

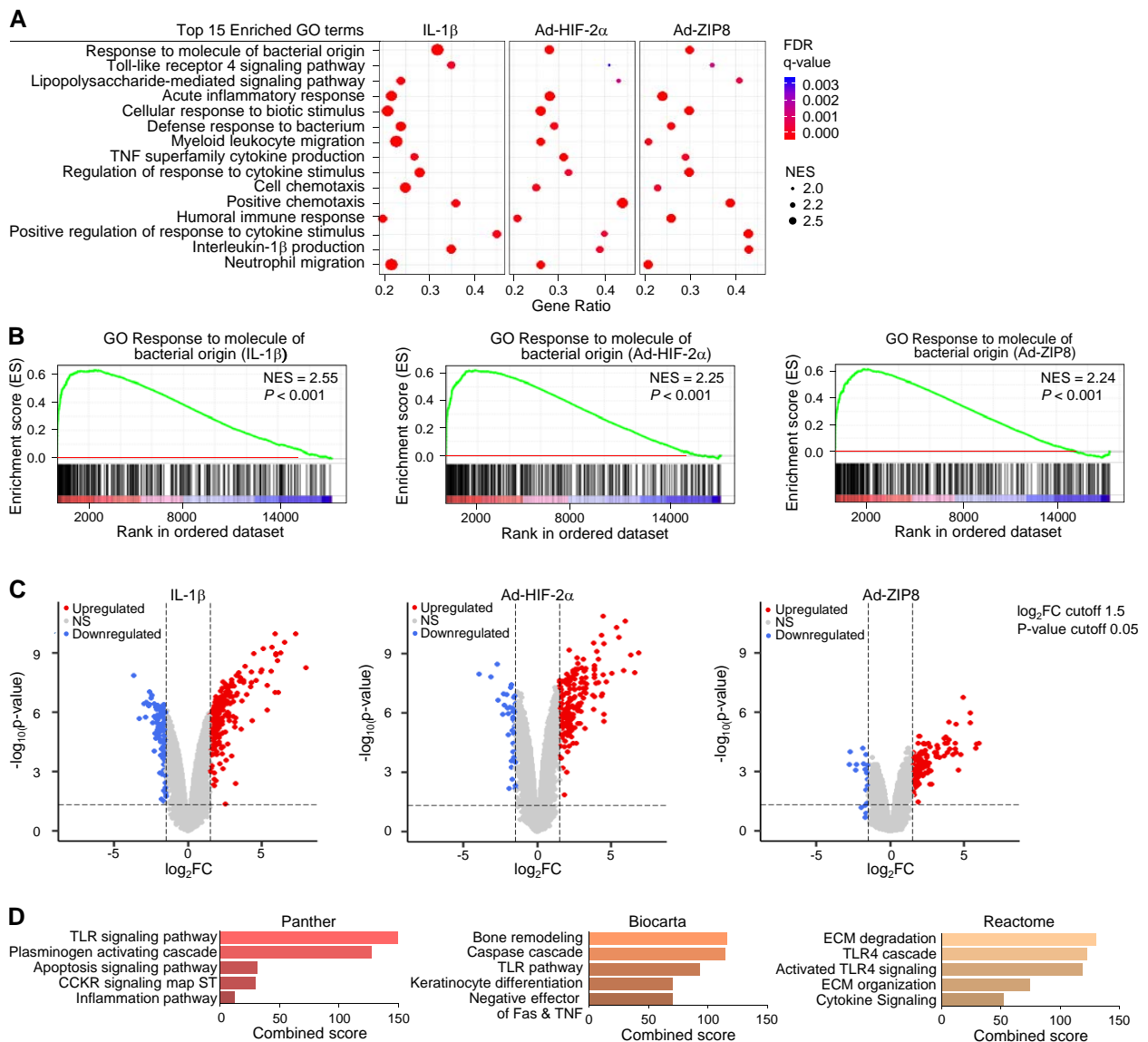
Supplementary Materials

LBP and CD14, cofactors of toll-like receptors, are essential for low-grade inflammation induced exacerbation of post-traumatic osteoarthritis

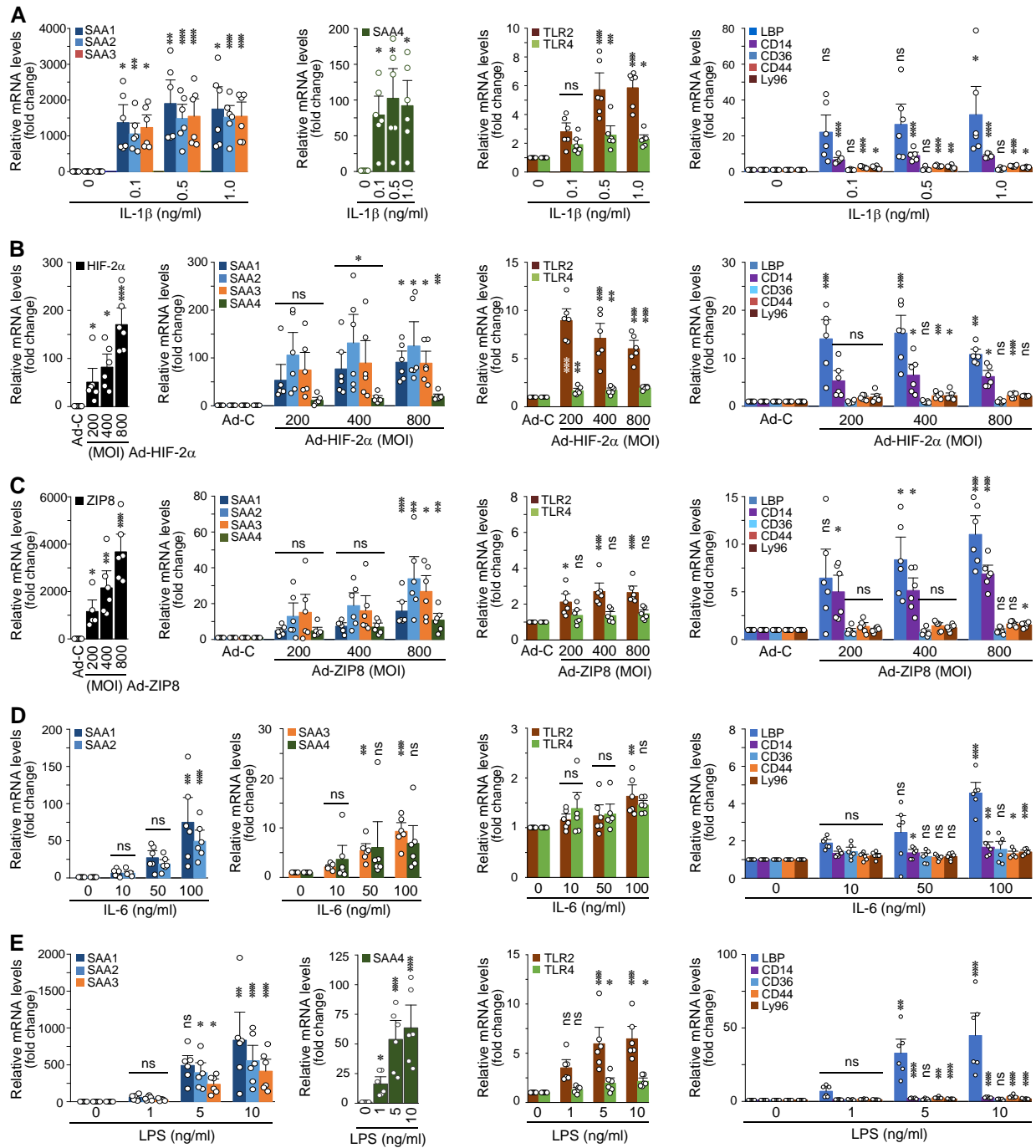
Yoonkyung Won, Jeong-In Yang, Seulki Park, Jang-Soo Chun

