinchona pseudo-NP scaffold structures.

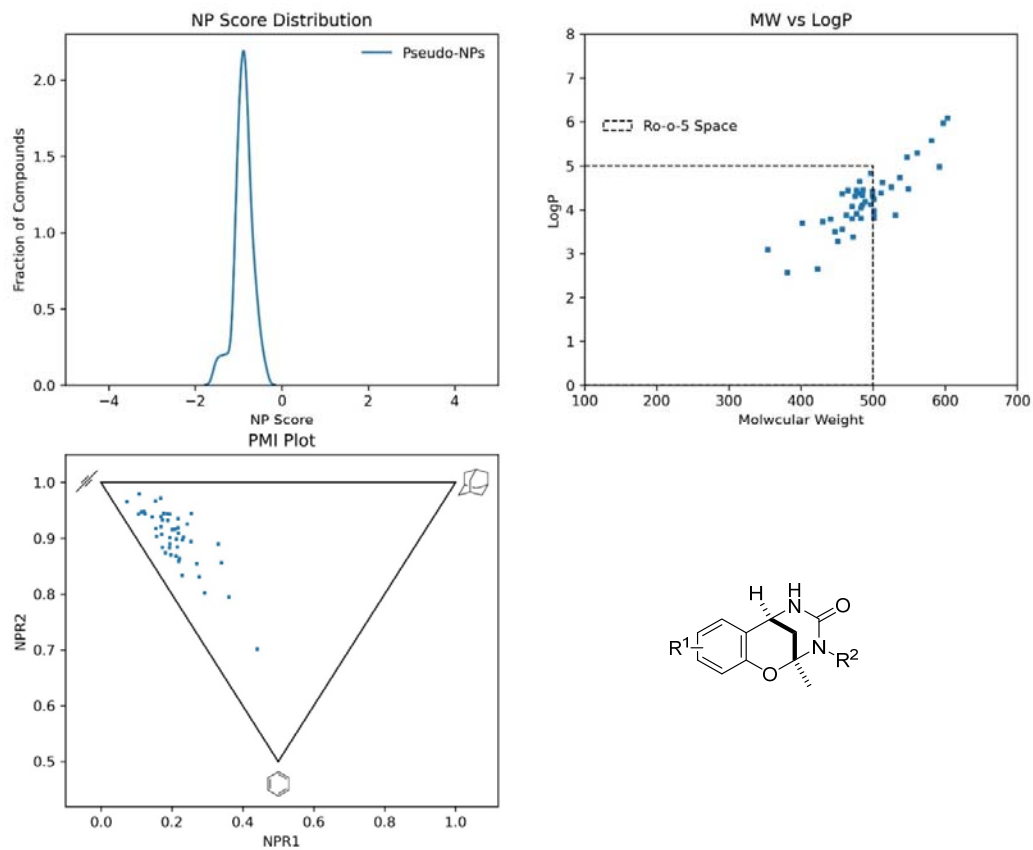
## Pseudo-NPs based on sinomenine, and griseofulvin<sup>[7]</sup>



**Figure S5:** Molecular properties of sinomenine, and griseofulvin pseudo-NPs. Top left: NP Score distribution. Top right: Molecular Weight against LogP. Dashed box denotes “Rule-of-five” space. Bottom left: PMI plot<sup>[3]</sup> of sinomenine, and griseofulvin pseudo-NP derivatives. Bottom right: sineomenine, and griseofulvin pseudo-NP scaffold structures.



