

Table S1 Detailed list of European farmed pigs incorporated in the present study.

Breed	origin	no of pigs		Reference
		SLA class I	SLA class II	
([LRxYS]xD)	DK	101	0	Pedersen <i>et al.</i> Vet. Immunol. Immunopathol. (2014) 162:108-116
	NL	8	8	Christine Jansen (2018, this study)
	UK	31	31	William T. Golde (2019-20, this study)
([LRxLW]xP)	Austria	9	9	Talker <i>et al.</i> Veterinary Research (2015) 46:52
		31	31	Talker <i>et al.</i> Journal of Virology (2016) 90:9364-9382
		8	8	Sabine Hammer (2012, this study)
LR x LW	UK	26	8	Franzoni <i>et al.</i> PLoS ONE (2013) 8:e84246
		10	10	Mokhtar <i>et al.</i> Frontiers in Immunology (2016) 7:40
	Spain	26	26	Burgara-Estrella <i>et al.</i> Viruses (2013) 5:663-677
		73	73	De León <i>et al.</i> Viruses (2020) 8:E513
YS x NL LR	UK	22	22	Mokhtar <i>et al.</i> Vaccine (2014) 32:6828-6837
YS x Ham	Sweden	44	0	Marianne Jensen-Waern (2015-16, this study)
YS		33	0	
GER LR	Germany	20	20	Ulrike Gimsa (2012, this study)
		65	65	Gimsa <i>et al.</i> Immunogenetics (2017) 69:39-47
AUT P	Austria	27	27	Essler <i>et al.</i> Animal Genetics (2013) 44:202-205
LW	UK	3	3	Simon Graham (2012, this study)
	CH	12	0	Artur Summerfield (2015, this study)
		549	341	Total no of pigs

LRYS/D, Animals were 25% Landrace (LR), 25% Yorkshire (YS), and 50% Duroc (D); LWLR/P, Animals were 25% Large White (LW), 25% Landrace (LR), and 50% Pietrain (P); LR x LW, Large White/Landrace crosses; YS x NL LR, Yorkshire/Dutch Landrace crosses; YS x Ham, Yorkshire/Hampshire crosses; GER LR, German Landrace; AUT P, Austrian Pietrain; LW, Large White.