Supplementary Figures 1–9

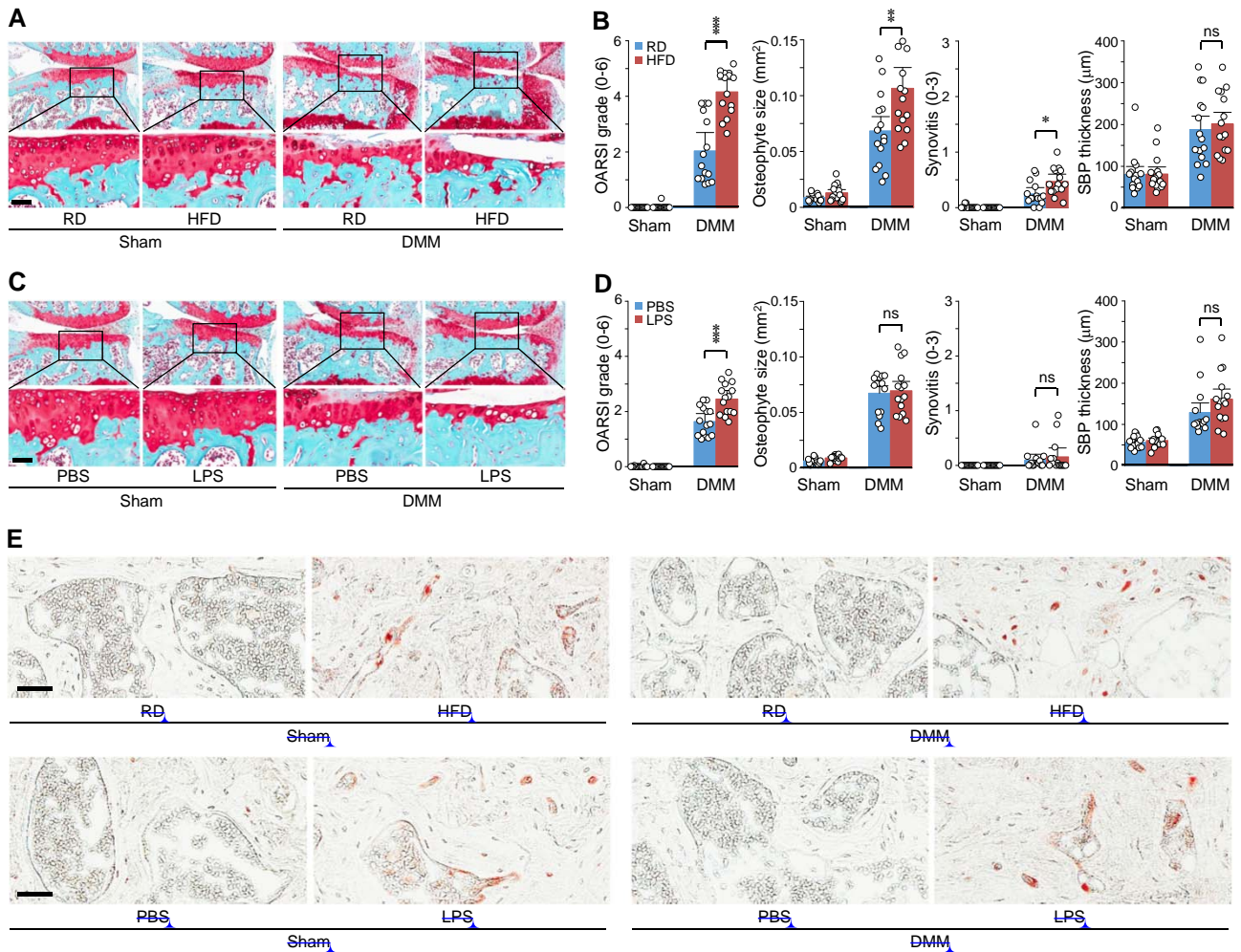
Supplementary Tables 1,2



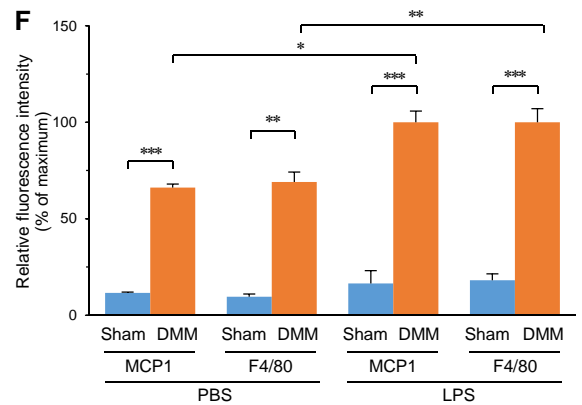
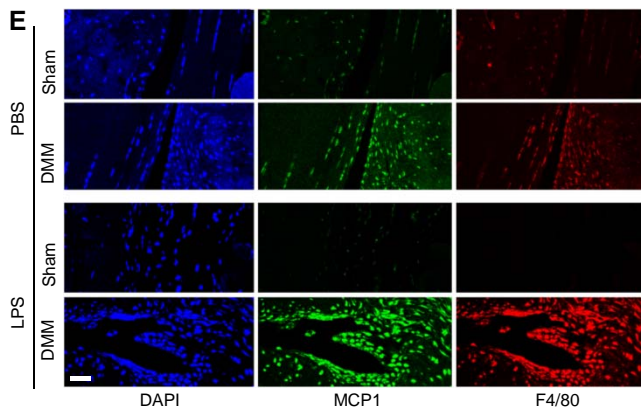
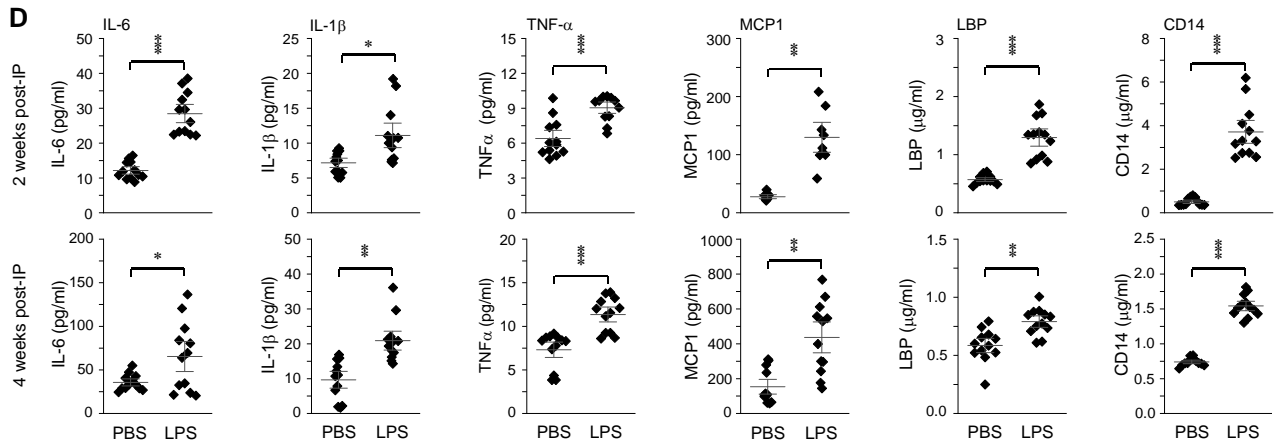
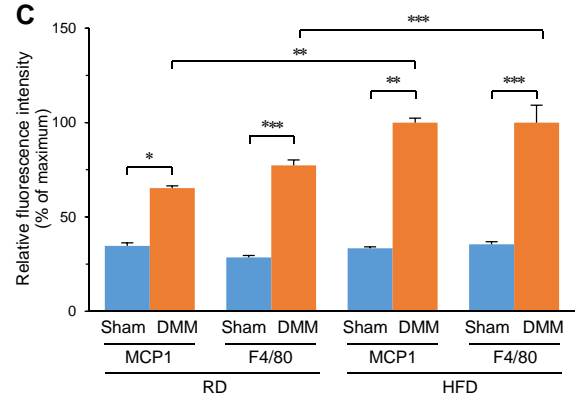
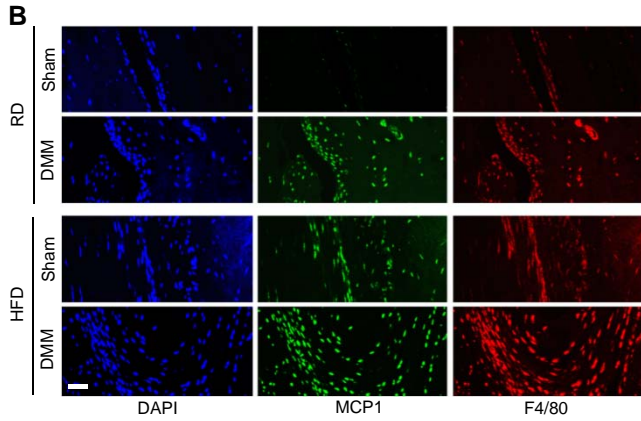
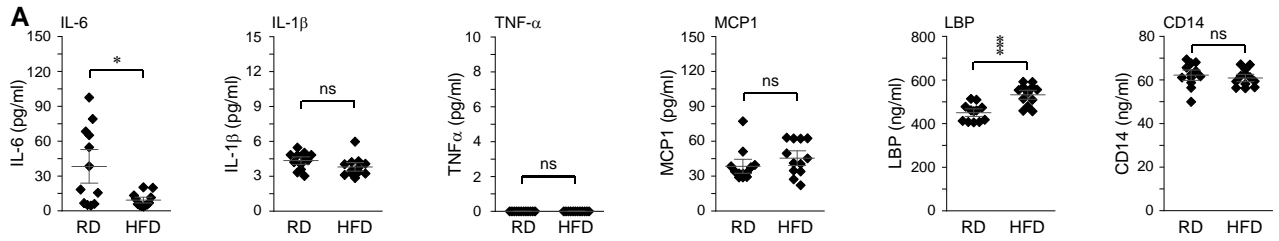
Supplementary Figure 1. Transcriptional profiling of mouse chondrocytes treated with OA-associated pathogenic catabolic mediators. Microarray data were obtained from primary-culture mouse articular chondrocytes that had been treated for 36 h with IL-1 β (1 ng/ml) or infected with 800 MOI of Ad-C, Ad-HIF-2 α , or Ad-ZIP8. **A**, Enrichment plot of the top 15 most highly enriched gene ontology (GO) biological process terms with normalized enrichment scores (NES). **B**, GSEA of microarray signals on GO biological processes. **C**, Volcano plots for upregulated and downregulated genes. $P < 0.05$ and absolute value of $\log_2(\text{fold change}) > 1.5$ were applied as threshold values. Upregulated gene transcripts are shown as red dots, downregulated genes are shown as blue dots, and genes with non-significant (NS) changes in expression levels are shown as gray dots. **D**, Five top-ranked annotations from Enrichr pathway analysis of the genes found to be upregulated in OA chondrocytes in the Panther, Biocarta, or Reactome pathway databases, and the corresponding combined scores are presented as bar graphs. The combined score is a representative parameter of the significance of the enrichment provided by Enrichr.



