Supplementary Information

Facile Access to C-Glycosyl Amino Acids and Peptides via Ni-Catalyzed Reductive Hydroglycosylation of Alkynes

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1. General Information

All commercially available chemicals were used as received without further purification, unless otherwise stated. The catalytic reactions were carried out in Schlenk flasks under N₂ atmosphere using pre-dried glassware. Thin layer chromatography (TLC) was performed on TLC silica gel 60 F254 (Merck). Preparative layer chromatography was performed on PLC silica gel 60 F254 (Merck, 0.5 mm). The TLC plates were visualized with UV light and/or by charring with EtOH/H₂SO₄ (8%, v/v) or staining with a basic solution of KMnO₄. NMR spectra were measured on Bruker AM 400, Bruker AM 500, Agilent 500, or 600 MHz NMR spectrometer using TMS as an internal standard. Chemical shifts were given relative to TMS (0.00 ppm), CDCl₃ (7.26 ppm for ¹H NMR, 77.16 ppm for ¹³C NMR), D₂O (4.79 ppm for ¹H NMR), or CD₃OD (3.31 ppm for ¹H NMR, 49.03 ppm for ¹³C NMR). The following abbreviations were used to describe peak splitting patterns when appropriate: br = broad, s = singlet, d =doublet, t = triplet, q = quartet, dd = doublet of doublet, m = multiplet. Coupling constants (J) were reported in hertz (Hz). High-resolution mass spectra were recorded with IonSpec 4.7 Tesla FTMS or APEXIII 7.0 Tesla FTMS. Mass spectroscopy data of the products were collected on an HRMS-TOF instrument using ESI ionization. Optical rotations were measured on an Anton Paar MCP5500 S2 polarimeter.

Experimental Section 2.

2.1 Preparation of glycosyl bromides

Monosaccharides

BzÓ

ΒzÓ

BzO

AcO

ÒВz

ÓAc OAc



Supplementary Figure 1. Glycosyl bromides used in this study.

ÓAc

S29

S33

Method A. The glycosyl bromides were prepared from the corresponding glycosyl acetate or benzoate via treatment with 33% HBr in HOAc¹. (pls check if the cited ref. 1 correct)



Supplementary Figure 2. General Method A for preparation of glycosyl bromides.

Unless otherwise stated, the fully protected glycosyl acetate/benzoate was dissolved in CH₂Cl₂ (0.6 M, x mL) and cooled to 0 °C, 33% HBr in HOAc (0.5x mL) was added dropwise under 0 °C. The ice bath was removed and the mixture was stirred at room temperature for 2–7 h, until the disappearance of the starting material as monitored by TLC. The mixture was diluted with CH_2Cl_2 and washed with ice-water (3 times), sat. aq. NaHCO₃ (3 times) or 0.5 M aq. KHCO₃ (3 times), and finally brine (3 times). The organic layer was dried over Na₂SO₄, filtered, and concentrated to give the glycosyl bromide. In most cases, the bromide product was pure and was used directly for the subsequent coupling reaction or stored at -20 °C.

Compounds **1a**, **1f** and **1i** are known compounds and were prepared according to the literature methods¹. (pls check if the cited ref. 1 correct)



Supplementary Figure 3. Preparation of GlcN bromide 1b.

Compound **1b** is a known compound and was prepared employing a modified literature procedure^{2,3}.

To a solution of glucosamine hydrochloride (4.3 g, 20 mmol) in MeOH (100 mL) were added sodium hydroxide (0.88 g, 22 mmol) and a trace amount of H₂O. After stirring rigorously at room temperature for 0.5 h, phthalic anhydride (1.63 g, 11 mmol) was added and the stirring was kept for ~0.5 h until a clear solution was resulted. To the solution was added Et₃N (3.0 mL, 22 mmol). The mixture was stirred for 15 min and white syrup appeared. The mixture was treated with a second portion of phthalic anhydride (1.63 g, 11 mmol) and stirred overnight at room temperature. The mixture was cooled to 0 °C, filtered, and washed with cold MeOH. The filtrates were dried under reduced pressure to yield a white solid (5.0 g, 58%).

The above solid was suspended in dry CH₂Cl₂ (40 mL), to which Et₃N (16 mL, 116 mmol) was added. BzCl (10 mL, 87 mmol) was then added slowly at 0 °C. After stirring at room temperature overnight, the mixture was diluted with CH₂Cl₂ (100 mL). The mixture was washed with 1 N aq. HCl (3×30 mL), sat. aq. NaHCO₃ (3×30 mL), and brine (3×30 mL) successively. The organic layer was dried over MgSO₄ and concentrated. The crude product was suspended in pyridine (20 mL), to which another portion of BzCl (1.3 mL, 11.6 mmol) was added. After refluxing for 4 h, the reaction mixture was washed with 1 N aq. HCl (3×30 mL), saturated NaHCO₃ (3×30 mL). The mixture was washed with 1 N aq. HCl (3×30 mL), saturated NaHCO₃ (3×30 mL), and brine (3×30 mL) successively. The organic layer was dried over MgSO₄ and concentrated. The crude product was purified by column chromatography (petroleum ether/ethyl acetate = 2/1) to provide 1,3,4,6-tetra-*O*-benzoyl-2-deoxy-2-phthalimido-D-glucopyranose **S1** (6.9 g, 82%) as a white solid: ¹H NMR (500 MHz, CDCl₃) δ 8.06 (dd, *J* = 8.3, 1.3 Hz, 2H), 7.97 (dd, *J* = 8.4, 1.2 Hz, 2H), 7.91 (dd, *J* = 8.4, 1.2 Hz, 2H), 7.48–

7.39 (m, 3H), 7.41–7.36 (m, 2H), 7.36–7.31 (m, 2H), 7.28 (ddt, *J* = 8.6, 7.3, 1.4 Hz, 2H), 6.94 (d, *J* = 8.8 Hz, 1H), 6.48 (dd, *J* = 10.7, 9.2 Hz, 1H), 5.84 (t, *J* = 9.6 Hz, 1H), 4.95 (dd, *J* = 10.7, 8.8 Hz, 1H), 4.69–4.63 (m, 1H), 4.55–4.47 (m, 2H).

Compound **S1** (3.63 g, 5 mmol) was dissolved in dry CH₂Cl₂ (7.5 mL), and the mixture was cooled to 0 $^{\circ}$ C, to which 33% HBr in HOAc (3.25 mL) was added dropwise. After warmed to room temperature, the mixture was stirred until the disappearance of starting material (TLC, 5–7 h). The mixture was diluted with CH₂Cl₂ (100 mL) and washed with ice-water (3×30 mL), 0.5 M aq. KHCO₃ (3×20 mL), and brine (3×30 mL) successively. The organic layer was dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give **1b** as a white solid, which was pure enough and was used directly. It was found that **1b** was stable for at least one year when stored at -20 $^{\circ}$ C.

3,4,6-Tri-*O*-acetyl-2-deoxy-2-phthalimido-D-galactopyranosyl bromide (1c, $\beta/\alpha = 2:1$)

Compound **1c** was prepared as a pale yellow solid from known 1,3,4,6-tetra-*O*-acetyl-2-deoxy-2-phthalimido- α/β -D-galactopyranose⁴ (955 mg, 2 mmol) in quantitative yield, employing Method A.

1cβ: ¹H NMR (500 MHz, CDCl₃) δ 7.88 (m, 2H), 7.77 (m, 2H), 6.40 (d, J = 9.5 Hz, 1H), 5.77 (dd, J = 11.1, 3.4 Hz, 1H), 5.54 (d, J = 3.4 Hz, 1H), 4.86–4.82 (m, 1H), 4.23–4.18 (m, 1H), 2.24 (s, 3H), 2.08 (s, 3H), 1.86 (s, 3H). **1ca**: ¹H NMR (500 MHz, CDCl₃) δ 7.88 (m, 2H), 7.77 (m, 2H), 6.69 (d, J = 3.6 Hz, 1H), 6.53 (dd, J = 12.0, 3.1 Hz, 1H), 5.72 (dd, J = 3.2, 1.3 Hz, 1H), 4.90–4.78 (m, 1H), 4.60 (t, J = 6.7 Hz, 1H), 4.27–4.13 (m, 2H), 2.18 (s, 3H), 2.08 (s, 3H), 1.91 (s, 3H).

2,3,4-Tri-O-(4-trifluoromethylbenzoyl)-α-D-xylopyranosyl bromide (1d)

Compound **1d** was prepared as a white solid from 1,3,4,6-tetra-O-(4-trifluoromethylbenzoyl)- α/β -D-xylopyranose (838 mg, 1 mmol) in quantitative yield, employing Method A.

1d: ¹H NMR (500 MHz, CDCl₃) δ 8.11 (t, J = 8.4 Hz, 4H), 8.04 (d, J = 8.2 Hz, 2H), 7.69 (d, J = 8.2 Hz, 4H), 7.62 (d, J = 8.3 Hz, 2H), 6.80 (d, J = 4.0 Hz, 1H), 6.23 (t, J = 9.8 Hz, 1H), 5.55 (dt, J = 10.2, 5.2 Hz, 1H), 5.35 (dd, J = 9.9, 4.0 Hz, 1H), 4.37 (dd, J = 11.4, 5.9 Hz, 1H), 4.18 (t, J = 11.2 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 164.56, 164.36, 164.17, 130.58, 130.44, 130.30, 125.88-125.75 (m), 87.25, 71.65, 70.91, 69.15, 62.86; ¹⁹F NMR (376 MHz, CDCl₃) δ -63.34, -63.38, -63.40.

3,4,6-Tri-*O*-acetyl-2-deoxy-2-phthalimido-α-D-mannopyranosyl bromide (1e)

Compound **1e** was prepared as a white solid, which should be stored at -20 °C, from known 1,3,4,6-tetra-*O*-acetyl-2-deoxy-2-phthalimido- α/β -D-mannopyranose⁴ (2.0 g, 4.2 mmol) in quantitative yield, employing Method A.

1e: ¹H NMR (500 MHz, CDCl₃) δ 7.88 (dd, J = 5.5, 3.1 Hz, 2H), 7.78 (dd, J = 5.5, 3.0 Hz, 2H), 6.82 (d, J = 3.5 Hz, 1H), 5.68 (dd, J = 6.7, 5.3 Hz, 1H), 5.58 (dd, J = 9.3, 6.7 Hz, 1H), 5.23 (dd, J = 5.3, 3.6 Hz, 1H), 4.46–4.41 (m, 1H), 4.36–4.29 (m, 2H), 2.17 (s, 3H), 2.12 (s, 3H), 1.95 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 170.85, 169.88, 169.74,

167.52, 134.79, 131.27, 124.00, 84.45, 72.66, 68.66, 67.96, 62.22, 55.85, 20.92, 20.89, 20.77.

(PGO)n SEt $\xrightarrow{Br_2}$ (PGO)n PGO)n Br extra dry CH₂Cl₂, 4 Å MS, r.t. 0.5 h-1.0 h, in the dark

Supplementary Figure 4. General Method B for preparation of glycosyl bromides.

Method B. The glycosyl bromides were prepared from the corresponding ethyl thioglycosides via treatment with stoichiometric liquid bromine⁵. A mixture of the ethyl thioglycoside (1.0 equiv.) and activated molecular sieves (4 Å, 100 mg/mL CH₂Cl₂) in CH₂Cl₂ (0.1 M) was stirred under argon for 0.5 h. A freshly prepared solution of Br₂ (0.5–1.2 equiv.) in CH₂Cl₂ (1/165, v/v) was added. The reaction flask was wrapped with aluminum foil and was stirred for 30–60 min at room temperature in the dark. After dilution with CH₂Cl₂ and filtration, the mixture was washed with sat. aq. Na₂S₂O₃, sat. aq. NaHCO₃, and brine (2 times) successively. The organic phase was dried over Na₂SO₄, concentrated, and azeotroped with toluene for three times to afford the glycosyl bromide. Generally, the product was pure enough as determined by ¹H NMR analysis and was used directly for the subsequent coupling reaction. **Note**: i) glycosyl bromides are generally not stable on TLC plates; ii) some glycosyl bromides have similar mobility on TLC plates with the thioglycosides, so the progress of the reactions was monitored by ¹H NMR.

Ethyl 6-*O-tert*-butyldiphenylsilyl-3,4-di-*O*-benzoyl-2-deoxy-2-phthalimido-1-thioβ-D-glucopyranosyl bromide (1g)



Supplementary Figure 5. Preparation of GlcN bromide 1g.

To a stirred solution of the commercially available 1,3,4,6-tri-O-acetyl-2-deoxy-2-phthalimido-D-glucopyranoside (**S2**) (5.0 g, 10.5 mmol) in extra dry CH₂Cl₂ (30 mL), was added EtSH (1.16 mL, 40 mmol). After which, BF₃ OEt₂ (5.08 ml, 40 mmol) was added slowly at 0 °C, and the mixture was stirred overnight. The reaction mixture was diluted with CH₂Cl₂ (100 mL), washed with water (3×50 mL), aq. NaHCO₃ (3×50 mL),

and brine $(3 \times 30 \text{ mL})$ successively. The organic phase was dried over MgSO₄. After concentration, the crude product was used directly in the next step.

To a stirred solution of the crude product in mixed solvents ($CH_2Cl_2/MeOH$, 1:10, 22 mL) was added MeONa (120 mg, 2.2 mmol). The mixture was stirred for 3 h. After completion, Amberlite[®] resin IR-120 (H⁺ form) was added to neutralize the solution. The resin was filtered off, and the filtrate was concentrated and dried over oil pump to give the crude product as a solid.

To a stirred solution of one half of the obtained crude product in dry DMF (10 mL) were added imidazole (680 mg, 10 mmol) and TBDPSCl (1.57 mL, 12 mmol). After stirring at rt for 24 h, the solution was diluted with ethyl acetate (100 mL), washed with water (3×30 mL), and brine (3×30 mL). After concentration, the crude product was redissolved in dry CH2Cl2 (15 mL), and Et3N (4.2 mL, 30 mmol) was added. BzCl (1.7 mL, 15 mmol) was added dropwise at 0 °C and the resulting mixture was stirred overnight at room temperature. The reaction mixture was diluted with CH₂Cl₂ (50 mL), washed with water (3×30 mL) and brine (3×30 mL), and was then concentrated to dryness. The residue was purified by column chromatography (petroleum ether/ethyl acetate = 5/1, v/v) to provide ethyl 6-O-tert-butyldiphenylsilyl-3,4-di-O-benzoyl-2deoxy-2-phthalimido-1-thio- β -D-glucopyranoside S3 (1.65 g, 42%) as a white solid: ¹H NMR (500 MHz, CDCl₃) δ 7.87 (dd, J = 13.8, 7.4 Hz, 3H), 7.77 (d, J = 7.4 Hz, 2H), 7.75–7.68 (m, 4H), 7.69–7.64 (m, 1H), 7.59 (d, J = 6.9 Hz, 2H), 7.51 (t, J = 7.4 Hz, 1H), 7.42 (t, J = 7.4 Hz, 1H), 7.38–7.32 (m, 6H), 7.28–7.25 (m, 2H), 7.20 (t, J = 7.5 Hz, 2H), 6.26 (t, J = 9.8 Hz, 1H), 5.69 (t, J = 9.7 Hz, 1H), 5.64 (d, J = 10.5 Hz, 1H), 4.65 (t, J = 10.4 Hz, 1H), 3.99 (dt, J = 10.1, 3.5 Hz, 1H), 3.86 (d, J = 3.5 Hz, 1H), 2.88– 2.76 (m, 1H), 2.76–2.63 (m, 1H), 1.28 (t, *J* = 7.4 Hz, 3H), 1.04 (s, 9H); ¹³C NMR (125) MHz, CDCl₃) δ 165.94, 165.13, 135.82, 135.67, 133.31, 133.19, 133.10, 129.95, 129.91, 129.76, 129.71, 129.37, 128.88, 128.46, 128.41, 127.76, 127.72, 123.85, 123.76, 80.92, 79.37, 72.55, 69.74, 63.05, 54.24, 26.75, 24.05, 19.30, 15.24; HRMS (ESI) calcd. for $C_{46}H_{45}NO_8Na (M + Na)^+ m/z 822.2527$, found 822.2526.

Compound **S3** (0.5 mmol) and activated molecular sieves (4Å, 800 mg) were suspended in extra dry CH₂Cl₂ (8 mL), and the mixture was stirred under argon for 0.5 h. To another round bottom flask wrapped with aluminum foil was charged with activated molecular sieves (4Å, 200 mg) and CH₂Cl₂ (2.1 mL), to which Br₂ (32 μ L, 0.25 mmol) was injected under stirring. After stirring for ~20 mins in the dark, the mixture was transferred into the first bottle via syringe. The resulting mixture was stirred for 30 mins in the dark. After dilution with CH₂Cl₂ (20 mL), the molecular sieves were filtered off and washed with CH₂Cl₂. The organic phase was washed with sat. aq. Na₂S₂O₃, sat. aq. NaHCO₃, and brine successively. After dried over Na₂SO₄ and concentrated under vaccum, the crude product was azeotroped with toluene for three times to give **1g**, which was pure enough, as determined by ¹H NMR analysis, and was used directly. **1g**: ¹H NMR (500 MHz, CDCl₃) δ 7.91–7.82 (m, 4H), 7.79–7.70 (m, 6H), 7.60 (d, *J* = 6.8 Hz, 2H), 7.52 (t, *J* = 7.4 Hz, 1H), 7.43 (t, *J* = 7.4 Hz, 1H), 7.36–7.31 (m, 5H), 7.31 (d, *J* = 7.5 Hz, 1H), 7.28 (d, *J* = 7.7 Hz, 2H), 7.22 (t, *J* = 7.5 Hz, 2H), 6.54 (d, *J* = 9.5 Hz, 1H), 6.18 (dd, *J* = 10.4, 9.3 Hz, 1H), 5.80 (t, *J* = 9.7 Hz, 1H), 4.86

(dd, J = 10.4, 9.5 Hz, 1H), 4.02 (ddd, J = 10.1, 4.5, 2.2 Hz, 1H), 3.88 (qd, J = 11.9, 3.4 Hz, 2H), 1.06 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 165.79, 164.94, 135.87, 135.75, 134.52, 133.47, 133.05, 133.03, 129.97, 129.94, 129.84, 129.78, 129.16, 128.59, 128.53, 128.47, 127.82, 127.75, 123.95, 80.04, 77.88, 71.62, 69.00, 62.70, 58.83, 26.81, 19.37.

4-*O*-Benzyl-**3**,**6**-di-*O*-benzoyl-**2**-deoxy-**2**-phthalimido-**1**-thio-β-D-glucopyranosyl bromide (**1**h)



Supplementary Figure 6. Preparation of GlcN bromide 1h.

To a solution of **S4**⁶ (0.82 g, 1.5 mmol) and BH₃ Et₃N (2.19 mL, 15 mmol) in CH₂Cl₂ (25 mL) and Et₂O (5 mL) was added AlCl₃ (0.4 g, 3.0 mmol) at 0 °C. After stirring for 30 min at room temperature, sat. aq. NaHCO₃ was added slowly. The mixture was filtered through a celite pad and the aqueous layer was extracted with CH₂Cl₂. The combined organic layer was washed with water and brine, and was dried over Na₂SO₄. After filtration and removal of the solvent, the residue was purified by silica gel column chromatography (petroleum/ethyl acetate = 4/1 to 2/1) to afford ethyl 4-*O*-benzyl-3-*O*-benzoyl-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside **S5** (0.72 g, 87%) as a white foam.

To a stirred solution of S5 in dry CH₂Cl₂ (4 mL) was added Et₃N (1.0 mL, 6.0 equiv.). BzCl (150 µL, 3.0 equiv.) was added dropwise at 0 °C. After stirring at room temperature for 4 h, the mixture was diluted with CH₂Cl₂ (20 mL), and was washed with water and brine. The organic layer was dried over Mg₂SO₄. After filtration and removal of the solvent, the residue was purified by silica gel column chromatography afford ethyl 4-O-benzyl-3,6-di-O-benzoyl-2-deoxy-2-phthalimido-1-thio-β-Dto glucopyranoside S6 (754 mg, 89%) as a white solid: ¹H NMR (500 MHz, CDCl₃) δ 8.09 (d, J = 7.2 Hz, 2H), 7.88 (d, J = 7.8 Hz, 3H), 7.73–7.63 (m, 3H), 7.61 (t, J = 7.4 Hz, 1H), 7.50 (q, J = 7.5 Hz, 3H), 7.36 (t, J = 7.7 Hz, 2H), 7.16–7.03 (m, 5H), 6.21 (dd, *J* = 10.4, 8.7 Hz, 1H), 5.61 (d, *J* = 10.5 Hz, 1H), 4.68 (dd, *J* = 12.0, 2.1 Hz, 1H), 4.61– 4.43 (m, 4H), 4.03 (ddd, J = 9.8, 4.7, 2.1 Hz, 1H), 3.99–3.91 (m, 1H), 2.78–2.54 (m, 1H), 1.20 (t, J = 7.4 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 166.31, 165.69, 136.93, 134.39, 134.19, 133.46, 133.31, 131.32, 129.99, 129.89, 129.85, 129.22, 128.58, 128.50, 128.27, 128.13, 123.77, 81.33, 77.41, 76.70, 74.92, 74.50, 63.66, 54.31, 24.65, 15.17; HRMS (ESI) calcd. for C₃₇H₃₃NO₈SNa (M + Na)⁺ m/z 674.1819, found 674.1825.

Compound **S6** (335 mg, 0.51 mmol) and activated molecular sieves (4 Å, 800 mg) were suspended in extra dry CH₂Cl₂ (8 mL), the mixture was stirred under argon for 0.5 h. Another round bottom flask wrapped with aluminum foil was charged with activated molecular sieves (4 Å, 300 mg) and CH₂Cl₂ (3.0 mL), to which Br₂ (32 µL, 0.25 mmol) was injected under stirring. After stirring for ~20 mins in the dark, the mixture was transferred into the first bottle via syringe. The resulting mixture was stirred for 30 mins in the dark. After dilution with CH₂Cl₂ (20 mL), the molecular sieves were filtered off and washed with CH₂Cl₂. The organic phase was washed with sat. aq. Na₂S₂O₃, sat. aq. NaHCO₃, and brine successively, and was then dried over Na₂SO₄. After concentration, the crude product was azeotroped with toluene for three times to give crude **1h**. ¹H NMR analysis showed a mixture containing **1h** (75%) together with the corresponding glycal (25%). The glycosyl bromide **1h** was not stable and was used directly in the next step. **1h**: ¹H NMR (500 MHz, CDCl₃) δ 8.11 (d, J = 7.3 Hz, 2H), 7.95–7.78 (m, 4H), 7.70 (s, 2H), 7.63 (t, J = 7.3 Hz, 1H), 7.55–7.51 (m, 3H), 7.40–7.33 (m, 3H), 7.13–7.03 (m, 4H), 6.53 (d, *J* = 9.4 Hz, 1H), 6.14 (dd, *J* = 10.5, 7.8 Hz, 1H), 4.79–4.72 (m, 1H), 4.68 (d, J = 10.8 Hz, 1H), 4.64–4.59 (m, 1H), 4.58–4.51 (m, 2H), 4.07–4.03 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 166.24, 165.53, 134.51, 133.63, 133.44, 130.01, 129.87, 128.64, 128.55, 128.41, 128.30, 123.94, 78.31, 77.94, 75.88, 75.04, 73.53, 63.02, 58.75.

