

Chemistry–A European Journal

Supporting Information

Soluble Congeners of Prior Insoluble Shape-Persistent Imine Cages

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1 General Remarks

Chemicals and Solvents

All utilised chemicals and solvents were purchased from *abcr*, *Acros Organics*, *Alfa Aesar*, *Carbolution*, *Degussa*, *Fisher Chemical*, *Grüssing*, *Honeywell*, *Merck*, *Sigma Aldrich*, *TCI* or *VWR*. They were used without further purification, if not mentioned otherwise. Dry DCM, THF or toluene were obtained from *Honeywell* and purified by a *Solvent Purification System MB SPS-800*.

Analytical and Purification Methods

Elemental analysis

The elemental analyses were performed on a *Vario EL Element Analyzer* by the microanalytical laboratory of University of Heidelberg.

IR Spectroscopy

Infrared spectroscopical measurements were conducted using a *Bruker Tensor 27 spectrometer* with ZnSe ATR crystal. The used abbreviations for describing the relative intensity of the obtained signals are *vw* (very weak, 0 – 10%), *w* (weak, 10 – 30%), *m* (medium, 30 – 60%), *s* (strong, 60 – 90%) and *vs* (very strong, 90 – 100%).

Mass Spectrometry

Mass spectrometric analyses were performed by the mass spectrometry department of University of Heidelberg under the supervision of Dr Jürgen H. Gross. The utilised devices were *Bruker AutoFlex Speed time-of-flight* spectrometer (MALDI-TOF) and *Bruker ApexQe hybrid 9.4T FT-ICR* spectrometer (ESI). For the MALDI analyses DCTB (*trans*-2-[3-(4-*tert*-Butylphenyl)-2-methylpropenylidene]malononitrile) was used as matrix. High resolution MALDI-TOF spectra were obtained by utilizing an internal polyethylene glycole (PEG) standard.

Melting Point Measurement

All measured melting points were obtained using a *Büchi Melting Point B-545* device.

NMR Spectroscopy

Nuclear magnetic resonance spectra were obtained using a *Bruker Avance III 300* (300 MHz), *Bruker Avance DRX 300* (300 MHz), *Bruker Fourier 300* (300 MHz), *Bruker Avance III 400* (400 MHz), *Bruker Avance III 500* (500 MHz) or *Bruker Avance III 600* (600 MHz) spectrometer. The chemical shifts δ are shown in parts per million (ppm) and the coupling constants J in Hertz (Hz). The spectra were recorded at 298 K, if not mentioned otherwise. The obtained signals of ^1H , ^2H and $^{13}\text{C}\{^1\text{H}\}$ NMR spectra were referred to known solvent signals (CDCl_3 : $^1\text{H}/^2\text{H}$ NMR 7.26 ppm, $^{13}\text{C}\{^1\text{H}\}$ NMR 77.2 ppm; THF- d_8 : ^1H NMR 3.58 ppm, 1.73 ppm, $^{13}\text{C}\{^1\text{H}\}$ NMR 67.6 ppm, 25.3 ppm).^[S1] To obtain a referable solvent signal in ^2H NMR spectroscopy CHCl_3 containing a low amount of CDCl_3 or a THF/THF- d_8 mixture has been used as solvent. Multiplicities were labelled with *s* (singlet), *d* (doublet), *dd* (doublet of doublet), *t* (triplet), *td* (triplet of doublet), *q* (quartet) and *m* (multiplet).

DOSY spectra were recorded at 298 K using THF- d_8 as solvent and referred to its self-diffusion coefficient ($D_{\text{THF}} = 2.33 \cdot 10^{-9} \text{ m}^2 \text{ s}^{-1}$).^[S2] The solvodynamic radii r of cage molecules were calculated using the Stokes-Einstein equation solved for r .

$$r = \frac{k_B T}{6\pi\eta D} \quad (\text{eq.1})$$

k_B is the Boltzmann constant, T the temperature, η the viscosity of THF and D the determined diffusion coefficient of the cage molecule.

Single Crystal X-Ray Diffraction

X-Ray diffraction measurements were proceeded using a *Stoe Stadivari* diffractometer containing a Cu micro source ($\lambda_{\text{K}\alpha} = 1.54178 \text{ \AA}$) and PILATUS detector or *Bruker APEX II Quazar* with Mo micro source ($\lambda_{\text{K}\alpha} = 0.71073 \text{ \AA}$). The structures were solved with *SHELXT-2014*^[S3] and refined against F^2 with a Full-matrix least-squares algorithm using the *SHELXL-2018/3*^[S4] software.

Size Exclusion Chromatography

The synthesised cage compounds were purified using a recycling size exclusion chromatography system from *Shimadzu* containing a control unit (*CBM-20A*), a degassing unit (*DGU-20A*), a pump system (*LC-20AD*), an autosampler (*SIL-20A HT*),

a column oven (CTO-20A), a high pressure switching valve (FCV-20AH2), a photo diode array (PDA) detector (SPC-M20A) and a fraction collector (FRC-10A). As stationary phase one 20 x 50 mm pre-column and three 20 x 300 mm columns containing SDV preparative 100 Å SEC material purchased from Polymer Standards Service. THF (chromatography GPC grade) from Fisher Scientific was used as mobile phase at a flow rate of 5 mL min⁻¹. During the separation the oven temperature was set to 40 °C. The shown chromatograms have been recorded with an absorption wavelength of 254 nm. To remove residual butylated hydroxytoluene (BHT, stabilizer in THF), the product containing fractions were washed with *n*-hexane (3 x 6 mL) and dried *in vacuo*.

Thin Layer and Flash Chromatography

Analytical thin layer chromatography was conducted using TLC Silica gel 60 F₂₅₄ plates purchased from Merck. The resulted spots were analysed using ultraviolet radiation (254 nm and 366 nm). Flash chromatography was performed using Silica gel 60 (40–63 µm / 230–400 mesh ASTM) from Machery-Nagel.

UV/vis Spectroscopy

UV/vis absorption spectra have been recorded using a Jasco V-730 spectrophotometer. Molar extinction coefficients (ϵ) were calculated by absorption measurements of five different concentrated solutions prepared by standard addition method. The illustrated absorption spectra have been obtained by normalizing the measured spectra and multiplying with the molar extinction coefficient of the global absorption maximum.

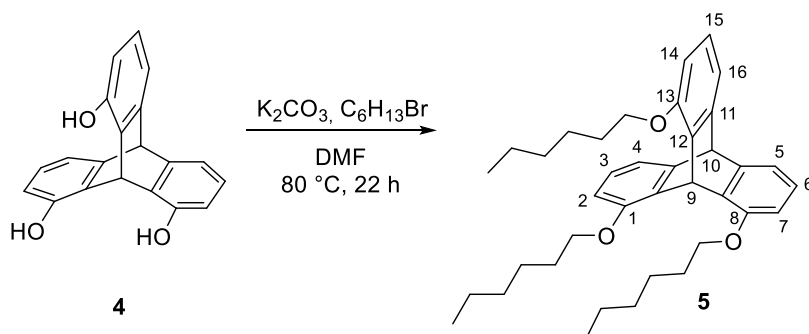
Synthesis of Literature-Known Starting Materials

1,8,13-Trihydroxytriptycene and 2,7,14-Triflormyl-1,8,13-trihydroxytriptycene have been synthesised by following the synthesis route described from Elbert *et al.*^[S5] 3,5-Diformyl-2,6-dihydroxytoluene was synthesised by a Duff formylation of 2,6-dihydroxytoluene.^[S6]

2 Experimental Procedures

2.1 Synthesis of 1,8,13-Trihexyloxy-3,6,15-triaminotriptycene Derivatives

2.1.1 1,8,13-Trihexyloxytriptycene 5

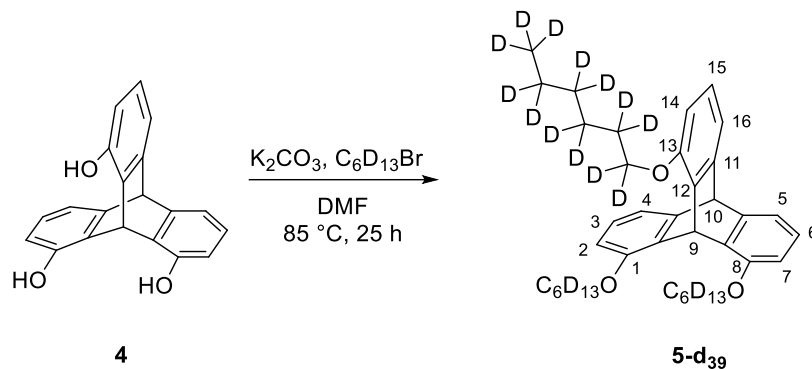


According to a modified procedure from Kumano *et al.*,^[S7] 1,8,13-trihydroxytriptycene **4** (300 mg, 0.99 mmol), potassium carbonate (823 mg, 5.95 mmol) and *N,N*-dimethylformamide (10 mL) were stirred for 1 h at $80\text{ }^\circ\text{C}$. To the emerged brown suspension 1-bromohexane (0.83 mL, 5.95 mmol) was added dropwise and the mixture was stirred for further 21 h at $80\text{ }^\circ\text{C}$. After cooling down to room temperature water (15 mL) was added to the light brown mixture and the precipitate collected by filtration. The solid was washed with water (2 x 30 mL) and methanol (2 x 5 mL) and dried *in vacuo* to give triptycene **5** in 92% yield (506 mg, 0.91 mmol) as off-white solid.

Melting point: $T = 226\text{ }^\circ\text{C}$. **$^1\text{H NMR}$:** (400 MHz, $CDCl_3$) δ (ppm) 7.00 (d, $J = 7.2\text{ Hz}$, 3H, ArH-4,5,16), 6.92 (s, 1H, H9), 6.88 (dd, $J = 8.2, 7.3\text{ Hz}$, 3H, ArH-3,6,15), 6.56 (dd, $J = 8.3, 0.9\text{ Hz}$, 3H, ArH-2,7,14), 5.37 (s, 1H, H10), 3.97 (t, $J = 6.4\text{ Hz}$, 6H, -O-CH₂-), 1.94 – 1.80 (m, 6H, -O-CH₂-CH₂-), 1.66-1.56 (m, 6H, -O-CH₂-CH₂-CH₂-), 1.47-1.33 (m, 12H, -CH₂-CH₂-CH₃), 1.03-0.88 (m, 9H, -CH₃). **$^{13}\text{C}\{^1\text{H}\}\text{ NMR}$:** (100 MHz, $CDCl_3$) δ (ppm) 154.3 (ArC-1,8,13), 148.7 (ArC-4a,10a,11), 134.0 (ArC-8a,9a,12), 125.6 (ArC-3,6,15), 116.6 (ArC-4,5,16), 110.4 (ArC-2,7,14), 69.0 (-O-CH₂-), 54.8 (C-10), 33.7 (C-9), 32.0 (-CH₂-CH₂-CH₃), 29.8 (-O-CH₂-CH₂-), 25.9 (-O-CH₂-CH₂-CH₂-), 22.8 (-CH₂-CH₂-CH₃), 14.3 (-CH₃). **HRMS (MALDI-TOF+, DCTB):** $[M]^+$: m/z calcd. for $C_{38}H_{50}O_3^+$: 554.3760, found: 554.3763. **IR:** (ATR, FT) $\tilde{\nu}$ (cm^{-1}) 3047 (w), 3032 (w), 3015 (w), 2949 (m), 2928 (m), 2866 (m), 1597 (s), 1485 (m), 1468 (s), 1439 (m), 1394 (m), 1377 (w), 1346 (w), 1323 (w), 1277 (vs), 1234 (m), 1198 (m), 1161 (w), 1124 (w), 1097 (s), 1065 (m), 1053 (m), 1024 (m), 991 (w), 949 (w), 916 (m), 858 (w), 824 (vw), 789 (s), 773 (m), 735 (vs), 683 (vw), 640 (w), 604 (w). **Elemental Anal.** Calcd. for $C_{38}H_{50}O_3 \cdot 1/5$

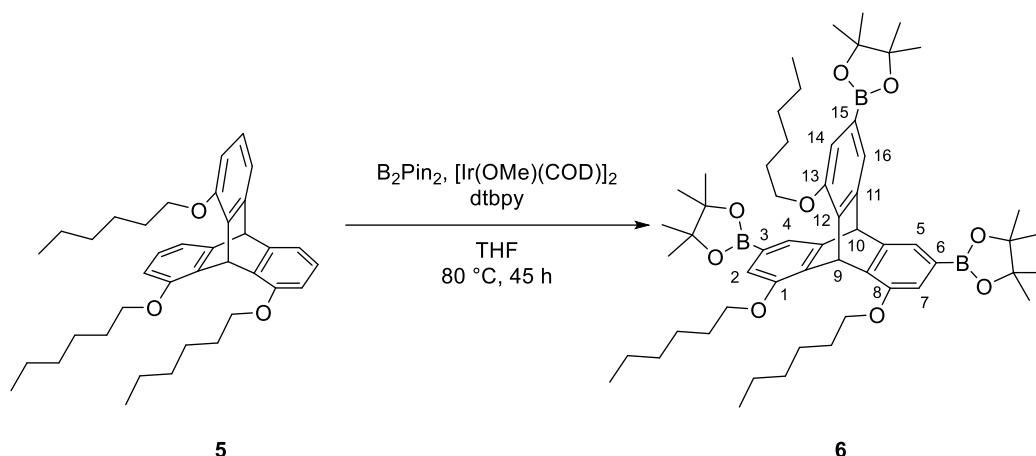
H₂O: C, 81.60; H, 9.10. Found: C, 81.66; H, 8.98. The analytical data is in accordance with the literature data.^[S7]

2.1.2 1,8,13-Trihexyloxytriptycene-d₃₉ 5-d₃₉



According to a modified procedure from Kumano *et al.* for the synthesis of 1,8,13-trihexyloxytriptycene,^[S7] 1,8,13-trihydroxytriptycene **4** (566 mg, 1.87 mmol), potassium carbonate (1.56 g, 11.2 mmol) and dry *N,N*-dimethylformamide (17 mL) were stirred under argon atmosphere for 1 h at 85 °C. 1-Bromohexane-*d*₁₃ (1.6 mL, 11.2 mmol) was added dropwise and the reaction mixture was stirred for further 24 h at 85 °C. After cooling down to room temperature water (40 mL) was added to the light brown suspension and the precipitate collected by filtration. The solid was washed with water (2 x 40 mL) and methanol (4 x 8 mL) and dried *in vacuo* to give triptycene **5-d₃₉** in 89% yield (987 mg, 1.66 mmol) as off-white solid. **Melting point:** *T* = 226 °C. **¹H NMR:** (400 MHz, CDCl₃) δ (ppm) 6.99 (dd, *J* = 7.2, 0.9 Hz, 3H, Ar*H*-4,5,16), 6.92 (s, 1H, *H*₉), 6.87 (dd, *J* = 8.2, 7.3 Hz, 3H, Ar*H*-3,6,15), 6.55 (dd, *J* = 8.2, 0.9 Hz, 3H, Ar*H*-2,7,14), 5.37 (s, 1H, *H*₁₀). **¹³C{¹H} NMR:** (100 MHz, CDCl₃) δ (ppm) 154.3 (Ar-C1,8,13), 148.7 (Ar-C4a,10a,11), 134.0 (Ar-C8a,9a,12), 125.6 (Ar-C3,6,15), 116.5 (Ar-C4,5,16), 110.4 (Ar-C2,7,14), 54.8 (C10), 33.7 (C9). **²H NMR:** (92 MHz, CHCl₃/CDCl₃) δ (ppm) 3.95 (s, 6D, -OCD₂-), 1.81 (s, 6D, -OCD₂-CD₂-), 1.53 (s, 6D, -OCD₂-CD₂-CD₂-), 1.33 (s, 12D, -CD₂-CD₂-CD₃), 0.89 (s, 9D, -CD₃). **HRMS (MALDI-TOF+, DCTB):** [M]⁺: *m/z* calcd. for C₃₈H₁₁D₃₉O₃⁺:593.6202, found: 593.6219. **IR:** (ATR, FT) $\tilde{\nu}$ (cm⁻¹) 2208 (w), 2097 (w), 2070 (vw), 1595 (m), 1479 (s), 1439 (w), 1325 (vw), 1281 (vs), 1248 (w), 1221 (w), 1165 (m), 1105 (s), 1059 (w), 1028 (m), 961 (vw), 922 (w), 856 (w), 824 (vw), 789 (s), 737 (vs), 644 (w). **Elemental Anal.** Calcd. for C₃₈H₁₁D₃₉O₃: C, 76.83; H+D, 8.53 (Value was corrected by detection difference of hydrogen and deuterium. Analyzing a compound containing a known deuterium amount resulted in detection of 50.4% of the apparent deuterium.). Found: C, 76.52; H+D, 8.68.

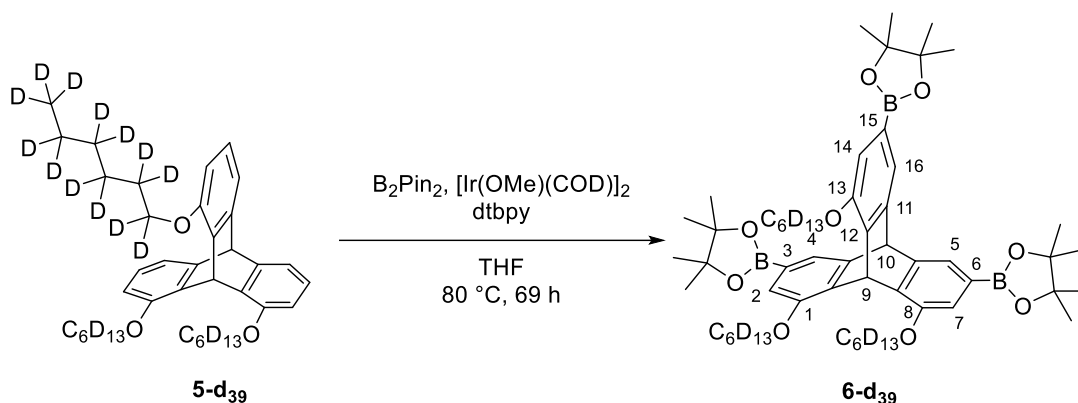
2.1.3 1,8,13-Trihexyloxytriptycene-3,6,15-triyltris(4,4,5,5-tetramethyl-1,3,2-dioxo)borolane **6**



In analogy to a literature procedure^[S8] 1,8,13-trihexyloxytriptycene **5** (3.42 g, 6.16 mmol) and bis(pinacolato)diboron (9.0 g, 35.44 mmol) were suspended in dry tetrahydrofuran (15 mL) under argon and stirred for 30 min. In another tube bis(pinacolato)diboron (440 mg, 1.73 mmol), 4,4'-di-tert-butyl-2,2'-dipyridine (538 mg, 2.0 mmol) and (1,5-cyclooctadiene)(methoxy)iridium(I) dimer (178 mg, 0.27 mmol) were filled under argon atmosphere and dry tetrahydrofuran (15 mL) was added. The mixture was stirred for 20 min at room temperature and turned dark red. The two solutions were combined and stirred for 41 h at 80 °C. After cooling to room temperature, solvent was evaporated *in vacuo* and methanol (50 mL) was added and the brownish suspension ultrasonicated for 10 min, before it was collected by filtration. Recrystallization using an ethanol-chloroform mixture gave boronic ester **6** in 70% yield (3.98 g, 4.26 mmol) as colourless solid. **Melting point:** $T = 314$ °C. **¹H NMR:** (400 MHz, CDCl₃) δ (ppm) 7.42 (s, 3H, Ar-*H*4,5,16), 6.98 (d, $J = 0.8$ Hz, 3H, Ar-*H*2,7,14), 6.94 (s, 1H, *H*9), 5.44 (s, 1H, *H*10), 3.99 (t, $J = 6.5$ Hz, 6H, -OCH₂-), 1.89-1.80 (m, 6H, -OCH₂-CH₂-), 1.62-1.50 (m, 6H, -OCH₂-CH₂-CH₂-), 1.41-1.34 (m, 12H, -CH₂-CH₂-CH₃), 1.28 (s, 36H, -B-O-C(CH₃)₂-), 0.98-0.89 (m, 9H, -CH₂-CH₃). **¹³C{¹H} NMR:** (100 MHz, CDCl₃) δ (ppm) 154.1 (Ar-C1,8,13), 148.0 (Ar-C4a,10a,11), 136.8 (Ar-C8a,9a,12), 126.6 (Ar-C3,6,15), 122.9 (Ar-C4,5,16), 116.5 (Ar-C2,7,14), 83.6 (-B-O-C(CH₃)₂-), 69.1 (-OCH₂-), 54.5 (C10), 34.5 (C9), 32.0 (-CH₂-CH₂-CH₃), 29.9 (-OCH₂-CH₂-), 25.9 (-OCH₂-CH₂-CH₂-), 24.9 (-B-O-C(CH₃)₂-), 22.8 (-CH₂-CH₂-CH₃), 14.3 (-CH₂-CH₃). **HRMS (+ESI):** [M+Na]⁺: m/z calcd. for C₅₆H₈₃B₃O₉Na⁺: 955.6208, found: 955.6217. **IR:** (ATR, FT) $\tilde{\nu}$ (cm⁻¹) 3082 (vw), 3065 (vw), 3045 (vw), 2972 (w), 2930 (w), 2864 (w), 1589 (w), 1574 (w), 1483 (m), 1412 (m), 1356 (vs), 1331 (m), 1298 (m), 1248 (s), 1215

(w), 1140 (s), 1107 (m), 1090 (m), 1001 (w), 964 (m), 937 (w), 916 (m), 856 (m), 831 (m), 768 (m), 714 (w), 696 (m), 667 (m), 648 (w), 609 (vw). **Elemental Anal.** Calcd. for $C_{56}H_{83}B_3O_9$: C, 72.11; H, 8.97. Found: C, 72.11; H, 9.00.

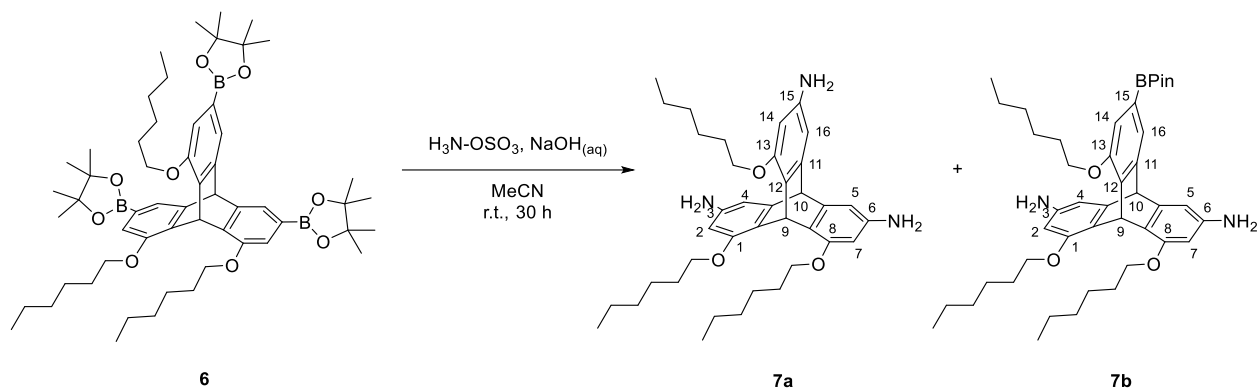
2.1.4 1,8,13-Trihexyloxytriptycene-3,6,15-triyltris(4,4,5,5-tetramethyl-1,3,2-dioxaborolane- d_{39} 6- d_{39})



In analogy to a literature procedure^[S8] 1,8,13-trihexyloxytriptycene- d_{39} **5- d_{39}** (901 mg, 1.52 mmol) and bis(pinacolato)diboron (1.62 g, 6.32 mmol) under argon atmosphere were suspended in dry tetrahydrofuran (3.6 mL) and stirred for 25 min. In another Schlenk tube bis(pinacolato)diboron (117 mg, 0.46 mmol), 4,4'-di-tert-butyl-2,2'-dipyridine (41 mg, 0.15 mmol) and (1,5-cyclooctadiene)(methoxy)iridium(I) dimer (51 mg, 0.08 mmol) were filled under argon atmosphere and dry tetrahydrofuran (3.6 mL) was added. The reaction mixture was stirred for 25 min at room temperature. The mixtures were combined and stirred for 69 h at 80 °C. The dark red reaction mixture was allowed to cool to room temperature, solvent was evaporated *in vacuo* and methanol (7 mL) was added. The resulting brown suspension was ultrasonicated for 10 min, before it was collected by filtration. Recrystallization from ethanol-chloroform gave boronic ester **6- d_{39}** in 70% yield (1.03 g, 1.05 mmol) as colourless crystals. **Melting point:** $T = 315$ °C. **1H NMR:** (700 MHz, $CDCl_3$) δ (ppm) 7.41 (s, 3H, Ar- $H_{4,5,16}$), 6.97 (s, 3H, Ar- $H_{2,7,14}$), 6.94 (s, 1H, H_9), 5.44 (s, 1H, H_{10}), 1.27 (s, 36H, -B-O-C(CH_3) $_2$ -). **$^{13}C\{^1H\}$ NMR:** (176 MHz, $CDCl_3$) δ (ppm) 154.0 (Ar- $C_{1,8,13}$), 148.0 (Ar- $C_{4a,10a,11}$), 136.7 (Ar- $C_{8a,9a,12}$), 126.5 (Ar- $C_{3,6,15}$), 122.8 (Ar- $C_{4,5,16}$), 116.4 (Ar- $C_{2,7,14}$), 83.6 (-B-O-C(CH_3) $_2$ -), 54.4 (C_{10}), 34.4 (C_9), 24.9 (-B-O-C(CH_3) $_2$ -). **2H NMR:** (92 MHz, $CHCl_3/CDCl_3$) δ (ppm) 3.97 (s, 6D, - OCd_2 -), 1.79 (s, 6D, - OCD_2-CD_2 -), 1.51 (s, 6D, - $OCD_2-CD_2-CD_2$ -), 1.31 (s, 12D, - $CD_2-CD_2-CD_3$), 0.88 (s, 9D, - CD_3). **HRMS (MALDI-TOF+, DCTB):** $[M]^+$: m/z calcd. for $C_{56}H_{44}D_{39}B_3O_9^+$: 971,8759, found:

971,8764. **IR:** (ATR, FT) $\tilde{\nu}$ (cm⁻¹) 2976 (w), 2932 (vw), 2208 (w), 2097 (vw), 1587 (w), 1479 (w), 1412 (w), 1362 (vs), 1333 (w), 1296 (m), 1254 (m), 1215 (w), 1140 (s), 1113 (m), 1082 (m), 1040 (w), 1005 (vw), 966 (m), 926 (w), 910 (vw), 856 (m), 831 (w), 766 (w), 716 (w), 696 (m), 667 (w), 646 (w), 617 (vw). **Elemental Anal.** Calcd. for C₅₆H₄₄D₃₉B₃O₉: C, 69.20; H+D, 8.64 (Value was corrected by detection difference of hydrogen and deuterium. Analyzing a compound containing a known deuterium amount resulted in detection of 50.4% of the apparent deuterium.). Found: C, 68.93; H+D, 8.72.

2.1.5 3,6,15-Triamino-1,8,13-trihexyloxytryptycene **7a** and 3,6-Diamino-1,8,13-trihexyloxytryptycene-15-yl(4,4,5,5-tetramethyl-1,3,2-dioxo)borolane **7b**



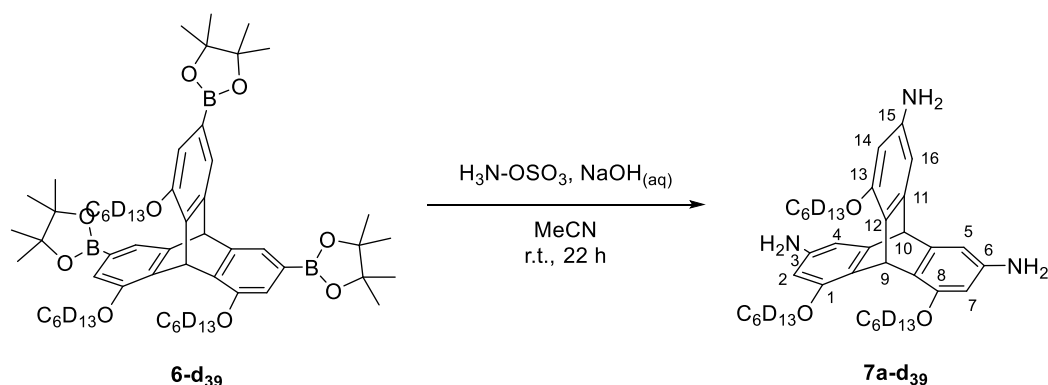
In analogy to a literature procedure^[S9] trisboronic ester **6** (1.01 g, 1.1 mmol), hydroxylamine-*O*-sulfonic acid (672 mg, 5.9 mmol), acetonitrile (32 mL) and aqueous sodium hydroxide solution (16 mL, 1 mol L⁻¹, 16 mmol) were stirred under argon atmosphere. After 24 h additional amounts of hydroxylamine-*O*-sulfonic acid (336 mg, 2.8 mmol) and aqueous sodium hydroxide solution (8 mL, 1 mol L⁻¹, 8 mmol) were added and stirred for additional 6 h at room temperature. Water (40 mL) was added and the mixture extracted with dichloromethane (3 x 60 mL). The combined organic phase was dried over sodium sulphate and solvent removed by rotary evaporation. Purification by silica column chromatography (dichloromethane/methanol 40:1) gave after drying in vacuum triamine **7a** in 62% yield (398 mg, 0.66 mmol) as off-white solid and **7b** in 9% yield (67 mg, 0.09 mmol) as dark brown solid.

Compound **7a**: **Melting point:** $T = 253$ °C. **Retention factor:** $R_f = 0.29$ (in dichloromethane/methanol 20:1 v/v). **¹H NMR:** (600 MHz, CDCl₃) δ (ppm) 6.44 (s, 1H, *H*₉), 6.37 (d, $J = 1.9$ Hz, 3H, Ar-*H*_{4,5,16}), 5.90 (s, $J = 1.9$ Hz, 3H, Ar-*H*_{2,7,14}), 4.93 (s, 1H, *H*₁₀), 3.88 (t, $J = 6.5$ Hz, 6H, -OCH₂-), 3.23 (br. s, 6H, -NH₂), 1.85 – 1.78 (m, 6H,

-OCH₂-CH₂-), 1.59 – 1.51 (m, 6H, -OCH₂-CH₂-CH₂-), 1.41 – 1.34 (m, 12H, -CH₂-CH₂-CH₃), 0.98 – 0.91 (m, 9H, -CH₃). **¹³C{¹H} NMR:** (150 MHz, CDCl₃) δ (ppm) 154.0 (Ar-C1,8,13), 149.2 (Ar-C4a,10a,11), 144.1 (Ar-C3,6,15), 126.0 (Ar-C8a,9a,12), 104.7 (Ar-C4,5,16), 97.9 (Ar-C2,7,14), 69.0 (-OCH₂-), 55.0 (C10), 32.2 (C9), 31.9 (-CH₂-CH₂-CH₃), 29.8 (-OCH₂-CH₂-), 25.9 (-OCH₂-CH₂-CH₂-), 22.8 (-CH₂-CH₂-CH₃), 14.3 (-CH₃). **HRMS (+ESI):** [M+H]⁺: *m/z* calcd. for C₃₈H₅₄N₃O₃⁺: 600.4160, found: 600.4166. **IR:** (ATR, FT) $\tilde{\nu}$ (cm⁻¹) 3396.49 (vw), 3335 (w), 2955 (m), 2930 (m), 2858 (w), 1603 (vs), 1495 (m), 1456 (m), 1394 (vw), 1375 (vw), 1325 (m), 1242 (w), 1196 (m), 1142 (vs), 1103 (m), 1076 (m), 1034 (w), 993 (vw), 976 (w), 914 (w), 816 (m), 766 (m), 725 (w), 681 (w), 635 (w), 619 (vw). **Elemental Anal.** Calcd. for C₃₈H₅₃N₃O₃ · 13/10 CH₃OH: C, 73.58; H, 9.14; N, 6.55. Found: C, 73.60; H, 9.32; N, 6.33.

Compound **7b**: **Melting point:** *T* = 254 °C. **Retention factor:** *R_f* = 0.5 (in dichloromethane/methanol 20:1 v/v). **¹H NMR:** (500 MHz, CDCl₃) δ (ppm) 7.42 (s, 1H, Ar-*H*16), 7.01 (s, 1H, Ar-*H*14), 6.62 (s, 1H, *H*9), 6.38 (d, *J* = 1.8 Hz, 2H, Ar-*H*4,5), 5.90 (d, *J* = 1.9 Hz, 2H, Ar-*H*2,7), 5.10 (s, 1H, *H*10), 3.99 (t, *J* = 6.5 Hz, 2H, PinB-Ar-OCH₂-), 3.88 (t, *J* = 6.5 Hz, 4H, H₂N-Ar-OCH₂-), 3.54 (br. s, 4H, -NH₂), 1.88 – 1.77 (m, 6H, -OCH₂-CH₂-), 1.60 – 1.51 (m, 6H, -OCH₂-CH₂-CH₂-), 1.42 – 1.34 (m, 12H, -CH₂-CH₂-CH₃), 1.29 (s, 12H, -B-O-C(CH₃)₂), 0.97 – 0.92 (m, 9H, -CH₃). **¹³C{¹H} NMR:** (126 MHz, CDCl₃) δ (ppm) 154.5 (Ar-C1,8), 153.2 (Ar-C13), 149.3 (Ar-C4a,10a), 147.8 (Ar-C11), 144.1 (Ar-C3,6), 138.8 (Ar-C12), 125.0 (Ar-C9a,8a), 122.6 (Ar-C14), 116.7 (Ar-C16), 104.8 (Ar-C4,5), 97.9 (Ar-C2,7), 83.6 (-B-O-C(CH₃)₂), 69.0 (-OCH₃-), 68.9 (-OCH₃-), 54.8 (C10), 32.9 (C9), 32.0 (-CH₂-CH₂-CH₃), 31.9 (-OCH₂-CH₂-), 29.8 (-OCH₂-CH₂-), 29.8 (-OCH₂-CH₂-), 25.9 (-OCH₂-CH₂-CH₂-), 25.9 (-OCH₂-CH₂-CH₂-), 24.9 (-B-O-C(CH₃)₂), 22.8 (-CH₂-CH₂-CH₃), 14.3 (-CH₃). (¹H,¹³C HMBC spectrum shows cross peak, which results from a ²*J* coupling between Ar-*H*16 and Ar-C15, at 7.42 ppm/125.8 ppm (¹H/¹³C). The signal for Ar-C15 is not apparent in ¹³C{¹H} NMR spectrum due to its low intensity resulting from coupling with bound boron nucleus.) **HRMS (+ESI):** [M+H]⁺: *m/z* calcd. for C₄₄H₆₄BN₂O₅⁺: 711.4903, found: 711.4911. **IR:** (ATR, FT) $\tilde{\nu}$ (cm⁻¹) 3464 (vw), 3410 (vw), 3337 (w), 2955 (m), 2930 (m), 2860 (m), 1599 (s), 1493 (m), 1468 (m), 1454 (m), 1402 (m), 1358 (vs), 1337 (s), 1296 (w), 1248 (m), 1196 (m), 1159 (m), 1130 (s), 1103 (s), 1084 (m), 966 (m), 932 (w), 916 (w), 854 (m), 839 (m), 824 (m), 814 (m), 768 (m), 725 (w), 698 (w), 673 (w), 629 (w). **Elemental Anal.** Calcd. for C₄₄H₆₃BN₂O₅: C, 74.35; H, 8.93; N, 3.94. Found: C, 74.11; H, 9.01; N, 3.65.

2.1.6 3,6,15-Triamino-1,8,13-trihexyloxytryptcene-d₃₉ **7a-d₃₉**

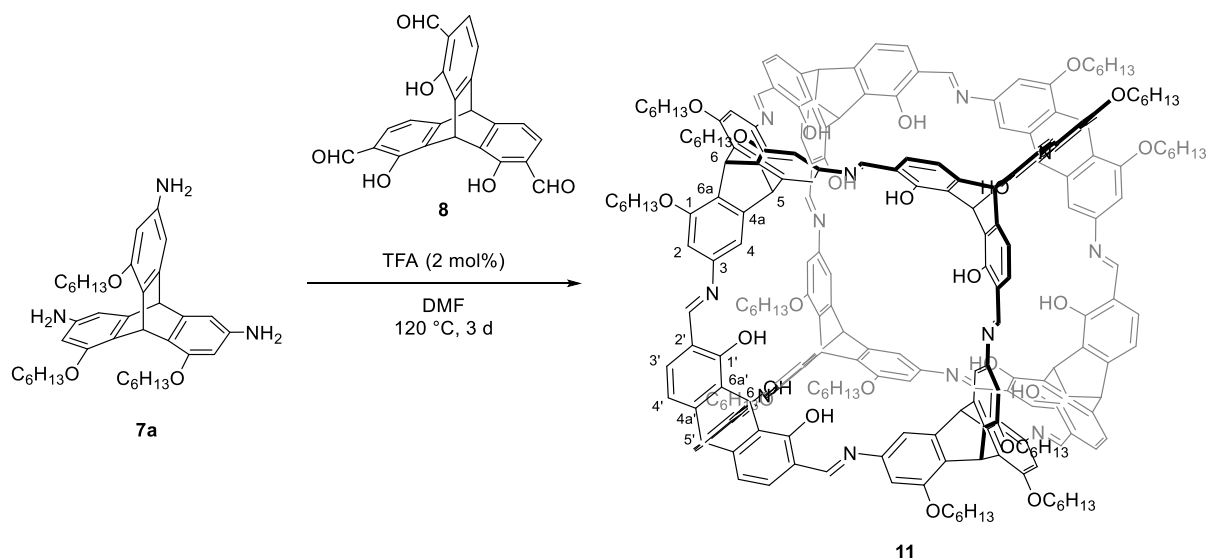


In analogy to a literature procedure^[S9] triboronic ester **6-d₃₉** (850 mg, 0.9 mmol), hydroxylamine-*O*-sulfonic acid (97 %-wt, 566 mg, 4.9 mmol), acetonitrile (27 mL) and aqueous sodium hydroxide solution (13 mL, 1 mol L⁻¹, 13 mmol) were stirred at room temperature under argon atmosphere. After 13 h additional amounts of hydroxylamine-*O*-sulfonic acid (283 mg, 2.4 mmol) and aqueous sodium hydroxide solution (6.5 mL, 1 mol L⁻¹, 6.5 mmol) were added and stirred for further 9 h at room temperature. Water (40 mL) was added and the mixture extracted with dichloromethane (3 x 60 mL). The combined organic phase was dried over sodium sulphate. After removal of the solvent by rotary evaporation the residue was purified by silica column chromatography (dichloromethane/methanol = 40:1) to give **7a-d₃₉** in 66% yield (370 mg, 0.58 mmol) as an off-white solid. **Melting point:** $T = 254$ °C. **Retention factor:** $R_f = 0.29$ (dichloromethane/methanol 20:1). **¹H NMR:** (600 MHz, CDCl₃) δ (ppm) 6.44 (s, 1H, *H*₉), 6.37 (d, $J = 1.9$ Hz, 3H, Ar-*H*_{4,5,16}), 5.89 (d, $J = 2.0$ Hz, 3H, Ar-*H*_{2,7,14}), 4.93 (s, 1H, *H*₁₀), 3.42 (br. s, 6H, -NH₂). **¹³C{¹H} NMR:** (151 MHz, CDCl₃) δ (ppm) 154.1 (Ar-C_{1,8,13}), 149.3 (Ar-C_{4a,10a,11}), 144.2 (Ar-C_{3,6,15}), 126.1 (Ar-C_{8a,9a,12}), 104.6 (Ar-C_{4,5,16}), 97.9 (Ar-C_{2,7,14}), 55.1 (C₁₀), 32.2 (C₉). **²H NMR:** (92 MHz, CHCl₃/CDCl₃) δ (ppm) 3.87 (s, 6D, -OCD₂-), 1.75 (s, 6D, -OCD₂-CD₂-), 1.48 (s, 6D, -OCD₂-CD₂-CD₂-), 1.31 (s, 12D, -CD₂-CD₂-CD₃), 0.88 (s, 9D, -CD₃). **HRMS (MALDI-TOF+, DCTB):** [M]⁺: m/z calcd. for C₃₈H₁₄N₃O₃⁺: 638.6529, found: 638.6540. **IR:** (ATR, FT) $\tilde{\nu}$ (cm⁻¹) 3605 (vw), 3460 (vw), 3412 (vw), 3371 (vw), 3339 (vw), 3217 (vw), 3020 (vw), 2208 (m), 2095 (w), 1601 (vs), 1493 (m), 1456 (m), 1360 (w), 1331 (m), 1244 (w), 1198 (m), 1169 (w), 1140 (s), 1107 (s), 1036 (m), 993 (w), 970 (vw), 916 (vw), 860 (w), 810 (m), 764 (m), 717 (w), 677 (w), 621 (w). **Elemental Anal.** Calcd. for C₃₈H₁₄D₃₉N₃O₃·6/10 H₂O: C, 70.23; H+D, 8.45 (Value was corrected by detection difference of hydrogen and deuterium. Analyzing a compound containing a known deuterium amount resulted in

detection of 50.4% of the apparent deuterium.); N, 6.47. Found: C, 70.27; H+D, 8.58; N 6.29.

2.2 Syntheses of Soluble Organic Cages

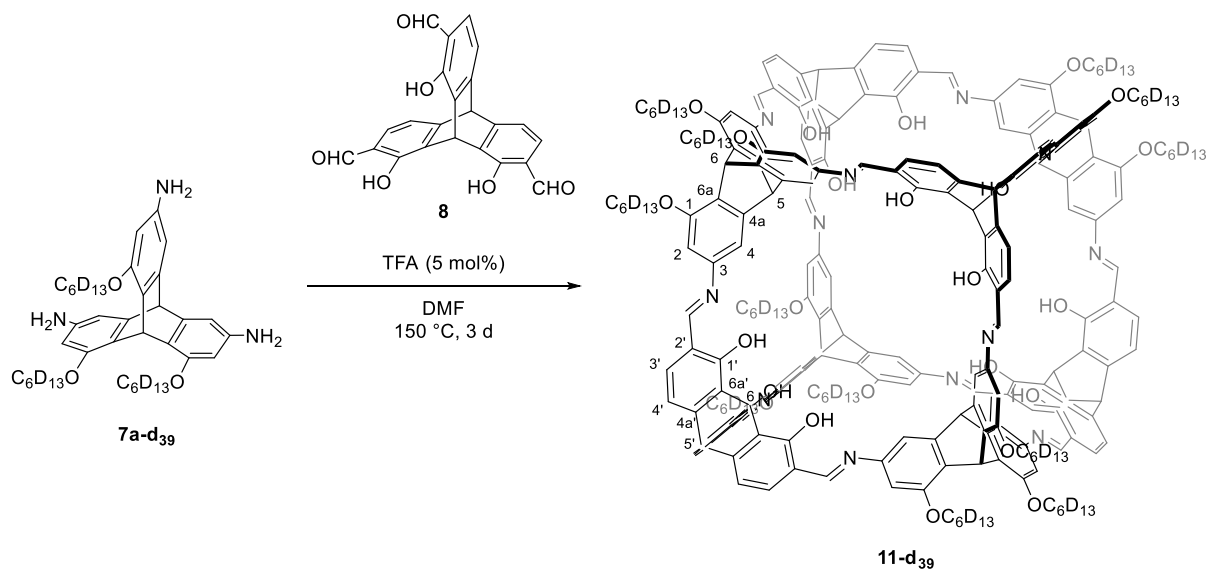
2.2.1 [4+4] Cube 11



In a 25 mL screw-capped vial triaminotriptycene **7a** (20.1 mg, 34 μmol) and triformyrtiptycene **8** (13.0 mg, 34 μmol), DMF (8 mL) and TFA (167 μL , 0.01 mol L⁻¹ in dry DMF, 1.67 μmol , 0.2 eq) were heated for 5 days at 150 °C. The orange suspension was cooled to room temperature molecular sieves (3 Å) added and stirred for additional 20 min. The mixture was filtration through Celite and the solvent was evaporated *in vacuo*. The residue was purified by rGPC (THF) to give **11** as yellow solid in 42% yield (13.0 mg, 3 μmol). **Melting point:** $T = 352$ °C (dec.). **¹H NMR:** (600 MHz, THF-d₈) δ (ppm) 15.21 (s, 12H, -OH), 9.02 (s, 12H, -CHN-), 7.51 (d, $J = 1.6$ Hz, 12H, *H*₄), 7.45 (s, 4H, *H*₆'), 7.07 – 7.02 (m, 24H, *H*₂, *H*₃'), 6.94 (d, $J = 7.7$ Hz, 12H, *H*₄'), 6.89 (s, 4H, *H*₆), 5.86 (s, 4H, *H*₅), 5.51 (s, 5H, *H*₅'), 4.08 (t, $J = 6.5$ Hz, 24H, -OCH₂-), 1.97 – 1.87 (m, 24H, -OCH₂-CH₂-), 1.70 – 1.60 (m, 24H, -OCH₂-CH₂-CH₂-), 1.49 – 1.38 (m, 48H, -CH₂-CH₂-CH₃), 1.01 – 0.91 (m, 36H, -CH₂-CH₂-CH₃). **¹³C{¹H} NMR:** (150 MHz, THF-d₈) δ (ppm) 158.8 (-CHN-), 158.3 (C1'), 157.8(C1'), 156.0 (C1), 152.0 (C4a'), 150.4 (C4a), 143.8 (C3), 134.4 (C6a), 134.3 (C6a'), 134.3 (C6a'), 129.4 (C3'), 119.3 (C4), 119.0 (C2'), 115.1 (C4'), 97.1 (C2), 69.8 (-OCH₂-), 56.6 (C5'), 55.5 (C5), 35.6 (C6), 33.5 (C6'), 32.9 (-CH₂-CH₂-CH₃), 30.8 (-OCH₂-CH₂-), 26.9 (-OCH₂-CH₂-CH₂-), 23.8 (-CH₂-CH₂-CH₃), 14.7 (-CH₂-CH₂-CH₃). **MS (MALDI-TOF+, DCTB):** [M+H]⁺: m/z calcd. for C₂₄₄H₂₄₅N₁₂O₂₄⁺: 3726.832, found: 3726.755. **IR:** (ATR, FT) $\tilde{\nu}$ (cm⁻¹) 2955 (w), 2928

(w) 2858 (w), 1620 (m), 1589 (vs), 1566 (m), 1470 (m), 1437 (m), 1364 (w), 1329 (w), 1281 (w), 1234 (m), 1163 (w), 1119 (s), 1099 (m), 1042 (w) 982 (w), 935 (vw), 872 (w), 827 (w), 808 (w), 781 (m), 762 (m), 731 (m), 662 (w). **UV/vis** (THF) λ_{max} (lg ϵ) 363 nm (5.54). **Elemental Anal.** Calcd. for C₂₄₄H₂₄₄N₁₂O₂₄·4THF: C, 77.74; H, 6.92; N, 4.18. Found: C, 77.69; H, 7.07; N, 3.99.

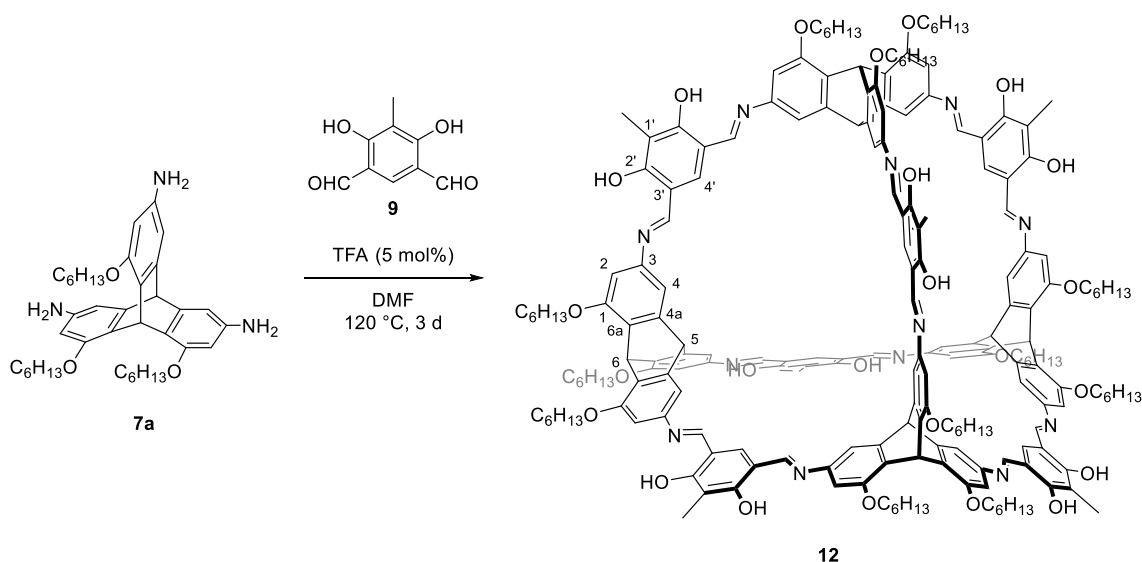
2.2.2 [4+4] Cube 11-d₁₅₆



In a 25 mL-screw-capped vial triaminotriptycene **7a-d₃₉** (21.3 mg, 34 μ mol), triformyltriptycene **8** (13.0 mg, 34 μ mol), dry DMF (8 mL) and TFA (167 μ L, 0.01 mol L⁻¹ in dry DMF, 1.67 μ mol) were stirred for 4 days at 150 °C under argon atmosphere. After cooling to room temperature MeCN (2 mL) was added and solids removed by filtration through a polyamide filter (0.45 μ m). The solvent of the filtrate was evaporated *in vacuo* and the remaining residue purified by rGPC (THF) to give **11-d₁₅₆** as yellow solid in 31% yield (10.0 mg, 3 μ mol). **Melting point:** $T = 354$ °C (dec.). **¹H NMR:** (600 MHz, THF-d₈) δ (ppm) 15.21 (s, 12H, -OH), 9.02 (s, 12H, -CHN-), 7.51 (d, $J = 1.8$ Hz, 12H, H_4), 7.45 (s, 4H, $H_{6'}$), 7.07-7.02 (m, 24H, H_2 , $H_{3'}$), 6.94 (d, $J = 7.7$ Hz, 12H, $H_{4'}$), 6.89 (s, 4H, H_6), 5.85 (s, 4H, H_5), 5.51 (s, 4H, $H_{5'}$). **¹³C{¹H} NMR:** (150 MHz, THF-d₈) δ (ppm) 158.8 (-CHN-), 158.3 (C1'), 156.0 (C1), 152.0 (C4a'), 150.4 (C4a), 143.8 (C3), 134.4 (C6a), 134.3 (C6a'), 129.3 (C3'), 119.3 (C4), 119.0 (C2'), 115.1 (C4'), 97.1 (C2), 69.3 – 68.5 (m, -OCD₂-), 56.6 (C5'), 55.5 (C5), 35.6 (C6), 33.5 (C6'), 32.1-31.1 (m, -CD₂-CD₂-CD₃), 30.0-29.0 (m, -OCD₂-CD₂-), 22.9 – 22.2 (m, -CD₂-CD₂-CD₃), 14.0 – 13.1 (m, -CD₂-CD₂-CD₃). **²H NMR:** (92 MHz, THF/THF-d₈) δ (ppm) 4.09 (s, 24D, -OCD₂-), 1.87 (s, 24D, -OCD₂-CD₂-), 1.59 (s, 24D, -OCD₂-CD₂-CD₂-), 1.37 (s, 48D, -

$CD_2-CD_2-CD_3$), 0.90 (s, 36D, $-CD_2-CD_2-CD_3$). **MS (MALDI-TOF+, DCTB):** $[M]^+$: m/z calcd. for $C_{244}H_{88}D_{156}N_{12}O_{24}^+$: 3882.803, found: 3882.182. **IR:** (ATR, FT) $\tilde{\nu}$ (cm^{-1}) 2210 (w), 2097 (w), 1620 (m), 1587 (vs), 1566 (s), 1474 (m), 1435 (m), 1364 (m), 1329 (w), 1279 (w), 1232 (m), 1163 (w), 1119 (s), 1086 (m), 1040 (m), 989 (w), 968 (w), 935 (w), 870 (m), 827 (m), 806 (m), 779 (m), 760 (m), 729 (m), 663 (w). **UV/vis** (THF) λ_{max} (lg ϵ) 363 nm (5.54). **Elemental Anal.** Calcd. for $C_{244}H_{88}D_{156}N_{12}O_{24} \cdot H_2O$: C, 75.08; H+D, 6.38 (Value was corrected by detection difference of hydrogen and deuterium. Analyzing a compound containing a known deuterium amount resulted in detection of 50.4% of the apparent deuterium.); N, 4.31. Found: C, 74.90; H+D, 6.24; N, 4.04.

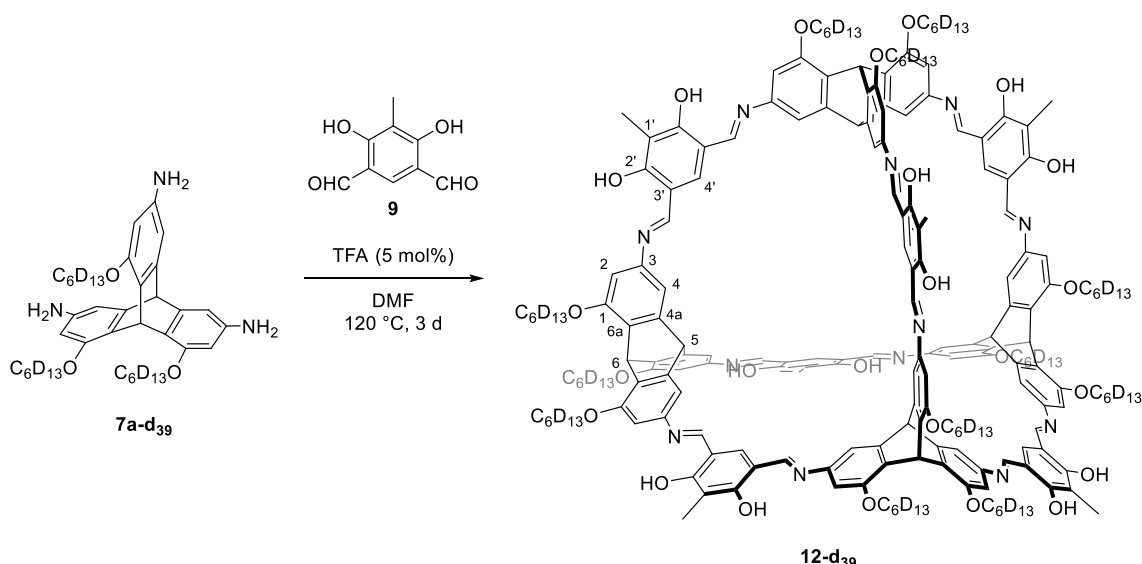
2.2.3 [4+6]-exo Cage 12



A 25 mL screw-capped vial was charged with triaminotryptcene **7a** (20.1 mg, 34 μ mol) and 3,5-diformyl-2,6-dihydroxytoluene **9** (9.0 mg, 50 μ mol) under argon atmosphere. Dry DMF (6 mL) and TFA (167 μ L, 0.01 mol L⁻¹ in dry DMF, 1.67 μ mol) were added and the orange solution was stirred for 3 days at 120 °C. The red solution was allowed to reach room temperature, the solvent was evaporated *in vacuo* and the residue was mixed with DMSO (2 mL) and ultrasonicated for 10 min. After filtration through a polyamide filter (0.45 μ m) the orange residue was washed with DMSO (4 mL) and methanol (6 mL). After further purification by rGPC (THF) exo-cage **12** was obtained as yellow solid in 63% yield (17.3 mg, 5 μ mol). **Melting point:** $T = 338$ °C (dec.). **¹H NMR:** (600 MHz, THF- d_8) δ (ppm) 15.01 (s, 12H, -OH), 9.00 (s, 12H, -CHN-), 7.46 (s, 6H, $H4'$), 7.36 (d, $J = 1.7$ Hz, 12H, $H4$), 6.97 (s, 4H, $H6$), 6.81 (d, $J = 1.7$ Hz, 12H, $H2$), 5.56 (s, 4H, $H5$), 4.11 (t, $J = 6.4$ Hz, 24H, $-OCH_2-$), 2.12 (s, 18H, $-C1'-CH_3$), 2.01-1.92 (m, 24H, $-OCH_2-CH_2-$), 1.76-1.67 (m, 24H, $-OCH_2-CH_2-CH_2-$), 1.52-1.43 (m, 48H, -

$\text{CH}_2\text{-CH}_2\text{-CH}_3$), 1.00 (t, $J = 6.8$ Hz, 36H, $-\text{CH}_2\text{-CH}_2\text{-CH}_3$). $^{13}\text{C}\{^1\text{H}\}$ NMR: (150 MHz, THF- d_8) δ (ppm) 165.7 (C2'), 159.2 (-CHN-), 155.8 (C1), 149.8 (C4a), 145.1 (C3), 135.8 (C4'), 133.0 (C6a), 113.7 (C3'), 112.9 (C1'), 110.1 (C2), 104.1 (C4), 69.5 (-OCH $_2$ -), 57.0 (C5), 34.4 (C6), 33.0 (-CH $_2$ -CH $_2$ -CH $_3$), 30.7 (-OCH $_2$ -CH $_2$ -), 26.9 (-OCH $_2$ -CH $_2$ -CH $_2$ -), 23.8 (-CH $_2$ -CH $_2$ -CH $_3$), 14.8 (-CH $_2$ -CH $_2$ -CH $_3$), 7.6 (C1'-CH $_3$). **MS (MALDI-TOF+, DCTB):** [M+H] $^+$: m/z calcd. for $\text{C}_{206}\text{H}_{237}\text{N}_{12}\text{O}_{24}^+$: 3262.769, found: 3262.884. **IR:** (ATR, FT) $\tilde{\nu}$ (cm $^{-1}$) 2953 (w), 2928 (w), 2858 (w), 1620 (m), 1576 (vs), 1464 (m), 1435 (m), 1393 (m), 1369 (m), 1296 (m), 1225 (w), 1165 (m), 1123 (s), 1099 (m), 1067 (w), 1028 (w), 972 (m), 914 (w), 872 (m), 845 (m), 800 (w), 764 (m), 741 (m); 636 (w), 606 (w), 573 (w). **UV/vis** (THF) λ_{max} (lg ϵ) 316.5 nm (5.31), 377.5 nm (5.50). **Elemental Anal.** Calcd. for $\text{C}_{206}\text{H}_{236}\text{N}_{12}\text{O}_{24} \cdot 10\text{H}_2\text{O}$: C, 71.84; H, 7.49; N 4.88. Found: C, 71.98; H, 7.26; N, 4.52.

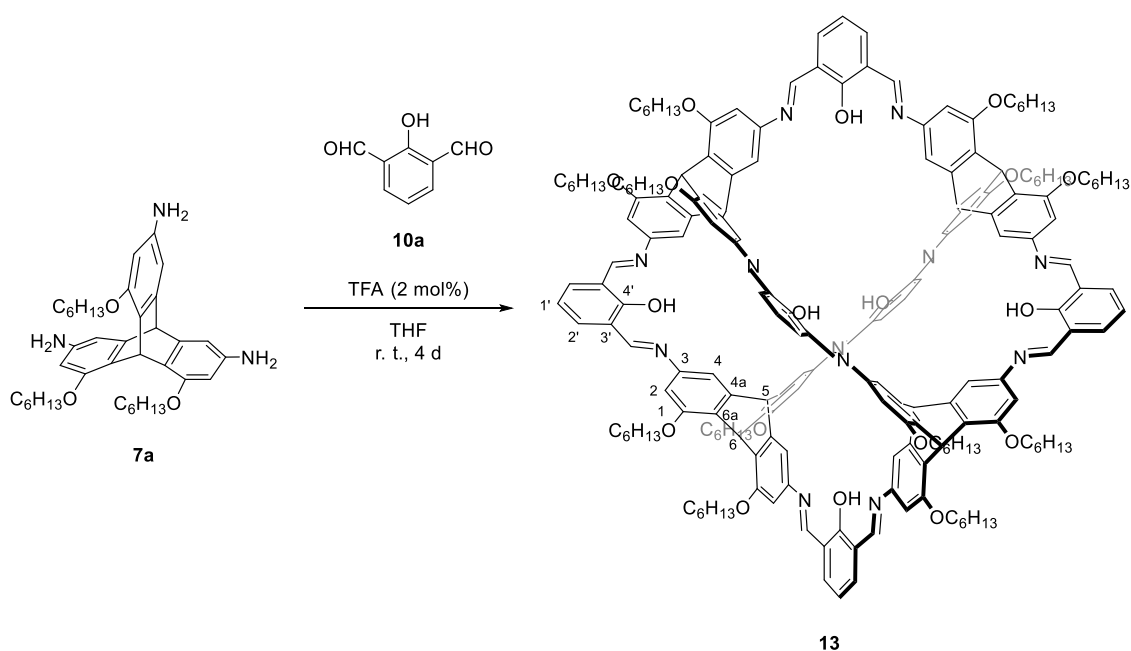
2.2.4 [4+6]-exo Cage 12-d $_{156}$



A 25 mL screw-capped vial was charged with triaminotriptycene **7a-d $_{39}$** (21.3 mg, 33 μmol) and 3,5-diformyl-2,6-dihydroxytoluene **9** (9.1 mg, 51 μmol) under argon atmosphere. Dry DMF (6 mL) and TFA (167 μL , 0.01 mol L $^{-1}$ in dry DMF, 1.67 μmol) were added and the orange solution was stirred for 3 days at 120 °C. The red solution was allowed to reach room temperature, the solvent was evaporated *in vacuo* and the residue was combined with DMSO (4 mL) and ultrasonicated for 10 min. After filtration through a polyamide filter (0.45 μm) the orange residue was washed with DMSO (4 mL) and methanol (6 mL). After further purification by rGPC (THF) cage **12-d $_{156}$** was obtained as yellow solid in 61% yield (17.3 mg, 5 μmol). **Melting point:** $T = 337$ °C

(dec.). **¹H NMR:** (600MHz, THF-d₈) δ (ppm) 15.01 (s, 12H, -OH), 8.99 (s, 12H, -CHN-), 7.45 (s, 6H, H4'), 7.36 (d, *J* = 1.7 Hz, 12H, H4), 6.97 (s, 4H, H6), 6.80 (d, *J* = 1.7 Hz, 12H, H2), 5.55 (s, 4H, H5), 2.12 (s, 18H, -C1'-CH₃). **¹³C{¹H} NMR:** (150 MHz, THF-d₈) δ (ppm) 165.7 (C2'), 159.1 (-CHN-), 155.8 (C1), 149.8 (C4a), 145.1 (C3), 135.8 (C4'), 133.0 (C6a), 113.7 (C3'), 112.9 (C1'), 110.1 (C2), 104.0 (C4), 69.1 – 68.3 (m, -OCD₂-), 57.0 (C5), 34.4 (C6), 32.131.1 (m, -CD₂-CD₂-CD₃), 30.1 – 29.1 (m, -OCD₂-CD₂-), 23.0-22.0 (m, -CD₂-CD₂-CD₃), 14.0-13.2 (m, -CD₂-CD₂-CD₃), 7.6 (C1'-CH₃). **²H NMR:** (92 MHz, THF/THF-d₈) δ (ppm) 4.12 (s, 24D, -OCD₂-), 1.91 (s, 24D, -OCD₂-CD₂-), 1.63 (s, 24D, -OCD₂-CD₂-CD₂-), 1.40 (s, 48H, -CD₂-CD₂-CD₃), 0.93 (s, 36H, -CD₂-CD₂-CD₃). **MS (MALDI-TOF+, DCTB):** [M]⁺: *m/z* calcd. for C₂₀₆H₈₀D₁₅₆N₁₂O₂₄⁺: 3418.741, found: 3418.847. **IR:** (ATR, FT) $\tilde{\nu}$ (cm⁻¹) 2210 (vw), 2097 (vw), 1620 (m), 1578 (vs), 1468 (w), 1435 (w), 1391 (w), 1369 (w), 1317 (w), 1296 (w), 1223 (vw), 1165 (m), 1124 (s), 1053 (vw), 993 (w), 970 (w), 874 (w), 849 (w), 798 (vw), 766 (w), 741 (w), 669 (vw), 638 (w). **UV/vis** (THF) λ_{max} (lg ϵ) 318.8 (5.28), 377.8 (5.46). **Elemental Anal.** Calcd. for C₂₀₆H₈₀D₁₅₆N₁₂O₂₄·2 H₂O: C, 71.57; H+D, 7.03 (Value was corrected by detection difference of hydrogen and deuterium. Analyzing a compound containing a known deuterium amount resulted in detection of 50.4% of the apparent deuterium.); N, 4.86. Found: C, 71.41; H+D, 7.20; N, 4.89.

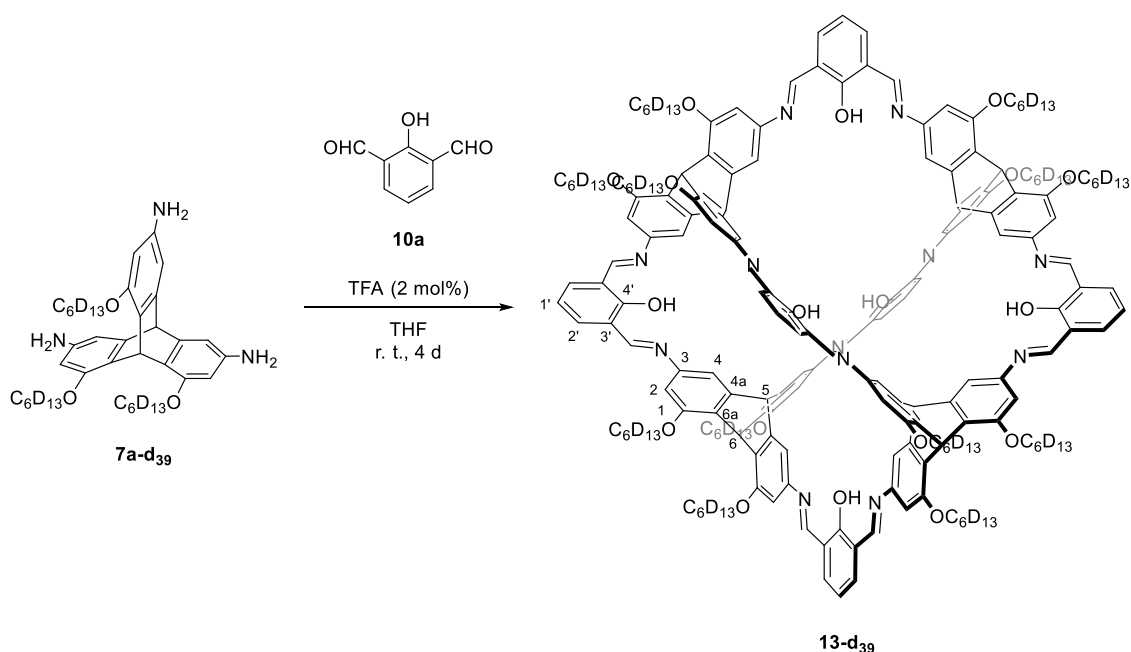
2.2.5 [4+6]-endo Cage 13



In an 8 mL screw-capped vial triaminotriptycene **7a** (20.1 mg, 34 μ mol) and 2,6-diformylphenol **10a** (7.8 mg, 52 μ mol) were combined under argon atmosphere. Dry

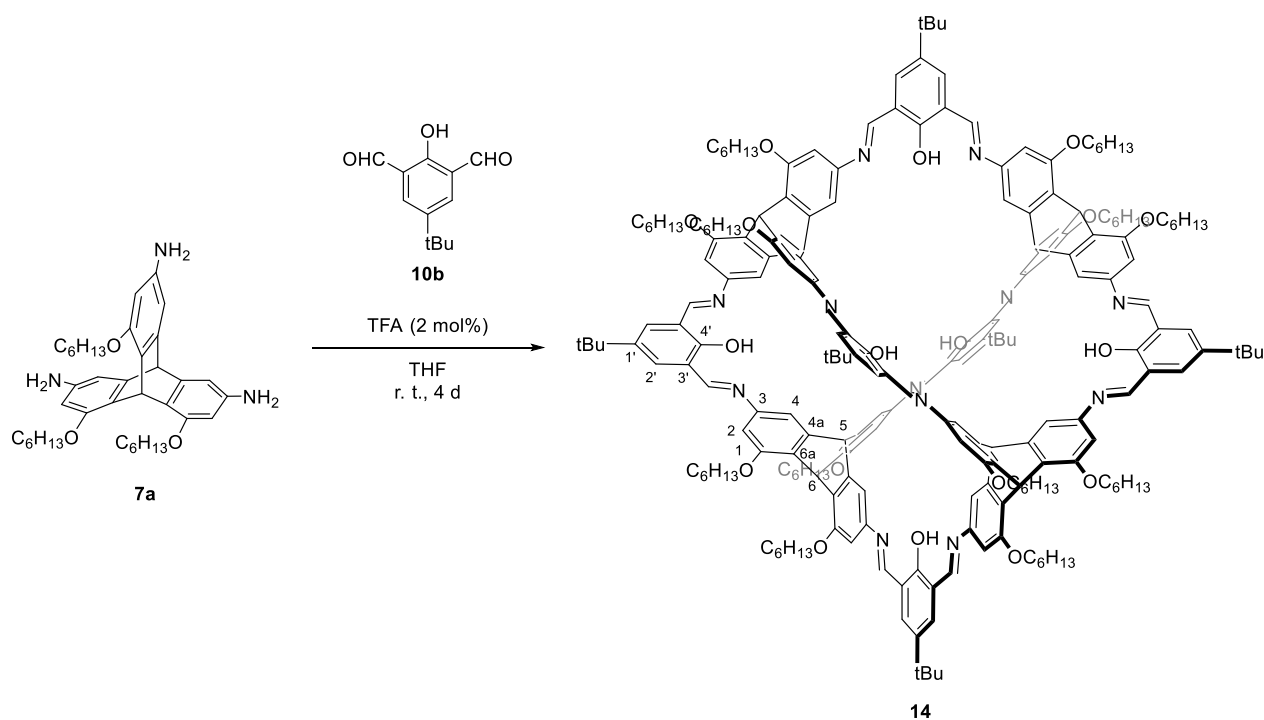
THF (5 mL) and TFA (67 μL , 0.01 mol L^{-1} in dry THF, $0.67 \mu\text{mol}$) were added and the red solution was stirred for 4 days at room temperature. Molecular sieves (3 \AA , 300 mg) was added and stirred for additional 20 min before the mixture was filtrated through Celite. Purification by rGPC (THF) gave cage **13** as yellow solid in 50% yield (12.9 mg, $4 \mu\text{mol}$). **Melting point:** $T = 357 \text{ }^\circ\text{C}$ (dec.). **$^1\text{H NMR}$:** (600 MHz, 328 K, THF- d_8) δ (ppm) 13.84 (s, 6H, -OH), 9.06 (s, 12H, -CHN-), 7.91 (s, 12H, $H2'$), 7.29 (s, 12H, $H4$), 6.96 (s, 4H, $H6$), 6.93 (t, $J = 7.7 \text{ Hz}$, 6H, $H1'$), 6.81 (s, 12H, $H2$), 5.73 (s, 4H, $H5$), 4.12 (t, $J = 6.5 \text{ Hz}$, 24H, -OCH $_2$ -), 2.00 – 1.92 (m, 24H, -OCH $_2$ -CH $_2$ -), 1.73 – 1.66 (m, 24H, -OCH $_2$ -CH $_2$ -CH $_2$ -), 1.53 – 1.42 (m, 48H, -CH $_2$ -CH $_2$ -CH $_3$), 0.99 (t, $J = 6.8 \text{ Hz}$, 36H, -CH $_3$). **$^{13}\text{C}\{^1\text{H}\} \text{NMR}$:** (150 MHz, 328 K, THF- d_8) δ (ppm) 162.4 ($C4'$), 155.7 ($C1$), 150.3 ($C4a$), 148.5 ($C3$), 133.8 ($C6a$, $C2'$), 123.7 ($C3'$), 119.7 ($C1'$), 110.8 ($C4$), 105.5 ($C2$), 70.0 (-OCH $_2$ -), 56.2 ($C5$), 34.9 ($C6$), 33.0 (-CH $_2$ -CH $_2$ -CH $_3$), 30.9 (-OCH $_2$ -CH $_2$ -), 26.9 (-OCH $_2$ -CH $_2$ -CH $_2$ -), 23.7 (-CH $_2$ -CH $_2$ -CH $_3$), 14.6 (-CH $_3$). **MS (MALDI-TOF+, DCTB):** $[\text{M}+\text{H}]^+$: m/z calcd. for $\text{C}_{200}\text{H}_{225}\text{N}_{12}\text{O}_{18}^+$: 3082.705, found: 3083.223. **IR:** (ATR, FT) $\tilde{\nu}$ (cm^{-1}) 2953 (w), 2928 (m), 2858 (w), 1620 (m), 1593 (vs), 1576 (vs), 1468 (m), 1435 (m), 1377 (w), 1358 (w), 1321 (m), 1300 (m), 1258 (w), 1229 (w), 1169 (w), 1124 (s), 1099 (s), 1076 (w), 1007 (m), 976 (m), 916 (vw), 858 (m), 793 (w), 750 (s), 723 (m), 640 (w). **UV/vis** (THF) λ_{max} ($\lg \epsilon$) 295.8 (5.22), 372.8 (5.35). **Elemental Anal.** Calcd. for $\text{C}_{200}\text{H}_{224}\text{N}_{12}\text{O}_{18} \cdot 5 \text{ H}_2\text{O}$: C, 75.68; H, 7.43; N, 5.30. Found: C, 75.73; H, 7.35; N, 4.99.