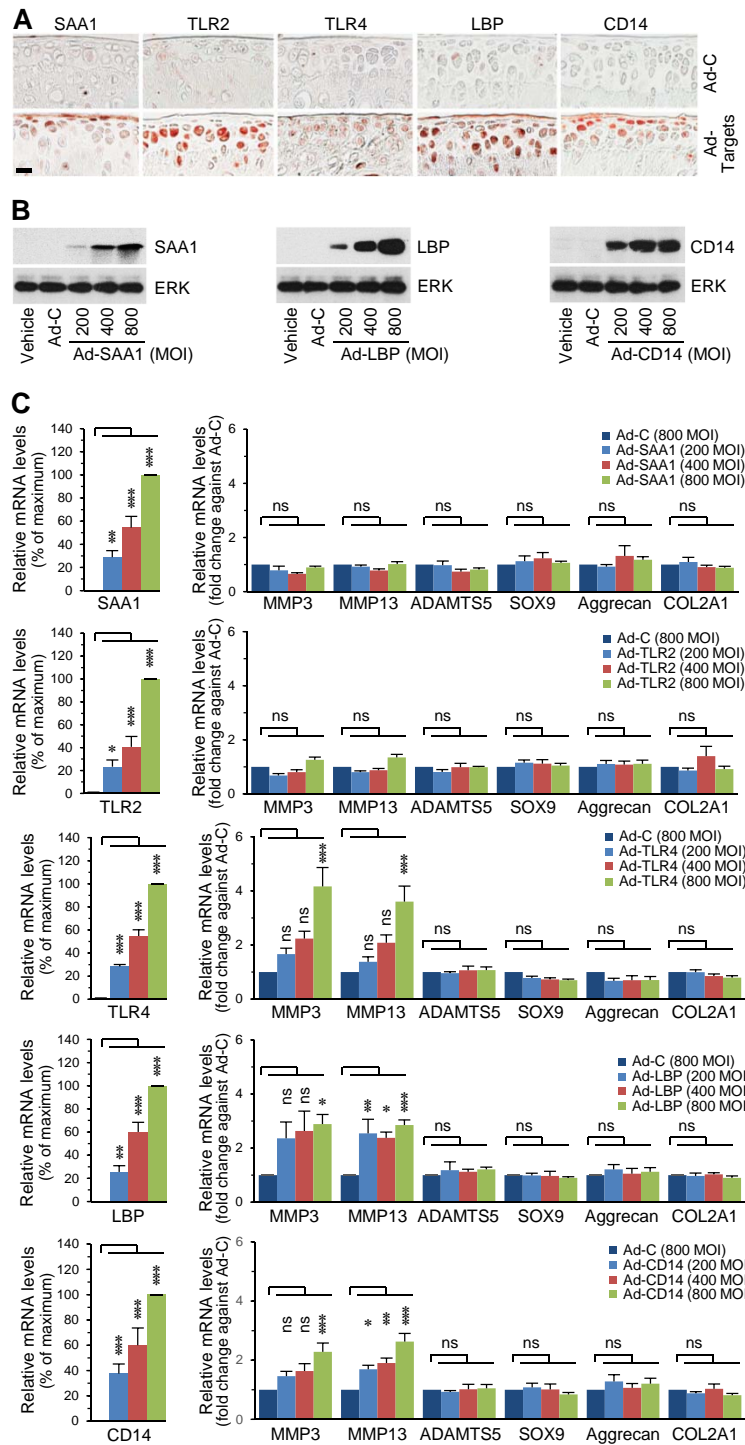
Supplementary Figure 2. Upregulation of TLR signaling components in chondrocytes stimulated with OA-associated catabolic signaling. **A–E**, Primary-culture mouse articular chondrocytes were treated for 36 h with the indicated concentrations of IL-1 β (**A**), Ad-HIF2 α (**B**), Ad-ZIP8 (**C**), IL-6 (**D**), or LPS (**E**). Relative mRNA levels (fold change against control) of the indicated molecules were determined by qRT-PCR analysis ($n = 6$). Means \pm s.e.m.; assessed by one-way ANOVA with *post-hoc* Bonferroni test. * = $P < 0.05$, ** = $P < 0.005$, *** = $P < 0.001$. ns, not significant.



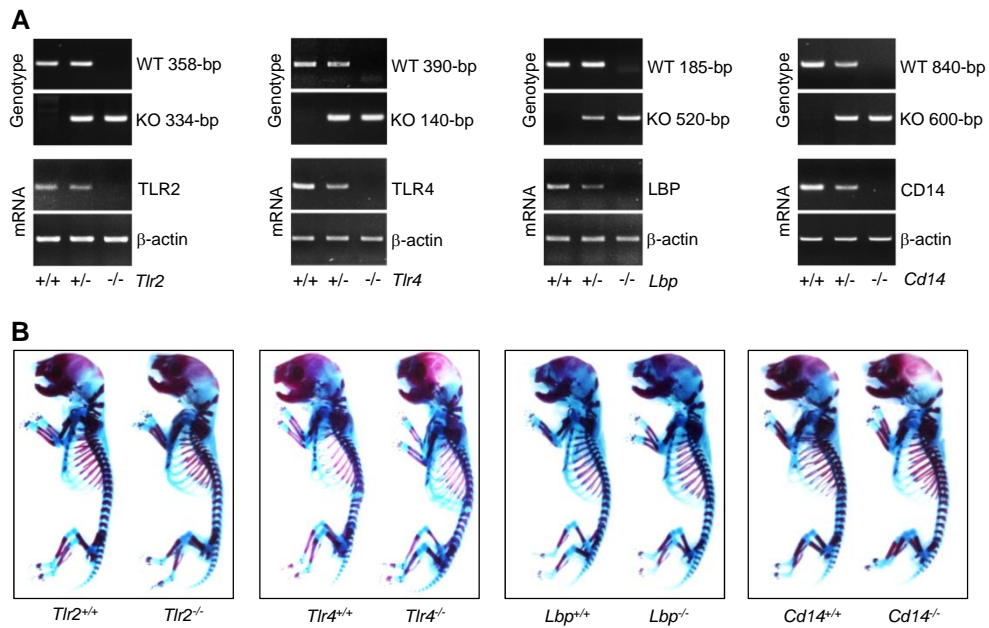
Supplementary Figure 3. HFD and metabolic endotoxemia enhance DMM-induced post-traumatic OA in mice. **A** and **B**, Mice were fed with RD or HFD, subject to DMM surgery or sham operation ($n = 15$ mice per group), and sacrificed at 6 weeks after the operation. Presented are representative Safranin-O staining images of joint sections (**A**) and scoring of OARSIS grade, osteophyte size, synovitis, and SBP thickness (**B**). **C** and **D**, Sham- or DMM-operated mice were administrated with PBS or LPS ($n = 15$ mice per group) for 4 weeks to induce metabolic endotoxemia and sacrificed at 8 weeks post-surgery. Presented are representative Safranin-O staining images of joint sections (**C**) and scoring of OA manifestations (**D**). **E**, Representative immunostaining images of Osterix in subchondral bone plate of the indicated mice. Values are means \pm 95% CI with Mann-Whitney U test for OARSIS grade and synovitis; means \pm s.e.m.; assessed by one-way ANOVA with *post-hoc* Bonferroni test for osteophyte size and SBP thickness. * = $P < 0.05$, ** = $P < 0.005$, *** = $P < 0.001$. ns, not significant. Scale bars, 50 μ m.