$\label{eq:2.1} \begin{array}{l} \textbf{4,6-Di-$O-benzoyl-2-deoxy-2-phthalimido-3-$O-(2,3,4,6-tetra-$O-acetyl-$\beta-D-galactopyranosyl}) \\ \textbf{-} \textbf{\beta-} \textbf{D-} \textbf{glucopyranosyl} \ bromide \ (1j) \end{array}$

Compound 1j was prepared employing a modified literature procedure^{7,8}.



Supplementary Figure 7. Preparation of disaccharide bromide **1j**. Reagents and conditions: a) DTBMP (1.0 equiv.), AgOTf (1.65 equiv.), CH₂Cl₂, 4 Å MS, 10 °C, 1 h, then 25 °C, 7.5 h, 96%; b) TMSOTf (5 mol%), CH₂Cl₂, 4Å MS, 18 h, 73%; c) aq. HOAc (HOAc/H₂O = 3:2), 60 °C, 0.5 h; d) BzCl (6.0 equiv.), DMAP (cat.), pyridine, 4 h, 60 °C, 87% for 2 steps; e) Br₂ (1.0 equiv.), CH₂Cl₂, 4 Å MS, 25 °C, in the dark, 0.5 h; this step was repeated twice to give **1j** quantitatively.

To a mixture of **S7**^{7,8} (882 mg, 2.0 mmol) and **S8** (1.233 g, 3.0 mmol) in dry CH₂Cl₂ (25 mL) were added activated molecular sieves (4 Å, 2.5 g) and DTBMP (2,6-di-tertbutyl-4-methylpyridine) (0.42 g, 2.0 mmol). The mixture was stirred at -30 °C for 30 mins, to which a solution of silver triflate (848 mg, 3.3 mmol) in dry toluene (8 mL) was added under argon. The mixture was stirred at 10 $\,^{\circ}$ C for 1 h, after which the mixture was warmed to room temperature and stirred for 7.5 h. TLC (petroleum ether/ethyl acetate = 2:1, $R_f = 0.3$) showed the complete consumption of the acceptor. The reaction was quenched with Et₃N (2.0 mL). After dilution with CH₂Cl₂ and filtration, the filtrate was successively washed with water and brine, and was dried over Na₂SO₄. After concentration, the residue was purified by column chromatography (petroleum ether/ethyl acetate = 2:1) to give **S9** (1.49 g, 96%) as a white foam: ¹H NMR (500 MHz, CDCl₃) *δ* 7.95–7.87 (m, 2H), 7.53–7.45 (m, 2H), 7.79–7.72 (m, 2H), 7.53–7.45 (m, 3H), 7.37–7.32 (m, 5H), 5.59 (d, J = 4.7 Hz, 1H), 5.57 (s, 1H), 5.39 (d, J = 10.7 Hz, 1H), 5.21 (t, J = 2.9 Hz, 1H), 4.72–4.65 (m, 2H), 4.41 (dd, J = 10.4, 4.5 Hz, 1H), 4.30 (dd, J = 10.7, 9.6 Hz, 1H), 4.09 (td, J = 6.6, 2.5 Hz, 1H), 4.02 (dd, J = 11.3, 6.7 Hz, 1H), 3.96 (dd, J = 11.3, 6.5 Hz, 1H), 3.81 (t, J = 9.9 Hz, 1H), 3.75–3.67 (m, 3H), 2.77–2.61 (m, 2H), 2.02 (s, 3H), 1.99 (s, 3H), 1.86 (s, 3H), 1.53 (s, 3H), 1.20 (t, J = 7.4 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 170.48, 169.79, 169.64, 168.66, 167.54, 137.25, 134.32, 134.20, 132.17, 131.68, 129.06, 128.30, 126.12, 123.73, 123.41, 121.79, 101.50, 97.55, 81.97, 80.56, 73.77, 71.28, 70.91, 70.88, 68.97, 68.73, 65.66, 61.19, 55.05, 24.84, 24.26, 20.81, 20.77, 20.60, 14.99.

A suspension of **S9** (1.49 g, 1.93 mmol) and molecular sieves (4 Å, 2.0 g) in extra dry CH₂Cl₂ (20 mL) was stirred at room temperature for 30 min, to which TMSOTf (~19 µL, 5 mol%) was added. The mixture was stirred for 18 h and was then diluted with CH₂Cl₂ (30 mL). After filtration and concentration, the crude product was purified by column chromatography (petroleum ether/ethyl acetate = 2:1 or petroleum ether/dichloromethane/ethyl acetate = 2:1:0.5) to give ethyl 4,6-O-benzylidene-2deoxy-2-phthalimido-3-O-(2,3,4,6-tetra-O-acetyl-\beta-D-galactopyranosyl)-1-thio-B-Dglucopyranoside S10 (1.07 g, 73%) as a white foam: ¹H NMR (500 MHz, CDCl₃) δ 7.92–7.81 (m, 2H), 7.75 (q, J = 3.5 Hz, 2H), 7.49–7.42 (m, 2H), 7.34 (dd, J = 5.1, 2.0Hz, 3H), 5.55 (s, 1H), 5.26 (d, J = 10.7 Hz, 1H), 5.17 (dd, J = 3.5, 1.1 Hz, 1H), 4.97 (dd, J = 10.4, 8.0 Hz, 1H), 4.75 (dd, J = 9.2, 8.1 Hz, 1H), 4.71 (dd, J = 9.6, 2.7 Hz, 1H),4.52 (d, J = 8.0 Hz, 1H), 4.41–4.30 (m, 2H), 4.01 (dd, J = 11.0, 8.1 Hz, 1H), 3.81 (d, J = 5.3 Hz, 1H), 3.79 (d, J = 3.0 Hz, 1H), 3.80–3.75 (m, 1H), 3.67 (td, J = 9.7, 4.8 Hz, 1H), 3.47 (ddd, J = 8.2, 5.5, 1.2 Hz, 1H), 2.73–2.56 (m, 2H), 2.03 (s, 3H), 1.88 (s, 3H), 1.81 (s, 3H), 1.53 (s, 3H), 1.13 (t, J = 7.4 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 170.22, 170.01, 170.00, 168.86, 168.35, 167.01, 137.00, 134.55, 134.19, 131.68, 131.52, 129.27, 128.34, 126.02, 123.97, 123.22, 101.40, 100.39, 81.69, 80.88, 77.36, 76.41, 70.95, 70.62, 70.28, 69.15, 68.63, 66.61, 60.78, 54.16, 23.86, 20.60, 20.55, 20.43, 20.10, 14.80.

A solution of **S10** (414.3 mg, 0.55 mmol) in acetic acid/water (7.5 mL, 3:2) was stirred at 60 $^{\circ}$ C for 0.5 h. After warming to room temperature, the solvents were

removed *in vacuo*. The residue diol was azeotroped with toluene for three times and was used directly in the next step.

To a solution of the diol in pyridine (1.0 mL) containing DMAP (10 mol%) was slowly added benzyl chloride (380 µL, 3.3 mmol) at room temperature. The mixture was heated to 65 °C and stirred for 3 h. After removal of solvent, the residue was purified by column chromatography (petroleum ether/ethyl acetate = 1:1) to afford ethvl 4,6-di-O-benzoyl-2-deoxy-2-phthalimido-3-O-(2,3,4,6-tetra-O-acetyl-β-Dgalactopyranosyl)-1-thio- β -D-glucopyranoside S11 (426 mg, 87%) as a white foam: ¹H NMR (500 MHz, CDCl₃) δ 8.03 (d, J = 8.3 Hz, 2H), 7.99 (d, J = 8.2 Hz, 2H), 7.87 (d, J = 17.5 Hz, 2H), 7.79 (dd, J = 5.3, 2.5 Hz, 2H), 7.53 (dt, J = 15.0, 7.4 Hz, 2H), 7.39 (dt, J = 15.8, 7.6 Hz, 4H), 5.42 (t, J = 9.5 Hz, 1H), 5.29 (d, J = 10.5 Hz, 1H), 5.00 (s, 1H), 4. 93-4.86 (dt, J = 18.0, 9.7 Hz, 2H), 4.60 (d, J = 12.0 Hz, 1H), 4.54 (d, J = 10.4 Hz, 1H), 4.47 (t, J = 10.4 Hz, 1H), 4.41 (dd, J = 12.2, 5.4 Hz, 2H), 4.20 (d, J = 7.8 Hz, 1H), 4.17–4.13 (m, 1H), 3.55–3.41 (m, 2H), 3.32 (ddd, *J* = 10.1, 7.1, 1.4 Hz, 1H), 2.74– 2.64 (m, 1H), 2.63–2.54 (m, 1H), 1.95 (s, 3H), 1.85 (s, 3H), 1.82 (s, 3H), 1.80 (s, 3H), 1.15 (t, J = 7.4 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 170.17, 170.08, 170.05, 169.13, 168.77, 167.17, 166.23, 164.64, 134.85, 134.71, 133.36, 133.12, 131.50, 131.29, 129.87, 129.84, 129.78, 129.69, 128.37, 123.83, 100.30, 81.32, 77.36, 76.21, 76.03, 70.94, 70.33, 69.02, 66.30, 63.66, 60.23, 54.72, 24.10, 20.68, 20.48, 20.44, 20.41, 14.97.

Compound S11 (134 mg, 0.15 mmol) was dissolved in extra dry CH₂Cl₂ (3.0 mL), and activated molecular sieves (4 Å, 300 mg) was added. The mixture was stirred under argon for 0.5 h. Another round bottom flask wrapped with aluminum foil was charged with activated molecular sieves (4 Å, 150 mg) and CH₂Cl₂ (1.3 mL), to which Br₂ (7.7 µL, 0.15 mmol) was injected under stirring. After stirring for ~20 mins in the dark, the mixture was transferred into the first bottle via syringe. The resulting mixture was stirred for 30 mins in the dark. After dilution with CH₂Cl₂ (20 mL), the molecular sieves were filtered off and washed with CH₂Cl₂. The organic phase was washed with sat. aq. Na₂S₂O₃, sat. aq. NaHCO₃, and brine successively, and was then dried over Na₂SO₄. After concentration, the crude product was azeotroped with toluene for three times to give crude 1j. ¹H NMR analysis showed that $\sim 20\%$ of the starting material was still left in the crude product. Thus, this procedure was repeated once again. ¹H NMR analysis showed completion of the reaction, and 1j was pure enough and was used directly. 1j: ¹H NMR (500 MHz, CDCl₃) δ 8.05 (t, J = 6.6 Hz, 4H), 7.92 (dd, J = 5.4, 3.0 Hz, 2H), 7.84 (dd, J = 5.5, 3.0 Hz, 2H), 7.60–7.53 (m, 2H), 7.43 (q, J = 8.0 Hz, 4H), 6.27 (d, J =9.6 Hz, 1H), 5.54 (t, J = 9.6 Hz, 1H), 5.02 (d, J = 3.5 Hz, 1H), 4.93–4.86 (m, 2H), 4.73 (dd, J = 10.4, 9.6 Hz, 1H), 4.65 (dd, J = 12.3, 2.7 Hz, 1H), 4.57 (dd, J = 10.4, 3.5 Hz)1H), 4.45 (dd, *J* = 12.4, 5.2 Hz, 1H), 4.26 (ddd, *J* = 9.9, 5.1, 2.7 Hz, 1H), 4.20 (d, *J* = 7.8 Hz, 1H), 3.55–3.46 (m, 2H), 3.38–3.30 (m, 1H), 1.98 (s, 3H), 1.90 (s, 3H), 1.83 (s, 3H), 1.82 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 170.11, 170.00, 169.08, 166.16, 164.46, 134.99, 133.47, 133.22, 131.15, 129.86, 129.57, 129.47, 128.40, 128.39, 123.99, 100.23, 78.00, 77.36, 77.06, 75.14, 70.83, 70.40, 69.65, 68.90, 66.23, 63.08, 60.17, 58.99, 20.64, 20.45, 20.42, 20.36.

3,6-Di-*O*-benzoyl-2-deoxy-2-phthalimido-4-*O*-(**3,4,6-tri**-*O*-acetyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)-β-D-glucopyranosyl bromide (1k)

Compound **1k** was prepared employing a modified literature procedure⁹.



Supplementary Figure 8. Preparation of disaccharide bromide **1k**. Reagents and conditions: a) MeONa, MeOH/CH₂Cl₂ (1:1), 25 °C, 4 h; b) PhCH(OMe)₂, *p*-TSA (10 mol%), DMF, 50 °C, 5 h; c) BzCl (5.0 equiv.), pyridine, 40 °C, 4 h; d) aq. HOAc (HOAc/H₂O = 4:1, v/v), 80 °C, 1.5 h, 67% for 4 steps; e) BzCl (1.1 equiv.), pyridine/CH₂Cl₂ (1:3), 0 °C \rightarrow 25 °C, 12 h; then another portion of BzCl (1.1 equiv.) was added, 25 °C, 5 h, 75%; f) TMSOTf (20 mol%), CH₂Cl₂, 4 Å MS, -40 °C \rightarrow 0 °C, 6 h, 60%; g) CAN (5.0 equiv.), CH₃CN/H₂O (4:1), 25 °C, 1h; h) AcCl (5.0 equiv.), pyridine, 0 °C \rightarrow 25 °C, 0.5 h, 75% for 2 steps; i) HBr in HOAc, CH₂Cl₂, 0 °C \rightarrow 25 °C, 3 h, quant.

To a stirred solution of **S12** (2.7 g, 5 mmol) in MeOH/CH₂Cl₂ (1:1, v/v, 20 mL) was added MeONa (108 mg, 40 mol%). After stirring at room temperature for 4 h, the mixture was neutralized with Amberlite[®] resign IR-120 (H⁺ form). The mixture was filtrated, concentrated, and dried over oil pump. The crude product was redissolved in DMF (10 mL). Benzaldehyde dimethyl acetal (2.3 mL, 15 mmol) was added and the reaction was kept rotating over vacuo evaporator at 50 °C for 5 h. After cooled to room temperature, the mixture was neutralized with Et₃N and concentrated. The residue was azeotroped with toluene for one time, and the crude *p*-methoxyphenyl 4,6-*O*-benzylidene-2-deoxy-2-phthalimido- β -D-glucopyranoside was dissolved in pyridine (10 mL). DMAP (10 mol%) was added, and benzyl chloride (1.7 mL, 15 mmol) was then added slowly at 0 °C. The mixture was removed *in vacuo*. The residue was dissolved in CH₂Cl₂ (50 mL), washed with 1 N aq. HCl (2×10 mL) and brine (2×10 mL). The organic phase was dried over MgSO₄ and concentrated to afford crude **S13**.

The crude **S13** was dissolved in acetic acid/water (4:1, v/v, 50 mL) and stirred at 80 $^{\circ}$ C for 1.5 h. The mixture was cooled to room temperature and neutralized carefully with sat. aq. NaHCO₃ at 0 $^{\circ}$ C. The mixture was extracted with ethyl acetate, dried over MgSO₄, and concentrated. The crude product was purified by column chromatography (petroleum ether/ethyl acetate = 2:1 then CH₂Cl₂/MeOH = 10:1) to afford *p*-

methoxyphenyl 3-*O*-benzoyl-2-deoxy-2-phthalimido- β -D-glucopyranoside (1.95 g, 67% over 4 steps).

To a stirred solution of *p*-methoxyphenyl 3-*O*-benzoyl-2-deoxy-2-phthalimido-β-D-glucopyranoside (1.04 g, 2.0 mmol) in pyridine/DCM (1:3, v/v, 28 mL) was added BzCl (253 µL, 1.0 equiv.) in CH₂Cl₂ (0.5 mL) dropwise under 0 °C. After 12 h, about 30% of the starting material was converted, and another portion of BzCl (253 µL, 1.0 equiv.) in CH₂Cl₂ (0.5 mL) was added. After 12 h, the mixture was concentrated and purified by column chromatography to give *p*-methoxyphenyl 3,6-di-*O*-benzoyl-2-deoxy-2-phthalimido-β-D-glucopyranoside **S14** (940 mg, 75%): ¹H NMR (500 MHz, CDCl₃) δ 8.09 (d, *J* = 7.1 Hz, 2H), 7.90 (d, *J* = 7.2 Hz, 2H), 7.80 (dd, *J* = 5.3, 3.2 Hz, 2H), 7.65 (dd, *J* = 5.5, 3.0 Hz, 2H), 7.60 (t, *J* = 7.4 Hz, 1H), 7.53–7.49 (m, 1H), 7.47 (t, *J* = 7.8 Hz, 1H), 7.35 (t, *J* = 7.8 Hz, 2H), 6.89 (d, *J* = 9.1 Hz, 2H), 6.66 (d, *J* = 9.1 Hz, 2H), 6.01 (dd, *J* = 10.7, 8.7 Hz, 1H), 5.97 (d, *J* = 8.5 Hz, 1H), 4.78–4.71 (m, 3H), 4.09 (ddd, *J* = 9.8, 5.4, 2.9 Hz, 1H), 3.96 (td, *J* = 9.3, 3.9 Hz, 1H), 3.69 (s, 3H), 3.59 (d, *J* = 4.9 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 167.26, 166.89, 155.72, 150.70, 134.43, 133.74, 133.41, 131.45, 130.08, 130.03, 129.84, 128.79, 128.61, 128.56, 123.79, 118.97, 114.51, 97.63, 74.62, 74.50, 70.72, 63.83, 55.68, 54.51.

To a stirred solution of 3,4,6-tri-O-acetyl-2-deoxy-2-phthalimido- β -D-glucopyranoside (1.5 mmol) in dry CH₂Cl₂ (30 mL) were added CCl₃CN (1.5 mL, 15 mmol) and K₂CO₃ (4.5 mmol). The mixture was stirred for 3 h, and was then filtrated. After concentration and azeotroped with toluene, the crude 3,4,6-tri-O-acetyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl trichloroacetimidate **S15** was obtained and was used directly.

A suspension of S14 (312 mg, 0.5 mmol), S15, and activated molecular sieves (4 Å, 1 g) in extra dry CH₂Cl₂ (10 mL) was stirred at -40 $^{\circ}$ C for 0.5 h. TMSOTf (18 μ L, 20 mol%) was added, and the resulting mixture was warmed to 0 °C slowly and stirred for 6 h. After quenching with triethylamine, the reaction mixture was concentrated. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 2:1) to give p-methoxyphenyl 3,6-di-O-benzoyl-2-deoxy-2-phthalimido-4-O-(3,4,6-tri-*O*-acetyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)-β-D-glucopyranoside **S16** (311 mg, 60%) as a white foam: ¹H NMR (500 MHz, CDCl₃) δ 7.96 (d, J = 7.7 Hz, 2H), 7.86 (d, J = 7.7 Hz, 2H), 7.75–7.63(br, 5H), 7.61–7.51 (m, 5H), 7.47–7.42 (m, 4H), 6.76 (d, J = 8.6 Hz, 2H), 6.59 (d, J = 8.7 Hz, 2H), 6.18 (dd, J = 10.4, 8.6 Hz, 1H), 5.87 (d, J = 8.5 Hz, 1H), 5.66 (d, J = 8.3 Hz, 1H), 5.58 (t, J = 9.9 Hz, 1H), 4.99 (t, J = 9.6 Hz)Hz, 1H), 4.59 (dd, J = 10.7, 8.3 Hz, 2H), 4.26 (q, J = 8.5 Hz, 2H), 4.07 (dd, J = 11.8, 4.9 Hz, 1H), 4.04–4.01 (m, 1H), 3.82 (dd, J = 12.2, 3.8 Hz, 1H), 3.65 (s, 1H), 3.55 (dd, J = 12.3, 2.2 Hz, 1H), 3.28 (dt, J = 10.2, 2.9 Hz, 1H), 2.00 (s, 3H), 1.88 (s, 3H), 1.77 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 170.50, 170.02, 169.17, 165.30, 164.99, 155.62, 150.37, 134.24, 134.19, 133.61, 133.08, 131.35, 130.85, 129.70, 129.66, 129.48, 128.89, 128.70, 128.29, 123.65, 123.57, 118.99, 114.31, 97.56, 97.50, 76.36, 72.53, 72.13, 71.77, 70.39, 67.97, 62.45, 60.94, 55.48, 54.91, 54.88, 20.69, 20.46, 20.29.

To a stirred solution of **S16** (311.6 mg, 0.3 mmol) in CH₃CN/H₂O (4:1, v/v, 3 mL), ammonium cerium (IV) nitrate (1.5 mmol) was added. After stirring at room temperature for 1 h, the mixture was diluted with EtOAc (100 mL) and washed with water (3×30 mL) and brine (3×30 mL). The organic layer was dried over Na₂SO₄ and concentrated *in vacuo* to give the corresponding lactol.