Table S1 – References

- Pedersen L.E., Jungersen G., Sorensen M.R., Ho C.-S. & Vadkær D.F. (2014) Swine leukocyte antigen (SLA) class I allele typing of Danish swine herds and identification of commonly occurring haplotypes using sequence specific low and high resolution primers. *Veterinary Immunology and Immunopathology* 162, 108-16. <https://doi.org/10.1016/j.vetimm.2014.10.007>
- Talker S.C., Koinig H., Stadler M., Graage R., Klingler E., Ladinig A., Mair K.H., Hammer S.E., Dürrwald R., Ritzmann M., Saalmüller A. & Gerner W. (2015) Magnitude and kinetics of multifunctional CD4⁺ and CD8⁺ T cells in pigs infected with swine influenza A virus. *Veterinary Research* 46, 52-67. <https://doi.org/10.1186/s13567-015-0182-3>.
- Talker S.C., Stadler M., Koinig H.C., Mair K.H., Rodriguez-Gomez I.M., Graage R., Zell R., Dürrwald R., Starick E., Harder T., Weissenböck H., Lamp B., Hammer S.E., Ladinig A., Saalmüller A. & Gerner W. (2016) Influenza A virus infection in pigs attracts multifunctional and cross-reactive T cells to the lung. *Journal of Virology* 90, 9364-82. <https://doi.org/10.1128/JVI.01211-16>.
- Franzoni G., Kurkure N.V., Essler S.E., Everett H.E., Bodman-Smith K., Crooke H.R. & Graham S.P. (2013) Proteome-wide screening reveals immunodominance in the CD8 T cell response against classical swine fever virus with antigen-specificity dependant on MHC class I haplotype. *PLoS One* 8, e84246. <https://doi.org/10.1371/journal.pone.0084246>
- Mokhtar H., Pedrera M., Frossard J.P., Biffar L., Choudhury B., Hammer S.E., Kvisgaard L.K., Larsen L.E., Stewart G.R., Somaravarapu S., Steinbach F. & Graham S.P. (2016) The non-structural protein 5 and matrix protein are major antigenic targets of T cell immunity to porcine reproductive and respiratory syndrome virus. *Frontiers in Immunology* 7, 40. <https://doi.org/10.3389/fimmu.2016.00040>.
- Burgara-Estrella A., Díaz I., Rodríguez-Gómez I., Essler S.E., Hernández J. & Mateu E. (2013) Predicted peptides from non-structural proteins of porcine reproductive and respiratory syndrome virus are able to induce INF- γ and IL-10. *Viruses* 5, 663-677. <https://doi.org/10.3390/v5020663>
- De León P., Cañas-Arranz R., Saez Y., Forner M., Defaus S., Bustos M.J., Torres E., Rodríguez-Pulido M., Andreu D., Blanco E., Sobrino F. & Hammer S.E. (2020) Association of porcine Swine Leukocyte Antigen (SLA) haplotypes with B- and T-cell immune response to foot-and-mouth disease virus (FMDV) peptides. Special issue: Evaluation of Vaccine Immunogenicity, *Vaccines (Basel)* 8, E513. <https://doi.org/10.3390/vaccines8030513>.
- Mokhtar H., Eck M., Morgan S.B., Essler S.E., Frossard J.P., Ruggli N. & Graham S.P. (2014) Proteome-wide screening of the European porcine reproductive and respiratory syndrome virus reveals a broad range of T cell antigen reactivity. *Vaccine* 32, 6828-37. <https://doi.org/10.1016/j.vaccine.2014.04.054>
- Gimsa U., Ho C.S. & Hammer S.E. (2017) Preferred SLA class I/class II haplotype combinations in German Landrace pigs. *Immunogenetics* 69, 39-47. <https://doi.org/10.1007/s00251-016-0946-6>.
- Essler S.E., Ertl W., Deutsch J., Ruetgen B.C., Groiss S., Stadler M., Wysoudil B., Gerner W., Ho C.-S. & Saalmüller A. (2013) Molecular characterization of swine leukocyte antigen gene diversity in purebred Pietrain pigs. *Animal Genetics* 44, 202-205. <https://doi.org/10.1111/j.1365-2052.2012.02375.x>

Table S2a Plate layout of the PCR primer panel for genotyping swine leukocyte antigen (SLA) class I alleles.