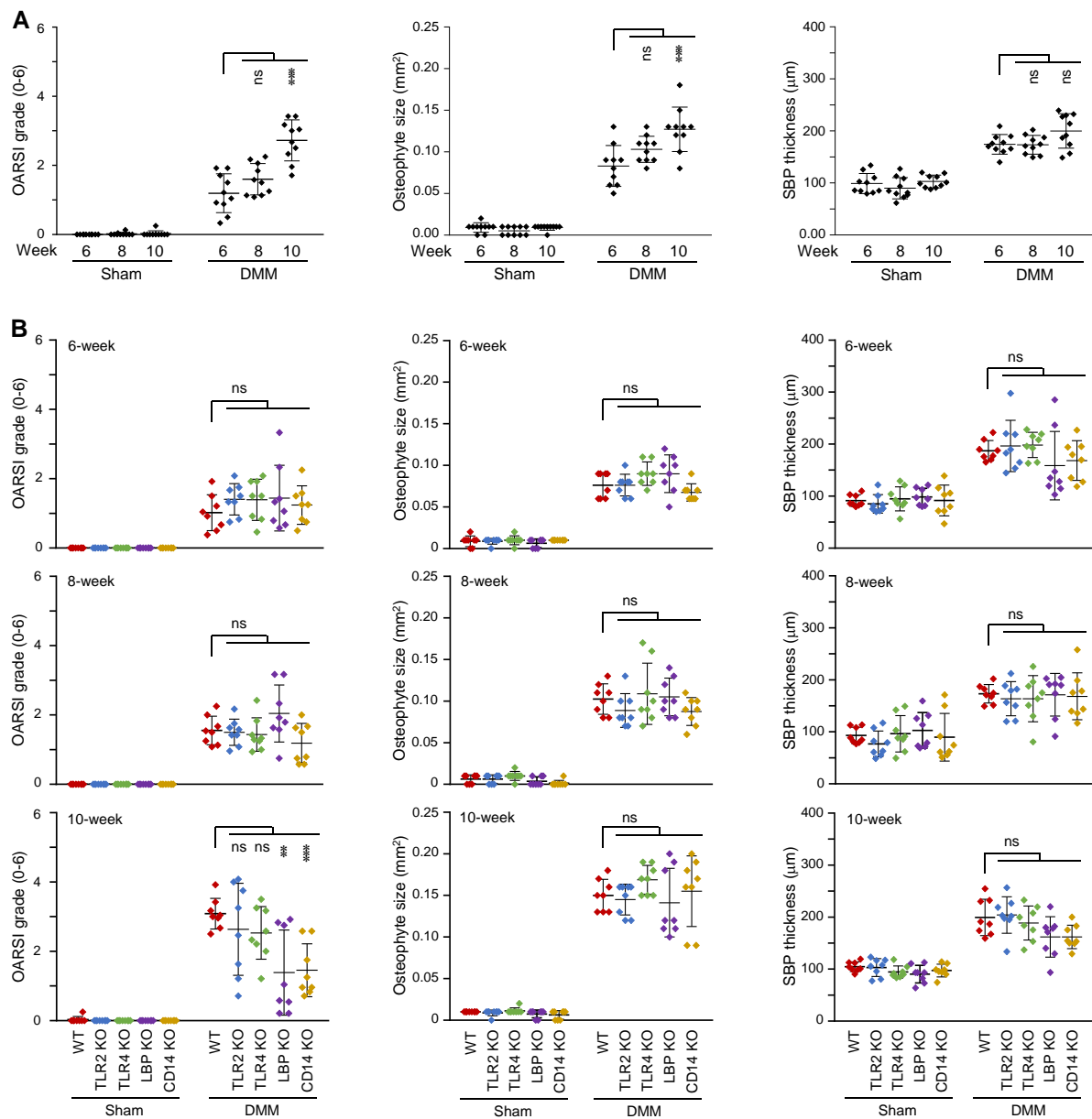
Supplementary Figure 4. Inflammatory responses in mice with HFD feeding or metabolic endotoxemia. **A–C**, Mice were fed with RD or HFD, subject to DMM surgery or sham operation, and sacrificed at 6 weeks after the operation. Concentrations of the indicated proteins in the sera were determined by ELISA (A, n = 12 mice per group). Synovial infiltration of MCP1- or F4/80-positive cells was determined by immunofluorescence microscopy (B, n = 4 mice per group) and quantified by using Image J software (C, n = 4 mice per group). **D–F**, Sham- or DMM-operated mice were administrated with PBS or LPS for 4 weeks and sacrificed at 8 weeks post-surgery. Serum concentrations of the indicated proteins at 2 or 4 weeks post-IP injection were determined by ELISA (D, n = 12 mice per group). Synovial infiltration of MCP1- or F4/80-positive cells was determined by immunofluorescence microscopy (B, n = 4 mice per group) and quantified by using Image J software (C, n = 4 mice per group) at 8 weeks post-DMM surgery. Values are presented as means \pm s.e.m. and were assessed by two-tailed *t*-test. Scale bars, 50 μ m.