To a stirred solution of the above crude product in pyridine (3.0 mL) was added Ac₂O (106 µL, 5.0 equiv.) in CH₂Cl₂ (0.9 mL) dropwise. The reaction was stirred at room temperature for 0.5 h. After evaporation of the solvent, the crude product was purified by column chromatography (petroleum ether/ethyl acetate = 2:1) to afford 3,6di-O-benzoyl-2-deoxy-2-phthalimido-4-O-(3,4,6-tri-O-acetyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)- β -D-glucopyranosyl acetate **S17** (977 mg, 75% over two steps): ¹H NMR (500 MHz, CDCl₃) δ 7.94 (d, J = 7.4 Hz, 2H), 7.87 (d, J = 7.6 Hz, 2H), 7.82– 7.62 (m, 4H), 7.63–7.55 (m, 4H), 7.47 (t, J = 7.7 Hz, 3H), 7.44 (t, J = 7.7 Hz, 3H), 6.56 (d, J = 8.9 Hz, 1H), 6.20 (dd, J = 10.4, 8.6 Hz, 1H), 5.66 (d, J = 8.4 Hz, 1H), 5.56 (dd, J = 10.4, 8.6 Hz, 1H), 5.66 (dd, J = 10.4, 8.6 Hz, 100 Hz), 5.66 (dd, J = 10.4, 8.6 Hz, 100 Hz), 5.66 (dd, J = 10.4, 8.6 Hz, 100 Hz), 5.66 (dd, J = 10.4, 8.6 Hz), 5.66 (dd, J =J = 10.7, 9.2 Hz, 1H), 4.98 (t, J = 9.6 Hz, 1H), 4.66–4.57 (m, 1H), 4.49 (dd, J = 10.4, 8.9 Hz, 1H), 4.32–4.27 (m, 2H), 4.25 (dd, J = 10.7, 8.5 Hz, 1H), 4.10–4.01 (m, 2H), 3.83 (dd, *J* = 12.3, 3.9 Hz, 1H), 3.53 (dd, *J* = 12.2, 2.2 Hz, 1H), 3.25 (ddd, *J* = 10.2, 3.9, 2.2 Hz, 1H), 2.02 (s, 3H), 1.91 (s, 3H), 1.87 (s, 3H), 1.76 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) § 170.59, 170.07, 169.21, 168.53, 167.56, 165.32, 164.97, 134.37, 134.26, 133.74, 133.18, 129.79, 129.70, 129.48, 128.87, 128.79, 128.38, 123.65, 97.55, 89.67, 75.69, 73.04, 72.07, 71.81, 70.40, 68.02, 62.22, 61.00, 54.91, 53.96, 20.76, 20.52, 20.35.

To a stirred solution of S17 (977 mg, 0.2 mmol) in dry CH₂Cl₂ (4.0 mL), 33% HBr in HOAc (0.25 mL) was added dropwise at 0 °C. After stirring for 2 h, the mixture was diluted with CH₂Cl₂ (50 mL), and washed with ice-water (3×20 mL), sat. aq. NaHCO₃ (3×10 mL), and brine (3×20 mL) successively. The organic layer was dried over Na₂SO₄, filtered, and concentrated to give **1k** as a white solid, which was pure enough and was used directly. 1k: ¹H NMR (500 MHz, CDCl₃) δ 7.91 (d, J = 7.5 Hz, 2H), 7.83 (d, J = 7.5 Hz, 2H), 7.78–7.54 (m, 10H), 7.45 (t, J = 7.8 Hz, 2H), 7.41 (t, J = 7.6 Hz, 2H), 6.43 (d, J = 9.5 Hz, 1H), 6.08 (dd, J = 10.2, 8.6 Hz, 1H), 5.63 (d, J = 8.4 Hz, 1H), 5.53 (dd, J = 10.6, 9.2 Hz, 1H), 4.95 (t, J = 9.6 Hz, 1H), 4.63 (t, J = 8.8 Hz, 2H), 4.60 (d, J = 8.7 Hz, 1H), 4.30 (t, J = 9.3 Hz, 1H), 4.22 (dd, J = 10.7, 8.4 Hz, 1H), 4.03 (dd, *J* = 12.2, 3.5 Hz, 1H), 4.01–3.95 (m, 1H), 3.79 (dd, *J* = 12.3, 3.7 Hz, 1H), 3.50 (dd, *J* = 12.3, 2.2 Hz, 1H), 3.21 (ddd, J = 10.1, 3.7, 2.2 Hz, 1H), 1.98 (s, 3H), 1.84 (s, 3H), 1.73 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 170.53, 170.05, 169.20, 165.28, 164.88, 134.45, 133.83, 133.26, 131.24, 130.83, 129.80, 129.70, 129.31, 128.83, 128.73, 128.39, 123.81, 123.67, 97.66, 77.57, 77.36, 77.26, 75.54, 72.16, 71.83, 70.38, 67.92, 62.21, 60.91, 58.60, 54.91, 20.76, 20.51, 20.34.

3,4-Di-*O*-benzoyl-2-*O*-(2,3,4-tri-*O*-benzoyl-α-L-rhamnopyranosyl)-α-L-rhamnopyranosyl bromide (11)



Supplementary Figure 9. Preparation of disaccharide bromide 11.

To a solution of $S18^{10}$ (130.6 mg, 0.15 mmol) in MeOH (1.5 mL) was added PdCl₂ (1.4 mg, 5 mol%). After stirring at room temperature for 12 h, the mixture was diluted with CH₂Cl₂, filtered, and concentrated to dryness. The crude product was dissolved in pyridine (2.0 mL), and Ac₂O (71 µL, 5.0 equiv.) was added under 0 °C. After stirring at room temperature for 1.0 h, the solvent was removed, and the product was purified by column chromatography to afford 3,4-di-O-benzoyl-2-O-(2,3,4-tri-O-benzoyl-α-Lrhamnopyranosyl)- α -L-rhamnopyranosyl acetate **S19** (116.5 mg, 89% yield over two steps): ¹H NMR (500 MHz, CDCl₃) δ 8.04–7.96 (m, 8H), 7.84 (d, J = 7.2 Hz, 2H), 7.58 (t, J = 7.4 Hz, 1H), 7.55-7.49 (m, 2H), 7.47-7.37 (m, 8H), 7.33 (t, J = 7.6 Hz, 3H),7.29–7.23 (m, 2H), 6.31 (d, J = 2.0 Hz, 1H), 5.95 (dd, J = 10.1, 3.5 Hz, 1H), 5.88 (dd, J = 3.5, 1.7 Hz, 1H), 5.81 (dd, J = 10.2, 3.2 Hz, 1H), 5.75–5.71 (m, 1H), 5.69 (t, J = 8.9 Hz, 1H), 5.19 (d, J = 1.7 Hz, 1H), 4.40–4.32 (m, 2H), 4.24 (dq, J = 9.5, 6.2 Hz, 1H), 2.25 (s, 3H), 1.43 (d, J = 6.2 Hz, 3H), 1.39 (d, J = 6.2 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) & 168.99, 165.97, 165.44, 165.36, 165.19, 133.52, 133.49, 133.35, 133.16, 130.11, 129.98, 129.95, 129.90, 129.87, 129.84, 129.42, 129.38, 129.27, 128.79, 128.65, 128.64, 128.57, 128.49, 128.37, 99.52, 92.40, 74.92, 71.72, 71.40, 70.74, 70.56, 69.74, 69.67, 67.97, 21.24, 17.87, 17.61.

To a stirred solution of **S19** (116.3 mg, 0.133 mmol) in dry CH₂Cl₂ (2.0 mL) was added 33% HBr in HOAc (0.5 mL) dropwise at 0 °C. After stirring for 1 h, the mixture was diluted with CH₂Cl₂ (20 mL) and washed with ice-water (2×10 mL), sat. aq. NaHCO₃ (2×10 mL), and brine (2×10 mL) successively. The organic layer was dried over Na₂SO₄, filtered, and concentrated to give **11** as a white solid, which was pure enough and was used directly. **11**: ¹H NMR (500 MHz, CDCl₃) δ 8.06–7.98 (m, 8H), 7.84 (d, *J* = 8.3 Hz, 2H), 7.57 (t, *J* = 7.5 Hz, 1H), 7.53 (q, *J* = 6.5 Hz, 2H), 7.46–7.38 (m, 8H), 7.34 (t, *J* = 7.7 Hz, 2H), 7.26 (t, *J* = 7.5 Hz, 2H), 6.62 (d, *J* = 1.5 Hz, 1H), 6.17 (dd, *J* = 10.2, 3.3 Hz, 1H), 5.93 (dd, *J* = 10.1, 3.4 Hz, 1H), 5.82–5.74 (m, 2H), 5.70 (t, *J* = 10.0 Hz, 1H), 5.16 (d, *J* = 1.7 Hz, 1H), 4.60 (dd, *J* = 3.1, 1.5 Hz, 1H), 4.43–4.34 (m, 1H), 4.33 (dt, *J* = 12.5, 6.2 Hz, 1H), 1.47 (d, *J* = 6.3 Hz, 3H), 1.40 (d, *J* = 6.2 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 165.95, 165.82, 165.47, 165.35, 165.18, 133.58, 133.54, 133.51, 133.48, 133.21, 130.13, 129.99, 129.97, 129.95, 129.83, 129.33, 129.30, 129.25, 128.83, 128.67, 128.60, 128.55, 128.39, 99.90, 86.14, 80.30, 77.42, 71.87,

71.63, 71.23, 70.47, 69.78, 69.69, 68.17, 17.80, 17.44; HRMS (ESI) calcd. for $C_{32}H_{35}N_2O_{13}Na (M + Na)^+ m/z$ 915.1623, found 915.1626.

4-*O*-Acetyl-3-*O*-(2,3,4-tri-*O*-acetyl-α-L-fucopyranosyl)-6-*O*-benzoyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl bromide (1m)

Compound 1m was prepared employing a literature procedure¹¹.



Supplementary Figure 10. Preparation of disaccharide bromide **1m**. Reagents and conditions: a) MeONa, MeOH/CH₂Cl₂ (10:1, v/v), 25 °C, 4 h; b) PhCH(OMe)₂, *p*-TSA (10 mol%), DMF, 50 °C, 5 h; c) TMSCl (5.2 equiv.), Et₃N (6.0 equiv.), DMF, 0 °C \rightarrow 25 °C, 5 h; d) TMSI (1.0 equiv.), 4 Å MS, CH₂Cl₂, 25 °C, 1 h; e) 2,6-di-*tert*-butylpyridine (5.0 equiv.), 25 °C, 36 h; f) MeOH/HOAc (6:1), 25 °C, 1 h; g) Ac₂O (50 equiv.), pyridine, 25 °C, overnight, 58% over 3 steps; h) HOAc/H₂O (3:2), 100 °C, 1.5 h, 80%; i) BzCl (6.0 equiv.), pyridine, 25 °C, 1 h, 82%; j) Ac₂O (10 equiv.), pyridine, 25 °C, overnight, 57% over 2 steps; l) 33% HBr in HOAc, CH₂Cl₂, 0 °C \rightarrow 25 °C, 1 h, quantitative.

To a stirred solution of L-fucose (6.7 g, 40 mmol) and Et_3N (33 mL, 0.24 mol, 6.0 equiv.) in dry DMF (200 mL) was added TMSCl (26.36 mL, 0.208 mol, 5.2 equiv.) slowly at 0 °C. The reaction mixture was stirred for 10 mins, after which the mixture was warmed to room temperature slowly and kept stirring for 5 h. After dilution with *n*-hexane (400 mL), the mixture was washed with water (4×400 mL) and brine (3×100 mL), and then dried over Na₂SO₄. After removal of solvent, pertrimethylsilyl fucose **S21** was obtained as a colorless oil (16.3 g, 90 %).

To a stirred solution of **S21** (13.58 g, 30 mmol) and 4Å MS (10 g) in dry CH_2Cl_2 (200 mL) was added TMSI (4.1 mL, 30 mmol, 1.0 equiv.) at room temperature. The

mixture was stirred for 60 min in the dark by wrapped in aluminum foil to give the corresponding iodide **S22**.

p-Methoxyphenyl 4,6-O-benzyliden-2-deoxy-2-phthalimido-β-D-glucopyranoside S20 (5 g, 10 mmol) was added into the above reaction mixture. 2,6-Di-tertbutylpyridine (11.4 mL, 50 mmol) was dissolved in dry CH₂Cl₂ (100 mL) and transferred into the above mixture. The mixture was stirred at room temperature for 36 h, the molecular sieves were filtered off, the filtrate was concentrated and dried over oil pump. The crude product was dissolved in MeOH/HOAc (6:1, v/v, 48 mL), and the mixture was stirred at room temperature for 1 h. After evaporation of the solvent, the crude product was dried over oil pump and was dissolved in pyridine (100 mL). Acetic anhydride (47 mL, 0.5 mol) was added, and the mixture was stirred at room temperature overnight. After removal of solvent, the residue was dissolved in AcOEt (100 mL), and was then washed with 1 N aq. HCl (2×20 mL) and brine (3×30 mL). The organic layer was dried over MgSO₄ and concentrated. The crude product was purified by column chromatography to yield p-methoxyphenyl 4,6-O-benzylidene-3-O-(2,3,4-tri-O-acetyl- α -L-fucopyranosyl)-2-deoxy-2-phthalimido- β -D-glucopyranoside S23 (4.5 g, 58 % over 3 steps): ¹H NMR (500 MHz, CDCl₃) δ 7.85 (dd, J = 5.6, 3.1 Hz, 2H), 7.76 (dd, J= 5.5, 3.0 Hz, 2H), 7.49 (dd, J = 7.7, 1.8 Hz, 2H), 7.41–7.32 (m, 3H), 6.77 (d, J = 9.2Hz, 2H), 6.71 (d, J = 9.2 Hz, 2H), 5.62 (d, J = 8.5 Hz, 1H), 5.59 (s, 1H), 5.26 (dd, J = 10.9, 3.3 Hz, 1H), 5.03 (dd, J = 3.4, 1.2 Hz, 1H), 4.94–4.86 (m, 2H), 4.78 (dd, J = 11.0, 3.9 Hz, 1H), 4.56 (dd, J = 10.3, 8.5 Hz, 1H), 4.42 (dd, J = 10.4, 4.4 Hz, 1H), 4.33 (q, J= 6.6 Hz, 1H), 3.87 (t, J = 10.0 Hz, 1H), 3.83 (t, J = 9.0 Hz, 1H), 3.80–3.78 (m, 1H), 3.71 (s, 3H), 2.01 (s, 3H), 1.96 (s, 3H), 1.92 (s, 3H); 13 C NMR (125 MHz, CDCl₃) δ 170.66, 170.65, 170.07, 155.77, 150.68, 137.01, 134.62, 129.61, 128.52, 126.61, 123.80, 118.68, 114.63, 102.52, 98.42, 95.99, 80.35, 72.13, 71.29, 68.85, 68.04, 67.95, 66.72, 64.74, 56.40, 55.73, 20.81, 20.79, 20.69, 15.19; HRMS (ESI) calcd. for $C_{40}H_{41}NO_{15}Na (M + Na)^+ m/z 798.2368$, found 798.2367.

A solution of **S23** (4.5 g, 5.8 mmol) in HOAc/H₂O (3:2, v/v, 60 mL) was stirred at 100 $^{\circ}$ C for 1.5 h. The reaction mixture was cooled to room temperature and neutralized carefully with sat. aq. NaHCO₃ under 0 $^{\circ}$ C. The mixture was extracted with ethyl acetate, washed with brine, and dried over MgSO₄. After concentration, the crude product was purified by a short silica gel column chromatography (firstly with petroleum ether/ethyl acetate = 2:1 to remove the generated benzaldehyde, then with CH₂Cl₂/MeOH = 10:1) to afford the crude diol (3.2 g, 80%), which and was used directly in the next step.

The above crude diol (2.43 g, 3.53 mmol) was dissolved in pyridine (15 mL) and BzCl (2.0 mL, 17.6 mmol, 5.0 equiv.) was added. The mixture was stirred at room temperature for 1 h. After completion, as monitored by TLC (petroleum ether/ethyl acetate = 1:1), the solvent was removed *in vacuo*. The residue was dissolved in AcOEt (150 mL) and was then washed with 1 N sat. aq. HCl (2×30 mL) and brine (2×30 mL) successively. The organic layer was dried over MgSO₄ and concentrated. The crude product was purified by column chromatography to give *p*-methoxyphenyl 3-*O*-(2,3,4-tri-*O*-acetyl- α -L-fucopyranosyl)-6-*O*-benzoyl-2-deoxy-2-phthalimido- β -D-glucopyranoside **S24** (2.3 g, 82%): ¹H NMR (500 MHz, CDCl₃) δ 8.07 (d, *J* = 7.7 Hz,

2H), 7.87 (s, 1H), 7.73 (dd, J = 4.9, 3.0 Hz, 2H), 7.59 (t, J = 7.4 Hz, 1H), 7.46 (t, J = 7.6 Hz, 2H), 6.80 (d, J = 9.0 Hz, 2H), 6.60 (d, J = 8.9 Hz, 2H), 5.63 (d, J = 7.7 Hz, 1H), 5.31 (d, J = 1.9 Hz, 1H), 5.22 (ddd, J = 9.0, 3.5, 1.6 Hz, 1H), 5.08 (m, 2H), 4.74 (dd, J = 11.9, 2.2 Hz, 1H), 4.66 (dd, J = 11.9, 6.1 Hz, 1H), 4.57–4.46 (m, 3H), 3.90 (ddd, J = 8.8, 6.3, 2.1 Hz, 1H), 3.69 (t, J = 8.0 Hz, 1H), 3.66 (s, 3H), 2.10 (s, 3H), 1.89 (s, 3H), 1.30 (s, 3H), 1.13 (d, J = 6.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 170.49, 170.05, 169.80, 166.71, 155.56, 150.71, 134.44, 133.33, 131.66, 129.91, 129.86, 128.52, 123.80, 118.72, 114.41, 97.60, 97.49, 80.70, 80.67, 74.04, 70.94, 70.89, 67.63, 67.55, 66.18, 63.92, 55.61, 54.78, 20.69, 20.65, 19.62, 15.86; HRMS (ESI) calcd. for C₄₀H₄₁NO₁₆Na (M + Na)⁺ m/z 814.2318, found 814.2320.

To a stirred solution of **S24** (396 mg, 0.5 mmol) in pyridine (3 mL) was added Ac₂O (0.49 mL, 10 equiv.). After stirring at room temperature overnight, methanol (0.7 mL) was added, and the mixture was stirred for another one hour. After removal of the solvent, the crude product was purified by column chromatography (petroleum ether/ethyl acetate = 1:1) to yield *p*-methoxyphenyl 4-O-acetyl-3-O-(2,3,4-tri-O-acetyl- α -L-fucopyranosyl)-6-O-benzoyl-2-deoxy-2-phthalimido- β -D-glucopyranoside S25 (366 mg, 88%): ¹H NMR (500 MHz, CDCl₃) δ 8.05 (d, J = 7.1 Hz, 2H), 7.78–7.70 (m, 2H), 7.59 (t, J = 8.0 Hz, 1H), 7.46 (t, J = 7.7 Hz, 2H), 6.81 (d, J = 9.1 Hz, 2H), 6.58 (d, *J* = 9.1 Hz, 2H), 5.73 (d, *J* = 8.0 Hz, 1H), 5.22 (dd, *J* = 3.4, 1.4 Hz, 1H), 5.15 (dd, *J* = 9.9, 8.3 Hz, 1H), 5.13 (dd, J = 11.0, 3.3 Hz, 1H), 5.05 (d, J = 3.4 Hz, 1H), 5.00 (dd, J = 11.0, 3.4 Hz, 1H), 4.65–4.48 (m, 3H), 4.36 (dd, J = 12.1, 7.3 Hz, 1H), 4.27 (q, J = 7.7, 7.0 Hz, 1H), 3.94 (ddd, J = 10.1, 7.3, 2.8 Hz, 1H), 3.66 (s, 3H), 2.10 (s, 3H), 2.09 $(s, 3H), 1.82 (s, 3H), 1.48 (s, 3H), 1.06 (d, J = 6.5 Hz, 3H); {}^{13}C NMR (125 MHz, CDCl_3)$ δ 170.52, 170.46, 170.11, 169.29, 166.21, 155.71, 150.63, 134.30, 133.33, 129.94, 129.91, 128.55, 118.96, 114.45, 97.54, 97.44, 72.59, 72.04, 71.43, 68.56, 67.20, 66.20, 63.40, 55.65, 55.13, 21.22, 20.74, 20.60, 20.11, 15.42; HRMS (ESI) calcd. for $C_{42}H_{43}NO_{17}Na (M + Na)^+ m/z 856.2423$, found 856.2422.

To a stirred solution of **S25** (366 mg, 0.44 mmol) in CH₃CN/H₂O (4:1, v/v, 5 mL) was added diammonium cerium (IV) nitrate (723 mg, 1.32 mmol). After stirring at room temperature for 1 h, the mixture was diluted with EtOAc (50 mL) and washed with water (3×15 mL) and brine (3×15 mL) successively. The organic layer was dried over Na₂SO₄ and concentrated *in vacuo*. The crude product was dissolved in pyridine (4 mL), and Ac₂O (0.6 mL, 15 equiv.) was added. After stirring at room temperature overnight, methanol (1.0 mL) was added, and the mixture was stirred for another one hour. After removal of solvent, the crude product was purified by column chromatography (petroleum ether/ethyl acetate = 1:1) to give 4-*O*-acetyl-3-*O*-(2,3,4-tri-*O*-acetyl-α-L-fucopyranosyl)-6-*O*-benzoyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl acetate **S26** (193 mg, 57% over 2 steps): ¹H NMR (500 MHz, CDCl₃) δ 8.09 (d, *J* = 7.2 Hz, 2H), 7.89 (dd, *J* = 5.1, 3.0 Hz, 2H), 7.76 (dd, *J* = 5.4, 3.0 Hz, 2H), 7.58 (t, *J* = 7.4 Hz, 1H), 7.47 (t, *J* = 7.7 Hz, 2H), 6.45 (d, *J* = 8.8 Hz, 1H), 5.20 (t, *J* = 9.4 Hz, 1H), 5.20 (d, *J* = 11.0, 3.4 Hz, 1H), 4.65 (dd, *J* = 10.6, 8.8 Hz, 1H), 4.51 (dd, *J* = 3.4 Hz, 1H), 4.99 (dd, *J* = 11.0, 3.4 Hz, 1H), 4.65 (dd, *J* = 10.6, 8.8 Hz, 1H), 4.51 (dd, *J* =

10.7, 9.0 Hz, 1H), 4.50 (dd, J = 11.7, 2.8 Hz, 1H), 4.32 (dd, J = 12.3, 5.4 Hz, 1H), 4.24 (q, J = 6.3 Hz, 1H), 3.99 (ddd, J = 9.9, 5.4, 2.7 Hz, 1H), 2.09 (s, 4H), 2.08 (s, 3H), 1.98 (s, 3H), 1.81 (s, 3H), 1.53 (s, 3H), 1.05 (d, J = 6.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 170.48, 170.39, 169.83, 169.30, 168.83, 167.67, 166.34, 134.38, 133.25, 131.59, 129.93, 129.82, 128.51, 124.01, 97.47, 89.78, 76.55, 73.14, 71.33, 71.28, 68.42, 67.09, 66.17, 62.85, 54.11, 21.15, 20.90, 20.68, 20.54, 20.14, 15.35; HRMS (ESI) calcd. for C₃₇H₃₉NO₁₇Na (M + Na)⁺ m/z 792.2110, found 792.2115.