## Chromopyrones<sup>[8]</sup>

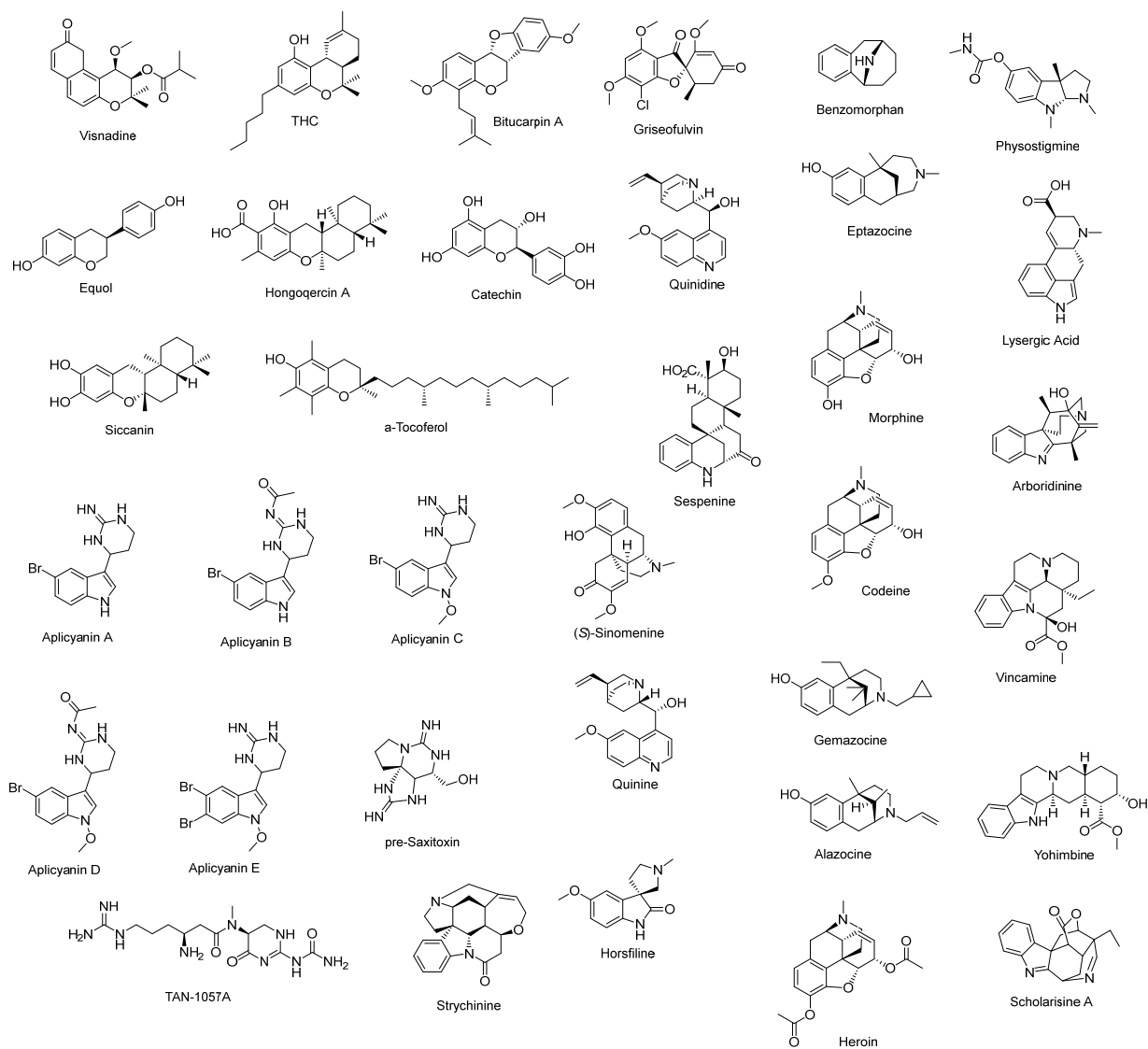


**Figure S6:** Molecular properties of chromopynone pseudo-NPs. Top left: NP Score distribution. Top right: Molecular Weight against LogP. Dashed box denotes “Rule-of-five” space. Bottom left: PMI plot<sup>[3]</sup> of chromopynone scaffold derivatives. Bottom right: chromopynone scaffold structure.

Table S1: Molecular properties of combined pseudo-NP libraries, averages and ranges.

	Mean	Range	Lit. Guidelines
Molecular Weight	426	205 - 686	150-500
LogP	4.26	0.47 - 7.61	0-5
Hydrogen-bond Acceptors	5	1 - 10	< 10
Hydrogen-bond Donors	1	0 - 3	< 5
Total Polar Surface Area (Å <sup>2</sup> )	64	8 - 136	< 140
Fraction of <i>sp</i> <sup>3</sup> C atoms	0.36	0.08 - 0.72	0.3-0.6

Literature guidelines referred in literature<sup>[9-13]</sup>



**Figure S7:** Structures and names for NPs and bioactive compounds used to prepare the PMI plot II in Figure 3.

## References:

- [1] J. D. Hunter, *Comput. Sci. Eng.* **2007**, 9, 90–95.
- [2] A. Christoforow, J. Wilke, A. Binici, A. Pahl, C. Ostermann, S. Sievers, H. Waldmann, *Angew. Chem. Int. Ed.* **2019**, 58, 14715–14723.
- [3] W. H. B. Sauer, M. K. Schwarz, *J. Chem. Inf. Comput. Sci.* **2003**, 43, 987–1003.
- [4] J. Ceballos, M. Schwalfenberg, G. Karageorgis, E. S. Reckzeh, S. Sievers, C. Ostermann, A. Pahl, M. Sellstedt, J. Nowacki, M. A. Carnero Corrales, et al., *Angew. Chem. Int. Ed.* **2019**, 58, 17016–17025.
- [5] H. Waldmann, J. Liu, G. S. Cremosnik, F. Otte, A. Pahl, S. Sievers, C. Strohmam, *Angew. Chem. Int. Ed.* **2020**, anie.202013731.
- [6] D. J. Foley, S. Zinken, D. Corkery, L. Laraia, A. Pahl, Y.-W. Wu, H. Waldmann, *Angew. Chem. Int. Ed.* **2020**, 59, 12470–12476.
- [7] M. Grigalunas, A. Burhop, S. Zinken, A. Pahl, S. Sievers, D. J. Foley, A. P. Antonchick, H. Waldmann, *Nat. Commun, Revis.* **n.d.**
- [8] G. Karageorgis, E. S. Reckzeh, J. Ceballos, M. Schwalfenberg, S. Sievers, C. Ostermann, A. Pahl, S. Ziegler, H. Waldmann, *Nat. Chem.* **2018**, 10, 1103–1111.
- [9] D. B. Kell, S. G. Oliver, *Front. Pharmacol.* **2014**, 5, 1–32.
- [10] C. A. Lipinski, F. Lombardo, B. W. Dominy, P. J. Feeney, *Adv. Drug Deliv. Rev.* **1997**, 23, 3–25.
- [11] D. F. Veber, S. R. Johnson, H.-Y. Cheng, B. R. Smith, K. W. Ward, K. D. Kopple, *J. Med. Chem.* **2002**, 45, 2615–2623.
- [12] F. Lovering, *Medchemcomm* **2013**, 4, 515–519.
- [13] F. Lovering, J. Bikker, C. Humblet, *J. Med. Chem.* **2009**, 52, 6752–6756.