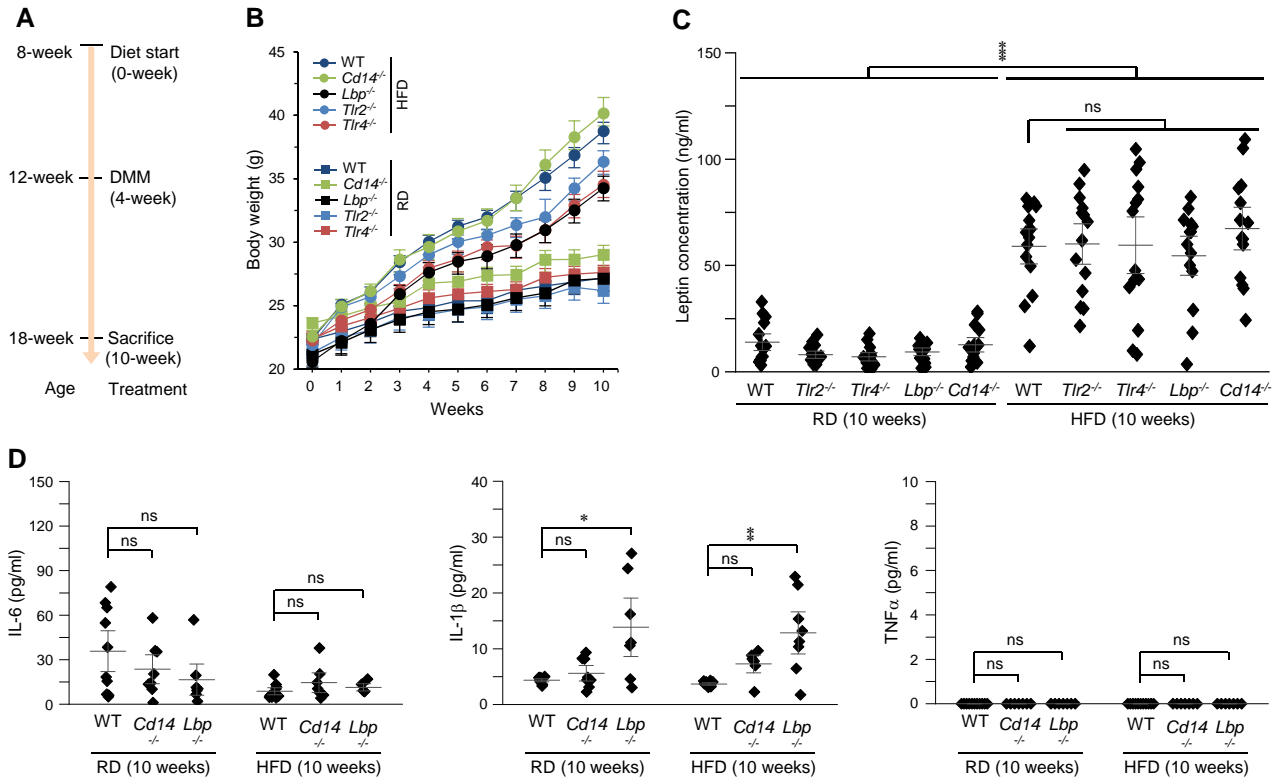
Supplementary Figure 5. Effect of overexpressing TLR signaling components in knee joint cartilage and primary-culture mouse articular chondrocytes. **A**, Representative immunostaining images of SAA1, TLR2, TLR4, LBP, and CD14 in knee-joint cartilage sections of mice that underwent IA injection (1×10^9 PFU in $10 \mu\text{l}$) of Ad-C, Ad-SAA1, Ad-TLR2, Ad-TLR4, Ad-LBP, or Ad-CD14 once weekly for 3 weeks ($n \geq 5$ mice per group). **B**, Western blot analysis of SAA1, LBP, and CD14 secreted by primary-culture mouse articular chondrocytes infected with Ad-C (800 MOI) or the indicated MOIs of Ad-SAA1, Ad-LBP, or Ad-CD14 for 36 h ($n \geq 5$). **C**, Relative mRNA levels of matrix-degrading enzymes and cartilage ECM molecules in chondrocytes infected with the indicated MOI of Ad-C, Ad-SAA1, Ad-TLR2, Ad-TLR4, Ad-LBP, or Ad-CD14 for 36 h ($n = 7$). Means \pm s.e.m.; assessed by one-way ANOVA with *post-hoc* Bonferroni test. * = $P < 0.05$, ** = $P < 0.005$, *** = $P < 0.001$. ns, not significant.



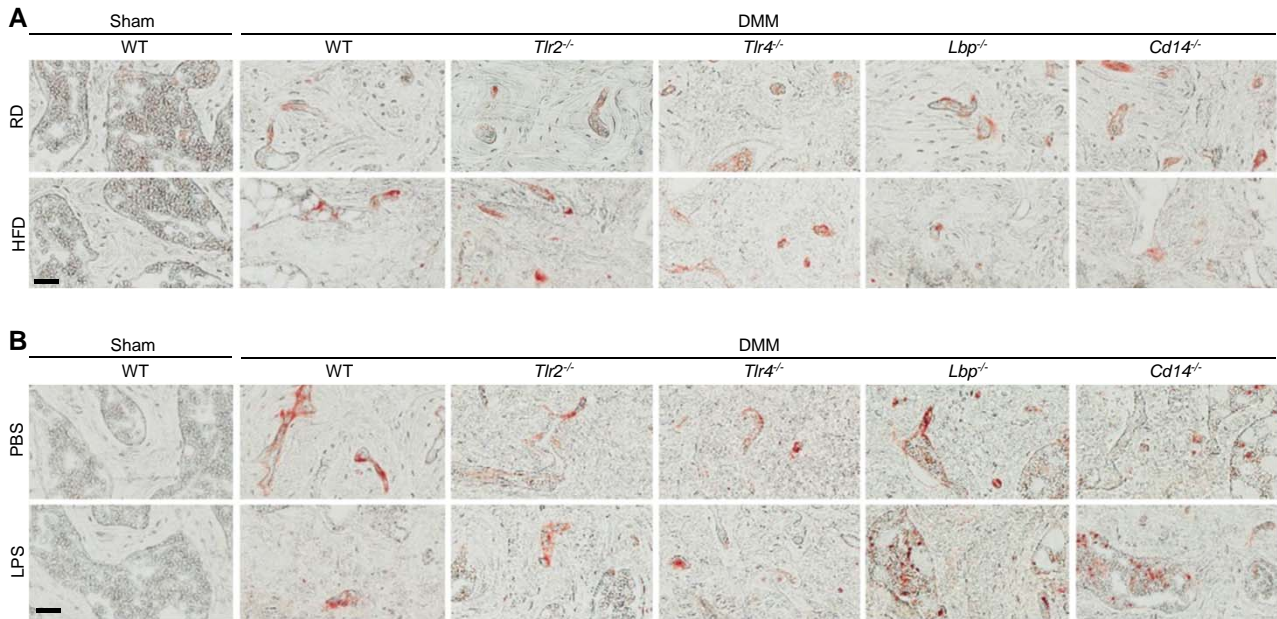
Supplementary Figure 6. Characterization of KO mice. **A**, Genotypes and mRNA levels of the KO-target genes in TLR2, TLR4, LBP, or CD14 heterozygous (+/-) and homozygous (-/-) KO mice and their WT (+/+) littermates ($n \geq 3$ mice per group). **B**, Representative skeletal staining images of E18.5 embryos of TLR2, TLR4, LBP, or CD14 homozygous (-/-) KO mice and their WT littermates ($n \geq 3$ mice per group).



Supplementary Figure 7. Characterization of DMM-induced post-traumatic OA in WT mice and KO mice of the indicated TLR signaling components. **A**, WT mice were subject to sham or DMM operation, and sacrificed at the indicated weeks after the operation. Presented are scoring of OARS I grade, osteophyte size, and SBP thickness ($n = 10$ mice per group). **B**, WT, *Tlr2*^{-/-}, *Tlr4*^{-/-}, *Lbp*^{-/-}, or *Cd14*^{-/-} mice were subject to sham or DMM operation, and sacrificed at the indicated weeks after the operation. Presented are scoring of OARS I grade, osteophyte size, and SBP thickness ($n = 8$ mice per group). Means \pm 95% CI; assessed by Mann-Whitney *U* test for OARS I grade or means \pm s.e.m.; assessed by one-way ANOVA with *post-hoc* Bonferroni test for osteophyte size and SBP thickness. * = $P < 0.05$, ** = $P < 0.005$, *** = $P < 0.001$. ns, not significant.



Supplementary Figure 8. Characterization of body weight and serum levels of inflammatory mediators in DMM-operated WT and KO mice fed with RD or HFD. **A**, Schematic illustration of HFD feeding schedule in mice. **B**, Body weights of the indicated mice fed with RD or HFD ($n = 15$ mice per group). **C**, Serum concentrations of leptin in WT and the indicated KO mice ($n = 14$ mice per group). **D**, Serum concentrations of the indicated inflammatory mediators in WT, *Cd14*^{-/-}, and *Lbp*^{-/-} mice ($n = 6\sim 8$ mice per group). Values are means \pm s.e.m.; assessed by one-way ANOVA with *post-hoc* Bonferroni test. * = $P < 0.05$, ** = $P < 0.005$, *** = $P < 0.001$. ns, not significant.



Supplementary Figure 9. Osterix expression in subchondral bone of WT and KO mice. **A**, WT and *Tlr2*^{-/-}, *Tlr4*^{-/-}, *Lbp*^{-/-}, or *Cd14*^{-/-} KO mice were fed with RD or HFD, subject to sham or DMM operation, and sacrificed 6 weeks after the operation. Presented are representative Osterix immunostaining images from subchondral bone of joint sections (n = 10 mice per group). **B**, Sham- or DMM-operated WT, *Tlr2*^{-/-}, *Tlr4*^{-/-}, *Lbp*^{-/-}, or *Cd14*^{-/-} mice were treated with PBS or a subclinical low-dose of LPS for 4 weeks, and sacrificed 8 weeks after the surgery. Presented are representative Osterix staining images from subchondral bone of joint sections (n = 10 mice per group). Scale bars, 50 μ m.