To a stirred solutio of **S26** (123 mg, 0.16 mmol) in CH₂Cl₂ (1.0 mL) was added 33% HBr in HOAc (0.2 mL) dropwise under 0 °C. The mixture was removed from the ice bath and stirred for 1 h at room temperature. The mixture was diluted with CH₂Cl₂ (50 mL), and was then washed with ice-water (3×20 mL), sat. aq. NaHCO₃ (3×10 mL), and brine (2×20 mL) successively. The organic layer was dried over Na₂SO₄, filtered, and concentrated to give 1m as a pale yellow solid, which was in high purity and was used directly in the coupling reaction. **1m**: ¹H NMR (500 MHz, CDCl₃) δ 8.10 (br, 2H), 7.90 (s, 2H), 7.77 (dd, *J* = 5.8, 2.9 Hz, 2H), 7.59 (t, *J* = 7.0 Hz, 1H), 7.48 (t, *J* = 7.4 Hz, 2H), 6.35 (d, J = 9.5 Hz, 1H), 5.24 (t, J = 9.4 Hz, 1H), 5.20 (d, J = 2.9 Hz, 1H), 5.08 (dd, J = 11.0, 3.2 Hz, 1H), 5.03 (d, J = 3.4 Hz, 1H), 4.98 (dd, J = 11.1, 3.3 Hz, 1H), 4.67 (t, J = 9.9 Hz, 1H), 4.54 (dd, J = 12.9, 3.1 Hz, 1H), 4.50 (t, J = 9.3 Hz, 1H), 4.34 (dd, J = 12.9, 3.1 Hz, 1H), 4.50 (t, J = 9.3 Hz, 1H), 4.54 (dd, J = 12.9, 3.1 Hz, 1H), 4.50 (t, J = 9.3 Hz, 1H), 4.54 (dd, J = 12.9, 3.1 Hz, 1H), 4.50 (t, J = 9.3 Hz, 1H), 4.54 (dd, J = 12.9, 3.1 Hz, 1H), 4.50 (t, J = 9.3 Hz, 1H), 4.54 (dd, J = 12.9, 3.1 Hz, 1H), 4.50 (t, J = 9.3 Hz, 1H), 4.54 (dd, J = 12.9, 3.1 Hz, 1H), 4.50 (t, J = 9.3 Hz, 1H), 4.50 (t, J = 9.3 Hz, 1H), 4.50 (t, J = 12.9, 3.1 Hz, 1H), 4.50 (t, J = 9.3 Hz, 1H), 4.50 (t, J = 9.3 Hz, 1H), 4.50 (t, J = 12.9, 3.1 Hz, 1H), 4.50 (t, J = 9.3 Hz, 1H), 4.50 (t 12.6, 5.6 Hz, 1H), 4.23 (q, J = 6.7 Hz, 1H), 3.95 (ddd, J = 9.0, 5.6, 2.3 Hz, 1H), 2.08 (s, 6H), 1.81 (s, 3H), 1.54 (s, 3H), 1.05 (d, J = 6.4 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 170.47, 170.44, 169.77, 169.24, 166.35, 134.47, 133.39, 131.65, 129.99, 129.73, 128.57, 124.13, 97.51, 77.81, 77.41, 76.91, 71.34, 71.17, 68.47, 67.05, 66.27, 63.00, 58.68, 21.14, 20.71, 20.56, 20.18, 15.40.

2,4-Di-O-benzoyl-3-O-(4-O-benzoyl-3-O-(3,4-di-O-benzoyl-2-O-(2,3,4-tri-O-benzoyl- α -L-rhamnopyranosyl)- α -L-rhamnopyranosyl)-2-O-(3,4,6-tri-O-acetyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)- α -L-rhamnopyranosyl)- α -L-rham



Supplementary Figure 11. Preparation of pentasaccharide bromide S29.

Allyl 2,4-di-O-benzoyl-3-O-(4-O-benzoyl-3-O-(3,4-di-O-benzoyl-2-O-(2,3,4-tri-O-benzoyl-α-L-rhamnopyranosyl)-α-L-rhamnopyranosyl)-2-O-(3,4,6-tri-O-acetyl-2deoxy-2-phthalimido-\beta-D-glucopyranosyl)-a-L-rhamnopyranosyl)-a-Lrhamnopyranoside **S27** was prepared employing a literature procedure¹⁰. **S27**: ¹H NMR (500 MHz, CDCl₃) δ 8.14 (d, J = 7.2 Hz, 2H), 8.08–8.01 (m, 4H), 7.98 (d, J = 7.3 Hz, 2H), 7.94 (ddd, J = 8.4, 3.9, 1.3 Hz, 4H), 7.85–7.80 (m, 2H), 7.80 (d, J = 7.3 Hz, 2H), 7.61 (td, J = 17.7, 16.5, 7.5 Hz, 5H), 7.54 (d, J = 7.7 Hz, 2H), 7.52–7.48 (m, 5H), 7.45 (t, J = 7.7 Hz, 2H), 7.42–7.34 (m, 8H), 7.27 (d, J = 8.2 Hz, 2H), 7.24 (d, J = 8.0 Hz, 2H), 7.01 (t, J = 7.8 Hz, 2H), 6.84 (t, J = 7.5 Hz, 1H), 6.02–5.91 (m, 2H), 5.81 (dd, J = 10.0, 3.4 Hz, 1H), 5.77 (dd, J = 3.5, 1.7 Hz, 1H), 5.74 (dd, J = 10.3, 3.1 Hz, 1H), 5.65 (d, J = 8.4 Hz, 1H), 5.49 (d, J = 9.7 Hz, 1H), 5.45 (d, J = 10.0 Hz, 1H), 5.41 (t, J = 9.9 Hz, 1H), 5.39–5.33 (m, H), 5.27 (dq, J = 10.5, 1.4 Hz, 1H), 5.04 (d, J = 2.0 Hz, 1H), 5.02 (d, J = 1.7 Hz, H), 4.85 (d, J = 1.7 Hz, 1H), 4.79 (d, J = 1.7 Hz, 1H), 4.67 (t, J = 9.8 Hz, 1H), 4.39 (dd, J = 9.8, 3.4 Hz, 1H), 4.29 (dd, J = 10.7, 8.3 Hz, 1H), 4.23 (ddt, J = 10.1, 7.4, 2.1 Hz, 2H), 4.10–3.97 (m, 5H), 3.89 (t, J = 2.3 Hz, 1H), 3.83 (dq, J = 9.8, 6.1 Hz, 1H), 3.58 (ddd, J = 9.7, 6.3, 3.3 Hz, 2H), 3.53 (d, J = 10.1 Hz, 1H), 2.06 (s, 3H), 2.03 (s, 3H), 1.83 (s, 3H), 1.28 (d, J = 6.1 Hz, 3H), 1.26 (d, J = 6.3 Hz, 3H), 0.82 (d, J = 6.3 Hz, 3H), 0.69 (d, J = 6.2 Hz, 3H).

To a stirred solutio of S27 (44.5 mg, 0.0235 mmol) in MeOH (0.5 mL) was added PdCl₂ (1.2 mg, 29 mol%). After stirring at room temperature for 12 h, the mixture was diluted with CH₂Cl₂, filtered, and concentrated to dryness. The crude product was dissolved in pyridine (2.0 mL), and Ac₂O (0.2 mL) was added at 0 °C. The reaction mixture was stirred at room temperature for 2 h. After quenching with MeOH (0.2 mL) under 0 °C and stirring for another 2.0 h, the solvent was removed. The crude product was purified by preparative TLC (petroleum ether/CH₂Cl₂/ethyl acetate = 2:1:1) to afford 2,4-di-O-benzoyl-3-O-(4-O-benzoyl-3-O-(3,4-di-O-benzoyl-2-O-(2,3,4-tri-Obenzoyl-α-L-rhamnopyranosyl)-α-L-rhamnopyranosyl)-2-O-(3,4,6-tri-O-acetyl-2deoxy-2-phthalimido-\beta-D-glucopyranosyl)-a-L-rhamnopyranosyl)-a-Lrhamnopyranosyl acetate S28 (23.1 mg, 52% over two steps): ¹H NMR (500 MHz, CDCl₃) δ 8.14 (d, J = 7.3 Hz, 2H), 8.08–8.04 (m, 4H), 7.99 (d, J = 7.3 Hz, 2H), 7.95 (dd, J = 7.2, 4.4 Hz, 4H), 7.86–7.82 (m, 2H), 7.80 (d, J = 7.3 Hz, 2H), 7.69–7.58 (m, 5H), 7.58–7.48 (m, 7H), 7.47–7.43 (m, 2H), 7.43–7.34 (m, 8H), 7.27 (m, 4H), 7.02 (t, J = 7.8 Hz, 2H), 6.86 (t, J = 7.5 Hz, 1H), δ 6.25 (d, J = 1.9 Hz, 1H), 5.99 (dd, J = 10.8, 9.0 Hz, 1H), 5.81 (dd, J = 10.1, 3.4 Hz, 1H), 5.78–5.73 (m, 2H), 5.66 (d, J = 8.4 Hz, 1H), 5.53 (t, J = 9.9 Hz, 1H), 5.47 (t, J = 10.0 Hz, 1H), 5.42 (t, J = 9.9 Hz, 1H), 5.37 (dd, J = 3.5, 2.1 Hz, 1H), 5.11-5.02 (m, 2H), 4.85 (d, J = 1.7 Hz, 1H), 4.80 (d, J = 1.6 Hz)Hz, 1H), 4.69 (t, J = 9.8 Hz, 1H), 4.40 (dd, J = 9.9, 3.4 Hz, 1H), 4.28 (dd, J = 10.8, 8.4 Hz, 1H), 4.22 (dd, J = 10.0, 2.5 Hz, 1H), 4.10 (dd, J = 9.9, 6.2 Hz, 1H), 4.07 (dd, J = 3.2, 1.6 Hz, 1H), 4.03-3.96 (m, 2H), 3.92 (t, J = 2.3 Hz, 1H), 3.85 (dq, J = 12.4, 6.1 Hz, 1H), 3.60 (ddt, J = 9.9, 7.7, 3.6 Hz, 3H), 2.21 (s, 3H), 2.05 (s, 3H), 2.04 (s, 3H), 1.83 (s, 3H), 1.32-1.22 (m, 6H), 0.83 (d, J = 6.3 Hz, 3H), 0.70 (d, J = 6.2 Hz, 3H); ${}^{13}C$ NMR (125 MHz, CDCl₃) δ 170.80, 169.96, 169.83, 168.47, 167.75, 165.90, 165.87, 165.71, 165.54, 165.38, 165.15, 165.07, 164.25, 134.17, 133.81, 133.79, 133.58, 133.52, 133.18, 133.16, 133.05, 133.02, 131.51, 130.08, 130.04, 129.99, 129.97, 129.93, 129.80, 129.65, 129.57, 129.54, 129.53, 129.47, 129.42, 129.41, 129.35, 128.99, 128.91, 128.85, 128.71, 128.51, 128.48, 128.45, 128.37, 128.34, 124.04, 101.37, 99.12, 98.91, 98.23, 90.48, 76.52, 76.08, 75.51, 73.80, 72.57, 72.52, 71.88, 71.72, 71.42, 71.04, 70.52, 70.28, 69.53, 69.22, 68.90, 68.41, 68.17, 67.42, 61.77, 54.66, 21.12, 20.92, 20.87, 20.68, 17.78, 17.65, 17.49, 17.28; HRMS (ESI) calcd. for C₁₀₂H₉₅NO₃₅Na (M + Na)⁺ m/z 1916.5577, found 1916.5582.

To a stirred solution of **S28** (23.1 mg, 0.0122 mmol) in dry CH₂Cl₂ (1.0 mL) was added 33% HBr in HOAc (0.2 mL) dropwise at 0 °C. The mixture was stirred at room temperature and was monitored by TLC. After completion (1 h), the mixture was diluted with CH₂Cl₂ (20 mL) and was washed with ice-water (2×5 mL), NaHCO₃ (2×5 mL), and brine (2×5 mL), successively. The organic layer was dried over Na₂SO₄ and filtered. The filtrate was concentrated under reduced pressure to give **S29** as a white solid, which was used directly for the following coupling reaction.

$\label{eq:2.3.4} 3-O-(2,3,4-Tri-O-acetyl-\alpha-L-fucopyranosyl)-4-O-(2,3,4,6-tetra-O-acetyl-\beta-D-galactopyranosyl)-6-O-benzoyl-2-deoxy-2-phthalimido-\beta-D-glucopyranosyl bromide (S33)$

Compound S33 was prepared employing a literature procedure¹¹.



Supplementary Figure 12. Preparation of Lewis^x trisaccharide bromide **S33**. Reagents and conditions: a) TMSOTf (20 mol%), CH₂Cl₂/Et₂O (1:5), 4 Å MS, -40 $\mathbb{C} \rightarrow 25 \mathbb{C}$, 8 h, 48 %; b) CAN (3.0 equiv.), CH₃CN/H₂O (4:1), 25 \mathbb{C} ; 1.5 h; c) Ac₂O (15.0 equiv.), pyridine, 25 \mathbb{C} , overnight; d) 33% HBr in HOAc, CH₂Cl₂, 0 $\mathbb{C} \rightarrow 25 \mathbb{C}$, 2 h, quantitative.

To a stirred solution of 2,3,4,6-tetra-*O*-acetyl-D-galactopyranose (1.67 g, 5 mmol) in CH_2Cl_2 (20 mL) and Cl_3CCN (6 mL, 60 mmol) was added K_2CO_3 (3.5 g, 25 mmol). The mixture was stirred at room temperature for 6 h. After filtration, the mixture was concentrated and azeotroped with toluene (2×20 mL) to give trichloroacetimidate 2,3,4,6-tetra-*O*-acetyl-D-galactopyranoside **S30**, which was used directly in the next step.

Compound S24 (529 mg, 0.67 mmol, 1.0 equiv.), S30 (3.88 mmol, 5.8 equiv.), and 4 Å MS (1 g) were suspended in dry CH₂Cl₂ (3 mL) and Et₂O (15 mL). The mixture was stirred under -40 °C for 0.5 h, after which TMSOTf (19 µL, 20 mol %) was injected. After stirring at -40 °C for 0.5 h, the reaction was allowed to warm to 0 °C slowly and stirred for 4 h at 0 °C. The reaction was guenched with Et₃N (0.5 mL). After filtration, the mixture was concentrated and purified by column chromatography (petroleum ether/ethyl acetate/ $CH_2Cl_2 = 2:1:0.5$ to 2:1:1.5) to give recovered **S30** (180 mg, 34%) and *p*-methoxyphenyl 3-O-(2,3,4-tri-O-acetyl-a-L-fucopyranosyl)-4-O-(2,3,4,6-tetra-O-acetyl-β-D-galactopyranosyl)-6-O-benzoyl-2-deoxy-2-phthalimido-β-Dglucopyranoside **S31** (360 mg, 48%): ¹H NMR (500 MHz, CDCl₃) δ 8.07 (d, J = 7.9 Hz, 2H), 7.85 (dd, J = 5.4, 3.1 Hz, 2H), 7.77 (dd, J = 5.5, 3.1 Hz, 2H), 7.64 (t, J = 7.4 Hz, 1H), 7.53 (t, J = 7.7 Hz, 2H), 6.72 (d, J = 9.1 Hz, 2H), 6.57 (d, J = 9.0 Hz, 2H), 5.53 (d, J = 8.6 Hz, 1H), 5.40 (d, J = 3.5 Hz, 1H), 5.39 (d, J = 3.5 Hz, 1H), 5.17 (dd, J = 11.0, 3.4 Hz, 1H), 5.15 (dd, J = 10.4, 8.0 Hz, 1H), 5.03-4.95 (m, 3H), 4.95 (dd, J = 10.3, 3.5 Hz, 1H), 4.85 (dd, J = 10.5, 9.0 Hz, 1H), 4.83 (dd, J = 10.9, 4.1 Hz, 1H), 4.66 (d, J = 8.2 Hz, 1H), 4.58 (dd, J = 11.6, 6.7 Hz, 1H), 4.54 (dd, J = 10.8, 8.8 Hz, 1H),

4.39 (dd, J = 11.8, 5.6 Hz, 1H), 4.31 (dd, J = 11.5, 7.5 Hz, 1H), 4.11 (t, J = 9.4 Hz, 1H),

3.93 (ddd, J = 10.0, 5.7, 1.9 Hz, 1H), 3.83 (t, J = 7.0 Hz, 1H), 3.66 (m, 3H), 2.19 (s, 3H), 2.14 (s, 3H), 2.11 (s, 3H), 2.09 (s, 3H), 2.07 (s, 3H), 1.97 (s, 3H), 1.93 (s, 3H), 1.23 (d, J = 6.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 170.84, 170.60, 170.59, 170.09, 169.89, 169.22, 165.79, 155.69, 150.70, 134.62, 133.69, 129.84, 129.53, 128.84, 123.86, 118.82, 114.45, 100.85, 97.86, 95.39, 75.82, 73.53, 71.47, 71.33, 71.30, 71.19, 68.99, 68.26, 67.87, 66.80, 64.44, 62.42, 60.82, 56.35, 55.63, 20.94, 20.94, 20.85, 20.81, 20.78, 20.68, 20.65, 16.01; HRMS (ESI) calcd. for C₅₄H₅₉NO₂₅Na (M + Na)⁺ m/z 1144.3268, found 1144.3254.

To a stirred solution of **S31** (360 mg, 0.321 mmol) in CH₃CN/H₂O (4:1, v/v, 3.2 mL) was added diammonium cerium (IV) nitrate (528 mg, 3.0 equiv.). After stirring at room temperature for 1.5 h, the mixture was diluted with EtOAc (50 mL) and washed with water (3×15 mL) and brine (3×15 mL) successively. The organic layer was dried over Na₂SO₄ and was concentrated. A short silica-gel column chromatography was used to remove the impurity with high mobility and insoluble impurities (petroleum ether/ethyl acetate = 2:1 to 1:1) to provide the crude product (273 mg, 83%). The crude product was dissolved in pyridine (3 mL), and Ac₂O (0.38 mL, 15.0 equiv.) was added. The mixture was stirred at room temperature overnight, after which methanol (0.5 mL) was added. The mixture was stirred for another one hour. After removal of solvent, the mixture was purified by column chromatography (petroleum ether/ethyl acetate = 1:1) to give 3-O-(2,3,4-tri-O-acetyl-α-L-fucopyranosyl)-4-O-(2,3,4,6-tetra-O-acetyl-β-Dgalactopyranosyl)-6-*O*-benzoyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl acetate **S32** (185 mg, 54% over 2 steps): ¹H NMR (500 MHz, CDCl₃) δ 8.09 (d, J = 7.3 Hz, 2H), 7.86 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.79 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.69–7.61 (m, 1H), 7.56 (t, J = 7.6 Hz, 2H), 6.25 (d, J = 8.8 Hz, 1H), 5.37 (d, J = 3.5 Hz, 2H), 5.19–5.09 (m, 2H), 4.98 (q, J = 7.0 Hz, 1H), 4.95-4.89 (m, 3H), 4.87 (dd, J = 10.3, 3.5 Hz, 1H), 4.81 (dd, J = 10.9, 4.1 Hz, 1H), 4.67 (d, J = 8.1 Hz, 1H), 4.56 (dd, J = 11.3, 6.2 Hz, 1H), 4.44 (dd, J = 10.4, 8.9 Hz, 1H), 4.37 (dd, J = 12.3, 3.6 Hz, 1H), 4.28 (dd, J = 11.6, 7.8 Hz, H), 4.16 (t, J = 9.4 Hz, 1H), 3.95 (ddd, J = 10.2, 3.9, 2.1 Hz, 1H), 3.71 (t, J = 7.0 Hz, 1H), 2.18 (s, 3H), 2.13 (s, 3H), 2.09 (s, 3H), 2.08 (s, 3H), 2.07 (s, 3H), 1.96 (s, 3H), 1.93 (s, 3H), 1.91 (s, 3H), 1.23 (d, J = 6.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 170.78, 170.71, 170.53, 170.43, 170.05, 169.84, 169.06, 168.85, 165.70, 134.71, 133.77, 131.23, 129.75, 129.40, 128.93, 123.94, 100.50, 95.31, 90.11, 77.16, 74.73, 73.86, 71.40, 71.29, 71.01, 68.86, 68.12, 67.88, 66.70, 64.45, 61.95, 60.69, 55.36, 20.89, 20.87, 20.81, 20.79, 20.79, 20.74, 20.65, 20.62, 15.97; HRMS (ESI) calcd. for $C_{49}H_{55}NO_{25}Na (M + Na)^+ m/z 1080.2955$, found 1080.2944.

To a stirred solution of **S32** (185 mg, 0.175 mmol) in dry CH₂Cl₂ (1.0 mL) was added 33% HBr in HOAc (0.2 mL) dropwise under 0 °C. The mixture was warmed to room temperature and was stirred for 1.5 h. After dilution with CH₂Cl₂ (50 mL), the mixture was washed with ice-water (3×20 mL), sat. aq. NaHCO₃ (3×10 mL), and brine (2×20 mL), successively. The organic layer was dried over Na₂SO₄, filtered, and concentrated to give **S33**, which was pure enough and was used directly in the coupling reaction. **S33**: ¹H NMR (500 MHz, CDCl₃) δ 8.10 (d, *J* = 7.9 Hz, 2H), 7.91–7.85 (m,

2H), 7.83–7.77 (m, 2H), 7.66 (t, J = 7.4 Hz, 1H), 7.56 (t, J = 7.1 Hz, 2H), 6.19 (d, J = 10.1 Hz, 1H), 5.37 (d, J = 3.5 Hz, 2H), 5.14 (dd, J = 7.7, 2.9 Hz, 1H), 5.12 (dd, J = 11.34, 7.38 Hz, 1H), 4.98 (d, J = 2.7 Hz, 1H), 4.96 (s, 1H), 4.91 (d, J = 2.9 Hz, 1H), 4.88 (d, J = 10.3, 3.4 Hz, 1H), 4.83 (dd, J = 10.9, 4.1 Hz, 1H), 4.76 (t, J = 9.6 Hz, 1H), 4.67 (d, J = 8.2 Hz, 1H), 4.62 (t, J = 9.8 Hz, 1H), 4.55 (dd, J = 11.1, 6.1 Hz, 1H), 4.38 (dd, J = 12.5, 3.8 Hz, 1H), 4.29 (dd, J = 11.2, 7.4 Hz, 1H), 4.20 (t, J = 9.5 Hz, 1H), 3.89 (dd, J = 10.0, 2.1 Hz, 1H), 3.73 (t, J = 7.1 Hz, 1H), 2.18 (s, 3H), 2.13 (s, 3H), 2.10 (s, 3H), 2.09 (s, 3H), 2.07 (s, 3H), 1.96 (s, 3H), 1.92 (s, 3H), 1.22 (d, J = 6.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 170.74, 170.72, 170.50, 170.41, 169.99, 169.78, 169.05, 165.63, 134.77, 133.80, 129.75, 129.25, 128.91, 124.00, 100.54, 95.41, 78.47, 78.19, 74.64, 71.77, 71.35, 71.32, 71.11, 68.85, 68.09, 67.66, 66.66, 64.50, 61.91, 60.69, 59.69, 59.50, 20.90, 20.87, 20.78, 20.77, 20.70, 20.60, 20.58, 15.94.