2.2.6 [4+6]-endo Cage 13-d₁₅₆



In an 8 mL screw-capped vial 3,6,15-triamino-1,8,13-trihexyloxytriptycene-d₃₉ **7a-d₃₉** (21.3 mg, 33 μmol) and 2,6-diformylphenol **10a** (7.8 mg, 52 μmol) were combined under argon atmosphere. Dry THF (5 mL) and TFA (67 μL, 0.01 mol L⁻¹ in dry THF, 0.67 μmol) were added and the red solution was stirred for 4 days at room temperature. The red solution was stirred with molecular sieves (3 Å, 300 mg) and the mixture was filtrated through Celite. The resulted red solution was purified by rGPC (THF) to give cage **13-d₁₅₆** as yellow solid in 56% yield (15.2 mg, 5 μmol). **Melting point:** *T* = 357 °C (dec.). **¹H NMR:** (700 MHz, THF- d₈) δ (ppm) 13.95 (s, 6H, -OH), 9.09 (s, 12H, -CHN-), 7.93 (s, 12H, H₂'), 7.31 (s, 12H, H₄), 6.97 (s, 4H, H₆), 6.95 (t, *J* = 7.7 Hz, 6H, H₁'), 6.83 (s, 12H, H₂), 5.77 (s, 4H, H₅). **¹³C{¹H} NMR:** (176 MHz, THF-d₈) 162.3 (-CHN-), 155.6 (C₁), 150.3 (C_{4a}), 133.6 (C_{6a}, C₂'), 123.6 (C₃'), 119.7 (C₁'), 69.1 – 68.3 (m, -OCD₂-), 55.9 (C₅), 34.6 (C₆), 32.0 – 31.3 (m, -CD₂-CD₂-CD₃), 30.0 – 29.3 (m, -OCD₂-CD₂-), 22.8 – 22.1 (m, -CD₂-CD₂-CD₃), 14.1 – 13.2 (m, -CD₃). **²H NMR:** (92 MHz, THF/THF- d₈) δ (ppm) 4.11 (s, 24D, -OCD₂-), 1.91 (s, 24D, -OCD₂-CD₂-), 1.64 (s, 24D, -OCD₂-CD₂-CD₂-), 1.40 (s, 48H, -CD₂-CD₂-CD₃), 0.93 (s, 36D, -CD₃). **MS (MALDI-TOF+, DCTB):** [M]⁺: *m/z* calcd. for C₂₀₀H₆₈D₁₅₆N₁₂O₁₈⁺: 3238.677, found: 3238.642. **IR:** (ATR, FT) $\tilde{\nu}$ (cm⁻¹) 2210 (w), 2098 (w), 1620 (m), 1593 (vs), 1576 (vs), 1474 (m), 1437 (m), 1360 (w), 1323 (m), 1302 (m), 1256 (w), 1231 (w), 1167 (w), 1124 (vs), 1105 (m), 1088 (m), 1053 (w), 1040 (w), 1005 (w), 982 (m), 918 (vw), 858 (m), 793 (w), 750 (s), 721 (w), 642 (w). **UV/vis** (THF) λ_{max} (lg ϵ) 295.8 (5.20), 372.8 (5.32). **Elemental Anal.** Calcd. for C₂₀₀H₆₈D₁₅₆N₁₂O₁₈·H₂O: C, 73.71; H+D, 7.02 (Value was corrected by detection difference of hydrogen and deuterium. Analyzing a compound containing a known deuterium amount resulted in detection of 50.4% of the apparent deuterium.); N, 5.16. Found: C, 73.62; H+D, 7.07; N, 4.87.

2.2.7 [4+6]-endo Cage 14

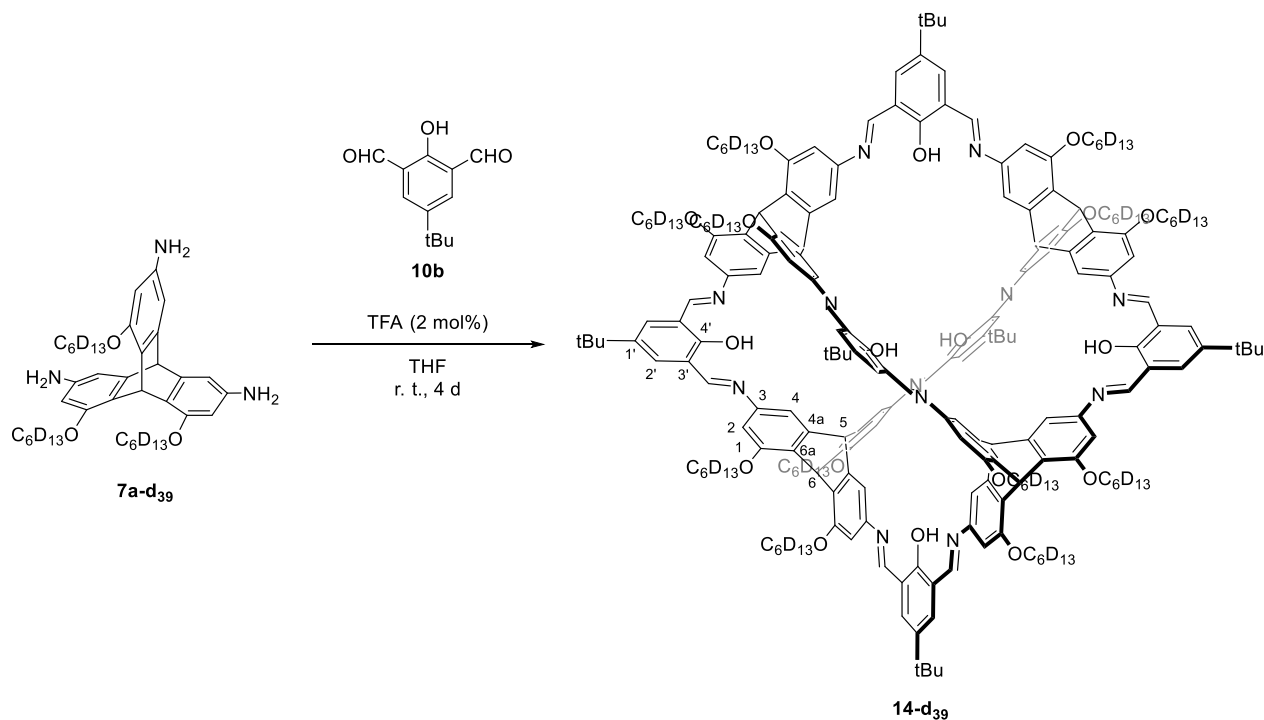


In an 8 mL screw-capped triaminotriptycene **7a** (20.0 mg, 33 μmol) and 2,6-diformyl-4-*tert*-butylphenol **10b** (10.3 mg, 50 μmol) were combined under argon atmosphere. Dry THF (5 mL) and TFA (67 μL , 0.01 mol L⁻¹ in dry THF, 0.67 μmol) were added and the red solution was stirred for 4 days at room temperature. Afterwards the solution was stirred with molecular sieves (3 Å, 300 mg) for 20 min and filtrated through Celite. The product was further purified by rGPC (THF) to give cage **14** as orange solid in 62% yield (17.6 mg, 5 μmol).

Melting point: $T = 380\text{ }^{\circ}\text{C}$ (dec.). **¹H NMR:** (700 MHz, THF- d_8) δ (ppm) 13.71 (s, 6H, -OH), 9.12 (s, 12H, -CHN-), 8.02 (s, 12H, $H2'$), 7.33 (s, 12H, $H4$), 6.98 (s, 4H, $H6$), 6.86 (s, 12H, $H2$), 5.79 (s, 4H, $H5$), 4.12 (t, $J = 6.5$ Hz, 24H, -OCH₂-), 2.04 – 1.92 (m, 24H, -OCH₂-CH₂-), 1.78 – 1.67 (m, 24H, -OCH₂-CH₂-CH₂-), 1.55 – 1.43 (m, 48H, -CH₂-CH₂-CH₃), 1.35 (s, 54H, -C(CH₃)₃), 1.06 – 0.95 (m, 36H, -CH₂-CH₂-CH₃). **¹³C{¹H} NMR:** (176 MHz, THF- d_8) δ (ppm) 160.3 ppm(-CHN-), 159.9 (C4'), 155.5 (C1), 150.3 (C4a), 148.4 (C3), 142.4 (C1'), 142.3 (C1'), 133.5 (C6a), 130.9 (C2'), 122.9 (C3'), 69.5 (-OCH₂-), 55.9 (C5), 35.0 (C1'-C(CH₃)₃), 34.6 (C6), 33.0 (-CH₂-CH₂-CH₃), 31.9 (-C1'-C(CH₃)₃), 30.9 (-OCH₂-CH₂-), 27.0 (-OCH₂-CH₂-CH₂-), 23.9 (-CH₂-CH₂-CH₃), 14.8 (-CH₂-CH₂-CH₃). **MS (MALDI-TOF+, DCTB):** [M+H]⁺: m/z calcd. for C₂₂₄H₂₇₃N₁₂O₁₈⁺: 3419.081, found: 3419.723. **IR:** (ATR, FT) $\tilde{\nu}$ (cm⁻¹) 2955 (m), 2930 (w), 2870 (w), 1624 (m), 1582 (vs), 1468 (s), 1433 (m), 1394 (w), 1377 (w), 1362 (w), 1300 (m), 1263 (m),

1227 (m), 1178 (w), 1124 (s), 1099 (s), 1065 (w), 1038 (w), 1015 (m), 976 (m), 918 (w), 887 (w), 858 (m), 825 (w), 762 (m), 744 (m), 729 (w), 650 (vw), 631 (w). **UV/vis** (THF) λ_{max} (lg ϵ) 298.4 nm (5.16), 377.8 nm (5.26). **Elemental Anal.** Calcd. for $C_{224}H_{272}N_{12}O_{18} \cdot 3THF$: C, 77.94; H, 8.20; N, 4.62. Found: C, 77.78; H, 8.40; N, 4.96.

2.2.8 [4+6]-endo Cage **14-d₁₅₆**



In an 8 mL screw-capped vial triaminotriptycene **7a-d₃₉** (21.3 mg, 33 μ mol) and 2,6-diformyl-4-*tert*-butylphenol **10b** (10.5 mg, 51 μ mol) were combined under argon atmosphere. Dry THF (5 mL) and TFA (67 μ L, 0.01 mol L⁻¹ in dry THF, 0.67 μ mol) were added and the red solution was stirred for 4 days at room temperature. Afterwards the solution was stirred with molecular sieves (3 Å, 300 mg) for 20 min and filtrated through Celite. Further purification by GPC (THF) gave cage **14-d₁₅₆** as orange solid in 66% yield (19.6 mg, 5 μ mol). **Melting point:** $T = 380$ °C (dec.) **¹H NMR:** (700 MHz, THF-*d*₈) δ (ppm) 13.69 (s, 6H, -OH), 9.11 (s, 12H, -CHN-), 8.01 (s, 12H, *H*2'), 7.31 (s, 12H, *H*4), 6.97 (s, 4H, *H*6), 6.84 (s, 12H, *H*2), 5.77 (s, 4H, *H*5), 1.34 (s, 54H, -C(CH₃)₃). **¹³C{¹H} NMR:** (176 MHz, THF- *d*₈) δ (ppm) 160.3 (-CHN-), 155.5 (C1), 150.3 (C4a), 148.4 (C3), 142.3 (C1'), 133.5 (C6a), 123.0 (C3'), 69.0 – 68.3 (m, -OCD₂-), 55.9 (C5), 35.0 (C1'-C(CH₃)₃), 34.6 (C6), 31.9 (-C1'-C(CH₃)₃) 31.8 – 31.3 (m, -CD₂-CD₂-CD₃), 30.0 – 29.3 (m, -OCD₂-CD₂-), 22.9 – 22.1 (m, -CD₂-CD₂-CD₃), 14.0 – 13.2 (m, (-CD₂-CD₂-CD₃). **²H NMR:** (92 MHz, THF/THF- *d*₈) δ (ppm) 4.13 (s, 24D, -OCD₂-), 1.92 (s, 24D, -OCD₂-

CD_2^-), 1.65 (s, 24D, $-OCD_2-CD_2-CD_2-$), 1.41 (s, 48D, $-CD_2-CD_2-CD_3$), 0.93 (s, 36D, $-CD_2-CD_2-CD_3$). **MS (MALDI-TOF+, DCTB):** $[M]^+$: m/z calcd. for $C_{224}H_{116}D_{156}N_{12}O_{18}^+$: 3575.053, found: 3575.511. **IR:** (ATR, FT) $\tilde{\nu}$ (cm^{-1}) 2961 (w), 2907 (vw), 2874 (vw), 2212 (w), 2098 (w), 1624 (m), 1583 (vs), 1474 (m), 1433 (m), 1362 (w), 1323 (w), 1304 (m), 1285 (w), 1229 (m), 1167 (w), 1126 (s), 1109 (m), 1040 (w), 1007 (w), 991 (w), 976 (w), 885 (w), 858 (w), 827 (w), 760 (w), 744 (w), 712 (vw), 652 (vw), 629 (w). **UV/vis** (THF) λ_{max} ($\lg \epsilon$) 298.2 (5.15), 376.8 (5.26). **Elemental Anal.** Calcd. for $C_{224}H_{116}D_{156}N_{12}O_{18} \cdot 4H_2O$: C, 73.72; H+D, 7.76 (Value was corrected by detection difference of hydrogen and deuterium. Analyzing a compound containing a known deuterium amount resulted in detection of 50.4% of the apparent deuterium.); N, 4.61. Found: C, 73.56; H+D, 7.66; N, 4.38.

3 NMR Spectra

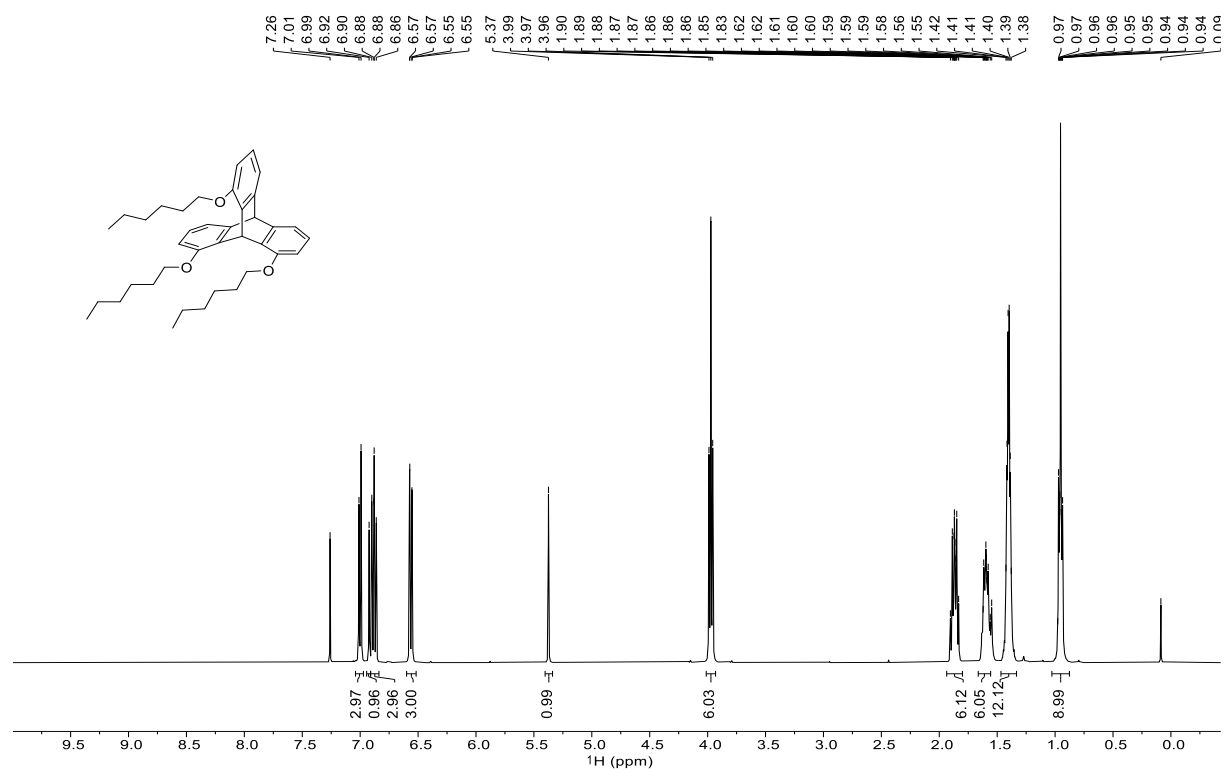


Figure S1. ¹H NMR spectrum of compound 5 (400 MHz, CDCl₃).

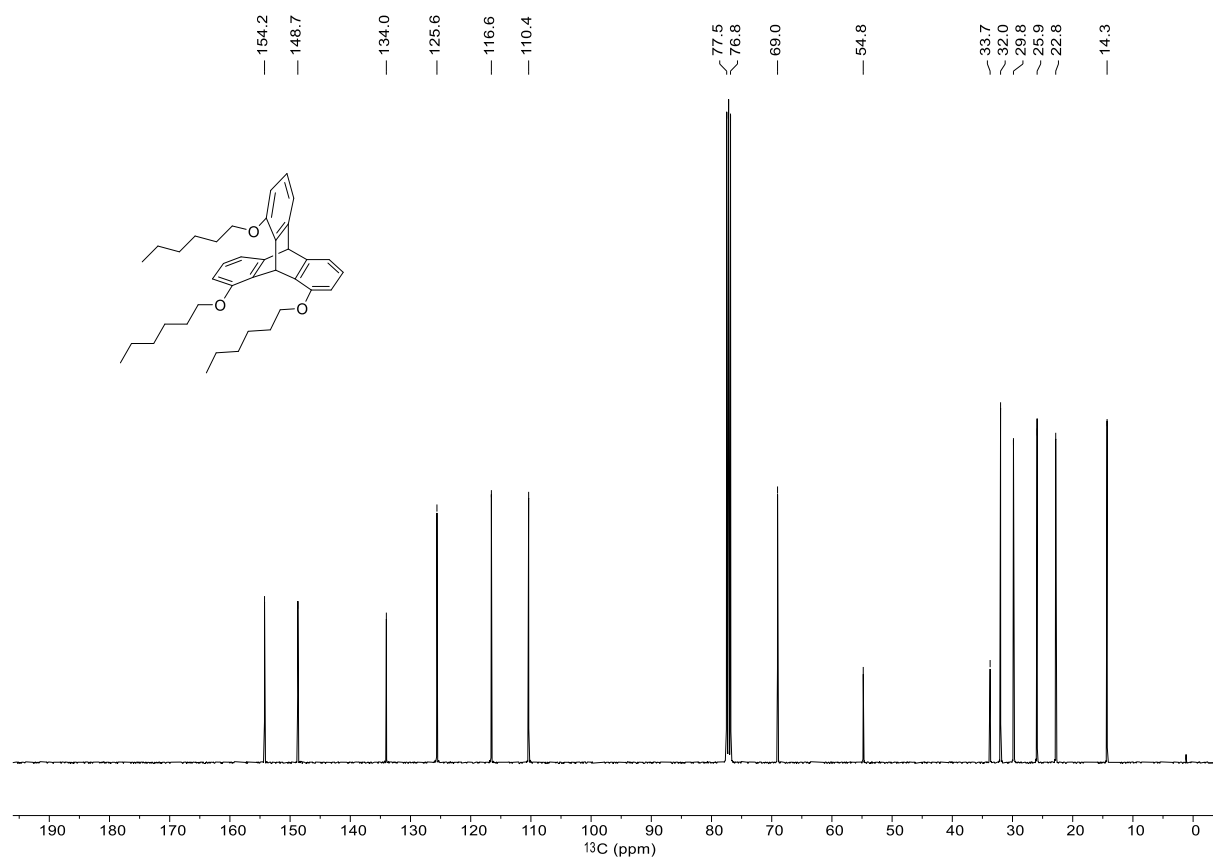


Figure S2. ¹³C{¹H} NMR spectrum of compound 5 (100 MHz, CDCl₃).

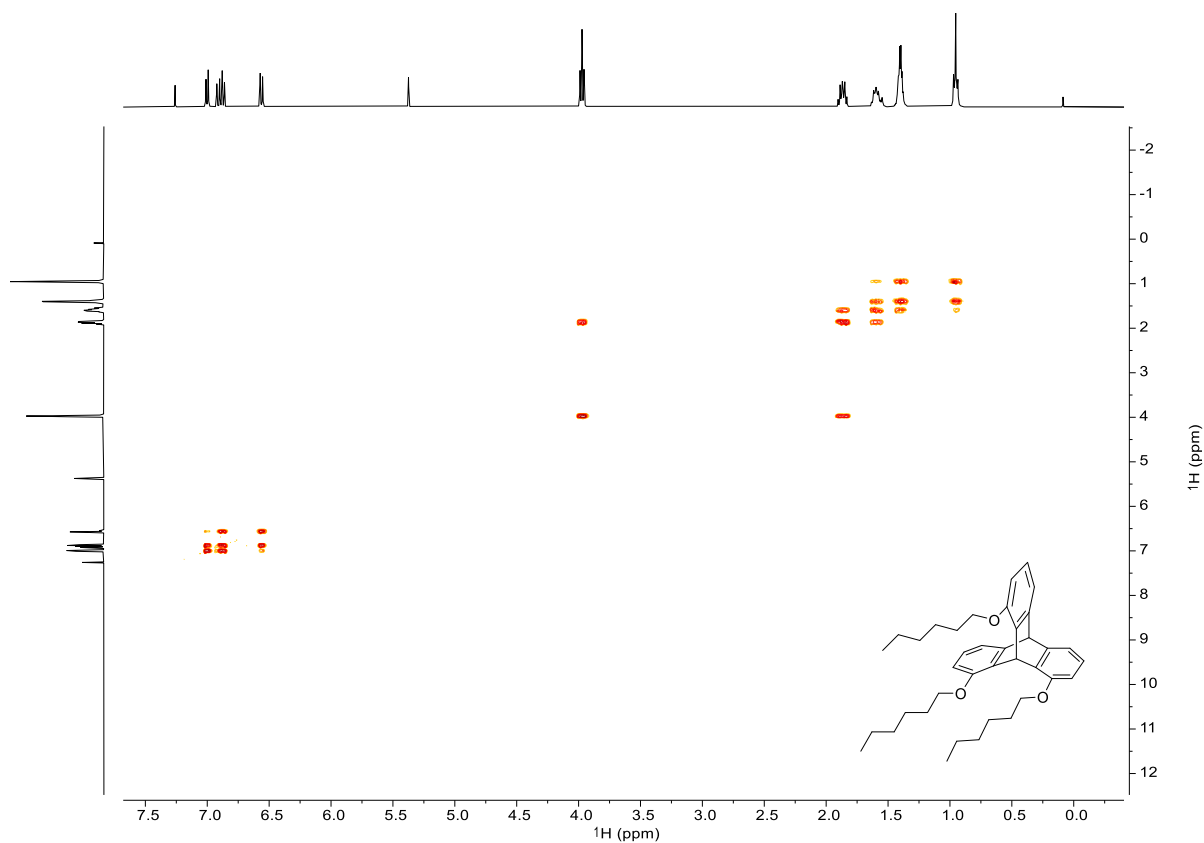


Figure S3. ¹H, ¹H-COSY spectrum of compound **5** (400 MHz, CDCl₃).

Table S1. Cross-peak assignment of ¹H, ¹H-COSY spectrum of compound **5**.

¹ H nucleus	δ_{1H} [ppm]	COSY
<i>a</i>	7.00	<i>c, d</i>
<i>b</i>	6.92	-
<i>c</i>	6.88	<i>a, d</i>
<i>d</i>	6.56	<i>a, c</i>
<i>e</i>	5.37	-
<i>f</i>	3.97	<i>g</i>
<i>g</i>	1.94 – 1.80	<i>f, h</i>
<i>h</i>	1.66 – 1.56	<i>g, i, j</i>
<i>i</i>	1.47 – 1.33	<i>h, j</i>
<i>j</i>	1.03 – 0.88	<i>h, i</i>

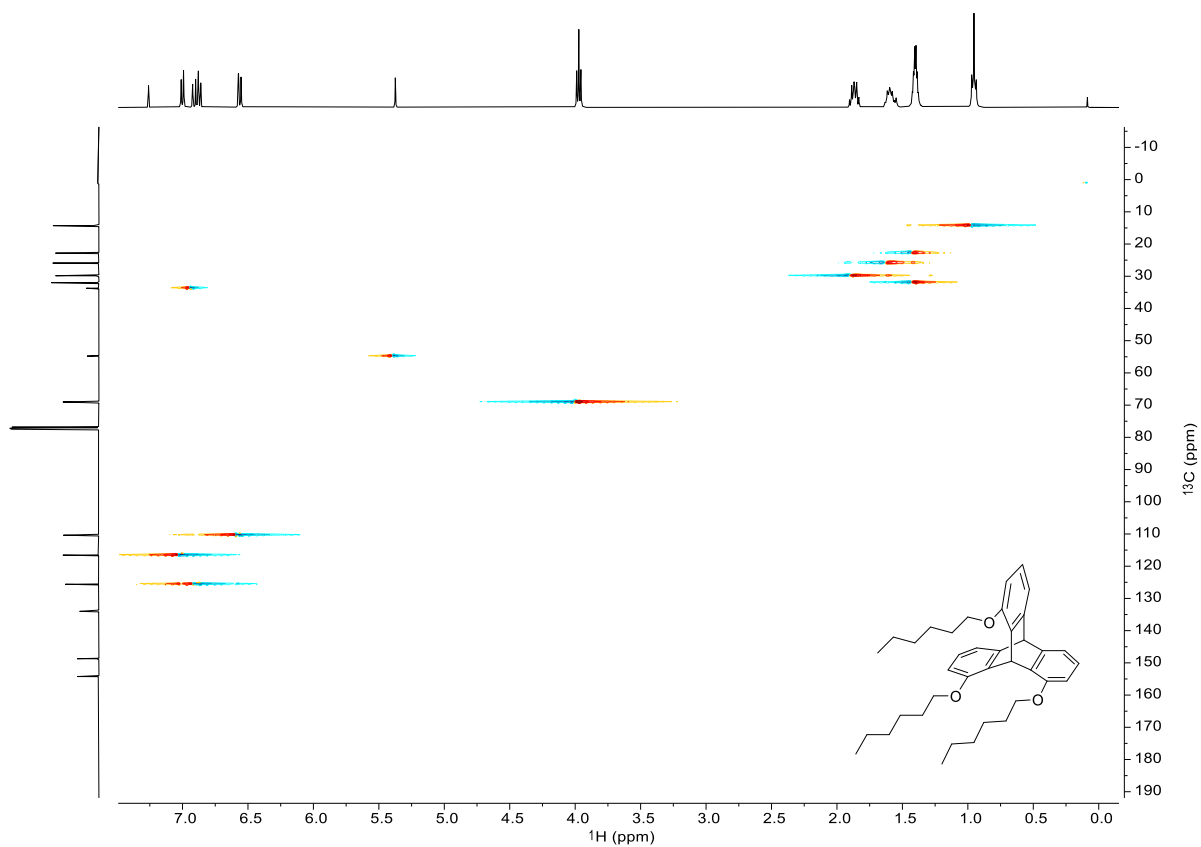


Figure S4. ^1H , ^{13}C -HSQC spectrum of compound **5** (400 MHz, 100 MHz, CDCl_3).

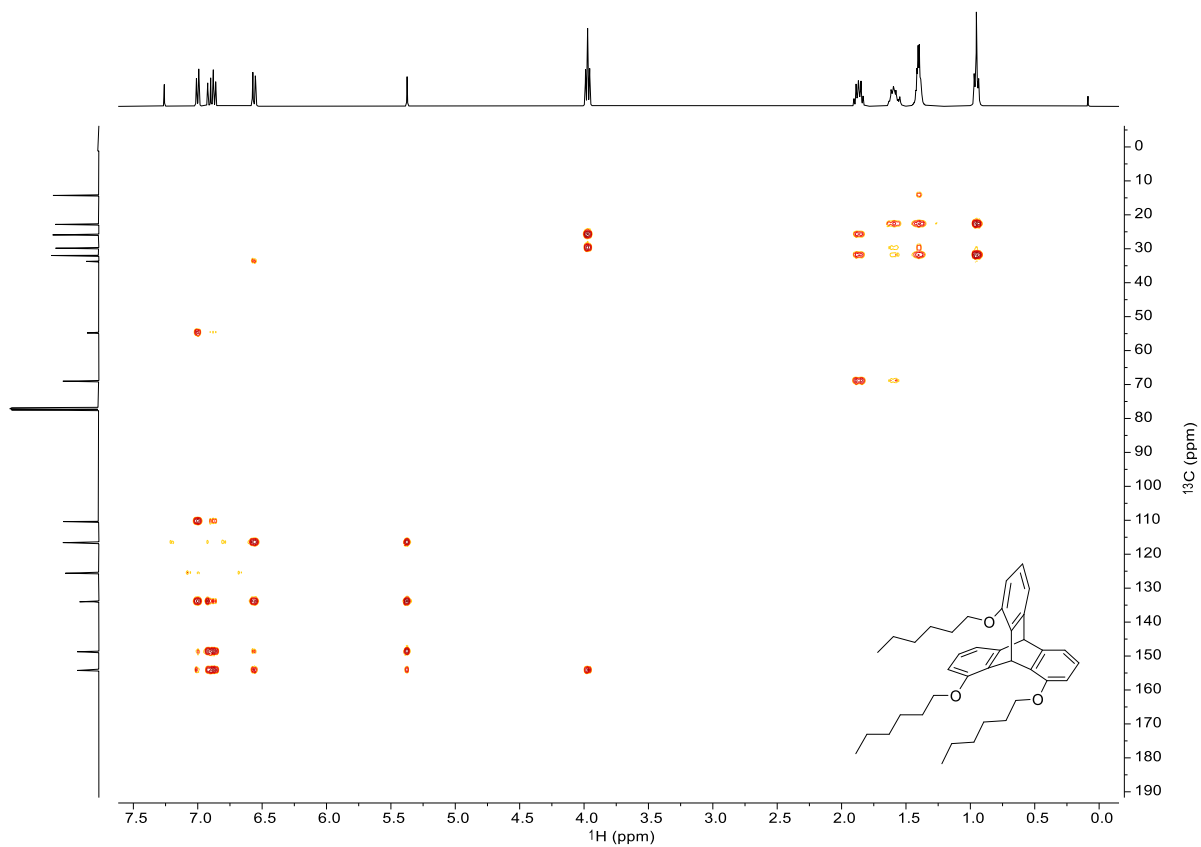


Figure S5. ^1H , ^{13}C -HMBC spectrum of compound **5** (400 MHz, 100 MHz, CDCl_3).

Table S2. Correlations of ^1H , ^{13}C -HSQC and ^1H , ^{13}C -HMBC spectra of compound **5**.

^{13}C nucleus	$\delta_{13\text{C}}$ [ppm]	^1H nucleus		
		HSQC	$\delta_{1\text{H}}$ [ppm]	HMBC
<i>A</i>	154.3	-	-	<i>a, b, c, d, e, f</i>
<i>B</i>	148.7	-	-	<i>a, b, c, d, e</i>
<i>C</i>	134.0	-	-	<i>a, b, c, d, e</i>
<i>D</i>	125.6	<i>c</i>	6.88	-
<i>E</i>	116.6	<i>a</i>	7.00	<i>d, e</i>
<i>F</i>	110.4	<i>d</i>	6.56	<i>a, c</i>
<i>G</i>	69.0	<i>f</i>	3.97	<i>g, h</i>
<i>H</i>	54.8	<i>e</i>	5.37	<i>a, c</i>
<i>I</i>	33.7	<i>b</i>	6.92	<i>d</i>
<i>J</i>	32.0	<i>i</i>	1.47 – 1.33	<i>g, h, i, j</i>
<i>K</i>	29.8	<i>g</i>	1.94 – 1.80	<i>f, h, i</i>
<i>L</i>	25.9	<i>h</i>	1.66 – 1.56	<i>f, g</i>
<i>M</i>	22.8	<i>i</i>	1.47 – 1.33	<i>h, i, j</i>
<i>N</i>	14.3	<i>j</i>	1.03 – 0.88	<i>i</i>

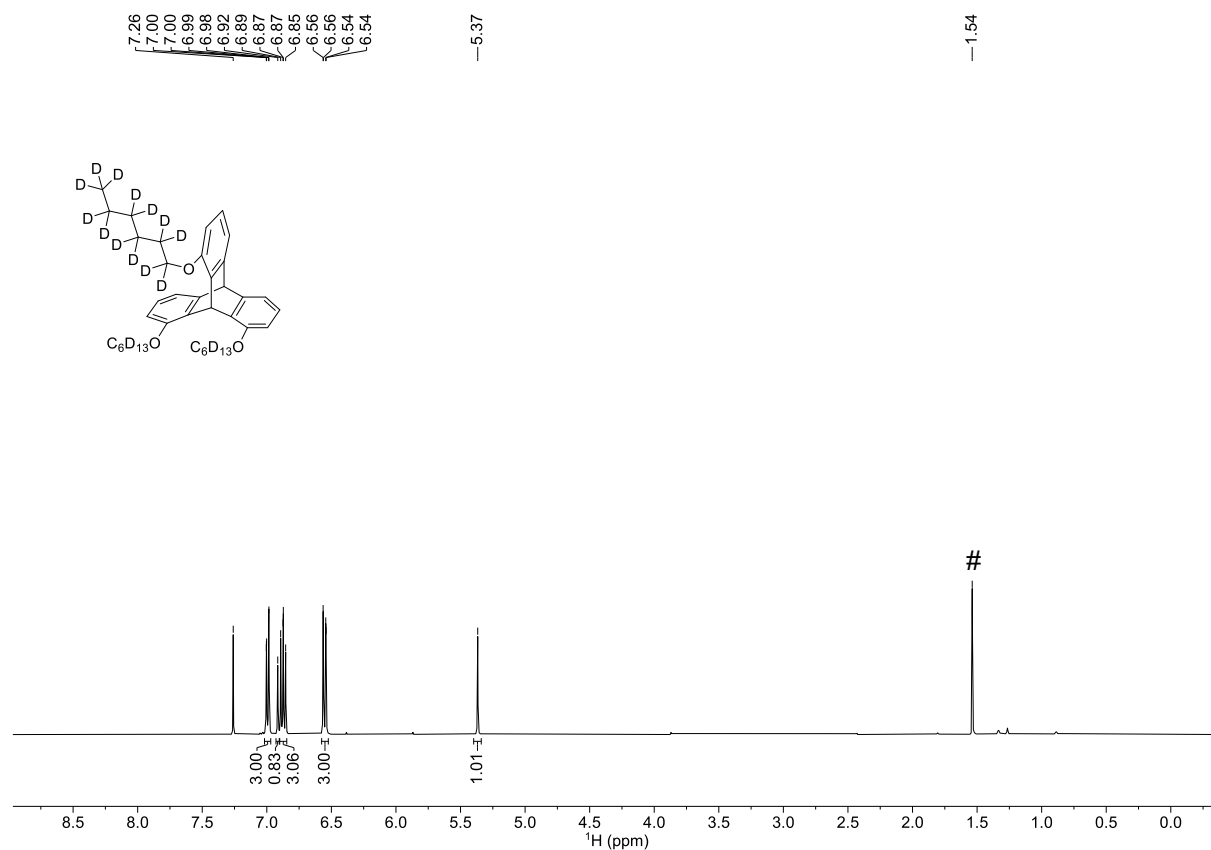


Figure S6. ^1H NMR spectrum of compound **5-d₃₉** (400 MHz, CDCl_3). #: H_2O .

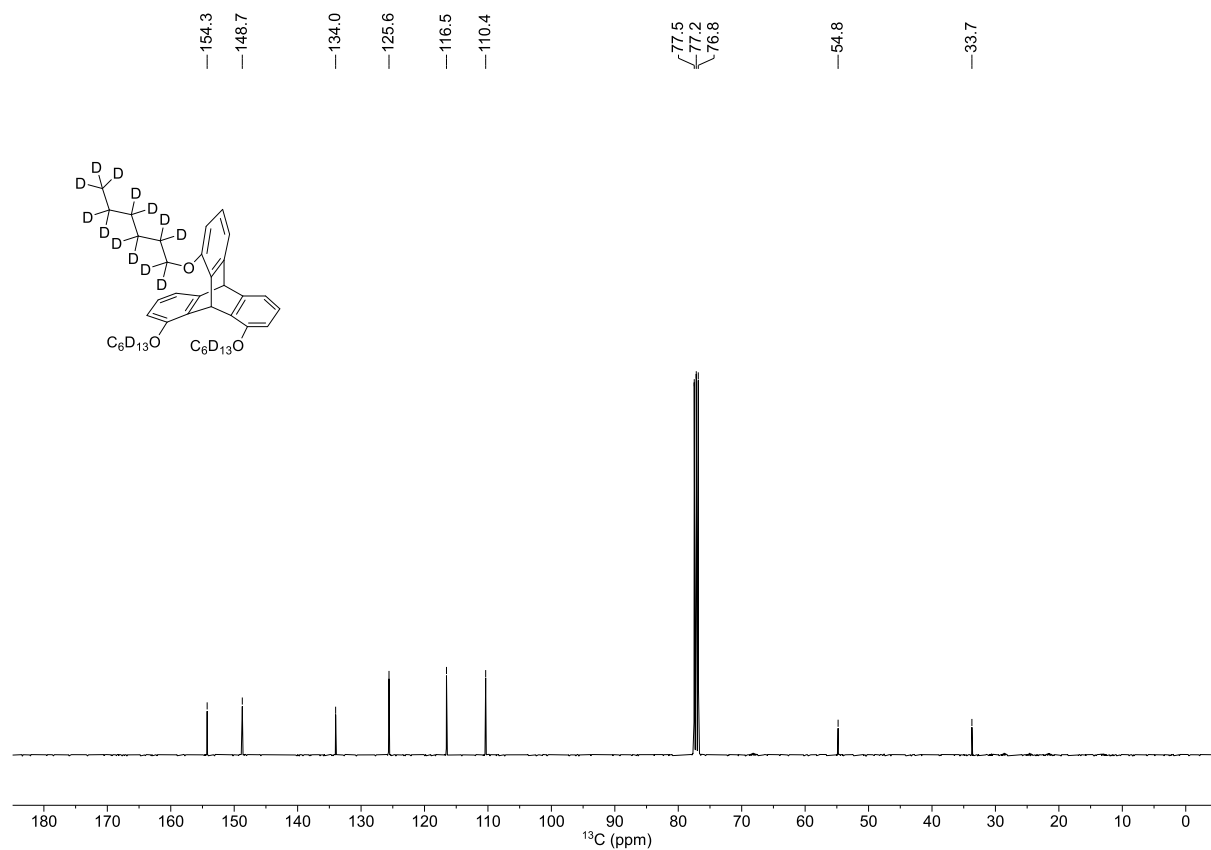


Figure S7. $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of compound **5-d₃₉** (100 MHz, CDCl_3).

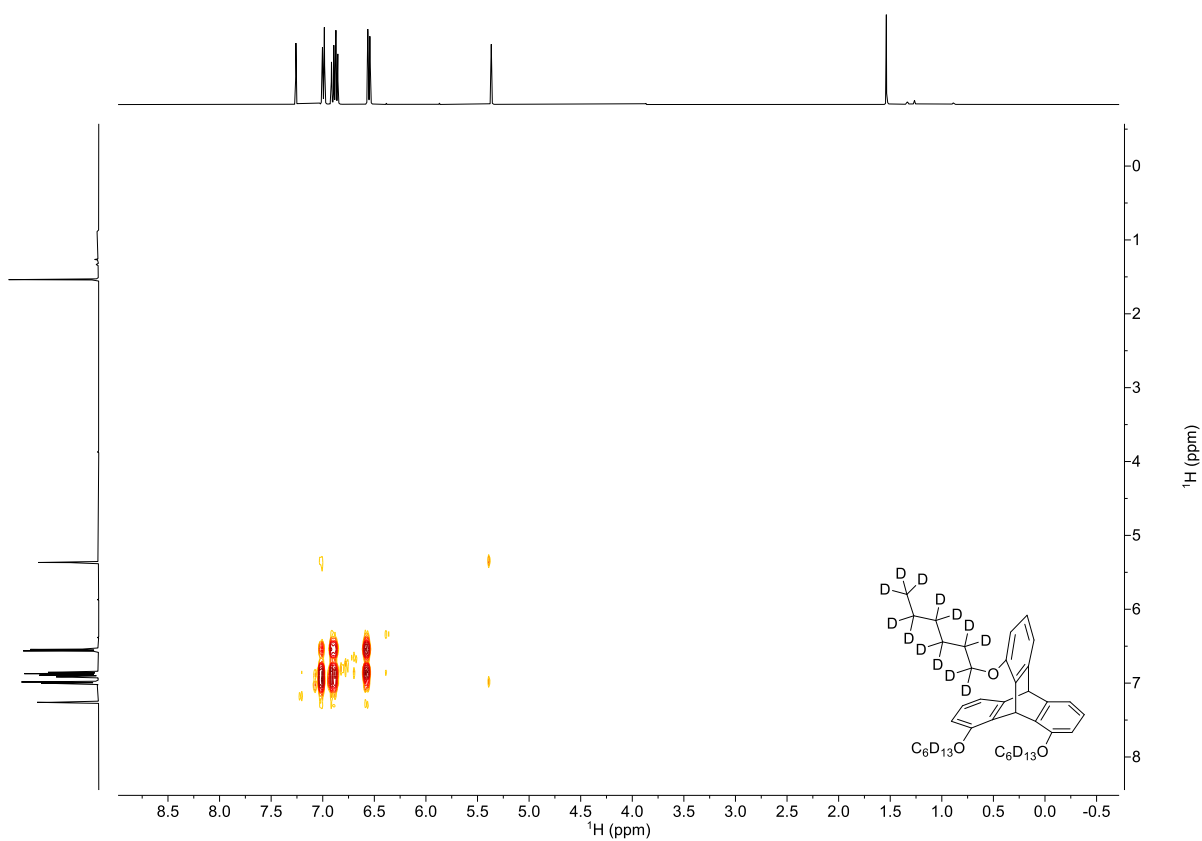


Figure S8. $^1\text{H}, ^1\text{H}$ -COSY spectrum of compound **5-d₃₉** (400 MHz, CDCl_3).

Table S3. Cross-peak assignment of $^1\text{H}, ^1\text{H}$ -COSY spectrum of compound **5-d₃₉**.

^1H nucleus	$\delta_{1\text{H}}$ [ppm]	COSY
<i>a</i>	6.99	<i>b, d, e</i>
<i>b</i>	6.92	-
<i>c</i>	6.87	<i>a, d</i>
<i>d</i>	6.55	<i>c</i>
<i>e</i>	5.37	<i>a</i>

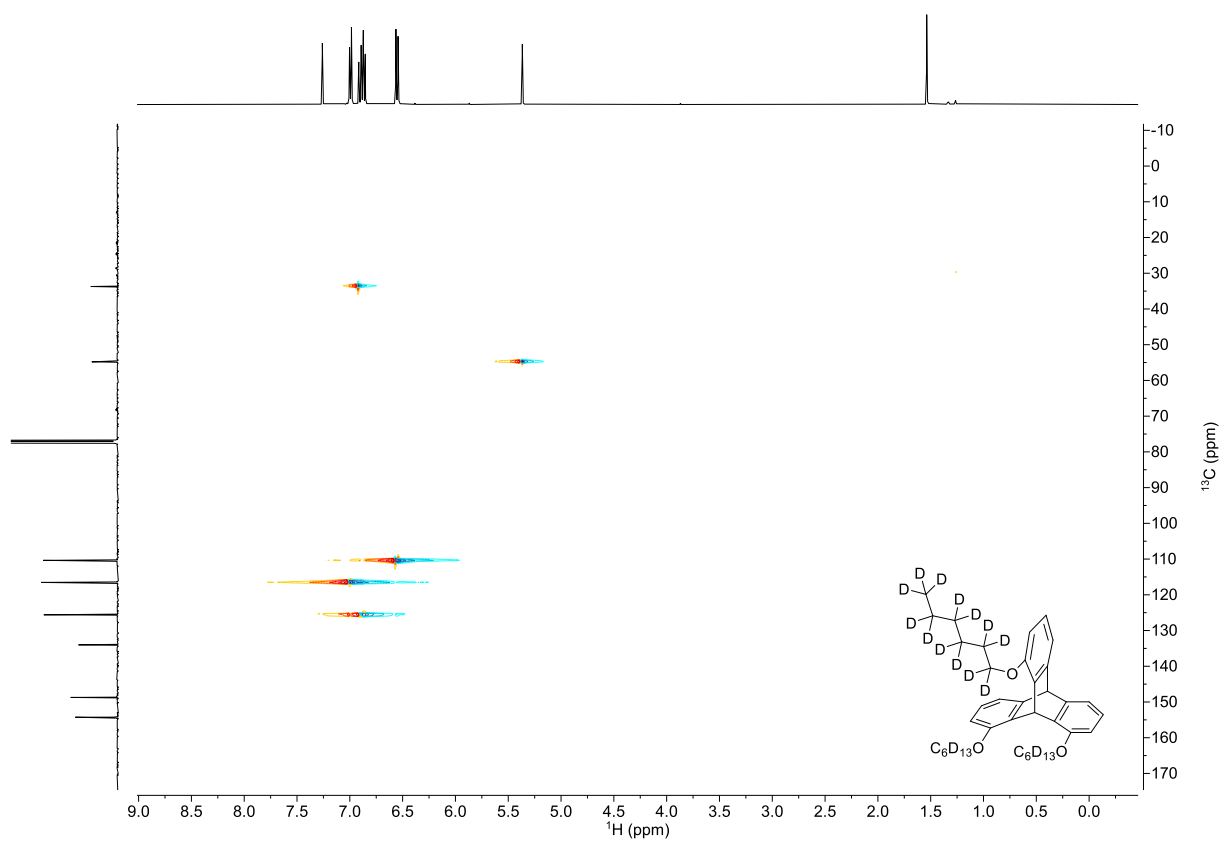


Figure S9. $^1\text{H},^{13}\text{C}$ -HSQC spectrum of compound **5-d₃₉** (400 MHz, 100 MHz, CDCl_3).

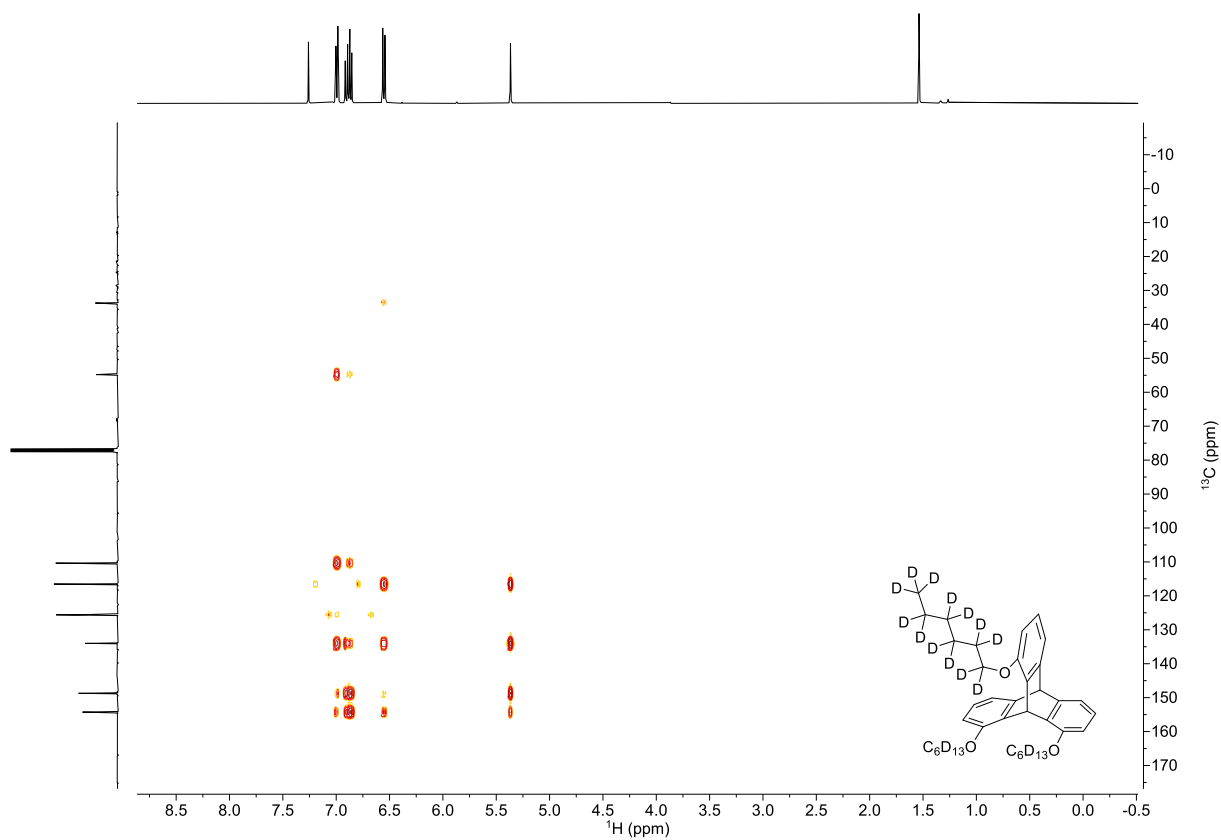


Figure S10. $^1\text{H},^{13}\text{C}$ -HMBC spectrum of compound **5-d₃₉** (400 MHz, 100 MHz, CDCl_3).

Table S4. Correlations of ^1H , ^{13}C -HSQC and ^1H , ^{13}C -HMBC spectra of compound **5-d₃₉**.

^{13}C nucleus	$\delta_{13\text{C}}$ [ppm]	^1H nucleus		
		HSQC	$\delta_{1\text{H}}$ [ppm]	HMBC
A	154.3	-	-	a, b, c, d, e
B	148.7	-	-	a, b, c, e
C	134.0	-	-	a, b, d, e
D	125.6	c	6.87	a
E	115.5	a	6.99	d, e
F	110.4	d	6.55	a, c
G	54.8	e	5.37	a
H	33.7	b	6.92	d

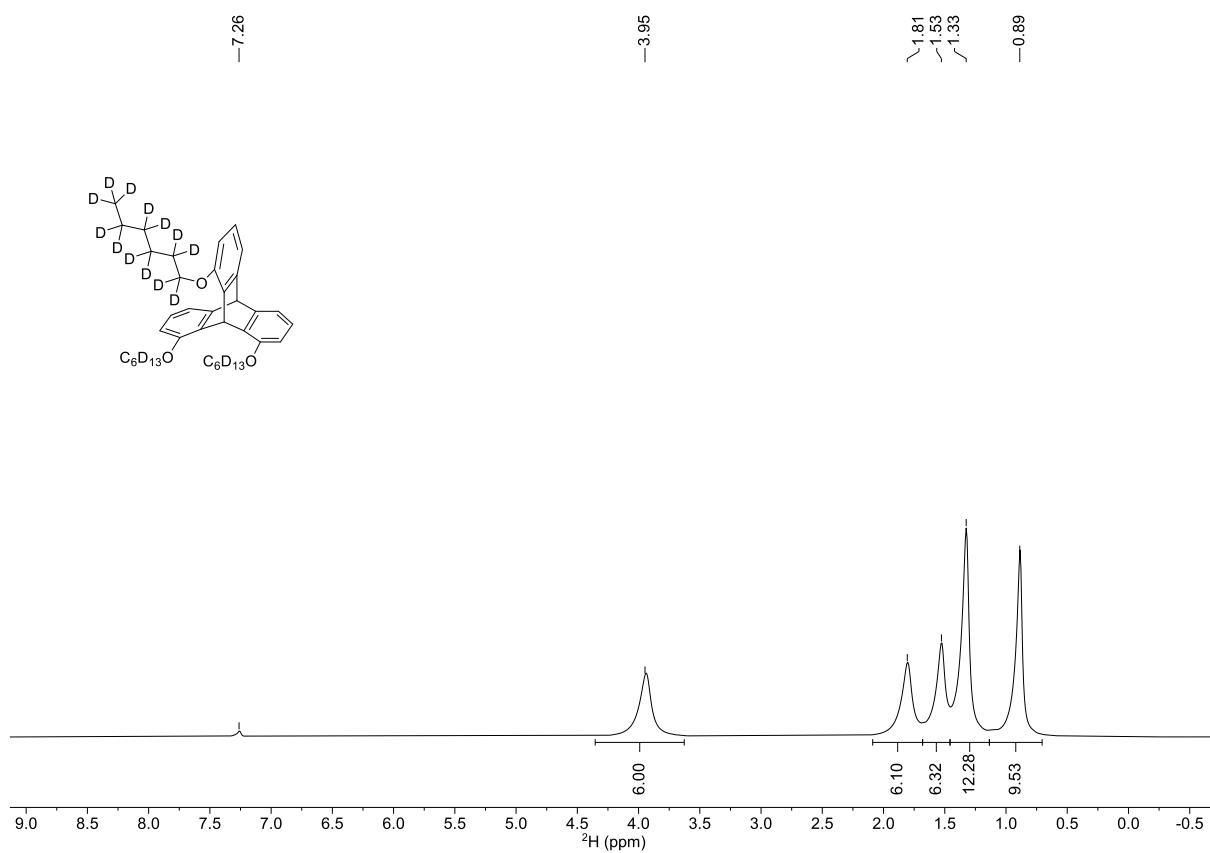


Figure S11. ^2H NMR spectrum of compound **5-d₃₉** (92 MHz, CHCl_3).

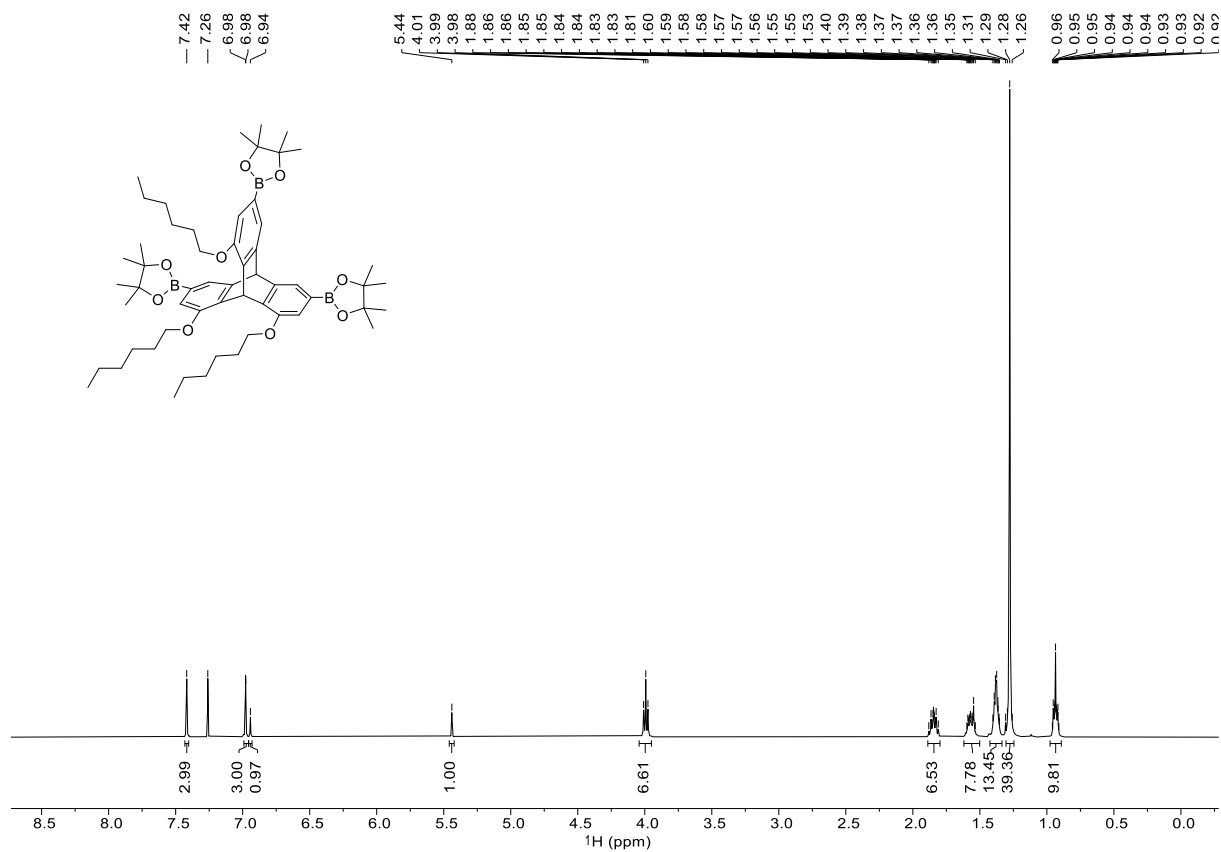


Figure S12. ^1H NMR spectrum of compound 6 (400 MHz, CDCl_3).

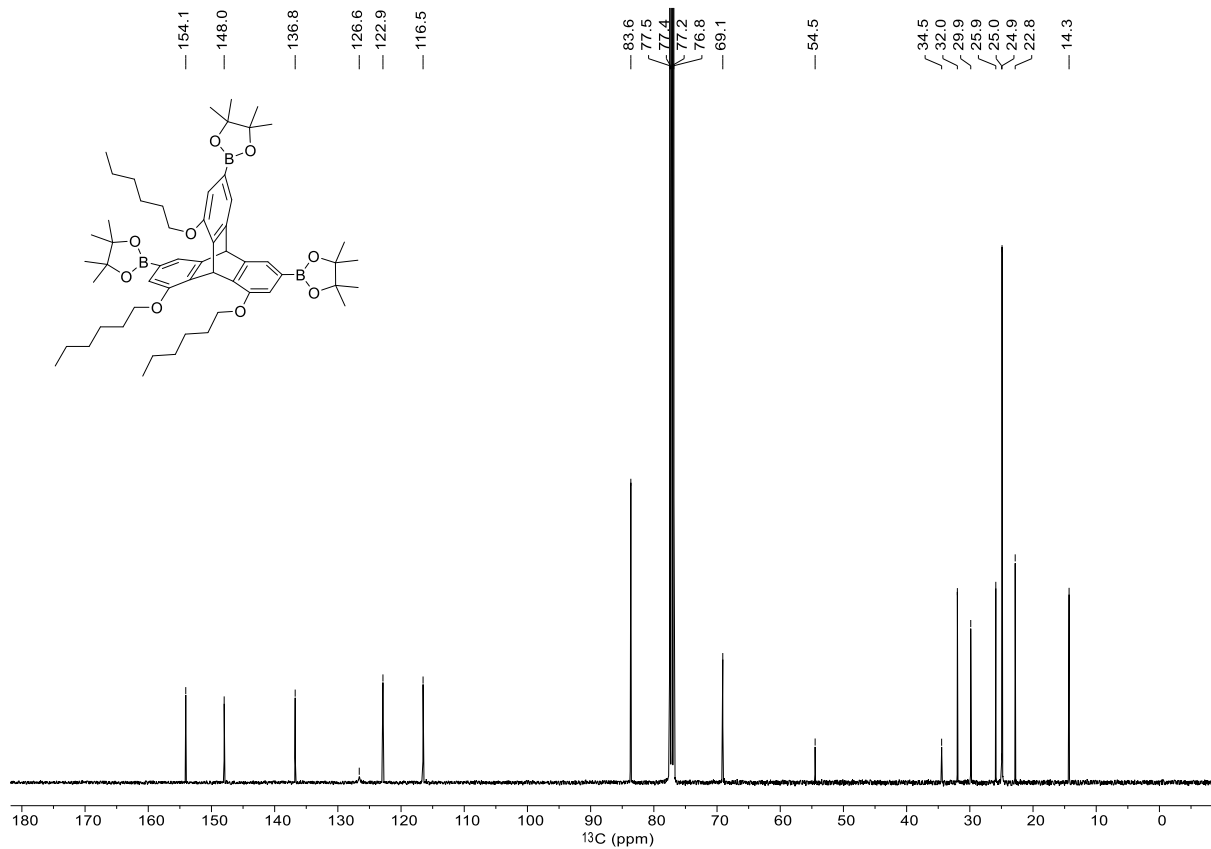


Figure S13. $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of compound 6 (100 MHz, CDCl_3).

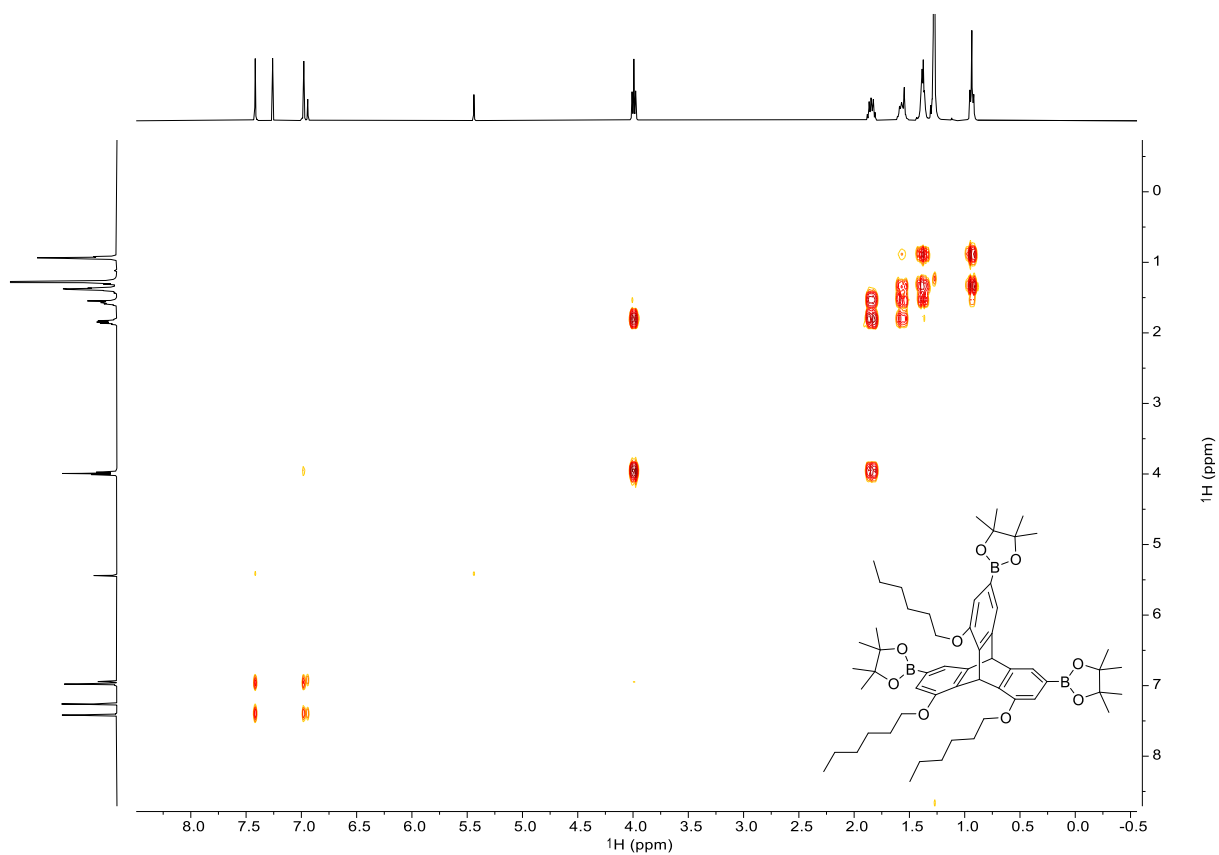


Figure S14. $^1\text{H},^1\text{H}$ -COSY spectrum of compound **6** (400 MHz, CDCl_3).

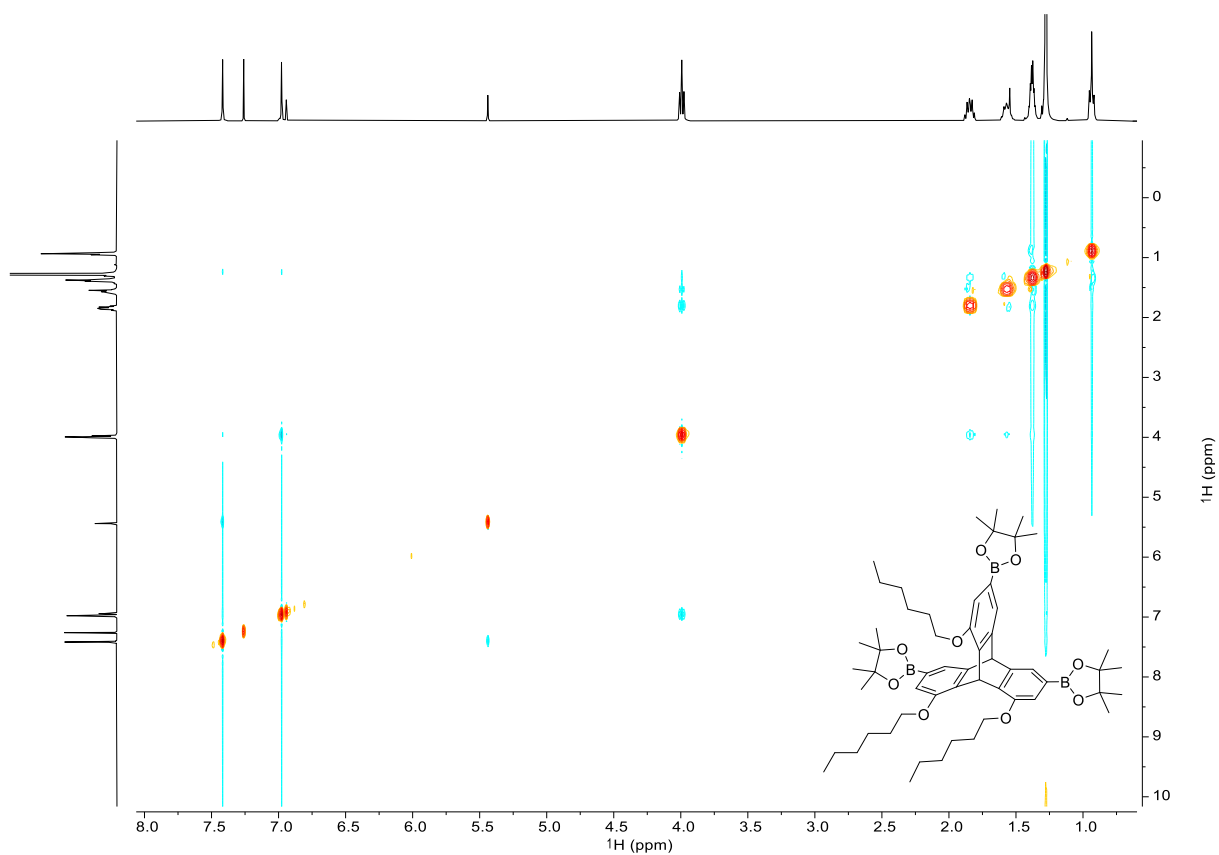


Figure S15. $^1\text{H},^1\text{H}$ -NOESY spectrum of compound **6** (400 MHz, CDCl_3).

Table S5. Cross-peak assignment of ¹H,¹H-COSY and ¹H,¹H-NOESY spectra of compound **6**.

¹ H nucleus	δ_{1H} [ppm]	COSY	NOESY
<i>a</i>	7.42	<i>b, d</i>	<i>d, i</i>
<i>b</i>	6.98	<i>a, e</i>	<i>e, f, g, h</i>
<i>c</i>	6.94	<i>a</i>	<i>e, f, g</i>
<i>d</i>	5.44	<i>a</i>	<i>a</i>
<i>e</i>	3.99	<i>f</i>	<i>b, c, f, g, h</i>
<i>f</i>	1.89 – 1.80	<i>e, g</i>	<i>b, c, e, h</i>
<i>g</i>	1.62 – 1.50	<i>f, h, j</i>	<i>b, c, e, h</i>
<i>h</i>	1.41 – 1.34	<i>g, j</i>	<i>b, e, f, g, j</i>
<i>i</i>	1.28	-	<i>a</i>
<i>j</i>	0.98 – 0.89	<i>g, h</i>	<i>h</i>

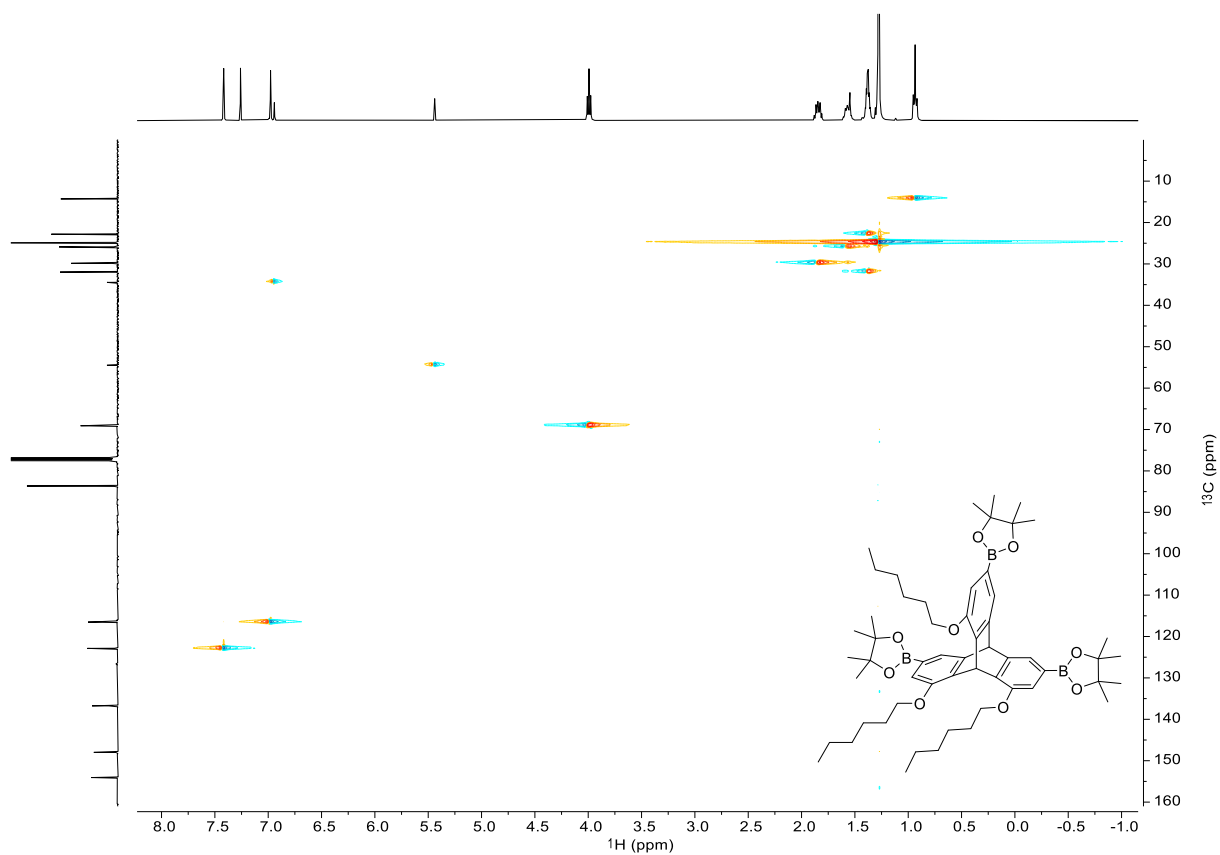


Figure S16. ^1H , ^{13}C -HSQC spectrum of compound **6** (400 MHz, 100 MHz, CDCl_3).