Supplementary Table 1. Sequences of primers.

Target (mouse)		Sequences	Purpose
ADAMTS5		S 5'-GCCATTGTAATAACCCTGCACC-3' AS 5'-TCAGTCCCATCCGTAACCTTTG-3'	RT-PCR
Aggrecan		S 5'-CTGTCTTTGTCACCCACACAT-3' AS 5'-GAAGACGACATCACCATCCAG-3'	RT-PCR
β-Actin		S 5'-ATATCGCTGCGCTGGTCGTC-3' AS 5'-AGGATGGCGTGAGGGAGAGC-3'	RT-PCR
CD14	WT Mutant Common	S 5'-CCAAGTTTTAGCGCTGCGTAAC-3' S 5'-CCGCTTCCATTGCTCAGCGG-3' AS 5'-GCCAGCCAAGGATACATAGCC-3'	Genotyping
CD14		S 5'-AATTGGGCGAGAGAGGACTGAT-3' AS 5'-AGCATCCCAGTGAATTGTGA-3'	RT-PCR
CD36		S 5'-TGTGGCTAAATGAGACTGGGAC-3' AS 5'-CACACACCACATTTCTTCTCC-3'	RT-PCR
CD44		S 5'-GAGGGCACAAGAAGAGTCATAC-3' AS 5'-CCAAGGCAAAGAGAAGGAAAGC-3'	RT-PCR
COL2A1		S 5'-CACACTGGTAAGTGGGGCAAGACCG-3' AS 5'-GGATTGTGTTGTTTCAGGGTTCGGG-3'	RT-PCR
HIF-2α (<i>Epas1</i>)		S 5'-CGAGAAGAACGACGTGGTGTTC-3' AS 5'-GTGAAGGCTGGCAGGCTCC-3'	RT-PCR
LBP	WT Mutant Common	S 5'-AGAGGTTTCCAGGTCCCCT-3' S 5'-CGTCCTGCAGTTCATTGAG-3' AS 5'-CCTCAGTTTTTCCTTGTCTGC-3'	Genotyping
LBP		S 5'-CCCTTCACTCCCCAGATATACA-3' AS 5'-GATCAGTCTCACTTGTGCCTTG-3'	RT-PCR
LBP		S 5'-GGACCTCTGCTCTTACATTGC-3' AS 5'-GCTGGTGGCTCTGCAGAGAGA-3'	RT-PCR (KO)
MD-2 (<i>Ly96</i>)		S 5'-GATGGTCTTCTGCGGAGTTTA-3' AS 5'-GAGGAGTTGCAGAACCACTGTT-3'	RT-PCR
MMP3		S 5'-TCCTGATGTTGGTGGCTTCAG-3' AS 5'-TGTCTTGGCAAATCCGGTGTA-3'	RT-PCR
MMP13		S 5'-TGATGGACCTTCTGGTCTTCTGG-3' AS 5'-CATCCACATGGTTGGGAAGTTCT-3'	RT-PCR
TLR2		S 5'-CAAACCTGGAGACTCTGGAAGCA-3' AS 5'-GCACCTACGAGCAAGATCAACA-3'	RT-PCR (mRNA/KO)
TLR2	WT Mutant Common	S 5'-ACGAGCAAGATCAACAGGAGA-3' S 5'-GGGCCAGCTCATTCTCCAC-3' AS 5'-CTTCTGAATTTGTCCAGTACA-3'	Genotyping
TLR4		S 5'-TGAGGACTGGGTGAGAAATGAG-3' AS 5'-ATACAATTCCACCTGCTGCCTC-3'	RT-PCR (mRNA/KO)
TLR4	WT Mutant	S 5'-ATATGCATGATCAACACCACAG-3' AS 5'-TTTCCATTGCTGCCCTATAG-3' S 5'-GCAAGTTTCTATATGCATTCTC-3' AS 5'-CCTCCATTTCCAATAGGTAG-3'	Genotyping

SAA1 (human)	S 5'-TGGAGAGCCTACTCTGACATGA-3' AS 5'-CACCATGGCCAAAGAATCTC-3	RT-PCR
SAA1	S 5'-ATGAAGGAAGCTAACTGGAAAACTC-3' AS 5'-TCCTCCTCAAGCAGTTACTACTGCAA-3'	RT-PCR
SAA2	S 5'-ATGAAGGAAGCTGGCTGGAAAGATGG-3' AS 5'-TCCTCCTCAAGCAGTTACTACTGCTC-3'	RT-PCR
SAA3	S 5'-GCCACCATGAAGCCTTCCATTGCCATCATT-3' AS 5'-TCAGTATCTTTAGGCAGGCCAGCAG-3'	RT-PCR
SAA4	S 5'-GTTCTTTGTTCTGGGAGTCAG-3' AS 5'-GTTTAGGAGGCCCTGGAAGTAT-3'	RT-PCR
SOX9	S 5'-ATGCTATCTTCAAGGCGCTG-3' AS 5'-GACGTCGAAGGTCTCAATGT-3'	RT-PCR
ZIP8	S 5'-GAACAATTGCCTGGATGATCACGC-3' AS 5'-AAGCCGGTTAACATCCCTGCATTC-3'	RT-PCR

S, sense; AS, antisense

Supplementary Table 2. mRNA levels of TLR signaling components in chondrocytes treated with IL-1 β (1 ng/ml, 36 h) or infected with 800 MOI (36 h) of Ad-HIF-2 α or Ad-ZIP8. Mean \pm s.e.m. (n = 4).