2.2 Preparation of alkyne derivatives of amino acids and peptides

Amino acids



Internal acetylenic amino acid



Supplementary Figure 13. Acetylenic amino acid and peptide derivatives used in this study.



Supplementary Figure 14. General procedures for methylation of amino acids.

Procedure A: To a stirred solution of Boc-L-propargylglycine (2.1 g, 10 mmol) in anhydrous DMF (15 mL) was added KHCO₃ (1.1 equiv.) at 0 °C. After stirring for 0.5 h at 0 °C, MeI (3.0 equiv.) was injected, and the mixture was stirred at room temperature overnight. After dilution with ethyl acetate (100 mL), the mixture was washed with water (3×30 mL), sat. aq. NaHCO₃ (3×20 mL), and brine, successively. The organic layer was dried over MgSO₄ and concentrated to afford **2a** (2.0 g, 88%), which was pure enough and used without further purification.

Procedure B: To a stirred solution of Boc-L-propargylglycine (2.1 g, 10 mmol) in anhydrous EtOH (30 mL) was added dicyclohexylamine (1.0 equiv.). After stirring at room temperature for 0.5 h, the solvent was removed *in vacuo*, and the crude product was dried over oil pump. The obtained white solid was suspended in dry DMF (50 mL), and MeI (3.2 equiv.) was added. The reaction mixture was stirred at room temperature for 30 h. After dilution with ethyl acetate (200 mL), the mixture was washed with water (3×50 mL), sat. aq. NaHCO₃ (3×50 mL), 1M aq. HCl (3×30 mL), and brine, successively. The organic layer was dried over MgSO₄ and concentrated to afford product **2a** (1.9 g, 83%), which was pure enough and was used without further purification.





Supplementary Figure 15. General procedure for Cbz/Fmoc protection of amino acids.

To a stirred solution of methyl Boc-L-propargylglycinate (454.5 mg, 2 mmol) in dry CH_2Cl_2 (4.5 mL) was added TFA (1.5 mL) slowly at 0 °C. The reaction mixture was stirred for 2–4 h (monitored by TLC) at room temperature to remove the Boc protecting

group. After completion, the reaction solution was concentrated and dried over oil pump. To a stirred solution of the crude product in THF (10 mL) was added sat. aq. NaHCO₃ (10 mL). After stirring for 0.5 h, CbzCl or FmocCl (1.3 equiv.) was added. After stirring for 4 h, the mixture was diluted with ethyl acetate. The organic phase was washed with water, 1 N aq. HCl, and brine, successively. After dried over anhydrous MgSO₄ and concentrated, the pure product was obtained by column chromatography.

N-Benzyloxycarbonyl-L-propargylglycine methyl ester (2b)¹³

CO₂Me 381 mg, 73%. ¹H NMR (500 MHz, CDCl₃) δ 7.38–7.31 (m, 5H), 5.62 (d, J = 8.4 Hz, 1H), 5.13 (s, 2H), 4.55 (dt, J = 8.9, 4.8 Hz, 1H), 3.79 (s, 3H), 2.78 (dt, J = 5.2, 2.6 Hz, 2H), 2.04 (t, J = 2.6 Hz, 1H).

N-(9-Fluorenylmethoxycarbonyl)-L-propargylglycine methyl ester (2c)¹⁴

 $\begin{array}{c} \text{CO}_2\text{Me} \\ \text{NHFmoc} \end{array} \begin{array}{c} 572 \text{ mg, } 82\%. \ ^1\text{H NMR (500 MHz, CDCl_3)} \ \delta \ 7.77 \ (\text{d}, \ J = 7.5 \ \text{Hz}, \\ 2\text{H}), \ 7.62 \ (\text{d}, \ J = 6.8 \ \text{Hz}, \ 3\text{H}), \ 7.41 \ (\text{t}, \ J = 7.4 \ \text{Hz}, \ 3\text{H}), \ 7.33 \ (\text{t}, \ J = \\ 7.5 \ \text{Hz}, \ 4\text{H}), \ 5.66 \ (\text{d}, \ J = 8.3 \ \text{Hz}, \ 1\text{H}), \ 4.56 \ (\text{dt}, \ J = 9.0, \ 4.8 \ \text{Hz}, \ 1\text{H}), \\ 4.41 \ (\text{d}, \ J = 7.2 \ \text{Hz}, \ 2\text{H}), \ 4.25 \ (\text{t}, \ J = 7.3 \ \text{Hz}, \ 1\text{H}), \ 3.81 \ (\text{s}, \ 3\text{H}), \ 2.80 \ (\text{dd}, \ J = 4.8, \ 2.6 \ \text{Hz}, \\ 2\text{H}), \ 2.08 \ (\text{t}, \ J = 2.6 \ \text{Hz}, \ 1\text{H}). \end{array}$

N-Tert-butoxycarbonyl-L-propargylglycine benzyl ester (2d)¹⁵

N-Tert-butoxycarbonyl-L-propargylglycine *tert*-butyl ester (2e)¹⁶





To a stirred solution of Boc-L-propargylglycine (530 mg, 2.5 mmol) and (Boc)₂O (1.4 equiv.) in THF (25 mL) was added DMAP (0.2 equiv.) slowly. The reaction mixture was stirred for 5 h at room temperature. After concentration, the pure product **2e** was obtained by column chromatography (280 mg, 42%): ¹H NMR (500 MHz, CDCl₃) δ 5.34 (d, *J* = 7.8 Hz, 1H), 4.34 (dt, *J* = 8.7, 4.6 Hz, 1H), 2.70 (dd, *J* = 7.6, 3.7 Hz, 2H), 2.02 (t, *J* = 2.6 Hz, 1H), 1.49 (s, 9H), 1.45 (s, 9H).

N,*N*-Phthaloyl-L-propargylglycine methyl ester (2f)¹⁷



Supplementary Figure 17. Preparation of acetylenic amino acid 2f.

To a stirred suspension of L-propargylglycine (250 mg, 2.2 mmol) in distilled water (3 ml) was added Na₂CO₃ (1.5 equiv.). After stirring for 10 mins, *N*-ethoxycarbonylphthalimide (1.5 equiv.) was added, and the reaction was continued until the solution became clear. The reaction mixture was cooled to 0 °C and aq. HCl (1 N, 15 mL) was added to acidify the solution. After extraction with ethyl acetate, the organic phase was washed with water and brine, and was then concentrated. The crude product was subjected directly to the Procedure B (as for the preparation of **2a**) to provide **2f** (384 mg, 68% yield over two steps): ¹H NMR (500 MHz, CDCl₃) δ 7.93–7.87 (m, 2H), 7.82–7.71 (m, 2H), 5.09 (dd, *J* = 11.2, 4.9 Hz, 1H), 3.76 (s, 3H), 3.31–3.19 (m, 1H), 3.17–3.02 (m, 1H), 1.91 (s, 1H).

N-(Tert-butoxycarbonyl)-O-(prop-2-yn-1-yl)-L-serine methyl ester (2g)^{18,19}



Supplementary Figure 18. Preparation of acetylenic amino acid 2g.

Compound **2g** was prepared from *N*-(*tert*-butoxycarbonyl)-*O*-(prop-2-yn-1-yl)-Lserine (0.72 g, 3 mmol) using Procedure A (as for the preparation of **2a**) (640 mg, 83%): ¹H NMR (500 MHz, CDCl₃) δ 5.38 (d, *J* = 8.8 Hz, 1H), 4.53–4.37 (m, 1H), 4.15 (dd, *J* = 2.4, 0.6 Hz, 2H), 3.96 (dd, *J* = 9.3, 3.2 Hz, 1H), 3.81–3.72 (m, 4H), 2.46 (td, *J* = 2.4, 0.6 Hz, 1H), 1.46 (s, 9H).





Supplementary Figure 19. Preparation of acetylenic amino acid 2h.

A mixture of Boc-L-Tyr-OMe (1.47 g, 5 mmol), propargyl bromide (1.1 mL, 1.3 mmol), and K_2CO_3 (2.0 equiv.) in DMF (10 mL) was stirred at room temperature overnight. The mixture was diluted with ethyl acetate. The organic phase was washed with 0.5 M HCl, water, and brine. After dried over MgSO₄, the solution was concentrated *in vacuo*

to give **2h** as a viscous oil (1.25 g, 75%): ¹H NMR (500 MHz, CDCl₃) δ 7.05 (d, J = 8.5 Hz, 2H), 6.90 (d, J = 8.6 Hz, 2H), 4.96 (d, J = 8.3 Hz, 1H), 4.67 (d, J = 2.4 Hz, 2H), 4.54 (q, J = 6.7 Hz, 1H), 3.71 (s, 1H), 3.03 (qd, J = 14.0, 5.9 Hz, 2H), 2.51 (t, J = 2.4 Hz, 1H), 1.42 (s, 9H).

 N^2 -(((9H-Fluoren-9-yl) methoxy) carbonyl)- N^6 -(hept-6-ynoyl)-L-lysine methyl ester (2i)



Supplementary Figure 20. Preparation of acetylenic amino acid 2i.

To a stirred solution of 6-heptynoic acid (946 mg, 7.5 mmol) in extra dry CH₂Cl₂ (15 mL) was added catalytic amount of DMF (0.05–0.1 mL), followed by addition of (COCl)₂ (1.27 mL, 2.0 equiv.) dropwise under 0 $\$ (Caution: The mixture bubbled vigorously!). The mixture was stirred vigorously for 0.5 h under 0 $\$ and 0.5 h at room temperature. The solvent and residue oxalyl chloride was removed *in vacuo* to give the crude acyl chloride S34, which was used directly in the next step.

To a stirred solution of N-Fmoc-N'-Boc-L-Lys (2.3 g, 5.0 mmol) in dry DMF (10 mL) was added KHCO₃ (1.5 g, 3.0 equiv.) at 0 °C. After vigorously stirring for 20 mins, MeI (0.65 mL, 2.0 equiv.) was added. The mixture was stirred for 7 h at room temperature. After dilution with ethyl acetate (50 mL), the mixture was washed with water and brine and was then dried over anhydrous MgSO₄. After concentration, the crude product was dissolved in dry CH₂Cl₂ (10 mL). TFA (5 mL) was added slowly under 0 °C, and the reaction mixture was stirred for 2 h at room temperature. The solvent was removed *in vacuo* and the residue was dried over oil pump. To a stirred solution of the above residue in dry CH₂Cl₂ (10 mL) was added Et₃N (2.1 mL, 3.0 equiv.) at 0 °C. The crude acyl chloride S34 was dissolved in extra dry CH₂Cl₂ (2 mL) and was injected to this reaction mixture. After stirring for 5 h at room temperature, the reaction mixture was diluted with CH₂Cl₂ (50 mL) and washed with 1 M aq. HCl (2×20 mL), sat. aq. NaHCO₃ (2×20 mL), and brine, successively. After concentration, the residue was purified by column chromatography (petroleum ether/ethyl acetate = 2:1 to 1:1) to yield **2i** (1.5 g, 60%): ¹H NMR (500 MHz, CDCl₃) δ 7.76 (d, J = 7.5 Hz, 2H), 7.60 (t, J = 6.4 Hz, 2H), 7.40 (t, J = 7.4 Hz, 2H), 7.31 (t, J = 7.4 Hz, 2H), 5.67 (t, J = 5.7 Hz, 1H), 5.51 (d, J = 8.2 Hz, 1H), 4.46–4.33 (m, 3H), 4.22 (t, J = 7.1 Hz, 1H), 3.75 (s, 3H), 3.24 (q, J = 6.7 Hz, 2H), 2.23–2.11 (m, 4H), 1.94 (s, 1H), 1.89–1.81 (m, 1H), 1.77–1.65 (m, 3H), 1.57–1.48 (m, 4H), 1.43–1.31 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 173.00, 172.89, 156.19, 143.96, 143.78, 141.39, 127.84, 127.17, 125.16, 120.11, 84.19, 68.75,

67.13, 53.68, 52.57, 47.25, 39.05, 36.18, 32.29, 29.07, 28.02, 24.88, 22.53, 18.26; HRMS (ESI) calcd. for $C_{29}H_{34}N_2O_5Na (M + Na)^+ m/z 513.2360$, found 513.2362.

Methyl N²-(tert-butoxycarbonyl)-N⁴-(prop-2-yn-1-yl)-L-asparaginate (2j)²¹



General Procedure for the preparation of peptides General Procedure I: preparation of dipeptides



Supplementary Figure 21. General Procedure I for the synthesis of dipeptides (2k as an example).

To a stirred suspension of Boc-L-propargylglycine (1.06 g, 5.0 mmol, 1.0 equiv.), methyl L-phenylalaninate hydrochloride (6 mmol, 1.2 equiv.), and HOBt (1.2 equiv.) in dry DMF (15 mL) was added DIPEA (2.61 mL, 3.0 eq) under -10 $^{\circ}$ C. After which, EDCI (1.2 equiv.) was added. The reaction mixture was stirred under -10 $^{\circ}$ C for 0.5 h. Then the mixture was warmed to room temperature and stirred overnight. After dilution with ethyl acetate, the organic phase was washed with water, 1M aq. HCl, and brine thrice. The organic phase was dried over anhydrous MgSO₄, and then concentrated to provide the crude product. The pure product was obtained by column chromatography.

N-(*Tert*-butoxycarbonyl)-L-*C*-propargylglycyl-L-phenylalanine methyl ester (2k)



1.47 g, 79%. ¹H NMR (500 MHz, CDCl₃) δ 7.30–7.23 (m, 3H), 7.14–7.10 (m, 2H), 6.74 (d, *J* = 7.5 Hz, 1H), 5.20 (s, 1H), 4.86 (dt, *J* = 7.6, 5.7 Hz, 1H), 4.27 (s, 1H), 3.71 (s, 3H), 3.21–3.06 (m, 2H), 2.79 (d, *J* = 17.0 Hz, 1H), 2.56 (ddd, *J* = 16.9, 6.7, 2.7 Hz, 1H), 2.02 (t, *J* = 2.7 Hz, 1H),

1.44 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 171.48, 169.90, 155.29, 135.74, 129.37, 128.59, 127.16, 80.50, 79.41, 71.79, 53.48, 52.81, 52.37, 37.89, 28.29, 22.31; HRMS (ESI) calcd. for C₂₀H₂₆N₂O₅Na (M + Na)⁺ m/z 397.1734, found 397.1737.

N-(Tert-butoxycarbonyl)-L-C-propargylglycyl-L-leucine methlyl ester (2l)



1.3 g, 78%. ¹H NMR (500 MHz, CDCl₃) δ 6.70 (s, 1H), 5.25 (s, 1H), 4.63 (td, J = 8.7, 4.7 Hz, 1H), 4.29 (s, 1H), 3.73 (s, 1H), 2.82 (ddd, J = 16.9, 5.4, 2.7 Hz, 1H), 2.60 (ddd, J = 16.9, 6.5, 2.4 Hz, 1H), 2.08 (t, J = 2.6 Hz, 1H), 1.71–1.61 (m, 1H), 1.61–1.52 (m, 1H), 1.47 (s, 1H), 0.93 (dd, J = 6.0, 5.4

5.0 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 173.05, 170.03, 80.77, 79.53, 71.77, 52.95, 52.46, 51.01, 41.79, 28.38, 24.86, 22.94, 22.23, 22.04; HRMS (ESI) calcd. for C₁₇H₂₈N₂O₅Na (M + Na)⁺ m/z 363.1890, found 363.1892.

N-(*Tert*-butoxycarbonyl)-L-*C*-propargylglycyl-L-isoleucine methyl ester (2m)



1.41 g, 83%. ¹H NMR (500 MHz, CDCl₃) δ 6.89 (d, J = 8.6 Hz, 1H), 5.32 (d, J = 7.2 Hz, 1H), 4.59 (dd, J = 8.6, 4.9 Hz, 1H), 4.31 (d, J = 7.2 Hz, 1H), 3.74 (s, 3H), 2.83 (ddd, J = 16.9, 5.4, 2.6 Hz, 1H), 2.60 (ddd, J = 16.9, 6.7, 2.7 Hz, 1H),

2.10 (t, J = 2.6 Hz, 1H), 1.91 (dqd, J = 9.4, 4.8, 2.5 Hz, 1H), 1.47 (s, 9H), 1.47–1.39 (m, 1H), 1.18 (ddq, J = 14.2, 9.0, 7.3 Hz, 1H), 0.92 (dd, J = 8.5, 7.0 Hz, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 171.97, 170.04, 155.43, 80.63, 79.54, 71.75, 56.72, 52.90, 52.19, 38.02, 28.33, 25.14, 22.14, 15.47, 11.62, 11.62; HRMS (ESI) calcd. for C₁₇H₂₈N₂O₅Na (M + Na)⁺ m/z 363.1890, found 363.1894.

N-(*Tert*-butoxycarbonyl)-L-*C*-propargylglycyl-L-proline methyl ester (2n)



1.29 g, 80%, occurring as a mixture of rotamers. ¹H NMR (500 MHz, CDCl₃) δ 5.33 (dd, J = 24.2, 9.0 Hz, 1H), 4.85– 4.62 (m, 1H), 4.58–4.51 (m, 1H), 3.79–3.71 (m, 5H), 2.70– 2065 (m, 1H), 2.61–2.54 (m, 1H), 2.26–2.20 (m, 1H), 2.12– 1.97 (m, 4H), 1.44 & 1.42 (s, 9H). The major rotamer: ¹³C

NMR (125 MHz, CDCl₃) δ 172.22, 169.49, 155.27, 80.07, 78.99, 71.00, 59.01, 52.35, 50.59, 47.21, 29.08, 28.41, 24.95, 22.99; HRMS (ESI) calcd. for C₁₆H₂₄N₂O₅Na (M + Na)⁺ m/z 347.1577, found 347.1579.

O-Benzyl-*N*-(*tert*-butoxycarbonyl)-L-threonyl-L-*C*-propargylglycine methyl ester (2p)



Supplementary Figure 22. Preparation of dipeptide 2p.

To a stirred solution of methyl L-Boc-*C*-propargylglycinate (650 mg, 2.86 mmol) in dry CH_2Cl_2 (6 mL) was added TFA (2 mL) slowly at 0 °C. The mixture was stirred at room temperature to remove the Boc group. After completion (monitored by TLC, 2–4 h), the reaction solution was concentrated and dried *in vacuo*.

To a stirred solution of the above crude product, *O*-benzyl-*N*-(*tert*-butoxycarbonyl)-L-threonine (1.06 g, 1.2 equiv.), and HOBt (1.2 equiv.) in dry DMF (10 mL) was added DIPEA (1.5 mL, 3.0 equiv.) under -10 °C. After which, EDCI (1.2 equiv.) was added. The reaction mixture was stirred under -10 °C for 0.5 h. Then the mixture was warmed to room temperature and stirred overnight. After dilution with ethyl acetate, the organic phase was washed with water, and 1M aq. HCl, and brine thrice. The organic phase was dried over anhydrous MgSO₄, and concentrated to provide the crude product, which was purified by column chromatography to yield **2p** (890 mg, 75%): ¹H NMR (500 MHz, CDCl₃) δ 7.48 (d, *J* = 7.7 Hz, 1H), 7.38–7.31 (m, 4H), 7.30–7.27 (m, 1H), 5.52 (d, *J* = 6.5 Hz, 1H), 4.73 (dt, *J* = 9.1, 4.8 Hz, 1H), 4.62 (q, *J* = 11.4 Hz, 2H), 4.36 (d, *J* = 4.6 Hz, 1H), 4.18 (d, *J* = 3.7 Hz, 1H), 3.76 (s, 3H), 2.81–2.62 (m, 2H), 1.79 (s, 1H), 1.47 (s, 9H), 1.24 (d, *J* = 6.3 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 170.49, 170.08, 155.89, 138.08, 128.47, 127.95, 127.83, 80.29, 78.21, 74.98, 71.88, 71.69, 57.60, 52.82, 50.93, 28.44, 22.44, 15.28.

*N-(Tert-*butoxycarbonyl)-L-tryptophanyl-L-*C*-propargylglycine methyl ester (20)



340 mg, 82%. ¹H NMR (500 MHz, CDCl₃) δ 8.16 (s, 1H), 7.64 (t, *J* = 5.4 Hz, 1H), 7.36 (t, *J* = 5.7 Hz, 1H), 7.23–7.04 (m, 3H), 6.60 (s, 1H), 5.18 (s, 1H), 4.59 (s, 1H), 4.50 (s, 1H), 3.70 (s, 3H), 3.43–3.15 (m, 2H), 2.67 (s, 3H), 1.87 (s, 1H), 1.44 (s, 9H); ¹³C NMR (125 MHz,

CDCl₃) δ 170.31, 136.32, 123.33, 122.36, 119.87, 118.93, 111.27, 110.55, 78.24, 71.68, 55.28, 52.87, 50.86, 28.43, 22.43; HRMS (ESI) calcd. for C₂₂H₂₇N₃O₅Na (M + Na)⁺ m/z 436.1843, found 436.1841.

N-(*Tert*-butoxycarbonyl)-L-phenylalanyl-L-*C*-propargylglycine methyl ester (2q)

H N E O CO₂Me

1.48 g, 79% (5 mmol scale). ¹H NMR (500 MHz, CDCl₃) δ 7.31 (t, J = 7.4 Hz, 2H), 7.2 6 (m, 1H), 7.21 (d, J = 7.6 Hz, 3H), 6.69 (d, J = 7.6 Hz, 1H), 4.97 (s, 1H), 4.67 (s, 1H), 4.42 (s, 1H), 3.76 (s, 3H), 3.10 (d, J = 6.6 Hz, 2H), 2.75 (dt, J =

4.8, 2.4 Hz, 2H), 1.97 (t, J = 2.6 Hz, 1H), 1.42 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 171.17, 170.39, 155.41, 129.44, 128.85, 127.10, 80.45, 78.27, 71.82, 55.67, 52.96, 50.89, 38.35, 28.38, 22.46; HRMS (ESI) calcd. for C₂₀H₂₆N₂O₅Na (M + Na)⁺ m/z 397.1734, found 397.1736.

General Procedure II: preparation of tripeptides



Supplementary Figure 23. General Procedure II for preparation of tripeptides (2r as an example).