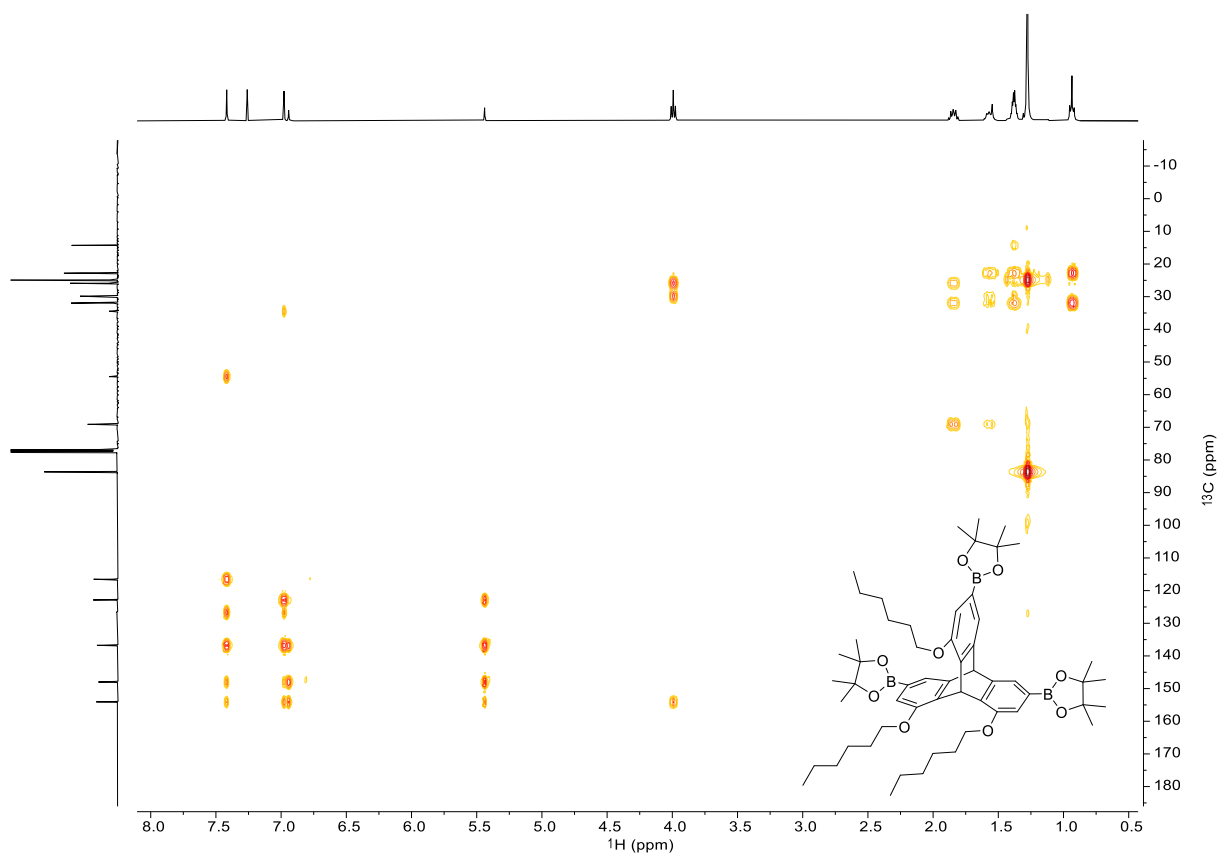
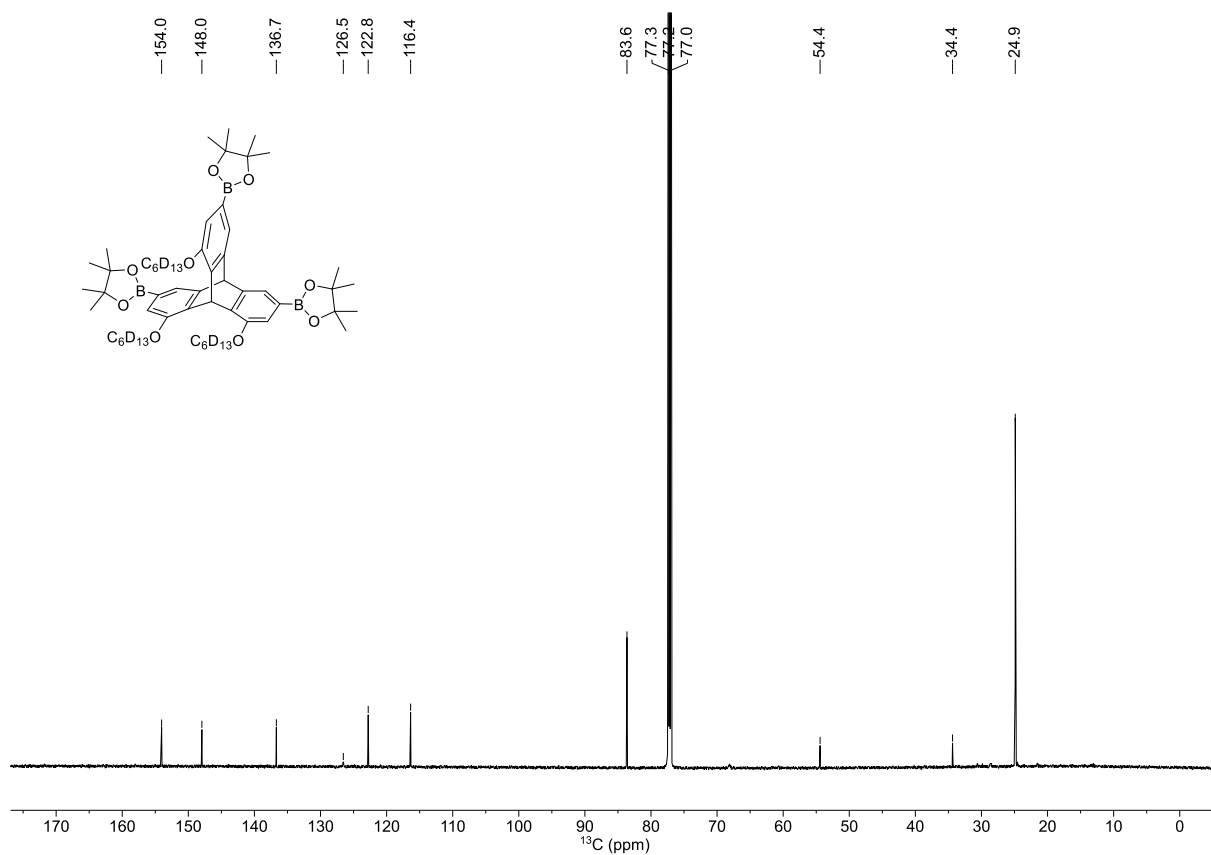
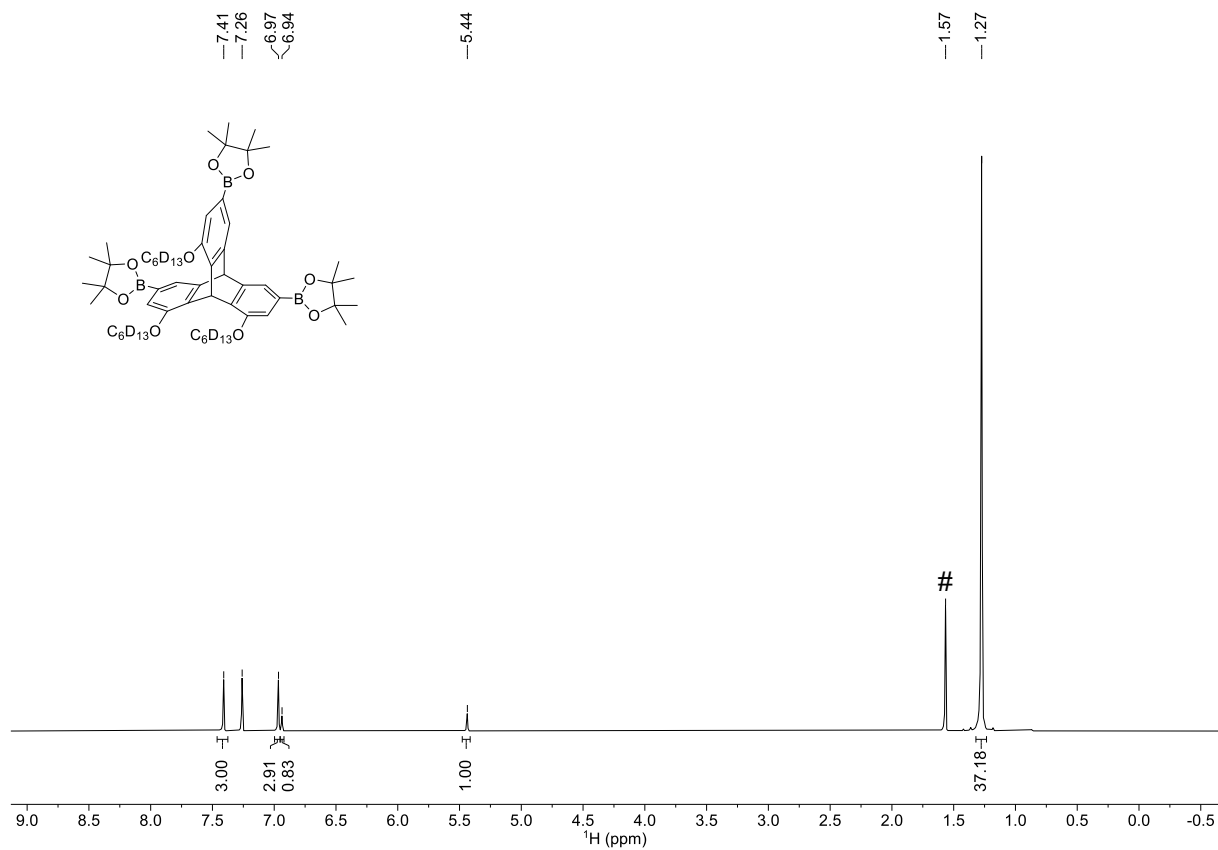
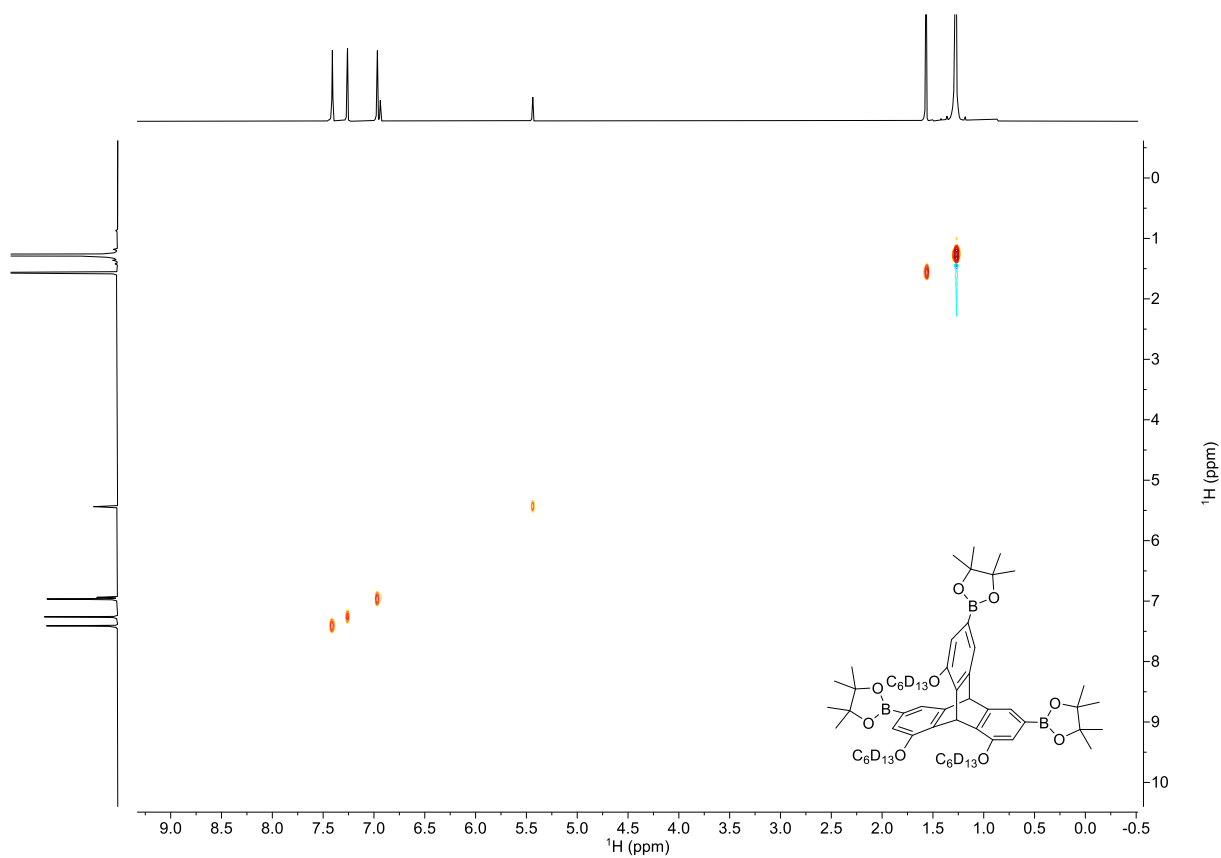
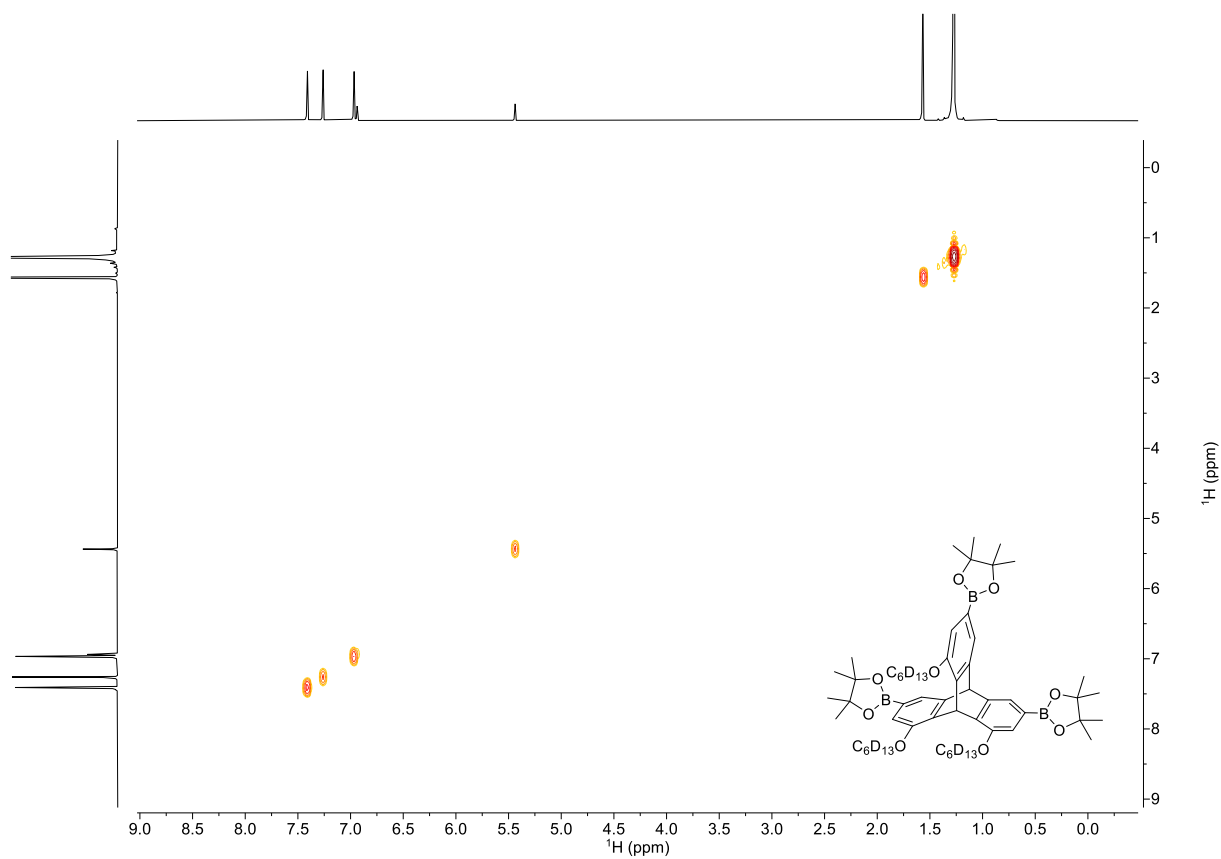


Figure S17. ^1H , ^{13}C -HMBC spectrum of compound **6** (400 MHz, 100 MHz, CDCl_3).

Table S6. Correlations of ^1H , ^{13}C -HSQC and ^1H , ^{13}C -HMBC spectra of compound **6**.

^{13}C nucleus	$\delta_{13\text{C}}$ [ppm]	^1H nucleus		
		HSQC	$\delta_{1\text{H}}$ [ppm]	HMBC
<i>A</i>	154.1	-	-	<i>a, b, c, d, e</i>
<i>B</i>	148.0	-	-	<i>a, b, c, d</i>
<i>C</i>	136.8	-	-	<i>a, b, c, d</i>
<i>D</i>	126.6	-	-	<i>a, b, i</i>
<i>E</i>	122.9	<i>a</i>	7.42	<i>b, c, d</i>
<i>F</i>	116.5	<i>b</i>	6.98	<i>a</i>
<i>G</i>	83.6	-	-	<i>i</i>
<i>H</i>	69.1	<i>e</i>	3.99	<i>f, g</i>
<i>I</i>	54.5	<i>d</i>	5.44	<i>a</i>
<i>J</i>	34.5	<i>c</i>	6.94	<i>a, b</i>
<i>K</i>	32.0	<i>h</i>	1.41 – 1.34	<i>f, g, h, j</i>
<i>L</i>	29.9	<i>f</i>	1.89 – 1.80	<i>e, g, h</i>
<i>M</i>	25.9	<i>g</i>	1.62 – 1.50	<i>e, g</i>
<i>N</i>	24.9	<i>i</i>	1.28	<i>i</i>
<i>O</i>	22.8	<i>h</i>	1.41 – 1.34	<i>g, h, j</i>
<i>P</i>	14.3	<i>j</i>	0.98 – 0.89	<i>h</i>





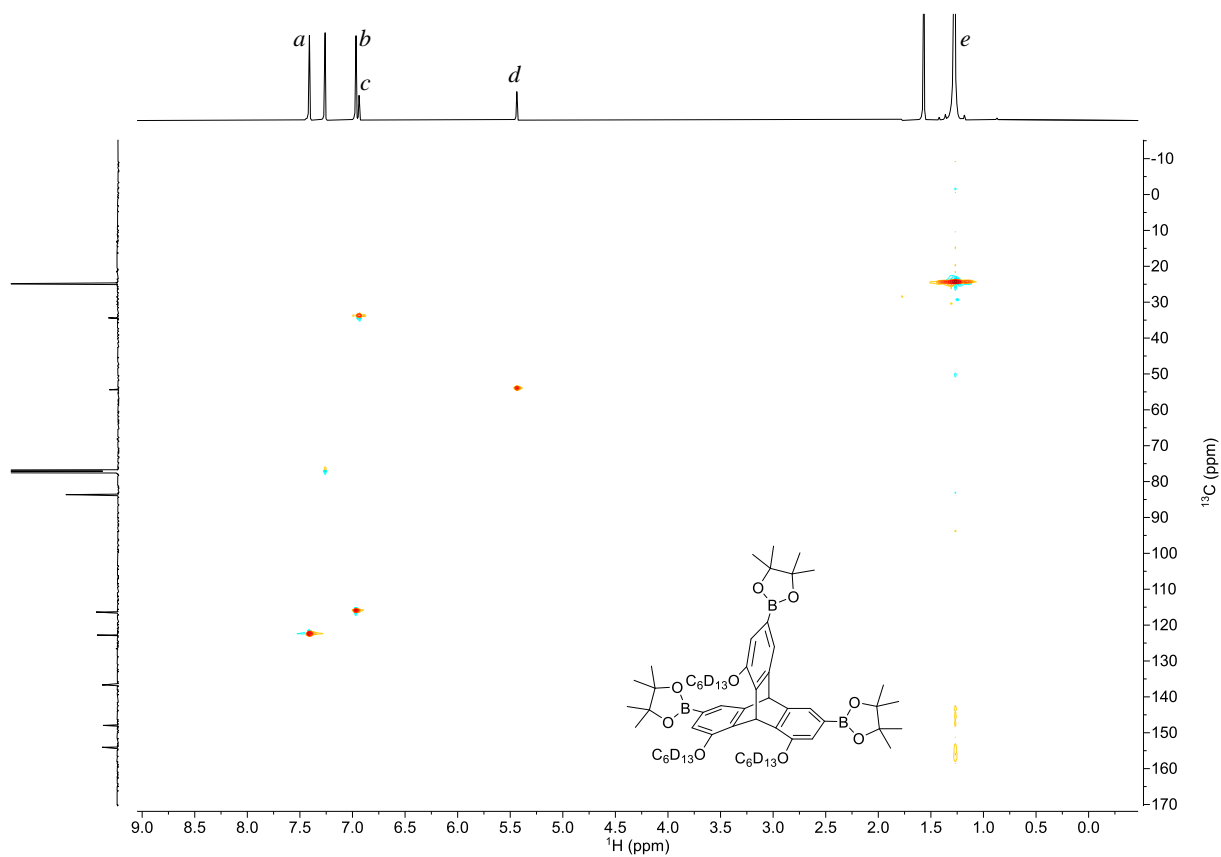


Figure S22. $^1\text{H},^{13}\text{C}$ -HSQC spectrum of compound **6-d₃₉** (700 MHz, 176 MHz, CDCl_3).

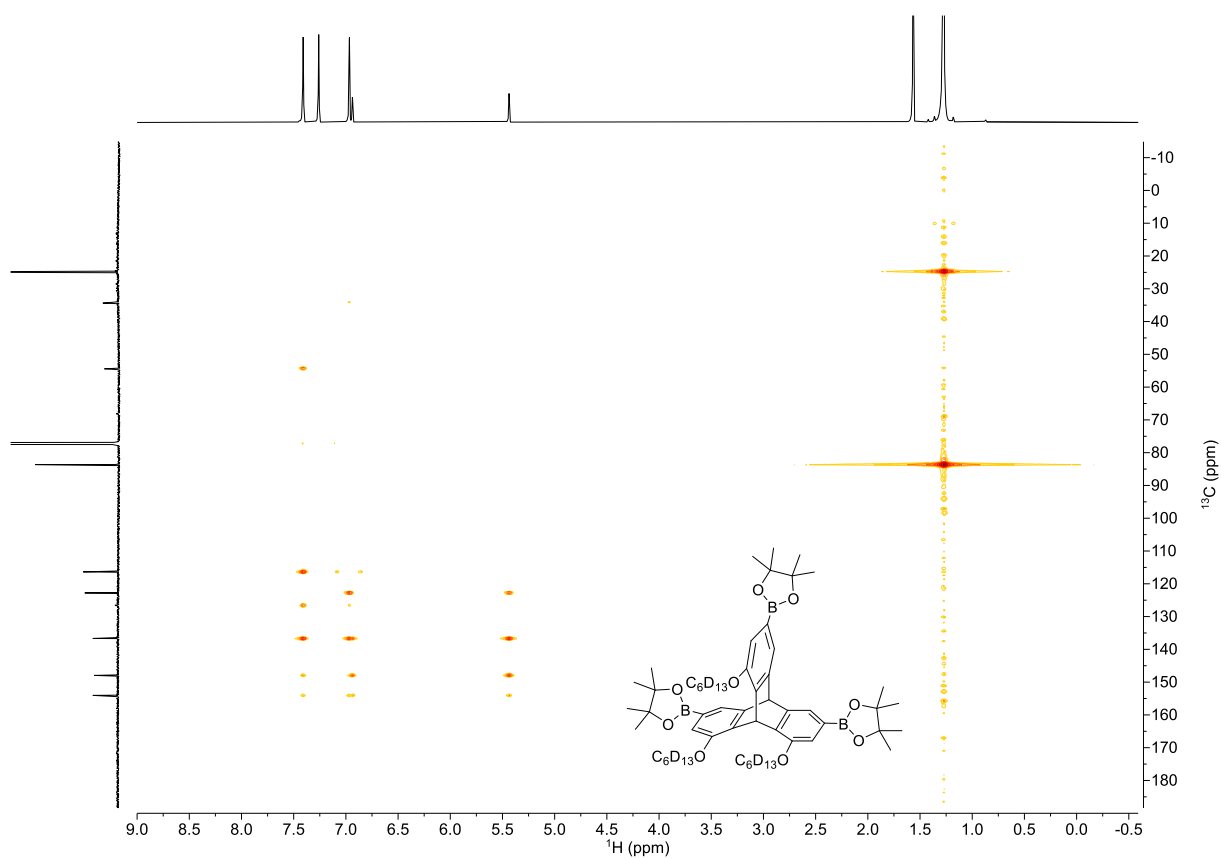


Figure S23. $^1\text{H},^{13}\text{C}$ -HMBC spectrum of compound **6-d₃₉** (700 MHz, 176 MHz, CDCl_3).

Table S7. Correlations of ^1H , ^{13}C -HSQC and ^1H , ^{13}C -HMBC spectra of compound **6-d₃₉**.

^{13}C nucleus	$\delta_{13\text{C}}$ [ppm]	^1H nucleus		
		HSQC	$\delta_{1\text{H}}$ [ppm]	HMBC
A	154.0	-	-	<i>a, b, c, d</i>
B	148.0	-	-	<i>a, c, d</i>
C	136.7	-	-	<i>a, b, c, d</i>
D	126.5	-	-	<i>a, b</i>
E	122.8	<i>a</i>	7.41	<i>b, d</i>
F	116.4	<i>b</i>	6.97	<i>a</i>
G	83.6	-	-	<i>e</i>
H	54.4	<i>d</i>	5.44	<i>a</i>
I	34.4	<i>c</i>	6.94	<i>b</i>
J	24.9	<i>e</i>	1.27	<i>e</i>

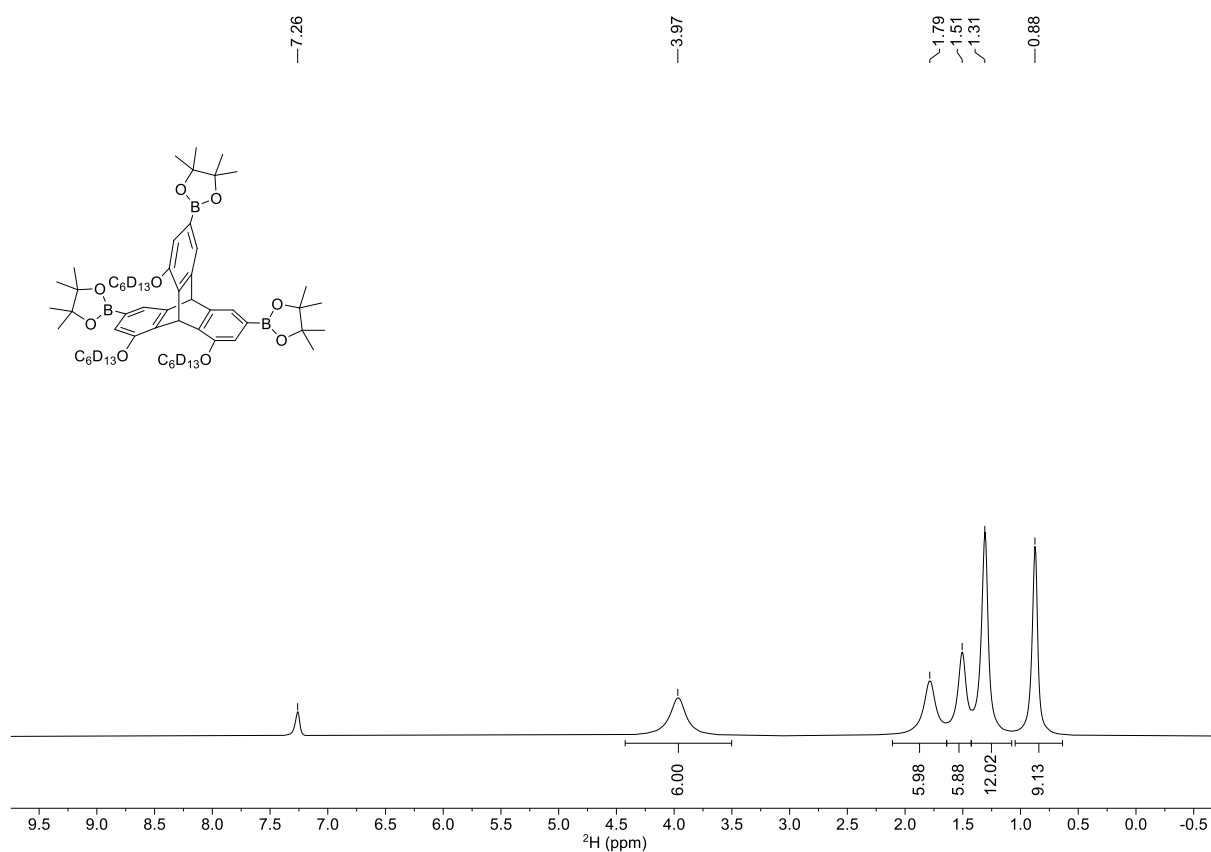


Figure S24. ^1H NMR spectrum of compound **6-d₃₉** (92 MHz, CHCl_3).

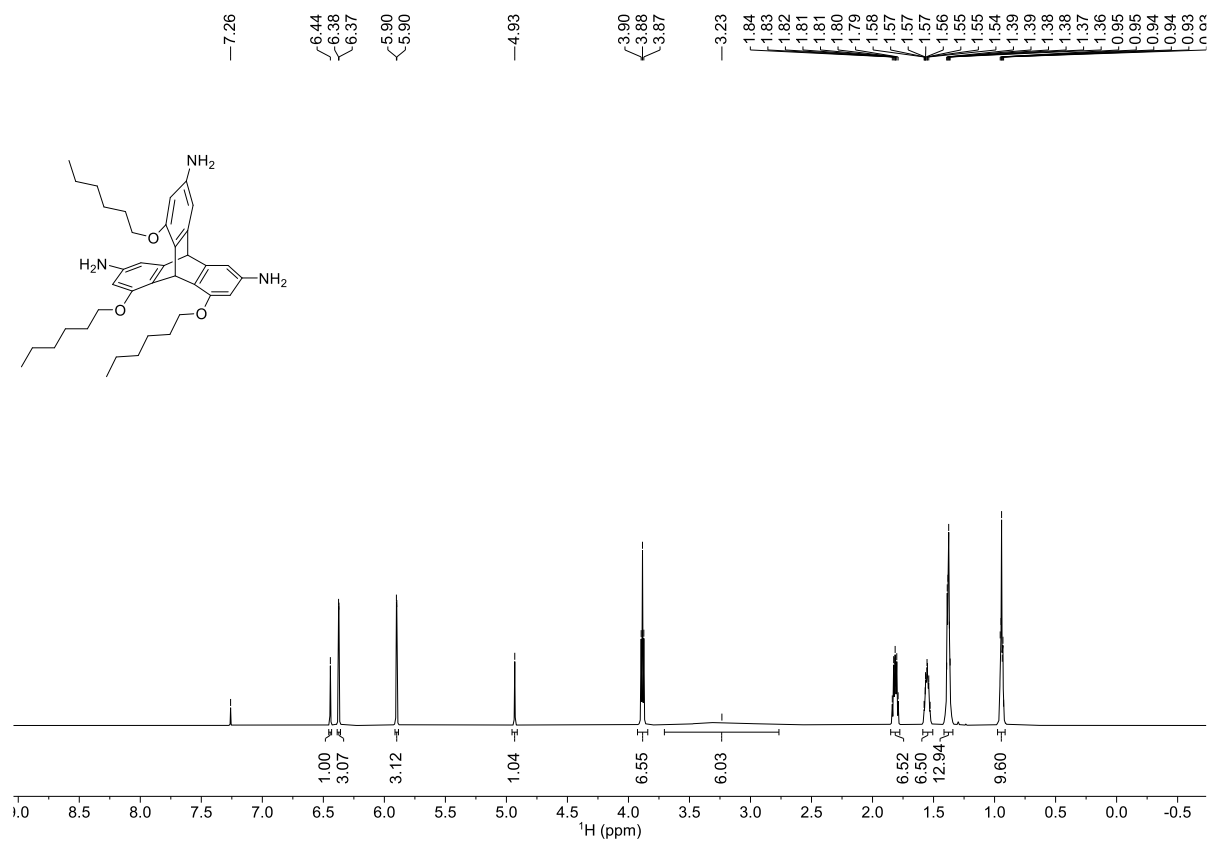


Figure S25. ^1H NMR spectrum of compound **7a** (600 MHz, CDCl_3).

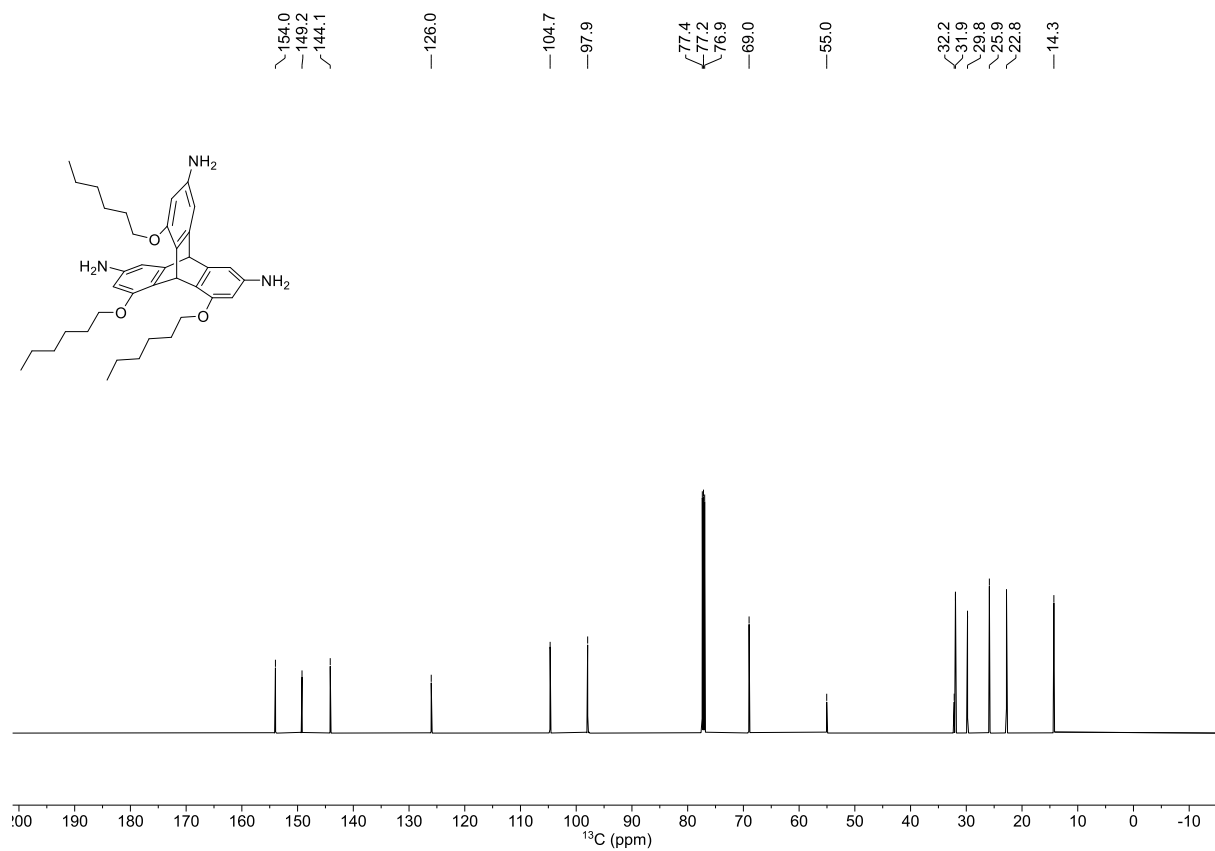


Figure S26. $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of compound **7a** (150 MHz, CDCl_3).

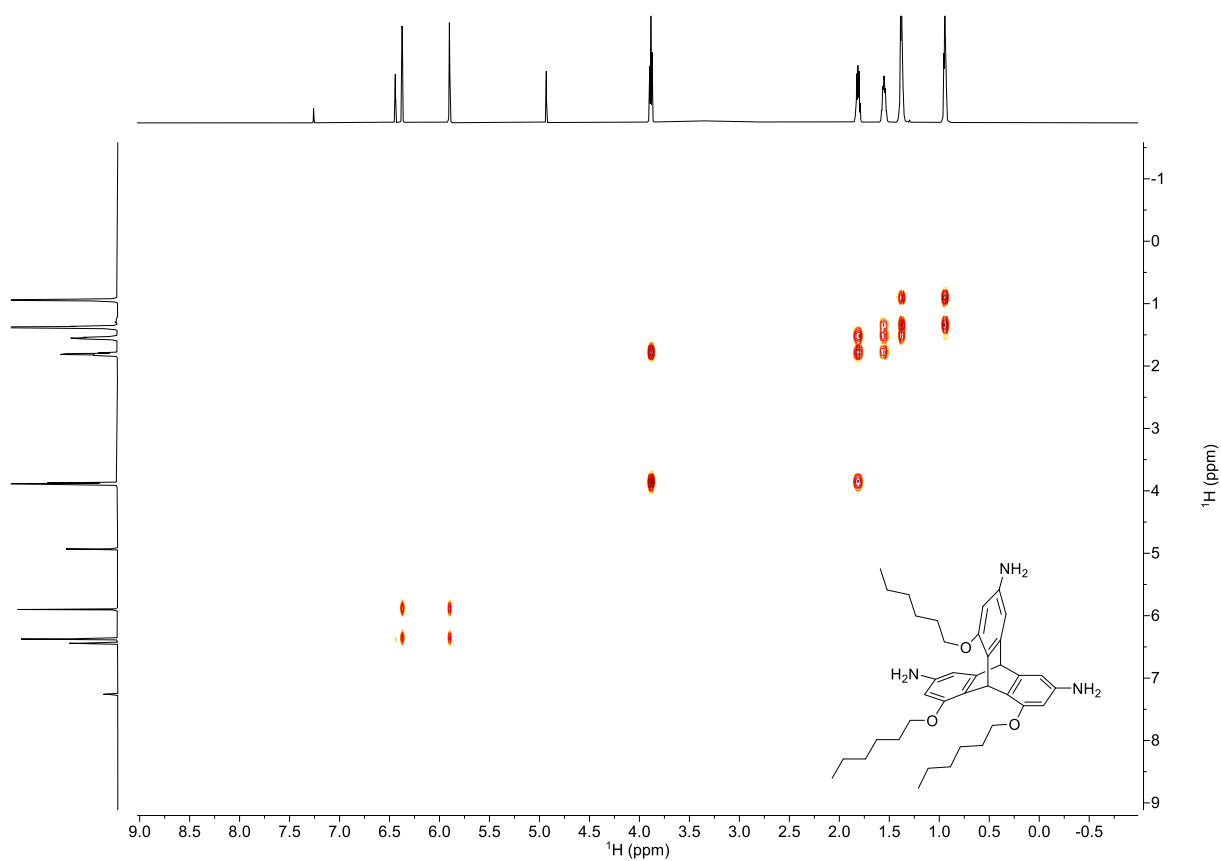


Figure S27. $^1\text{H}, ^1\text{H}$ -COSY spectrum of compound **7a** (600 MHz, CDCl_3).

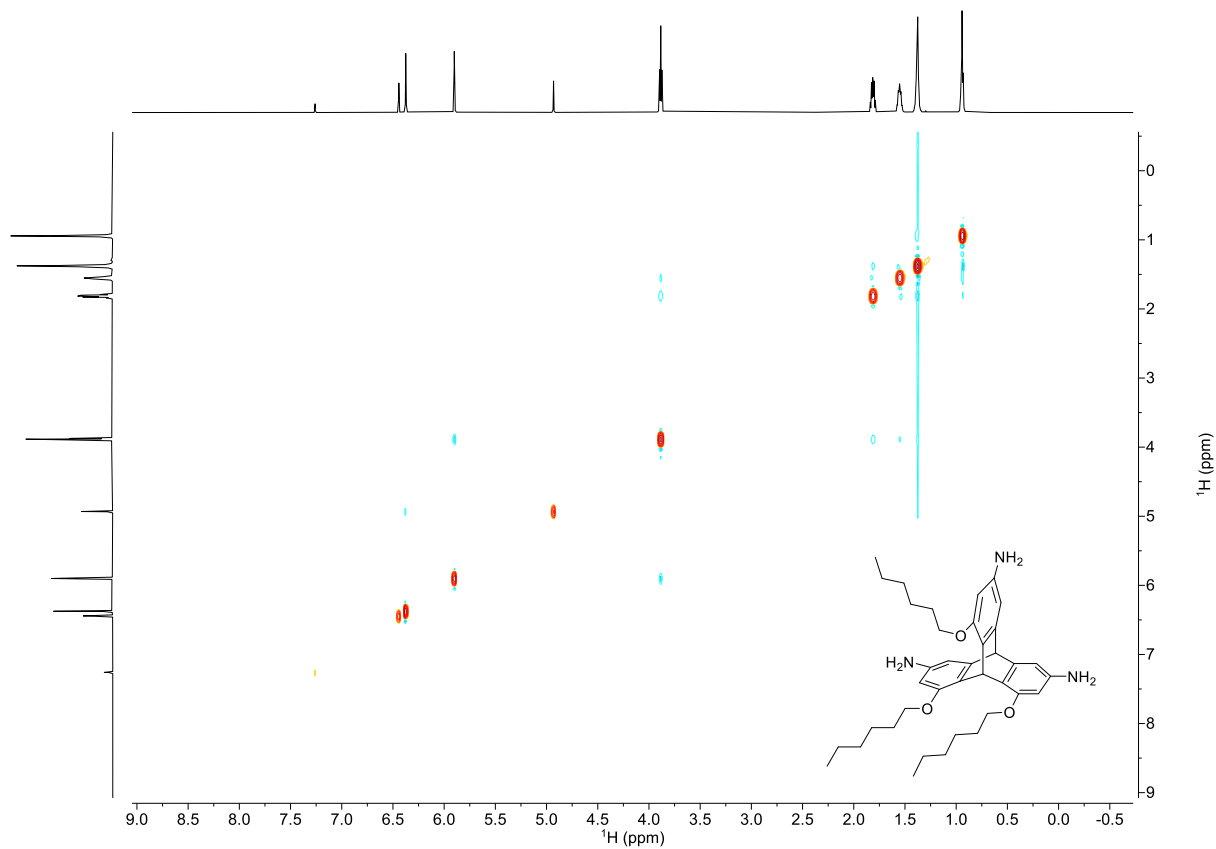


Figure S28. $^1\text{H}, ^1\text{H}$ -NOESY spectrum of compound **7a** (600 MHz, CDCl_3).

Table S8. Cross-peak assignment of ^1H , ^1H -COSY spectrum of compound **7a**.

^1H nucleus	$\delta_{1\text{H}}$ [ppm]	COSY
<i>a</i>	6.45	-
<i>b</i>	6.38	<i>c, d</i>
<i>c</i>	5.90	<i>b, d</i>
<i>d</i>	4.94	-
<i>e</i>	3.88	<i>g</i>
<i>f</i>	3.40	-
<i>g</i>	1.86 – 1.74	<i>e, h, i</i>
<i>h</i>	1.60 – 1.47	<i>g, i</i>
<i>i</i>	1.42 – 1.30	<i>g, h, j</i>
<i>j</i>	0.97 – 0.88	<i>i</i>

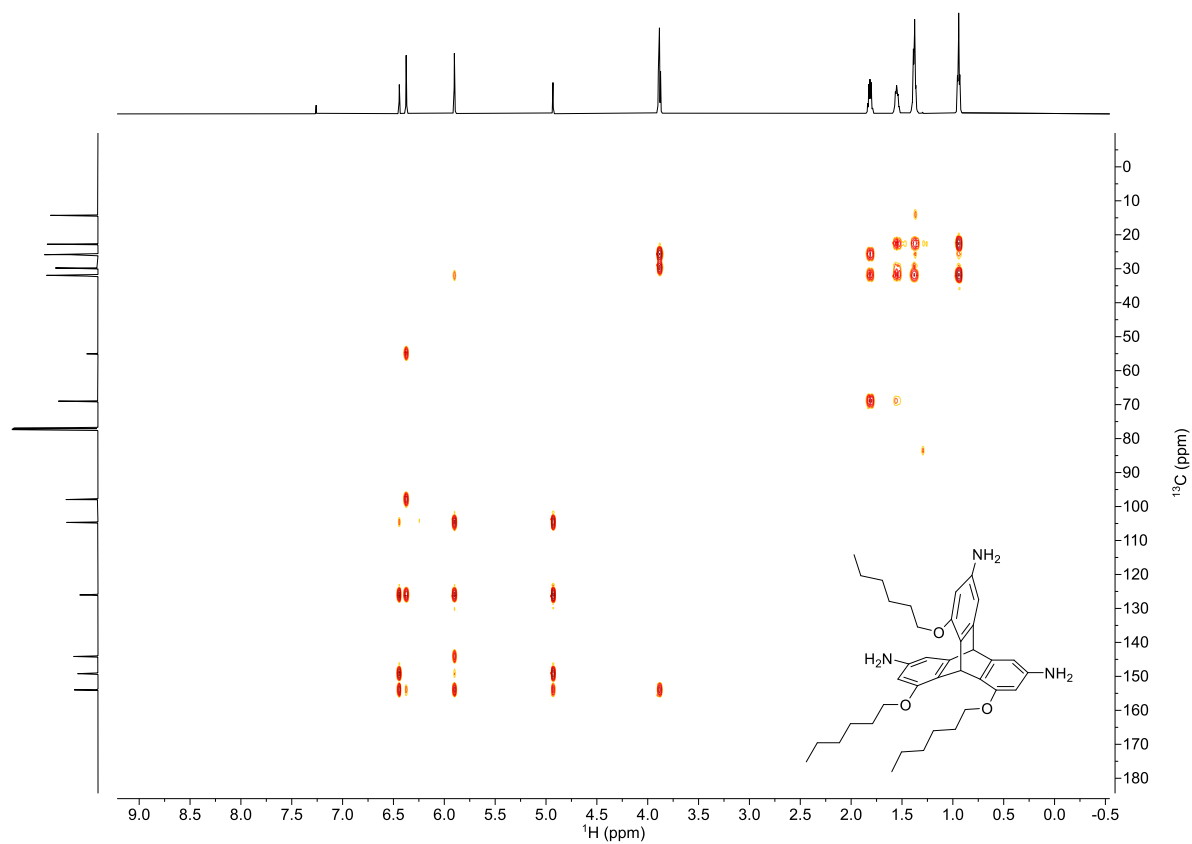
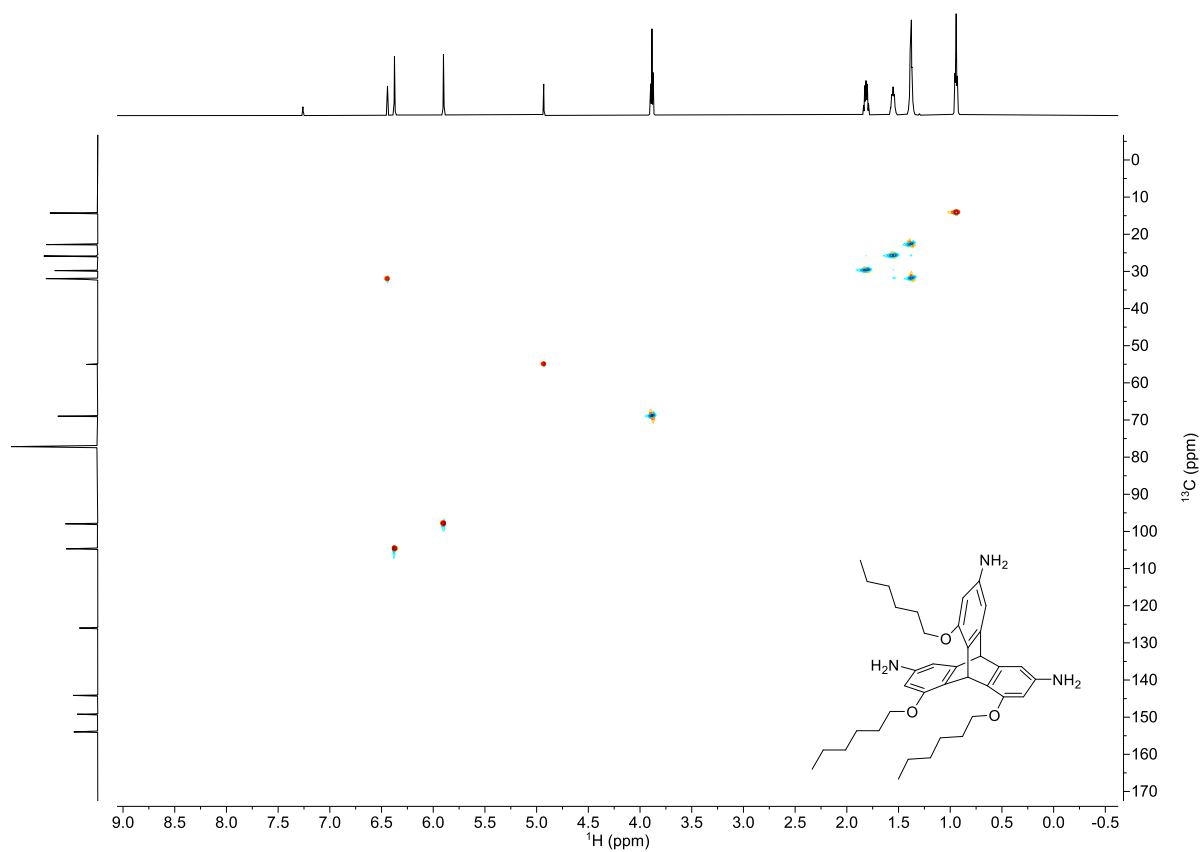


Table S9. Correlations of ^1H , ^{13}C -HSQC and ^1H , ^{13}C -HMBC spectra of compound **7a**.

^{13}C nucleus	$\delta_{13\text{C}}$ [ppm]	^1H nucleus		
		HSQC	$\delta_{1\text{H}}$ [ppm]	HMBC
<i>A</i>	154.0	-	-	<i>a, c, e</i>
<i>B</i>	149.2	-	-	<i>a, d</i>
<i>C</i>	144.1	-	-	<i>c</i>
<i>D</i>	126.1	-	-	<i>a, b, c, d</i>
<i>E</i>	104.7	<i>b</i>	6.38	<i>c, d</i>
<i>F</i>	98.0	<i>c</i>	5.90	<i>b</i>
<i>G</i>	69.0	<i>e</i>	3.88	<i>g, h</i>
<i>H</i>	55.1	<i>d</i>	4.94	<i>b</i>
<i>I</i>	32.2	<i>a</i>	6.45	-
<i>J</i>	32.0	<i>i</i>	1.42 – 1.30	<i>g, i, j</i>
<i>K</i>	29.8	<i>g</i>	1.86 – 1.74	<i>e</i>
<i>L</i>	25.9	<i>h</i>	1.60 – 1.47	<i>e, g</i>
<i>M</i>	22.8	<i>i</i>	1.42 – 1.30	<i>h, i, j</i>
<i>O</i>	14.3	<i>j</i>	0.97 – 0.88	-

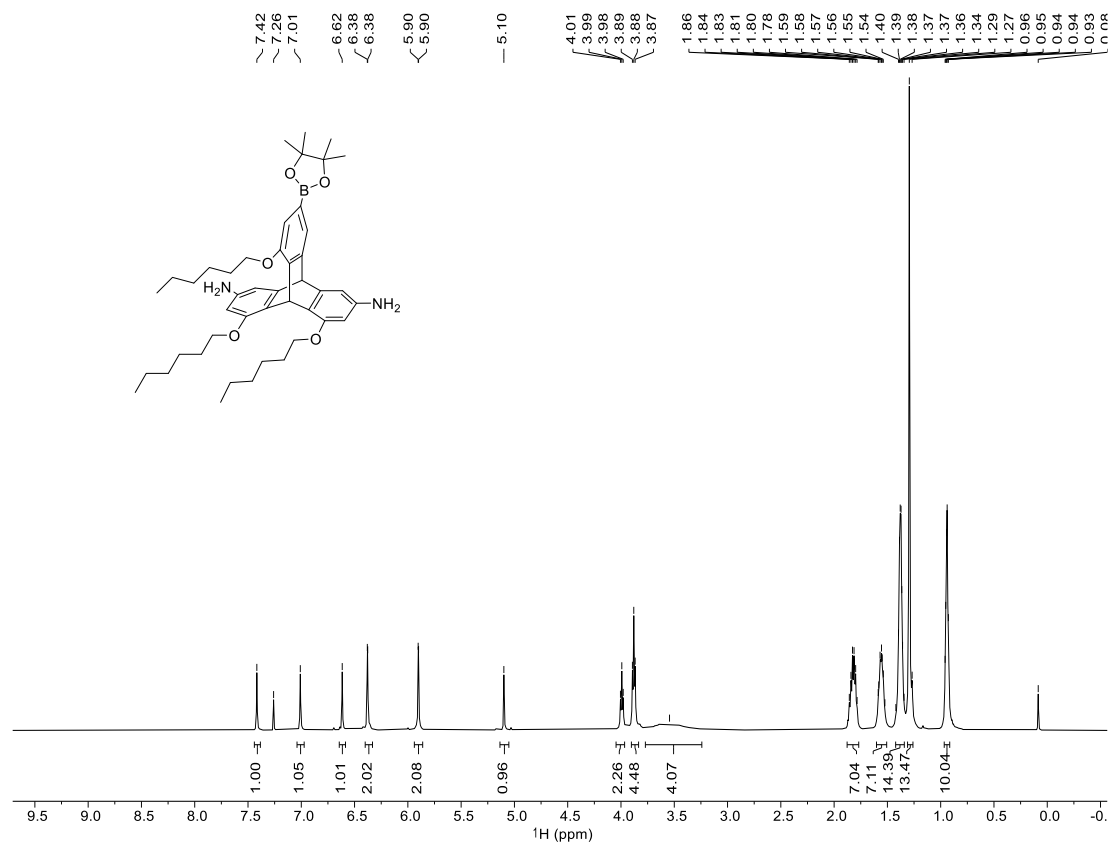


Figure S31. ^1H NMR spectrum of compound **7b** (500 MHz, CDCl_3).

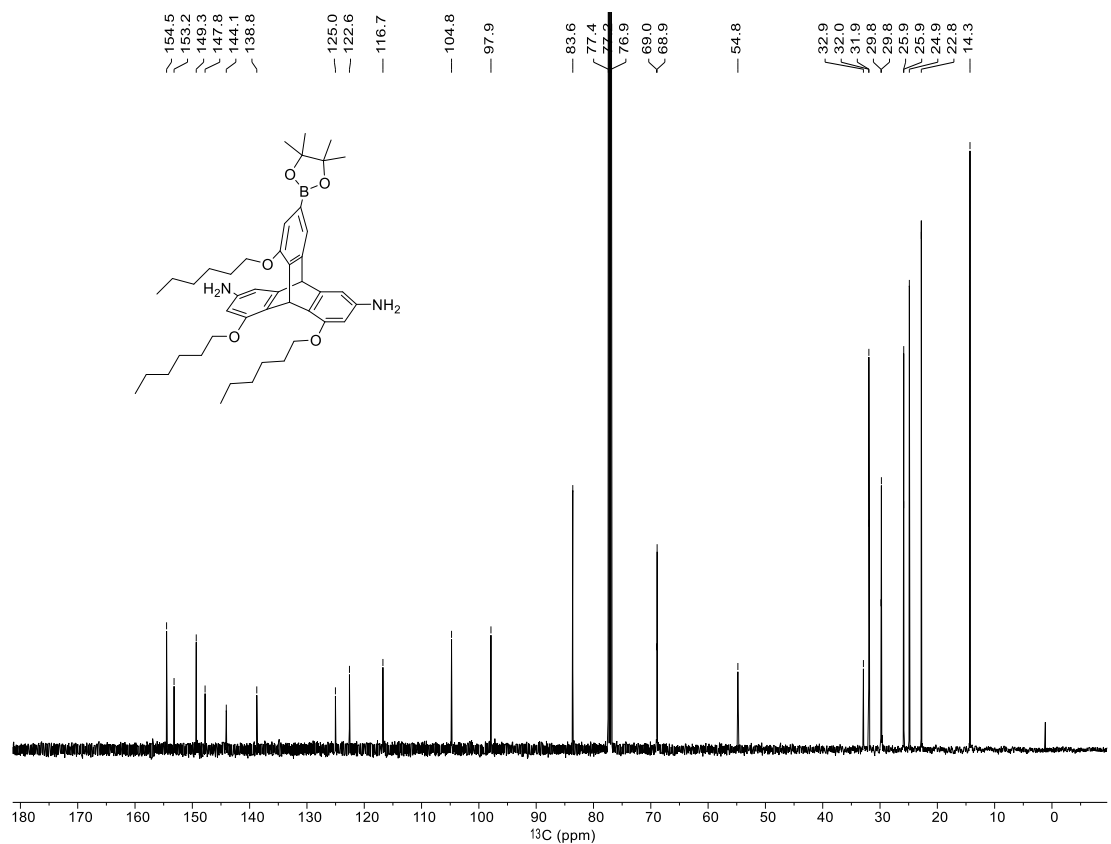


Figure S32. $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of compound **7b** (125 MHz, CDCl_3).

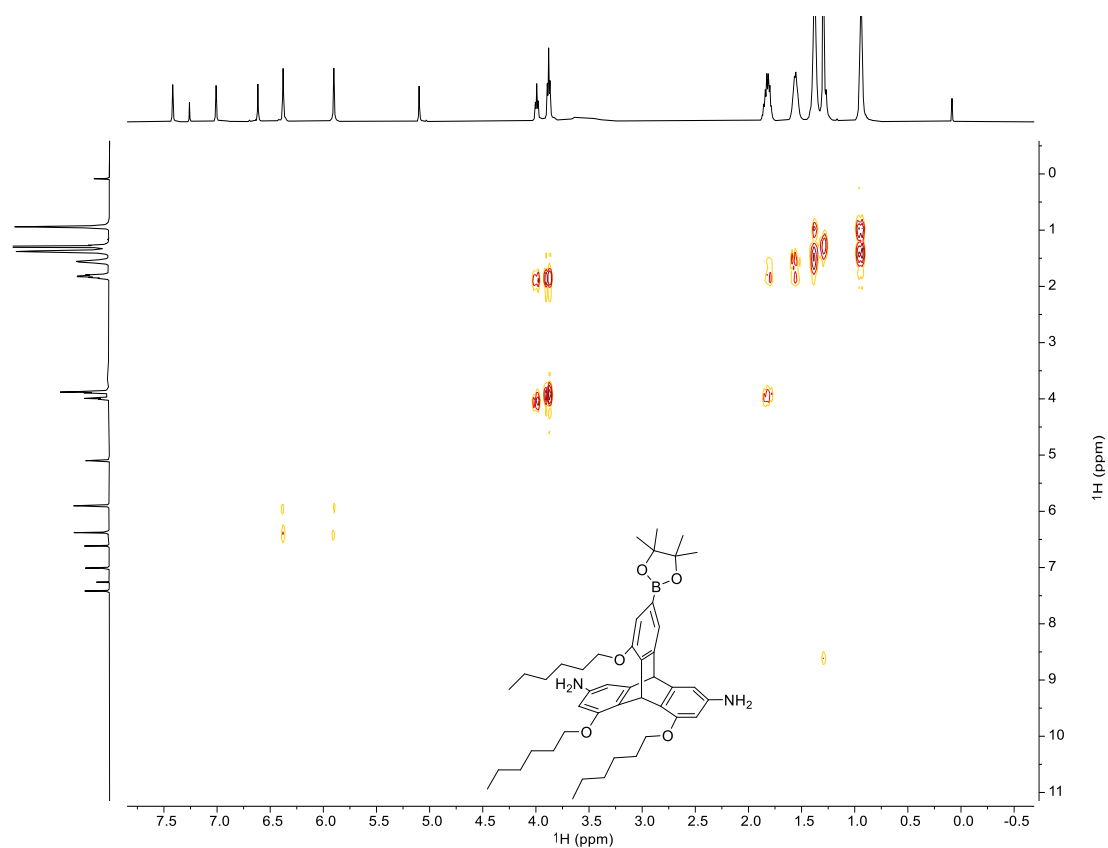


Figure S33. $^1\text{H}, ^1\text{H}$ -COSY spectrum of compound **7b** (500 MHz, 125 MHz, CDCl_3).

Table S10. Cross-peak assignment of $^1\text{H}, ^1\text{H}$ -COSY spectrum of compound **7b**.

^1H nucleus	$\delta_{1\text{H}}$ [ppm]	COSY
<i>a</i>	7.42	-
<i>b</i>	7.01	-
<i>c</i>	6.62	-
<i>d</i>	6.38	<i>e</i>
<i>e</i>	5.90	<i>d</i>
<i>f</i>	5.10	-
<i>g</i>	3.99	<i>j</i>
<i>h</i>	3.88	<i>j</i>
<i>i</i>	3.54	-
<i>j</i>	1.88 – 1.77	<i>g, h, k, l, n</i>
<i>k</i>	1.60 – 1.51	<i>j, l, n</i>
<i>l</i>	1.42 – 1.34	<i>j, k, n</i>
<i>m</i>	1.29	-
<i>n</i>	0.97 – 0.92	<i>j, k, l</i>

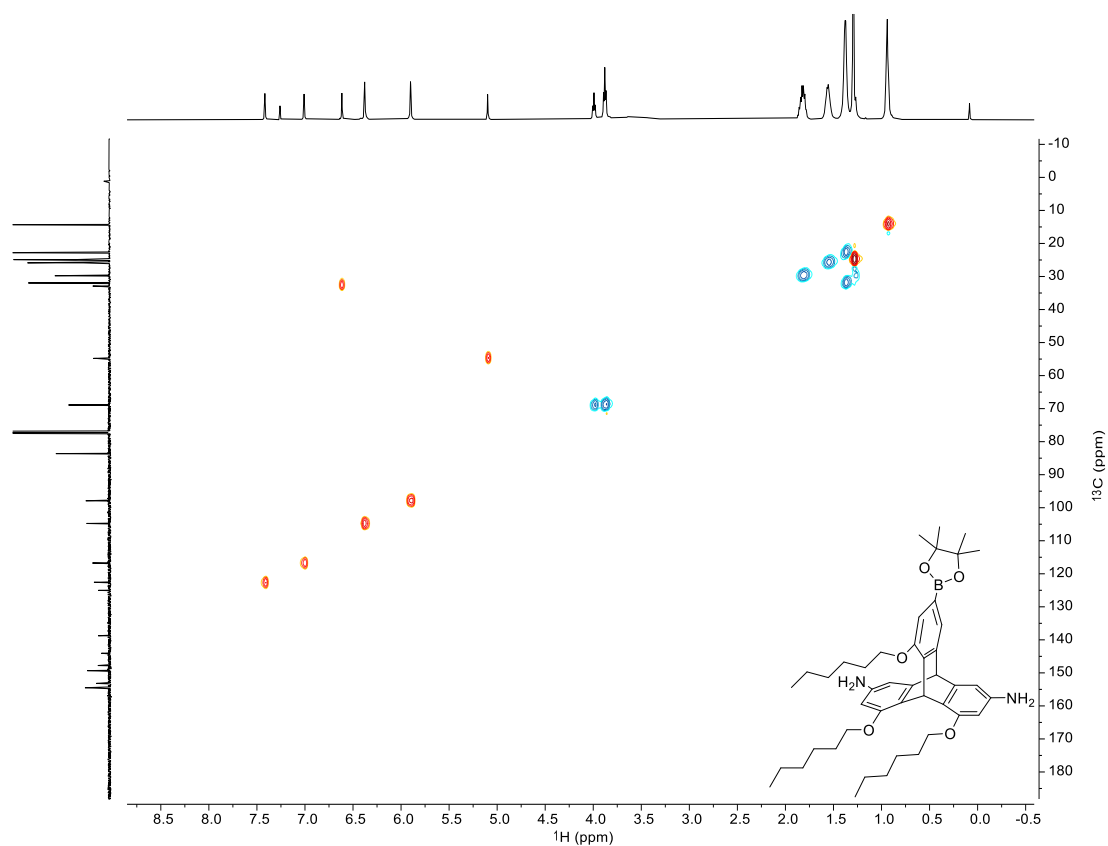


Figure S34. ^1H , ^{13}C -HSQC spectrum of compound **7b** (500 MHz, 125 MHz, CDCl_3).

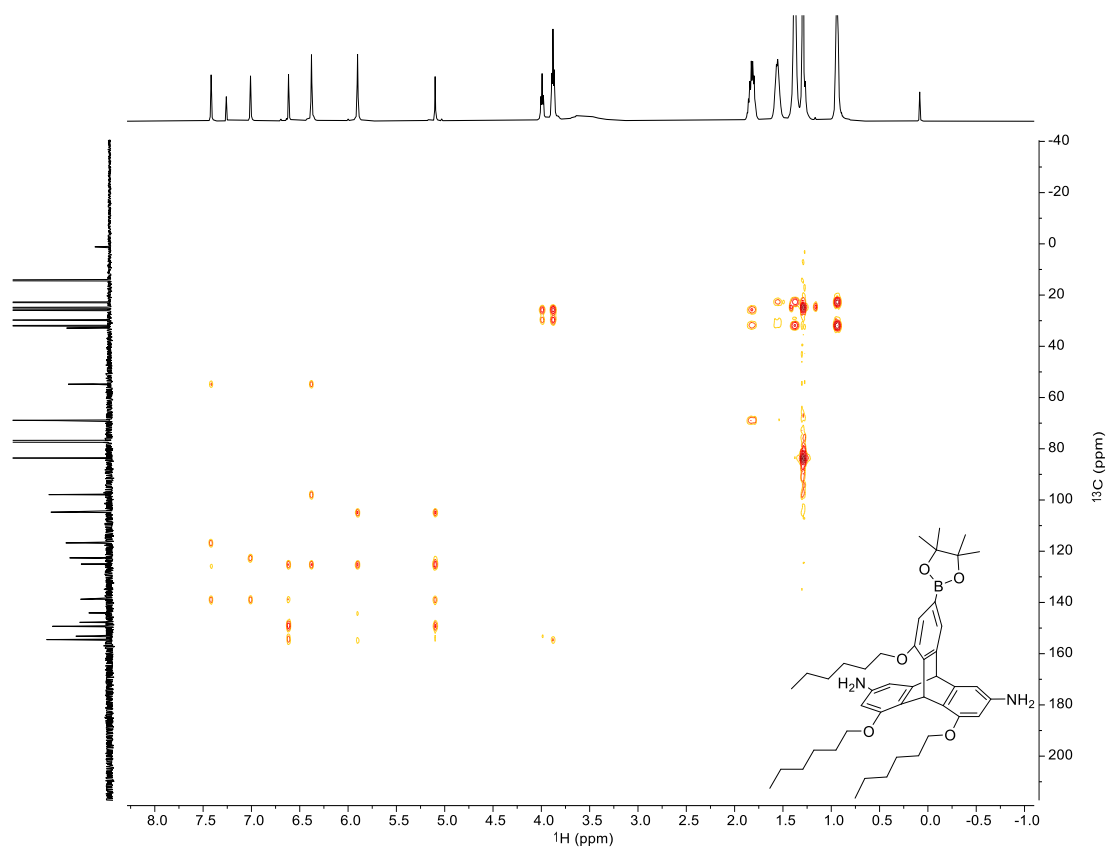


Figure S35. ^1H , ^{13}C -HMBC spectrum of compound **7b** (500 MHz, 125 MHz, CDCl_3).

Table S11. Correlations of ^1H , ^{13}C -HSQC and ^1H , ^{13}C -HMBC spectra of compound **7b**.

^{13}C nucleus	$\delta_{13\text{C}}$ [ppm]	^1H nucleus		
		HSQC	$\delta_{1\text{H}}$ [ppm]	HMBC
A	154.5	-	-	<i>c, e, f, h</i>
B	153.2	-	-	<i>b, f, g</i>
C	149.3	-	-	<i>c, f</i>
D	147.8	-	-	<i>c, f</i>
E	144.1	-	-	<i>e</i>
F	138.8	-	-	<i>a, b, c, f</i>
G	125.0	-	-	<i>c, d, e, f</i>
H	122.6	<i>a</i>	7.42	<i>b</i>
I	116.7	<i>b</i>	7.01	<i>a</i>
J	104.8	<i>d</i>	6.38	<i>e, f</i>
K	97.9	<i>e</i>	5.90	<i>d</i>
L	83.6	-	-	<i>m</i>
M	69.0	<i>g</i>	3.99	<i>j, k</i>
O	68.9	<i>h</i>	3.88	<i>j, k</i>
P	54.8	<i>f</i>	5.10	<i>a, d</i>
Q	32.9	<i>c</i>	6.62	-
R	32.0	<i>l</i>	1.42 – 1.34	<i>j, k, l, n</i>
S	31.9	<i>l</i>	1.42 – 1.34	<i>j, k, l, n</i>
T	29.8	<i>j</i>	1.88 – 1.77	<i>g, h, k</i>
U	29.8	<i>j</i>	1.88 – 1.77	<i>g, h, k</i>
V	25.9	<i>k</i>	1.60 – 1.51	<i>g, h, j</i>
W	25.9	<i>k</i>	1.60 – 1.51	<i>g, h, j</i>
X	24.9	<i>m</i>	1.29	<i>m</i>
Y	22.8	<i>l</i>	1.42 – 1.34	<i>k, l, n</i>
Z	14.3	<i>n</i>	0.97 – 0.92	-

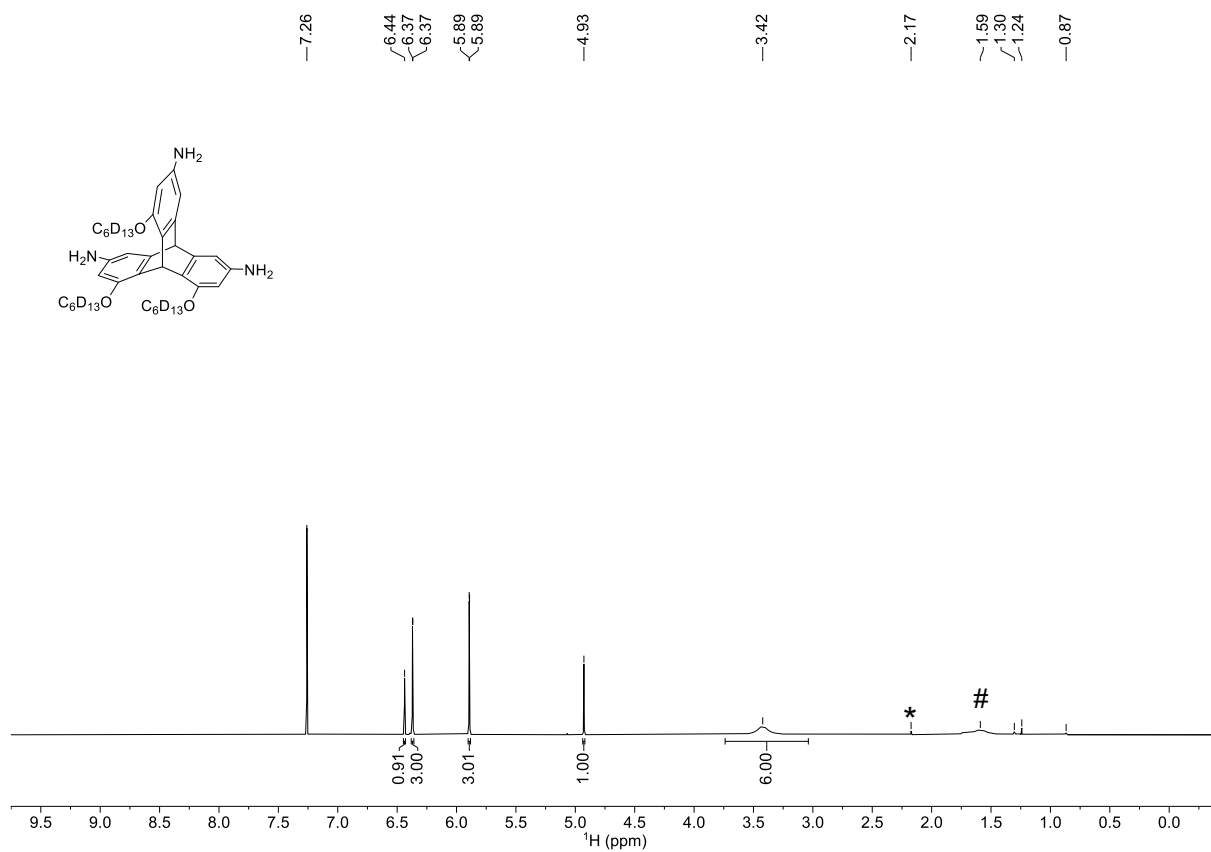


Figure S36. 1H NMR spectrum of compound **7a-d₃₉** (600 MHz, $CDCl_3$). #: H_2O . *: Acetone.

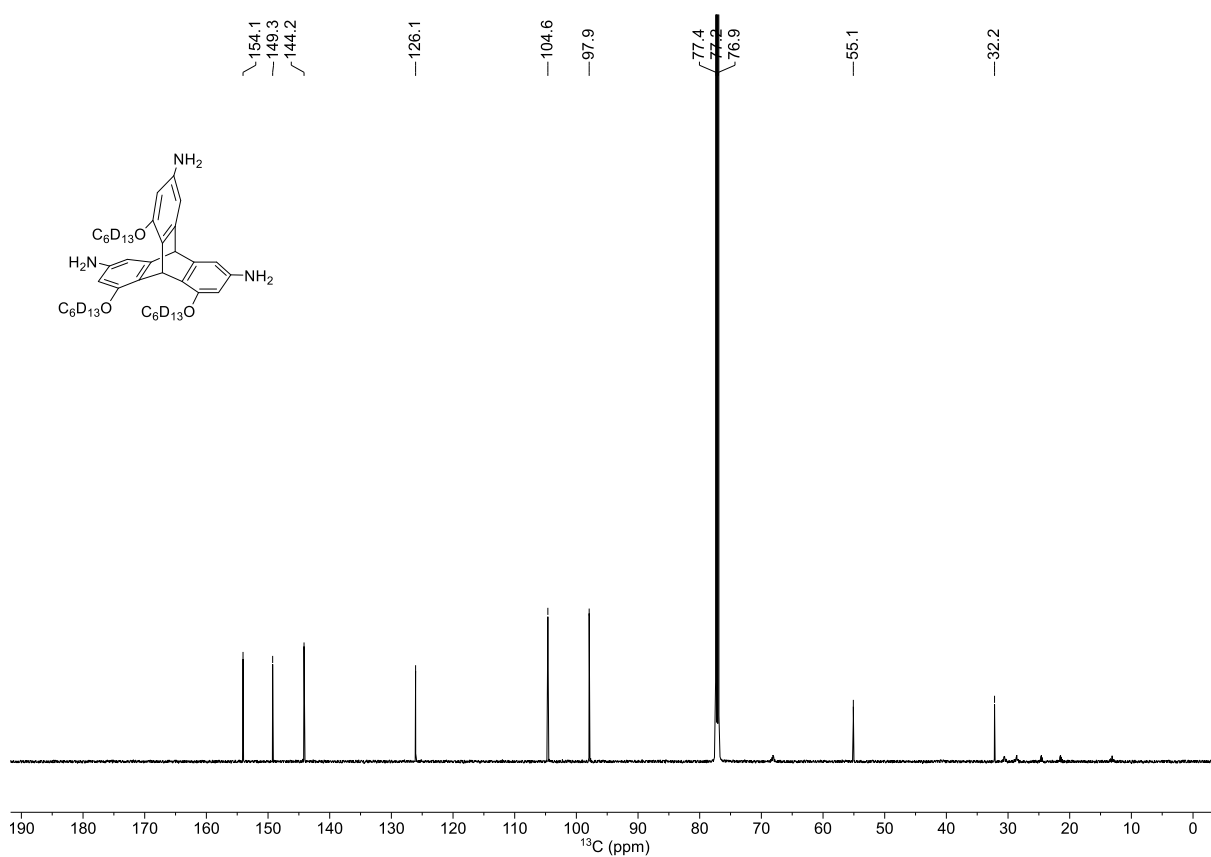


Figure S37. $^{13}C\{^1H\}$ NMR spectrum of compound **7a-d₃₉** (150 MHz, $CDCl_3$).

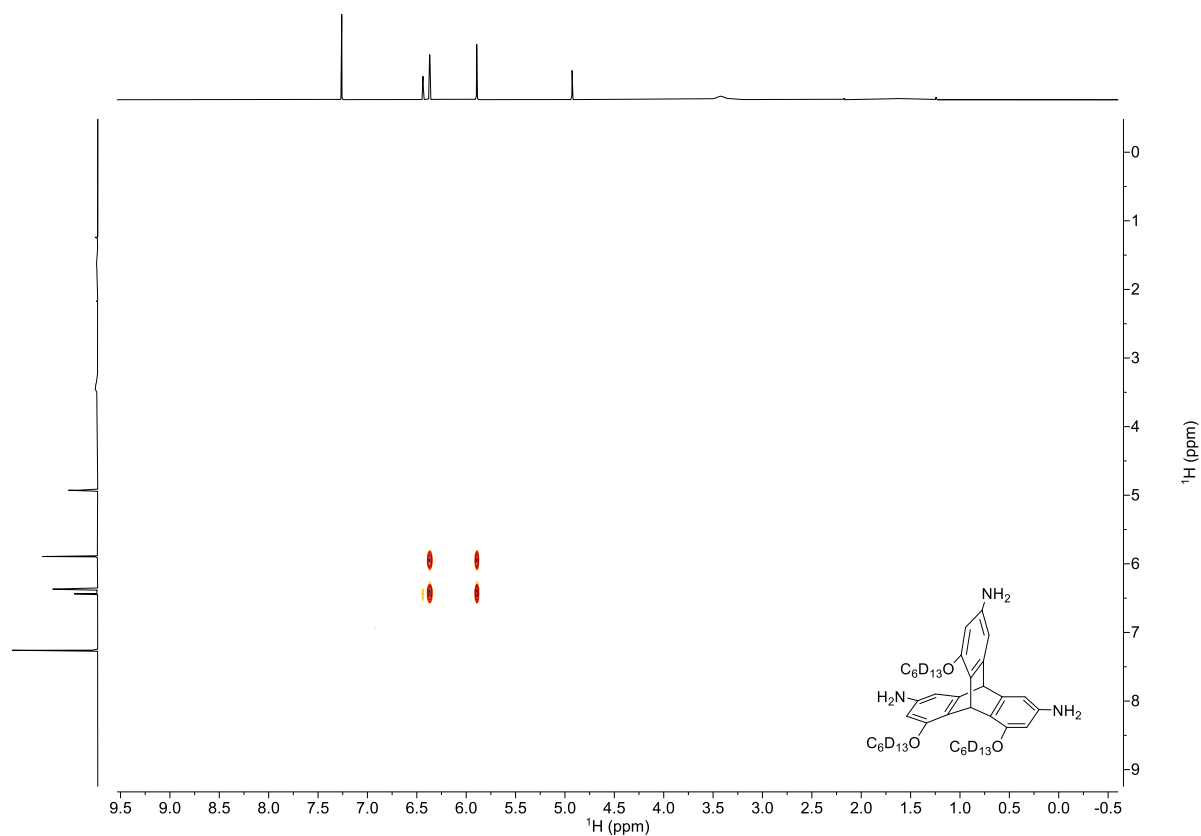


Figure S38. $^1\text{H}, ^1\text{H}$ -COSY spectrum of compound **7a-d₃₉** (600 MHz, CDCl_3).