	H	G	F	E	D	C	B	A
1	Negative	209 bp	147 bp	181 bp	220 bp	163 bp	220 bp	138 bp
	Control	SLA-1*01XX(all)	SLA-1*02XX(all)	SLA-1*04XX(all)	SLA-1*05XX(all); SLA-1*16:02	SLA-1*06XX(all); SLA-1*13:01	SLA-1*07XX(all)	SLA-1*08XX(all)
2	195 bp	180 bp	182 bp	119 bp	211 bp	219 bp	253 bp	173 bp
	SLA-1*09XX(all)	SLA-1*10XX(all)	SLA-1*11XX(all)	SLA-1*12XX(all)	SLA-1*13XX(all)	SLA-1*14XX(all)	SLA-1*15XX(all); SLA-1*14:02; SLA-2*01XX(all)	SLA-1*16XX(all)
3	134 bp	208 bp	193 bp	130 bp	196 bp	177 bp	183 bp	192 bp
	SLA-1*16XX(all); SLA-1*16:02;	SLA-1*17:01	SLA-1*07:03; SLA-2*11:04	SLA-1*07:03; SLA-1*07:05	SLA-1*18:01	SLA-3*01XX(all)	SLA-3*03XX(all); SLA-3*08XX(all)	SLA-3*04XX(all); SLA-3*04:04 SLA-2*15XX(all)
	SLA-2*03XX(all); SLA-2*11:04							
4	138 bp	187 bp	152 bp	152 bp	139 bp	139 bp	172 bp	138 bp
	SLA-3*05XX(all)	SLA-3*06XX(all); SLA-3*07XX(all)	SLA-3*07XX(all)	SLA-3*06:01	SLA-3*06:02	SLA-1*11:03; SLA-3*04XX(all); 04:04 SLA-3*03XX excl. 03:06	SLA-2*01XX(all)	SLA-2*02XX(all)
5	89 bp	311 bp	127 bp	125 bp	199 bp	126 bp	177 bp	104 bp
	SLA-2*03XX(all)	SLA-2*04XX(all)	SLA-2*05XX(all)	SLA-2*06XX(all)	SLA-2*07XX(all)	SLA-2*08XX(all)	SLA-2*09XX(all)	SLA-2*10XX(all)
6	123 bp	160 bp	117 bp	131 bp	90 bp	196 bp	138 bp	175 bp
	SLA-2*11XX(all)	SLA-1*14:02;	SLA-1*11:03;	SLA-1*09XX(all);	SLA-2*15XX(all);	SLA-2*06:01~02 / 06:06;	SLA-1*07XX(all)	SLA-2*16:02
		SLA-2*12XX(all)	SLA-2*13XX(all)	SLA-2*14XX(all) 16XX(all); 16:02	SLA-2*11:04	09:01/09:02; 16XX(all)	SLA-2*02XX(all) SLA-2*16XX(all); SLA-2*17:01	

Reference: Ho *et al.* Animal Genetics (2009) 40:468-78. <https://doi.org/10.1111/j.1365-2052.2009.01860.x>.

Table S2b Plate layout of the PCR primer panel for genotyping swine leukocyte antigen (SLA) class II alleles.

	H	G	F	E	D	C	B	A
7	Negative	162 bp	203 bp	115 bp	180 bp	206 bp	172 bp	122 bp
	Control	DRB1*01XX(all)	DRB1*01XX(all)	DRB1*02XX(all)	DRB1*03XX(all)	DRB1*04XX(all)	DRB1*05XX(all)	DRB1*06XX(all)
		DRB1*13:02/01:03	DRB1*13:02					
		DRB1*01:04/07:04	DRB1*01:04/01:05					
8	133 bp	108 bp	105 bp	157 bp	135 bp	109 bp	186 bp	182 bp
	DRB1*07XX(all)	DRB1*08XX(all)	DRB1*09XX(all)	DRB1*09XX(all)	DRB1*10XX(all)	DRB1*11XX(all)	DRB1*12XX(all)	DRB1*13XX(all)
		DRB1*03:02	DRB1*09:05/09:06	DRB1*16:02/17:01	DRB1*10:06/10:07			
		DRB1*16:01/16:03	DRB1*09:04	DRB1*09:06/09:07				
9	113 bp	160 bp	134 bp	202 bp	197 bp	117 bp	160 bp	118 bp
	DRB1*14XX(all)	DRB1*14:02	DRB1*15:01	DRB1*17:01	DRB1*10:07/10:08	DRB1*04:03~04	DRB1*04:01~02	DRB1*04:01~02
				DRB1*15:02	DRB1*09:07	DRB1*11:01/11:03	DRB1*04:05/04:06	DRB1*04:05/04:06
				DRB1*15:03		DRB1*14:02		
10	165 bp	180 bp	146 bp	166 bp	197 bp	193 bp	204 bp	154 bp
	DQB1*01XX(all)	DQB1*01XX(all)	DQB1*02XX(all)	DQB1*03XX(all)	DQB1*04XX(all)	DQB1*05XX(all)	DQB1*06XX(all)	DQB1*07XX(all)
		DQB1*11:01					DQB1*06:03	
11	148 bp	146 bp	180 bp	161 bp	193 bp	176 bp	133 bp	165 bp
	DQB1*08XX(all)	DQB1*09XX(all)	DQB1*09XX(all)	DQB1*10:01	DQB1*02XX	DQB1*02:02 / 02:06	DQB1*02XX	DQB1*02:03/02:06
	DQB1*08:06		DQB1*03:04	DQB1*10:02	DQB1*03:04/08:06			DQB1*03:01
12	173 bp	141 bp	210 bp	160 bp	124 bp	148 bp	111 bp	120 bp
	DQB1*02:02/02:04	DQA*01XX(all)	DQA*02XX(all)	DQA*03XX(all)	DQA*04XX(all)	DQA*04XX(all)	DQA*05XX(all)	DQA*06:01
	DQB1*08:06		DQA*06:01					