To a suspension of Boc-L-alanine (1.13 g, 6.0 mmol, 1.0 eq), methyl L-valinate hydrochloride (6 mmol, 1.0 equiv.), and HOBt (1.2 eq) in dry DMF (15 mL) was added DIPEA (3.13 mL, 3.0 eq) under -10 °C. After which, EDCI (1.2 eq) was added. The reaction mixture was stirred under -10 °C for 0.5 h. Then the mixture was warmed to room temperature and stirred overnight. After dilution with ethyl acetate, the organic phase was washed with water, and 1 N aq. HCl, and brine thrice. The organic phase was dried over anhydrous MgSO₄ and concentrated to provide the crude dipeptide. The crude product was dissolved in dry CH₂Cl₂ (15 mL) and cooled to 0 °C. TFA (5 mL) was added slowly and the mixture was stirred for 2–4 h (monitored by TLC) at room temperature to remove thr Boc group. After completion, the reaction solution was concentrated and dried *in vacuo*.

To a stirred solution of the above crude product, Boc-*L*-propargylglycine (1.06 g, 5 mmol), and HOBt (1.2 eq) in dry DMF (20 mL) was added DIPEA (3.13 mL, 3.0 eq) under -10 $\$ C. After which, EDCI (1.2 eq) was added. The reaction mixture was stirred under -10 $\$ C for 0.5 h. Then the mixture was warmed to room temperature and stirred overnight. After dilution with ethyl acetate, the organic phase was washed with water, 1N aq. HCl, and brine thrice. The organic phase was dried over anhydrous MgSO₄ and concentrated. The pure product was obtained by column chromatography.

N-(*Tert*-butoxycarbonyl)-L-*C*-propargylglycyl-L-alanyl-L-valine methyl ester (2r)

 $\begin{array}{c} 1.83 \text{ g}, 91\%. {}^{1}\text{H} \text{ NMR} (500 \text{ MHz}, \text{CDCl}_{3}) \delta 7.03 (d, \\ J = 7.4 \text{ Hz}, 1\text{H}), 6.88 (d, J = 8.7 \text{ Hz}, 1\text{H}), 5.41 (d, J \\ = 8.1 \text{ Hz}, 1\text{H}), 4.61 (p, J = 7.0 \text{ Hz}, 1\text{H}), 4.52 (dd, J \\ = 8.8, 5.0 \text{ Hz}, 1\text{H}), 4.33 (d, J = 7.0 \text{ Hz}, 1\text{H}), 3.75 (s, \\ \end{array}$

3H), 2.80 (ddd, J = 16.8, 5.6, 2.7 Hz, 1H), 2.60 (ddd, J = 16.9, 6.8, 2.6 Hz, 1H), 2.17 (pd, J = 6.9, 5.0 Hz, 1H), 2.09 (t, J = 2.6 Hz, 1H), 1.46 (s, 9H), 1.40 (d, J = 7.0 Hz, 3H), 0.93 (d, J = 6.9 Hz, 3H), 0.90 (d, J = 6.9 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.25, 171.75, 170.19, 155.41, 80.72, 79.30, 72.08, 57.40, 53.01, 52.32, 49.29, 31.28, 28.39, 22.62, 19.10, 18.35, 17.91; HRMS (ESI) calcd. for C₁₉H₃₁N₃O₆Na (M + Na)⁺ m/z 420.2105, found 420.2108.

*N-(Tert-*butoxycarbonyl)-L-*C*-propargylglycyl-L-tertleucyl-L-alanine methyl ester (2s)



1.82 g, 88%. ¹H NMR (500 MHz, CDCl₃) δ 7.25 (d, J = 9.2 Hz, 1H), 7.15 (d, J = 7.3 Hz, 1H), 5.59 (d, J = 8.4 Hz, 1H), 4.57 (p, J = 7.2 Hz, 1H), 4.46 (d, J = 9.3 Hz, 1H), 4.41 (d, J = 7.2 Hz, 1H), 3.74 (s, 3H), 2.79 (ddd, J = 16.9, 5.7, 2.6 Hz, 1H), 2.65–2.55 (m,

1H), 2.09 (d, J = 2.6 Hz, 1H), 1.46 (s, 9H), 1.39 (d, J = 7.2 Hz, 3H), 1.02 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 173.29, 170.34, 169.76, 155.45, 80.44, 79.52, 77.16, 71.85, 60.52, 53.18, 52.42, 47.98, 35.05, 28.34, 26.61, 22.49, 18.02; HRMS (ESI) calcd. for C₂₀H₃₃N₃O₆Na (M + Na)⁺ m/z 434.2262, found 434.2265.

*N-(Tert-*butoxycarbonyl)-L-*C*-propargylglycyl-L-alanyl-*O-(tert-*butyl)-Lthreonine methyl ester (2t)



Supplementary Figure 24. Preparation of tripeptide 2t.

To a stirred solution of N-Fmoc-L-alanine (1.09 g, 3.5 mmol), methyl O-(tert-butyl)-Lthreoninate hydrochloride (1.1 equiv.), and 3-(diethoxyphosphoryloxy)-1,2,3benzotrizin-4(3H)-one (DEPBT) (1.2 equiv.) in dry CH₂Cl₂ (8 mL) was added DIPEA (1.5 eq) under 0 °C. The mixture was stirred under 0 °C for 0.5 h, then warmed to room temperature slowly and stirred for 4 h. After dilution with CH₂Cl₂, the organic phase was washed with water and brine thrice. The organic phase was dried over anhydrous Na₂SO₄ and concentrated. The crude product was dissolved in MeOH (22 mL), and Et_2NH (11 mL) was added. The mixture was stirred for 1 h at room temperature to remove the Fmoc. After completion, the solvent and residue Et₂NH were removed in *vacuo*. To a stirred solution of the crude product, Boc-L-propargylglycine (1.2 equiv.) and 3-(diethoxyphosphoryloxy)-1,2,3-benzotrizin-4(3H)-one (DEPBT) (1.2 equiv.) in dry DMF (15 mL) was added DIPEA (1.5 eq) at 0 °C. The mixture was stirred under 0 ° for 0.5 h, then warmed to room temperature slowly and stirred for 4 h. After dilution with ethyl acetate, the organic phase was washed with water and brine thrice. The organic phase was dried over anhydrous Na₂SO₄ and concentrated, and the residue was purified by column chromatography to yield 2t (546 mg, 34%): ¹H NMR (500 MHz, CDCl₃) δ 7.00 (d, J = 7.3 Hz, 1H), 6.49 (d, J = 9.2 Hz, 1H), 5.32 (d, J = 8.1 Hz, 1H), 4.58 (p, J = 7.0 Hz, 1H), 4.48 (dd, J = 9.2, 1.8 Hz, 1H), 4.35–4.29 (m, 1H), 4.24 (qd, J = 6.3, 1.8 Hz, 1H), 3.71 (s, 3H), 2.82 (d, J = 17.5 Hz, 1H), 2.59 (ddd, J = 16.9, 6.8, 2.6 Hz, 1H), 2.10 (t, J = 2.6 Hz, 1H), 1.46–1.45 (m, 12H)1.16 (d, J = 6.3 Hz, 3H), 1.12 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.31, 171.05, 169.76, 155.36, 80.65, 79.43, 74.37, 72.03, 67.36, 58.00, 52.95, 52.40, 49.27, 28.45, 28.40, 22.74, 21.11, 19.09; HRMS (ESI) calcd. for C₂₂H₃₇N₃O₇Na (M + Na)⁺ m/z 478.2524, found 478.2526.

Methyl *N*-(9-fluorenylmethoxycarbonyl)-*O*-(*tert*-butyl)-L-threonyl-L-*C*-propargylglycyl-L-alloisoleucinate (2u)



Supplementary Figure 25. Preparation of tripeptide 2u.

To a stirred solution of methyl (tert-butoxycarbonyl)-L-propargylglycyl-Lalloisoleucinate (**2m**) (578 mg, 1.7 mmol) in dry CH₂Cl₂ (6 mL) was added TFA (3 mL) slowly at 0 $^{\circ}$ C. The mixture was stirred for 2–4 h at room temperature to remove the Boc group (monitored by TLC). After completion, the reaction solution was concentrated and dried over oil pump to give the crude **S37**.

To a stirred solution of the above crude S37, N-(9-fluorenylmethoxycarbonyl)-O-(tert-butyl)-L-threonine (0.79 g, 2.0 mmol) and 3-(diethoxyphosphoryloxy)-1,2,3benzotrizin-4(3H)-one (DEPBT) (1.2 equiv.) in dry CH₂Cl₂ (8 mL) was added DIPEA (1.5 eq) under 0 \mathbb{C} . The mixture was stirred under 0 \mathbb{C} for 0.5 h, then warmed to room temperature slowly and stirred for 2 h. After dilution with CH₂Cl₂, the organic phase was washed with water and brine thrice. The organic phase was dried over anhydrous Na₂SO₄ and concentrated, and the residue was purified by column chromatography to yield **2u** (154 mg, 25%): ¹H NMR (500 MHz, CDCl₃) δ 7.76 (d, J = 7.5 Hz, 2H), 7.61 (dd, J = 10.2, 7.9 Hz, 3H), 7.40 (t, J = 7.3 Hz, 2H), 7.32 (t, J = 7.4 Hz, 2H), 6.88 (d, J = 8.8 Hz, 1H), 6.00 (d, J = 5.4 Hz, 1H), 4.73 (dt, J = 8.8, 5.5 Hz, 1H), 4.62 (dd, J = 8.7, 5.1 Hz, 1H), 4.40 (d, J = 7.2 Hz, 2H), 4.28 (t, J = 4.7 Hz, 1H), 4.25–4.17 (m, 2H), 3.72 (s, 3H), 2.96 (ddd, J = 16.9, 5.1, 2.6 Hz, 1H), 2.59 (ddd, J = 17.0, 6.0, 2.6 Hz, 1H), 2.07 (t, J = 2.6 Hz, 1H), 1.97 - 1.83 (m, 1H), 1.45 (tdt, J = 10.7, 7.9, 3.1 Hz, 1H), 1.32 (s, 9H),1.28–1.17 (m, 1H), 1.12 (d, J = 6.4 Hz, 3H), 0.91 (t, J = 7.3 Hz, 6H); ¹³C NMR (125) MHz, CDCl₃) δ 171.83, 169.53, 169.41, 156.06, 143.96, 143.74, 141.43, 141.40, 127.85, 127.17, 127.15, 125.24, 125.19, 120.13, 120.10, 79.41, 76.02, 71.99, 67.15, 66.78, 58.87, 56.86, 52.19, 51.71, 47.26, 37.88, 28.27, 25.16, 22.24, 17.08, 15.55, 11.60; HRMS (ESI) calcd. for $C_{35}H_{45}N_3O_7Na (M + Na)^+ m/z 642.3150$, found 642.3152.


2.3 Optimization of Reaction Conditions with Mannose Donors



Supplementary Figure 26. Optimization of the base. The yield was determined by ¹H NMR using CH_2Br_2 as an internal standard. dtbbpy = 4,4'-di-*tert*-butyl-2,2'-bipyridine, DMI = 1,3-dimethyl-2-imidazolidinone.

2-(2,3,4,6-Tetra-O-benzoyl-α-D-mannopyranosyl)-1-hexene (3a-1)

¹H NMR (500 MHz, CDCl₃) δ 8.09 (t, J = 8.0 Hz, 4H), 7.96 (d, J = 7.2 Hz, 2H), 7.87 (d, J = 7.2 Hz, 2H), 7.59 (q, J = 7.6 Hz, 2H), 7.51 (t, J = 7.4 Hz, 1H), 7.47–7.40 (m, 5H), 7.36 (t, J = 7.8 Hz, 2H), 7.29 (t, J = 7.8 Hz, 2H), 6.23 (dd, J = 3.1, 1.9 Hz, 1H), 6.08 (t, J = 9.9 Hz, 1H), 5.78 (dd, J = 9.9, 3.1 Hz, 1H), 5.58 (d, J = 2.0 Hz, 1H), 5.41 (q, J = 1.4 Hz, 1H), 4.73 (d, J = 1.9 Hz, 1H), 4.65 (dd, J = 12.0, 2.5 Hz, 1H), 4.52 (dd, J = 12.0, 5.9 Hz, 1H), 4.16 (ddd, J = 9.9, 5.9, 2.5 Hz, 1H), 2.29 (ddd, J = 15.3, 9.8, 5.4 Hz, 1H), 2.19 (ddd, J = 15.5, 9.8, 6.2 Hz, 1H), 1.59–1.52 (m, 1H), 1.48 (dddd, J = 13.1, 9.6, 6.5, 3.1 Hz, 1H), 1.31 (tt, J = 14.2, 7.4 Hz, 2H), 0.90 (t, J = 7.3 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 166.32, 166.14, 165.80, 165.58, 143.98, 133.58, 133.48, 133.39, 133.22, 129.97, 129.90, 129.84, 129.68, 129.08, 128.65, 128.56, 128.51, 128.47, 115.10, 77.19, 71.32, 71.24, 70.34, 67.62, 63.69, 33.14, 29.74, 22.58, 14.07; HRMS (ESI) calcd. for C₄₀H₃₈O₉Na (M + Na)⁺ m/z 685.2408, found 385.2409.

2,3,4,6-Tetra-O-benzoyl-D-glucal 4a²³

¹H NMR (500 MHz, CDCl₃) δ 8.07–7.97 (m, 6H), 7.59–7.51 (m, 3H), 7.46–7.36 (m, 6H), 6.61 (dd, J = 6.2, 1.2 Hz, 1H), 5.85–5.75 (m, 1H), 5.71 (ddd, J = 5.4, 3.7, 1.1 Hz, 1H), 5.12 (dd, J = 6.2, 3.5 Hz, 1H), 4.73–4.63 (m, 3H).



Supplementary Figure 27. Optimization of the solvent. The yield was determined by ¹H NMR using CH₂Br₂ as an internal standard. dtbbpy = 4,4'-di-*tert*-butyl-2,2'-bipyridine, DME = 1,2-dimethoxyethane, DMAc = N,N-dimethylacetamide.



Supplementary Figure 28. Further optimization of the solvent. The yield was determined by ¹H NMR using CH₂Br₂ as an internal standard. dtbbpy = 4,4'-di-*tert*-butyl-2,2'-bipyridine, DME = 1,2-dimethoxyethane, DMAc = N,N-dimethylacetamide.

BZO BZO BZO BZO Br	+	NiCl ₂ (DME) (10 mol%) dtbbpy (12 mol%), (<i>R</i>)-BINAP (x mol%) (EtO) ₂ MeSiH (2.0 equiv.), Na ₂ CO ₃ (2.0 equiv.) DME:DMAc (10:1) 33–35 °C, Ar, 16 h				BzO BzO BzO BzO
iu		Entry	x	Yiel	d (%)	3a-1
		1	0	42	31	
		2	5	67	13	
		3	10	72	13	
		4	15	43	38	
		5	20	56	26	
		6 ^[a]	15	75	7	

Supplementary Figure 29. Optimization of the equivalents of (*R*)-BINAP. The yield was determined by ¹H NMR using CH₂Br₂ as an internal standard. ^[a]22 h. dtbbpy = 4,4'-di-*tert*-butyl-2,2'-bipyridine, DME = 1,2-dimethoxyethane, DMAc = N,N-dimethylacetamide.



Supplementary Figure 30. Optimization of the P-based additive. The yield was determined by ¹H NMR using CH₂Br₂ as an internal standard. ^[a]Without dtbbpy. ^[b]Isolated yield. dtbbpy = 4,4'-di-*tert*-butyl-2,2'-bipyridine, DME = 1,2-dimethoxyethane, DMAc = N,N-dimethylacetamide.



Supplementary Figure 31. Optimized Conditions I and control experiments for the synthesis of **3aa**. The yield was determined by ¹H NMR using CH₂Br₂ as an internal standard. ^[a]Isolated yield. ^[b]Average of two isolated yields for entry 6 and entry 8.

2,3,4,6-Tetra-O-benzoyl-α/β-D-mannopyranose (1a-OH)

¹H NMR (500 MHz, CD₃Cl) δ 8.12 (d, *J* = 7.3 Hz, 2H), 8.02 (d, *J* = 7.4 Hz, 2H), 7.96 (d, *J* = 7.4 Hz, 2H), 7.85 (d, *J* = 7.4 Hz, 2H), 7.56 (t, *J* = 7.0 Hz, 2H), 7.49 (t, *J* = 7.4 Hz, 1H), 7.41 (q, *J* = 7.8 Hz, 3H), 7.35 (q, *J* = 7.6 Hz, 4H), 7.26 (t, *J* = 7.3 Hz, 2H), 6.19 (t, *J* = 10.1 Hz, 1H), 6.02 (dd, *J* = 10.2, 3.2 Hz, 1H), 5.75 (dd, *J* = 3.2, 1.9 Hz, 1H), 5.54 (dd, *J* = 3.9, 1.9 Hz, 1H), 4.77 (dd, *J* = 12.2, 2.7 Hz, 1H), 4.68 (dt, *J* = 10.1, 3.1 Hz, 1H), 4.44 (dd, *J* = 12.2, 3.5 Hz, 1H), 4.30 (d, *J* = 4.1 Hz, 1H).

2.4 Optimization of Reaction Conditions with Glucosamine Donors (GlcN)

We found that the judicious choice of protecting groups (i.e., N-Phth and O-Bz/Ac) on the GlcN substrates was critical for the present coupling reaction.



Supplementary Figure 32. Optimization of the *N*-ligand. The yield was determined by ¹H NMR using CH₂Br₂ as an internal standard. DME = 1,2-dimethoxyethane, DMAc = N,N-dimethylacetamide.

2-(3,4,6-Tri-*O***-acetyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)-1-hexene (3b-1)** Colorless syrup. ¹H NMR (500 MHz, CDCl₃) δ 7.86–7.79 (m, 2H), 7.76–7.70 (m, 2H), 5.87 (dd, J = 10.4, 9.1 Hz, 1H), 5.18 (t, J = 9.6 Hz, 1H), 4.94 (s, 1H), 4.84 (d, J = 10.7 Hz, 2H), 4.45 (t, J = 10.4 Hz, 1H), 4.27 (dd, J = 12.2, 4.5 Hz, 1H), 4.18 (dd, J = 12.2, 2.2 Hz, 1H), 3.87 (ddd, J = 10.1, 4.5, 2.4 Hz, 1H), 2.18–2.08 (m, 4H), 2.04 (s, 3H), 2.00 (dd, J = 10.1, 5.3 Hz, 1H), 1.86 (s, 3H), 1.41 (ddt, J = 16.5, 12.1, 6.0 Hz, 1H), 1.30 (qt, J = 12.6, 5.6 Hz, 3H), 0.87 (t, J = 7.2 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 170.93, 170.34, 169.79, 167.99, 167.43, 144.79, 134.49, 134.30, 131.75, 131.14, 123.69, 115.02, 79.59, 75.72, 71.89, 69.41, 62.58, 52.93, 29.98, 29.75, 22.56, 20.95, 20.84, 20.66, 14.07; HRMS (ESI) calcd. for C₂₆H₃₁NO₉Na (M + Na)⁺ m/z 524.1891, found 524.1895.

3,4,6-Tri-O-acetyl-1,5-anhydro-2-deoxy-2-phthalimido-D-glucitol (5a)²⁴

¹H NMR (500 MHz, CDCl₃) δ 7.85 (dd, J = 5.5, 3.0 Hz, 2H), 7.74 (dd, J = 5.5, 3.1 Hz, 2H), 5.84 (dd, J = 10.4, 9.1 Hz, 1H), 5.13 (dd, J = 10.1, 9.1 Hz, 1H), 4.56 (ddd, J = 11.5, 10.5, 5.4 Hz, 1H), 4.35 (t, J = 11.3 Hz, 1H), 4.27 (dd, J = 12.3, 4.8 Hz, 1H), 4.18 (dd, J = 12.3, 2.2 Hz, 1H), 3.95 (dd, J = 11.2, 5.5 Hz, 1H), 3.79 (ddd, J = 10.1, 4.8, 2.2 Hz, 1H), 2.13 (s, 3H), 2.04 (s, 3H), 1.87 (s, 3H).



+

<u></u>____Bu

1

2

3

4

5

6



5

6

Ar, 24 h. 20 mol% for L1, L2, L3; 10 mol% for L4, L5

Using (EtO)₂MeSiH (2.0 equiv), 33-35 °C,

 L_4

 L_5

65

58

8

17

Using PMHS (2.5 equiv), r.t. (18-23 °C), Ar, 24 h. 20 mol% for L1, L2, L3; 10 mol% for L4, L5

 L_4

 L_5

68 (70)^[a]

70 (70)^[a] trace

trace

OAc

0

5a

NPhth



Supplementary Figure 33. Optimization of the P-based additive and silanes. The yield was determined by ¹H NMR using CH₂Br₂ as an internal standard. ^[a]Average of two isolated yields for entry 5 and entry 6. dtbbpy = 4,4'-di-tert-butyl-2,2'-bipyridine, PMHS = poly(methylhydrosiloxane).



Supplementary Figure 34. Control experiments using **1b** as glycosyl donor. The yield was determined by ¹H NMR using CH₂Br₂ as an internal standard. dtbbpy = 4,4'-di*tert*-butyl-2,2'-bipyridine, PMHS = poly(methylhydrosiloxane).

$\begin{array}{l} 2\text{-}(3,4,6\text{-}Tri\text{-}\textit{O}\text{-}benzoyl\text{-}2\text{-}deoxy\text{-}2\text{-}phthalimido-\beta\text{-}D\text{-}glucopyranosyl)\text{-}1\text{-}hexene~(3b-2) \end{array}$

¹H NMR (500 MHz, CDCl₃) δ 8.05 (d, J = 7.2 Hz, 2H), 7.91 (d, J = 7.2 Hz, 2H), 7.86 (d, J = 7.8 Hz, 1H), 7.76 (d, J = 7.2 Hz, 2H), 7.68 (dt, J = 6.6, 3.1 Hz, 2H), 7.64 (d, J = 7.2 Hz, 1H), 7.55 (t, J = 7.4 Hz, 1H), 7.47 (t, J = 7.4 Hz, 1H), 7.41 (q, J = 8.0 Hz, 3H), 7.32 (t, J = 7.8 Hz, 2H), 7.25 (t, J = 7.8 Hz, 2H), 6.36 (dd, J = 10.4, 9.3 Hz, 1H), 5.73 (t, J = 9.6 Hz, 1H), 5.07 (d, J = 10.4 Hz, 1H), 5.01 (s, 1H), 4.86 (d, J = 1.6 Hz, 1H), 4.70 (t, J = 10.5 Hz, 1H), 4.66 (dd, J = 12.2, 3.1 Hz, 1H), 4.47 (dd, J = 12.1, 4.5 Hz, 1H), 4.25 (ddd, J = 10.1, 4.5, 3.1 Hz, 1H), 2.20 (ddd, J = 15.7, 10.0, 5.4 Hz, 1H), 2.04 (ddd, J = 15.9, 10.0, 5.7 Hz, 1H), 1.43 (m, 1H), 1.40–1.32 (m, 1H), 1.27 (m, 2H), 0.83 (t, J = 7.3 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 168.18, 167.42, 166.28, 165.85, 165.38, 144.88, 134.35, 134.16, 133.44, 133.29, 133.13, 131.74, 131.13, 129.93, 129.86, 129.84, 129.05, 128.84, 128.46, 128.45, 128.37, 123.66, 123.63, 115.07, 79.79, 75.91, 72.31, 70.46, 63.36, 53.25, 30.05, 29.91, 22.57, 14.02; HRMS (ESI) calcd. for C₄₁H₃₇NO₉Na (M + Na)⁺ m/z 710.2361, found 710.2364.