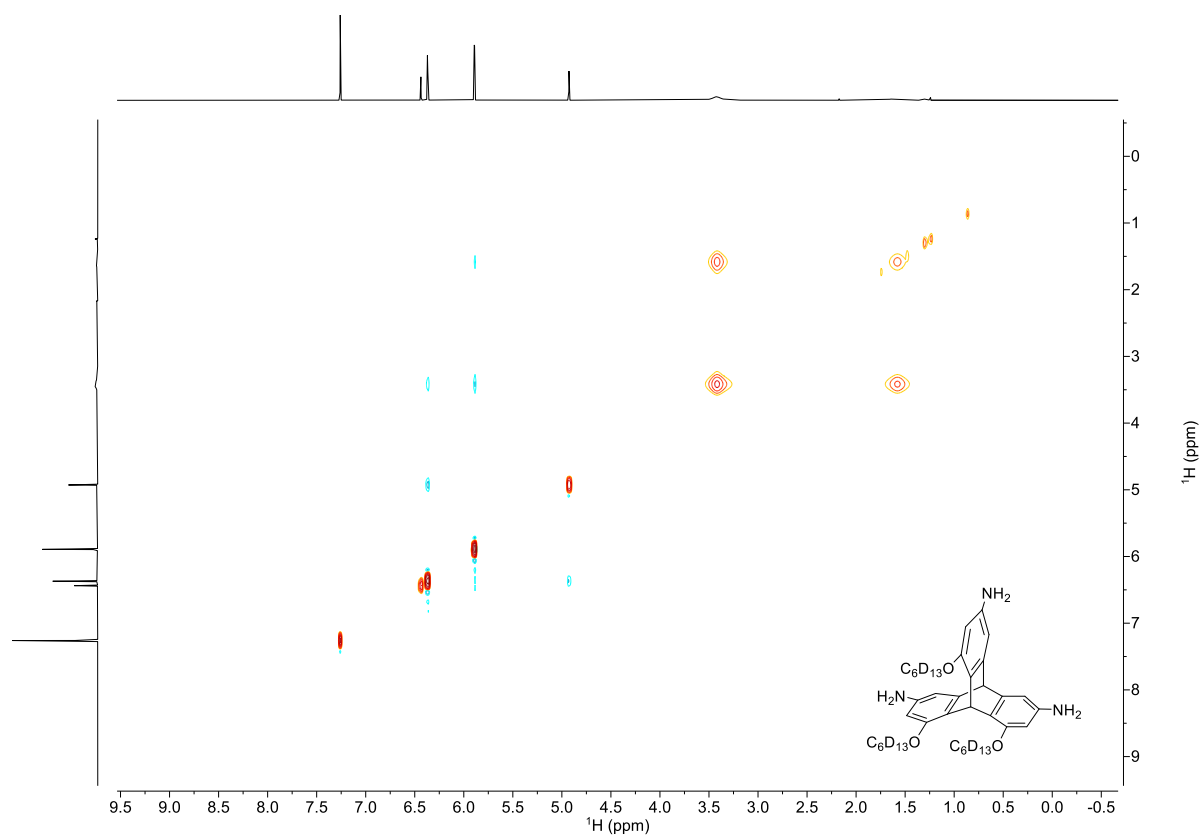


Figure S39. $^1\text{H}, ^1\text{H}$ -NOESY spectrum of compound **7a-d₃₉** (600 MHz, CDCl_3).

Table S12. Cross-peak assignment of ^1H , ^1H -COSY and ^1H , ^1H -NOESY spectrum of compound **7a-d**₃₉.

^1H nucleus	$\delta_{1\text{H}}$ [ppm]	COSY	NOESY
<i>a</i>	6.44	-	-
<i>b</i>	6.37	<i>c</i>	<i>d</i> , <i>e</i>
<i>c</i>	5.89	<i>b</i>	<i>e</i> , H ₂ O
<i>d</i>	4.93	-	<i>b</i>
<i>e</i>	3.42	-	H ₂ O

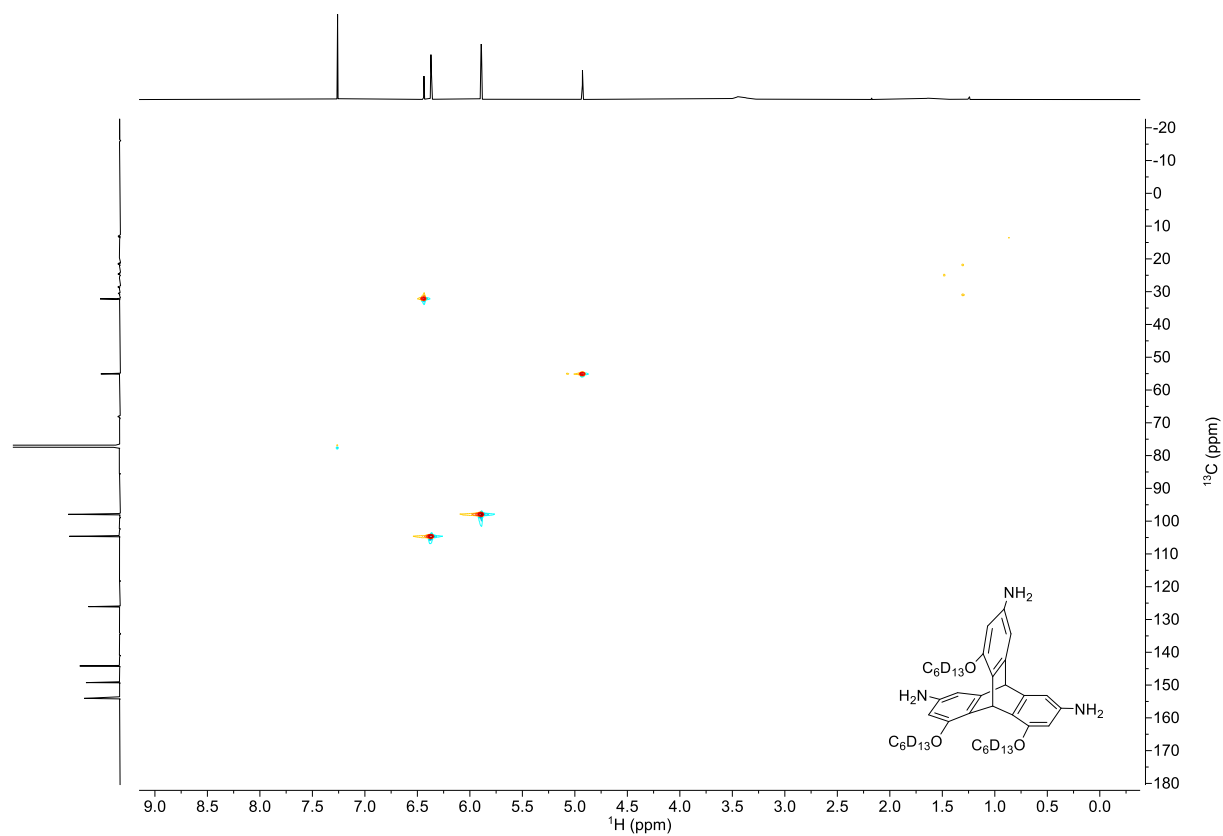


Figure S40. ^1H , ^{13}C -HSQC spectrum of compound **7a-d₃₉** (600 MHz, 150 MHz, CDCl_3).

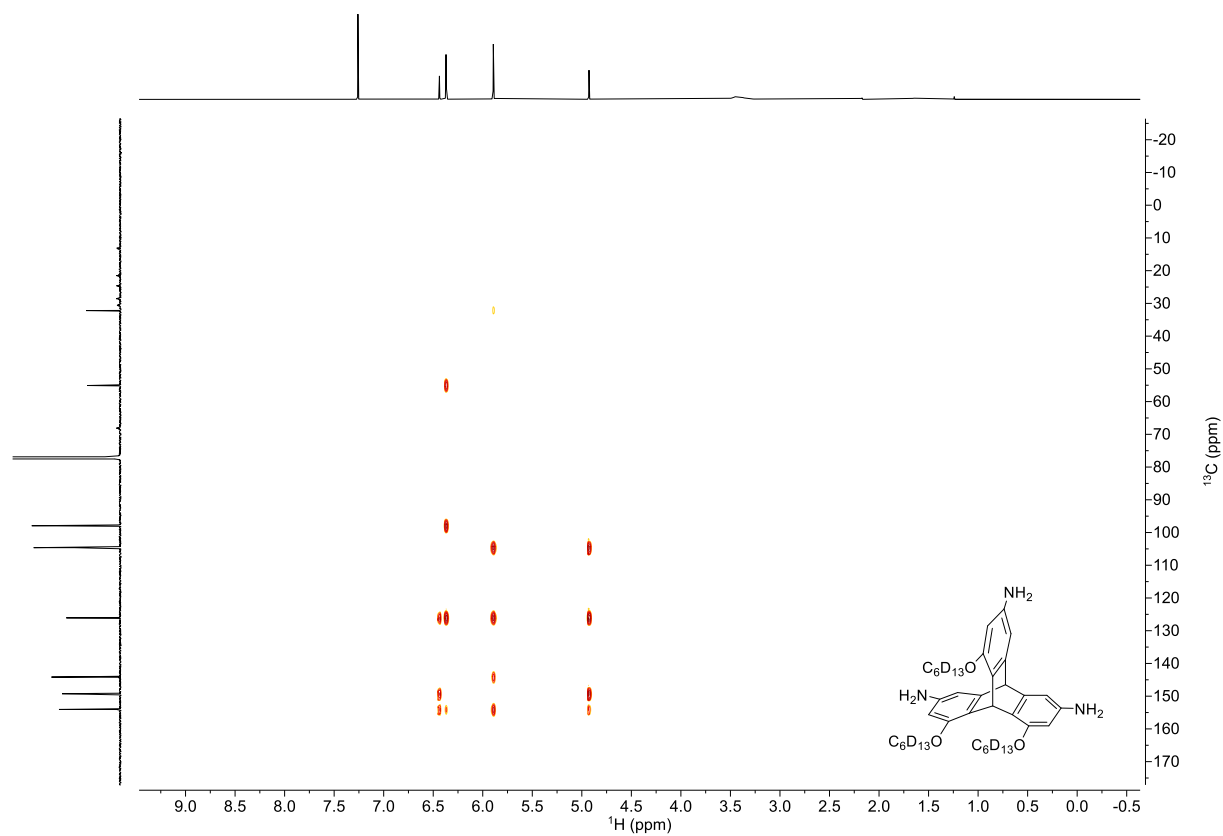


Figure S41. ^1H , ^{13}C -HMBC spectrum of compound **7a-d₃₉** (600 MHz, 150 MHz, CDCl_3).

Table S13. Correlations of ^1H , ^{13}C -HSQC and ^1H , ^{13}C -HMBC spectra of compound **7a-d**₃₉.

^{13}C nucleus	$\delta_{13\text{C}}$ [ppm]	^1H nucleus		
		HSQC	$\delta_{1\text{H}}$ [ppm]	HMBC
A	154.1	-	-	<i>a, b, c, d</i>
B	149.3	-	-	<i>a, d</i>
C	144.2	-	-	<i>c</i>
D	126.1	-	-	<i>a, b, c, d</i>
E	104.6	<i>b</i>	6.37	<i>c, d</i>
F	97.9	<i>c</i>	5.89	<i>b</i>
G	55.1	<i>d</i>	4.93	<i>b</i>
H	32.2	<i>a</i>	6.44	<i>c</i>

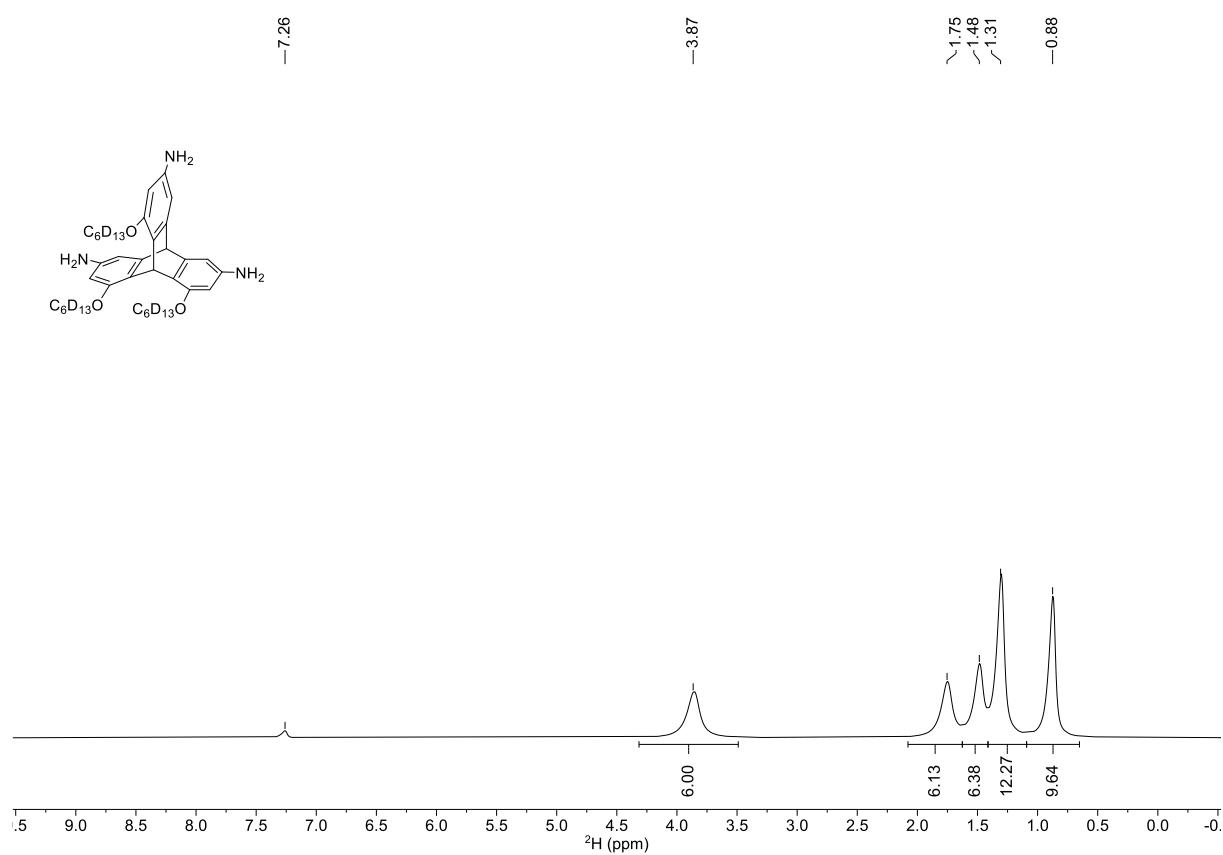


Figure S42. ^2H NMR spectrum of compound **7a-d**₃₉ (92 MHz, CHCl_3).

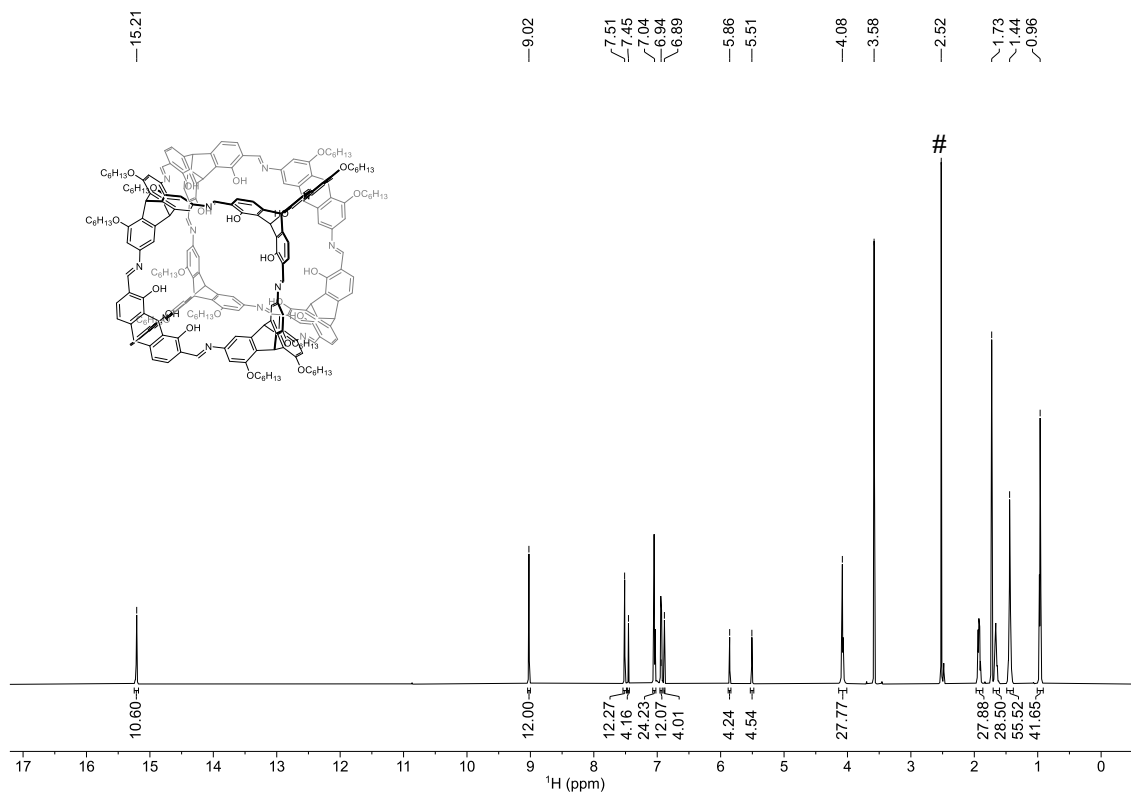


Figure S43. ^1H NMR spectrum (600 MHz, THF-d_8) of [4+4] cube 11. #: H_2O .

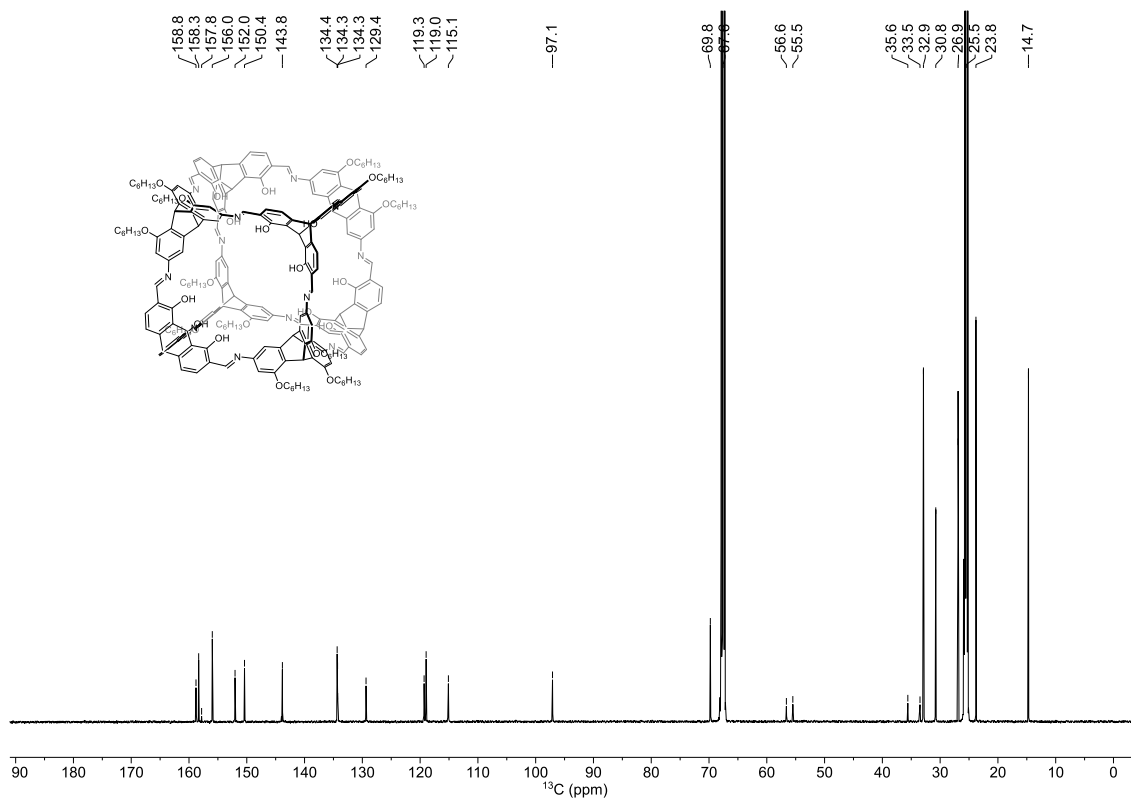


Figure S44. $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (150 MHz, THF-d_8) of [4+4] cube 11.

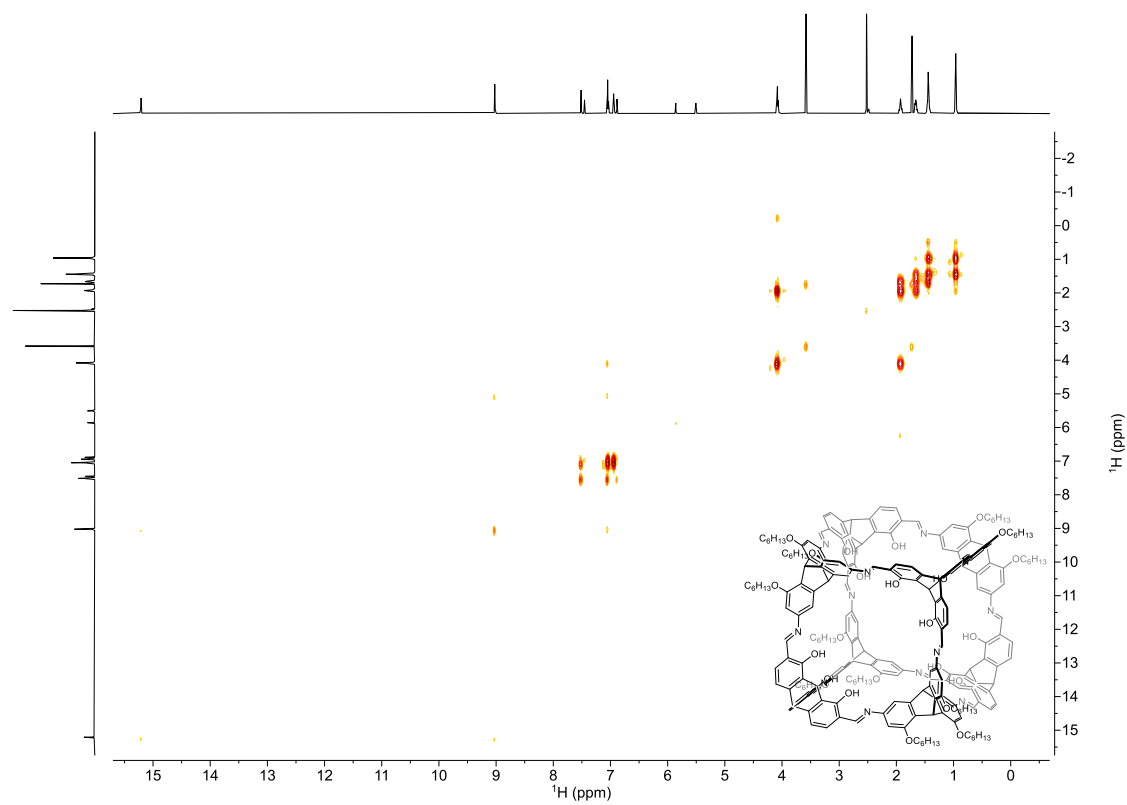


Figure S45. $^1\text{H}, ^1\text{H}$ COSY spectrum (600 MHz, THF-d_8) of [4+4] cube **11**.

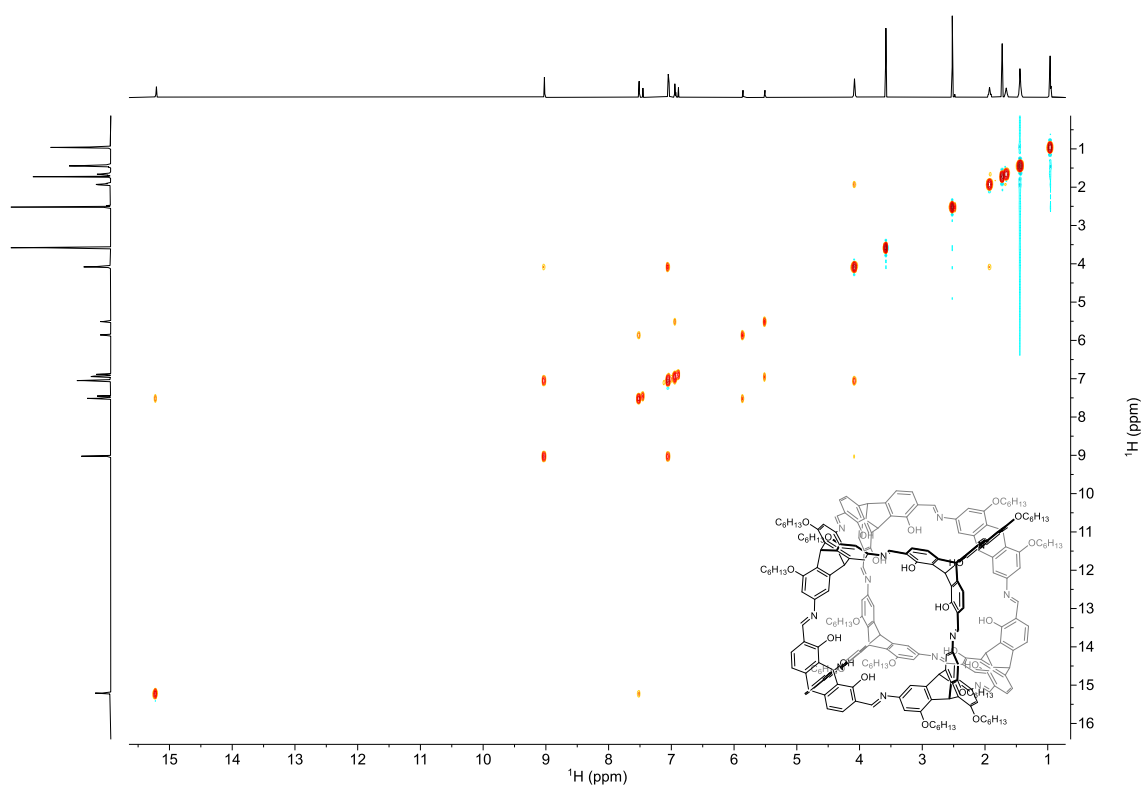


Figure S46. $^1\text{H}, ^1\text{H}$ NOESY spectrum (600 MHz, THF-d_8) of [4+4] cube **11**.

Table S14. Cross-peak assignment of ¹H,¹H-COSY and ¹H,¹H-NOESY spectra of [4+4] cube **11**.

¹ H nucleus	δ_{1H} [ppm]	COSY	NOESY
<i>a</i>	15.25 – 15.18	-	<i>c</i>
<i>b</i>	9.02	-	<i>e, j</i>
<i>c</i>	7.51	<i>e</i>	<i>a, h</i>
<i>d</i>	7.45	-	-
<i>e</i>	7.07 – 7.02	<i>c</i>	<i>b, j</i>
<i>f</i>	6.94	-	<i>i</i>
<i>g</i>	6.89	-	-
<i>h</i>	5.86	-	<i>c</i>
<i>i</i>	5.51	-	<i>f</i>
<i>j</i>	4.08	<i>k</i>	<i>b, e, k</i>
<i>k</i>	1.97 – 1.87	<i>j, l</i>	<i>j, l</i>
<i>l</i>	1.70 – 1.60	<i>k, m</i>	-
<i>m</i>	1.49 – 1.38	<i>l, n</i>	-
<i>n</i>	1.01 – 0.91	<i>m</i>	-

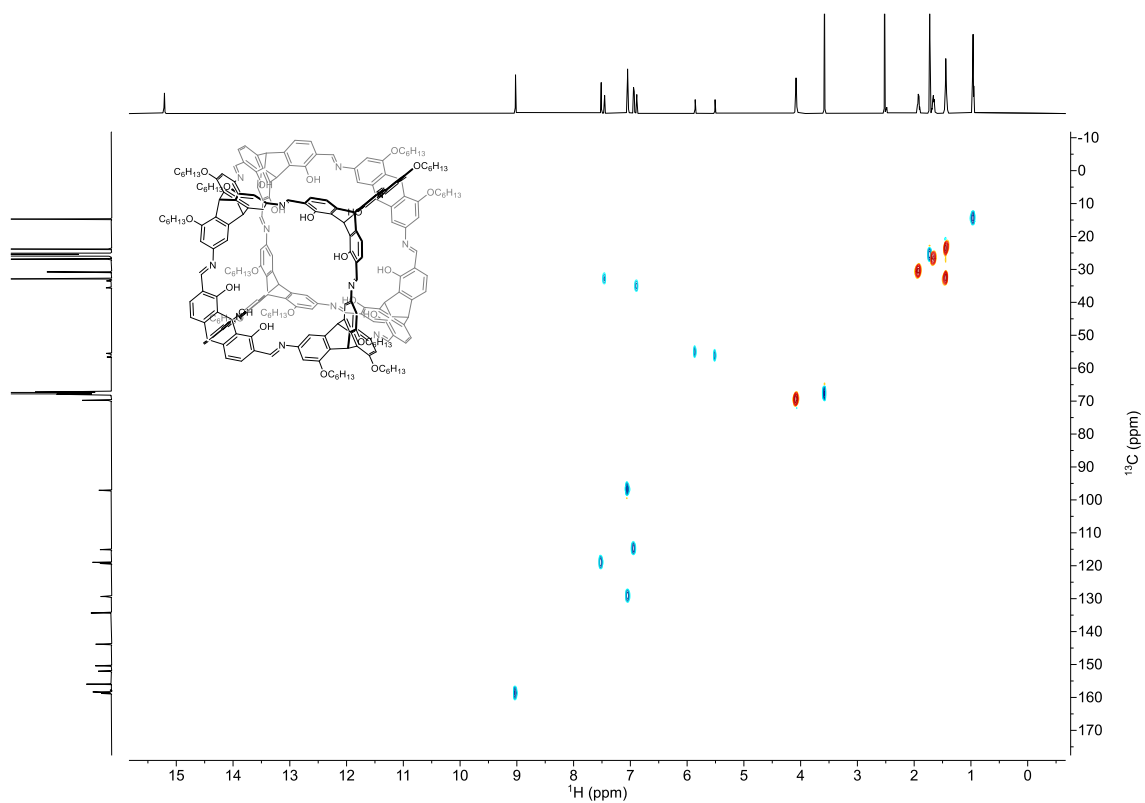


Figure S47. ^1H , ^{13}C HSQC spectrum (600 MHz, 150 MHz, THF-d_8) of [4+4] cube **11**.

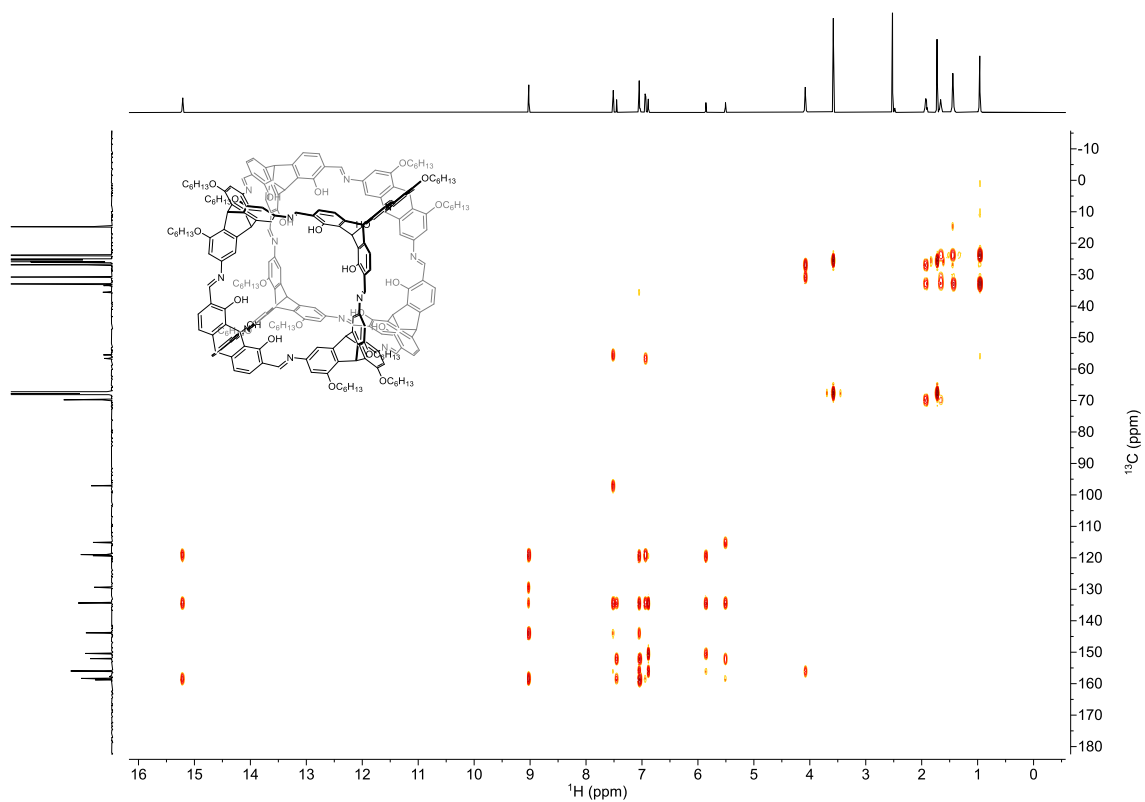


Figure S48. ^1H , ^{13}C HMBC spectrum (600 MHz, 150 MHz, THF-d_8) of [4+4] cube **11**.

Table S15. Correlations of ^1H , ^{13}C -HSQC and ^1H , ^{13}C -HMBC spectra of [4+4] cube **11**.

^{13}C nucleus	$\delta_{13\text{C}}$ [ppm]	^1H nucleus		
		HSQC	$\delta_{1\text{H}}$ [ppm]	HMBC
A	158.8	<i>b</i>	9.02	<i>e</i>
B	158.3	-	-	<i>a, b, d, e</i>
C	157.8	-	-	-
D	156.0	-	-	<i>c, e, g, h, j</i>
E	152.0	-	-	<i>d, e, i</i>
F	150.4	-	-	<i>g, h</i>
G	143.8	-	-	<i>b, c, e</i>
H	134.4	-	-	<i>a, b, c, d, e, f, g, h, i</i>
I	134.3	-	-	<i>a, b, c, d, e, f, g, h, i</i>
J	134.3	-	-	<i>a, b, c, d, e, f, g, h, i</i>
K	129.4	<i>e</i>	7.07 – 7.02	<i>b</i>
L	119.3	<i>c</i>	7.51	<i>e, h</i>
M	119.0	-	-	<i>a, b, f</i>
O	115.1	<i>f</i>	6.94	<i>i</i>
P	97.1	<i>e</i>	7.07 – 7.02	<i>c</i>
Q	69.8	<i>j</i>	4.08	<i>k</i>
R	56.6	<i>i</i>	5.51	<i>f</i>
S	55.5	<i>h</i>	5.86	<i>c</i>
T	35.6	<i>g</i>	6.89	<i>e</i>
U	33.5	<i>d</i>	7.45	-
V	32.9	<i>m</i>	1.49 – 1.38	<i>k, l, m, n</i>
W	30.8	<i>k</i>	1.97 – 1.87	<i>j</i>
X	26.9	<i>l</i>	1.70 – 1.60	<i>j, k</i>
Y	23.8	<i>m</i>	1.49 – 1.38	<i>l, m, n</i>
Z	14.7	<i>n</i>	1.01 – 0.91	<i>m</i>

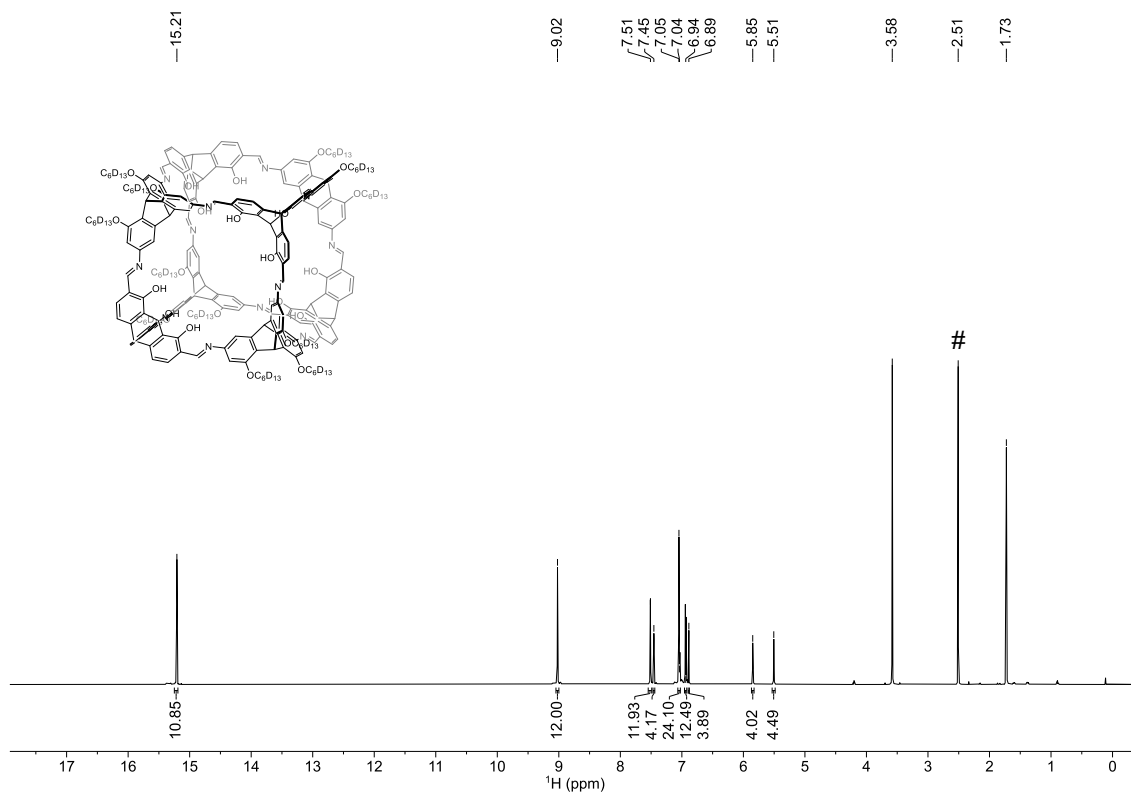


Figure S49. ¹H NMR spectrum (600 MHz, THF-d₈) of [4+4] cube 11-d₁₅₆. #: H₂O.

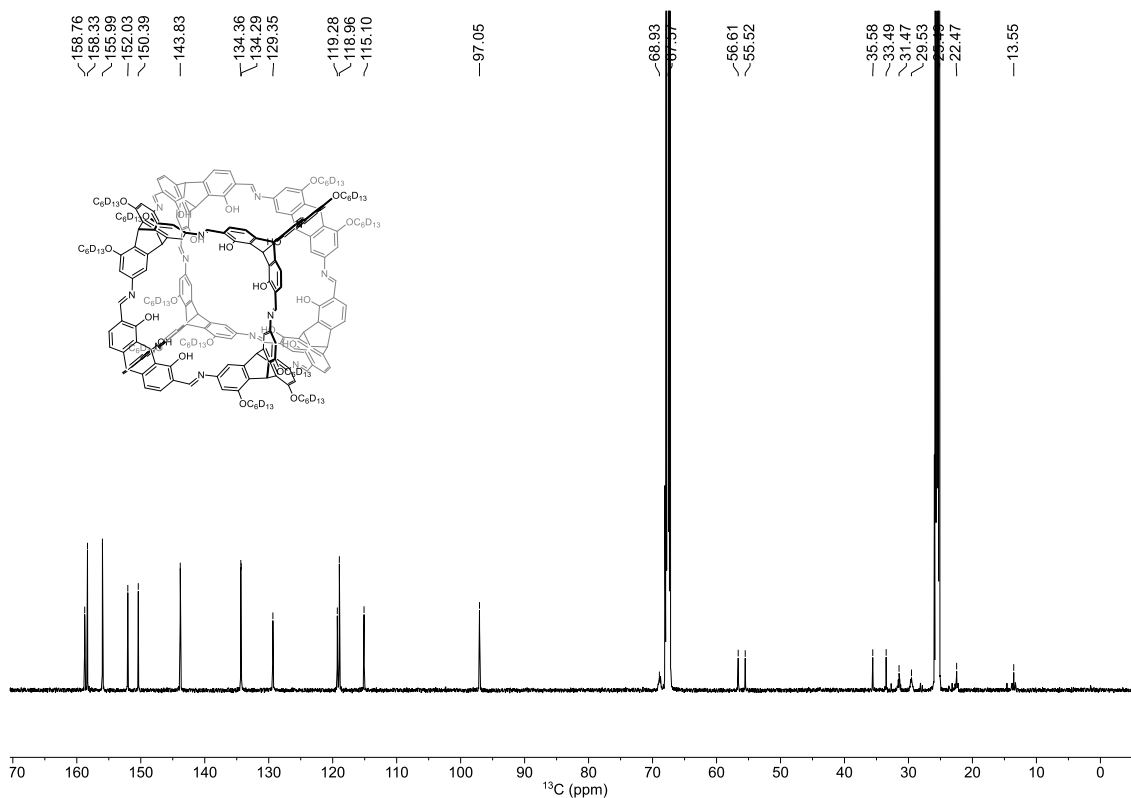


Figure S50. ¹³C{¹H} NMR spectrum (150 MHz, THF-d₈) of [4+4] cube 11-d₁₅₆.

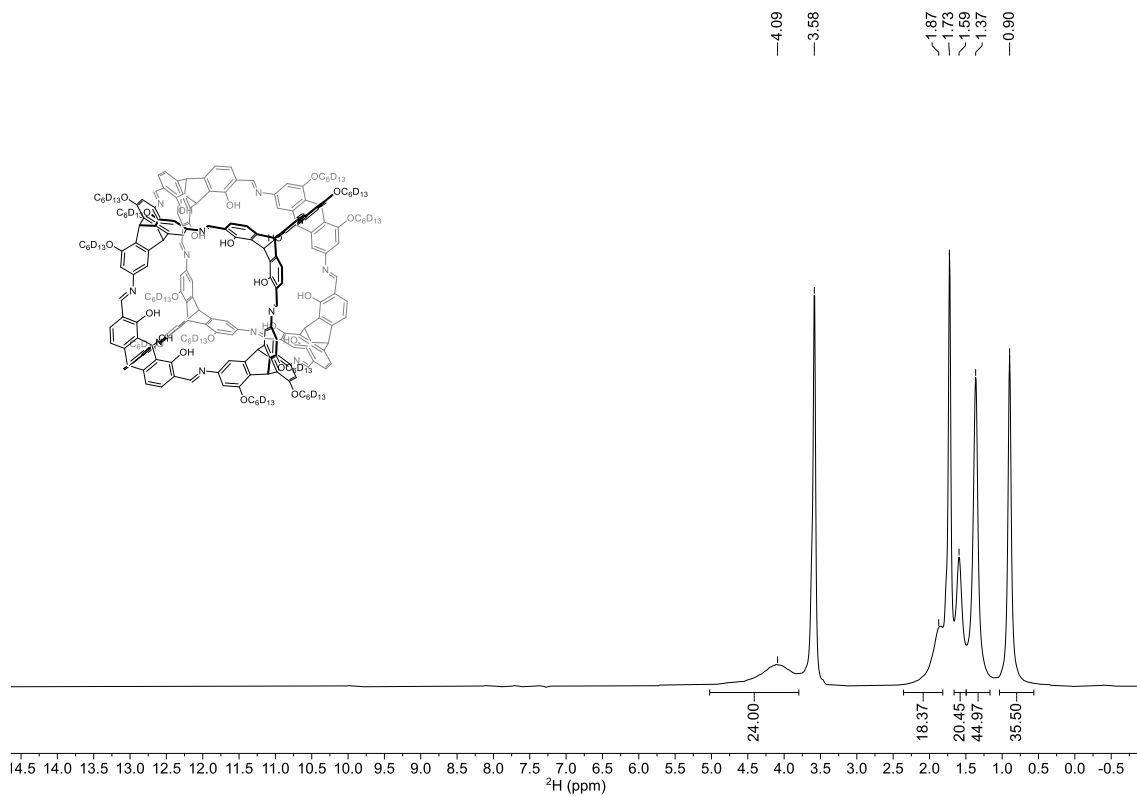


Figure S51. ²H NMR spectrum (92 MHz, THF/THF-d₈) of [4+4] cube **11-d₁₅₆**.

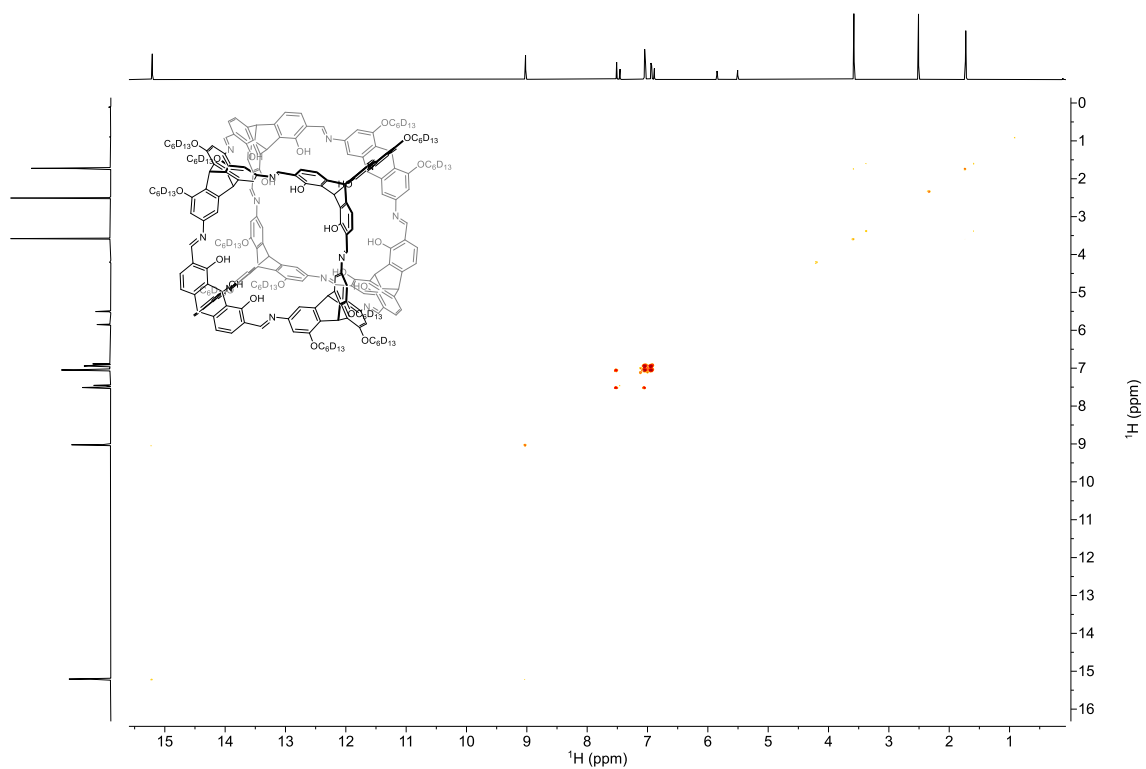


Figure S52. ¹H, ¹H COSY spectrum (600 MHz, THF-d₈) of [4+4] cube **11-d₁₅₆**.

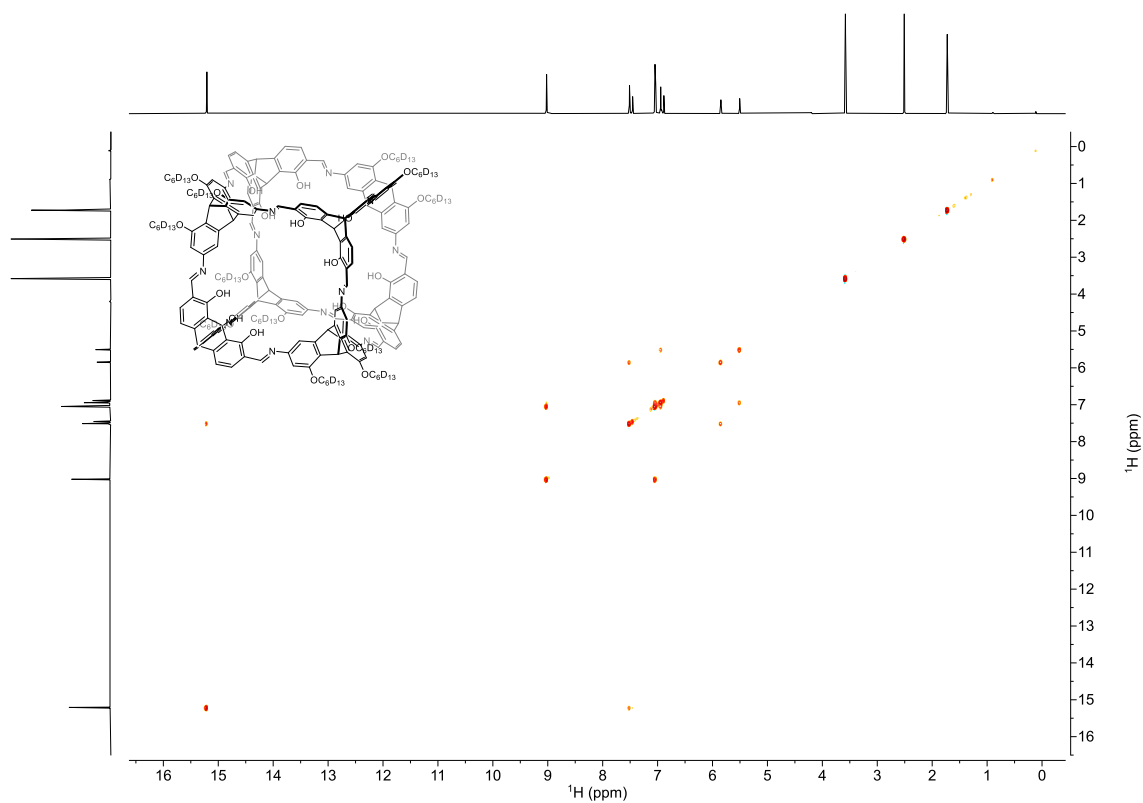


Figure S53. $^1\text{H}, ^1\text{H}$ NOESY spectrum (600 MHz, THF-d_8) of [4+4] cube **11-d₁₅₆**.

Table S16. Cross-peak assignment of $^1\text{H}, ^1\text{H}$ -COSY and $^1\text{H}, ^1\text{H}$ -NOESY spectra of [4+4] cube **11-d₁₅₆**.

^1H nucleus	$\delta_{1\text{H}}$ [ppm]	COSY	NOESY
<i>a</i>	15.21	-	<i>c</i>
<i>b</i>	9.02	-	<i>f</i>
<i>c</i>	7.51	<i>e</i>	<i>h</i>
<i>d</i>	7.45	-	-
<i>e</i>	7.07 – 7.02	<i>c, f</i>	<i>b, f</i>
<i>f</i>	6.94	<i>e</i>	<i>e, i</i>
<i>g</i>	6.89	-	-
<i>h</i>	5.85	-	<i>c</i>
<i>i</i>	5.51	-	<i>f</i>

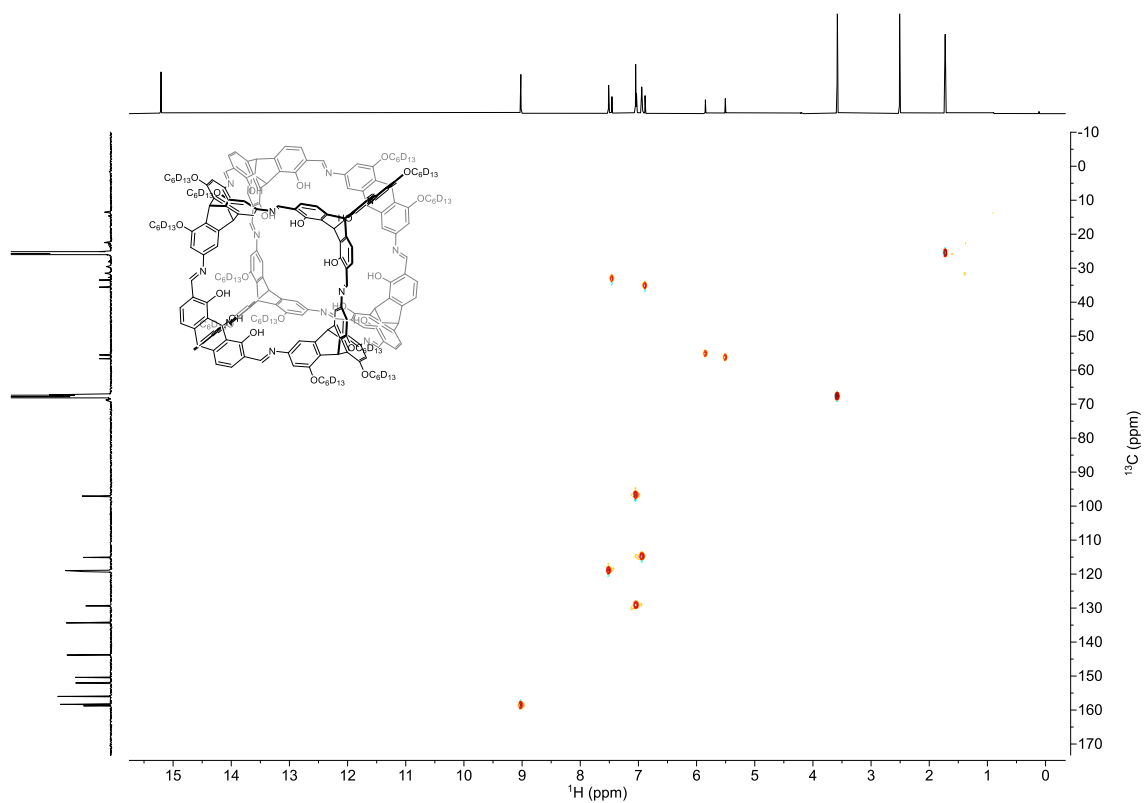


Figure S54. ^1H , ^{13}C HSQC spectrum (600 MHz, 150 MHz, THF-d_8) of [4+4] cube **11-d₁₅₆**.

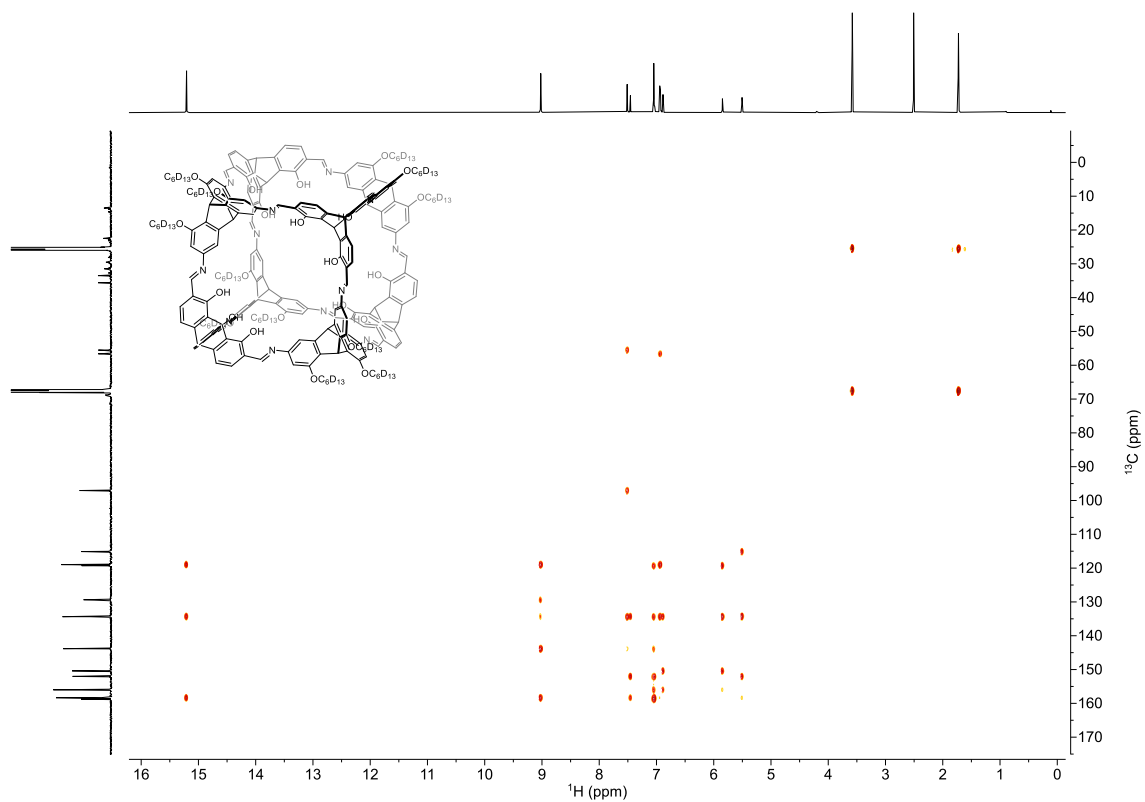


Figure S55. ^1H , ^{13}C HMBC spectrum (600 MHz, 150 MHz, THF-d_8) of [4+4] cube **11-d₁₅₆**.

Table S17. Correlations of ^1H , ^{13}C -HSQC and ^1H , ^{13}C -HMBC spectra of [4+4] cube **11-d**₁₅₆.

^{13}C nucleus	$\delta_{13\text{C}}$ [ppm]	^1H nucleus		
		HSQC	$\delta_{1\text{H}}$ [ppm]	HMBC
<i>A</i>	158.8	<i>b</i>	9.02	<i>a, b, d, e</i>
<i>B</i>	158.3	-	-	<i>a, b, d, e, i</i>
<i>C</i>	156.0	-	-	<i>e, g, h</i>
<i>D</i>	152.0	-	-	<i>d, e, i</i>
<i>E</i>	150.4	-	-	<i>g, h</i>
<i>F</i>	143.8	-	-	<i>b, c, e</i>
<i>G</i>	134.4	-	-	<i>a, b, c, d, e, f, g, h, i</i>
<i>H</i>	134.3	-	-	<i>a, b, c, d, e, f, g, h, i</i>
<i>I</i>	129.3	<i>e</i>	7.07 – 7.02	<i>b</i>
<i>J</i>	119.3	-	-	<i>e, h</i>
<i>K</i>	119.0	<i>c</i>	7.51	<i>a, b, f</i>
<i>L</i>	115.1	<i>f</i>	6.94	<i>i</i>
<i>M</i>	97.1	<i>e</i>	7.07 – 7.02	<i>c</i>
<i>O</i>	69.3 – 68.5	-	-	-
<i>P</i>	56.6	<i>i</i>	5.51	<i>f</i>
<i>Q</i>	55.5	<i>h</i>	5.85	<i>c</i>
<i>R</i>	35.6	<i>g</i>	6.89	-
<i>S</i>	33.5	<i>d</i>	7.45	-
<i>T</i>	32.1 – 31.1	-	-	-
<i>U</i>	30.0 – 29.0	-	-	-
<i>V</i>	22.9 – 22.2	-	-	-
<i>W</i>	14.0 – 13.1	-	-	-

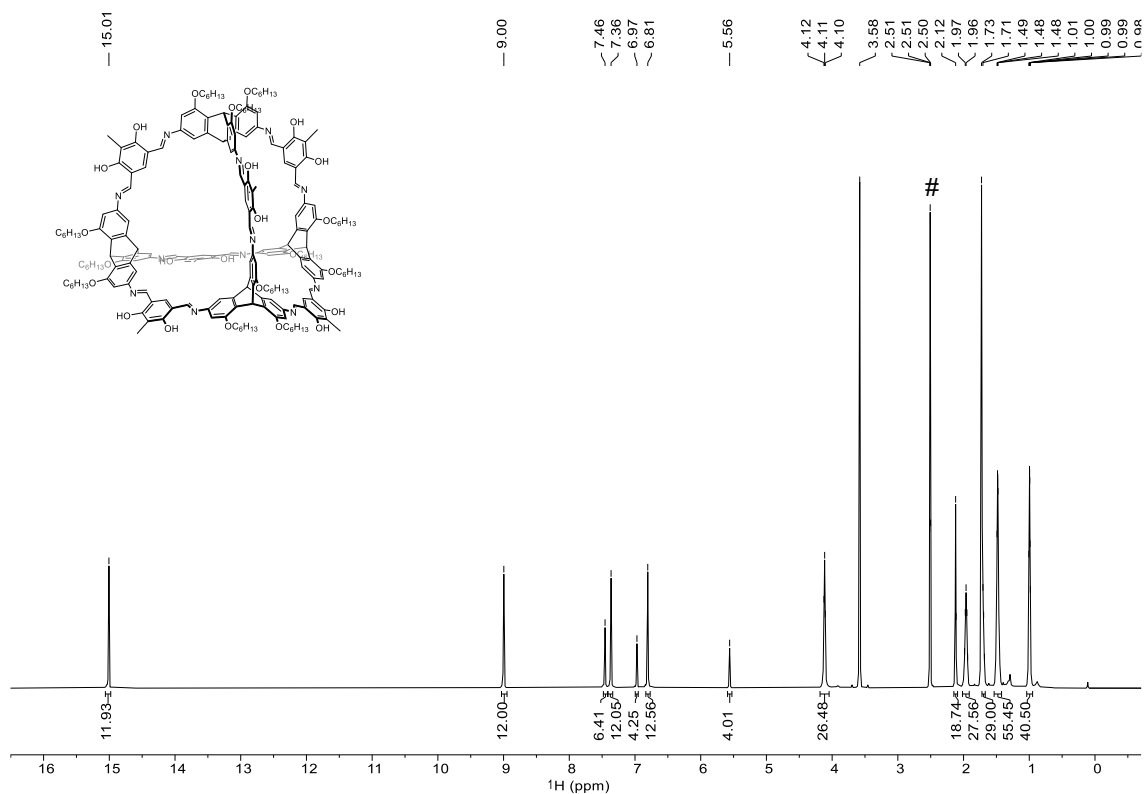


Figure S56. ^1H NMR spectrum (600 MHz, THF-d_8) of [4+6]-exo cage 12. #: H_2O .

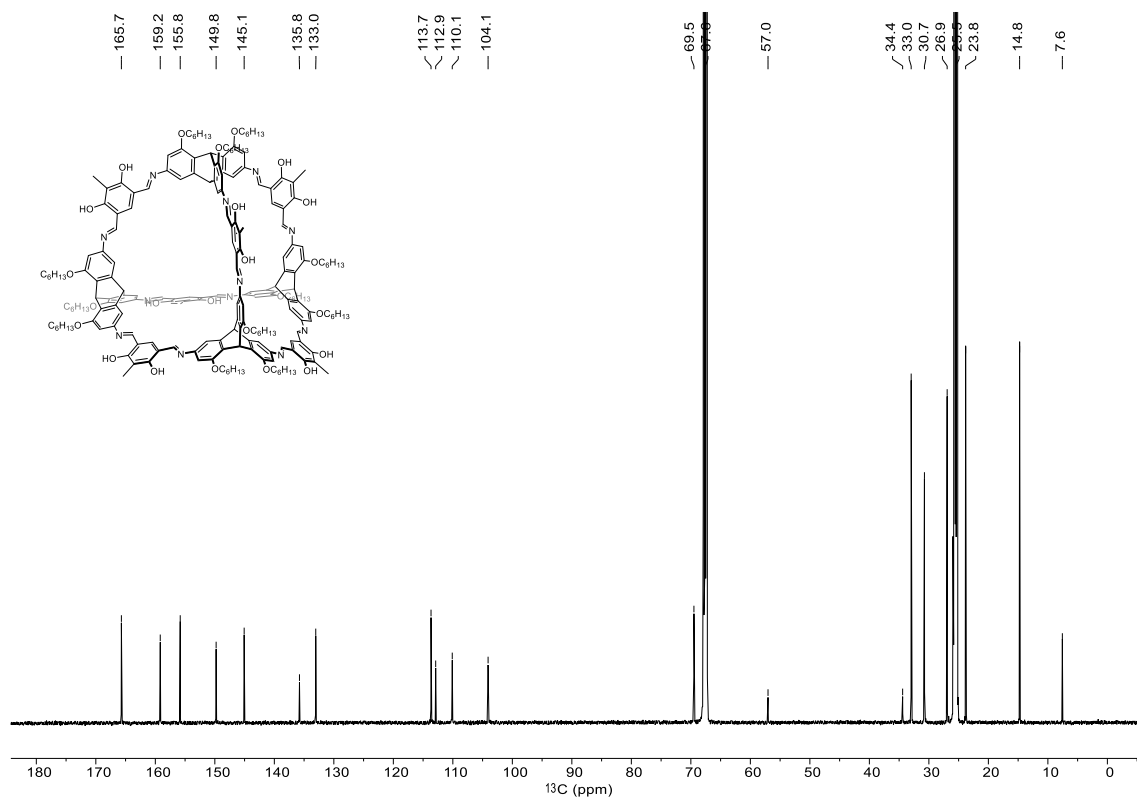


Figure S57. $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (150 MHz, THF-d_8) of [4+6]-exo cage 12.

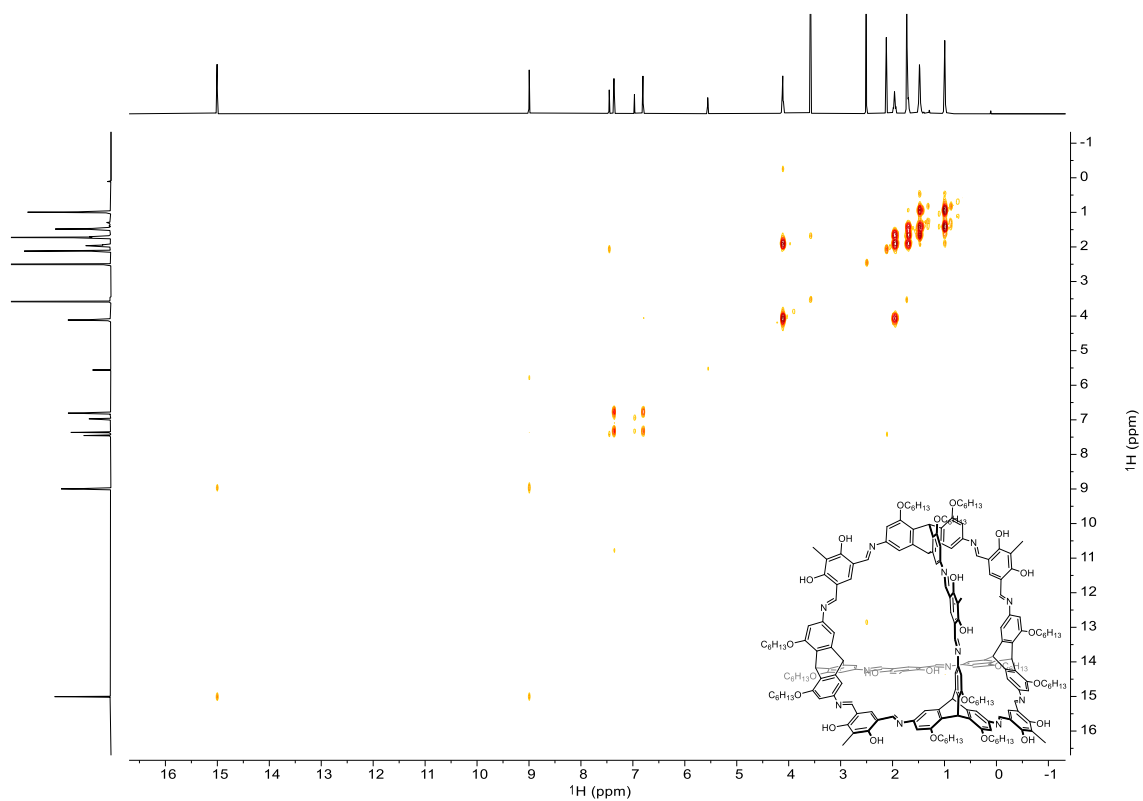


Figure S58. ^1H , ^1H COSY spectrum (600 MHz, THF-d_8) of [4+6]-exo cage 12.

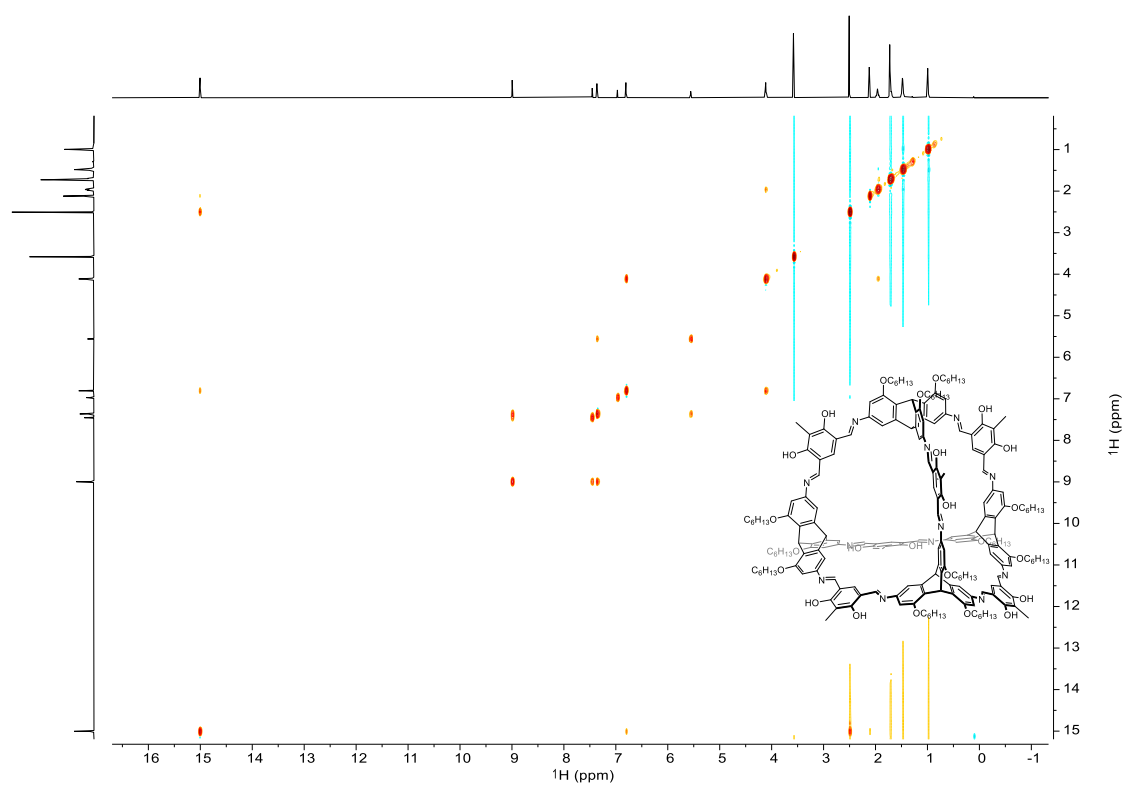


Figure S59. ^1H , ^1H NOESY spectrum (600 MHz, THF-d_8) of [4+6]-exo cage 12.

Table S18. Cross-peak assignment of ¹H,¹H-COSY and ¹H,¹H-NOESY spectra of [4+6]-exo cage **12**.

¹ H nucleus	δ_{1H} [ppm]	COSY	NOESY
<i>a</i>	15.01	<i>b</i>	<i>f, i, H₂O</i>
<i>b</i>	9.00	<i>a</i>	<i>c, d</i>
<i>c</i>	7.46	-	<i>b</i>
<i>d</i>	7.36	<i>f</i>	<i>b, g</i>
<i>e</i>	6.97	-	-
<i>f</i>	6.81	<i>d</i>	<i>h</i>
<i>g</i>	5.56	-	<i>d</i>
<i>h</i>	4.11	<i>j</i>	<i>f, j</i>
<i>i</i>	2.12	-	<i>a</i>
<i>j</i>	2.01 – 1.92	<i>h, k</i>	<i>h</i>
<i>k</i>	1.76 – 1.67	<i>j, l</i>	-
<i>l</i>	1.52 – 1.43	<i>k, m</i>	-
<i>m</i>	1.00	<i>l</i>	-

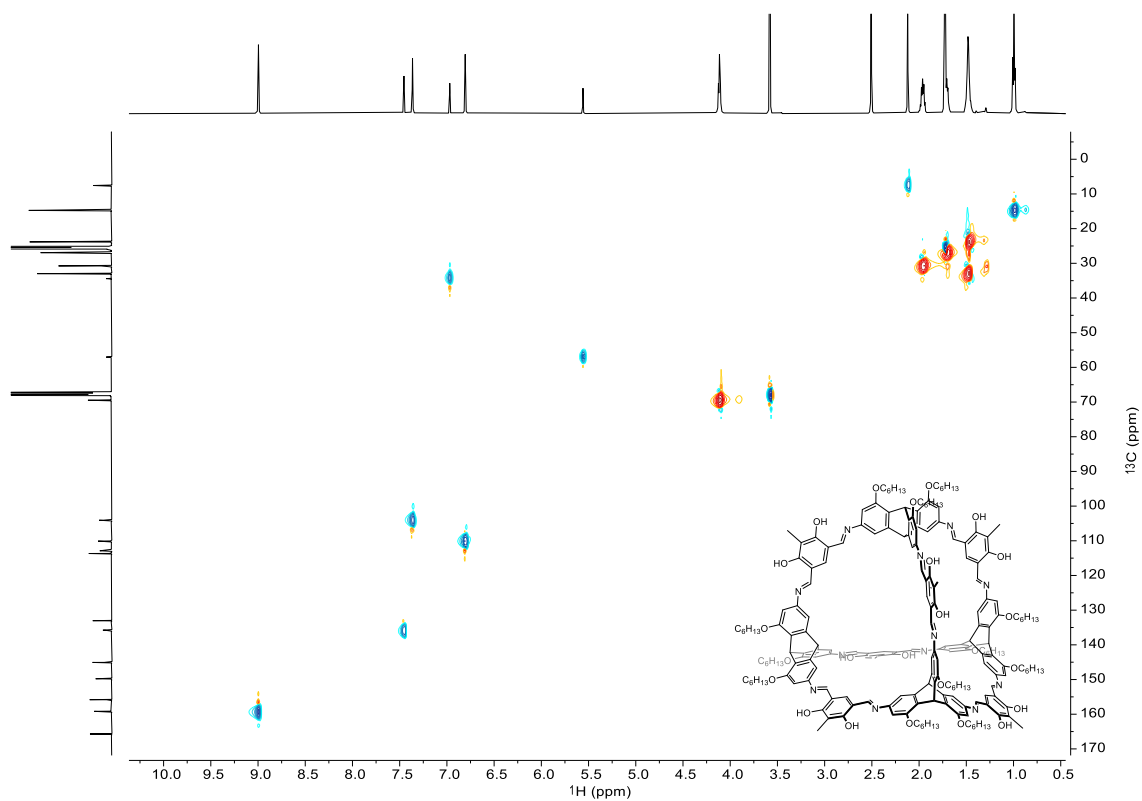


Figure S60. ^1H , ^{13}C HSQC spectrum (600 MHz, 150 MHz, THF-d_8) of [4+6]-exo cage **12**.