Reference: Ho *et al.* Animal Genetics (2010) 41:428-32. <https://doi.org/10.1111/j.1365-2052.2010.02019.x>.

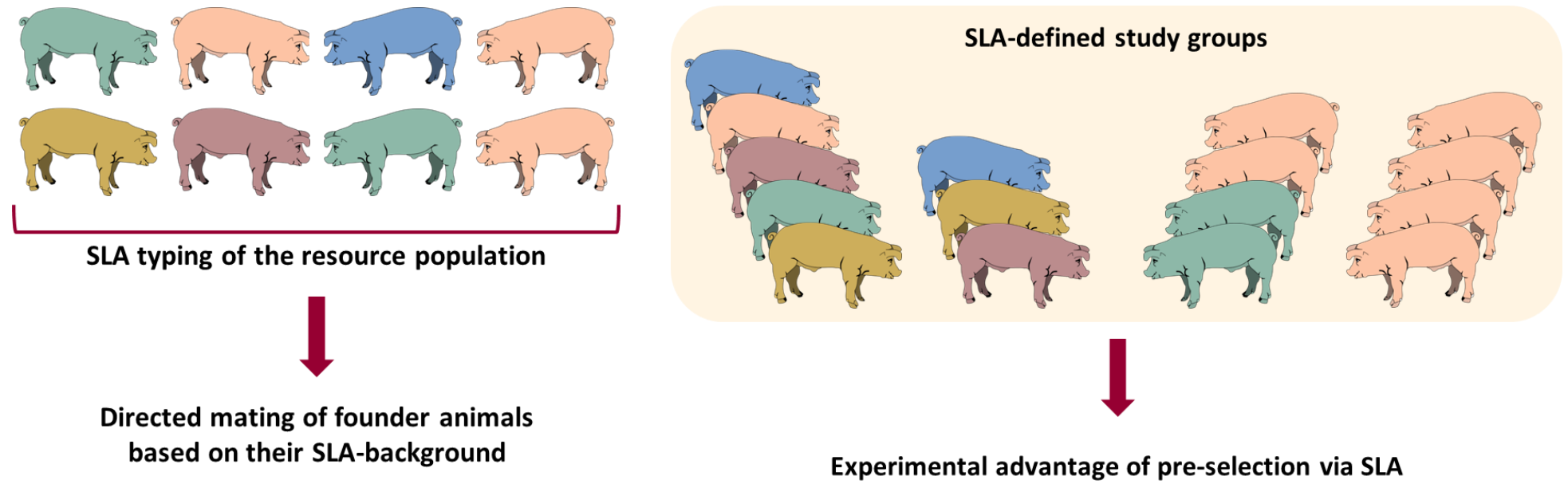


Figure S1 Two basics concepts for Swine leukocyte antigen (SLA) haplotyping-assisted animal trials in vaccine and transplantation research.