Supplementary Figure 35. Optimized Conditions II and control experiments for the synthesis of **3ba**. The yield was determined by ¹H NMR using CH₂Br₂ as an internal standard. ^[a]Isolated yield.

3,4,6-Tri-O-benzoyl-2-phthalimido-glucal (4b)

¹H NMR (500 MHz, CDCl₃) δ 8.15 (d, J = 8.3 Hz, 2H), 8.02 (d, J = 8.4 Hz, 2H), 7.90 (d, J = 8.3 Hz, 2H), 7.87–7.82 (m, 2H), 7.74–7.68 (m, 2H), 7.60 (t, J = 7.4 Hz, 1H), 7.55 (t, J = 7.4 Hz, 1H), 7.48 (t, J = 6.4 Hz, 3H), 7.41 (t, J = 7.7 Hz, 2H), 7.35 (t, J = 7.7 Hz, 2H), 6.95 (s, 1H), 6.00–5.95 (m, 1H), 5.86 (dt, J = 3.5, 1.9 Hz, 1H), 5.04–4.98 (m, 1H), 4.97–4.90 (m, 1H), 4.80 (dd, J = 11.3, 5.3 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 167.82, 166.14, 165.72, 165.17, 148.75, 134.40, 133.76, 133.52, 133.38, 131.87, 130.23, 129.91, 129.87, 129.45, 129.13, 129.01, 128.70, 128.60, 128.55, 123.85, 110.12, 105.52, 74.32, 67.67, 66.27, 61.70; HRMS (ESI) calcd. for C₃₅H₂₅NO₉Na (M + Na)⁺ m/z 626.1422, found 626.1424.

3,4,6-Tri-*O*-benzoyl-1,5-anhydro-2-deoxy-2-phthalimido-D-glucitol (5b)

¹H NMR (500 MHz, CDCl₃) δ 8.08 (dd, J = 8.1, 1.4 Hz, 2H), 7.92 (dd, J = 8.1, 1.5 Hz, 2H), 7.80 (s, 2H), 7.79–7.73 (m, 2H), 7.69 (dd, J = 5.5, 3.0 Hz, H), 7.56 (t, J = 7.4 Hz, 1H), 7.51–7.39 (m, 4H), 7.34 (t, J = 7.8 Hz, 2H), 7.29–7.24 (m, 2H), 6.31 (dd, J = 10.5, 9.2 Hz, 1H), 5.69 (t, J = 9.6 Hz, 1H), 4.81 (td, J = 10.9, 5.4 Hz, 1H), 4.65 (dd, J = 12.2, 2.7 Hz, 1H), 4.55 (t, J = 11.4 Hz, 1H), 4.46 (dd, J = 12.2, 5.0 Hz, 1H), 4.17 (ddd, J = 10.1, 4.9, 2.7 Hz, 1H), 4.08 (dd, J = 11.3, 5.4 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 166.39, 165.96, 165.38, 134.41, 133.51, 133.34, 133.24, 131.53, 129.97, 129.96, 129.88, 129.80, 129.03, 128.83, 128.52, 128.41, 123.74, 76.96, 72.23, 70.26, 65.88, 63.34, 50.71; HRMS (ESI) calcd. for C₃₅H₂₇NO₉Na (M + Na)⁺ m/z 628.1578, found 628.1581.

[1,1'-binaphthalene]-2,2'-diylbis(di-p-tolylphosphine oxide) (di-oxidized (*R*)-Tol-BINAP(O)₂)²⁵

¹H NMR (500 MHz, CDCl₃) δ 7.84–7.74 (m, 4H), 7.48 (dd, *J* = 12.0, 8.0 Hz, 4H), 7.43 (dd, *J* = 11.6, 8.6 Hz, 2H), 7.37 (ddd, *J* = 8.1, 5.9, 2.0 Hz, 2H), 7.27 (dd, *J* = 11.3, 8.1, 4H), 7.03 (d, *J* = 6.4 Hz, 4H), 6.98 (d, *J* = 6.1 Hz, 4H), 6.89-6.85 (m, 4H), 2.32 (s, 6H), 2.27 (s, 6H); ³¹P NMR (162 MHz, CDCl₃) δ 29.20.

2.5 Ni-Catalyzed synthesis of vinyl C-glycosyl amino acids and glycopeptides

General Procedure A (unless otherwise stated)



To an oven-dried 10 mL Schlenk tube (Titan, TF891910) containing a Teflon coated magnetic stirring bar were added glycosyl bromide **1** (0.1 mmol), NiCl₂(DME) (10 mol%), dtbbpy (12 mol%), PPh₃ (20 mol%), and Na₂CO₃ (2.5 equiv). The tube was sealed with a rubber cap then with parafilm and evacuated then refilled with Ar for at least five cycles. The acetylenic amino acid and peptide derivative 2 was dissolved in solvent (DME/DMAc = 1:1, 1.0 mL) and injected into the reaction tube (this substrate was added directly with glycosyl bromide if it was solid). When stirring, (EtO)₂MeSiH (40 μ L, 2.5 equiv.) was injected via microliter syringe. Otherwise noted, the tube was moved to an oil bath preheated to 33–35 °C and kept stirring for 36 h. The reaction mixture was diluted with CH₂Cl₂ (20 mL) and filtered. After concentration, the residue was purified by column chromatography on silica gel or preparative TLC to afford the desired product **3**.

General Procedure B (unless otherwise stated)

To an oven-dried 10 mL Schlenk tube (Titan, TF891910) containing a Teflon coated magnetic stirring bar were added glycosyl bromide **1** (0.1 mmol), NiCl₂(DME) (10 mol%), dtbbpy (12 mol%), (*R*)-Tol-BINAP (10 mol%), and Na₂CO₃ (2.5 equiv). The tube was sealed with a rubber cap then with parafilm and evacuated then refilled with Ar for at least five cycles. The acetylenic amino acid and peptide derivative **2** was dissolved in THF (1.0 mL) and injected into the reaction tube (this substrate was added directly with glycosyl bromide if it was solid). When stirring, PMHS (32 μ L, 2.5 equiv.) was injected via microliter syringe. Otherwise noted, the tube was kept stirring under 30 °C for 36 h. The reaction mixture was diluted with CH₂Cl₂ (20 mL) and filtered. After concentration, the residue was purified by flash column chromatography on silica gel or preparative TLC to afford the desired product **3**.

N-(*Tert*-butoxycarbonyl)-3-(1-(2,3,4,6-tetra-O-benzoyl- α -D-mannopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (3aa)



General Procedure A. White foam, 62.5 mg, 77%. $[\alpha]_D^{25}$ –57.5 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.09 (d, *J* = 7.4 Hz, 2H), 8.05 (d, *J* = 7.3 Hz, 2H), 7.96 (d, *J* = 7.3 Hz, 2H), 7.86 (d, *J* = 7.3 Hz, 2H), 7.59 (q, *J* = 7.6 Hz, 2H), 7.51 (t, *J* = 7.4 Hz, 2H), 7.44 (dq, *J* = 15.6,

7.8 Hz, 5H), 7.36 (t, J = 7.8 Hz, 2H), 7.30 (t, J = 7.8 Hz, 2H), 6.21 (t, J = 2.6 Hz, 1H), 6.07 (t, J = 9.6 Hz, 1H), **5.74 (s, 1H, vinyl H)**, 5.72 (dd, J = 9.8, 3.1 Hz, 1H), **5.51 (s, 1H, vinyl H)**, 5.11 (d, J = 8.2 Hz, 1H), **4.75 (s, 1H, anomeric H)**, 4.66 (dd, J = 12.1, 2.6 Hz, 1H), 4.55 (dd, J = 12.3, 5.5 Hz, 2H), 4.14 (ddd, J = 9.7, 5.5, 2.7 Hz, 1H), 3.69 (s, 3H), 2.79-2.65 (m, 2H), 1.42 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.94, 166.29, 166.16, 165.67, 165.51, 139.37, 133.62, 133.54, 133.47, 133.25, 129.99, 129.92, 129.87, 129.85, 129.59, 129.07, 129.02, 128.68, 128.61, 128.59, 128.52, 119.38, 80.25, 77.16, 71.65, 71.08, 69.78, 67.41, 63.45, 52.51, 52.40, 36.20, 28.39; HRMS (ESI) calcd. for C₄₅H₃₅NO₁₃Na (M + Na)⁺ m/z 830.2783, found 830.2786.

N-Benzyloxycarbonyl-3-(1-(2,3,4,6-tetra-O-benzoyl- α -D-mannopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (3ab)



General Procedure A. White foam, 65.7 mg, 78%. $[\alpha]_D^{25}$ –52.1 (*c* 0.7, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.08 (d, *J* = 7.4 Hz, 2H), 8.05 (dd, *J* = 8.2, 1.1 Hz, 2H), 7.96 (dd, *J* = 8.3, 1.2 Hz, 2H), 7.87 (dd, *J* = 8.3, 1.1 Hz, 2H), 7.59 (t, *J* = 7.5 Hz, 1H), 7.54 (t, *J* = 7.2 Hz, 1H),

7.51 (t, J = 6.9 Hz, 1H), 7.46 (t, J = 7.5 Hz, 1H), 7.44–7.39 (m, 4H), 7.36 (t, J = 7.8 Hz, 2H), 7.34–7.26 (m, 7H), 6.18 (s, 1H), 6.06 (t, J = 9.5 Hz, 1H), 5.71 (d, J = 6.1 Hz, 2H), 5.48 (s, 1H), 5.42 (d, J = 8.2 Hz, 1H), 5.08 (s, 2H), 4.78 (d, J = 2.1 Hz, 1H), 4.68–4.54 (m, 3H), 4.14 (ddd, J = 11.3, 5.1, 2.4 Hz, 1H), 3.69 (s, 3H), 2.79 (dd, J = 14.6, 7.1 Hz, 1H), 2.71 (dd, J = 14.6, 6.5 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 172.52, 166.27, 166.10, 165.66, 165.51, 155.82, 139.21, 136.26, 133.63, 133.54, 133.54, 133.48, 133.25, 129.98, 129.91, 129.85, 129.82, 129.53, 129.03, 128.99, 128.67, 128.61, 128.58, 128.27, 128.18, 119.47, 76.62, 71.73, 70.95, 69.71, 67.42, 67.19, 63.36, 52.87, 52.59, 36.24, 29.38; HRMS (ESI) calcd. for C₄₈H₄₃NO₁₃Na (M + Na)⁺ m/z 864.2627, found 864.2628.

N-(9-Fluorenylmethoxycarbonyl)-3-(1-(2,3,4,6-tetra-*O*-benzoyl-α-D-mannopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (3ac)



7.46 (t, *J* = 7.5 Hz, 1H), 7.42–7.33 (m, 8H), 7.32–7.26 (m, 4H), 6.20 (t, *J* = 2.8 Hz, 1H), 6.07 (t, *J* = 9.5 Hz, 1H), 5.78–5.69 (m, 2H), 5.52–5.47 (m, 2H), 4.81 (s, 1H), 4.70–4.62

(m, 2H), 4.59 (dd, J = 12.3, 5.8 Hz, 1H), 4.40–4.31 (m, 2H), 4.23–4.14(m, 2H), 3.70 (s, 3H), 2.80 (dd, J = 14.7, 7.2 Hz, 1H), 2.73 (dd, J = 14.6, 6.3 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 172.54, 166.29, 166.11, 165.69, 165.52, 155.85, 143.92, 143.85, 141.38, 139.27, 133.63, 133.54, 133.49, 133.26, 129.97, 129.90, 129.86, 129.82, 129.51, 129.02, 128.98, 128.67, 128.59, 128.58, 128.53, 127.80, 127.19, 125.23, 120.05, 119.48, 76.65, 71.79, 70.94, 69.73, 67.46, 67.25, 63.39, 52.94, 52.63, 47.22, 36.26; HRMS (ESI) calcd. for C₅₅H₄₇NO₁₃Na (M + Na)⁺ m/z 952.2940, found 952.2941.

N-(*Tert*-butoxycarbonyl)-3-(1-(2,3,4,6-tetra-*O*-benzoyl-α-D-mannopyranosyl)eth-1-en-1-yl)-L-alanine benzyl ester (3ad)



General Procedure A. White foam, 70.9 mg, 80%. [α] $_{D}^{25}$ -69.7 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.10–8.04 (m, 4H), 7.95 (d, *J* = 7.3 Hz, 2H), 7.86 (d, *J* = 7.2 Hz, 2H), 7.58 (dt, *J* = 15.4, 7.5 Hz, 2H), 7.50 (t, *J* = 7.4 Hz, 1H), 7.46–7.39 (m, 5H), 7.38–7.33 (m, 3H),

7.32–7.27 (m, J = 8.1, 7.7 Hz, 6H), 6.19 (t, J = 2.6 Hz, 1H), 6.08 (t, J = 9.7 Hz, 1H), 5.70 (dd, J = 9.8, 3.1 Hz, 1H), 5.65 (s, 1H), 5.43 (s, 1H), 5.17–5.11 (m, J = 6.7 Hz, 3H), 4.76 (s, 1H), 4.64 (dd, J = 12.2, 2.7 Hz, 1H), 4.52 (dd, J = 12.1, 5.3 Hz, 1H), 4.11 (ddd, J = 9.6, 5.3, 2.6 Hz, 1H), 2.76 (dd, J = 14.5, 7.0 Hz, 1H), 2.69 (dd, J = 14.5, 6.6 Hz, 1H), 1.42 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.20, 166.22, 166.09, 165.56, 165.46, 155.22, 139.07, 135.25, 133.58, 133.51, 133.42, 133.17, 129.96, 129.92, 129.87, 129.83, 129.82, 129.56, 129.02, 128.99, 128.65, 128.54, 128.47, 119.42, 80.19, 76.91, 71.57, 71.07, 69.70, 67.37, 67.30, 63.37, 52.40, 36.14, 28.36; HRMS (ESI) calcd. for C₅₁H₄₉NO₁₃Na (M + Na)⁺ m/z 906.3096, found 906.3098.

N-(*Tert*-butoxycarbonyl)-3-(1-(2,3,4,6-tetra-*O*-benzoyl-α-D-mannopyranosyl)eth-1-en-1-yl)-L-alanine *tert*-butyl ester (3ae)



General Procedure A. White foam, 70 mg, 82%. $[\alpha]_D^{25}$ –58.8 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.10 (d, *J* = 7.3 Hz, 2H), 8.06 (d, *J* = 7.5 Hz, 2H), 7.95 (d, *J* = 7.7 Hz, 2H), 7.86 (d, *J* = 8.1 Hz, 2H), 7.58 (q, *J* = 7.7 Hz, 2H), 7.50 (t, *J* = 7.4 Hz, 1H), 7.42 (h, *J* = 8.8,

8.0 Hz, 5H), 7.35 (t, J = 7.7 Hz, 2H), 7.29 (t, J = 7.7 Hz, 2H), 6.24 (t, J = 2.5 Hz, 1H), 6.11 (t, J = 9.8 Hz, 1H), 5.77–5.70 (m, 2H), 5.51 (s, 1H), 5.12 (d, J = 8.2 Hz, 1H), 4.81 (s, 1H), 4.69 (dd, J = 12.1, 2.5 Hz, 1H), 4.55 (dd, J = 11.9, 4.7 Hz, 1H), 4.43 (q, J = 7.2 Hz, 1H), 4.14 (ddd, J = 10.0, 5.2, 2.6 Hz, 1H), 2.78 (dd, J = 14.6, 6.3 Hz, 1H), 2.67 (dd, J = 14.4, 7.0 Hz, 1H), 1.43 (s, 18H); ¹³C NMR (125 MHz, CDCl₃) δ 171.42, 166.27, 166.18, 165.55, 165.47, 155.19, 139.51, 133.56, 133.47, 133.42, 133.17, 129.93, 129.89, 129.87, 129.84, 129.60, 129.07, 129.02, 128.64, 128.55, 128.48, 119.32, 82.62, 79.90, 76.71, 71.46, 71.20, 69.84, 67.34, 63.47, 52.68, 36.47, 28.38, 27.94; HRMS (ESI) calcd. for C₄₈H₃₉NO₁₃Na (M + Na)⁺ m/z 872.3253, found 871.3258.

N,*N*-Phthaloyl-3-(1-(2,3,4,6-tetra-*O*-benzoyl-α-D-mannopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (3af)



General Procedure A. White foam, 61.2 mg, 73%. $[\alpha]_D^{25}$ -82.0 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.07 (d, *J* = 7.0 Hz, 2H), 8.02 (d, *J* = 6.9 Hz, 2H), 7.98 (d, *J* = 7.0 Hz, 2H), 7.87 (d, *J* = 7.0 Hz, 2H), 7.80 (dt, *J* = 5.5, 2.8 Hz, 2H), 7.68 (dt, *J* = 5.6, 2.8 Hz, 2H), 7.59

(t, J = 7.4 Hz, 1H), 7.57–7.49 (m, 2H), 7.46 (t, J = 7.4 Hz, 2H), 7.43–7.35 (m, 5H), 7.30 (t, J = 7.1 Hz, 2H), 6.14 (q, J = 2.7 Hz, 1H), 6.06 (t, J = 9.0 Hz, 1H), 5.71 (d, J = 9.5 Hz, 1H), 5.66 (s, 1H), 5.47 (s, 1H), 5.24 (ddd, J = 10.4, 4.9, 2.3 Hz, 1H), 4.74 (s, 1H), 4.63 (dt, J = 12.2, 2.6 Hz, 1H), 4.57 (ddd, J = 12.2, 5.4, 2.4 Hz, 1H), 4.22 (ddt, J = 8.4, 5.3, 2.6 Hz, 1H), 3.70 (s, 3H), 3.39–3.22 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 169.28, 167.76, 166.29, 166.03, 165.63, 165.46, 139.43, 134.29, 133.60, 133.50, 133.45, 133.10, 131.79, 129.96, 129.94, 129.90, 129.86, 129.52, 129.04, 128.99, 128.63, 128.58, 128.52, 128.50, 123.72, 118.24, 77.32, 71.60, 70.93, 69.72, 67.32, 63.29, 53.10, 50.86, 32.57; HRMS (ESI) calcd. for C₄₈H₃₉NO₁₃Na (M + Na)⁺ m/z 860.2314, found 860.2316.

N-(*Tert*-butoxycarbonyl)-*O*-(2-(2,3,4,6-tetra-*O*-benzoyl-α-D-mannopyranosyl) prop-2-en-1-yl)-L-serine methyl ester (3ag)



General Procedure A. White foam, 59.8 mg, 71%. $[\alpha]_D^{25}$ –37.0 (*c* 1.2, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.08 (t, *J* = 8.8 Hz, 4H), 7.96 (d, *J* = 7.9 Hz, 2H), 7.87 (d, *J* = 7.9 Hz, 2H), 7.59 (q, *J* = 7.3, 6.8 Hz, 2H), 7.51 (t, *J* = 7.1 Hz, 1H), 7.47–

7.4 (m, 5H), 7.36 (t, J = 7.0 Hz, 2H), 7.29 (t, J = 7.0 Hz, 2H), 6.17 (s, 1H), 6.09 (t, J = 10.5 Hz, 1H), 5.74–5.68 (m, 2H), 5.64–5.58 (m, 2H), 4.79 (s, 1H), 4.64 (d, J = 11.9 Hz, 1H), 4.55 (ddd, J = 12.0, 5.6, 1.9 Hz, 1H), 4.46 (d, J = 7.5 Hz, 1H), 4.21 (ddt, J = 10.1, 5.0, 2.1 Hz, 1H), 4.16 (d, J = 12.8 Hz, 1H), 4.10 (d, J = 12.6 Hz, 1H), 3.87 (d, J = 9.0 Hz, 1H), 3.71 (s, 3H), 1.43 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 171.07, 166.24, 166.15, 165.65, 165.55, 155.71, 139.66, 133.62, 133.54, 133.46, 133.29, 129.98, 129.91, 129.87, 129.82, 129.57, 129.03, 129.01, 128.67, 128.59, 128.49, 117.48, 80.05, 75.87, 71.87, 71.46, 71.17, 70.04, 67.44, 63.47, 54.07, 52.59, 28.42; HRMS (ESI) calcd. for C₄₆H₄₇NO₁₄Na (M + Na)⁺ m/z 860.2889, found 860.2893.

$\label{eq:linear} N-(\textit{Tert-butoxycarbonyl})-O-(2-(2,3,4,6-tetra-O-benzoyl-\alpha-D-mannopyranosyl) prop-2-en-1-yl)-L-tyrosine methyl ester (3ah)$



General Procedure A. White foam, 81.5 mg, 89%. $[\alpha]_D^{25}$ –32.9 (*c* 1.3, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.07 (td, *J* = 8.2, 1.4 Hz, 4H), 7.97 (dd, *J* = 8.4, 1.2 Hz, 2H), 7.88 (dd, *J* = 8.3, 1.2 Hz, 2H), 7.62–7.55 (m, 2H), 7.51 (t, *J* = 7.4 Hz, 1H), 7.47–7.41 (m,

3H), 7.40–7.35 (m, 4H), 7.30 (t, *J* = 7.8 Hz, 2H), 6.99 (d, *J* = 8.5 Hz, 2H), 6.84 (d, *J* = 8.6 Hz, 2H), 6.26 (dd, *J* = 3.1, 2.1 Hz, 1H), 6.09 (t, *J* = 9.6 Hz, 1H), 5.88 (d, *J* = 10.4 Hz, 2H), 5.84 (dd, *J* = 9.7, 3.1 Hz, 1H), 4.99 (d, *J* = 8.3 Hz, 1H), 4.96 (s, 1H), 4.68–

4.56 (m, 4H), 4.58–4.51 (m, 1H), 4.28 (ddd, J = 9.0, 5.8, 2.8 Hz, 1H), 3.70 (s, 3H), 3.02 (qd, J = 14.0, 5.9 Hz, 2H), 1.42 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.49, 166.24, 166.10, 165.64, 165.53, 157.53, 155.21, 139.16, 133.63, 133.55, 133.46, 133.26, 130.44, 129.96, 129.89, 129.84, 129.80, 129.75, 129.51, 128.98, 128.97, 128.66, 128.58, 128.57, 128.49, 118.43, 114.91, 80.02, 75.87, 71.71, 71.10, 69.83, 68.14, 67.53, 63.46, 54.61, 52.30, 37.55, 28.41; HRMS (ESI) calcd. for C₅₂H₅₁NO₁₄Na (M + Na)⁺ m/z 936.3202, found 936.3203.