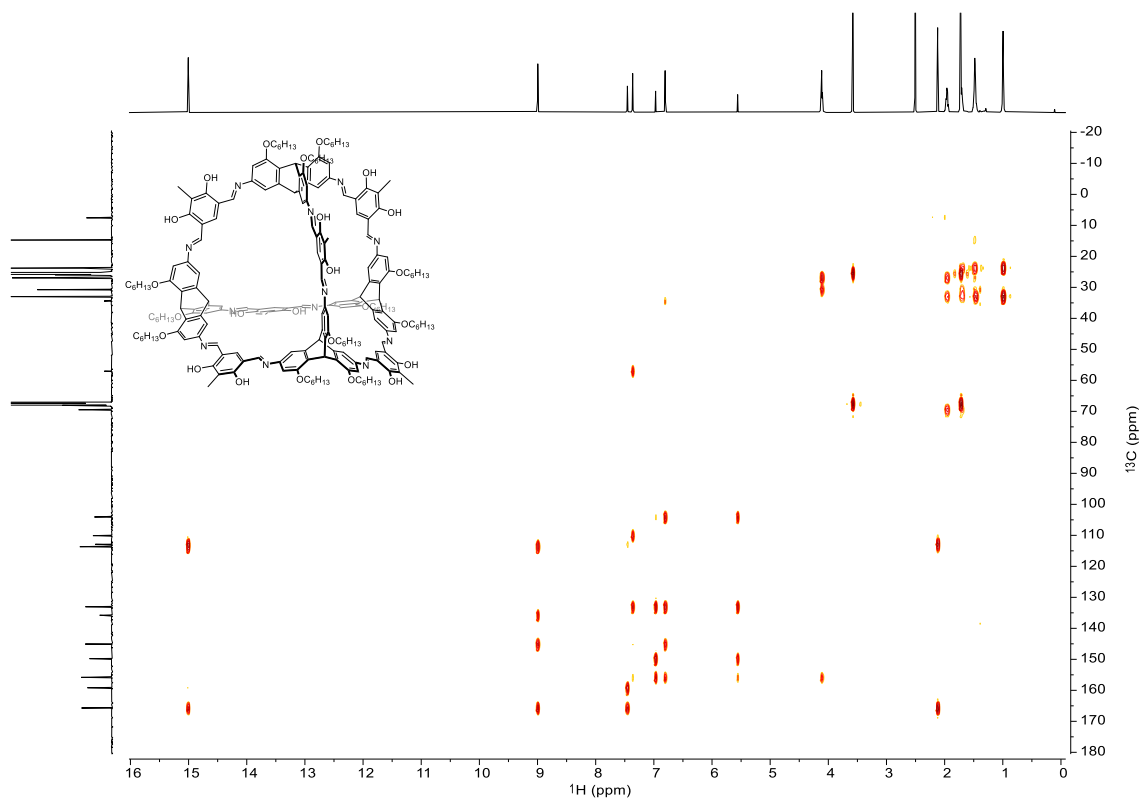


Figure S61. ^1H , ^{13}C HMBC spectrum (600 MHz, 150 MHz, THF-d_8) of [4+6]-exo cage **12**.

Table S19. Correlations of ^1H , ^{13}C -HSQC and ^1H , ^{13}C -HMBC spectra of [4+6]-exo cage **12**.

^{13}C nucleus	$\delta_{13\text{C}}$ [ppm]	^1H nucleus		
		HSQC	$\delta_{1\text{H}}$ [ppm]	HMBC
A	165.7	-	-	<i>a, b, c, i</i>
B	159.2	<i>b</i>	9.00	<i>c</i>
C	155.8	-	-	<i>e, f, h</i>
D	149.8	-	-	<i>e, g</i>
E	145.1	-	-	<i>b, f</i>
F	135.8	<i>c</i>	7.46	<i>b</i>
G	133.0	-	-	<i>d, e, f, g</i>
H	113.7	-	-	<i>a, b, i</i>
I	112.9	-	-	<i>a, b, i</i>
J	110.1	<i>f</i>	6.81	<i>d</i>
K	104.1	<i>d</i>	7.36	<i>f, g</i>
L	69.5	<i>h</i>	4.11	<i>j</i>
M	57.0	<i>g</i>	5.51	<i>d</i>
O	34.4	<i>e</i>	6.97	-
P	33.0	<i>l</i>	1.52 – 1.43	<i>j, k, l, m</i>
Q	30.7	<i>j</i>	2.01 – 1.92	<i>h</i>
R	26.9	<i>k</i>	1.76 – 1.67	<i>h, j</i>
S	23.8	<i>l</i>	1.52 – 1.43	<i>k, l, m</i>
T	14.8	<i>m</i>	1.00	<i>l</i>
U	7.6	<i>i</i>	2.12	-

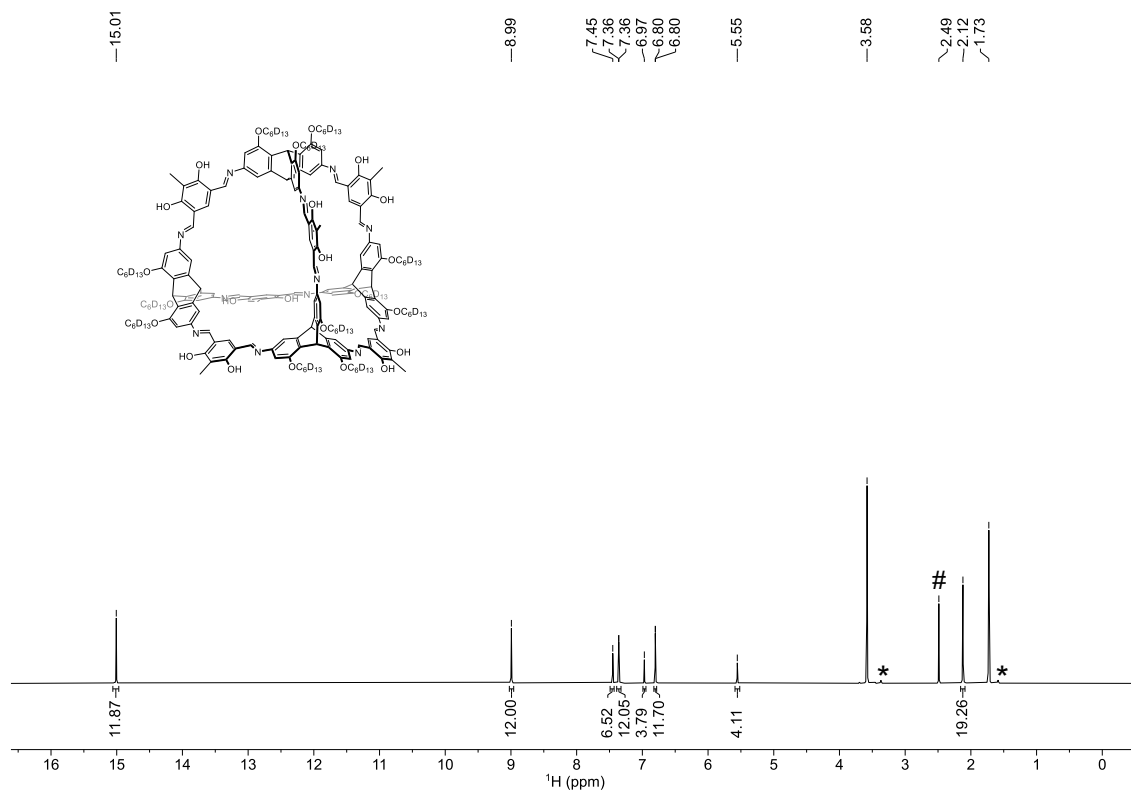


Figure S62. ¹H NMR spectrum (600 MHz, THF-d₈) of [4+6]-exo cage 12-d₁₅₆. #: H₂O, *: THF.

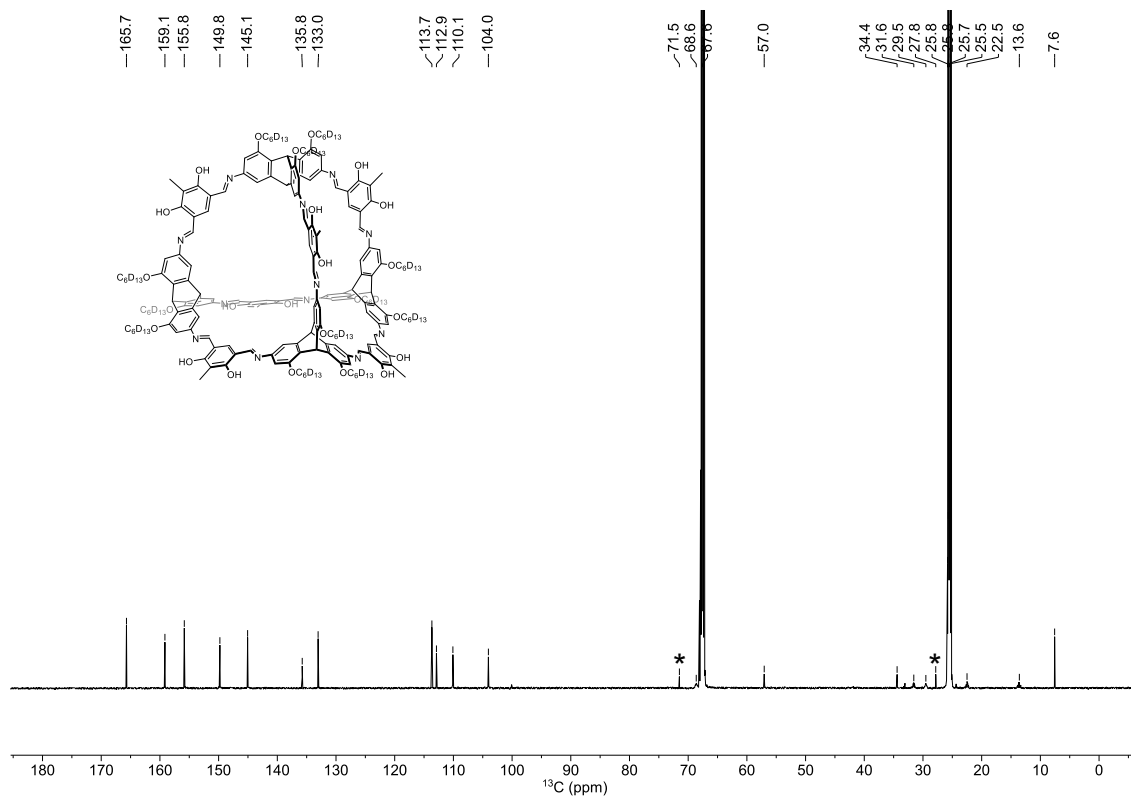


Figure S63. ¹³C{¹H} NMR spectrum (150 MHz, THF-d₈) of [4+6]-exo cage 12-d₁₅₆. *: THF.

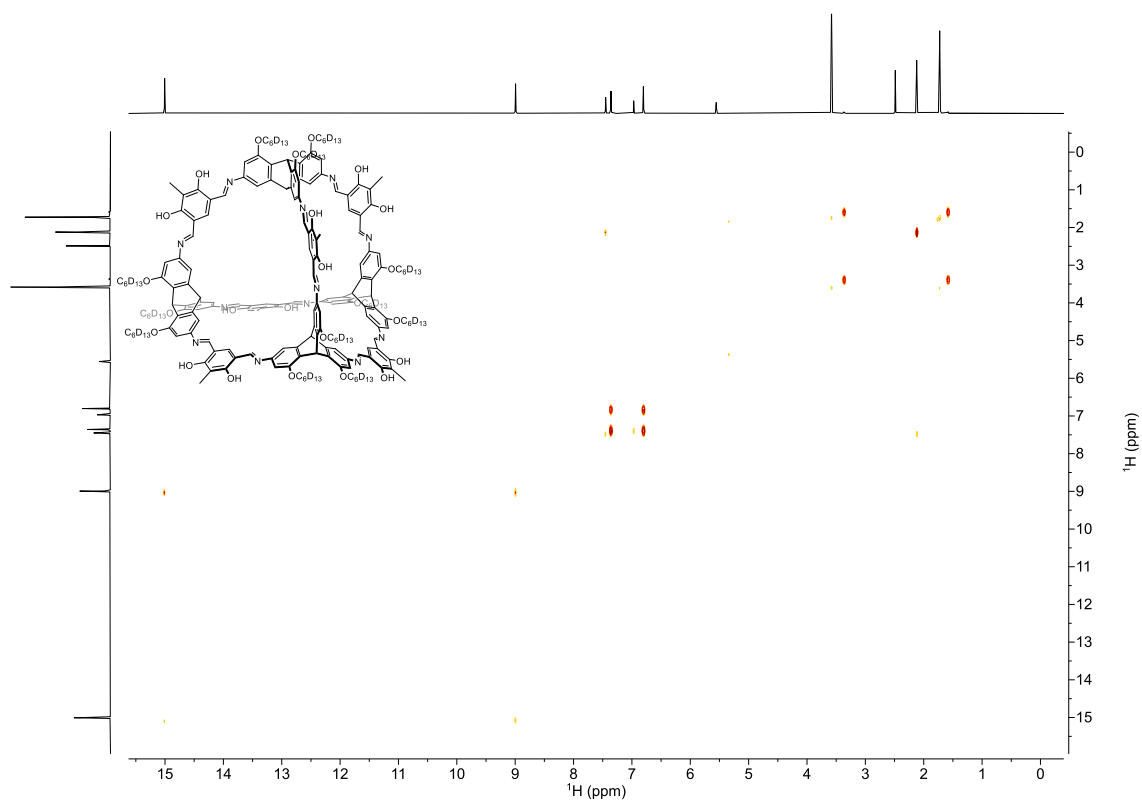


Figure S64. $^1\text{H}, ^1\text{H}$ COSY spectrum (600 MHz, THF-d_8) of [4+6]-exo cage **12-d₁₅₆**.

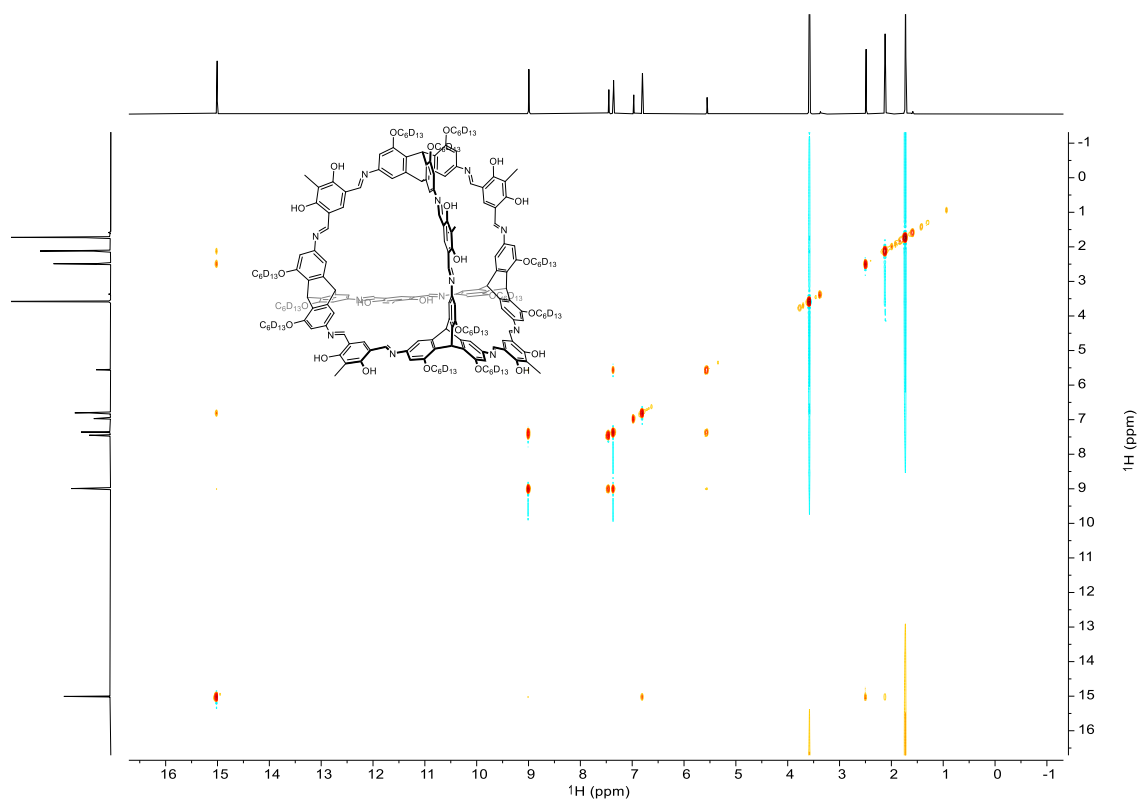


Figure S65. $^1\text{H}, ^1\text{H}$ NOESY spectrum (600 MHz, THF-d_8) of [4+6]-exo cage **12-d₁₅₆**.

Table S20. Cross-peak assignment of ^1H , ^1H -COSY and ^1H , ^1H -NOESY spectra of [4+6]-exo cage **12-d**₁₅₆.

^1H nucleus	$\delta_{1\text{H}}$ [ppm]	COSY	NOESY
<i>a</i>	15.01	-	<i>f</i> , H ₂ O, <i>h</i>
<i>b</i>	8.99	-	<i>c</i> , <i>d</i>
<i>c</i>	7.45	-	<i>b</i>
<i>d</i>	7.36	<i>f</i>	<i>b</i> , <i>g</i>
<i>e</i>	6.97	-	-
<i>f</i>	6.80	<i>d</i>	-
<i>g</i>	5.55	-	<i>d</i>
<i>h</i>	2.12	-	<i>a</i>

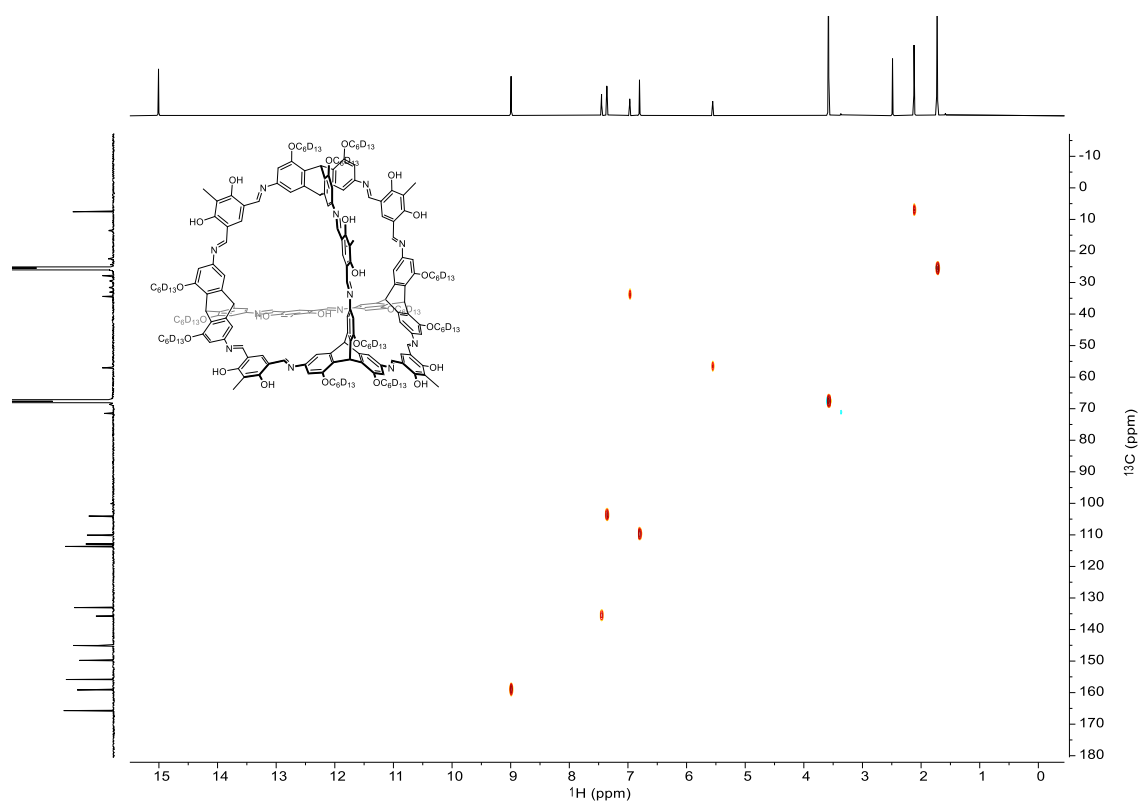


Figure S66. ^1H , ^{13}C HSQC spectrum (600 MHz, 150 MHz, THF- d_8) of [4+6]-exo cage **12-d₁₅₆**.

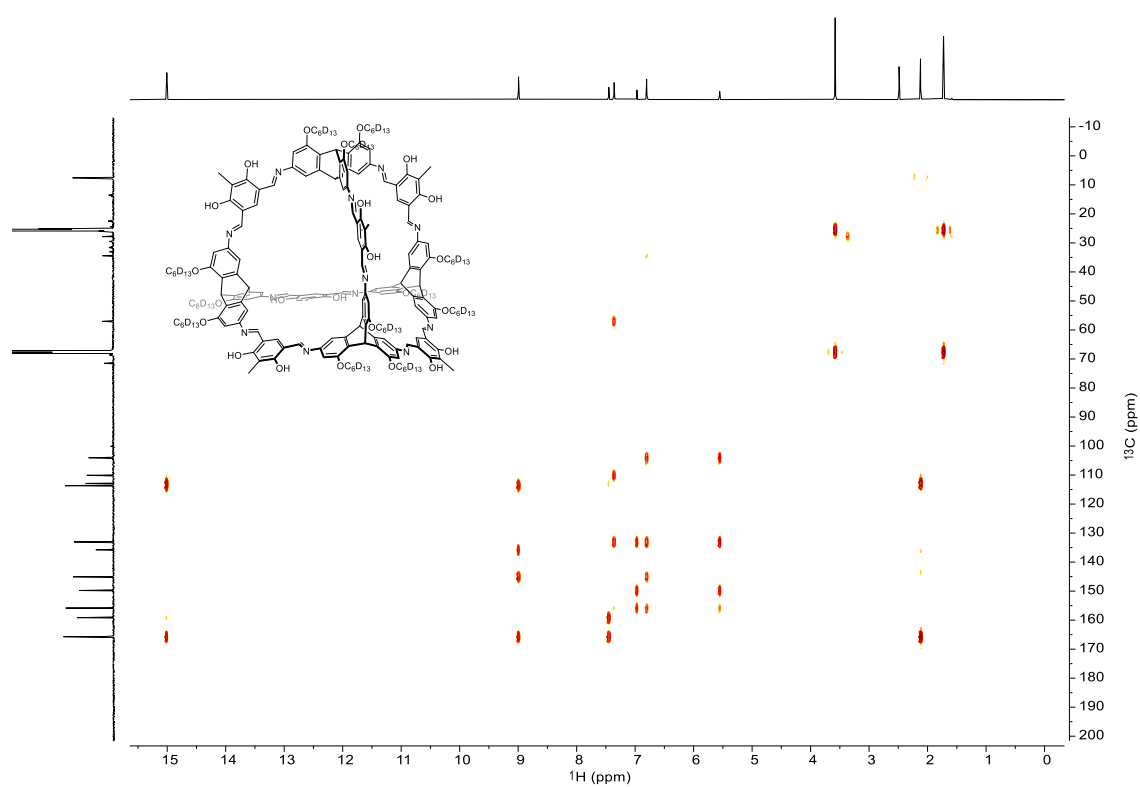


Figure S67. ^1H , ^{13}C HMBC spectrum (600 MHz, 150 MHz, THF- d_8) of [4+6]-exo cage **12-d₁₅₆**.

Table S21. Correlations of ^1H , ^{13}C -HSQC and ^1H , ^{13}C -HMBC spectra of [4+6]-exo cage **12-d**₁₅₆.

^{13}C nucleus	$\delta_{13\text{C}}$ [ppm]	^1H nucleus		
		HSQC	$\delta_{1\text{H}}$ [ppm]	HMBC
A	165.7	<i>b</i>	8.99	<i>a, b, c, h</i>
B	159.1	-	-	<i>c</i>
C	155.8	-	-	<i>d, e, f, g</i>
D	149.8	-	-	<i>e, g</i>
E	145.1	-	-	<i>b, f</i>
F	135.8	<i>c</i>	7.45	<i>b</i>
G	133.0	-	-	<i>d, e, f, g</i>
H	113.7	-	-	<i>a, b, h</i>
I	112.9	-	-	<i>a, b, h</i>
J	110.1	<i>f</i>	6.80	<i>d</i>
K	104.0	<i>d</i>	7.36	<i>f, g</i>
L	69.1 – 68.3	-	-	-
M	57.0	<i>g</i>	5.55	<i>d</i>
O	34.4	<i>e</i>	6.97	-
P	32.1 – 31.1	-	-	-
Q	30.1 – 29.1	-	-	-
R	23.0 – 22.0	-	-	-
S	14.0 – 13.2	-	-	-
T	7.6	<i>h</i>	2.12	-

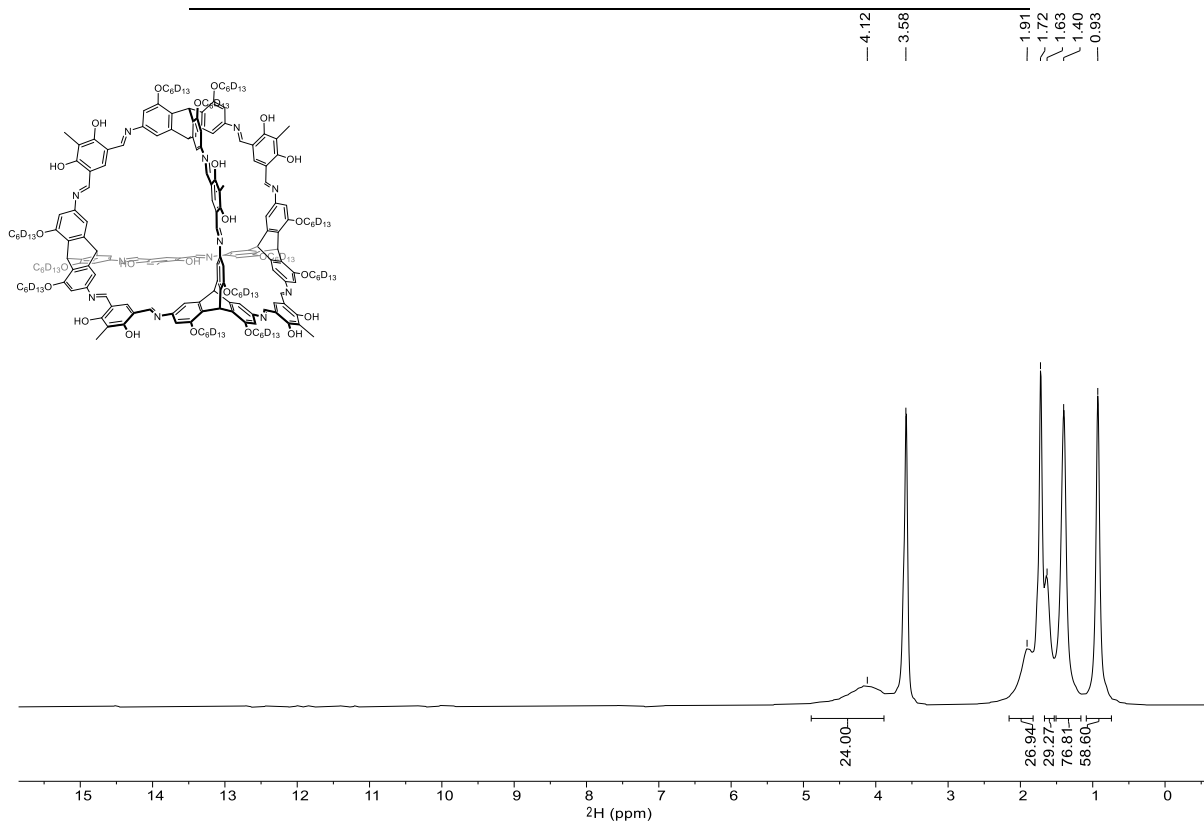


Figure S68. ^2H NMR spectrum (92 MHz, THF/THF- d_8) of [4+6]-exo cage **12-d**₁₅₆

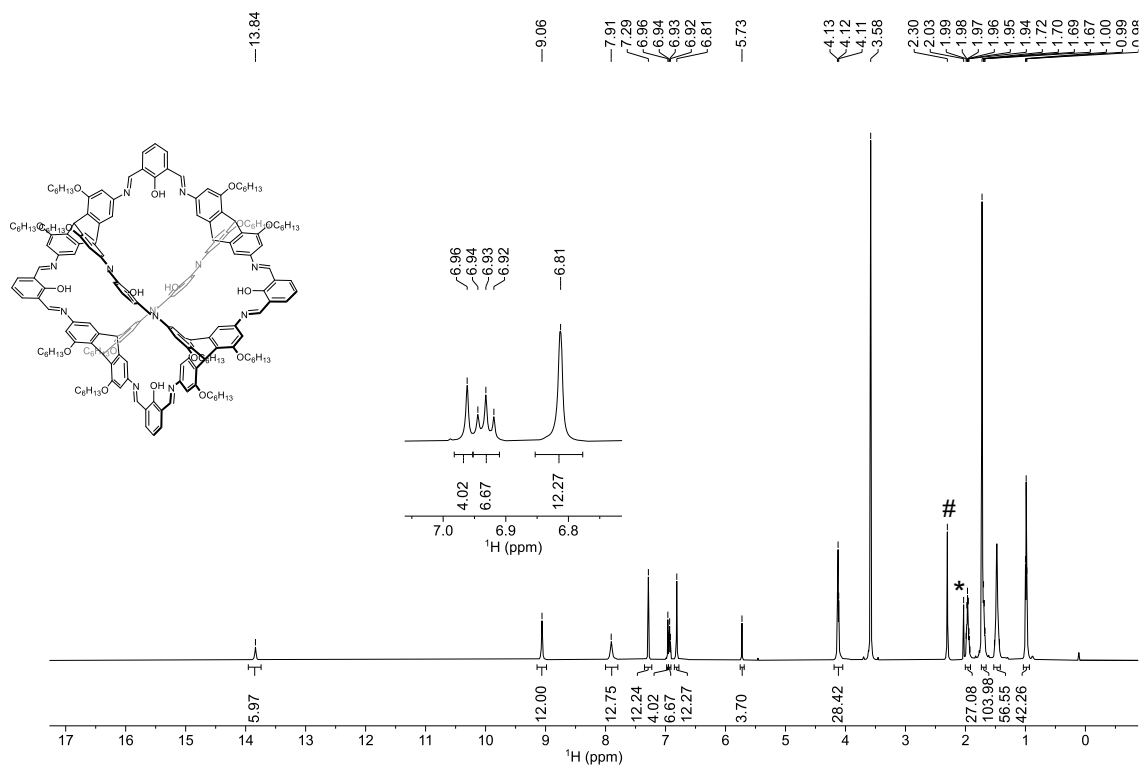


Figure S69. ^1H NMR spectrum (600 MHz, 328 K, THF-d_8) of [4+6]-endo cage 13. #: H_2O , *: Acetone.

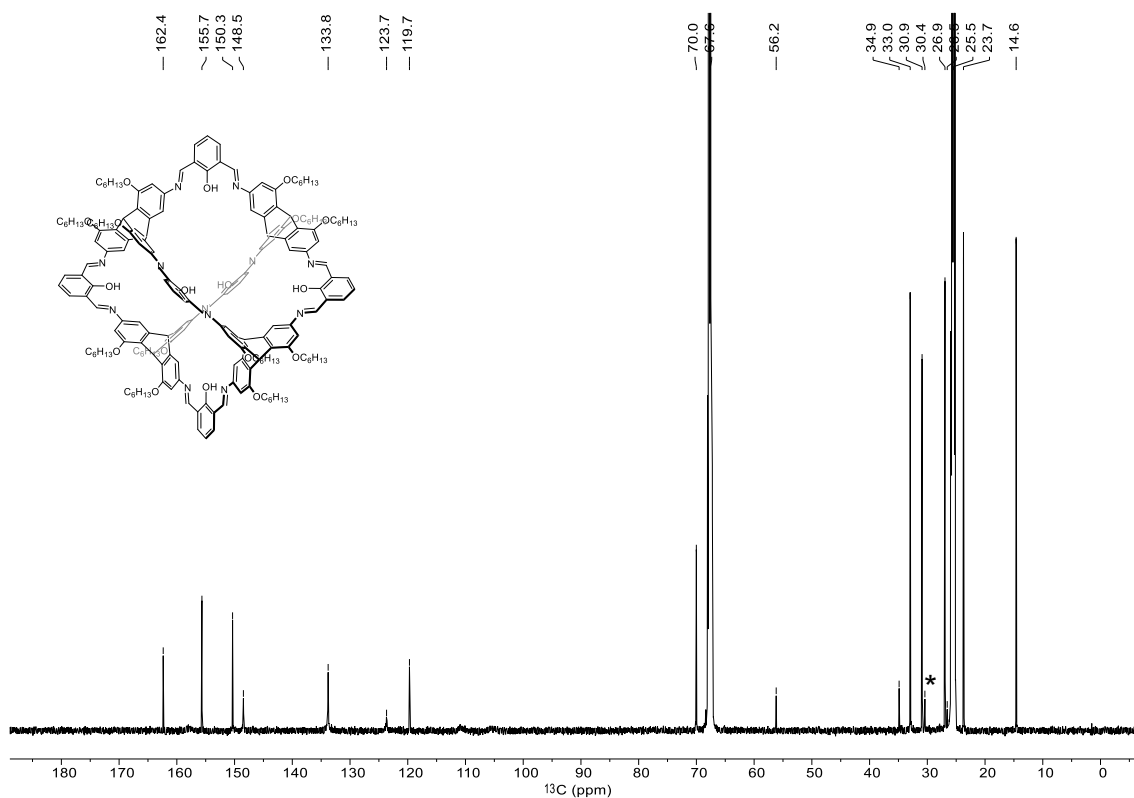


Figure S70. $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (150 MHz, 328 K, THF-d_8) of [4+6]-endo cage 13. *: Acetone

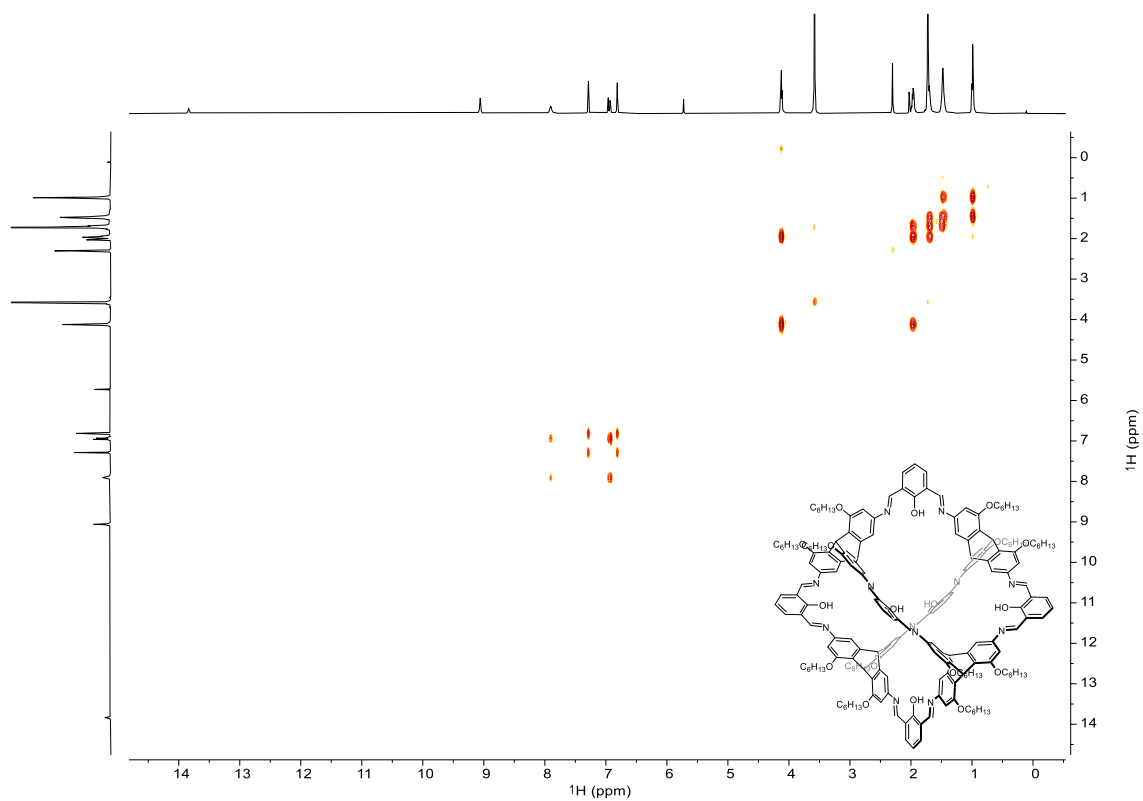


Figure S71. $^1\text{H},^1\text{H}$ COSY spectrum (600 MHz, 328 K, THF-d_8) of [4+6]-endo cage **13**.

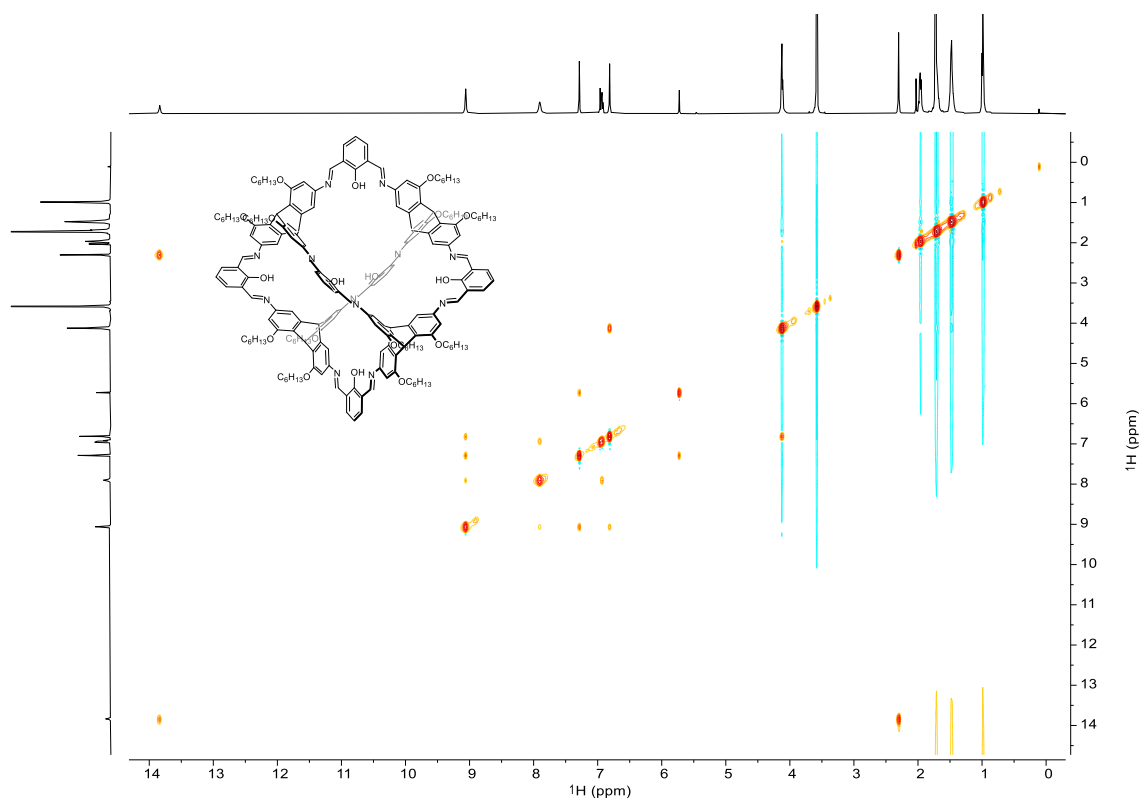


Figure S72. $^1\text{H},^1\text{H}$ NOESY spectrum (600 MHz, 328 K, THF-d_8) of [4+6]-endo cage **13**.

Table S22. Cross-peak assignment of ¹H,¹H-COSY and ¹H,¹H-NOESY spectra of [4+6]-*endo* cage **13**.

¹ H nucleus	δ_{1H} [ppm]	COSY	NOESY
<i>a</i>	13.84	-	H ₂ O
<i>b</i>	9.06	-	<i>c, d, g</i>
<i>c</i>	7.91	<i>f</i>	<i>b, f</i>
<i>d</i>	7.29	<i>g</i>	<i>b, h</i>
<i>e</i>	6.96	-	-
<i>f</i>	6.93	<i>c</i>	<i>c</i>
<i>g</i>	6.81	<i>d</i>	<i>b, i</i>
<i>h</i>	5.73	-	<i>d</i>
<i>i</i>	4.12	<i>j</i>	<i>g</i>
<i>j</i>	2.00 – 1.92	<i>i, k</i>	-
<i>k</i>	1.73 – 1.66	<i>j, l</i>	-
<i>l</i>	1.53 – 1.42	<i>k, m</i>	-
<i>m</i>	0.99	<i>l</i>	-

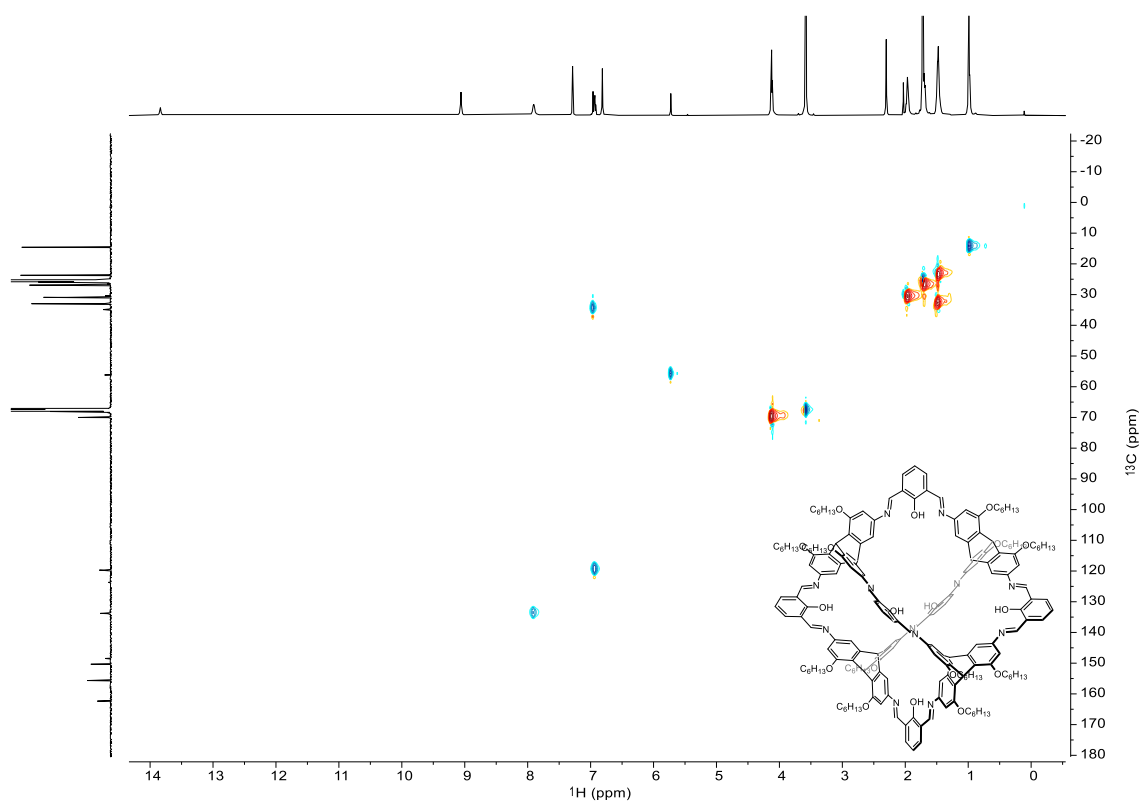


Figure S73. ^1H , ^{13}C HSQC spectrum (600 MHz, 150 MHz, 328 K, THF-d_8) of [4+6]-endo cage **13**.

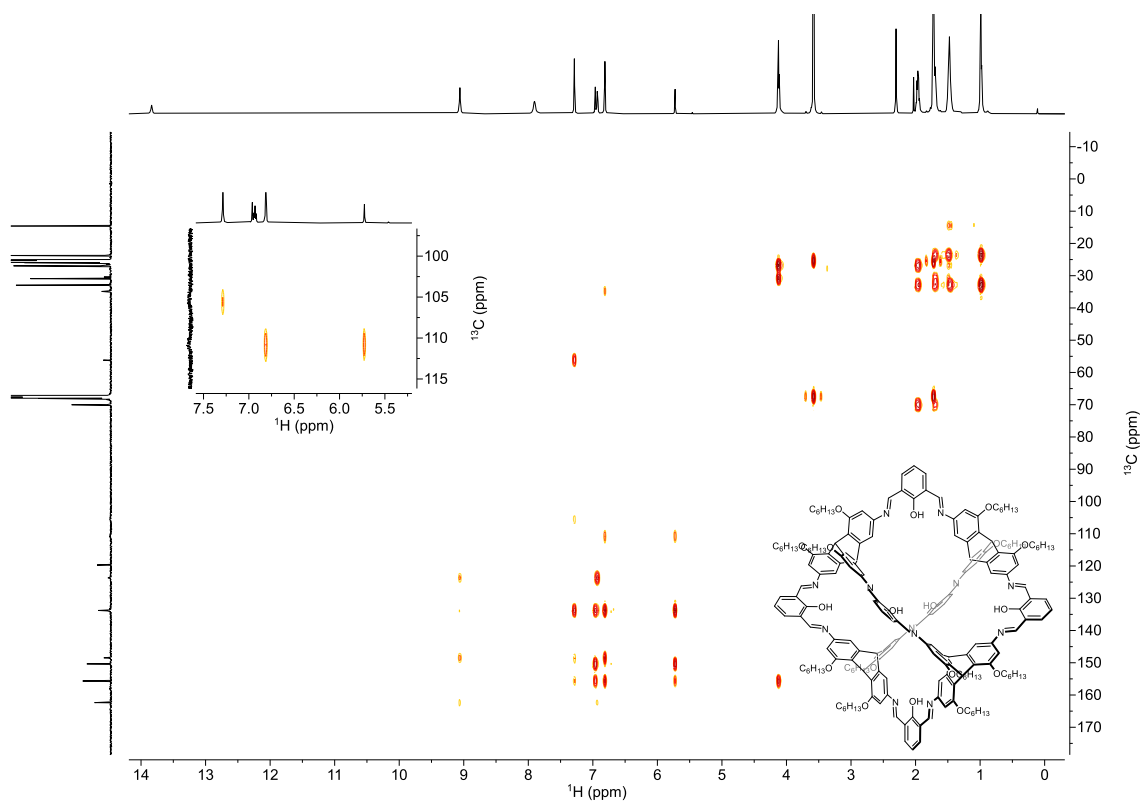


Figure S74. ^1H , ^{13}C HMBC spectrum (600 MHz, 150 MHz, 328 K, THF-d_8) of [4+6]-endo cage **13**. Also, a zoom into the region of 7.5 – 5.5 ppm (^1H) and 115 – 100 ppm (^{13}C) is illustrated to show the chemical shifts of the hydrogen bond aromatic carbon nuclei of the triptycene building block.

Table S23. Correlations of ^1H , ^{13}C -HSQC and ^1H , ^{13}C -HMBC spectra of [4+6]-endo cage **13**.

^{13}C nucleus	$\delta_{13\text{C}}$ [ppm]	^1H nucleus		
		HSQC	$\delta_{1\text{H}}$ [ppm]	HMBC
<i>A</i>	162.4	-	-	<i>b, f</i>
<i>B</i>	155.7	-	-	<i>d, e, g, h, i</i>
<i>C</i>	150.3	-	-	<i>e, h</i>
<i>D</i>	148.5	-	-	<i>b, d, g</i>
<i>E</i>	133.8	<i>c</i>	7.91	<i>b, d, e, f, g, h</i>
<i>F</i>	123.7	-	-	<i>b, f</i>
<i>G</i>	119.7	<i>f</i>	6.93	-
<i>H</i>	110.8	-	-	<i>g, h</i>
<i>I</i>	105.5	-	-	<i>d</i>
<i>J</i>	70.0	<i>i</i>	4.12	<i>j, k</i>
<i>K</i>	56.2	<i>h</i>	5.73	<i>d</i>
<i>L</i>	34.9	<i>e</i>	6.96	<i>g</i>
<i>M</i>	33.0	<i>l</i>	1.53 - 1.42	<i>j, k, l, m</i>
<i>O</i>	30.9	<i>j</i>	2.00 - 1.92	<i>i, k</i>
<i>P</i>	26.9	<i>k</i>	1.73 - 1.66	<i>i, j, l</i>
<i>Q</i>	23.7	<i>l</i>	1.53 - 1.42	<i>k, l, m</i>
<i>R</i>	14.6	<i>m</i>	0.99	<i>l</i>

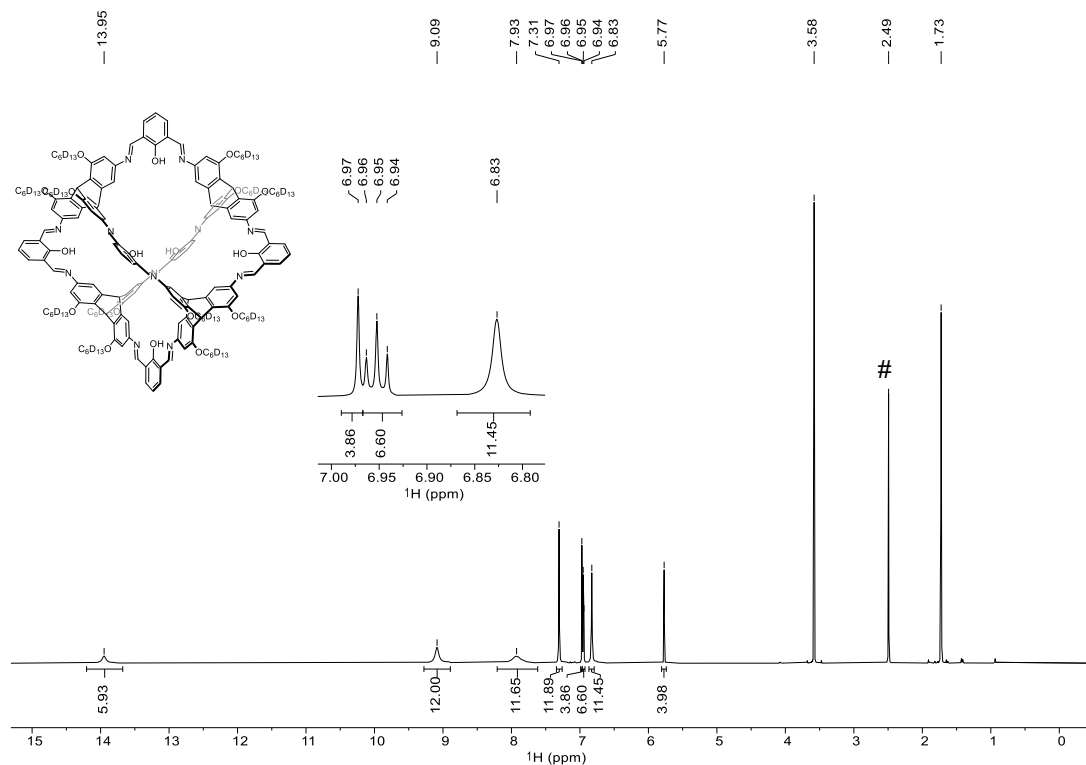


Figure S75. ¹H NMR spectrum (700 MHz, THF-d₈) of [4+6]-endo cage 13-d₁₅₆. #: H₂O.

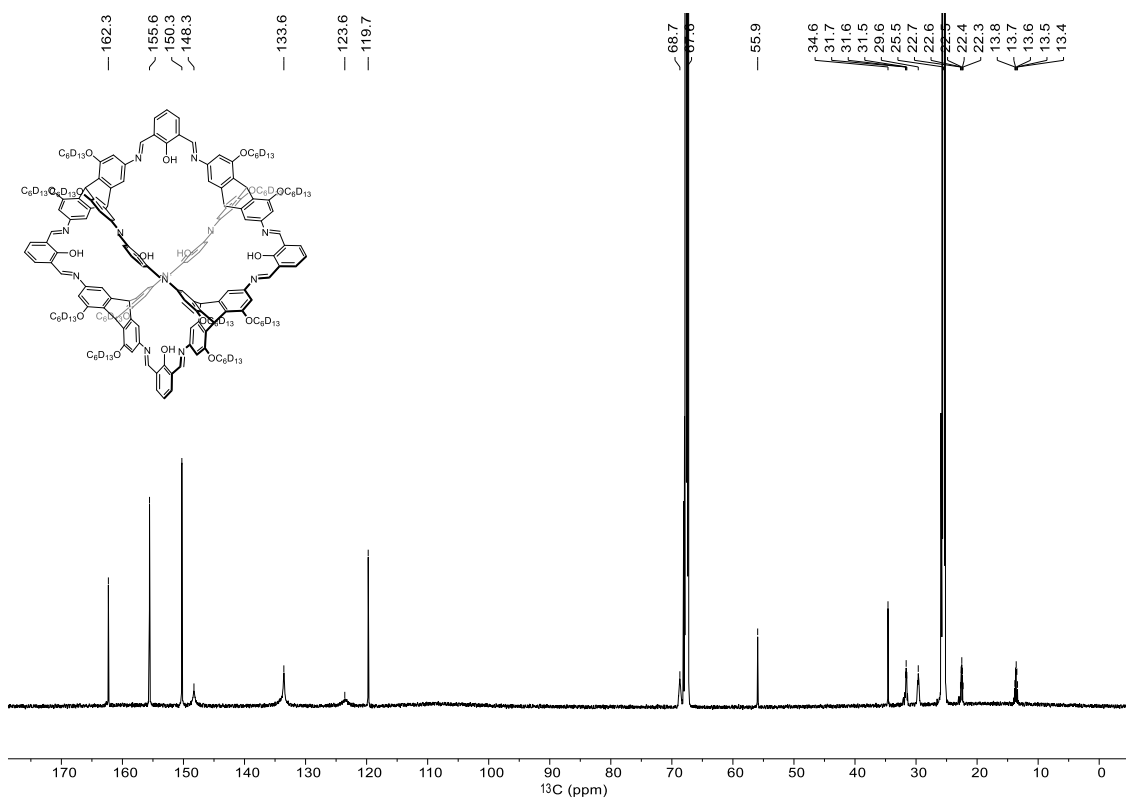


Figure S76. ¹³C{¹H} NMR spectrum (176 MHz, THF-d₈) of [4+6]-endo cage 13-d₁₅₆.

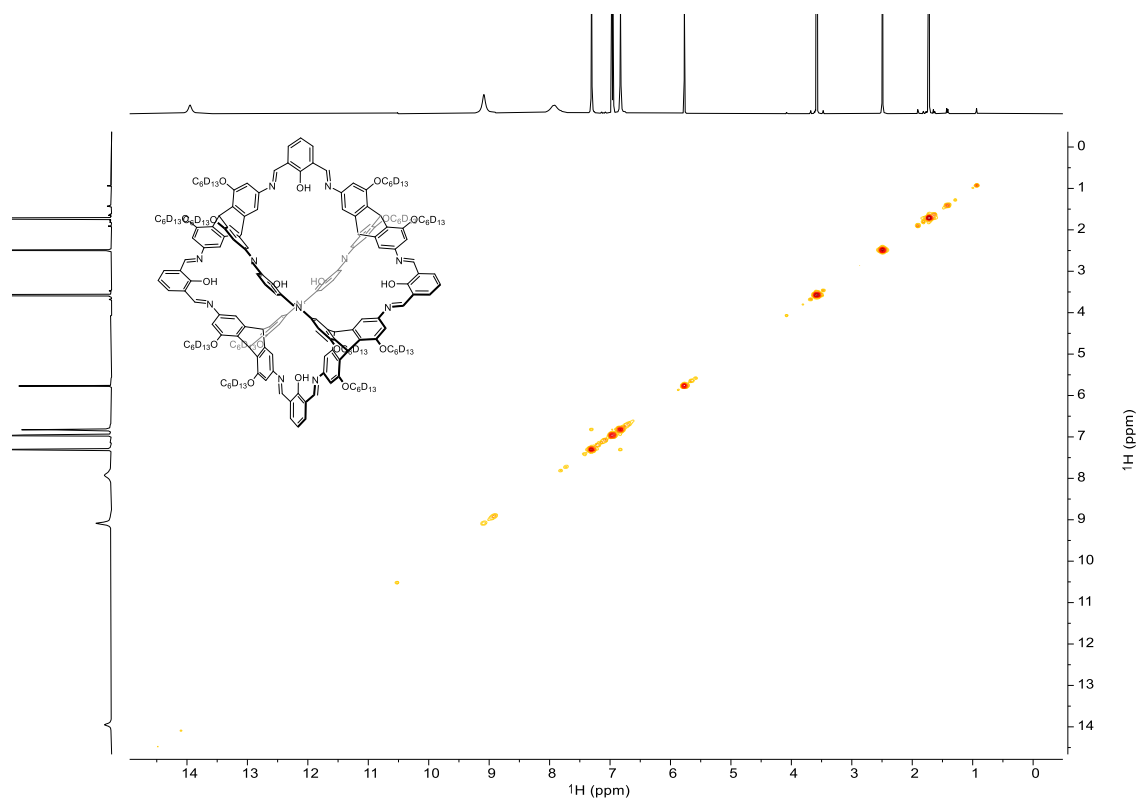


Figure S77. $^1\text{H}, ^1\text{H}$ COSY spectrum (700 MHz, THF-d_8) of [4+6]-*endo* cage **13-d₁₅₆**.

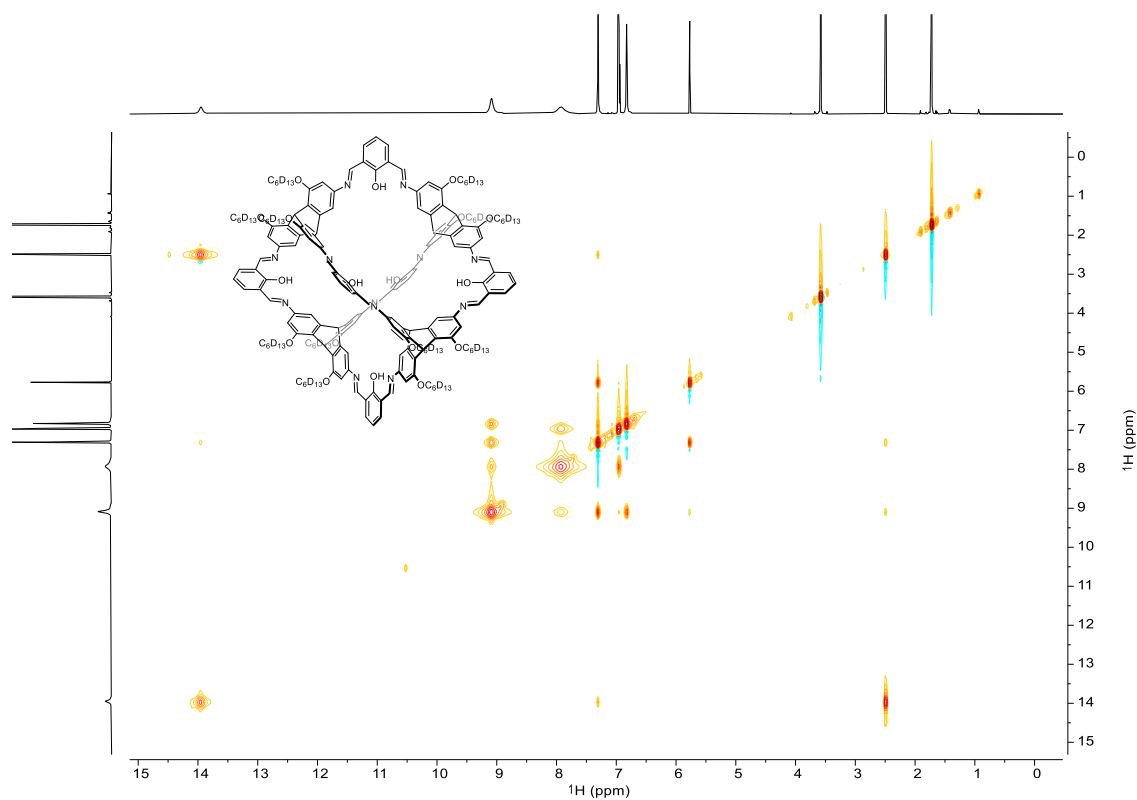


Figure S78. $^1\text{H}, ^1\text{H}$ NOESY spectrum (700 MHz, THF-d_8) of [4+6]-*endo* cage **13-d₁₅₆**.

Table S24. Cross-peak assignment of ^1H , ^1H -COSY and ^1H , ^1H -NOESY spectra of [4+6]-*endo* cage **13-d**₁₅₆.

^1H nucleus	$\delta_{1\text{H}}$ [ppm]	COSY	NOESY
<i>a</i>	13.95	-	<i>d</i> , H ₂ O
<i>b</i>	9.09	-	<i>c</i> , <i>d</i> , <i>g</i>
<i>c</i>	7.93	-	<i>b</i> , <i>f</i>
<i>d</i>	7.31	<i>g</i>	<i>b</i> , <i>h</i>
<i>e</i>	6.97	-	-
<i>f</i>	6.95	-	<i>c</i>
<i>g</i>	6.83	<i>d</i>	<i>b</i>
<i>h</i>	5.77	-	<i>d</i>

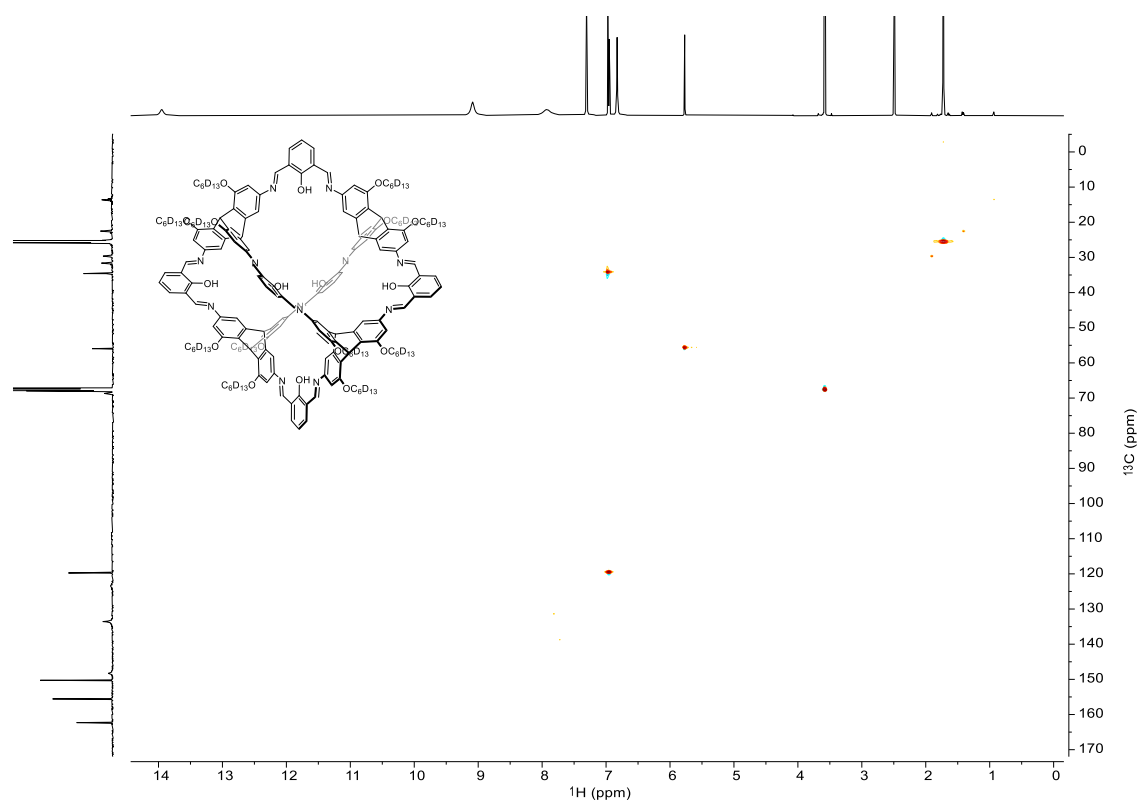


Figure S79. ^1H , ^{13}C HSQC spectrum (700 MHz, 176 MHz, THF-d_8) of [4+6]-*endo* cage **13-d**₁₅₆.

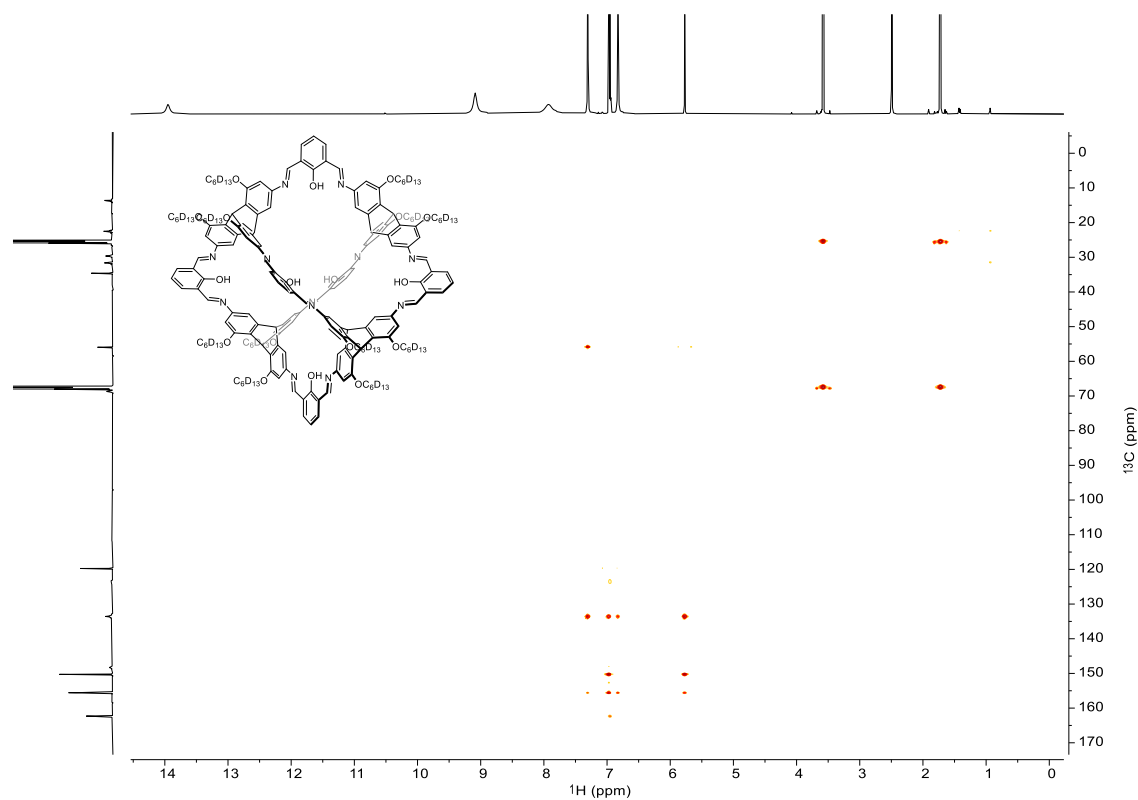


Figure S80. ^1H , ^{13}C HMBC spectrum (700 MHz, 176 MHz, THF-d_8) of [4+6]-*endo* cage **13-d**₁₅₆.

Table S25. Correlations of ^1H , ^{13}C -HSQC and ^1H , ^{13}C -HMBC spectra of [4+6]-*endo* cage **13-d**₁₅₆.

^{13}C nucleus	$\delta_{13\text{C}}$ [ppm]	^1H nucleus		
		HSQC	$\delta_{1\text{H}}$ [ppm]	HMBC
A	162.3	-	-	<i>f</i>
B	155.6	-	-	<i>d, e, g, h</i>
C	150.3	-	-	<i>e, h</i>
D	133.6	-	-	-
E	123.6	-	-	<i>d, e, g, h</i>
F	119.7	<i>f</i>	6.83	-
G	69.1 – 68.3	-	-	-
H	55.9	<i>h</i>	5.77	<i>d</i>
I	34.6	<i>e</i>	6.95	-
J	32.0 – 31.3	-	-	-
K	30.0 – 29.3	-	-	-
L	22.8 – 22.1	-	-	-
M	14.1 – 13.2	-	-	-

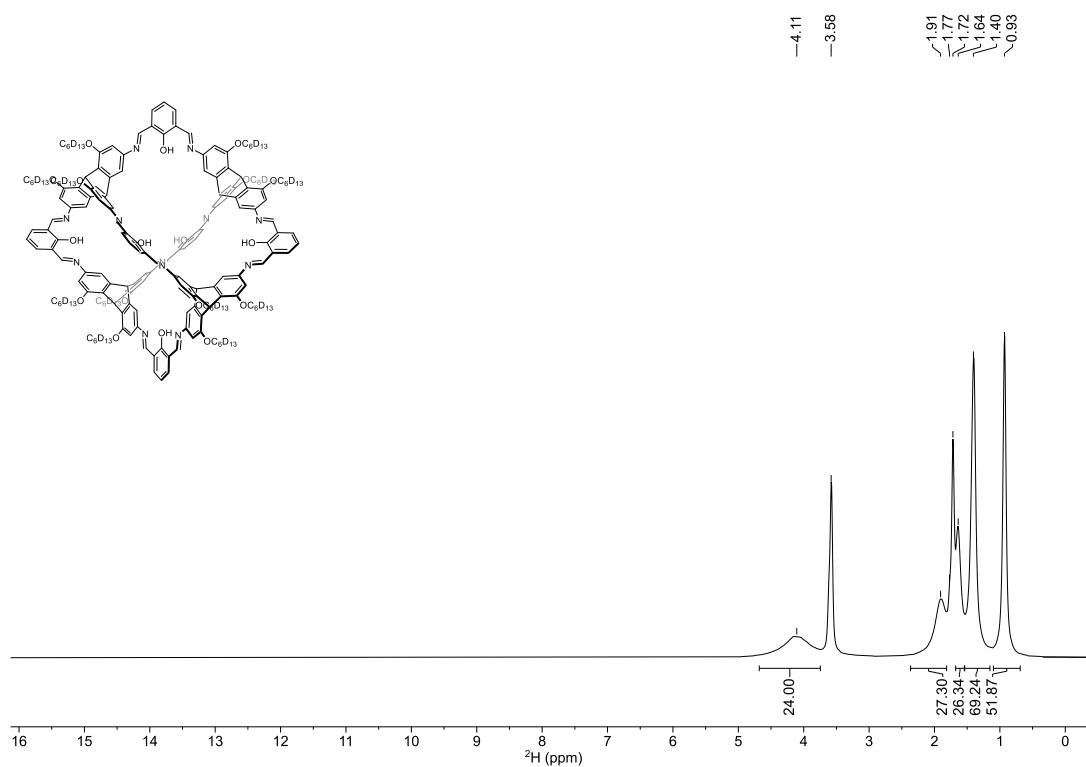
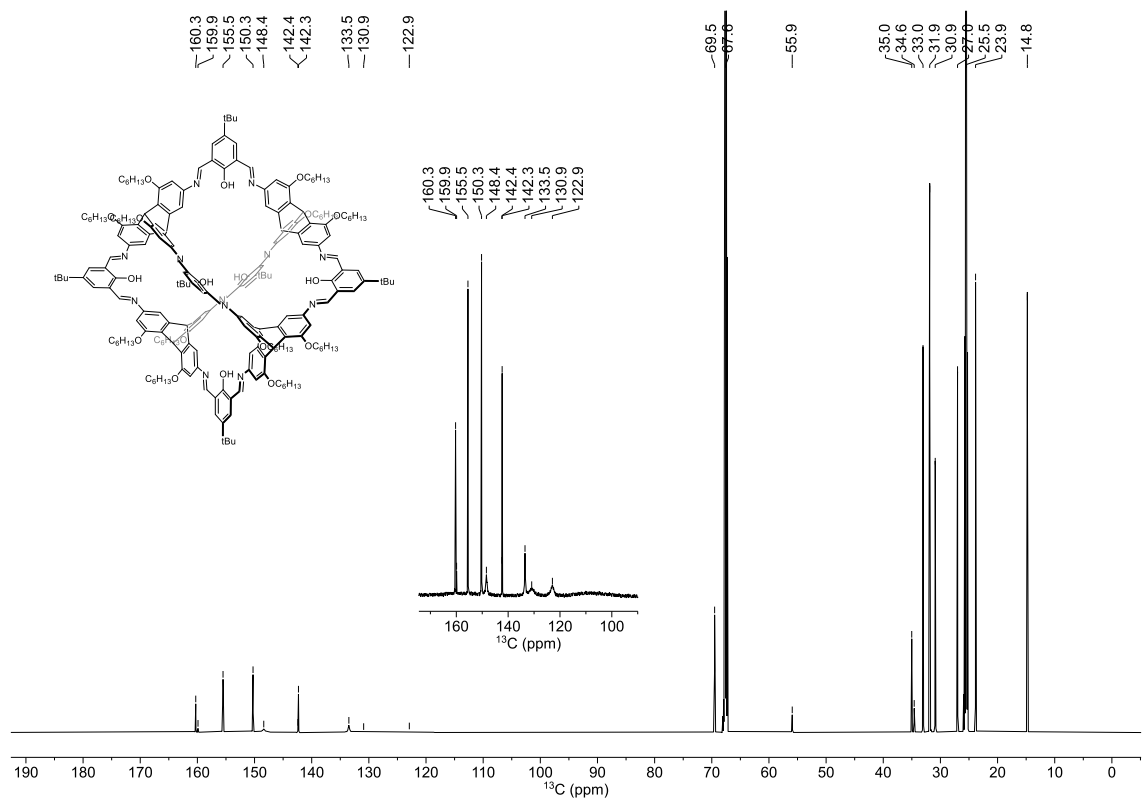
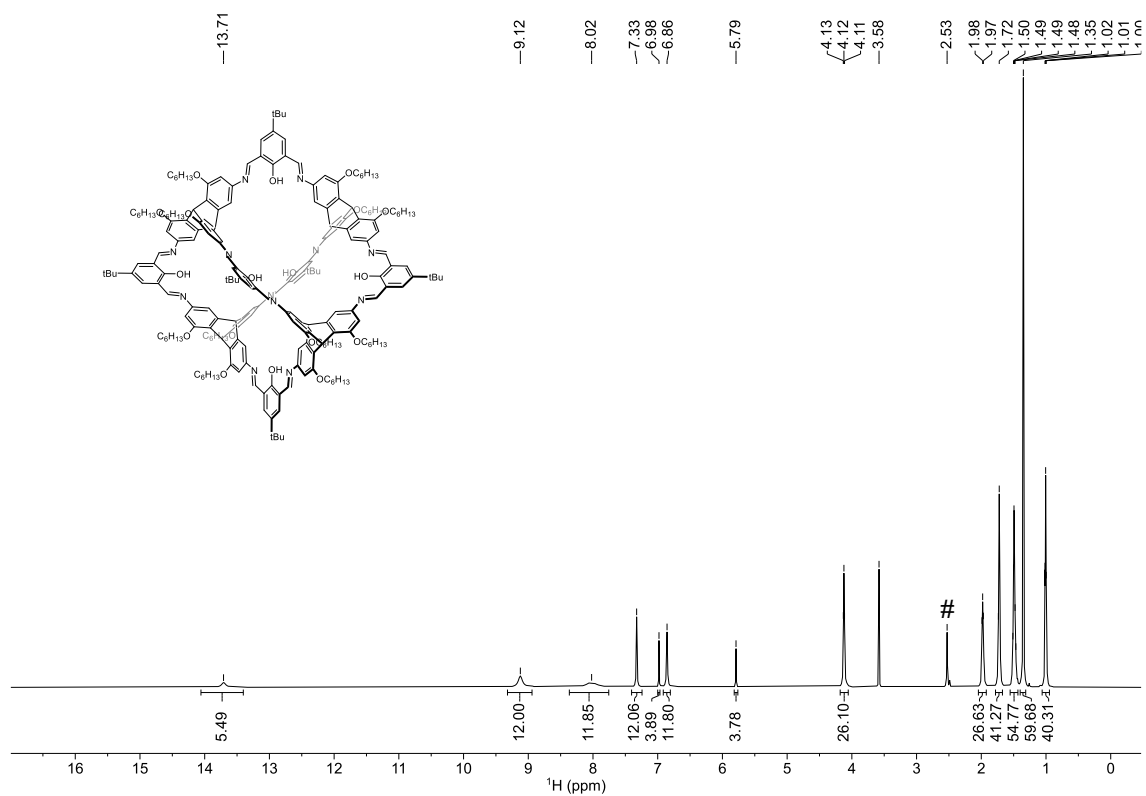


Figure S81. ^2H NMR spectrum (92 MHz, THF/THF- d_8) of [4+6]-*endo* cage **13-d**₁₅₆.