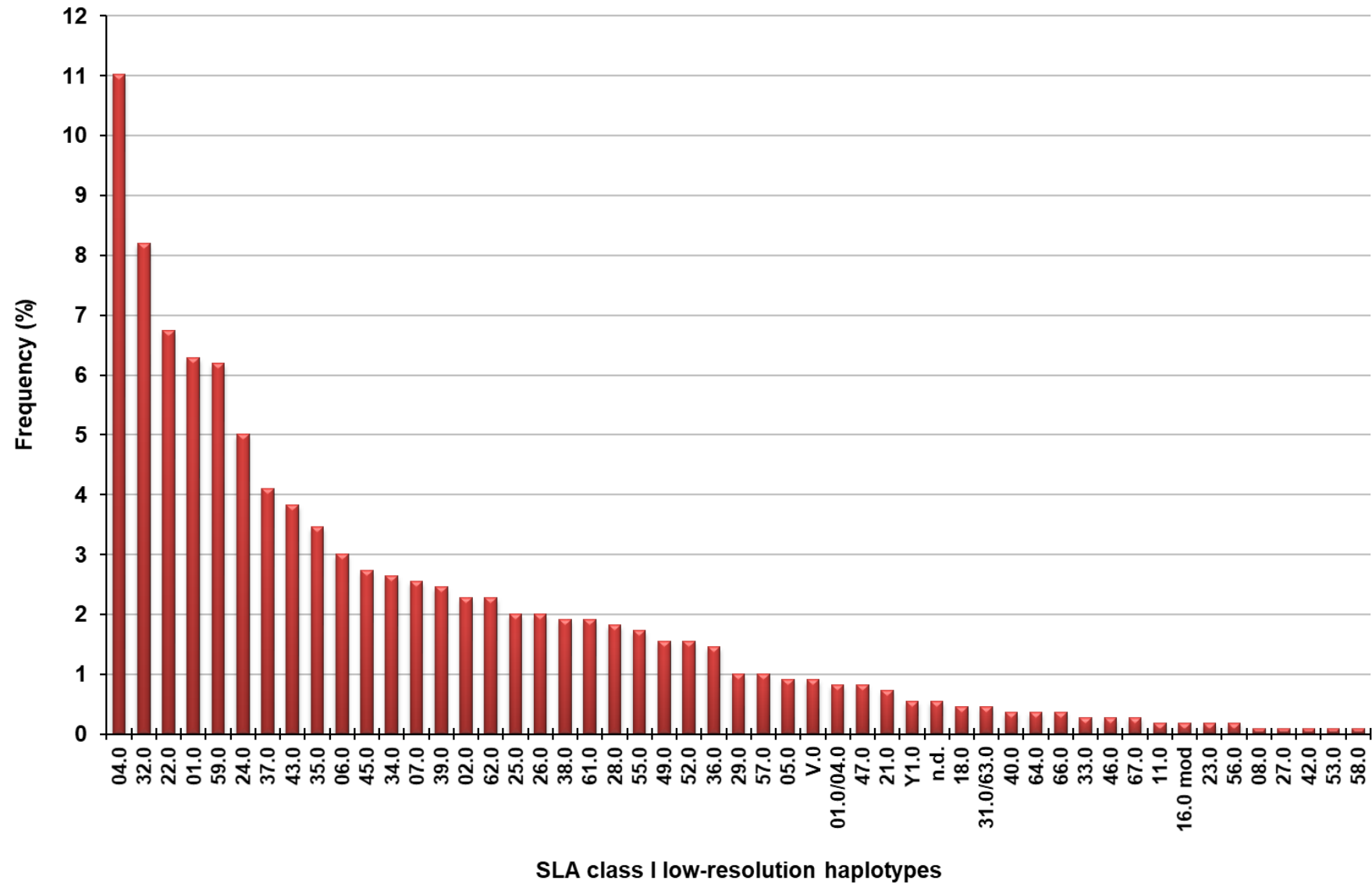


Figure S2a Frequency of swine leukocyte antigen (SLA) class I low-resolution haplotypes identified in 549 European farmed pigs.