Group	Gene symbol	Definition	Fold change			
			IL-1 β	Ad-HIF-2 α	Ad-ZIP8	
TLRs	<i>Tlr1</i>	Toll-like receptor 1	0.93 \pm 0.15	0.92 \pm 0.14	0.84 \pm 0.12	
	<i>Tlr2</i>	Toll-like receptor 2	2.83 \pm 0.49	2.62 \pm 0.48	2.83 \pm 0.47	
	<i>Tlr3</i>	Toll-like receptor 3	2.20 \pm 0.72	1.96 \pm 0.56	1.75 \pm 0.48	
	<i>Tlr4</i>	Toll-like receptor 4	1.65 \pm 0.17	2.00 \pm 0.20	2.03 \pm 0.26	
	<i>Tlr5</i>	Toll-like receptor 5	1.66 \pm 0.21	0.93 \pm 0.14	0.76 \pm 0.07	
	<i>Tlr6</i>	Toll-like receptor 6	1.15 \pm 0.23	0.87 \pm 0.14	0.80 \pm 0.14	
	<i>Tlr7</i>	Toll-like receptor 7	1.03 \pm 0.09	0.94 \pm 0.24	0.91 \pm 0.22	
	<i>Tlr8</i>	Toll-like receptor 8	1.02 \pm 0.12	1.11 \pm 0.19	1.13 \pm 0.22	
	<i>Tlr9</i>	Toll-like receptor 9	0.95 \pm 0.18	1.03 \pm 0.12	0.85 \pm 0.07	
	<i>Tlr10</i>	Toll-like receptor 10	-	-	-	
	<i>Tlr11</i>	Toll-like receptor 11	0.97 \pm 0.12	1.04 \pm 0.31	0.90 \pm 0.14	
	<i>Tlr12</i>	Toll-like receptor 12	1.10 \pm 0.15	0.94 \pm 0.07	0.91 \pm 0.19	
	<i>Tlr13</i>	Toll-like receptor 13	0.85 \pm 0.15	1.13 \pm 0.20	1.18 \pm 0.23	
Accessory molecules	<i>Lbp</i>	LPS binding protein	5.27 \pm 2.14	23.06 \pm 4.36	22.3 \pm 4.37	
	<i>Cd14</i>	CD14 antigen	3.19 \pm 1.12	3.65 \pm 0.76	2.69 \pm 0.63	
	<i>Ly96</i>	Lymphocyte antigen 96	2.35 \pm 0.42	2.71 \pm 0.73	1.97 \pm 0.48	
	<i>Cd36</i>	CD36 antigen	1.01 \pm 0.15	1.01 \pm 0.15	0.96 \pm 0.14	
	<i>Cd44</i>	CD44 antigen	2.59 \pm 0.19	2.62 \pm 0.29	2.36 \pm 0.36	
	<i>Hmgb1</i>	High mobility group box 1	0.91 \pm 0.09	0.99 \pm 0.05	1.06 \pm 0.06	
	<i>Camp</i>	Cathelicidin antimicrobial peptide	1.62 \pm 0.24	1.72 \pm 0.50	1.55 \pm 0.36	
	<i>Tril</i>	TLR4 interactor leucine-rich repeats	0.96 \pm 0.27	0.86 \pm 0.19	0.88 \pm 0.18	
	<i>Grn</i>	Granulin	1.11 \pm 0.10	1.18 \pm 0.15	1.16 \pm 0.15	
	<i>Hsp90b1</i>	Heat shock protein 90	1.04 \pm 0.06	1.01 \pm 0.03	0.98 \pm 0.04	
	<i>Cnpy3</i>	Canopy 3 homolog	1.00 \pm 0.04	1.00 \pm 0.06	1.08 \pm 0.08	
<i>Unc93b1</i>	Unc-93 homolog B1	1.65 \pm 0.26	1.10 \pm 0.16	1.07 \pm 0.15		
TLR agonists: fatty acids, lipoproteins	Amyloid β	<i>Apbb1ip</i>	Amyloid beta (A4) precursor protein binding, family A, member 1	0.80 \pm 0.08	1.13 \pm 0.11	0.90 \pm 0.09
		<i>Apba1</i>	Amyloid beta (A4) precursor protein binding, family A, member 2	0.97 \pm 0.08	0.99 \pm 0.13	0.98 \pm 0.12
		<i>Apbb3</i>	Amyloid beta (A4) precursor protein binding, family B, member 1	1.06 \pm 0.08	0.84 \pm 0.12	0.81 \pm 0.08
		<i>Apba2</i>	Amyloid beta (A4) precursor protein binding, family B, member 2	1.03 \pm 0.04	0.93 \pm 0.09	0.96 \pm 0.06
		<i>Aplp2</i>	Amyloid beta (A4) precursor protein binding, family B, member 3	1.02 \pm 0.21	1.01 \pm 0.07	0.93 \pm 0.12
		<i>Aplp1</i>	Amyloid beta (A4) precursor protein	1.07 \pm 0.03	0.85 \pm 0.05	0.93 \pm 0.05
		<i>Apbb2</i>	Amyloid beta (A4) precursor-like protein 1	1.21 \pm 0.07	0.93 \pm 0.06	0.92 \pm 0.06
		<i>Appbp2</i>	Amyloid beta (A4) precursor-like protein 2	1.16 \pm 0.04	0.96 \pm 0.06	1.03 \pm 0.09
		<i>App</i>	Amyloid beta (A4) precursor protein binding, family B	0.82 \pm 0.13	0.93 \pm 0.17	1.00 \pm 0.17
		<i>Iapp</i>	Islet amyloid polypeptide	0.94 \pm 0.07	0.99 \pm 0.14	1.03 \pm 0.10

Oxidized LDL	<i>Apob</i>	Apolipoprotein B	1.06 ± 0.16	1.05 ± 0.14	1.17 ± 0.26	
	<i>Apobec1</i>	Apolipoprotein B mRNA editing enzyme, catalytic polypeptide 1	0.90 ± 0.17	0.99 ± 0.23	1.04 ± 0.19	
	<i>Apobec2</i>	Apolipoprotein B mRNA editing enzyme, catalytic polypeptide 2	0.93 ± 0.15	0.83 ± 0.22	0.91 ± 0.23	
	<i>Apobec3</i>	Apolipoprotein B mRNA editing enzyme, catalytic polypeptide 3	1.73 ± 0.19	1.35 ± 0.22	1.23 ± 0.22	
	<i>Loxhd1</i>	Lipoxygenase homology domains 1	1.01 ± 0.11	1.02 ± 0.18	1.01 ± 0.18	
	<i>Mpo</i>	Myeloperoxidase	0.79 ± 0.16	1.15 ± 0.30	1.17 ± 0.24	
Serum amyloid A	<i>Saa1</i>	Serum amyloid A 1	49.92 ± 9.69	61.55 ± 22.96	29.49 ± 12.18	
	<i>Saa2</i>	Serum amyloid A 2	60.63 ± 3.66	104.4 ± 45.45	60.92 ± 27.31	
	<i>Saa3</i>	Serum amyloid A 3	44.84 ± 18.76	21.49 ± 7.88	20.66 ± 7.59	
	<i>Saa4</i>	Serum amyloid A 4	19.82 ± 3.63	14.71 ± 3.71	13.05 ± 2.90	
	<i>Apcs</i>	Serum amyloid P-component	0.95 ± 0.05	0.95 ± 0.14	0.86 ± 0.13	
TLR agonists: peptides, proteins	Fibrinogen	<i>Fga</i>	Fibrinogen alpha chain	0.98 ± 0.17	1.00 ± 0.09	1.01 ± 0.13
		<i>Fgb</i>	Fibrinogen beta chain	0.98 ± 0.14	0.92 ± 0.11	1.01 ± 0.14
		<i>Fgg</i>	Fibrinogen gamma chain	1.03 ± 0.08	1.02 ± 0.13	0.91 ± 0.17
		<i>Fibcd1</i>	Fibrinogen C domain containing 1	0.92 ± 0.15	0.70 ± 0.13	0.84 ± 0.09
		<i>Fgl1</i>	Fibrinogen-like protein 1	1.50 ± 0.17	1.33 ± 0.16	1.45 ± 0.35
		<i>Fsbp</i>	fibrinogen silencer binding protein	1.00 ± 0.07	1.14 ± 0.11	1.15 ± 0.09
	S100	<i>S100a8</i>	S100 calcium binding protein A8	1.35 ± 0.50	0.94 ± 0.23	1.02 ± 0.16
<i>S100a9</i>		S100 calcium binding protein A9	0.92 ± 0.24	1.09 ± 0.25	1.09 ± 0.21	
TLR down-stream signaling molecules	TIRAP	<i>Tirap</i>	Toll-interleukin 1 receptor (TIR) domain-containing adaptor protein	1.01 ± 0.07	0.99 ± 0.11	0.96 ± 0.1
	MyD88	<i>Myd88</i>	Myeloid differentiation primary response gene 88	1.11 ± 0.23	1.61 ± 0.21	1.62 ± 0.25
	IRAK	<i>Irak1</i>	IL-1 receptor-associated kinase 1	1.26 ± 0.04	1.01 ± 0.06	1.02 ± 0.07
		<i>Irak2</i>	IL-1 receptor-associated kinase 2	1.68 ± 0.16	1.71 ± 0.33	1.30 ± 0.18
		<i>Irak3</i>	IL-1 receptor-associated kinase 3	6.45 ± 0.99	5.73 ± 0.96	4.04 ± 0.69
		<i>Irak4</i>	IL-1 receptor-associated kinase 4	1.14 ± 0.09	1.04 ± 0.11	1.02 ± 0.09
		<i>Irak1bp1</i>	IL-1 receptor-associated kinase 1 binding protein 1	1.46 ± 0.16	1.27 ± 0.12	1.23 ± 0.13
	TRAF family	<i>Traf1</i>	TNF receptor-associated factor 1	1.45 ± 0.18	1.34 ± 0.34	1.12 ± 0.26
		<i>Traf2</i>	TNF receptor-associated factor 2	1.51 ± 0.07	1.59 ± 0.13	1.56 ± 0.15
		<i>Traf3</i>	TNF receptor-associated factor 3	1.03 ± 0.11	1.09 ± 0.06	1.04 ± 0.13
		<i>Traf4</i>	TNF receptor-associated factor 4	1.12 ± 0.07	0.79 ± 0.11	0.87 ± 0.10
		<i>Traf5</i>	TNF receptor-associated factor 5	0.84 ± 0.11	1.01 ± 0.10	0.86 ± 0.05
		<i>Traf6</i>	TNF receptor-associated factor 6	1.10 ± 0.13	0.94 ± 0.12	0.88 ± 0.13
<i>Traf7</i>		TNF receptor-associated factor 7	1.21 ± 0.05	1.06 ± 0.06	1.00 ± 0.06	
<i>Trafd1</i>		TRAF Zn finger domain containing 1	1.06 ± 0.08	0.85 ± 0.06	0.93 ± 0.06	
<i>Traf3ip1</i>		TRAF3 interacting protein 1	1.01 ± 0.06	1.11 ± 0.12	1.03 ± 0.08	
<i>Traf3ip2</i>	TRAF3 interacting protein 2	2.83 ± 0.25	2.76 ± 0.33	2.02 ± 0.24		
TAK1	<i>Map3k7</i>	Mitogen-activated protein kinase kinase kinase 7	1.09 ± 0.11	1.06 ± 0.03	1.03 ± 0.03	
TAB	<i>Tab1</i>	TGF-beta activated kinase 1/MAP3K7 binding protein 1	0.74 ± 0.11	0.87 ± 0.12	0.94 ± 0.09	