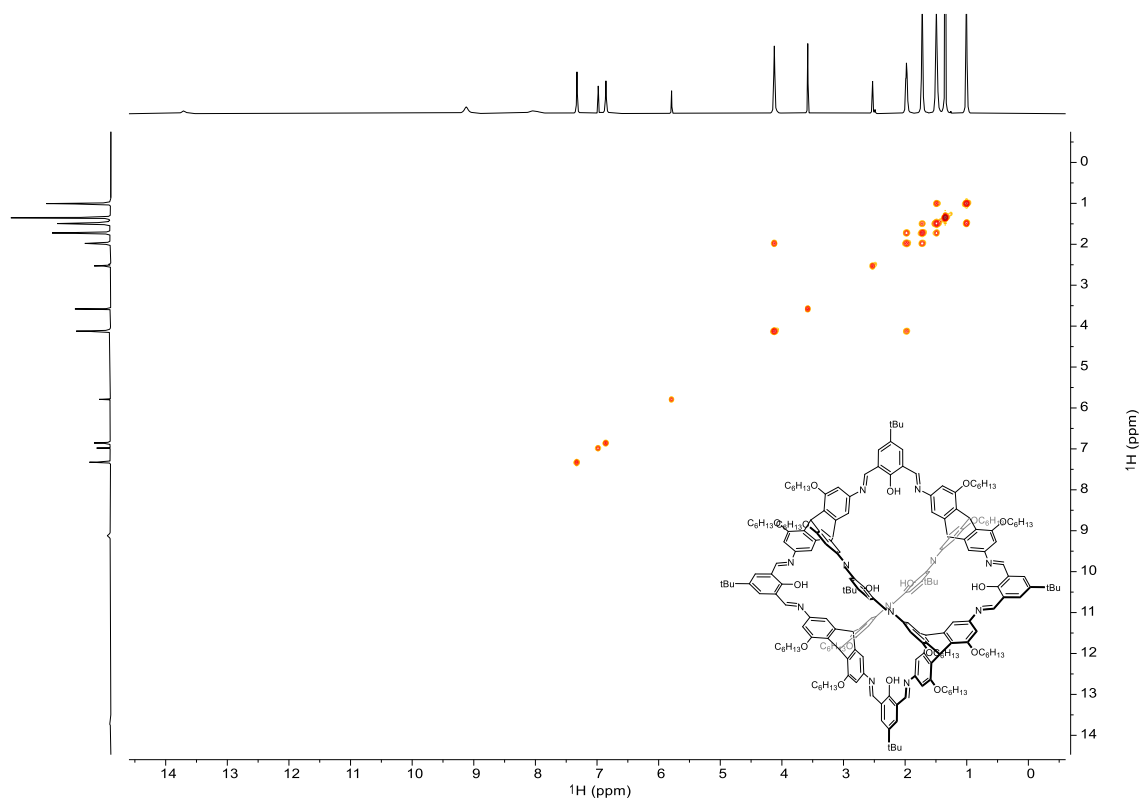


Figure S84. $^1\text{H}, ^1\text{H}$ COSY spectrum (700 MHz, THF-d_8) of *tert*-butylated [4+6]-*endo* cage **14**.

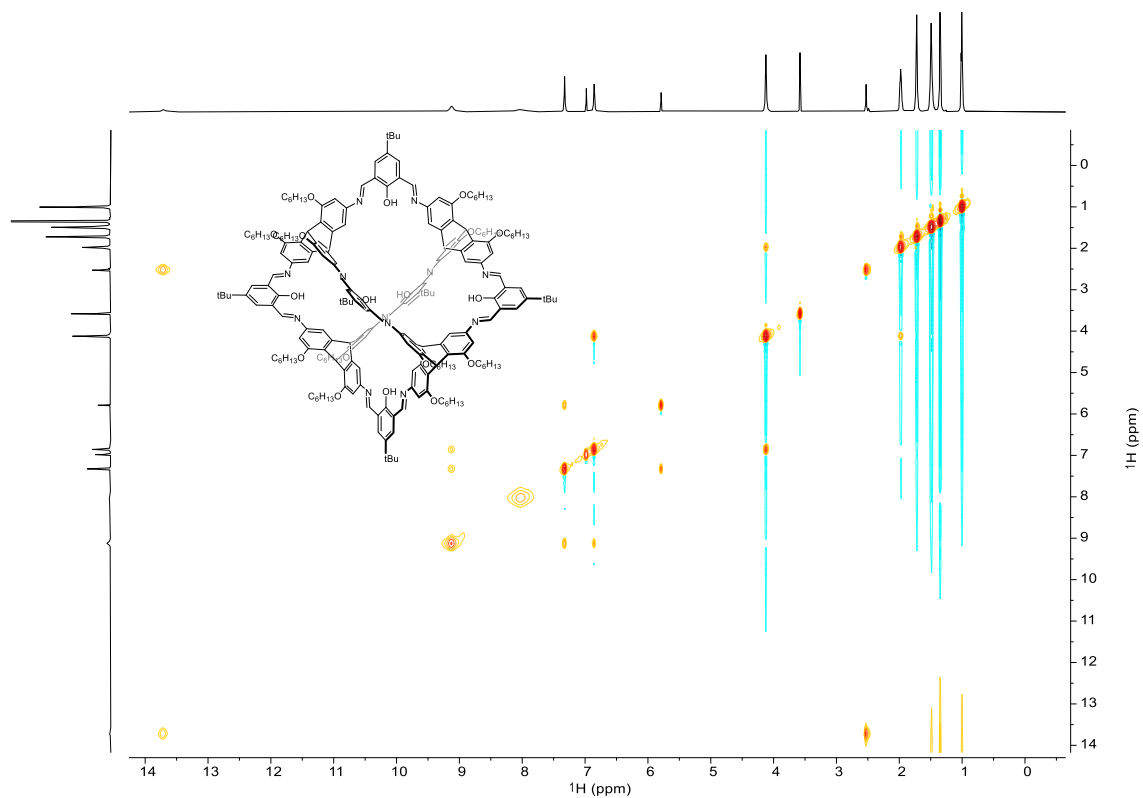


Figure S85. $^1\text{H}, ^1\text{H}$ NOESY spectrum (700 MHz, THF-d_8) of *tert*-butylated [4+6]-*endo* cage **14**.

Table S26. Cross-peak assignment of ^1H , ^1H -COSY and ^1H , ^1H -NOESY spectra of *tert*-butylated [4+6]-*endo* cage 14.

^1H nucleus	$\delta_{1\text{H}}$ [ppm]	COSY	NOESY
<i>a</i>	13.71	-	H ₂ O
<i>b</i>	9.12	-	<i>d, f</i>
<i>c</i>	8.02	-	-
<i>d</i>	7.33	-	<i>b, g</i>
<i>e</i>	6.98	-	-
<i>f</i>	6.86	-	<i>b, h</i>
<i>g</i>	5.79	-	<i>d</i>
<i>h</i>	4.12	<i>i</i>	<i>f, i</i>
<i>i</i>	2.04 – 1.92	<i>h, j</i>	<i>h</i>
<i>j</i>	1.78 – 1.67	<i>i, k</i>	-
<i>k</i>	1.55 – 1.43	<i>j, m</i>	-
<i>l</i>	1.35	-	-
<i>m</i>	1.06 – 0.95	<i>k</i>	-

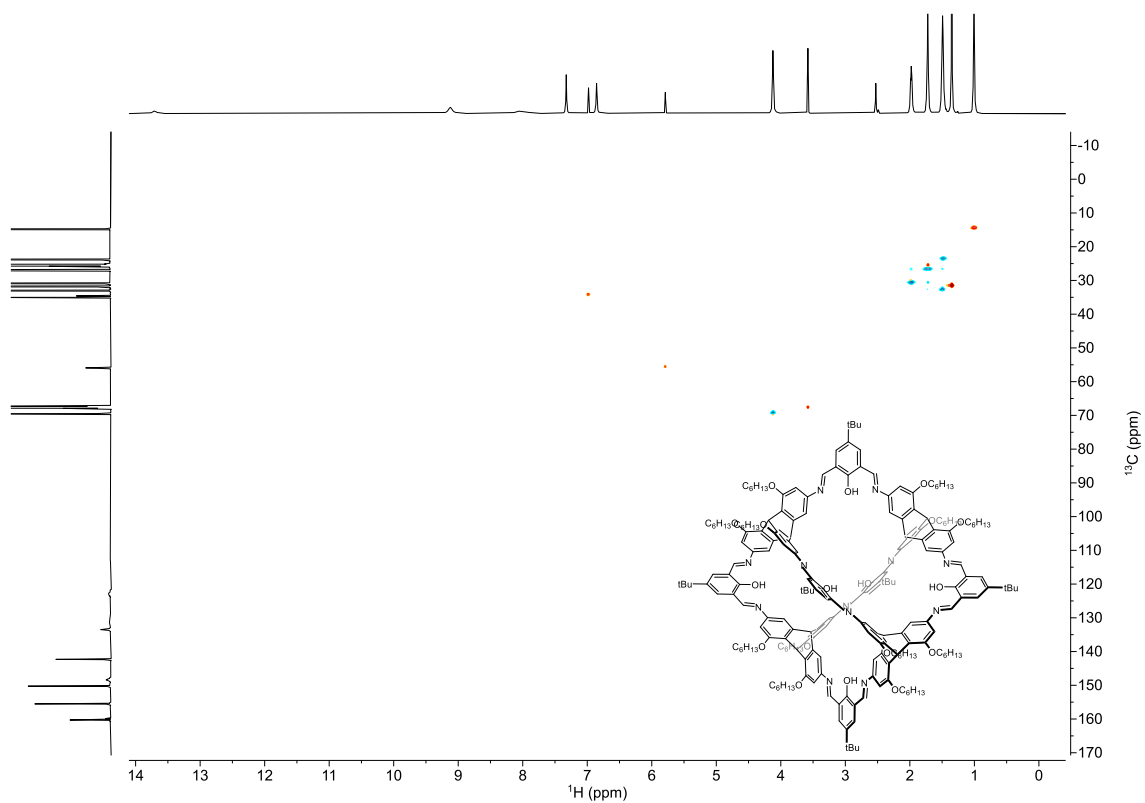


Figure S86. ^1H , ^{13}C HSQC spectrum (700 MHz, 176 MHz, THF-d_8) of *tert*-butylated [4+6]-endo cage **14**.

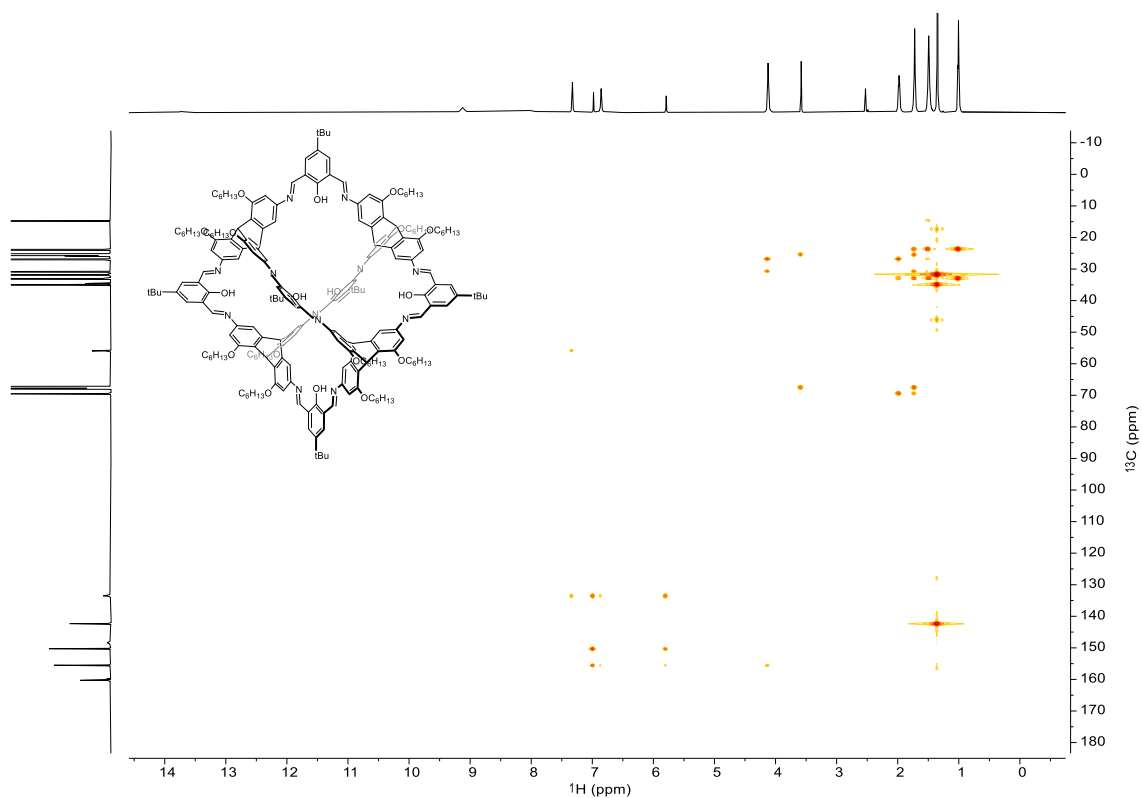


Figure S87. ^1H , ^{13}C HMBC spectrum (700 MHz, 176 MHz, THF-d_8) of *tert*-butylated [4+6]-endo cage **14**.

Table S27. Correlations of ^1H , ^{13}C -HSQC and ^1H , ^{13}C -HMBC spectra of *tert*-butylated [4+6]-*endo* cage **14**.

^{13}C nucleus	$\delta_{13\text{C}}$ [ppm]	^1H nucleus		
		HSQC	$\delta_{1\text{H}}$ [ppm]	HMBC
<i>A</i>	160.3	-	-	-
<i>B</i>	159.9	-	-	-
<i>C</i>	155.5	-	-	<i>d, e, g, h</i>
<i>D</i>	150.3	-	-	<i>e, g</i>
<i>E</i>	148.4	-	-	-
<i>F</i>	142.4	-	-	<i>l</i>
<i>G</i>	142.3	-	-	<i>l</i>
<i>H</i>	133.5	-	-	<i>d, e, f, g</i>
<i>I</i>	130.9	-	-	-
<i>J</i>	122.9	-	-	-
<i>K</i>	69.5	<i>h</i>	4.12	<i>i, j</i>
<i>L</i>	55.9	<i>g</i>	5.79	<i>d</i>
<i>M</i>	35.0	<i>e</i>	6.98	<i>l</i>
<i>O</i>	34.6	-	-	-
<i>P</i>	33.0	<i>k</i>	1.55 – 1.43	<i>i, j, k, m</i>
<i>Q</i>	31.9	<i>l</i>	1.35	<i>l</i>
<i>R</i>	30.9	<i>i</i>	2.04 – 1.92	<i>h, j</i>
<i>S</i>	27.0	<i>j</i>	1.78 – 1.67	<i>h, i, k</i>
<i>T</i>	23.9	<i>k</i>	1.55 – 1.43	<i>j, k, m</i>
<i>U</i>	14.8	<i>m</i>	1.06 – 0.95	<i>k</i>

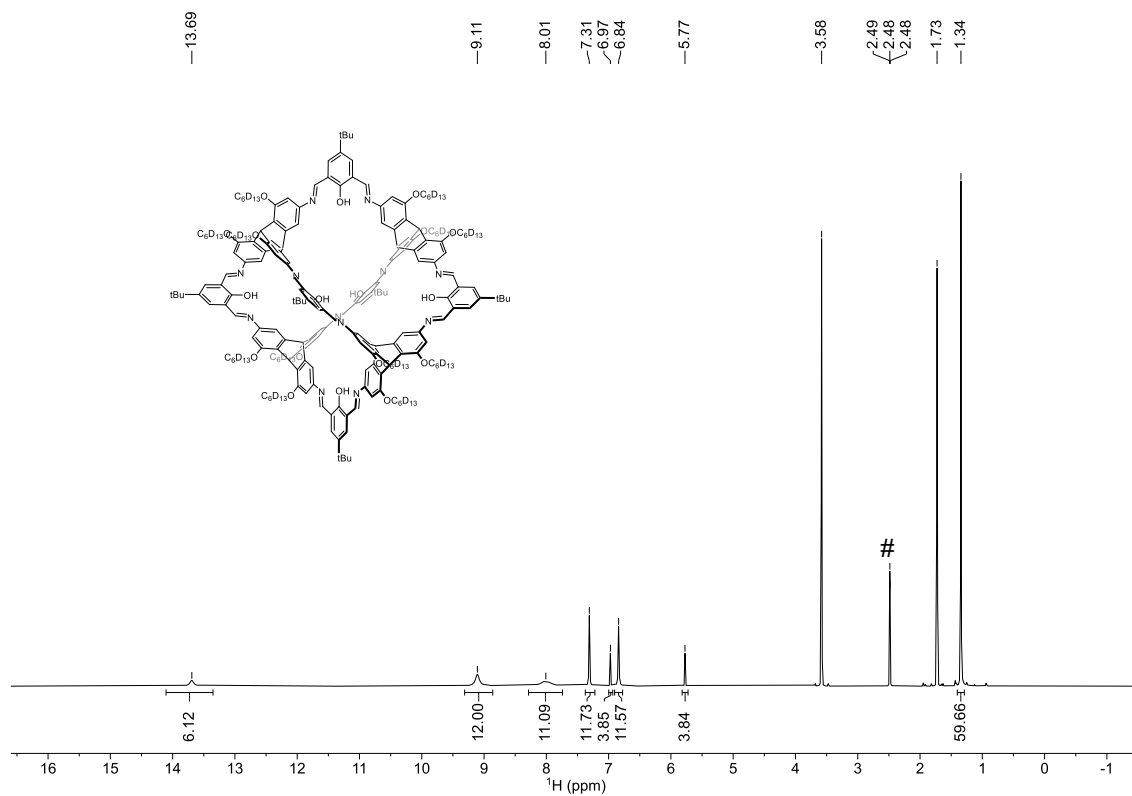


Figure S88. ¹H NMR spectrum (700 MHz, THF-d₈) of *tert*-butylated [4+6]-endo cage **14-d₁₅₆**. #: H₂O.

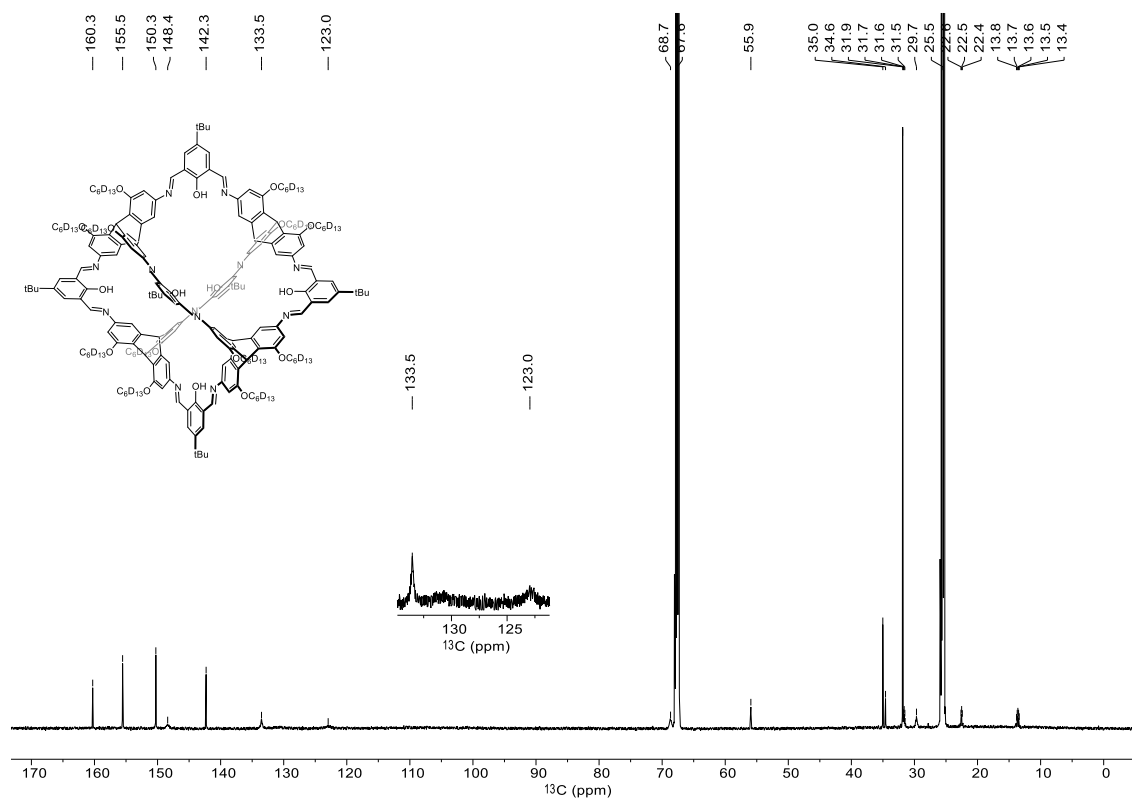


Figure S89. ¹³C{¹H} NMR spectrum (176 MHz, THF-d₈) of *tert*-butylated [4+6]-endo cage **14-d₁₅₆**.

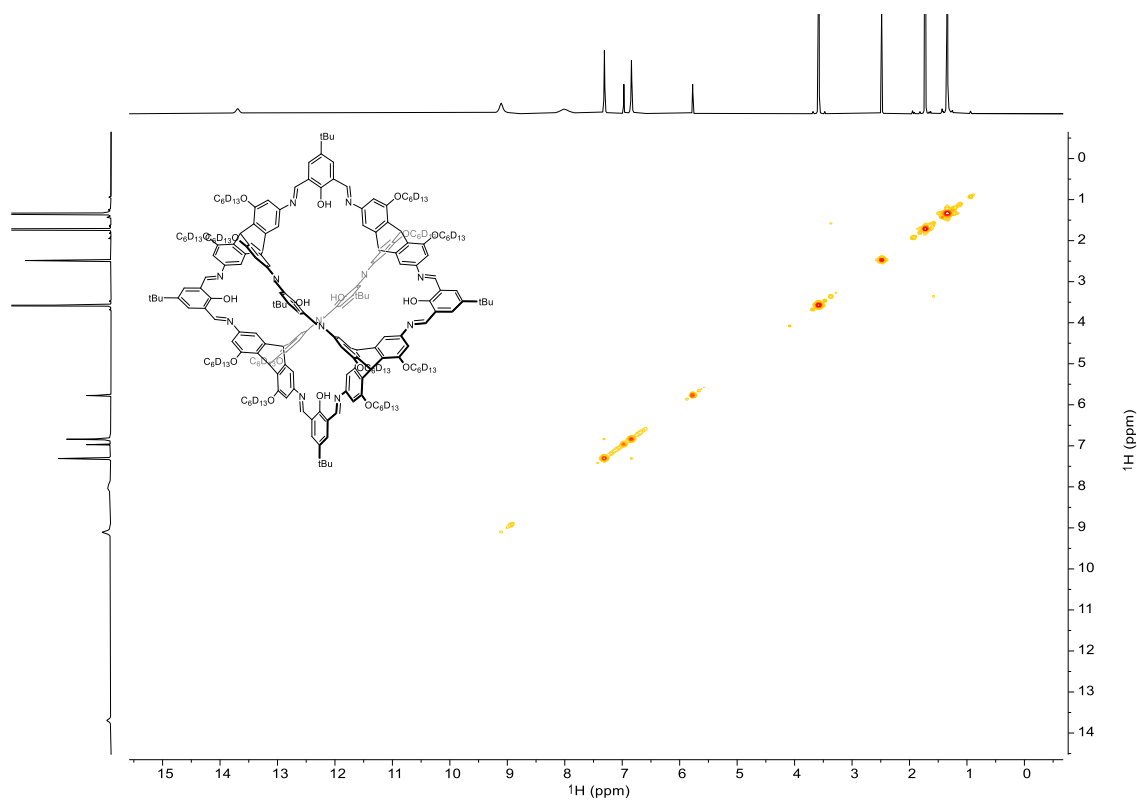


Figure S90. $^1\text{H}, ^1\text{H}$ COSY spectrum (700 MHz, THF-d_8) of *tert*-butylated [4+6]-*endo* cage **14-d**₁₅₆.

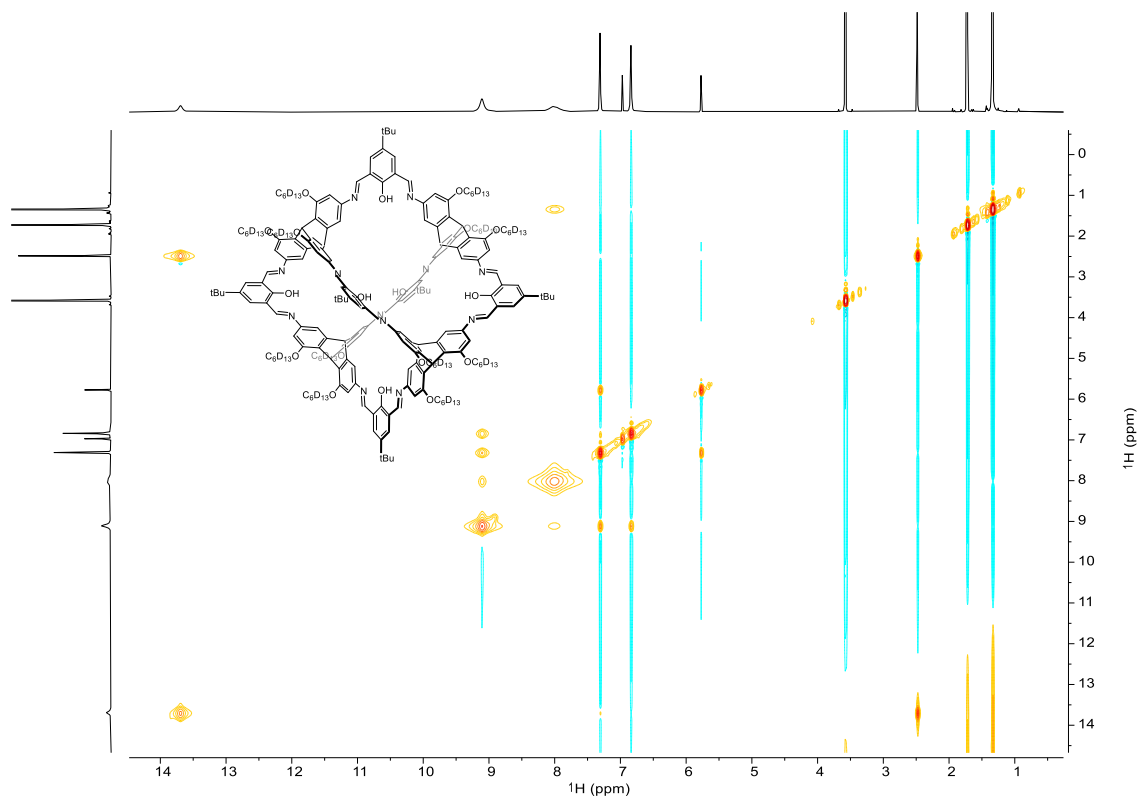


Figure S91. $^1\text{H}, ^1\text{H}$ NOESY spectrum (700 MHz, THF-d_8) of *tert*-butylated [4+6]-*endo* cage **14-d**₁₅₆.

Table S28. Cross-peak assignment of ^1H , ^1H -COSY and ^1H , ^1H -NOESY spectra of *tert*-butylated [4+6]-*endo* cage **14-d**₁₅₆.

^1H nucleus	$\delta_{1\text{H}}$ [ppm]	COSY	NOESY
<i>a</i>	13.69	-	H ₂ O
<i>b</i>	9.11	-	<i>c, d, f</i>
<i>c</i>	8.01	-	<i>h</i>
<i>d</i>	7.31	-	<i>b, g</i>
<i>e</i>	6.97	-	-
<i>f</i>	6.84	-	<i>b</i>
<i>g</i>	5.77	-	<i>d</i>
<i>h</i>	1.34	-	<i>c</i>

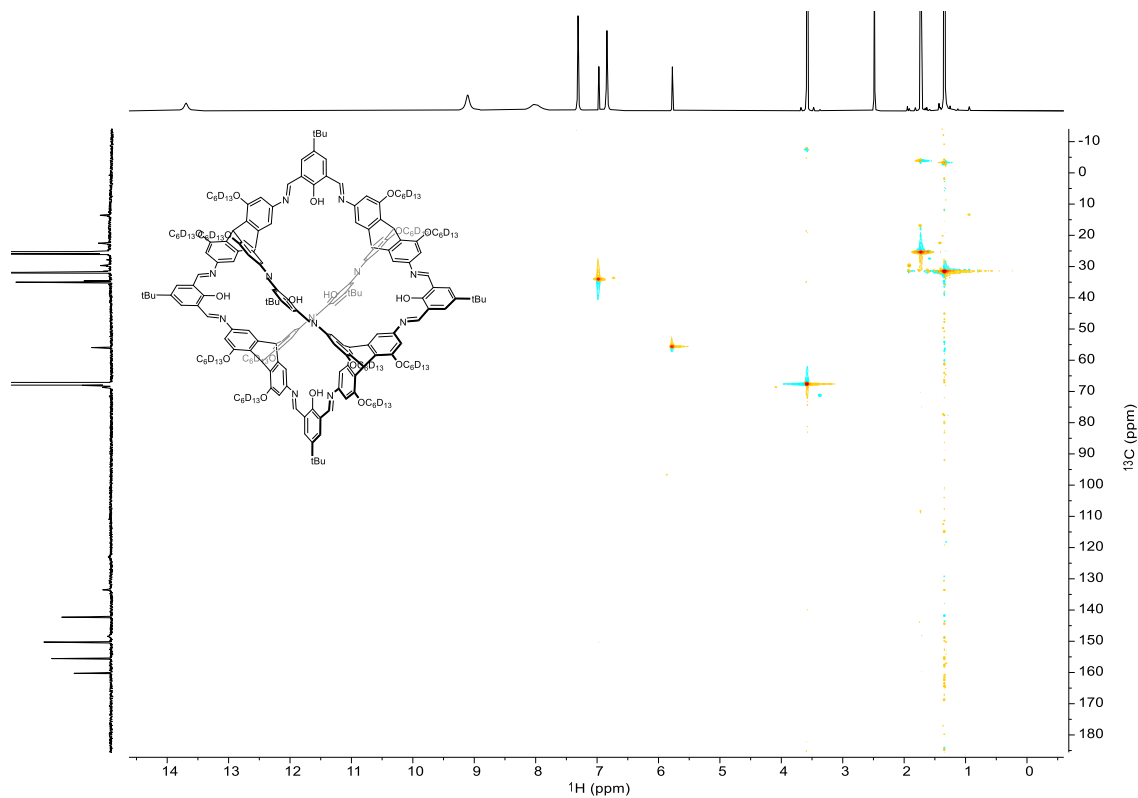


Figure S92. ^1H , ^{13}C HSQC spectrum (700 MHz, 176 MHz, THF-d_8) of *tert*-butylated [4+6]-*endo* cage **14-d156**.

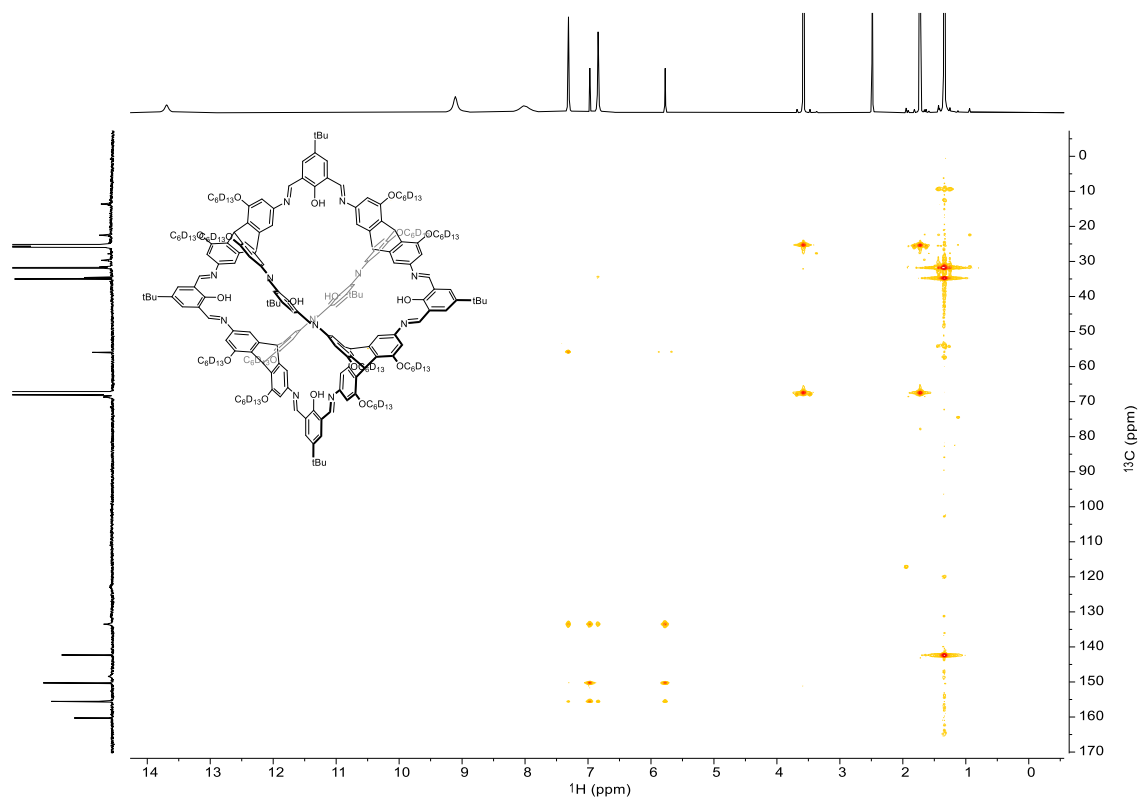


Figure S93. ^1H , ^{13}C HMBC spectrum (700 MHz, 176 MHz, THF-d_8) of *tert*-butylated [4+6]-*endo* cage **14-d156**.

Table S29. Correlations of ^1H , ^{13}C -HSQC and ^1H , ^{13}C -HMBC spectra of *tert*-butylated [4+6]-*endo* cage **14-d**₁₅₆.

^{13}C nucleus	$\delta_{13\text{C}}$ [ppm]	^1H nucleus		
		HSQC	$\delta_{1\text{H}}$ [ppm]	HMBC
A	160.3	-	-	-
B	155.5	-	-	<i>d, e, f, g</i>
C	150.3	-	-	<i>e, g</i>
D	148.4	-	-	-
E	142.3	-	-	<i>h</i>
F	133.5	-	-	<i>d, e, f, g</i>
G	123.0	-	-	-
H	69.0 – 68.3	-	-	-
I	55.9	<i>g</i>	5.77	<i>d</i>
J	35.0	-	-	<i>h</i>
K	34.6	<i>e</i>	6.97	<i>f</i>
L	31.9	<i>h</i>	1.34	<i>h</i>
M	31.8 – 31.3	-	-	-
N	30.0 – 29.3	-	-	-
O	22.9 – 22.05	-	-	-
P	14.01 – 13.2	-	-	-

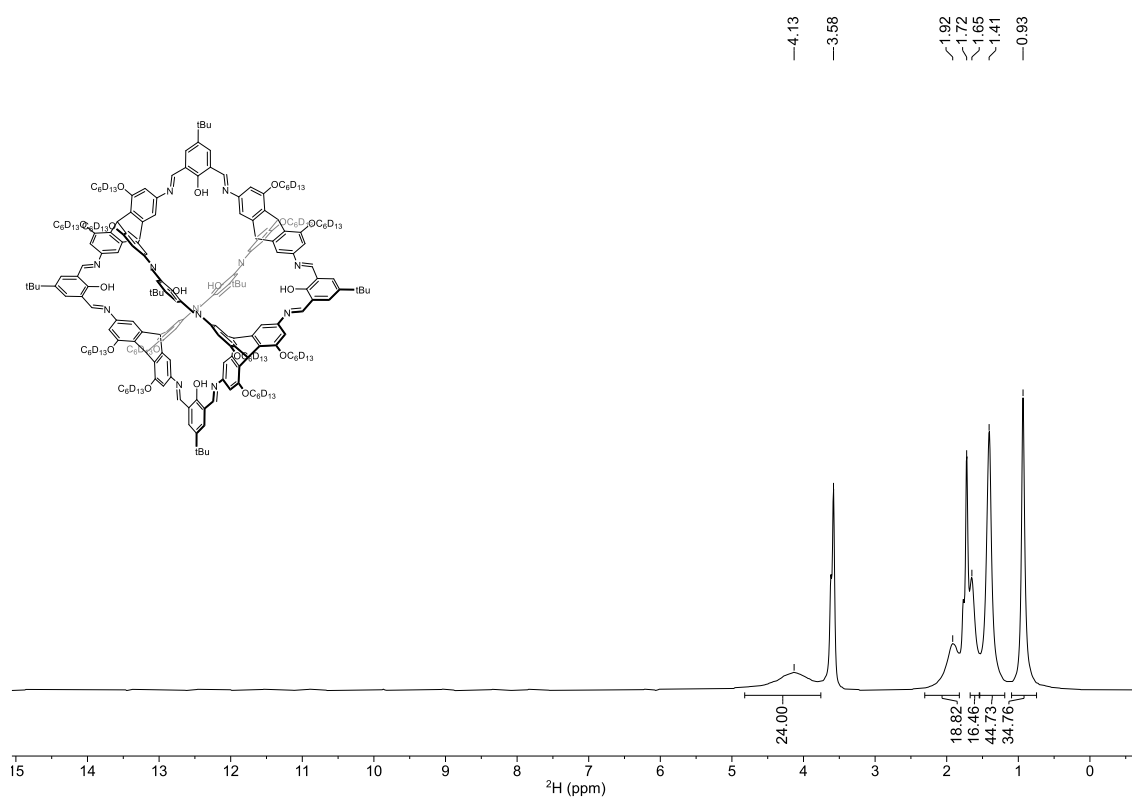


Figure S94. ^2H NMR spectrum (92 MHz, THF/THF- d_8) of *tert*-butylated [4+6]-*endo* cage **14-d**₁₅₆.

4 DOSY Spectra

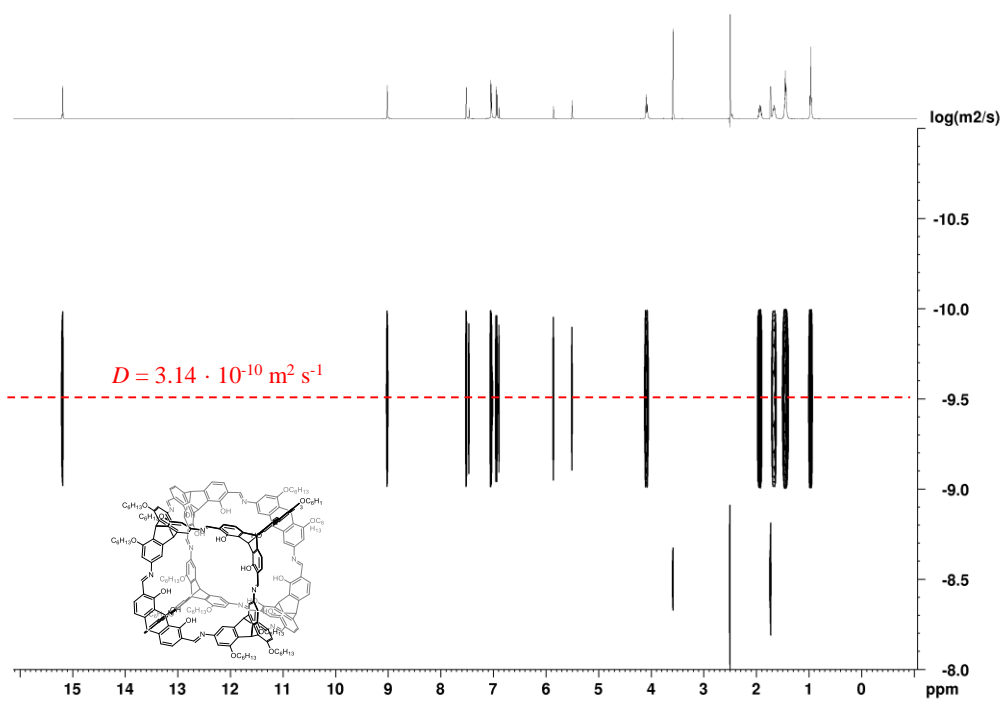


Figure S95. ¹H-DOSY spectrum (400 MHz, THF-d₈) of [4+4] cube 11.

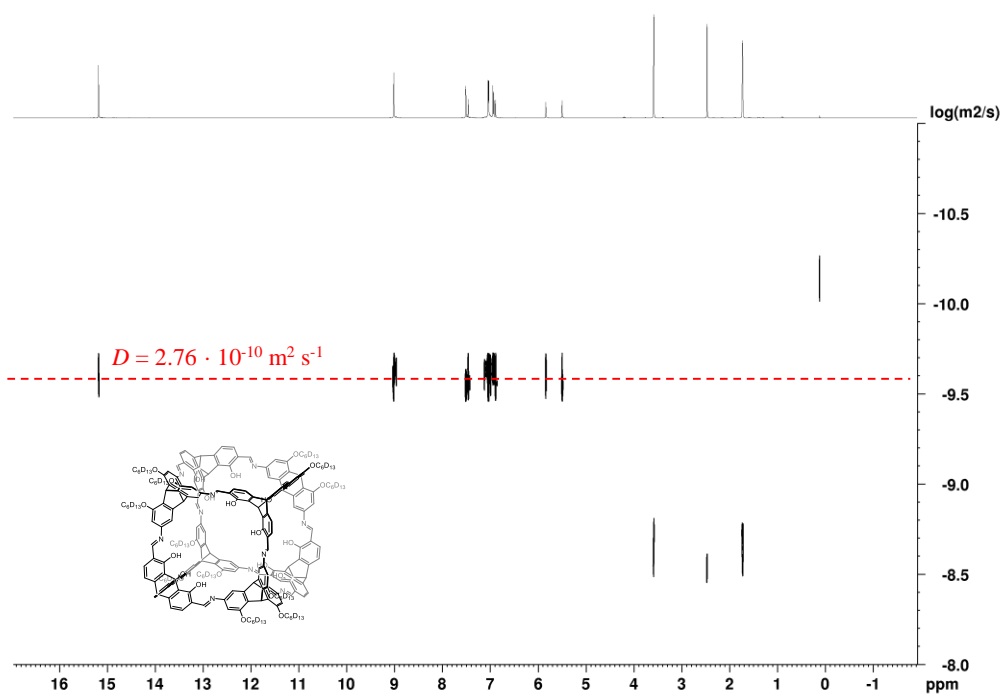


Figure S96. ¹H-DOSY spectrum (400 MHz, THF-d₈) of [4+4] cube 11-d₁₅₆.

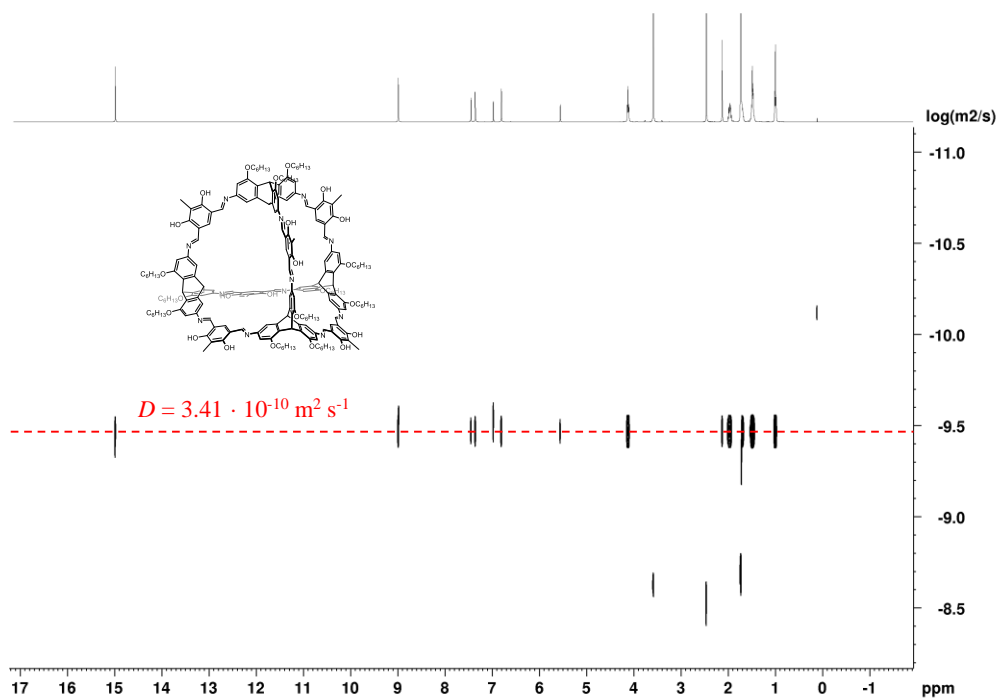


Figure S97. ^1H -DOSY spectrum (400 MHz, THF-d_8) of [4+6]-exo cage 12.

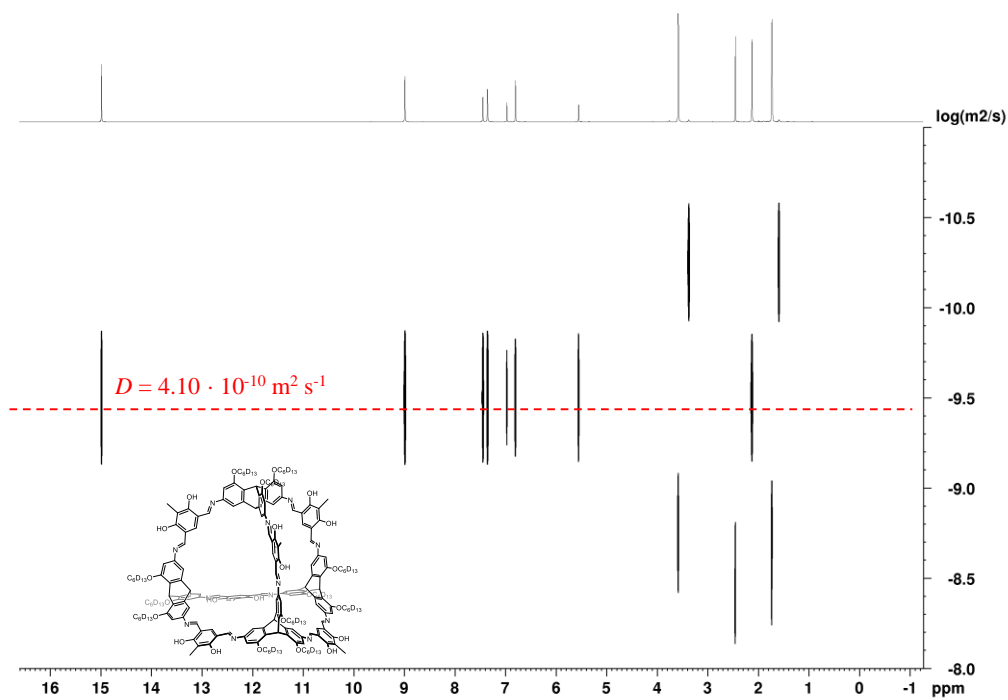


Figure S98. ^1H -DOSY spectrum (400 MHz, THF-d_8) of [4+6]-exo cage 12- d_{156} .

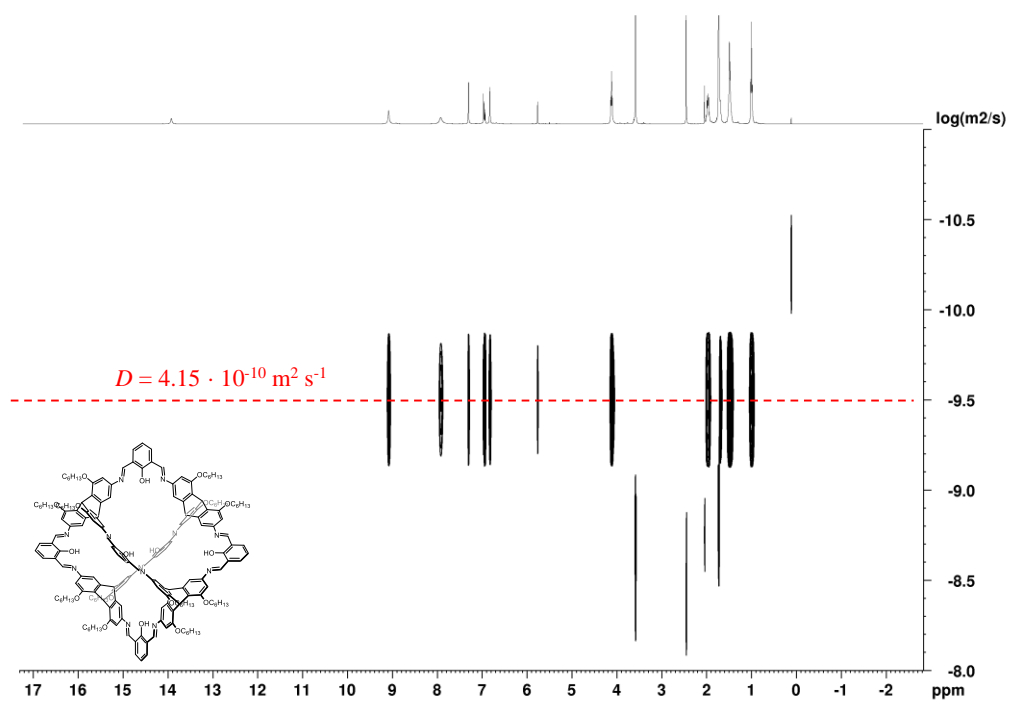


Figure S99. ^1H -DOSY spectrum (400 MHz, THF-d_8) of [4+6]-*endo* cage **13**.

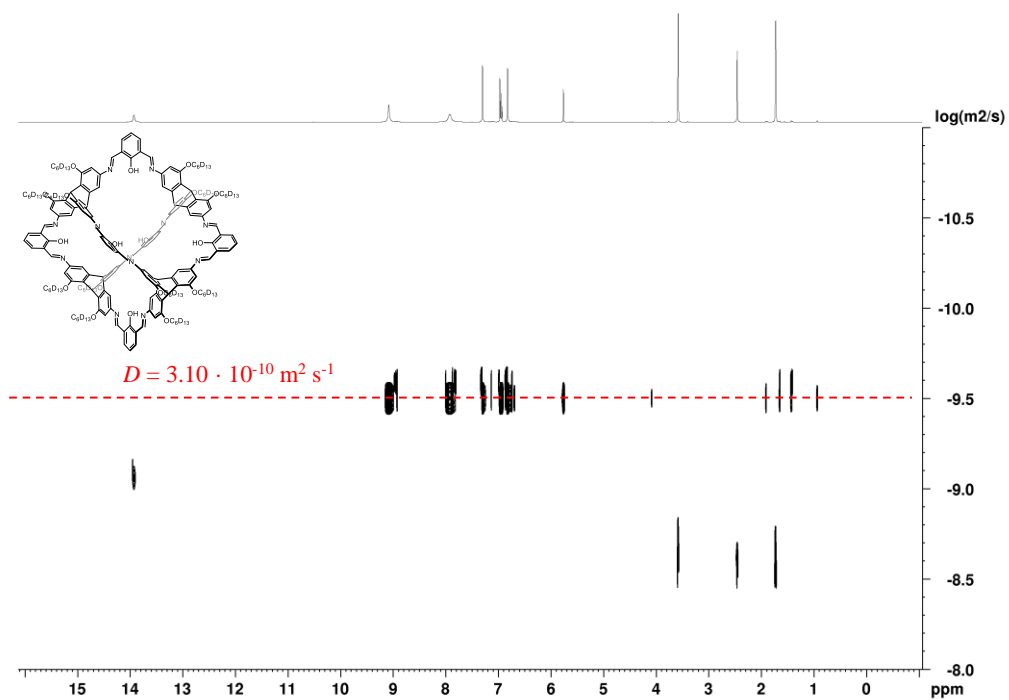


Figure S100. ^1H -DOSY spectrum (400 MHz, THF-d_8) of [4+6]-*endo* cage **13-d**₁₅₆.

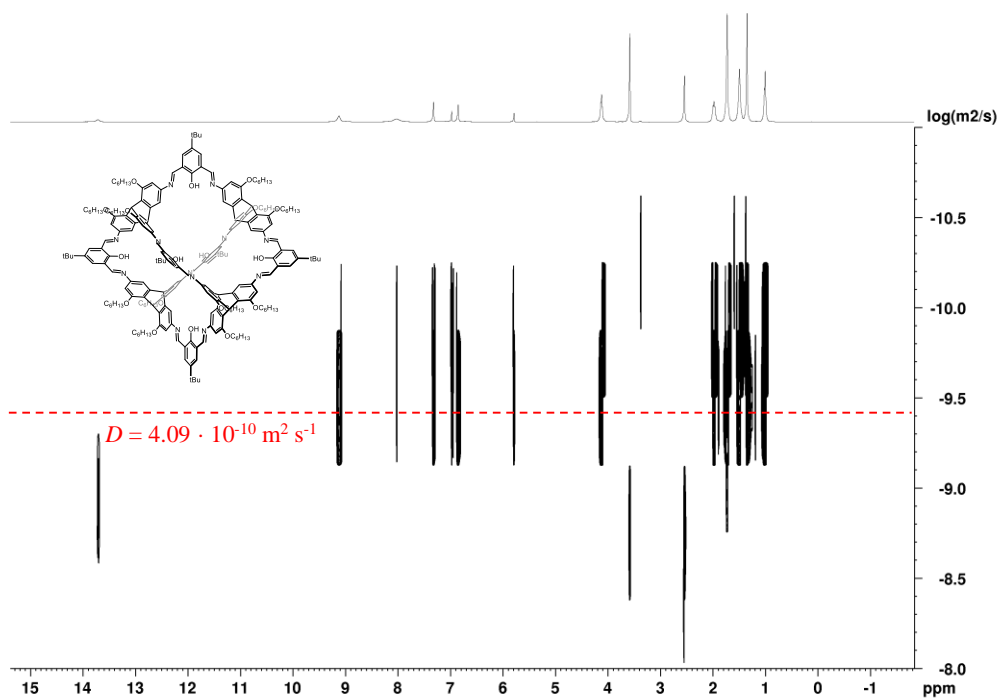


Figure S101. ^1H -DOSY spectrum (400 MHz, THF-d_8) of *tert*-butylated [4+6]-*endo* cage **14**.

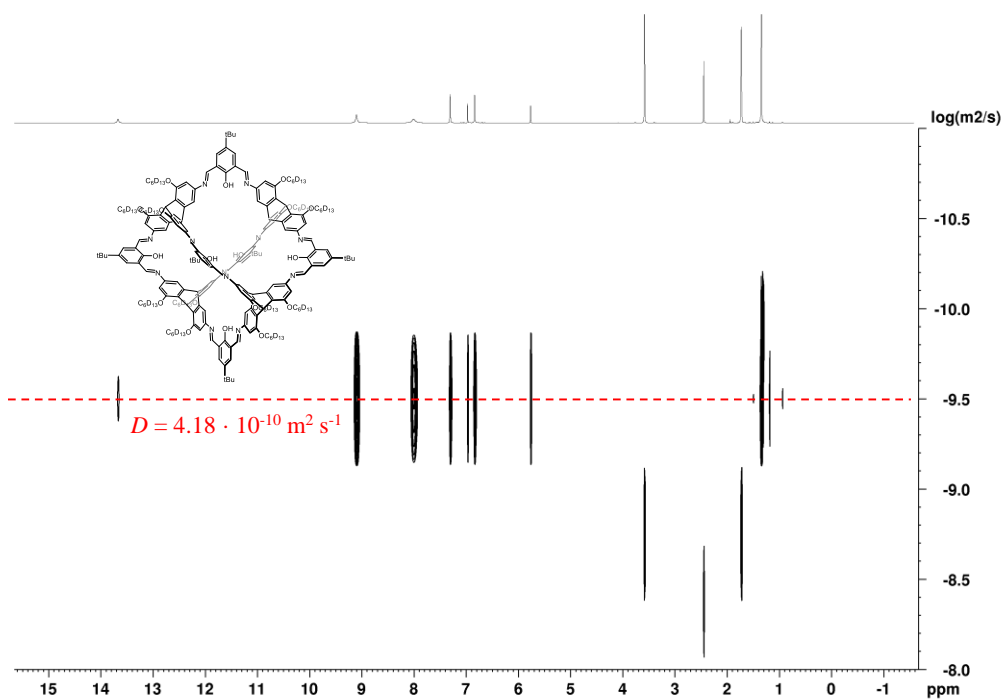


Figure S102. ^1H -DOSY spectrum (400 MHz, THF-d_8) of *tert*-butylated [4+6]-*endo* cage **14-d**₁₅₆.

5 Mass Spectra of Cage Compounds

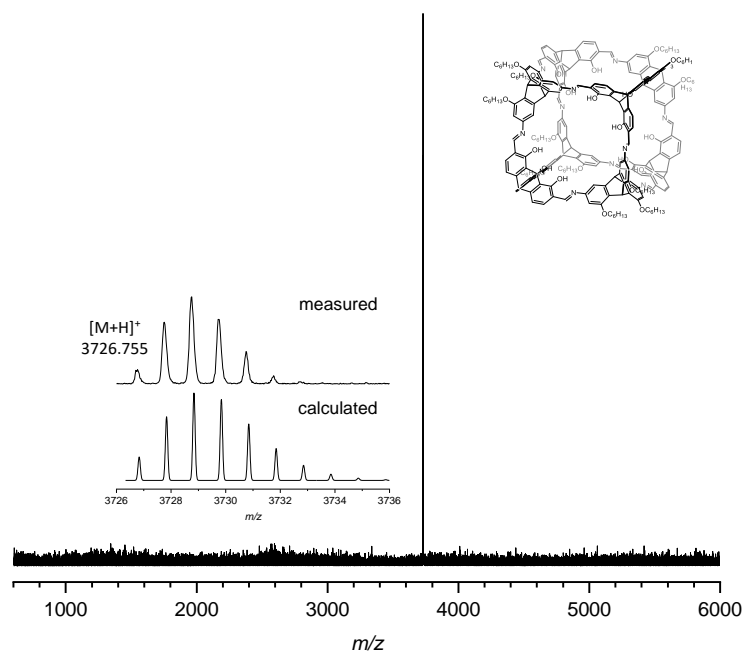


Figure S103. MALDI-TOF mass spectrum of the [4+4] cube **11** (DCTB matrix, reflector mode, positive polarisation).

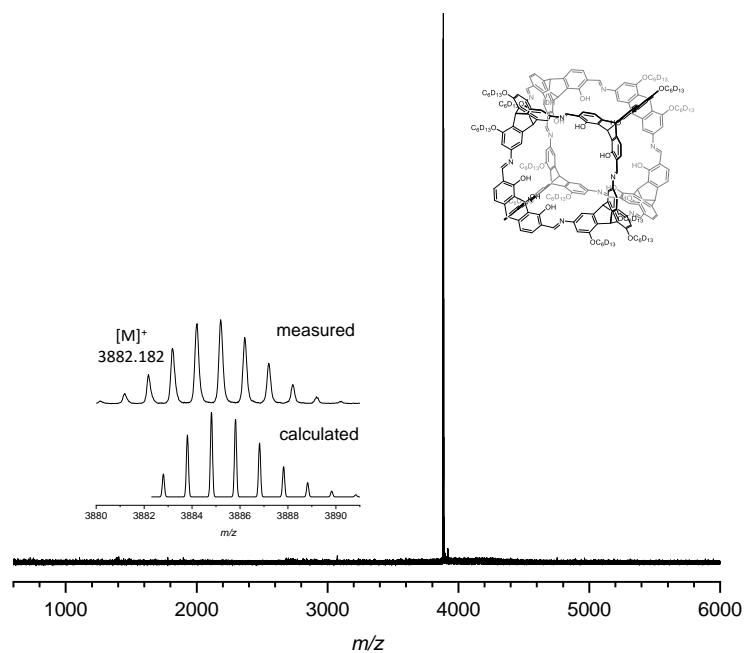


Figure S104. MALDI-TOF mass spectrum of the [4+4] cube **11-d₁₅₆** (DCTB matrix, reflector mode, positive polarisation).

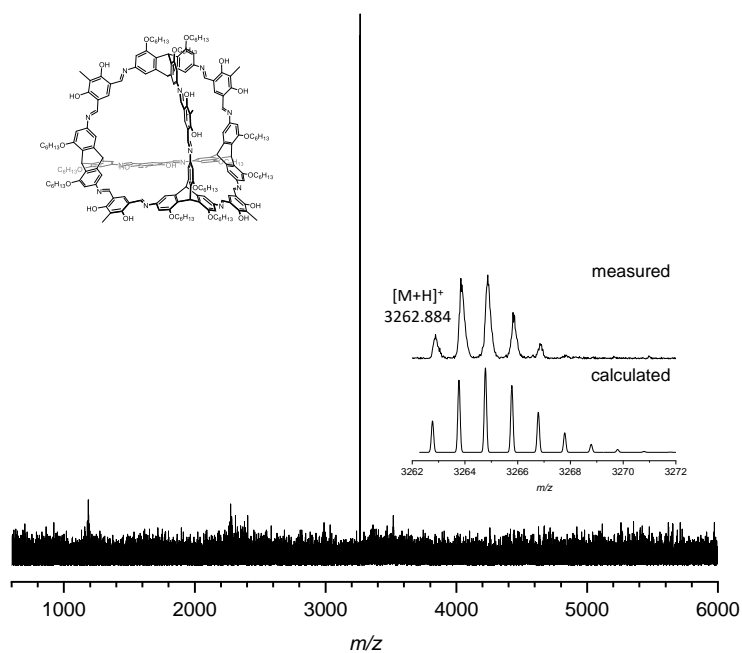


Figure S105. MALDI-TOF mass spectrum of the [4+6]-exo cage **12** (DCTB matrix, reflector mode, positive polarisation).

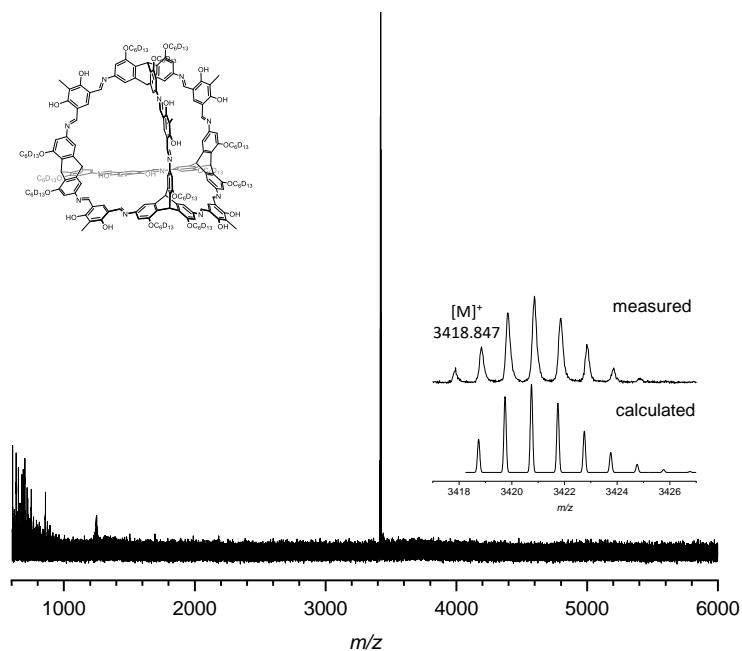


Figure S106. MALDI-TOF mass spectrum of the [4+6]-exo cage **12-d₁₅₆** (DCTB matrix, reflector mode, positive polarisation).

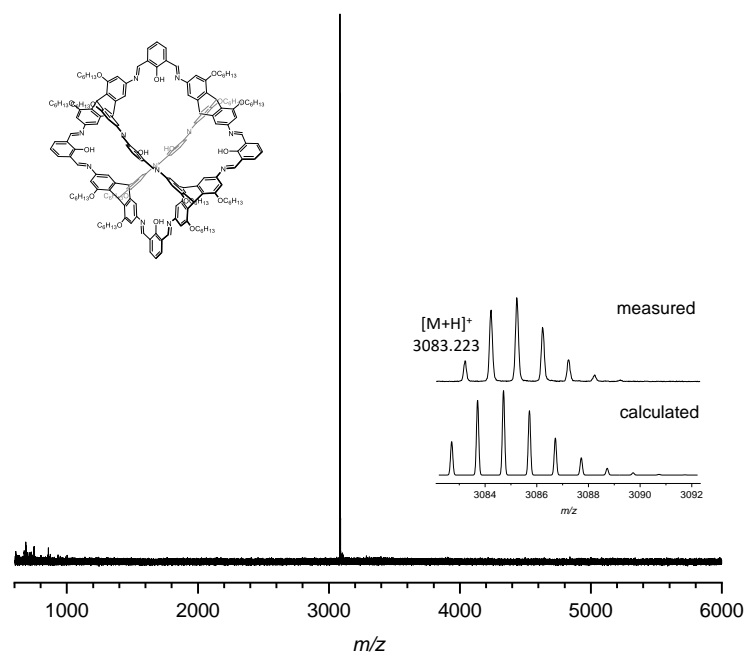


Figure S107. MALDI-TOF mass spectrum of the [4+6]-*endo* cage **13** (DCTB matrix, reflector mode, positive polarisation).

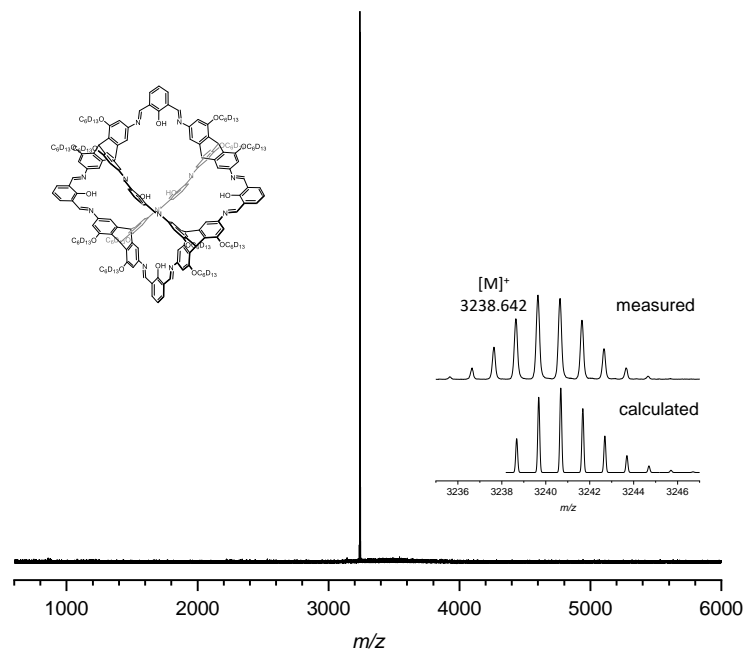


Figure S108. MALDI-TOF mass spectrum of the [4+6]-*endo* cage **13-d₁₅₆** (DCTB matrix, reflector mode, positive polarisation).

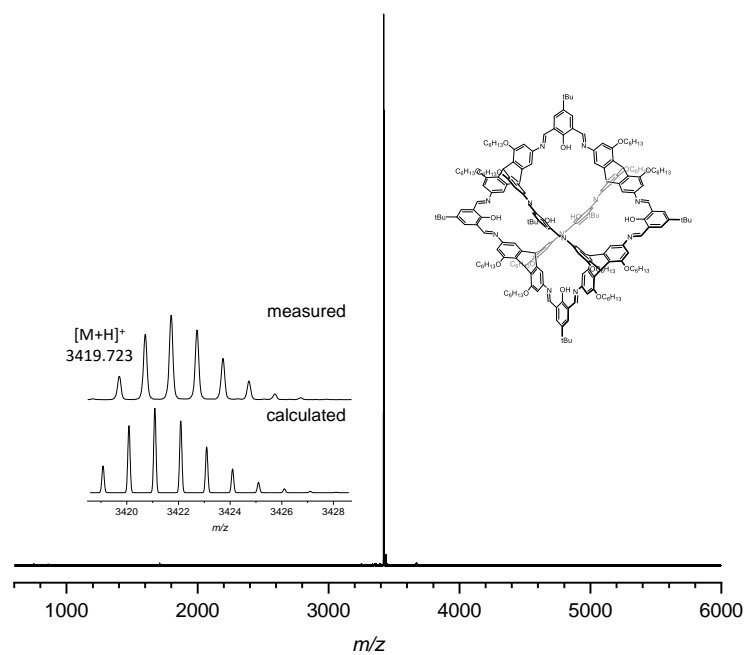


Figure S109. MALDI-TOF mass spectrum of the *tert*-butyl substituted [4+6]-*endo* cage **14** (DCTB matrix, reflector mode, positive polarisation).

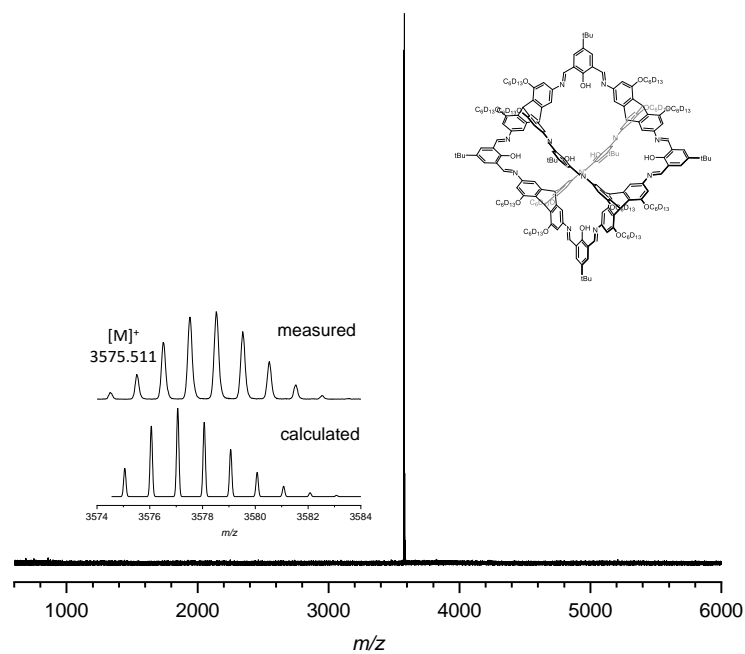


Figure S110. MALDI-TOF mass spectrum of the *tert*-butyl substituted [4+6]-*endo* cage **14-d₁₅₆** (DCTB matrix, reflector mode, positive polarisation).

6 IR Spectra

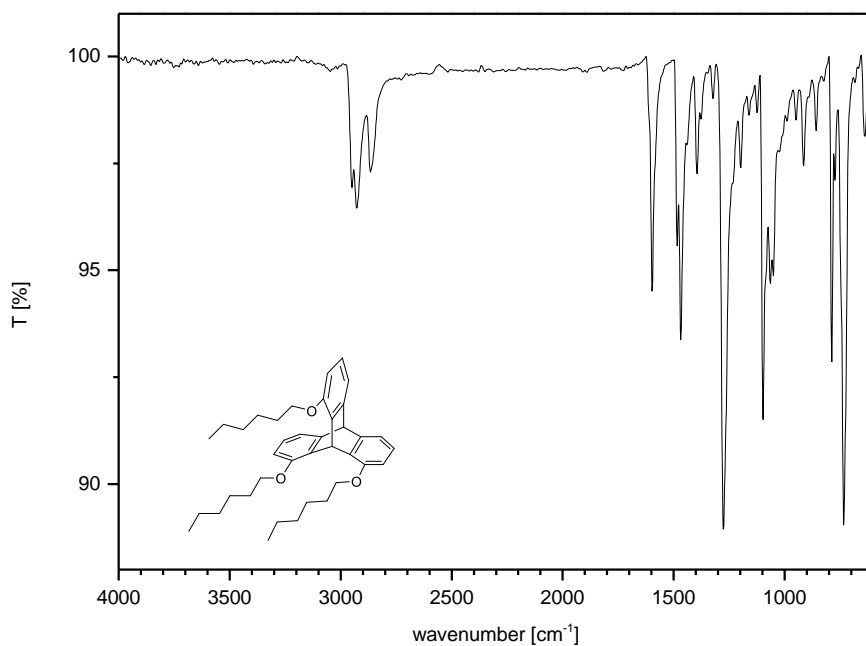


Figure S111. FT-IR spectrum of compound **5** (ATR, ZnSe).

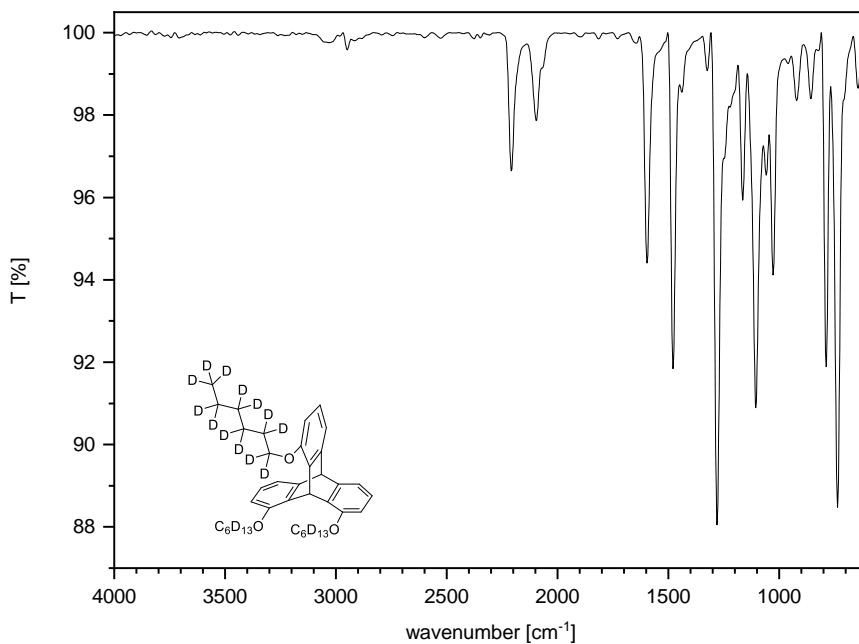


Figure S112. FT-IR spectrum of compound **5-d₃₉** (ATR, ZnSe).