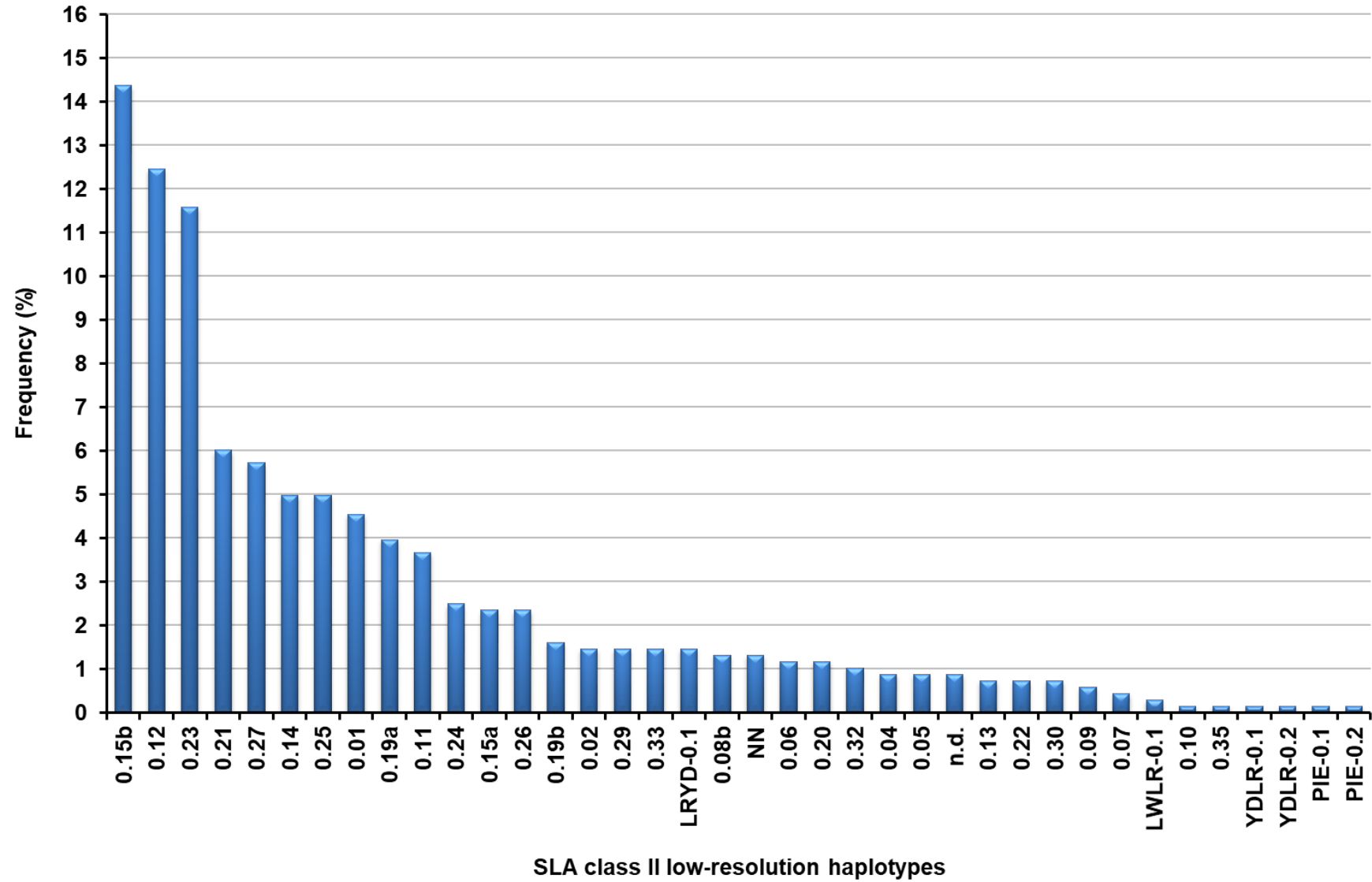


Figure S2b Frequency of swine leukocyte antigen (SLA) class II low-resolution haplotypes identified in 341 European farmed pigs.