	<i>Tab2</i>	TGF-beta activated kinase 1/MAP3K7 binding protein 2	0.78 ± 0.11	0.76 ± 0.06	0.81 ± 0.05
	<i>Tab3</i>	TGF-beta activated kinase 1/MAP3K7 binding protein 3	1.06 ± 0.03	1.01 ± 0.11	0.90 ± 0.08
TOLLIP	<i>Tollip</i>	Toll interacting protein	0.94 ± 0.11	1.05 ± 0.15	1.00 ± 0.18
RIP	<i>Ripk1</i>	Receptor-interacting serine-threonine kinase 1	1.10 ± 0.10	1.06 ± 0.05	1.04 ± 0.06
	<i>Ripk2</i>	Receptor-interacting serine-threonine kinase 2	1.99 ± 0.28	1.39 ± 0.12	1.47 ± 0.15
	<i>Ripk3</i>	Receptor-interacting serine-threonine kinase 3	1.76 ± 0.13	2.15 ± 0.67	1.76 ± 0.51
	<i>Ripk4</i>	Receptor-interacting serine-threonine kinase 4	1.00 ± 0.14	1.00 ± 0.13	1.02 ± 0.11
PI3K	<i>Pik3ca</i>	PI3-kinase, catalytic, alpha polypeptide	1.01 ± 0.07	1.05 ± 0.06	0.97 ± 0.04
	<i>Pik3cb</i>	PI3-kinase, catalytic, beta polypeptide	0.67 ± 0.08	0.73 ± 0.08	0.80 ± 0.07
	<i>Pik3r2</i>	PI3-kinase, regulatory subunit, polypeptide 2 (p85 beta)	0.94 ± 0.08	0.95 ± 0.06	0.96 ± 0.06
	<i>Pik3r1</i>	PI3-kinase, regulatory subunit, polypeptide 1 (p85 alpha)	2.24 ± 0.30	1.44 ± 0.11	1.52 ± 0.16
	<i>Pik3c2a</i>	PI3-kinase, C2 domain containing, alpha polypeptide	1.25 ± 0.27	1.15 ± 0.12	1.04 ± 0.12
	<i>Pik3c2g</i>	PI3-kinase, C2 domain containing, gamma polypeptide	0.99 ± 0.24	0.93 ± 0.16	0.94 ± 0.09
	<i>Smg1</i>	SMG1 homolog, PI3-kinase-related kinase	1.12 ± 0.07	0.96 ± 0.17	0.96 ± 0.15
IKKs	<i>Chuk</i>	Conserved helix-loop-helix ubiquitous kinase	0.87 ± 0.12	0.78 ± 0.07	0.81 ± 0.11
	<i>Ikkbb</i>	Inhibitor of kappaB kinase beta	1.16 ± 0.16	1.03 ± 0.11	0.97 ± 0.12
	<i>Ikkbg</i>	Inhibitor of kappaB kinase gamma	1.07 ± 0.05	1.01 ± 0.13	1.02 ± 0.07
	<i>Ikkbe</i>	Inhibitor of kappaB kinase epsilon	2.03 ± 0.19	1.66 ± 0.31	1.60 ± 0.36
NF-κB	<i>Rela</i>	v-rel reticuloendotheliosis viral oncogene homolog A (avian)	1.39 ± 0.14	1.17 ± 0.11	1.04 ± 0.10
	<i>Relb</i>	Avian reticuloendotheliosis viral (v-rel) oncogene related B	1.59 ± 0.18	1.43 ± 0.11	1.45 ± 0.13
	<i>Rel</i>	Reticuloendotheliosis oncogene	1.88 ± 0.43	1.51 ± 0.15	1.30 ± 0.10
	<i>Nfkb1</i>	NFκ light polypeptide gene enhancer in B cells 1, p105	1.66 ± 0.09	2.23 ± 0.16	1.97 ± 0.15
	<i>Nfkb2</i>	NFκ light polypeptide gene enhancer in B cells 2, p49/p100	1.51 ± 0.12	1.58 ± 0.08	1.50 ± 0.10
NF-κB inhibitor	<i>Nfkbia</i>	NFκ light polypeptide gene enhancer in B cells inhibitor, alpha	5.10 ± 0.13	2.44 ± 0.33	2.27 ± 0.33
	<i>Nfkbib</i>	NFκ light polypeptide gene enhancer in B cells inhibitor, beta	0.85 ± 0.10	1.01 ± 0.05	1.05 ± 0.08
	<i>Nfkbid</i>	NFκ light polypeptide gene enhancer in B cells inhibitor, delta	1.13 ± 0.11	1.44 ± 0.32	1.47 ± 0.37
	<i>Nfkbiz</i>	NFκ light polypeptide gene enhancer in B cells inhibitor, zeta	9.78 ± 0.88	6.00 ± 0.90	5.17 ± 0.82
	<i>Nfkbil1</i>	NFκ light polypeptide gene enhancer in B cells inhibitor like 1	0.83 ± 0.06	0.77 ± 0.10	0.85 ± 0.07
TRIF	<i>Ticam1</i>	TLR adaptor molecule 1	1.04 ± 0.09	0.98 ± 0.14	0.85 ± 0.10
TRAM	<i>Ticam2</i>	TLR adaptor molecule 2	1.00 ± 0.19	0.96 ± 0.12	1.22 ± 0.20
TBK1	<i>Tbk1</i>	TANK-binding kinase 1	1.33 ± 0.14	1.36 ± 0.13	1.22 ± 0.10
API	<i>Jun</i>	Jun proto-oncogene	2.37 ± 0.22	1.52 ± 0.29	1.10 ± 0.14
	<i>Junb</i>	Jun B proto-oncogene	1.79 ± 0.19	2.36 ± 0.23	2.25 ± 0.22
	<i>Jund</i>	Jun D proto-oncogene	1.07 ± 0.06	0.93 ± 0.10	0.94 ± 0.09
	<i>Fos</i>	FBJ osteosarcoma oncogene	1.40 ± 0.29	2.29 ± 0.59	1.84 ± 0.35
	<i>Fra1</i>	Fos-like antigen 1	8.13 ± 1.24	7.51 ± 2.46	3.36 ± 0.97

<i>Fra2</i>	Fos-like antigen 2	2.78 ± 0.20	2.42 ± 0.23	1.89 ± 0.20
<i>Batf</i>	Basic leucine zipper transcription factor, ATF-like	1.70 ± 0.26	3.03 ± 0.40	2.96 ± 0.41
<i>Atf1</i>	Activating transcription factor 1	0.97 ± 0.18	1.20 ± 0.22	1.20 ± 0.22
<i>Atf3</i>	Activating transcription factor 3	0.61 ± 0.04	0.59 ± 0.08	0.83 ± 0.10
<i>Atf4</i>	Activating transcription factor 4	0.69 ± 0.06	0.80 ± 0.11	0.81 ± 0.06
<i>Atf6</i>	Activating transcription factor 6	0.85 ± 0.10	0.75 ± 0.05	0.82 ± 0.06
<i>Atf6b</i>	Activating transcription factor 6 beta	0.95 ± 0.10	0.99 ± 0.04	1.01 ± 0.03
<i>Atf7</i>	Activating transcription factor 7	1.28 ± 0.03	0.97 ± 0.08	1.00 ± 0.06
<i>Atf7ip</i>	Activating transcription factor 7 interacting protein	1.01 ± 0.08	1.10 ± 0.09	1.03 ± 0.08
<i>Atf7ip2</i>	Activating transcription factor 7 interacting protein 2	0.85 ± 0.16	1.11 ± 0.33	1.13 ± 0.19
<i>Jdp2</i>	Jun dimerization protein 2	0.49 ± 0.02	0.70 ± 0.06	0.78 ± 0.07