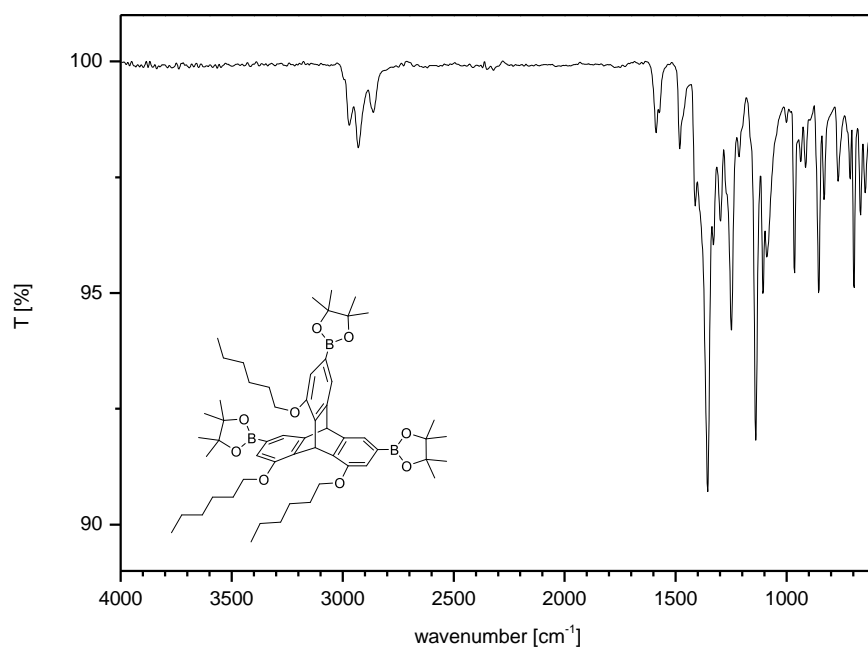


Figure S113. FT-IR spectrum of compound **6** (ATR, ZnSe).

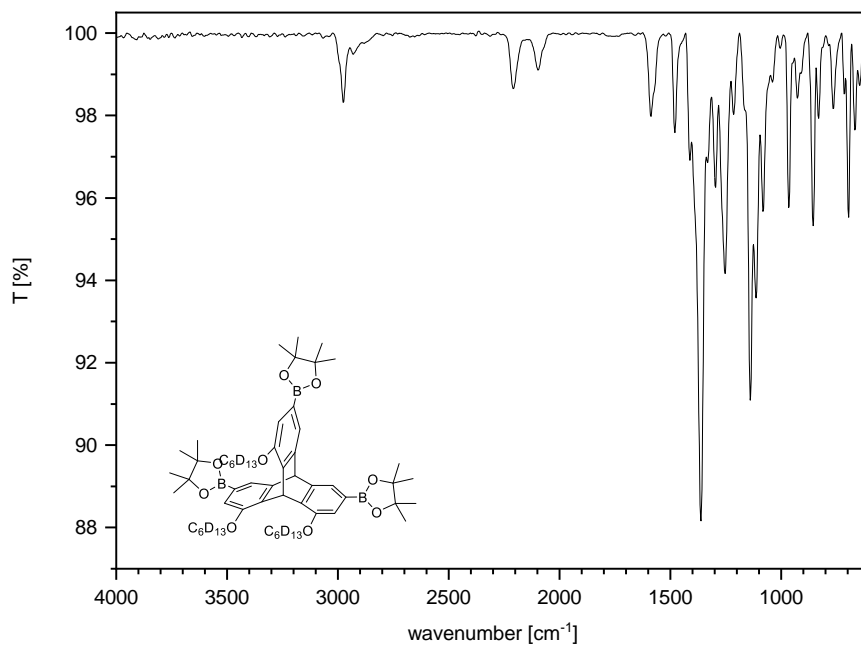


Figure S114. FT-IR spectrum of compound **6-d₃₉** (ATR, ZnSe).

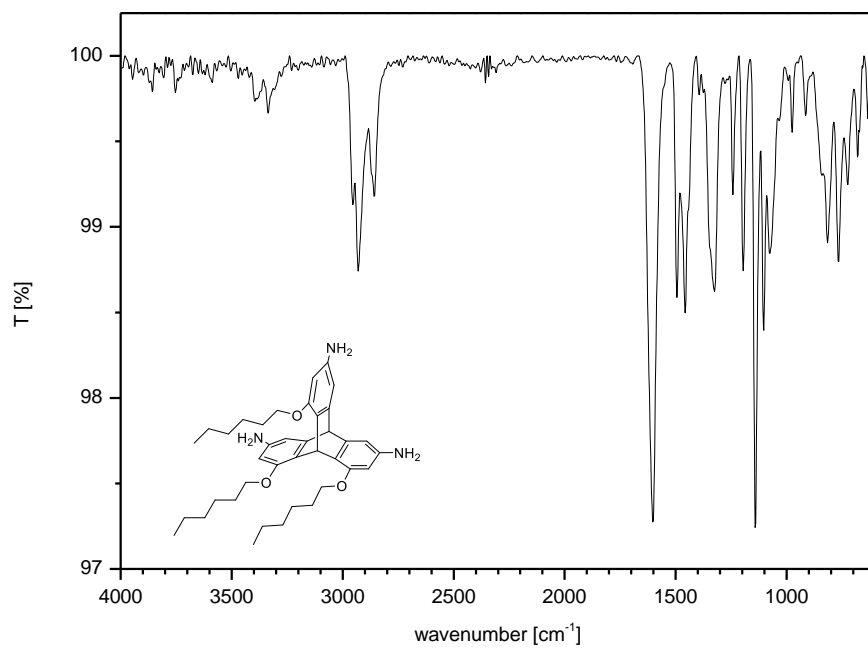


Figure S115. FT-IR spectrum of compound **7a** (ATR, ZnSe).

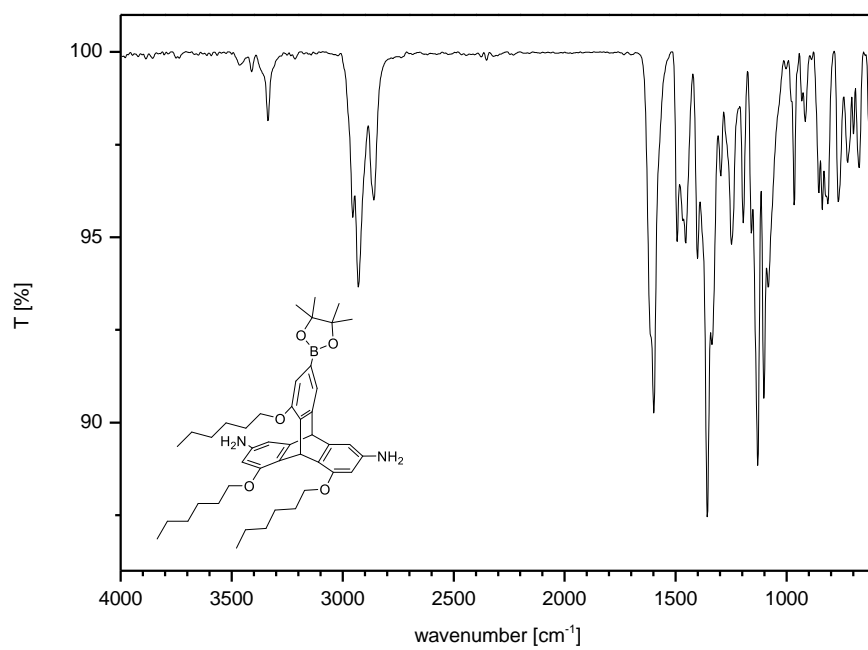


Figure S116. FT-IR spectrum of compound **7b** (ATR, ZnSe).

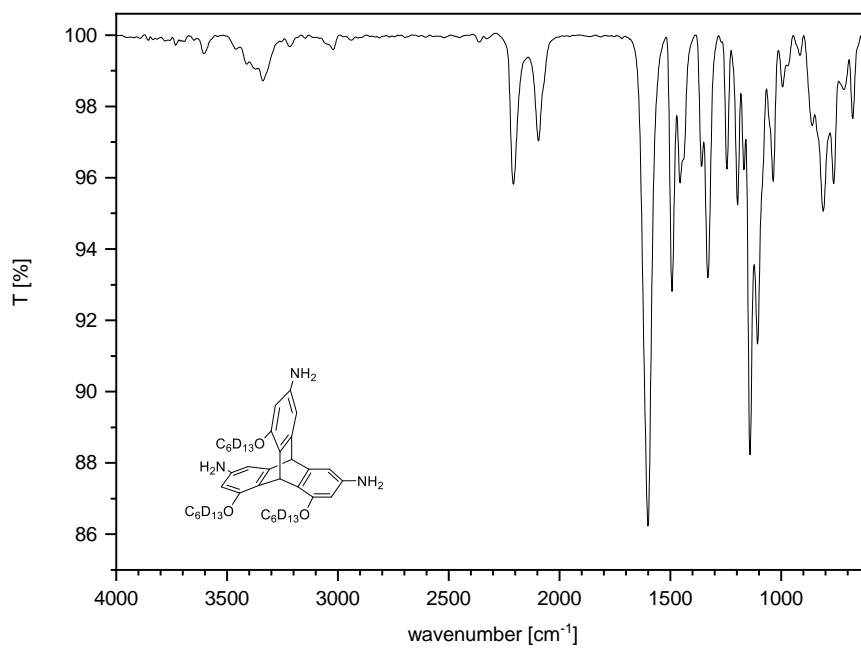


Figure S117. FT-IR spectrum of compound **7a-d₃₉** (ATR, ZnSe).

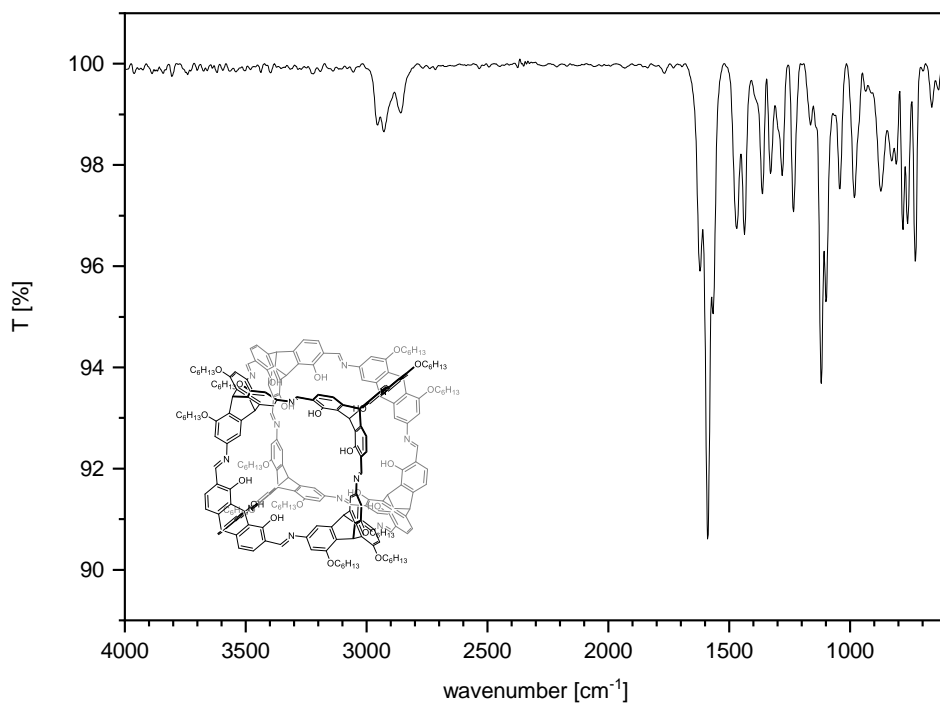


Figure S118. FT-IR spectrum of [4+4] cube **11** (ATR, ZnSe).

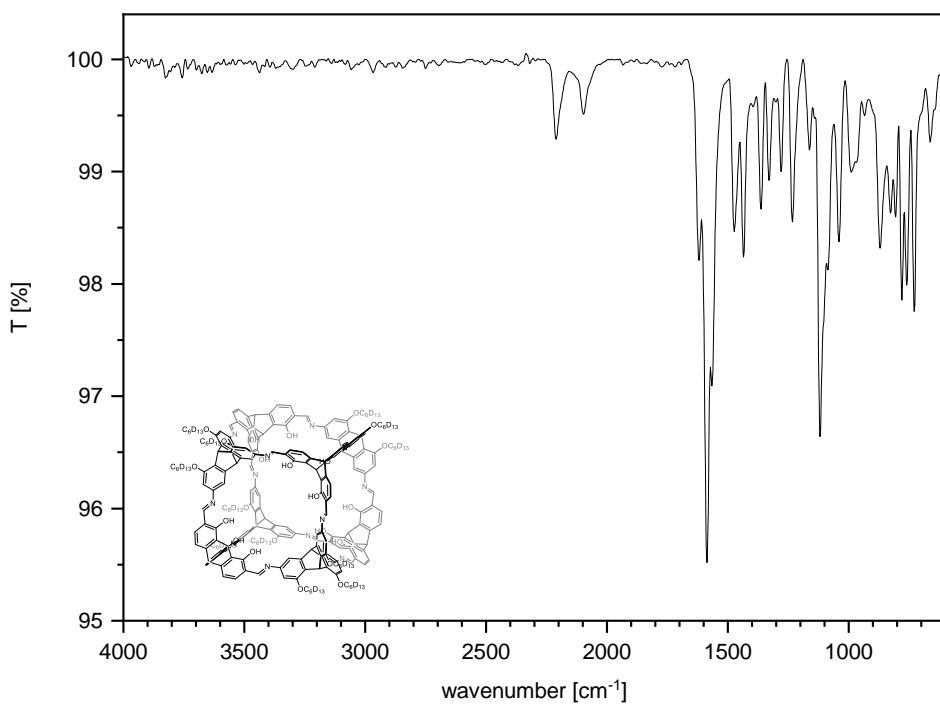


Figure S119. FT-IR spectrum of [4+4] cube **11-d₁₅₆** (ATR, ZnSe).

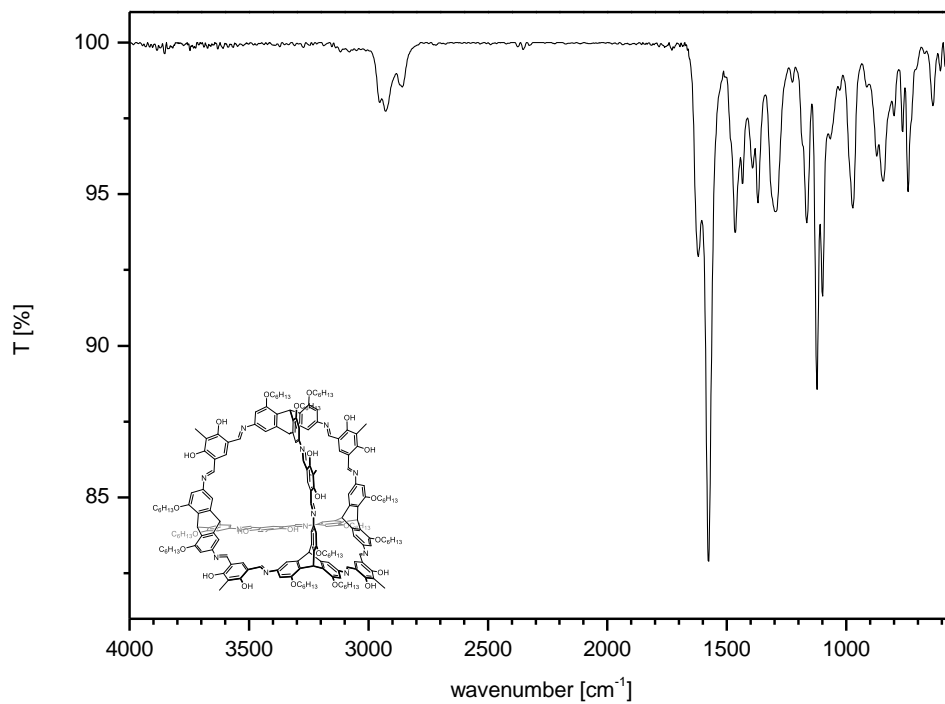


Figure S120. FT-IR spectrum of [4+6]-exo cage **12** (ATR, ZnSe).

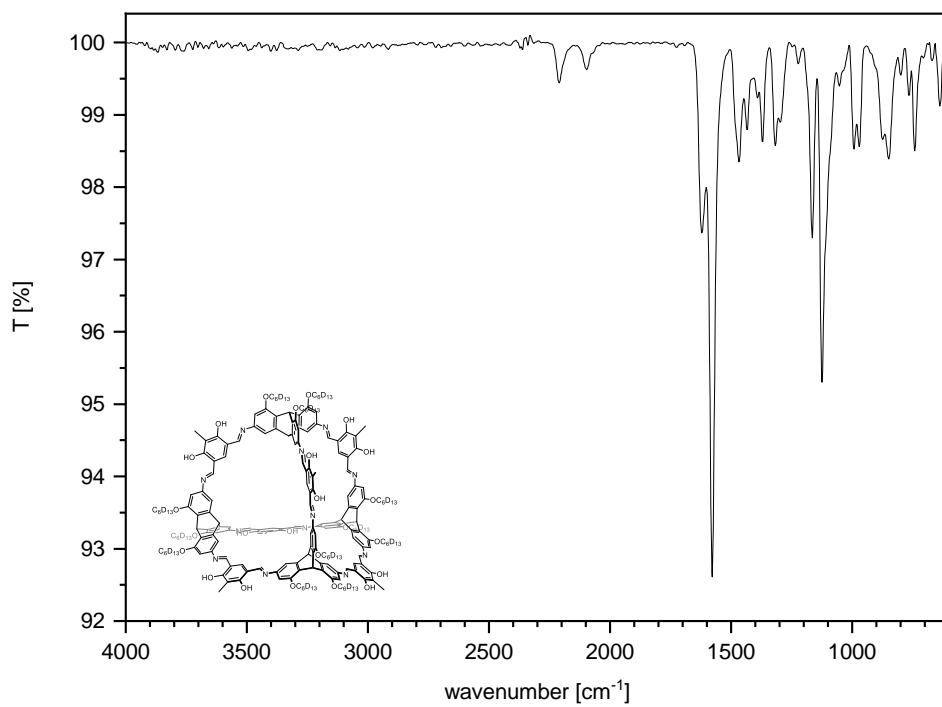


Figure S121. FT-IR spectrum of [4+6]-exo cage **12-d₁₅₆** (ATR, ZnSe).

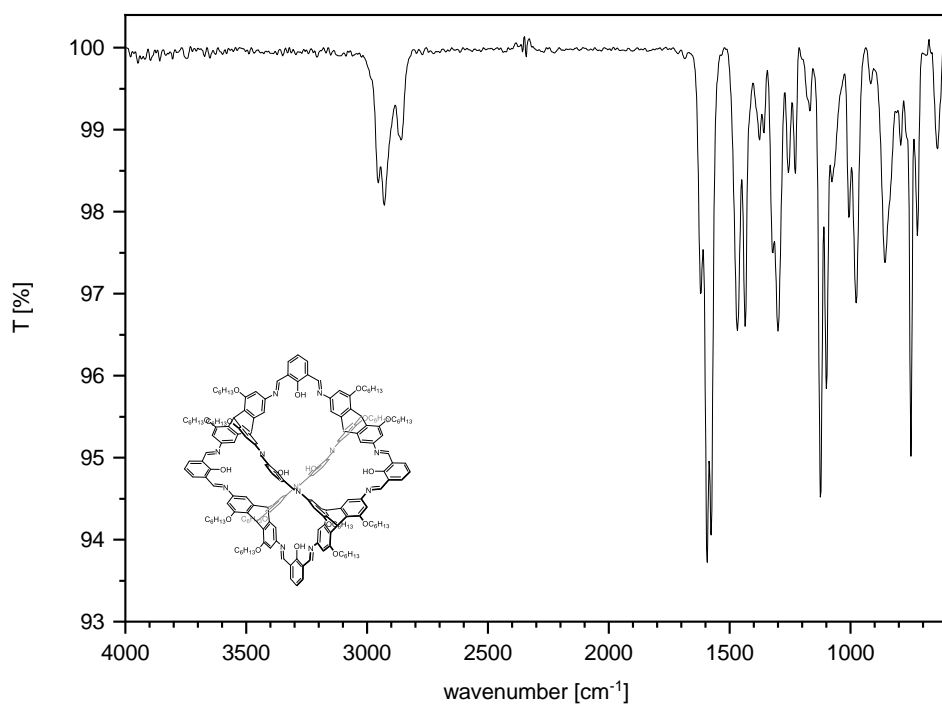


Figure S122. FT-IR spectrum of [4+6]-*endo* cage **13** (ATR, ZnSe).

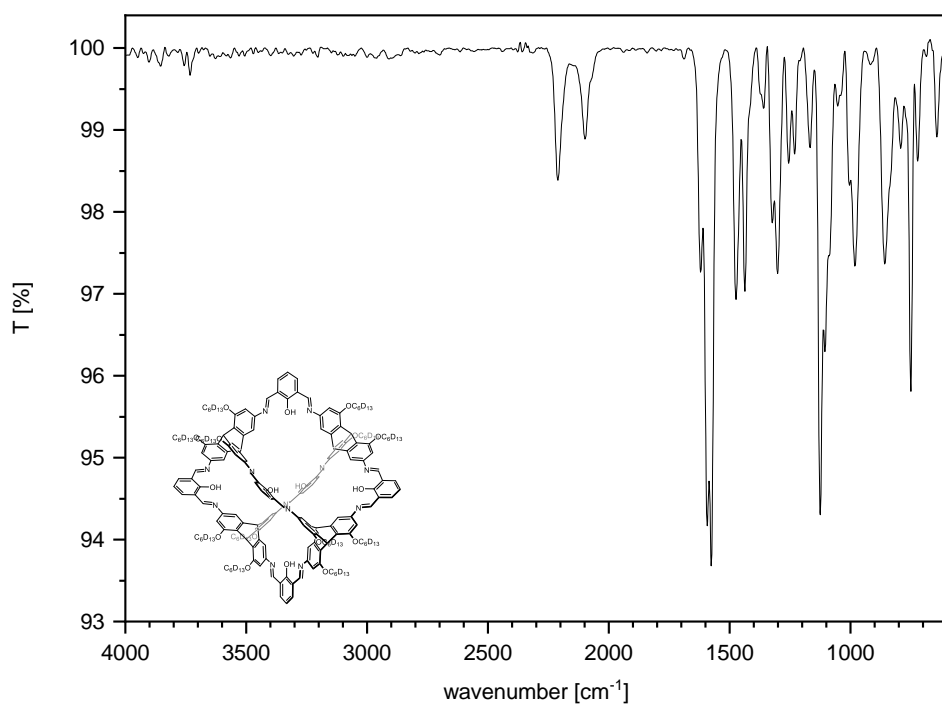


Figure S123. FT-IR spectrum of [4+6]-*endo* cage **13-d₁₅₆** (ATR, ZnSe).

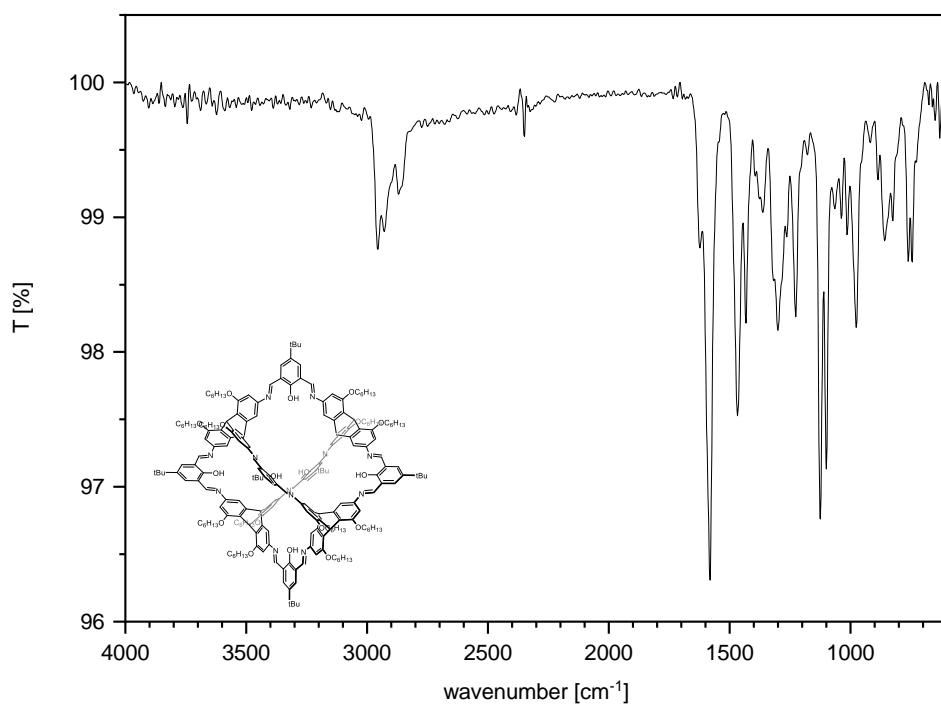


Figure S124. FT-IR spectrum of *tert*-butylated [4+6]-*endo* cage **14** (ATR, ZnSe).

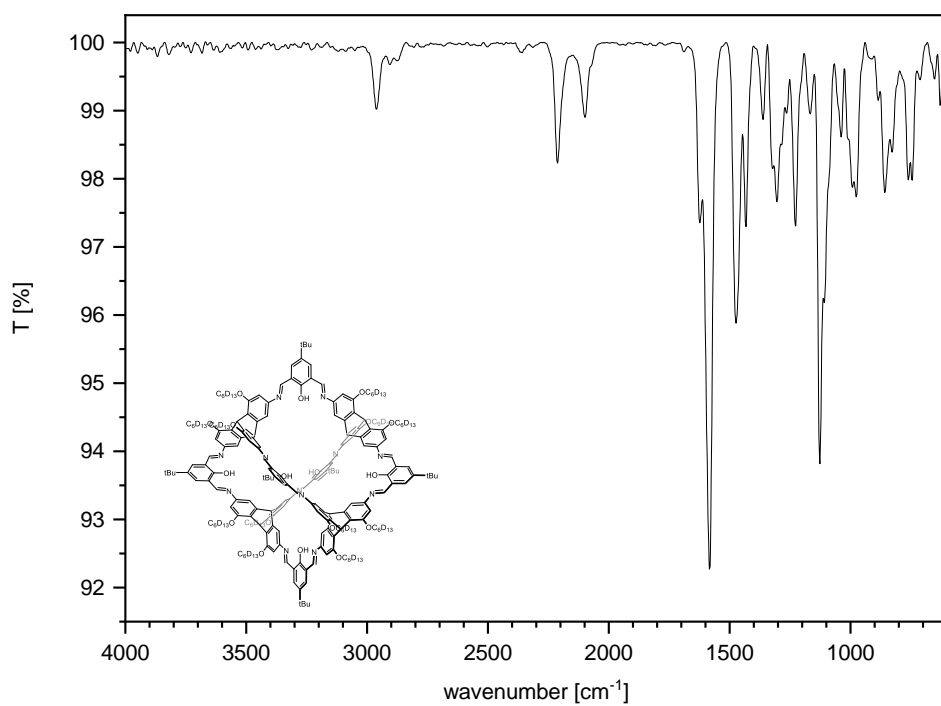


Figure S125. FT-IR spectrum of *tert*-butylated [4+6]-*endo* cage **14-d₁₅₆** (ATR, ZnSe).

7 UV/vis spectra

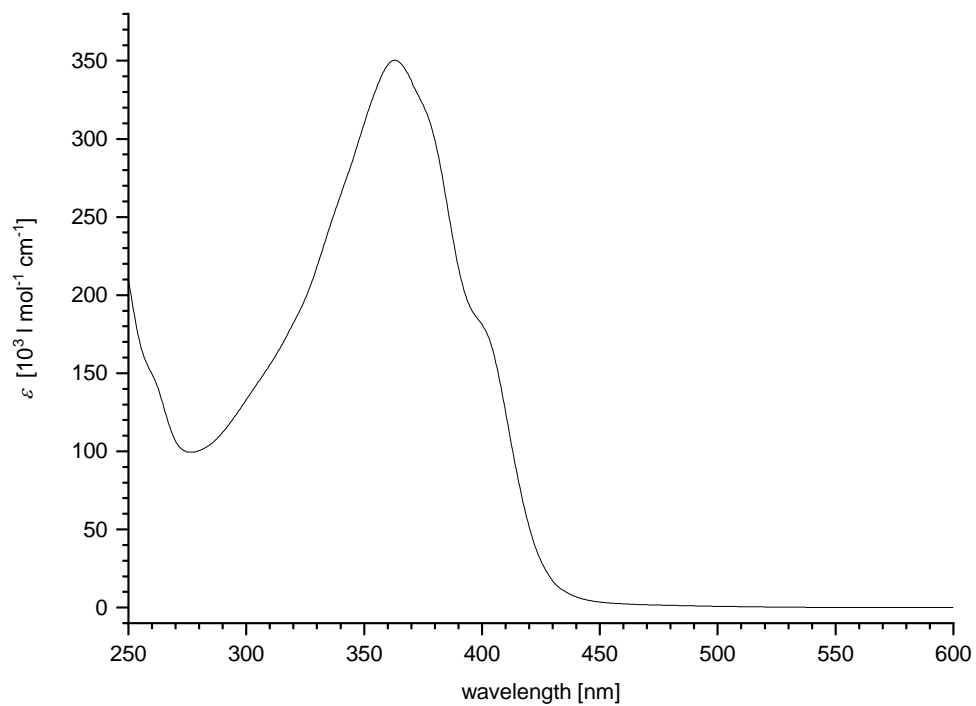


Figure S126. UV/vis spectrum of [4+4] cube **11** in THF.

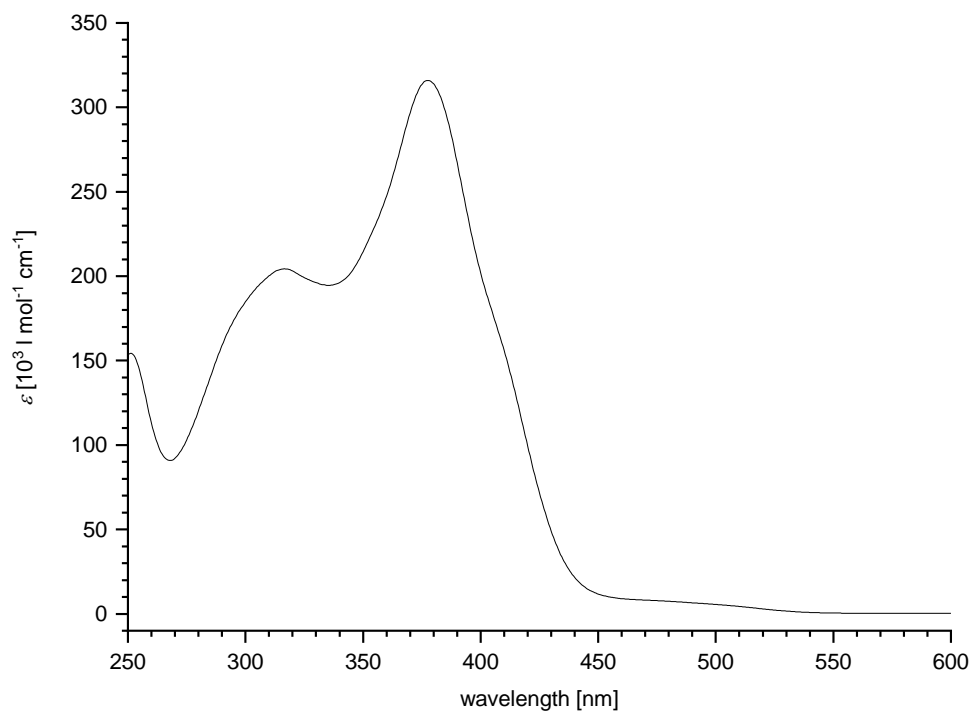


Figure S127. UV/vis spectrum of [4+6]-exo cage **12** in THF.

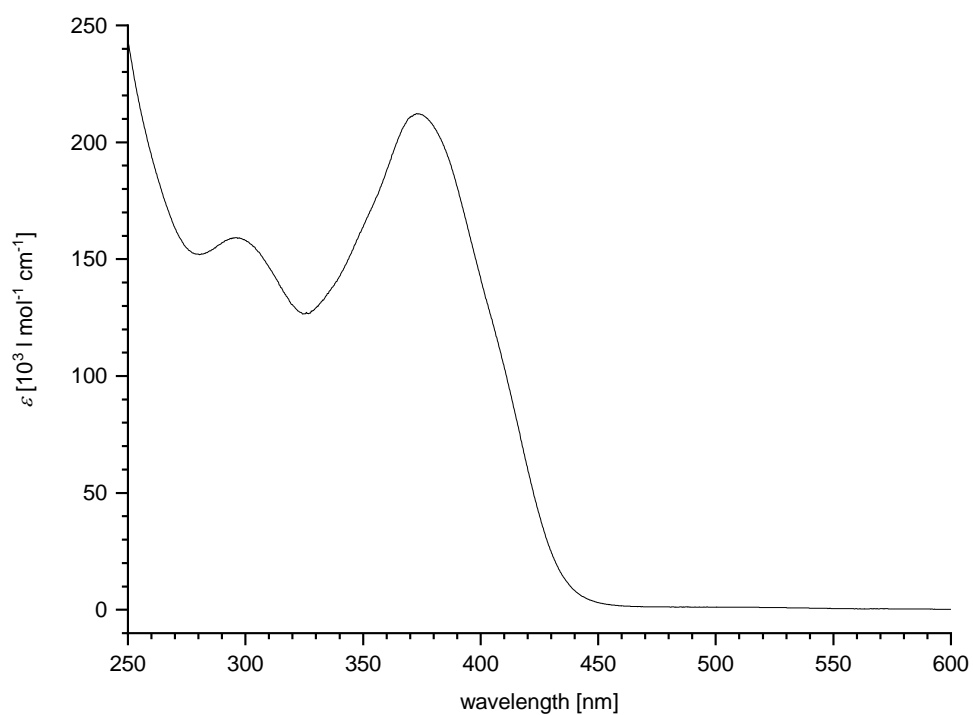


Figure S128. UV/vis spectrum of [4+6]-endo cage 13 in THF.

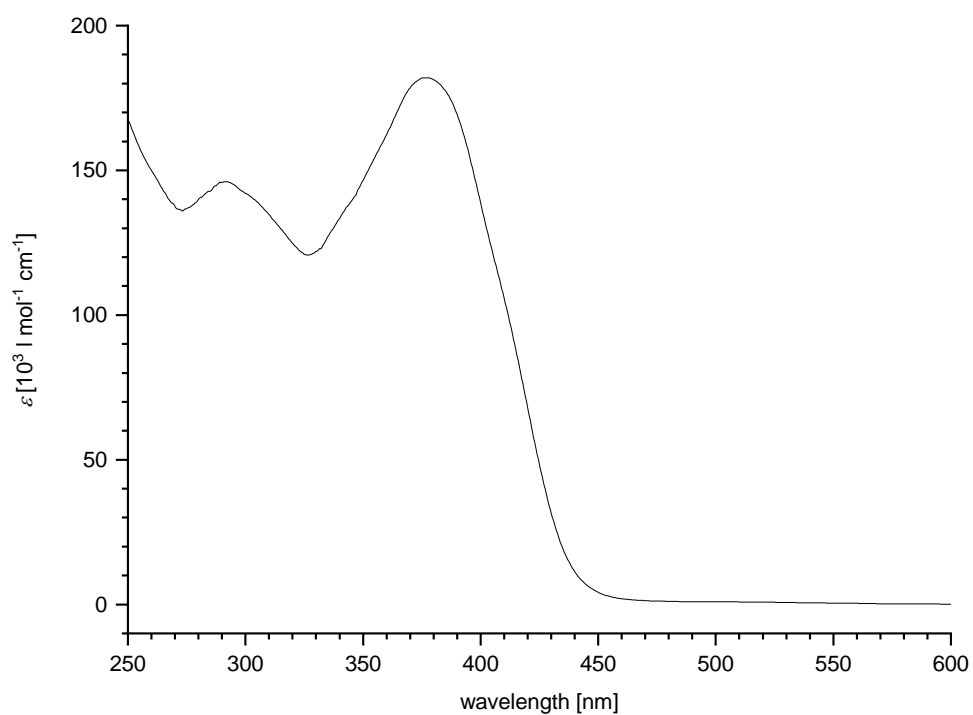


Figure S129. UV/vis spectrum of *tert*-butylated [4+6]-endo cage 14 in THF.

8 GPC Chromatograms

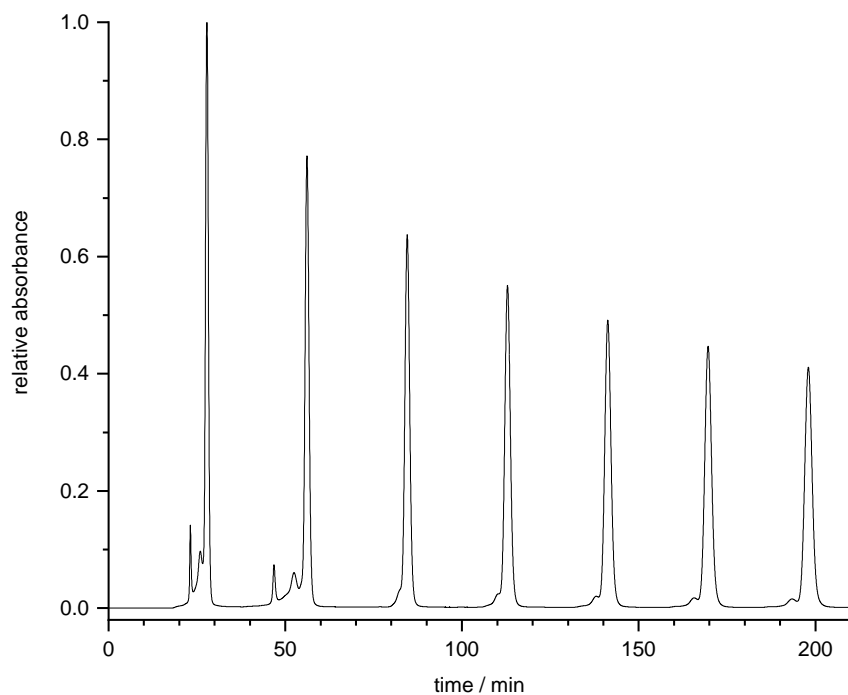


Figure S130. GPC chromatogram of crude [4+4] cube **11** using THF as mobile phase.

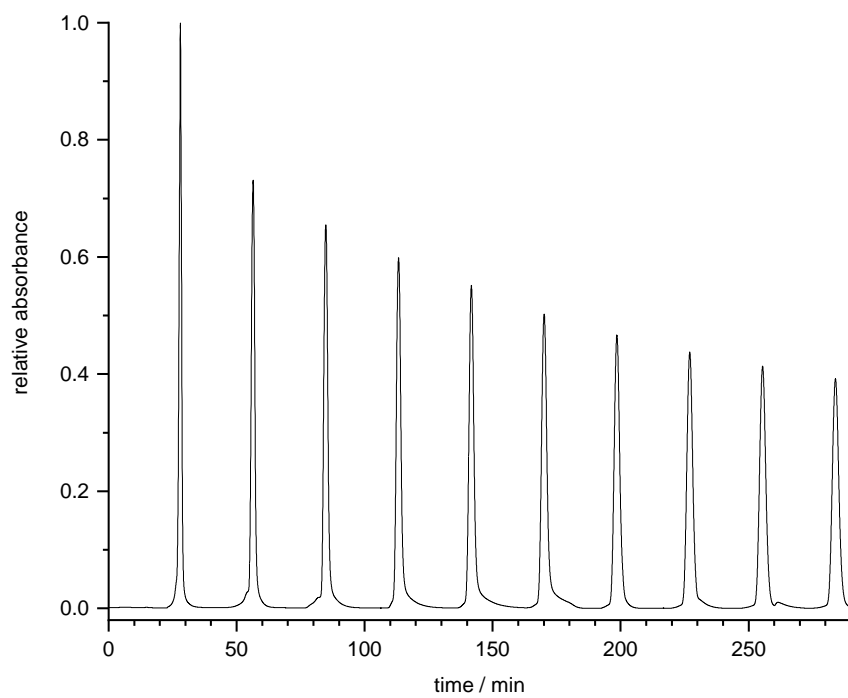


Figure S131. GPC chromatogram of [4+6]-exo cage **12** after washing with DMSO and methanol using THF as mobile phase.

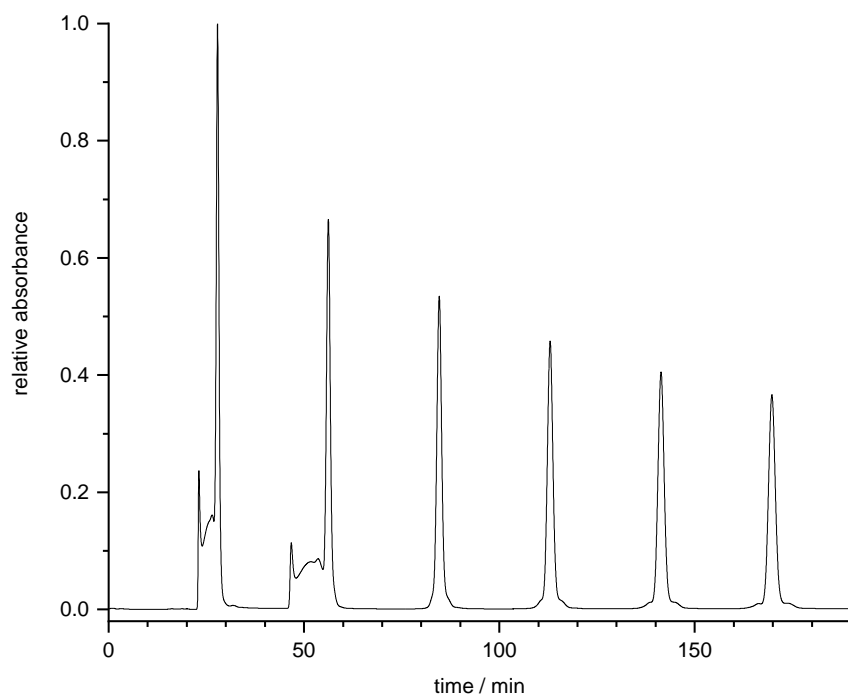


Figure S132. GPC chromatogram of crude [4+6]-*endo* cage **13** after stirring with molecular sieves (3 Å) and filtration through diatomaceous earth powder using THF as mobile phase.

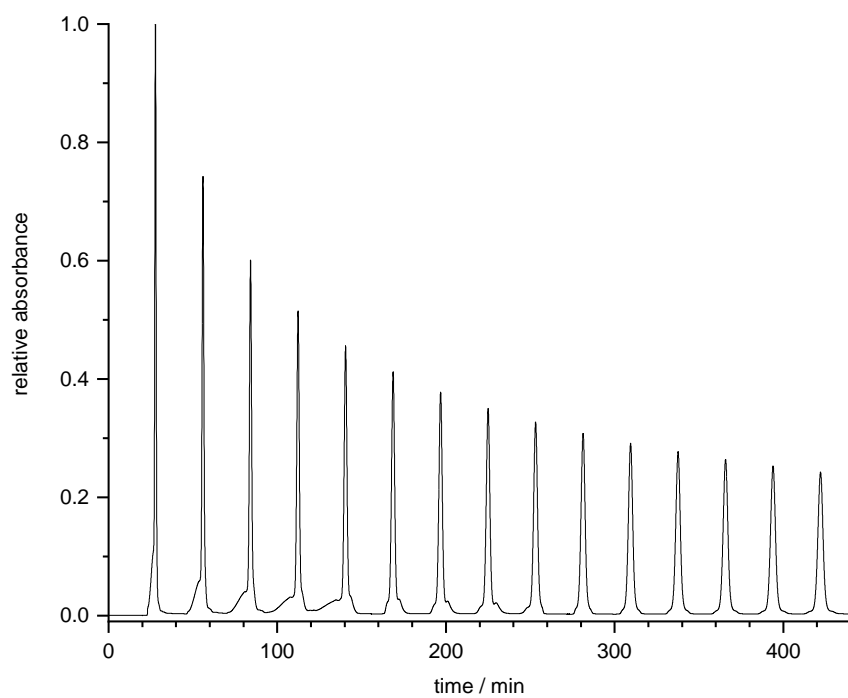


Figure S133. GPC chromatogram of *tert*-butylated [4+6]-*endo* cage **14** after stirring with molecular sieves (3 Å) and filtration through Celite.

9 Crystallographic Data

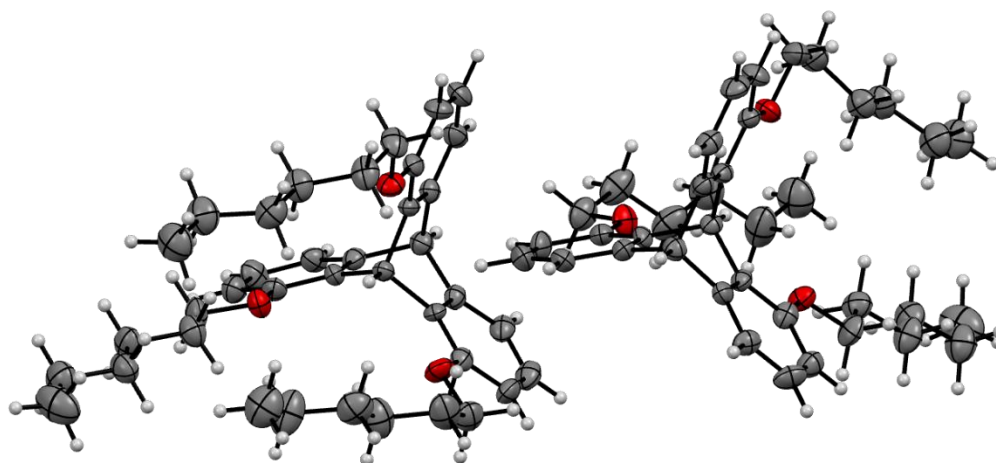


Figure S134. X-ray crystal structure of compound **5**. Atoms are illustrated as thermal ellipsoids with 50% probability. Grey: carbon, white: hydrogen, red: oxygen. Crystals were obtained via slow evaporation of a 5 containing CDCl₃ solution in an NMR tube at room temperature.

CCDC	2060523
Empirical formula	C ₃₈ H ₅₀ O ₃
Formula weight	554.78
Temperature [K]	200(2)
λ (CuK α) [Å]	1.54178
Crystal system	monoclinic
Space group	P2 ₁
Z	2
a [Å]	13.9339(12)
b [Å]	8.0969(6)
c [Å]	14.6140(13)
α [°]	90
β [°]	100.695(7)
γ [°]	90
Volume [Å ³]	1620.1(2)
ρ (calculated) [mg mm ⁻³]	1.14
μ [mm ⁻¹]	0.54
Crystal shape	plate
Crystal size [mm ³]	0.057 x 0.041 x 0.012
Crystal colour	colourless
Θ range for data collection [°]	3.1 to 69.5
Index ranges	-16 \leq h \leq 10, -9 \leq k \leq 8, -16 \leq l \leq 17
Reflections collected	11486
Independent reflections	4884 (R_{int} = 0.1139)
Observed reflections	2824 ($I > 2\sigma(I)$)
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	1.41 and 0.73
Refinement method	Full-matrix least-squares on F^2
Data/restraints/parameters	4884 / 1 / 373
Goodness-of-fit on F^2	1.07
Final R indices ($I > 2\sigma(I)$)	R_1 = 0.073, wR_2 = 0.122
Largest diff. peak and hole [e Å ⁻³]	0.24 and -0.25
Resolution [Å]	0.82

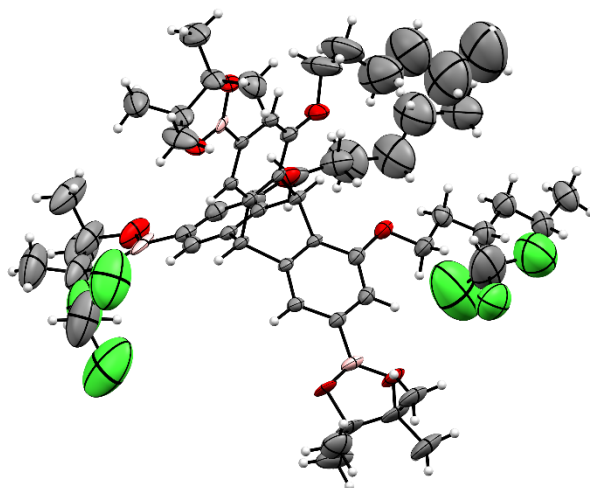


Figure S135. X-ray crystal structure of compound **6** co-crystallised with two chloroform molecules. Atoms are illustrated as thermal ellipsoids with 50% probability. Grey: carbon, white: hydrogen, red: oxygen, pink: boron, green: chlorine. Crystals were obtained via thermal recrystallization of **6** in ethanol by adding chloroform under reflux conditions.

CCDC	2060524
Empirical formula	C ₅₈ H ₈₅ B ₃ Cl ₆ O ₉
Formula weight	1171.39
Temperature [K]	200(2)
λ (MoK α) [Å]	0.71073 Å
Crystal system	triclinic
Space group	P $\bar{1}$
Z	2
a [Å]	12.652(4)
b [Å]	12.778(4)
c [Å]	22.927(7)
α [°]	77.132(8)
β [°]	79.236(8)
γ [°]	60.982(7)
Volume [Å ³]	3146.4(17)
ρ (calculated) [mg mm ⁻³]	1.24
μ [mm ⁻¹]	0.32
Crystal shape	polyhedron
Crystal size [mm ³]	0.200 x 0.110 x 0.080
Crystal colour	colourless
Θ range for data collection [°]	0.9 to 19.4
Index ranges	-11 ≤ h ≤ 11, -11 ≤ k ≤ 11, -21 ≤ l ≤ 21
Reflections collected	17297
Independent reflections	5362 ($R_{int} = 0.1292$)
Observed reflections	3063 ($I > 2\sigma(I)$)
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	0.96 and 0.66
Refinement method	Full-matrix least-squares on F^2
Data/restraints/parameters	5362 / 1644 / 685
Goodness-of-fit on F^2	1.75
Final R indices ($I > 2\sigma(I)$)	$R_1 = 0.202$, $wR_2 = 0.491$
Largest diff. peak and hole [e Å ⁻³]	0.91 and -0.92
Resolution [Å]	1.07

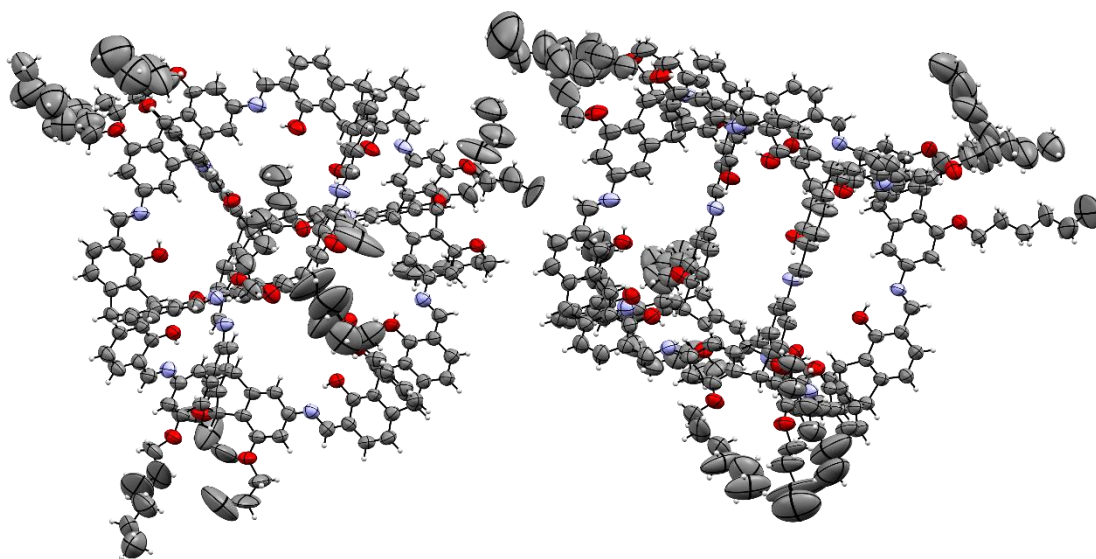


Figure S136: X-ray crystal structure of [4+4] cube **11**. Atoms are illustrated as thermal ellipsoids with 50% probability. Grey: carbon, white: hydrogen, blue: nitrogen, red: oxygen. Crystals were obtained via gas phase diffusion of methanol in a solution of [4+4] cube in ethyl acetate at room temperature.

CCDC	2060526
Empirical formula	$C_{488}H_{488}N_{24}O_{48}$
Formula weight	7457.00
Temperature [K]	200(2)
$\lambda(\text{CuK}\alpha)$ [Å]	1.54178
Crystal system	triclinic
Space group	$P\bar{1}$
Z	2
a [Å]	25.1707(5)
b [Å]	32.4180(6)
c [Å]	40.0124(8)
α [°]	85.230(2)
β [°]	85.835(2)
γ [°]	71.082(1)
Volume [Å ³]	30742.8(11)
ρ (calculated) [mg mm ⁻³]	0.81
μ [mm ⁻¹]	0.41
Crystal shape	brick
Crystal size [mm ³]	0.158 x 0.133 x 0.068
Crystal colour	orange
Θ range for data collection [°]	2.2 to 49.1
Index ranges	$-24 \leq h \leq 24, -31 \leq k \leq 24, -39 \leq l \leq 39$
Reflections collected	154010
Independent reflections	59090 ($R_{int} = 0.0799$)
Observed reflections	26500 ($I > 2\sigma(I)$)
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	1.44 and 0.66
Refinement method	Full-matrix least-squares on F^2
Data/restraints/parameters	59090 / 26804 / 4870
Goodness-of-fit on F^2	1.45
Final R indices ($I > 2\sigma(I)$)	$R_1 = 0.099, wR_2 = 0.281$
Largest diff. peak and hole [$e \text{ \AA}^{-3}$]	0.60 and -0.26
Resolution [Å]	1.02

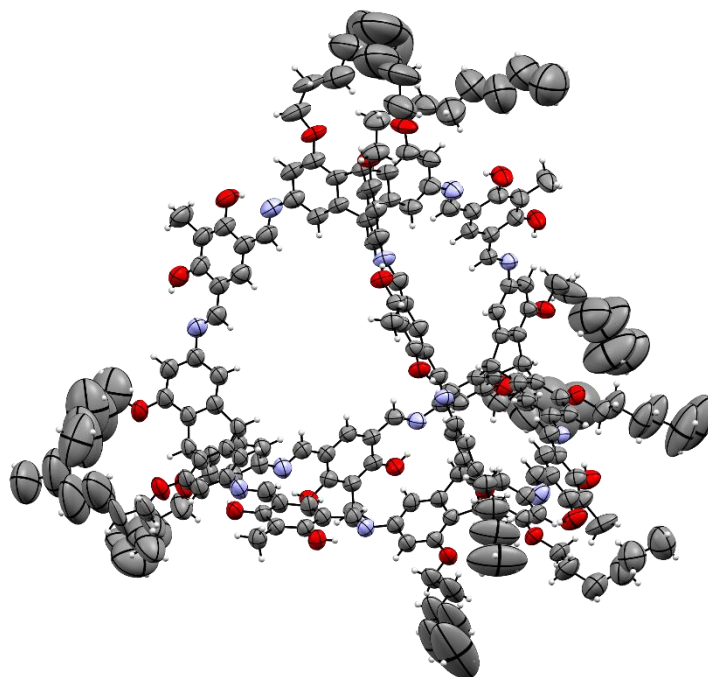


Figure S137. X-ray crystal structure of [4+6]-exo-cage **12**. Atoms are illustrated as thermal ellipsoids with 50% probability. Grey: carbon, white: hydrogen, blue: nitrogen, red: oxygen. Crystals were obtained via slow solvent evaporation of a solution of **12** in DCM at room temperature.

CCDC	2060525
Empirical formula	C ₂₁₀ H ₂₃₆ N ₁₂ O ₂₄
Formula weight	3312.09
Temperature [K]	200(2)
$\lambda(\text{CuK}\alpha)$ [Å]	1.54178
Crystal system	triclinic
Space group	P $\bar{1}$
Z	2
a [Å]	23.9115(7)
b [Å]	26.0674(8)
c [Å]	27.8715(8)
α [°]	63.665(2)
β [°]	74.083(2)
γ [°]	84.127(2)
Volume [Å ³]	14968.9(8)
ρ (calculated) [mg mm ⁻³]	0.74
μ [mm ⁻¹]	0.38
Crystal shape	brick
Crystal size [mm ³]	0.136 x 0.106 x 0.100
Crystal colour	orange
Θ range for data collection [°]	2.4 to 52.6
Index ranges	-24 \leq h \leq 22, -26 \leq k \leq 19, -28 \leq l \leq 27
Reflections collected	95908
Independent reflections	33955 ($R_{int} = 0.0498$)
Observed reflections	21587 ($I > 2\sigma(I)$)
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	1.56 and 0.58
Refinement method	Full-matrix least-squares on F^2
Data/restraints/parameters	33955 / 9312 / 2257
Goodness-of-fit on F^2	1.37
Final R indices ($I > 2\sigma(I)$)	$R_1 = 0.112$, $wR_2 = 0.333$
Largest diff. peak and hole [e Å ⁻³]	0.72 and -0.36
Resolution [Å]	0.97

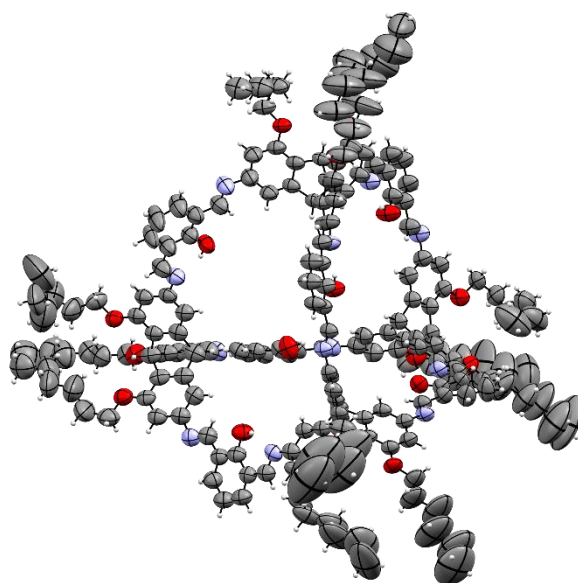


Figure S138: X-ray crystal structure of [4+6]-endo cage **13**. Atoms are illustrated as thermal ellipsoids with 50% probability. Grey: carbon, white: hydrogen, blue: nitrogen, red: oxygen. Crystals were obtained via gas phase diffusion of acetonitrile in a solution of [4+6]-endo cage in DMF at room temperature.

CCDC	2060527
Empirical formula	C ₂₀₀ H ₂₂₄ N ₁₂ O ₁₈
Formula weight	3083.90
Temperature [K]	200(2)
$\lambda(\text{CuK}\alpha)$ [Å]	1.54178
Crystal system	triclinic
Space group	P $\bar{1}$
Z	2
a [Å]	21.9869(7)
b [Å]	23.7687(6)
c [Å]	29.5082(8)
α [°]	78.767(2)
β [°]	76.553(2)
γ [°]	63.044(2)
Volume [Å ³]	13297.5(7)
ρ (calculated) [mg mm ⁻³]	0.77
μ [mm ⁻¹]	0.39
Crystal shape	brick
Crystal size [mm ³]	0.280 x 0.210 x 0.130
Crystal colour	orange
Θ range for data collection [°]	2.1 to 51.1
Index ranges	-21 \leq h \leq 22, -24 \leq k \leq 22, -27 \leq l \leq 29
Reflections collected	76777
Independent reflections	28074 ($R_{int} = 0.0464$)
Observed reflections	17341 ($I > 2\sigma(I)$)
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	1.30 and 0.78
Refinement method	Full-matrix least-squares on F^2
Data/restraints/parameters	28074 / 9681 / 2255
Goodness-of-fit on F^2	2.37
Final R indices ($I > 2\sigma(I)$)	$R_1 = 0.135$, $wR_2 = 0.358$
Largest diff. peak and hole [e Å ⁻³]	0.55 and -0.43
Resolution [Å]	0.99

10 Exchange Experiments

Exchange between [4+4] cages **11** and **11-d₁₅₆**

In an 8 mL screw-capped vial cage **11** (1.0 mg, 0.3 μmol , 1 eq) and cage **11-d₁₅₆** (1.0 mg, 0.3 μmol , 1 eq) were combined under argon atmosphere with 0.49 mL dry DMF and TFA, p-toluidine or water (each 10.29 μL , 0.01 mol L⁻¹ in dry DMF, 0.4 eq). In an additional experiment TFA (10.29 μL , 0.01 mol L⁻¹ in dry DMF, 0.4 eq) and water (61.8 μL , 0.1 mol L⁻¹ in dry DMF, 24 eq) were used. The orange solutions were stirred at 150 °C. After 3 and 7 days aliquots were analysed by mass spectrometry.

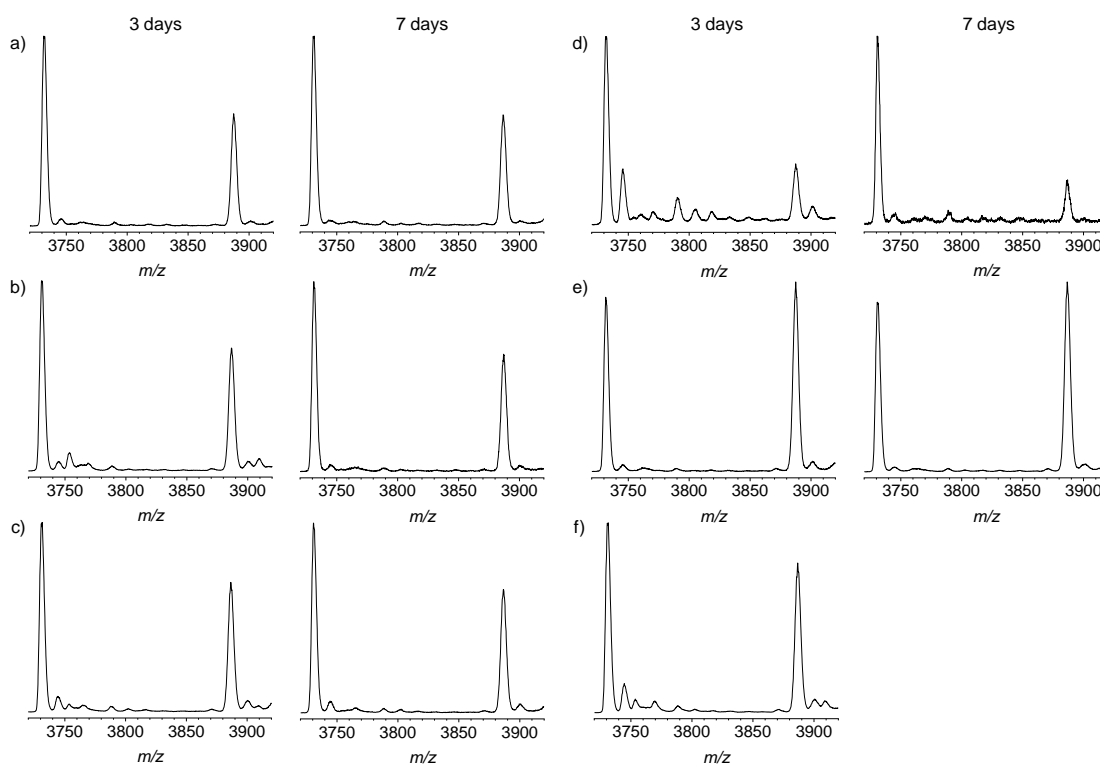


Figure S139. MALDI-TOF mass spectra of building block exchange between [4+4] cages **11** and **11-d₁₅₆** after 3 and 7 days (DCTB matrix, linear mode, positive polarisation) using a) TFA, b) TFA and water (0.4 eq), c) TFA and p-toluidine, d) p-toluidine and water, e) p-toluidine and f) TFA and water (24 eq) as additives.

Exchange between [4+6]-exo cages **12** and **12-d₁₅₆**

In an 8 mL screw-capped vial cage **12** (1.1 mg, 0.3 μmol , 1 eq) and cage **12-d₁₅₆** (1.1 mg, 0.3 μmol , 1 eq) were combined under argon atmosphere with 0.48 mL dry DMF and TFA, p-toluidine or water (each 13.48 μL , 0.01 mol L⁻¹ in dry DMF, 0.4 eq). The orange solutions were stirred at 120 °C. After 3 and 7 days aliquots were analysed by mass spectrometry.

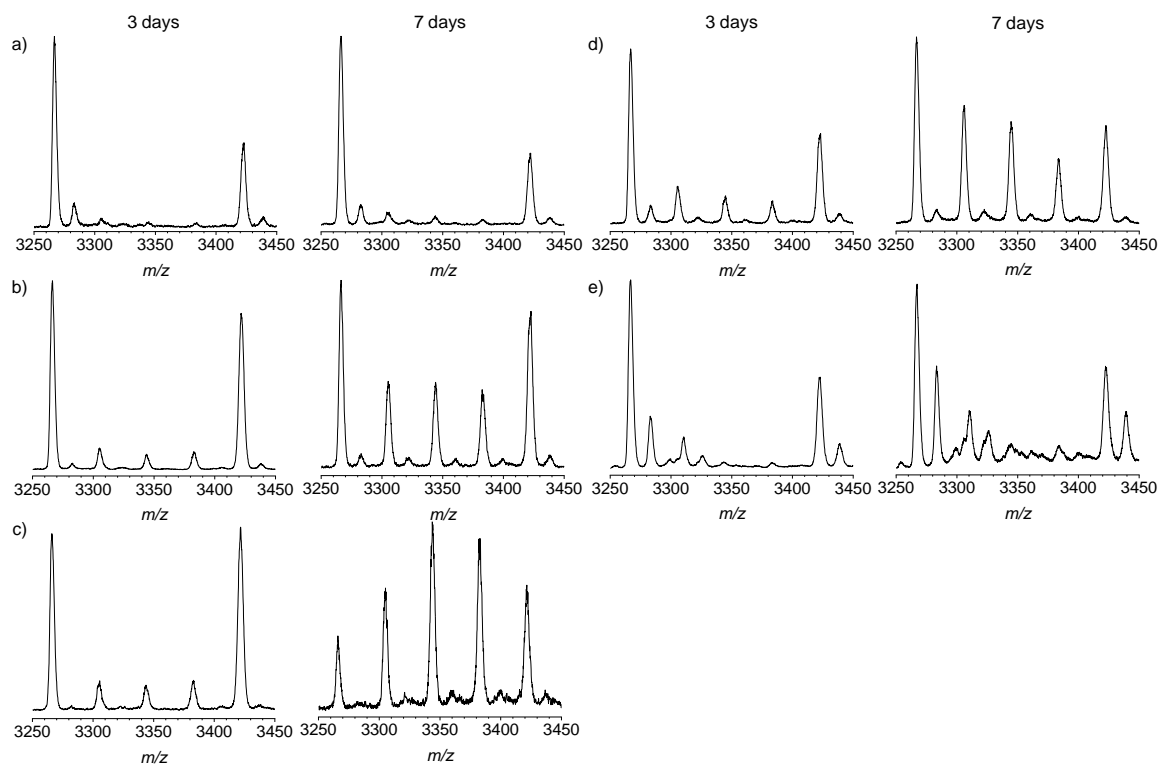


Figure S140. MALDI-TOF mass spectra of building block exchange between [4+6]-exo cages **12** and **12-d₁₅₆** after 3 and 7 days (DCTB matrix, linear mode, positive polarisation) using a) TFA, b) TFA and water, c) TFA and p-toluidine, d) p-toluidine and water and e) p-toluidine as additives.

Exchange between [4+6]-endo cages **14** and **14-d₁₅₆**

In an 8 mL screw-capped vial cage **14** (1.0 mg, 0.3 μmol , 1 eq) and cage **14-d₁₅₆** (1.0 mg, 0.3 μmol , 1 eq) were combined under argon atmosphere with 0.35 mL dry THF and TFA, p-toluidine or water (each 4.68 μL , 0.01 mol L⁻¹ in dry THF, 0.16 eq). In an additional experiment TFA (4.68 μL , 0.01 mol L⁻¹ in dry THF, 0.16 eq) and water (70.2 μL , 0.1 mol L⁻¹ in dry THF, 24 eq) were used. The orange solutions were stirred at room temperature. After 3 and 7 days aliquots were analysed by mass spectrometry.

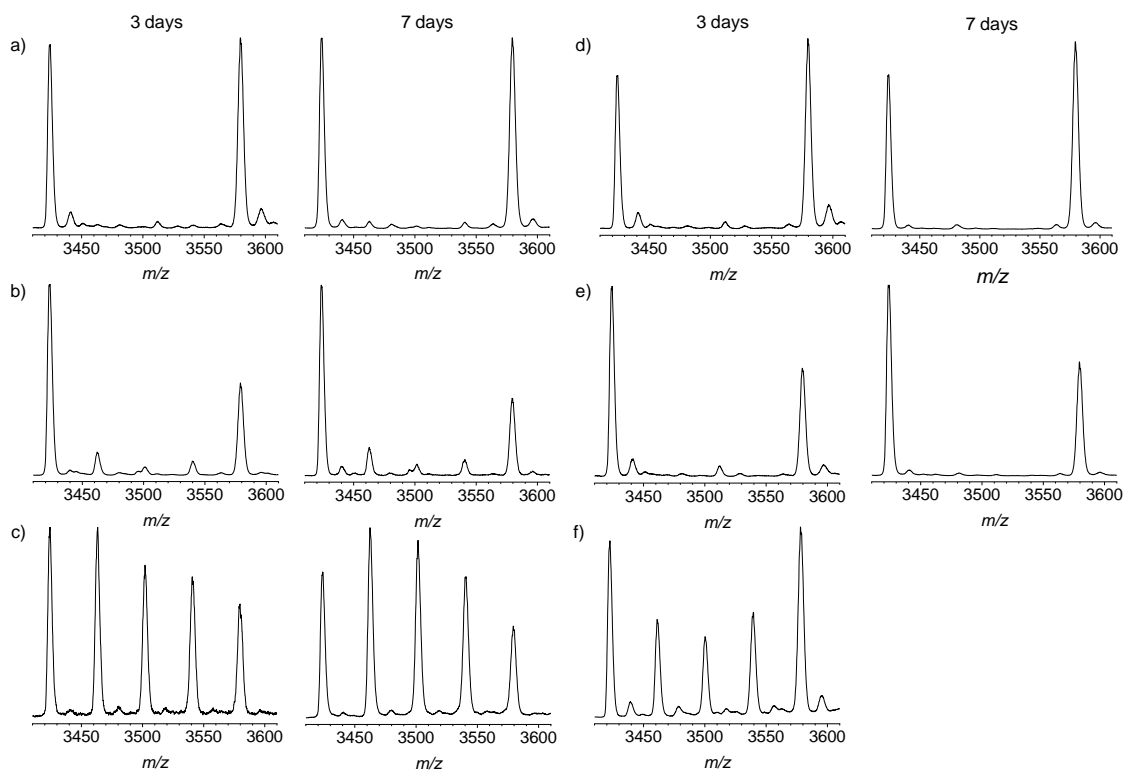


Figure S141. MALDI-TOF mass spectra of building block exchange between *tert*-butylated [4+6]-*endo* cages **14** and **14-d₁₅₆** after 3 and 7 days (DCTB matrix, linear mode, positive polarisation) using a) TFA, b) TFA and water (0.16 eq), c) TFA and *p*-toluidine, d) *p*-toluidine and water, e) *p*-toluidine and f) TFA and water (24 eq) as additives.

11 Monitoring of Cage Formation in Deuterated Solvents

Formation of [4+4] cube 11

A 25 mL screw-capped vial was charged with triaminotriptycene **7a** (20.1 mg, 34 μmol), triformyltriptycene **8** (13.0 mg, 34 μmol) and 1,3,5-trimethoxybenzene (1.6 mg, 9.51 μmol) as internal standard. Under argon atmosphere DMF- d_7 (8 mL) and TFA (167 μL , 0.01 mol L⁻¹ in CDCl₃, 1.67 μmol) were added and the orange solution was stirred at 150 °C. After 1 h, 2 h, 4 h, 8 h, 24 h, 48 h, 72 h and 96 h aliquots of 0.5 mL were taken and analysed by GPC and ¹H NMR spectroscopy.

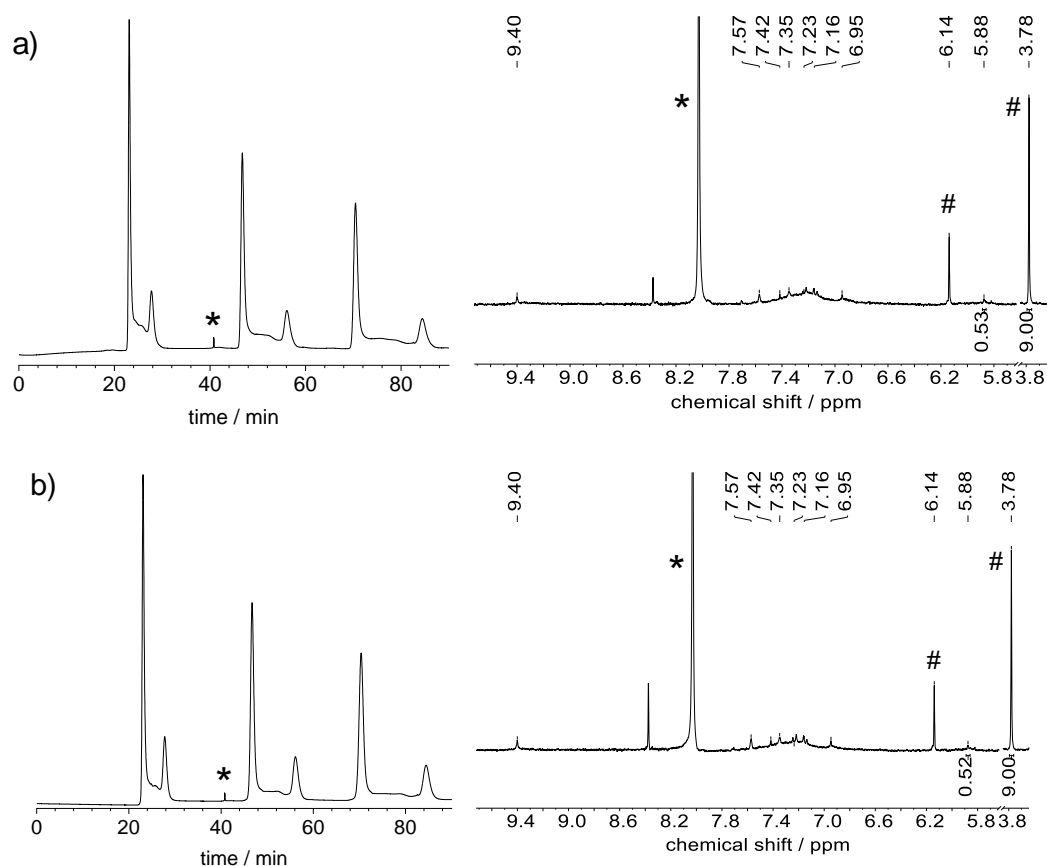


Figure S142. GPC chromatograms and ¹H-NMR spectra (300 MHz, DMF- d_7) of samples from reaction mixtures for synthesis of cage 11 after a) 1 h and b) 2 h. *: DMF- d_7 . #: 1,3,5-Trimethoxybenzene. ¹H NMR spectra show a range from 9.72 to 3.64 ppm with a cut out from 5.70 to 3.80 ppm.