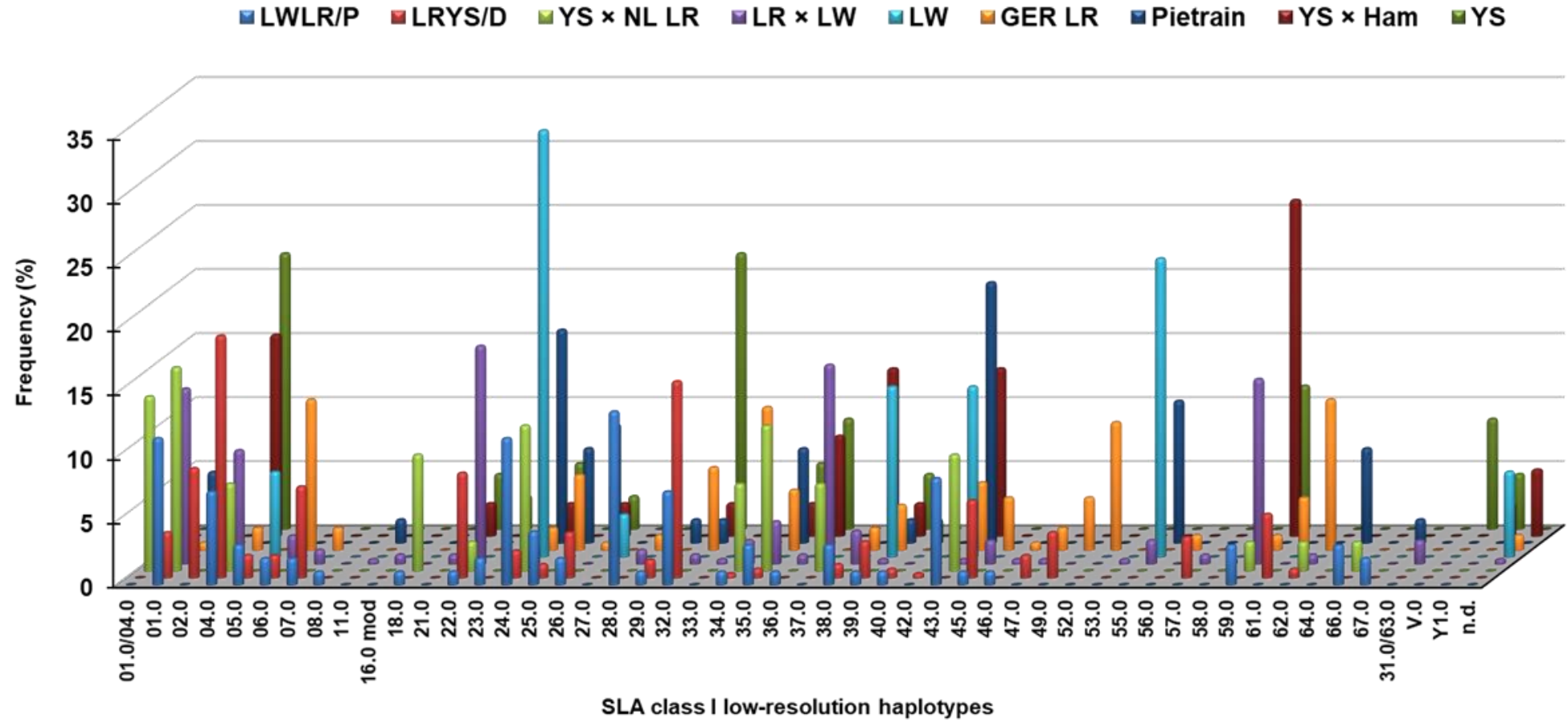


Figure S3a Swine leukocyte antigen (SLA) class I low-resolution haplotype diversity in nine European commercial pig populations. LWLR/P, Animals were 25% Large White (LW), 25% Landrace (LR), and 50% Pietrain (P); LRYS/D, Animals were 25% Landrace (LR), 25% Yorkshire (YS), and 50% Duroc (D); YS x NL LR, Yorkshire/ Dutch Landrace crosses; LR x LW, Large White/Landrace crosses; GER LR, German Landrace; YS x Ham, Yorkshire/Hampshire crosses.