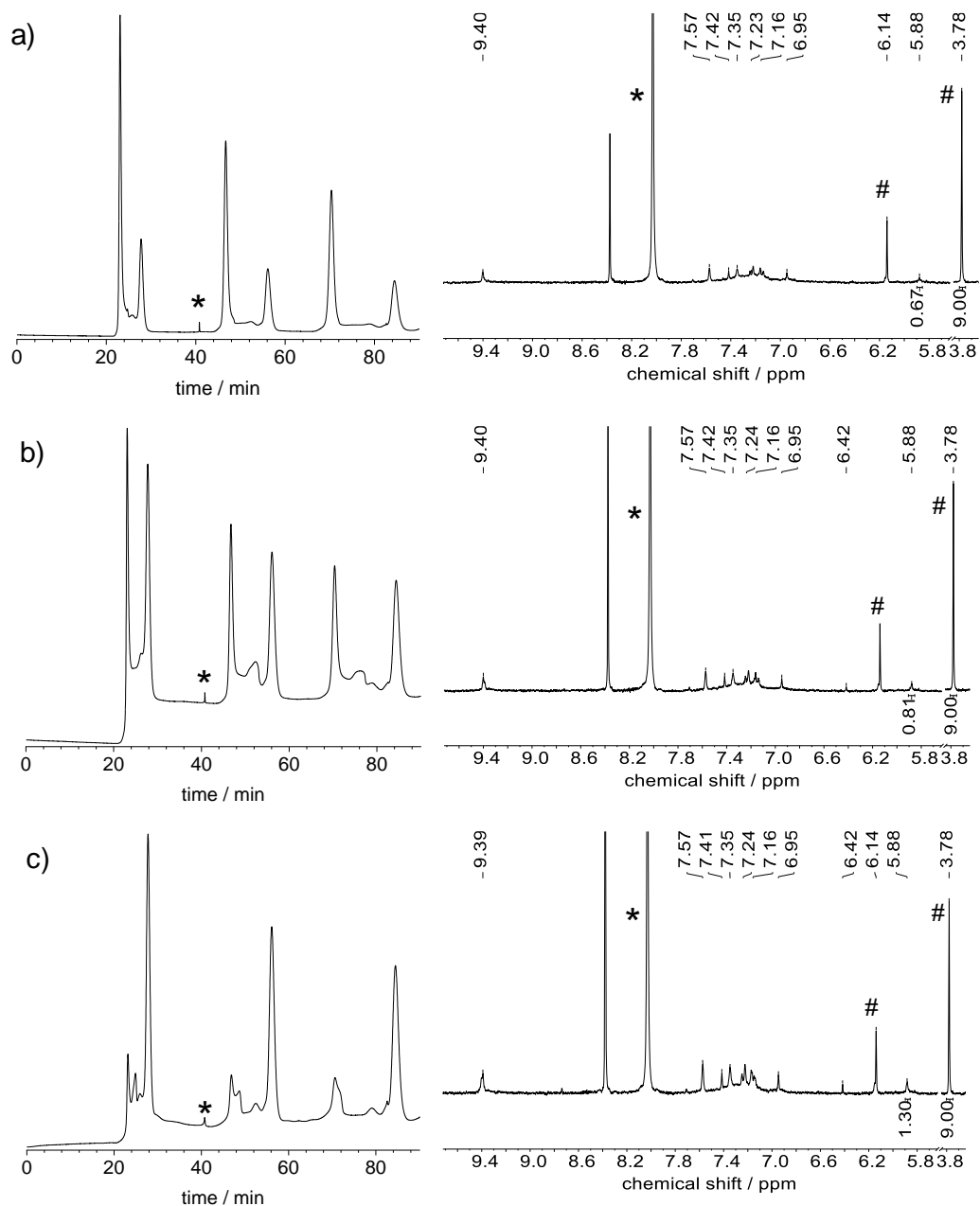


Figure S143. GPC chromatograms and ¹H-NMR spectra (300 MHz, DMF-d₇) of samples from reaction mixtures for synthesis of cage 11 after a) 4 h, b) 8 h and c) 24 h. *: DMF-d₇. #: 1,3,5-Trimethoxybenzene. ¹H NMR spectra show a range from 9.72 to 3.64 ppm with a cut out from 5.70 to 3.80 ppm.

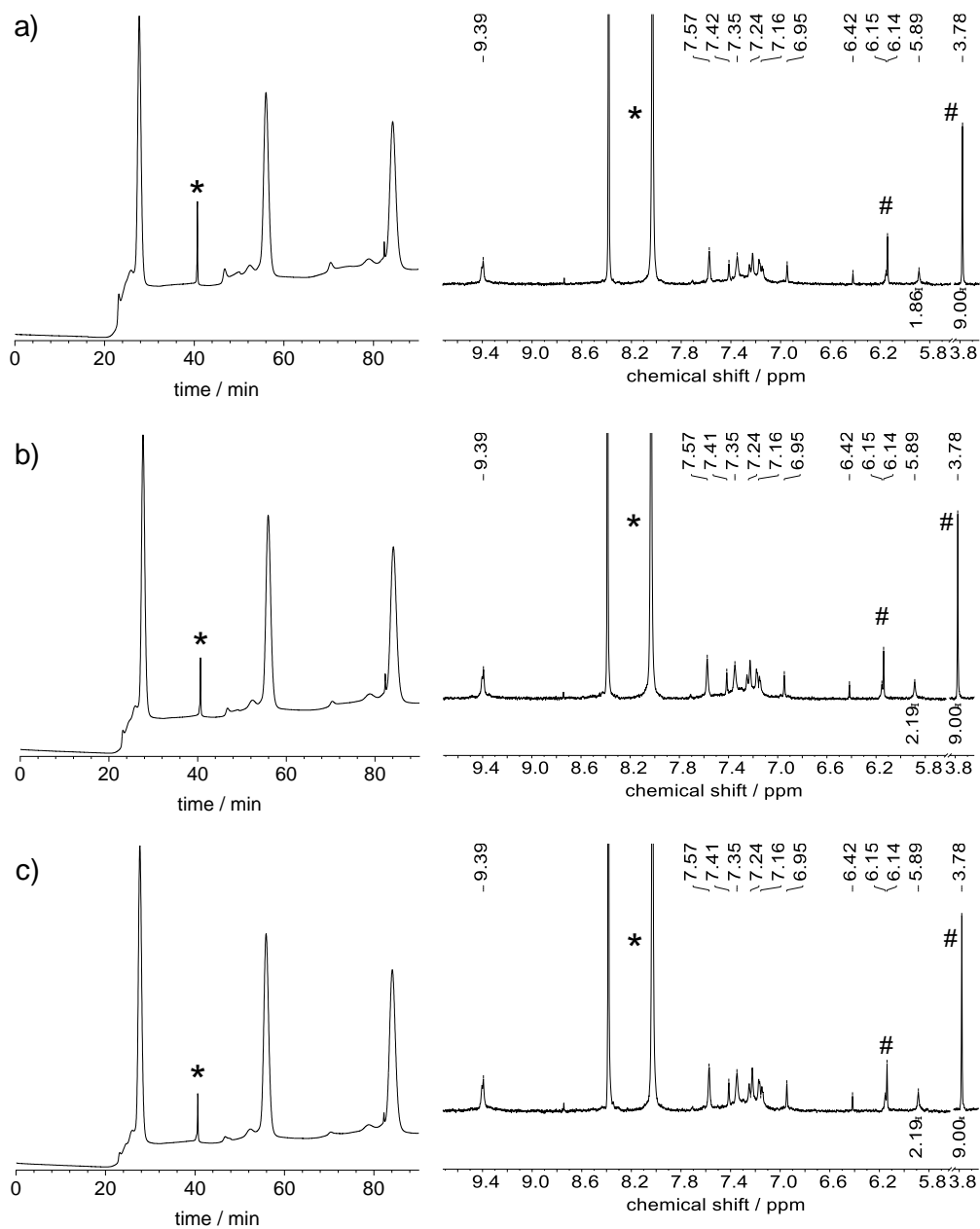


Figure S144. GPC chromatograms and $^1\text{H-NMR}$ spectra (300 MHz, DMF-d_7) of samples from reaction mixtures for synthesis of cage **11** after a) 48 h, b) 72 h and c) 96 h. *: DMF-d_7 . #: 1,3,5-Trimethoxybenzene. $^1\text{H-NMR}$ spectra show a range from 9.72 to 3.64 ppm with a cut out from 5.70 to 3.80 ppm.

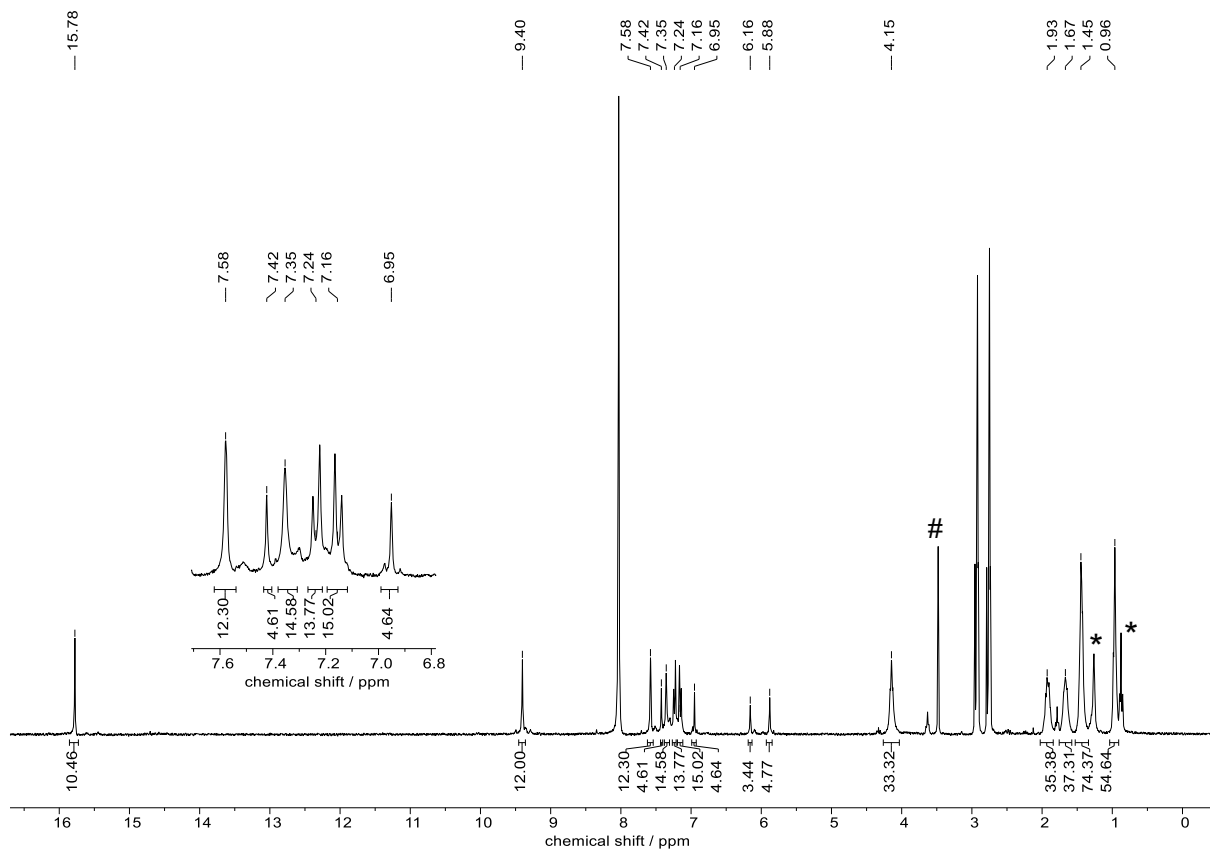


Figure S145. ^1H NMR spectrum (300 MHz, DMF-d_7) of [4+4] cube 11. #: H_2O . *: *n*-hexane.

Formation of [4+6] exo cage 12

An 8 mL screw-capped vial was charged with triaminotriptycene **7a** (20.0 mg, 33 μmol), 3,5-diformyl-2,6-dihydroxytoluene **9** (9.0 mg, 50 μmol) and 1,3,5-trimethoxybenzene (1.8 mg, 10.7 μmol) as internal standard. Under argon atmosphere DMF- d_7 (6 mL) and TFA (167 μL , 0.01 mol L $^{-1}$ in CDCl_3 , 1.67 μmol) were added and the orange solution was stirred at 120 $^\circ\text{C}$. After 1h, 2 h, 4h, 8 h, 24 h, 48 h, 72 h and 96 h aliquots of 0.5 mL were taken and analysed by GPC and ^1H NMR spectroscopy.

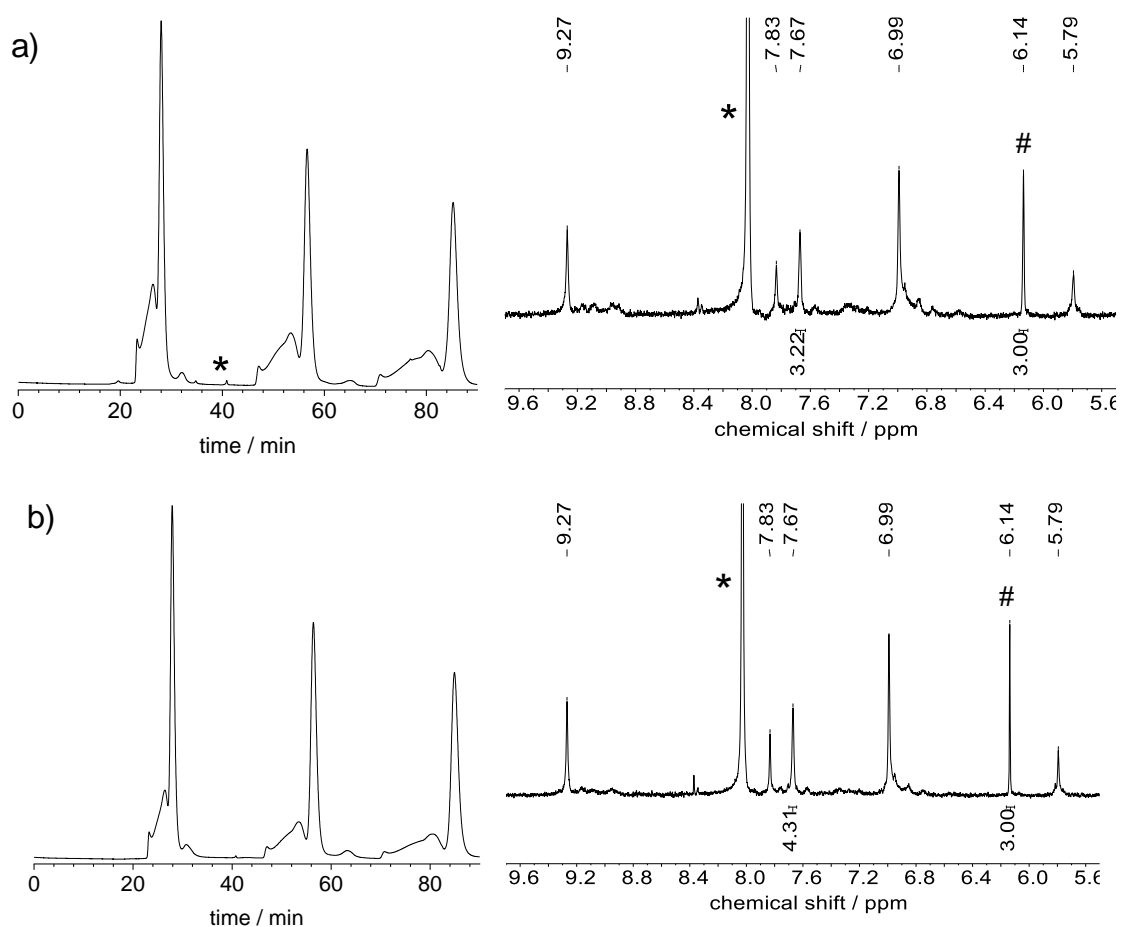


Figure S146. GPC chromatograms and ^1H -NMR spectra (300 MHz, DMF- d_7) of samples from reaction mixtures for synthesis of [4+6]-exo cage **12** after a) 1 h and b) 2 h. *: DMF- d_7 . #: 1,3,5-Trimethoxybenzene. ^1H NMR spectra show a range from 9.70 to 5.50 ppm.

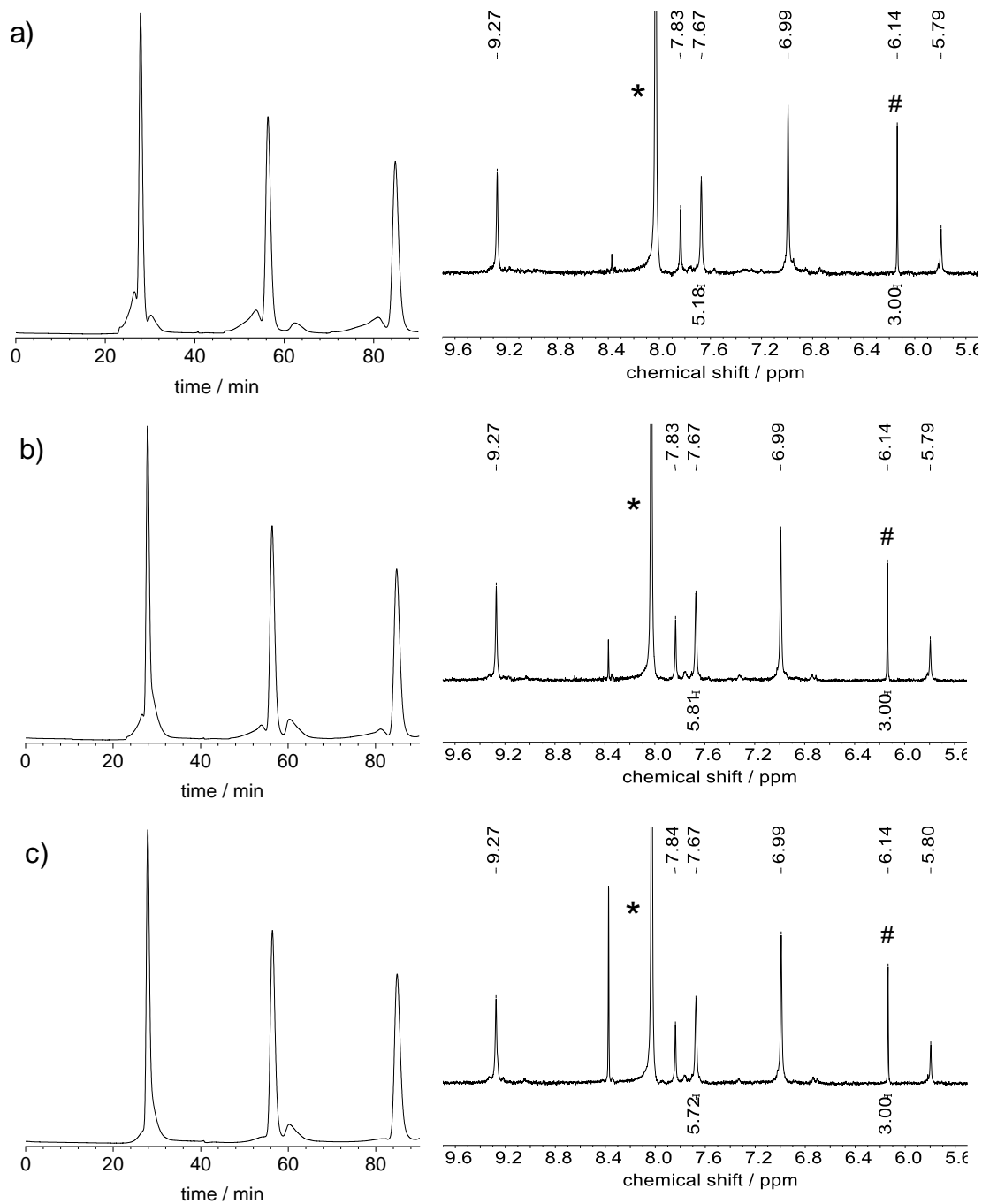


Figure S147. GPC chromatograms and ¹H-NMR spectra (300 MHz, DMF-d₇) of samples from reaction mixtures for synthesis of [4+6]-exo cage **12** after a) 4 h, b) 8 h and c) 24 h. *: DMF-d₇. #: 1,3,5-Trimethoxybenzene. ¹H NMR spectra show a range from 9.70 to 5.50 ppm.

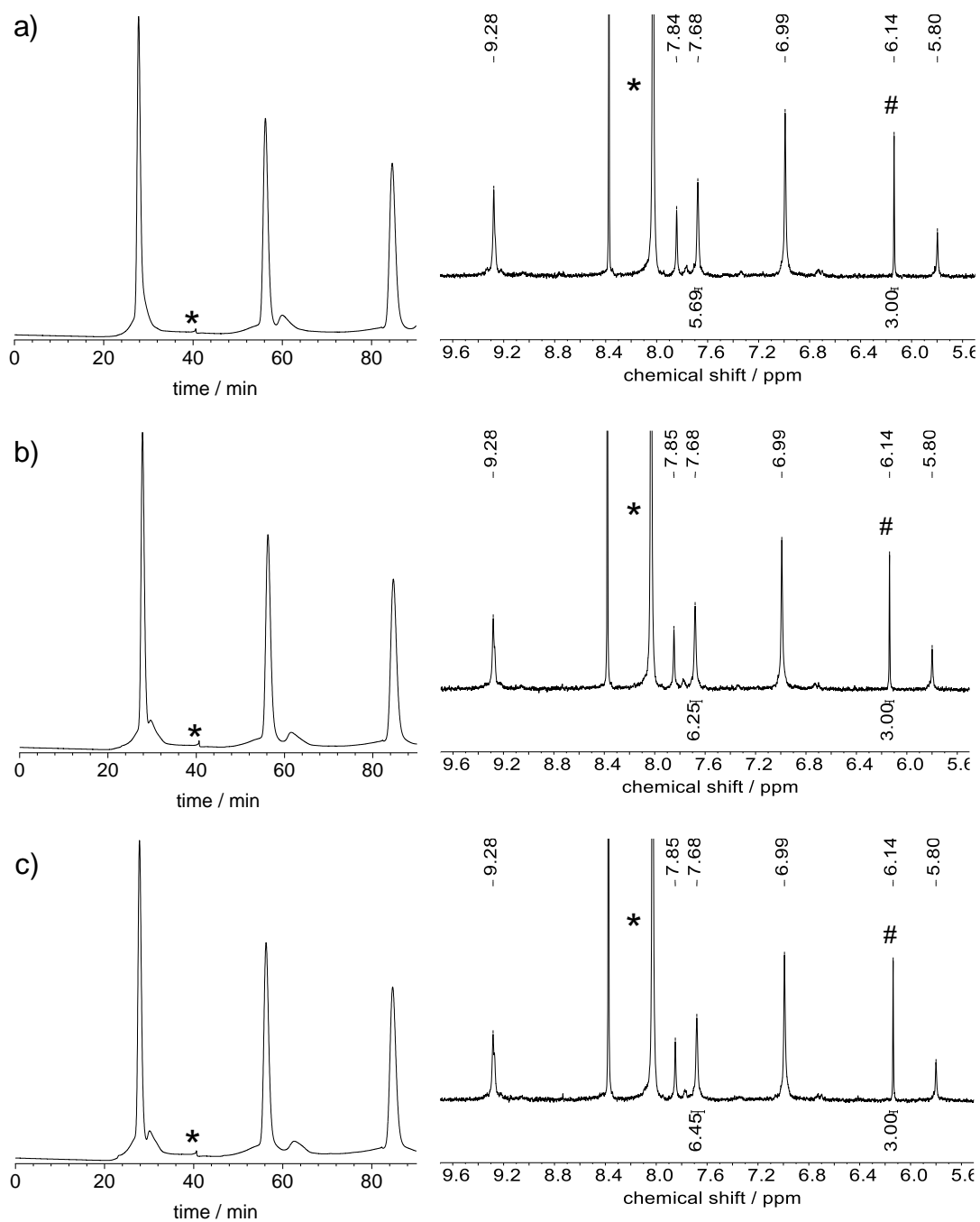


Figure S148. GPC chromatograms and ¹H-NMR spectra (300 MHz, DMF-d₇) of samples from reaction mixtures for synthesis of [4+6]-exo cage **12** after a) 48 h, b) 72 h and c) 96 h. *: DMF-d₇. #: 1,3,5-Trimethoxybenzene. ¹H NMR spectra show a range from 9.70 to 5.50 ppm.

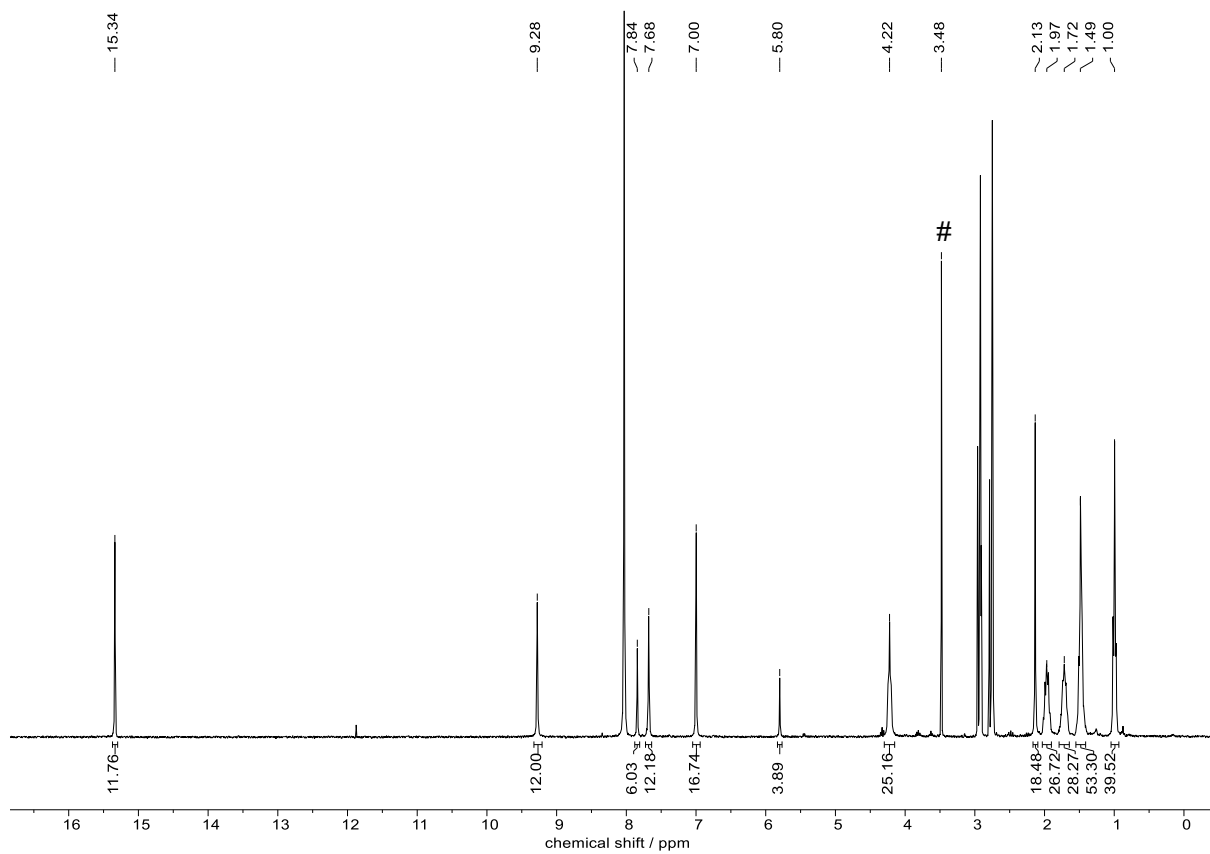


Figure S149. ^1H NMR spectrum (300 MHz, DMF-d_7) of [4+6]-exo cage 12. #: H_2O .

Formation of [4+6] *endo* cage **13**

An 8 mL screw-capped vial was charged with triaminotriptycene **7a** (20.1 mg, 34 μmol), 2,6-diformylphenol **10a** (7.8 mg, 52 μmol) and 1,3,5-trimethoxybenzene (1.7 mg, 10.11 μmol) as internal standard. Under argon atmosphere THF- d_8 (5 mL) and TFA (67 μL , 0.01 mol L $^{-1}$ in CDCl_3 , 0.67 μmol) were added and the orange solution was stirred at room temperature. After 1h, 2 h, 4h, 8 h, 24 h, 48 h, 72 h and 96 h aliquots of 0.5 mL were taken and analysed by GPC and ^1H NMR spectroscopy.

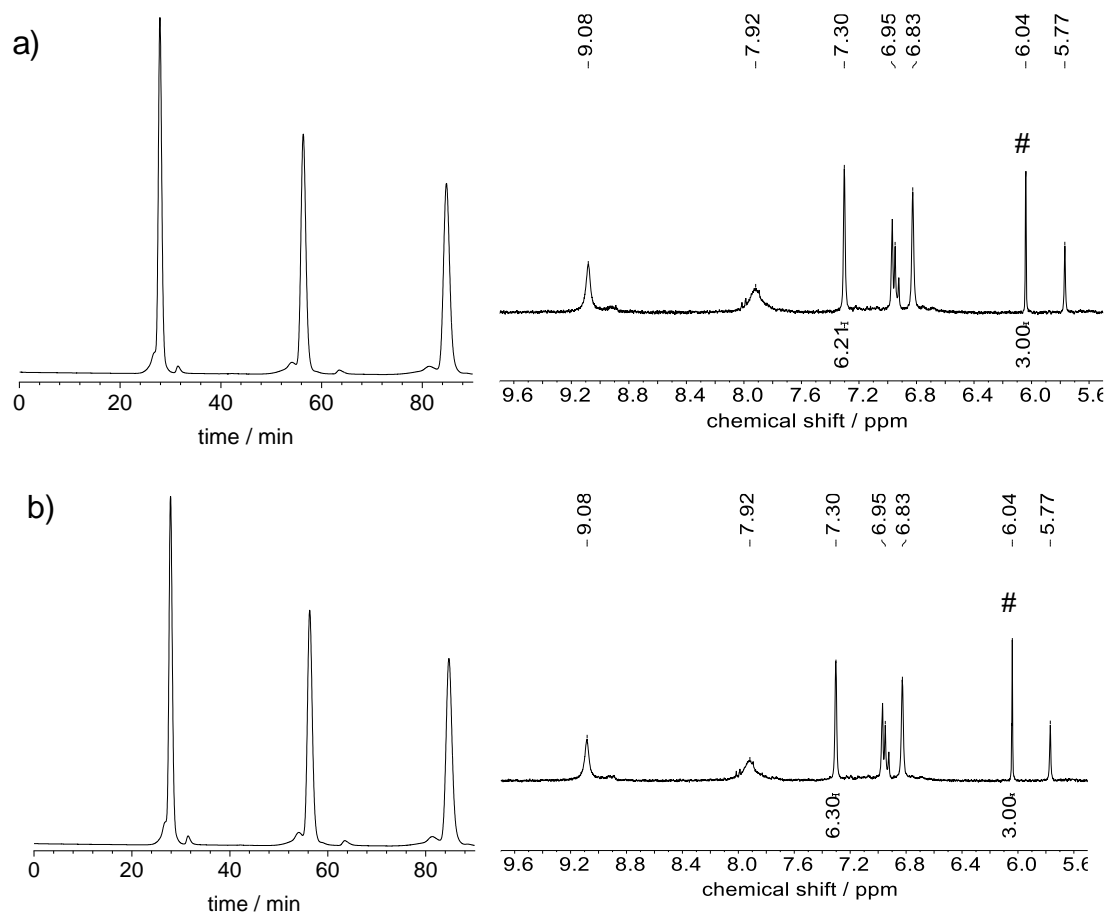


Figure S150. GPC chromatograms and ^1H -NMR spectra (300 MHz, THF- d_8) of samples from reaction mixtures for synthesis of [4+6]-*endo* cage **13** after a) 1 h and b) 2 h. #: 1,3,5-Trimethoxybenzene. ^1H NMR spectra show a range from 9.70 to 5.50 ppm.

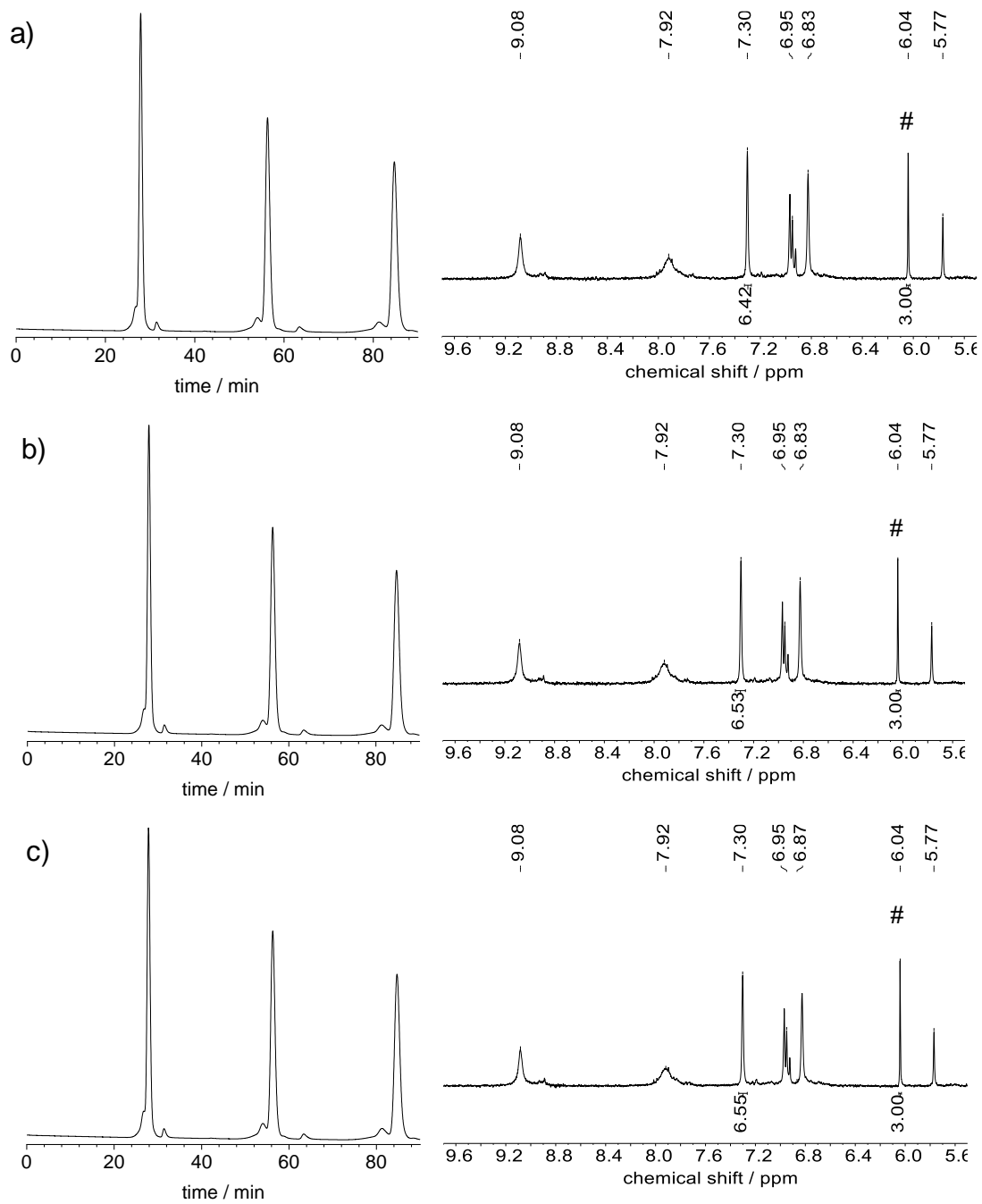


Figure S151. GPC chromatograms and ¹H-NMR spectra (300 MHz, THF-d₈) of samples from reaction mixtures for synthesis of [4+6]-*endo* cage **13** after a) 4 h, b) 8 h and c) 24 h. *: #: 1,3,5-Trimethoxybenzene. ¹H NMR spectra show a range from 9.70 to 5.50 ppm.

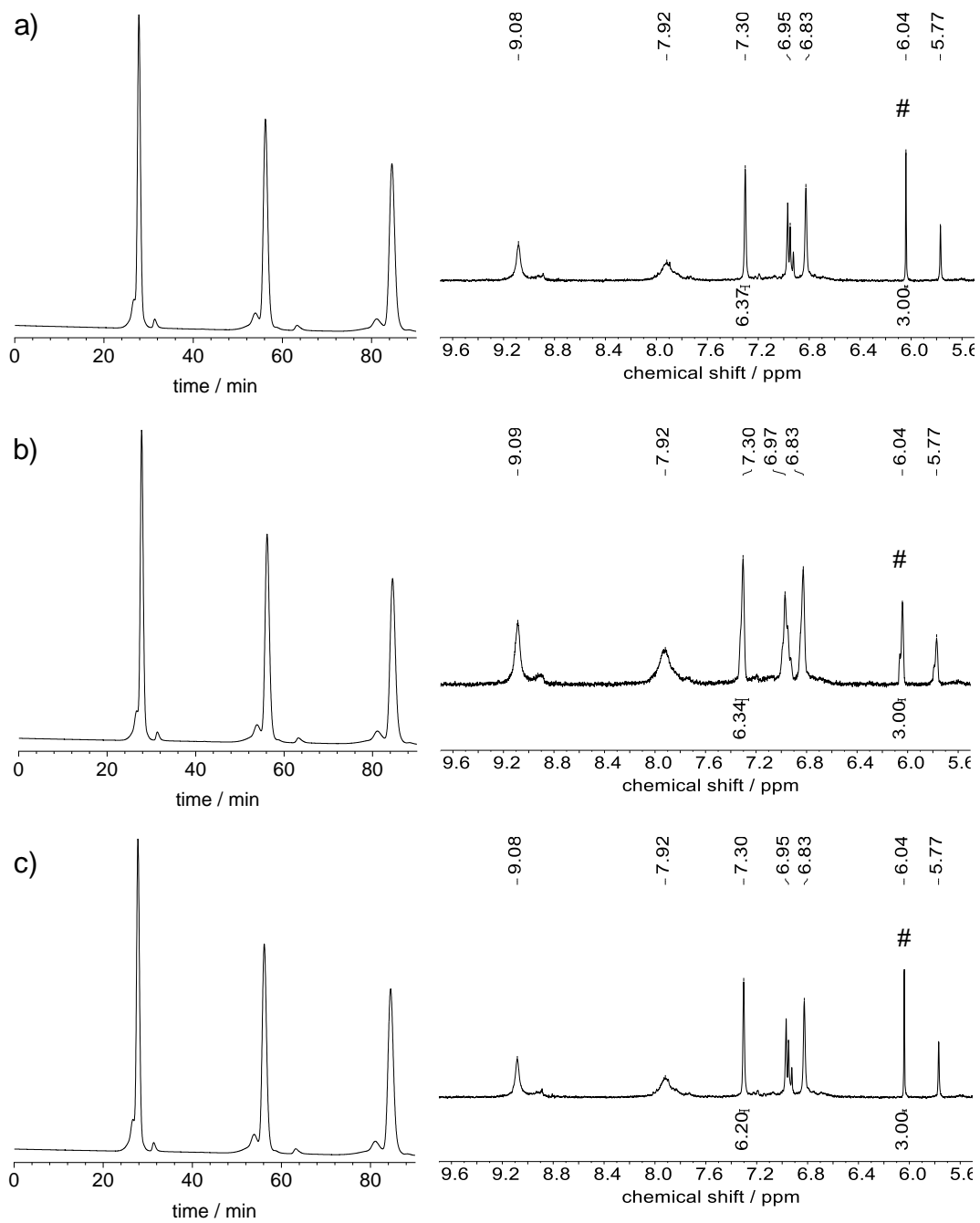


Figure S152. GPC chromatograms and ¹H-NMR spectra (300 MHz, THF-d₈) of samples from reaction mixtures for synthesis of [4+6]-endo cage **13** after a) 48 h, b) 72 h and c) 96 h. #: 1,3,5-Trimethoxybenzene. ¹H NMR spectra show a range from 9.70 to 5.50 ppm.

Formation of [4+6] *endo* cage **14**

An 8 mL screw-capped vial was charged with triaminotriptycene **7a** (20.0 mg, 33 μmol), 2,6-diformyl-4-*tert*-butylphenol **10b** (10.3 mg, 50 μmol) and 1,3,5-trimethoxybenzene (1.8 mg, 10.7 μmol) as internal standard. Under argon atmosphere THF- d_8 (5 mL) and TFA (67 μL , 0.01 mol L $^{-1}$ in CDCl_3 , 0.67 μmol) were added and the orange solution was stirred at room temperature. After 1 h, 2 h, 4 h, 8 h, 24 h, 48 h, 72 h and 96 h aliquots of 0.5 mL were taken and analysed by GPC and ^1H NMR spectroscopy.

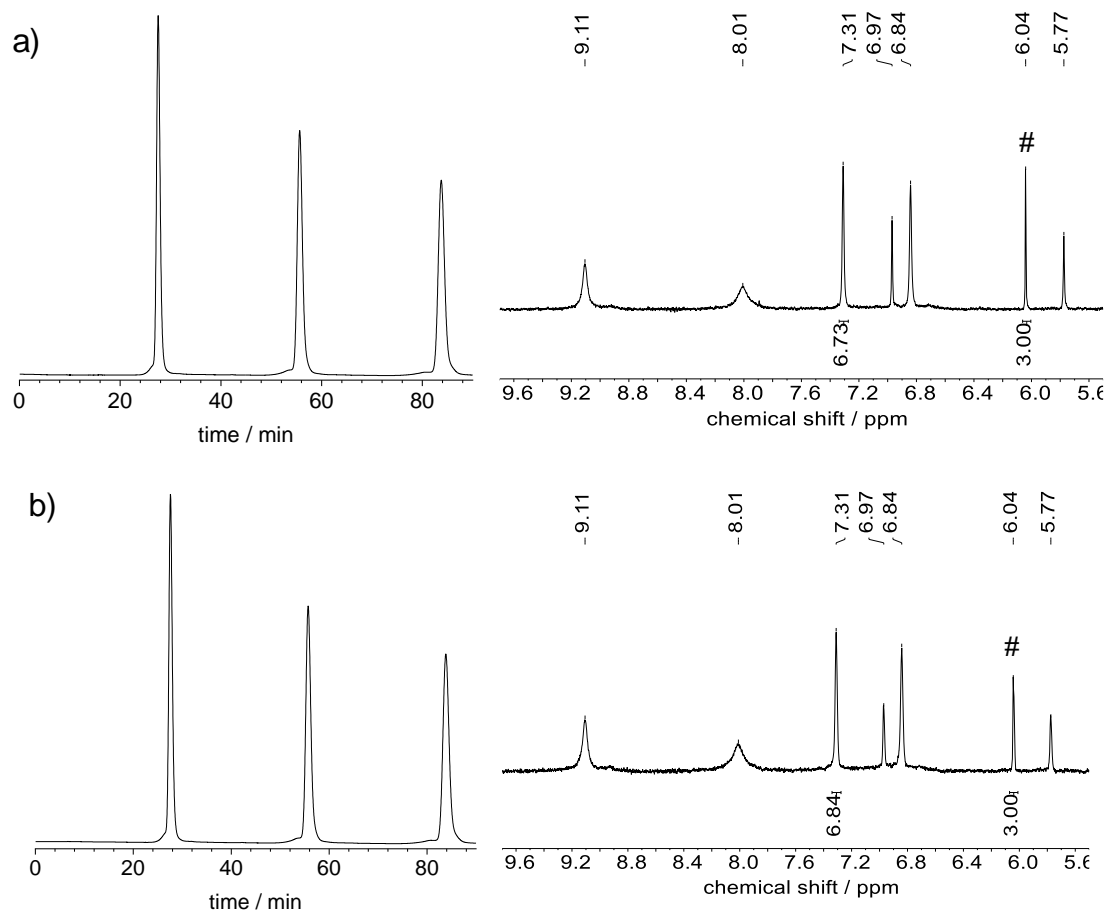


Figure S153. GPC chromatograms and ^1H -NMR spectra (300 MHz, THF- d_8) of samples from reaction mixtures for synthesis of [4+6]-*endo* cage **14** after a) 1 h and b) 2 h. #: 1,3,5-Trimethoxybenzene. ^1H NMR spectra show a range from 9.70 to 5.50 ppm.

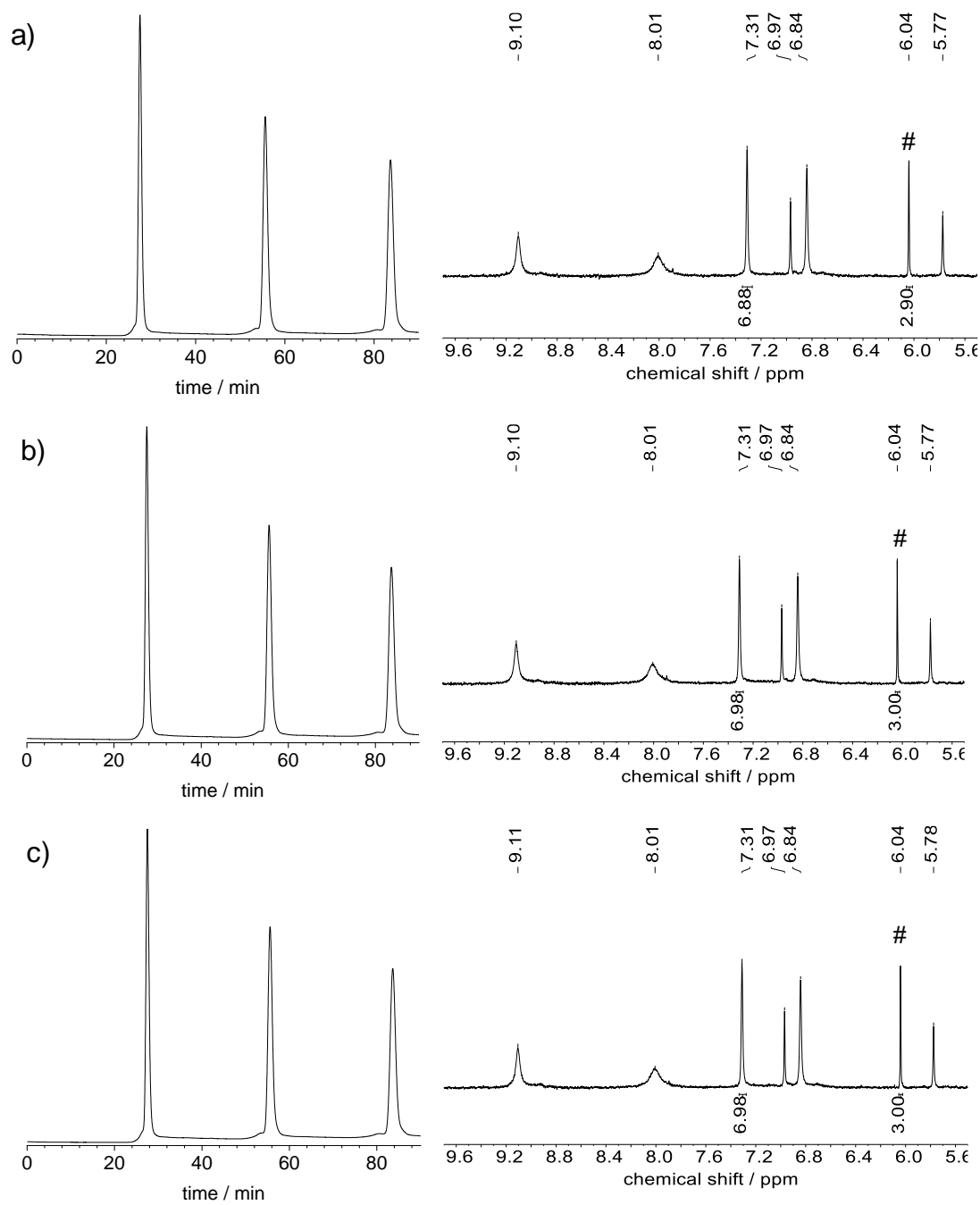


Figure S154. GPC chromatograms and $^1\text{H-NMR}$ spectra (300 MHz, THF-d_8) of samples from reaction mixtures for synthesis of [4+6]-endo cage **14** after a) 4 h, b) 8 h and c) 24 h. *: #: 1,3,5-Trimethoxybenzene. $^1\text{H NMR}$ spectra show a range from 9.70 to 5.50 ppm.

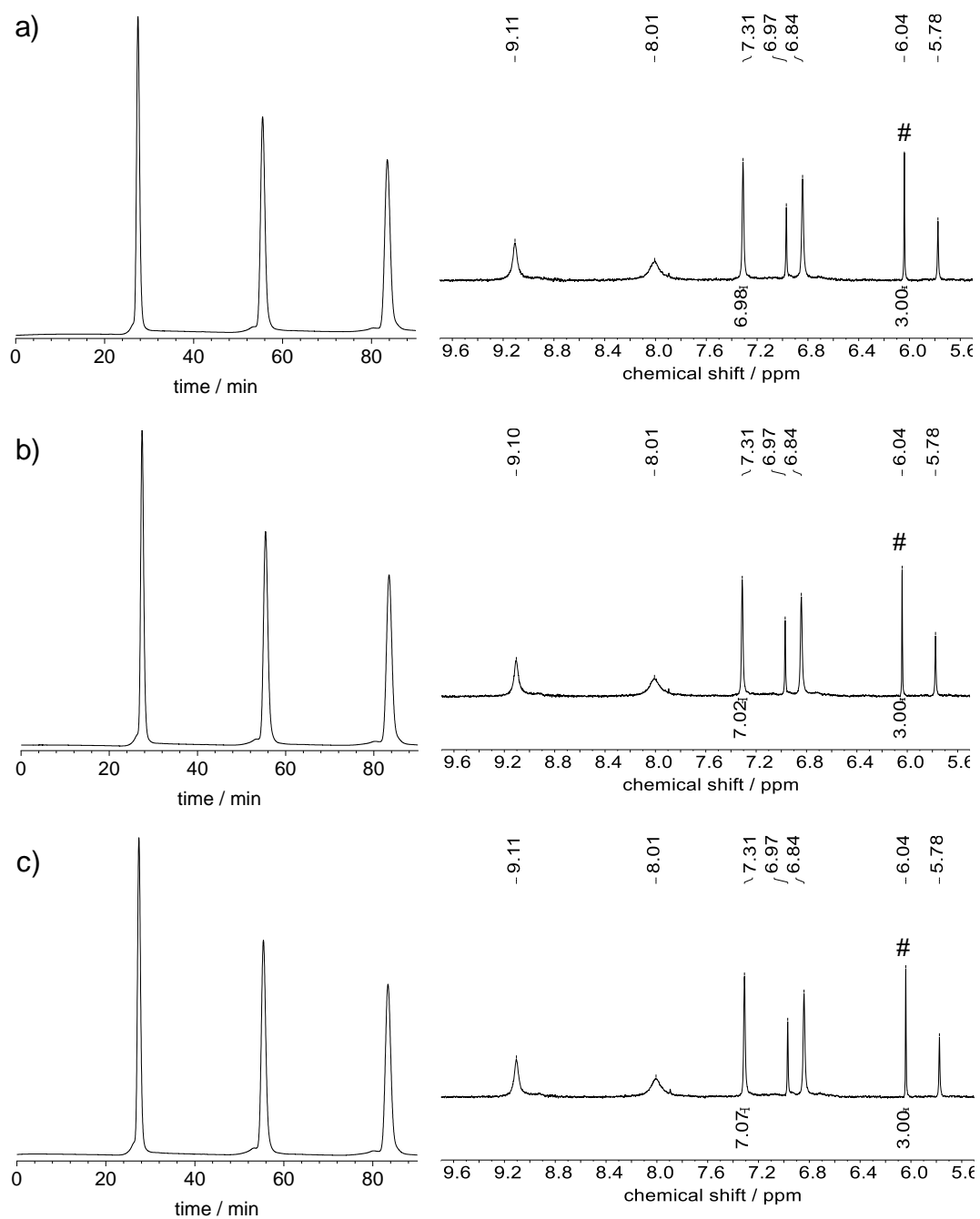


Figure S155. GPC chromatograms and $^1\text{H-NMR}$ spectra (300 MHz, THF-d_8) of samples from reaction mixtures for synthesis of [4+6]-endo cage **14** after a) 48 h, b) 72 h and c) 96 h. #: 1,3,5-Trimethoxybenzene. $^1\text{H NMR}$ spectra show a range from 9.70 to 5.50 ppm.

Determination of Yields by Using an Internal Standard

Yields have been calculated from measured ^1H NMR spectra of cages **11** – **14** using the following formula

$$\eta = \frac{I_1 n_{\text{standard}}}{I_0 n_{\text{theor.}}} * 100\%. \quad (2)$$

η is the yield, I_1 describes the measured integral of a cage signal in ^1H NMR spectrum, I_0 is the integral of the same signal, if the cage and the standard would have a 1:1 stoichiometry, n_{standard} is the amount of added standard and $n_{\text{theor.}}$ resembles the amount of cage molecules at 100% yield. Uncertainties were calculated applying propagation of uncertainty by using the error of weighing. Since the amount of standard has been calculated by its mass and the 100% yield amount of cage has been calculated by the amount of added 3,6,15-triamino-1,8,13-trihexyloxytriptycene divided by 4 the variables n_{standard} and $n_{\text{theor.}}$ can be exchanged using $n = m M^{-1}$:

$$\eta = \frac{I_1 m_{\text{standard}} M_{\text{amine}}}{I_0 M_{\text{standard}} m_{\text{amine}}} * 100\% = 400\% * \frac{I_1 M_{\text{amine}} m_{\text{standard}}}{I_0 M_{\text{standard}} m_{\text{amine}}}. \quad (3)$$

m_{standard} and m_{amine} are the added masses of 1,3,5-trimethoxybenzen and 3,6,15-triamino-1,8,13-trihexyloxytriptycene, respectively, as well as M_{standard} and M_{amine} are their molecular masses. The error of the yield $\Delta\eta$ can be calculated using propagation of uncertainty:

$$\Delta\eta = \frac{\delta\eta}{\delta m_{\text{standard}}} \Delta m_{\text{standard}} + \frac{\delta\eta}{\delta m_{\text{amine}}} \Delta m_{\text{amine}}. \quad (4)$$

Since the errors of the weighing is for both compounds the same value ($\Delta m = 0.0001$ g), equation (4) can be changed to:

$$\Delta\eta = \Delta m \frac{\delta\eta}{\delta m_{\text{standard}}} + \frac{\delta\eta}{\delta m_{\text{amine}}}. \quad (5)$$

From equation (5) results equation (6).

$$\Delta\eta = 400\% * \frac{I_1 M_{\text{amine}} \Delta m}{I_0 M_{\text{standard}}} \left(\frac{1}{m_{\text{amine}}} - \frac{m_{\text{standard}}}{m_{\text{amine}}^2} \right) \quad (6)$$

Table S30. Calculated yields and errors of cages **11 - 14** by ^1H NMR spectroscopy using 1,3,5-trimethoxybenzene as internal standard. Yields and errors were calculated applying equations (2) and (6). $\log t$ is the logarithm of time.

Cage		11		12		13		14	
time / h	$\log t$	$\eta / \%$	$\Delta\eta / \%$	$\eta / \%$	$\Delta\eta / \%$	$\eta / \%$	$\Delta\eta / \%$	$\eta / \%$	$\Delta\eta / \%$
1	0.00	15	1	34	2	63	3	72	4
2	0.30	15	1	46	2	63	3	73	4
4	0.60	19	1	55	3	65	3	74	4
8	0.90	23	1	62	3	66	4	75	4
24	1.38	37	2	61	3	66	4	75	4
48	1.68	53	3	61	3	64	3	75	4
72	1.86	62	4	67	3	64	3	75	4
96	1.98	62	4	69	3	62	3	76	4

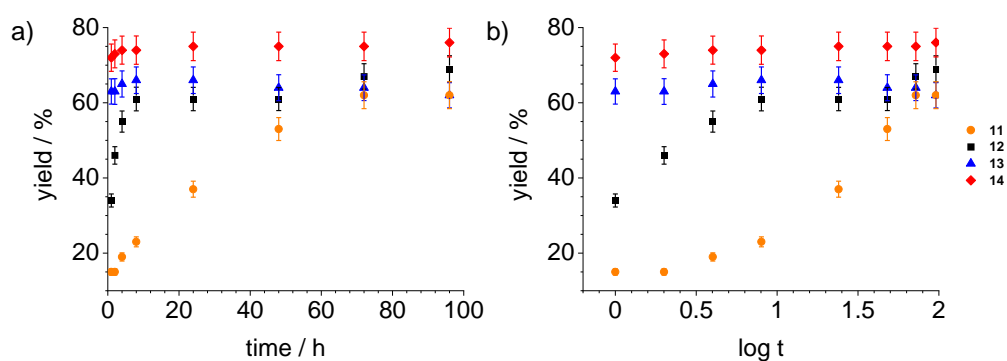


Figure S156. Diagram of yields of cages **11 – 14** determined by ^1H NMR spectroscopy in a) linear and b) logarithmic time axis. 1,3,5-Trimethoxybenzene has been used as internal standard. For every obtained yield an error bar has been illustrated.

12 VT NMR Spectra of [4+6]-*endo* cages 13 and 14

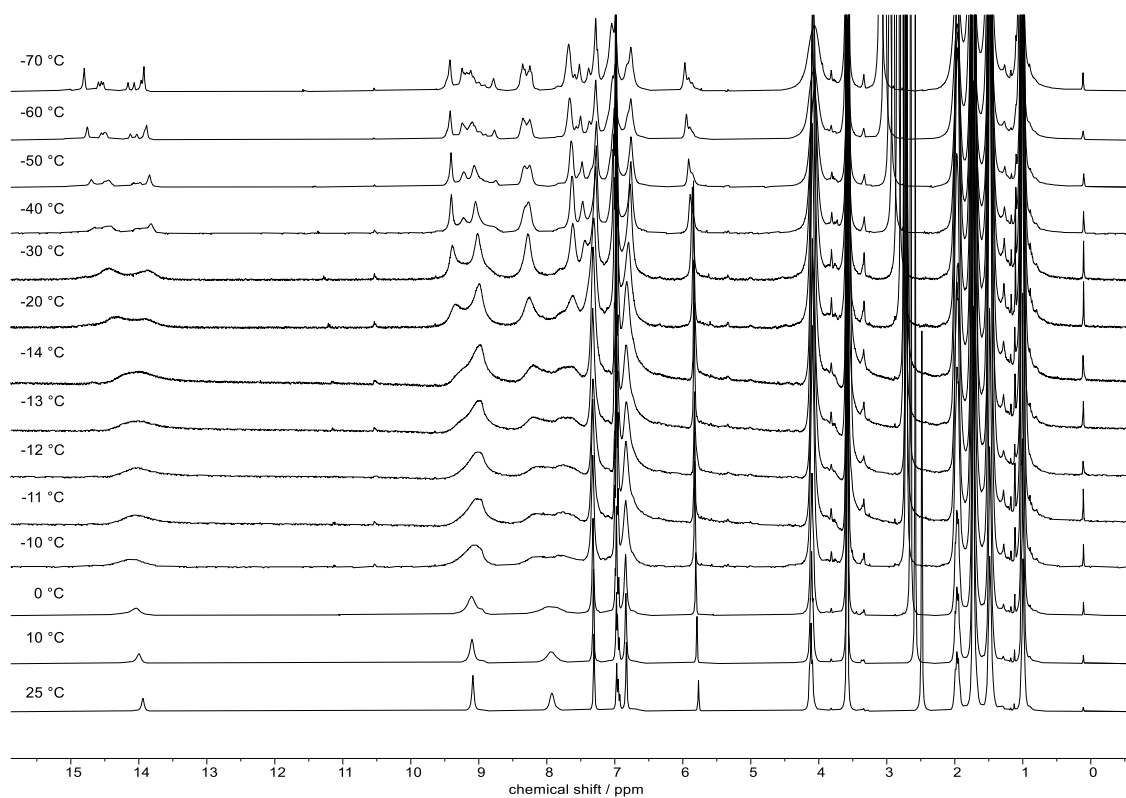


Figure S157. ¹H NMR spectra (300 MHz, THF-*d*₈) of [4+6]-*endo* cage 13 at different temperatures.

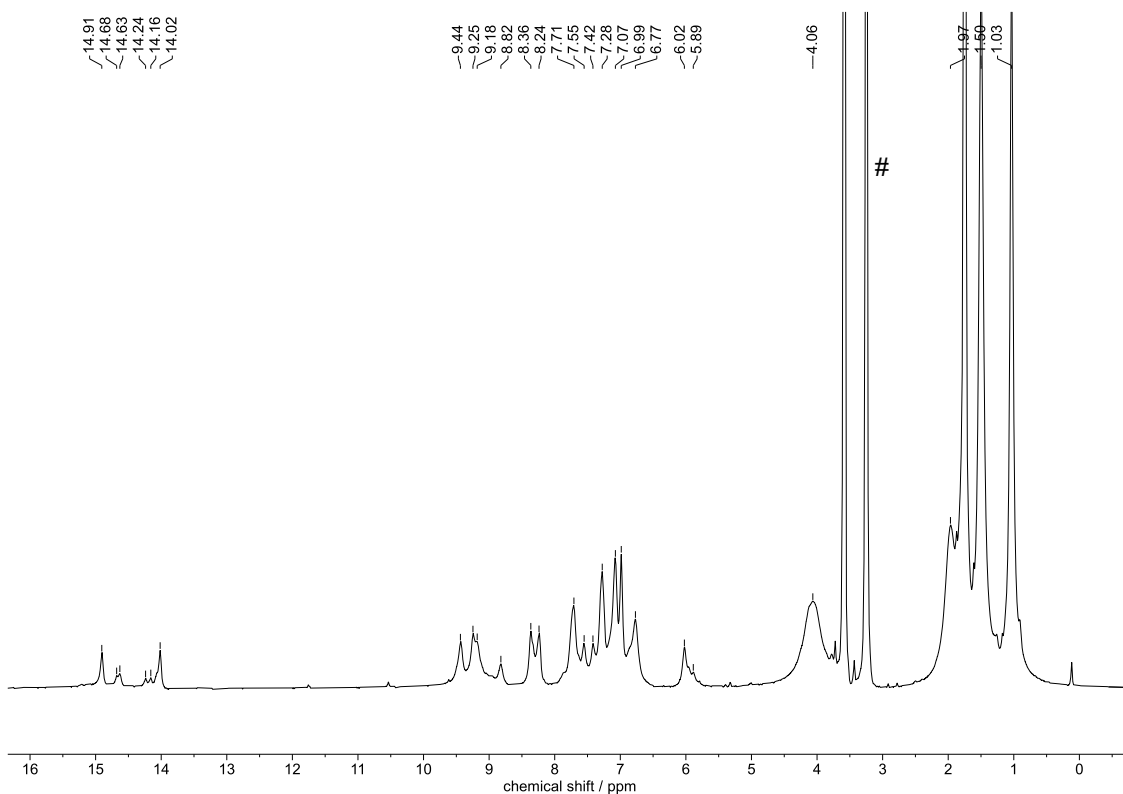


Figure S158. ¹H NMR spectrum (500 MHz, 203K, THF-*d*₈) of [4+6]-*endo* cage 13 at -70 °C. #: H₂O.

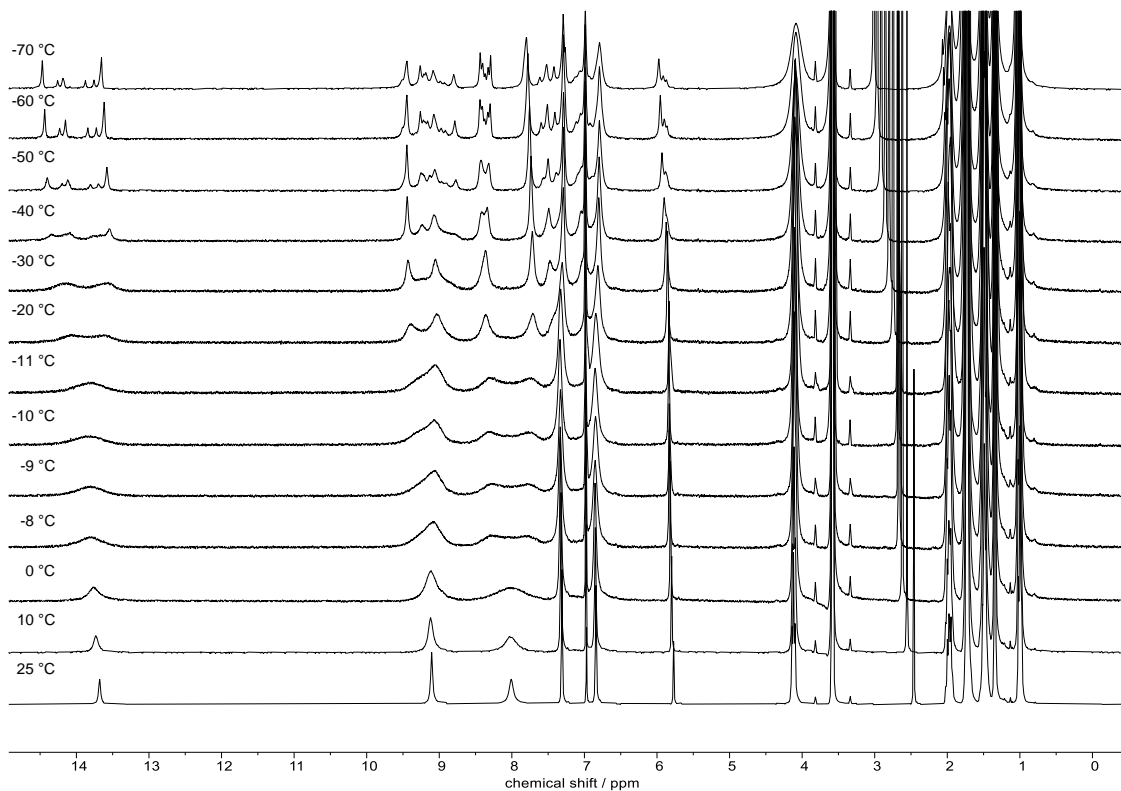


Figure S159. ¹H NMR spectra (300 MHz, THF-d₈) of *tert*-butylated [4+6]-*endo* cage **14** at different temperatures.

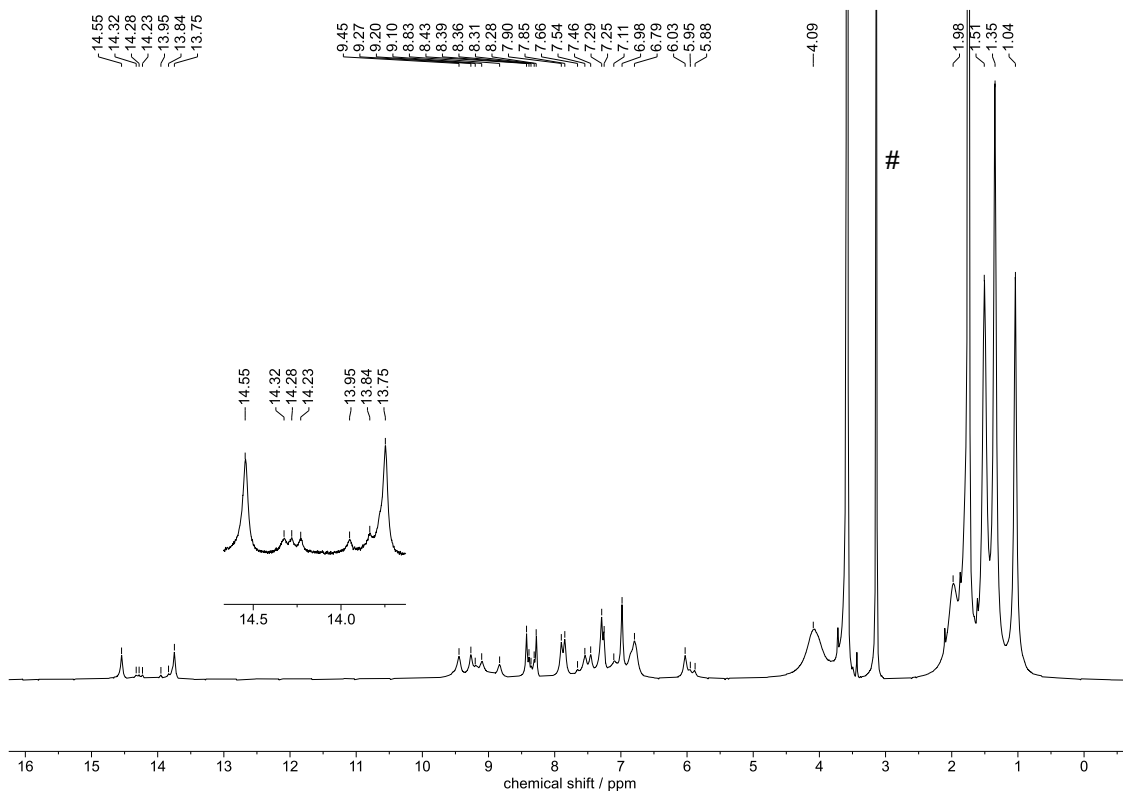


Figure S160. ¹H NMR spectrum (500 MHz, 203 K, THF-d₈) of *tert*-butylated [4+6]-*endo* cage **14** at -70 °C. #: H₂O.

13 Screening of Reaction Conditions

Synthesis of [4+4] cube 11

Table S31. Screening conditions for synthesis of [4+4] cube 11. eq_{TFA} is the used equivalents of TFA, V is the volume of used solvent, T is the reaction temperature, t is the reaction duration and η is the obtained yield after GPC purification.

No. ^a	eq_{TFA}	Solvent ^b	V / mL	T / °C	t / d	η / %
1	0.08	DMF	3	100	3	11
2	0.08	DMF	3	100	4	5
3	0.08	DMF	3	120	4	6
4	0.08	DMF	3	120	2	3
5	0.08	DCM	3	80	2	4
6	0.08	DCM	3	50	1	8
7	0.08	DMF	4	120	3	31
8	0.00	DMF	4	120	3	17
9	0.20	DMF	4	120	3	19
10	0.20	DMF	4.5	120	3	3
11	0.08	THF	5	80	2	6
12	0.20	DMF	5	120	3	8
13	0.20	THF	7	r. t.	11	1
14	0.20	THF	7	80	12	2
15	0.20	CHCl ₃	7	80	3	5
16	0.20	toluene	7	120	3	4
17	0.20	DMSO	7	120	3	8
18	0.00	DMF	7	120	3	6
19	0.08	DMF	7	120	3	17
20	0.20	DMF	7	120	4	6
21	0.20	DMF	7	120	12	14
22	0.28	DMF	7	120	3	5
23	0.20	DMF	7	140	3	3
24	0.20	DMF	7.5	120	3	4
25	0.20	1,4-dioxane	8	120	3	7
26	0.08	DMF	8	120	3	6
27	0.20	DMF	8	120	3	34
28	0.20	DMF	8	150	3	37
29	0.20	DMF	8	150	5	42
30	0.20	DMF	8	180	3	20
31	0.20	DMF	9	120	3	13
32	0.20	DMF	15	120	3	17
33	0.20	DMF	20	120	3	2

a) General conditions: 3,6,15-triamino-1,8,13-trihexyloxytryptcene (20.0 mg, 33 μ mol, 4.0 eq), 2,7,14-triformyl-1,8,13-trihydroxytryptcene (13.0 mg, 34 μ mol, 4.0 eq), TFA (0.01 mol L⁻¹ in dry DMF).

b) Dry solvents have been used.

Synthesis of [4+6]-exo cage 12

Table S32. Screening conditions for synthesis of [4+6]-exo cage 12. eq_{TFA} is the used equivalents of TFA, V is the volume of used dry DMF, T is the reaction temperature, t is the reaction duration and η is the obtained yield after GPC purification.

No. ^a	eq_{TFA}	V / mL	$T / ^\circ C$	t / d	$\eta / \%$
1	0.08	3	100	3	36
2	0.20	4	120	3	55
3	0.08	5	120	3	47
4	0.20	5	120	3	51
5	0.20	5	120	4	53
6	0.20	5	120	5	55
7	0.28	5	120	3	47
8	0.40	5	120	3	45
9	0.20	6	120	3	63
10	0.20	6	120	4	54
11	0.20	6	120	6	57
12	0.20	7	120	3	48

- a) General conditions: 3,6,15-triamino-1,8,13-trihexyloxytryptycene (20.0 mg, 33 μ mol, 4.0 eq), 3,5-diformyl-2,6-dihydroxytoluene (9.0 mg, 50 μ mol, 6.0 eq), TFA (0.01 mol L⁻¹ in dry DMF), dry DMF.

Synthesis of [4+6]-endo cage 13

Table S33. Screening conditions for synthesis of [4+6]-endo cage 13. $eq_{Aldehyde}$ is the used equivalents of 2,6-diformylphenol, V is the volume of used dry THF, t is the reaction duration and η is the obtained yield after SEC purification.

No. ^a	$eq_{Aldehyde}$	V / mL	t / d	$\eta / \%$
1	6.1	2	4	25
2	6.1	3	4	29
3	6.2	5	4	50
4	6.0	6	4	21

- a) General conditions: 3,6,15-triamino-1,8,13-trihexyloxytryptycene (20.0 mg, 33 μ mol, 4.0 eq), 2,6-diformylphenol, TFA (67 μ L, 0.01 mol L⁻¹ in dry THF, 0.08 eq), dry THF, r. t.

Synthesis of [4+6]-endo cage 14

Table S34. Screening conditions for synthesis of [4+6]-endo cage 14. *V* is the volume of used dry THF, *t* is the reaction duration and η is the obtained yield after SEC purification.

No. ^a	<i>V</i> / mL	<i>t</i> / d	η / %
1	3.0	3	41
2	4.5	4	46
3	5.0	4	62
4	7.0	4	47

- a) General conditions: 3,6,15-triamino-1,8,13-trihexyloxytriptycene (20.0 mg, 33 μ mol, 4.0 eq), 2,6-diformyl-4-*tert*-butylphenol (10.3 mg, 50 μ mol, 6.0 eq), TFA (67 μ L, 0.01 mol L⁻¹ in dry THF, 0.08 eq), dry THF, r. t.

14 References

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