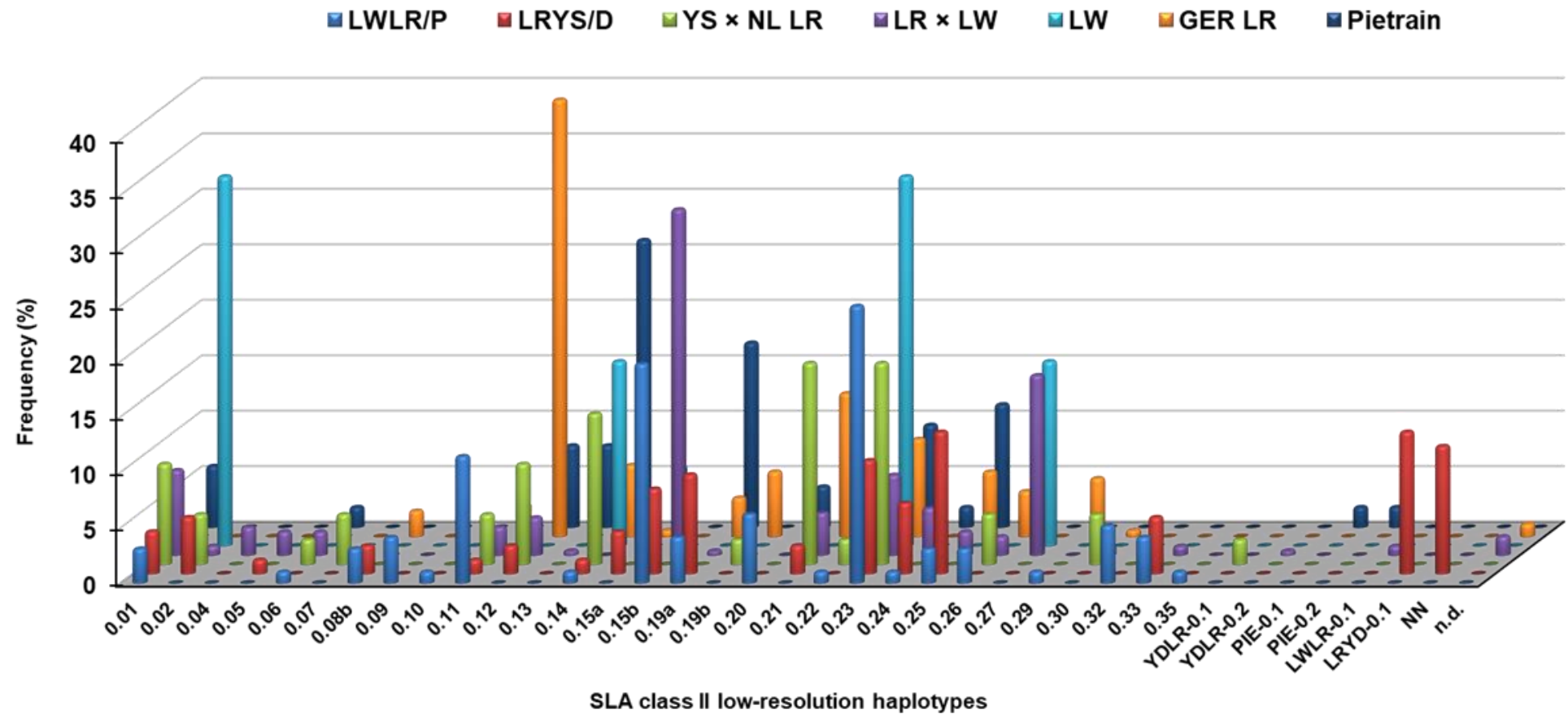


Figure S3b Swine leukocyte antigen (SLA) class II low-resolution haplotype diversity in seven European commercial pig populations. LWLR/P, Animals were 25% Large White (LW), 25% Landrace (LR), and 50% Pietrain (P); LRYS/D, Animals were 25% Landrace (LR), 25% Yorkshire (YS), and 50% Duroc (D); YS x NL LR, Yorkshire/ Dutch Landrace crosses; LR x LW, Large White/Landrace crosses; GER LR, German Landrace.