N^2 -(9-Fluorenylmethoxycarbonyl)- N^6 -(6-(2,3,4,6-tetra-O-benzoyl- α -D-mannopyranosyl)hept-6-enoyl)-L-lysine methyl ester (3ai)



General Procedure A (at 25 °C). White foam, 75 mg, 70%. $[\alpha]_D^{25}$ –39.5 (*c* 1.5, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.07 (ddd, *J* = 13.1, 8.3, 1.2 Hz, 4H), 7.96 (dd, *J* = 8.3, 1.2 Hz, 2H),

7.86 (dd, J = 8.3, 1.2 Hz, 2H), 7.75 (d, J = 7.5 Hz, 2H), 7.62–7.54 (m, 4H), 7.50 (t, J = 7.4 Hz, 1H), 7.47–7.33 (m, 9H), 7.29 (q, J = 7.4 Hz, 4H), 6.20 (dd, J = 3.2, 1.9 Hz, 1H), 6.09 (t, J = 9.8 Hz, 1H), 5.73 (dd, J = 9.9, 3.1 Hz, 1H), 5.69 (t, J = 5.7 Hz, 1H), 5.57 (s, 1H), 5.47 (d, J = 8.1 Hz, 1H), 5.38 (s, 1H), 4.66 (dd, J = 12.1, 2.6 Hz, 1H), 4.51 (dd, J = 12.0, 5.5 Hz, 1H), 4.44–4.31 (m, 3H), 4.21 (t, J = 7.0 Hz, 1H), 4.15 (ddd, J = 9.7, 5.6, 2.7 Hz, 1H), 3.74 (s, 3H), 3.22 (q, J = 6.9 Hz, 2H), 2.32-2.26 (m, J = 14.6, 9.2, 5.3 Hz, 1H), 2.24–2.11 (m, 3H), 1.89–1.79 (m, 1H), 1.73–1.45 (m, 7H), 1.44–1.30 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 173.00, 166.32, 166.17, 165.81, 165.57, 156.18, 143.97, 143.80, 143.31, 141.39, 133.60, 133.54, 133.44, 133.28, 129.93, 129.89, 129.86, 129.84, 129.82, 129.60, 129.05, 129.02, 128.67, 128.58, 128.49, 127.83, 127.18, 125.20, 125.17, 120.10, 115.51, 77.01, 71.29, 71.25, 70.29, 67.53, 67.12, 63.54, 53.71, 52.55, 47.26, 39.06, 36.42, 33.04, 32.24, 29.16, 27.07, 25.40, 22.54; HRMS (ESI) calcd. for C₆₃H₆₂N₂O₁₄Na (M + Na)⁺ m/z 1093.4093, found 1093.4095.

N^2 -(*Tert*-butoxycarbonyl)- N^4 -(2-(2,3,4,6-tetra-*O*-benzoyl- α -D-mannopyranosyl) prop-2-en-1-yl)-L-asparagine methyl ester (3aj)



General Procedure A (at 25 °C). Colorless oil, 50 mg, 58%. $[\alpha]_D^{25}$ –33.3 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.10 (d, *J* = 7.3 Hz, 2H), 8.04 (d, *J* = 7.2 Hz, 2H), 7.98 (d, *J* = 7.3 Hz, 2H), 7.87 (d, *J* = 7.3 Hz, 2H), 7.60–7.57 (m, 3H), 7.52 (t, *J* = 7.4 Hz, 1H), 7.48-7..36 (m, 7H), 7.30

(t, J = 7.8 Hz, 2H), 6.26 (t, J = 6.2 Hz, 1H), 6.16 (t, J = 2.7 Hz, 1H), 6.10 (t, J = 9.6 Hz, 1H), 5.82 (d, J = 8.5 Hz, 1H), 5.72 (dd, J = 9.9, 3.4 Hz, 1H), 5.70 (s, 1H), 5.57 (s, 1H), 4.77 (s, 1H), 4.66 (dd, J = 12.1, 2.8 Hz, 1H), 4.59 (dd, J = 12.1, 5.3 Hz, 1H), 4.57–4.52 (m, 1H), 4.30–4.19 (m, 2H), 3.85 (dd, J = 16.4, 5.5 Hz, 1H), 3.74 (s, 3H), 2.96 (dd, J = 15.7, 3.5 Hz, 1H), 2.75 (dd, J = 15.7, 4.5 Hz, 1H), 1.44 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.04, 170.24, 166.36, 166.19, 165.77, 165.54, 155.81, 139.96, 133.68, 133.63, 133.54, 133.31, 129.97, 129.94, 129.89, 129.87, 129.79, 129.41, 128.99,

128.92, 128.69, 128.62, 128.55, 116.63, 80.16, 76.25, 71.71, 71.12, 69.80, 67.29, 63.28, 52.81, 50.53, 41.75, 38.05, 28.44; HRMS (ESI) calcd. for $C_{47}H_{48}N_2O_{14}Na (M + Na)^+$ m/z 887.2998, found 887.3000.

N-(*Tert*-butoxycarbonyl)-3-(1-(2,3,4,6-tetra-O-benzoyl- α -D-mannopyranosyl)eth-1-en-1-yl)-L-alanyl-L-phenylalanine methyl ester (3ak)



General Procedure A. White foam, 61.3 mg, 64%. $[\alpha]_D^{25}$ –39.9 (*c* 0.9, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.09 (d, *J* = 7.2 Hz, 2H), 8.04 (d, *J* = 7.3 Hz, 2H), 7.98 (d, *J* = 7.2 Hz, 2H), 7.87 (d, *J* = 8.4 Hz, 2H), 7.58 (q, *J* = 7.1, 6.3 Hz, 2H), 7.51 (t, *J* = 7.4 Hz, 1H), 7.48– 7.35 (m, 7H), 7.30 (t, *J* = 7.8 Hz, 2H), 7.25

(m, 2H), 7.20 (t, J = 7.3 Hz, 1H), 7.12 (d, J = 6.9 Hz, 2H), 6.82 (d, J = 7.9 Hz, 1H), 6.18 (s, 1H), 6.10 (t, J = 9.5 Hz, 1H), 5.70 (dd, J = 9.6, 3.1 Hz, 1H), 5.65 (s, 1H), 5.43 (s, 1H), 5.08 (d, J = 8.2 Hz, 1H), 4.87–4.80 (m, 2H), 4.80 (s, 1H), 4.68 (dd, J = 12.1, 2.9 Hz, 1H), 4.54 (dd, J = 12.1, 5.2 Hz, 1H), 4.41 (s, 1H), 4.23 (p, J = 3.4 Hz, 1H), 3.65 (s, 3H), 3.14 (dd, J = 13.9, 5.8 Hz, 1H), 3.07 (dd, J = 13.8, 6.5 Hz, 1H), 2.83–2.73 (m, 1H), 2.53 (dd, J = 15.3, 8.4 Hz, 1H), 1.40 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 171.58, 171.06, 166.29, 166.07, 165.75, 165.51, 155.53, 139.21, 135.94, 133.59, 133.54, 133.46, 133.19, 129.98, 129.94, 129.87, 129.86, 129.50, 129.35, 129.06, 129.02, 128.67, 128.64, 128.58, 128.50, 127.20, 118.51, 80.30, 77.26, 71.56, 70.94, 69.83, 67.41, 63.27, 53.50, 53.19, 52.39, 38.04, 36.23, 28.34; HRMS (ESI) calcd. for C₅₄H₅₄N₂O₁₄Na (M + Na)⁺ m/z 977.3467, found 977.3471.

N-(*Tert*-butoxycarbonyl)-3-(1-(2,3,4,6-tetra-*O*-benzoyl-α-D-mannopyranosyl)eth-1-en-1-yl)-L-alanyl-L-leucine methyl ester (3al)



General Procedure A. White foam, 57.7 mg, 63%. $[\alpha]_D^{25}$ –74.7 (*c* 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.10 (d, *J* = 7.7 Hz, 2H), 8.05 (d, *J* = 7.7 Hz, 2H), 7.98 (d, *J* = 7.5 Hz, 2H), 7.87 (d, *J* = 7.6 Hz, 2H), 7.58 (q, *J* = 7.4 Hz, 2H), 7.51 (t, *J* = 7.4 Hz, 1H), 7.48–7.34 (m, 7H), 7.30 (t, *J* = 7.7 Hz, 2H), 6.82 (d, *J* = 8.3 Hz, 1H), 6.21

(s, 1H), 6.11 (t, J = 9.6 Hz, 1H), 5.72 (dd, J = 9.6, 3.1 Hz, 1H), 5.67 (s, 1H), 5.47 (s, 1H), 5.14 (d, J = 8.4 Hz, 1H), 4.85 (s, 1H), 4.71 (dd, J = 12.4, 2.9 Hz, 1H), 4.63–4.55(m, 2H), 4.48–4.39 (m, 1H), 4.34–4.24 (m, 1H), 3.67 (s, 3H), 2.85 (dd, J = 15.1, 5.8 Hz, 1H), 2.56 (dd, J = 15.1, 8.4 Hz, 1H), 1.72–1.55 (m, 3H), 1.40 (s, 9H), 0.91 (d, J = 6.0 Hz, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 173.01, 171.26, 166.34, 166.09, 165.89, 165.54, 155.70, 139.13, 133.60, 133.51, 133.46, 133.20, 129.99, 129.95, 129.91, 129.89, 129.87, 129.49, 129.06, 129.03, 128.66, 128.57, 128.50, 118.46, 80.30, 71.53, 70.97, 70.03, 67.44, 63.35, 53.02, 52.37, 50.91, 41.28, 36.28, 28.32, 24.83, 22.98, 21.84; HRMS (ESI) calcd. for C₃₁H₅₆N₂O₁₄Na (M + Na)⁺ m/z 943.3624, found 943.3625.

N-(*Tert*-butoxycarbonyl)-3-(1-(2,3,4,6-tetra-O-benzoyl- α -D-mannopyranosyl)eth-1-en-1-yl)-L-alanyl-L-isoleucine methyl ester (3am)



General Procedure A. White foam, 57 mg, 62%. $[\alpha]_D^{25}$ –93.1 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.10 (d, *J* = 7.4 Hz, 2H), 8.05 (d, *J* = 7.4 Hz, 2H), 7.98 (d, *J* = 7.2 Hz, 2H), 7.87 (d, *J* = 7.2 Hz, 2H), 7.62–7.54 (m, 2H), 7.41 (m, 4H), 7.30 (t, *J* = 7.8 Hz, 2H), 6.93

(d, J = 8.7 Hz, 2H), 6.20 (s, 1H), 6.11 (t, J = 9.5 Hz, 1H), 5.73 (dd, J = 9.7, 3.1 Hz, 1H), 5.66 (s, 1H), 5.46 (s, 1H), 5.11 (d, J = 8.3 Hz, 1H), 4.84 (s, 1H), 4.69 (dd, J = 12.2, 2.8 Hz, 1H), 4.61–4.53 (m, 2H), 4.49–4.39 (m, 1H), 4.29 (s, 1H), 3.67 (s, 3H), 2.84 (dd, J = 15.3, 5.9 Hz, 1H), 2.56 (dd, J = 15.2, 8.2 Hz, 1H), 1.91 (tdt, J = 9.0, 6.8, 3.4 Hz, 1H), 1.41 (s, 10H), 1.20 (tdt, J = 16.5, 9.2, 4.2 Hz, 1H), 0.88 (m, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 171.97, 171.20, 166.29, 166.04, 165.84, 165.53, 155.69, 139.25, 133.57, 133.43, 133.17, 129.97, 129.93, 129.87, 129.85, 129.50, 129.05, 129.03, 128.65, 128.55, 128.48, 118.21, 80.29, 77.30, 77.16, 71.56, 70.91, 70.00, 67.42, 63.37, 56.70, 53.08, 52.14, 37.79, 35.94, 28.32, 25.03, 15.59, 11.65; HRMS (ESI) calcd. for C₅₁H₅₆N₂O₁₄Na (M + Na)⁺ m/z 943.3624, found 943.3627.

N-(*Tert*-butoxycarbonyl)-3-(1-(2,3,4,6-tetra-*O*-benzoyl-α-D-mannopyranosyl)eth-1-en-1-yl)-L-alanyl-L-proline methyl ester (3an)



General Procedure A. White foam, 57.4 mg, 63%. $[\alpha]_D^{25}$ -64.4 (*c* 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.13 (d, *J* = 7.3 Hz, 2H), 8.04 (d, *J* = 7.3 Hz, 2H), 7.97 (d, *J* = 7.3 Hz, 2H), 7.87 (d, *J* = 7.2 Hz, 2H), 7.58 (q, *J* = 6.6, 5.8 Hz, 2H), 7.51 (t, *J* = 7.4 Hz, 1H), 7.48–7.42 (m, 3H), 7.40–7.34

(m, 4H), 7.30 (t, J = 7.8 Hz, 2H), 6.23 (t, J = 2.5 Hz, 1H), 6.16 (t, J = 9.8 Hz, 1H), 5.78 (s, 1H), 5.72 (dd, J = 9.8, 3.1 Hz, 1H), 5.55 (s, 1H), 5.29 (d, J = 9.1 Hz, 1H), 4.85 (s, 1H), 4.76 (dd, J = 12.3, 2.5 Hz, 1H), 4.74–4.66 (m, 1H), 4.54–4.47 (m, 2H), 4.34–4.27 (m, 1H), 3.82 (dt, J = 10.6, 6.7 Hz, 1H), 3.78–3.72 (m, 1H), 3.66 (s, 3H), 2.74 (dd, J = 14.5, 5.1 Hz, 1H), 2.59 (dd, J = 14.7, 8.7 Hz, 1H), 2.23-2.18 (m, 1H), 2.10–1.93 (m, 3H), 1.40 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.32, 170.63, 166.26, 166.18, 165.63, 165.48, 155.46, 138.74, 133.56, 133.41, 133.16, 130.04, 129.98, 129.91, 129.90, 129.86, 129.66, 129.15, 129.09, 128.60, 128.55, 128.50, 120.00, 79.90, 77.67, 71.33, 71.26, 69.97, 67.36, 63.29, 59.11, 52.33, 51.07, 47.26, 36.56, 29.16, 28.40, 25.08; HRMS (ESI) calcd. for C₅₀H₅₂N₂O₁₄Na (M + Na)⁺ m/z 927.3311, found 927.3315.

N-(*Tert*-butoxycarbonyl)-L-tryptophanyl-3-(1-(2,3,4,6-tetra-O-benzoyl- α -D-mannopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (3ao)



White foam, 66.6 mg, 67%. $[\alpha]_D^{25}$ -65.1 (*c* 1.1, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.84–8.77 (m, 1H), 8.08 (t, *J* = 7.3 Hz, 4H), 7.95 (d, *J* = 7.6 Hz, 2H), 7.89 (d, *J* = 7.5 Hz, 2H), 7.76 (d, *J* = 7.9 Hz, 1H), 7.59 (dt, *J* = 16.2, 7.4 Hz, 2H), 7.48 (dt, *J* = 15.9,

7.4 Hz, 2H), 7.44–7.38 (m, 5H), 7.35 (t, J = 7.8 Hz, 2H), 7.31 (t, J = 7.7 Hz, 2H), 7.19 (t, J = 7.5 Hz, 1H), 7.13 (t, J = 7.2 Hz, 2H), 6.21 (d, J = 7.8 Hz, 1H), 6.12 (t, J = 2.5 Hz, 1H), 6.08 (t, J = 9.7 Hz, 1H), 5.62 (dd, J = 9.8, 3.1 Hz, 1H), 5.34 (s, 1H), 5.06 (s, 1H), 4.70–4.62 (m, 2H), 4.61 (s, 1H), 4.53 (s, 1H), 4.50 (dd, J = 12.1, 5.3 Hz, 1H), 4.03 (ddd, J = 8.8, 5.2, 2.6 Hz, 1H), 3.61 (s, 3H), 3.42 (d, J = 14.4 Hz, 1H), 3.10 (dd, J = 14.5, 8.2 Hz, 1H), 2.70–2.64 (m, 1H), 2.46 (dd, J = 14.6, 6.3 Hz, 1H), 1.45 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.01, 171.54, 166.32, 166.17, 166.08, 165.49, 155.60, 138.51, 136.49, 133.75, 133.65, 133.57, 133.34, 130.05, 129.86, 129.80, 129.70, 129.30, 128.94, 128.86, 128.72, 128.61, 128.59, 128.55, 127.33, 124.07, 122.35, 119.83, 119.70, 119.03, 111.56, 110.28, 80.07, 76.48, 71.46, 70.96, 70.00, 67.27, 63.31, 54.83, 52.50, 50.57, 35.41, 28.84, 28.43; HRMS (ESI) calcd. for C₅₆H₅₅N₃O₁₄Na (M + Na)⁺ m/z 1016.3576, found 1016.3579.

O-Benzyl-*N*-(*tert*-butoxycarbonyl)-L-threonyl-3-(1-(2,3,4,6-tetra-*O*-benzoyl-α-D-mannopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (3ap)



General Procedure A. White foam, 80 mg, 80%. [α] $_{D}^{25}$ -50.5 (*c* 1.4, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.09–8.04 (m, 4H), 7.95 (d, *J* = 7.3 Hz, 2H), 7.87 (d, *J* = 7.3 Hz, 2H), 7.63–7.54 (m, 2H), 7.50 (t, *J* = 7.4 Hz, 1H), 7.48–7.39 (m, 5H), 7.38–7.34 (m, 2H), 7.29 (m, 7H), 7.25–7.21 (m, 1H),

6.10 (t, J = 2.7 Hz, 1H), 6.04 (t, J = 9.6 Hz, 1H), 5.67 (dd, J = 9.7, 3.1 Hz, 1H), 5.54 (d, J = 7.1 Hz, 1H), 5.52 (s, 1H), 5.33 (s, 1H), 4.77 (q, J = 7.1 Hz, 1H), 4.71 (d, J = 2.1 Hz, 1H), 4.68–4.59 (m, 2H), 4.53 (m, 2H), 4.33 (dd, J = 6.9, 3.2 Hz, 1H), 4.19–4.05 (m, 2H), 3.71 (dd, J = 8.6, 4.8 Hz, 1H), 3.66 (s, 3H), 2.70 (dd, J = 14.7, 7.4 Hz, 1H), 2.64 (dd, J = 14.7, 7.0 Hz, 1H), 1.45 (s, 9H), 1.19 (d, J = 6.4 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 171.92, 169.96, 166.22, 165.99, 165.50, 165.45, 155.77, 139.00, 137.95, 133.57, 133.49, 133.41, 133.22, 129.93, 129.84, 129.79, 129.74, 129.53, 129.08, 128.96, 128.62, 128.54, 128.51, 128.46, 128.28, 127.93, 127.88, 80.13, 76.52, 74.99, 71.59, 71.52, 70.84, 69.65, 67.36, 63.39, 57.35, 52.43, 51.32, 35.78, 28.38, 14.82; HRMS (ESI) calcd. for C₅₆H₅₈N₂O₁₅Na (M + Na)⁺ m/z 1021.3729, found 1021.3733.

N-(*Tert*-butoxycarbonyl)-L-phenylalanyl--3-(1-(2,3,4,6-tetra-O-benzoyl- α -D-mannopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (3aq)



White foam, 65.9 mg, 69%. $[\alpha]_D^{25}$ –51.8 (*c* 1.1, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.09 (d, *J* = 7.3 Hz, 2H), 8.04 (d, *J* = 7.4 Hz, 2H), 7.96 (d, *J* = 7.3 Hz, 2H), 7.87 (d, *J* = 7.3 Hz, 2H), 7.58 (t, *J* = 7.4 Hz, 2H), 7.51 (t, *J* = 7.4 Hz, 1H), 7.50–

7.35 (m, 7H), 7.30 (t, J = 7.8 Hz, 3H), 7.24 (d, J = 7.1 Hz, 2H), 7.20 (t, J = 5.9 Hz, 3H), 6.58 (d, J = 7.6 Hz, 1H), 6.13 (t, J = 2.7 Hz, 1H), 6.08 (t, J = 9.6 Hz, 1H), 5.70 (dd, J = 9.7, 3.1 Hz, 1H), 5.66 (d, J = 1.9 Hz, 1H), 5.42 (s, 1H), 5.19–5.09 (m, 1H), 4.78 (q, J = 7.0 Hz, 1H), 4.71 (s, 1H), 4.67 (dd, J = 12.0, 2.7 Hz, 1H), 4.56 (dd, J = 12.1, 5.3 Hz, 1H), 4.44–4.32 (m, 1H), 4.17–4.09 (m, 1H), 3.66 (s, 3H), 3.12 (dd, J = 13.9, 6.3 Hz, 1H), 3.01 (d, J = 10.1 Hz, 1H), 2.77-2.66 (m, 2H), 1.37 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 171.85, 171.26, 166.28, 166.10, 165.72, 165.48, 155.52, 139.03, 136.77, 133.63, 133.58, 133.49, 133.30, 129.96, 129.89, 129.84, 129.74, 129.45, 129.43, 128.96, 128.92, 128.69, 128.66, 128.61, 128.58, 128.52, 126.99, 119.51, 80.24, 76.58, 71.65, 70.91, 69.80, 67.27, 63.32, 55.93, 52.61, 51.22, 38.34, 35.83, 28.31; HRMS (ESI) calcd. for C₅₄H₅₄N₂O₁₄Na (M + Na)⁺ m/z 977.3467, found 977.3470.

N-(*Tert*-butoxycarbonyl)-3-(1-(2,3,4,6-tetra-*O*-benzoyl-α-D-mannopyranosyl)eth-1-en-1-yl)-L-alanyl-L-valine methyl ester (3ar)



General Procedure A. White foam, 54.5 mg, 56%. $[\alpha]_D^{25}$ –58.3 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.10 (d, *J* = 7.3 Hz, 2H), 8.03 (d, *J* = 7.4 Hz, 2H), 7.98 (d, *J* = 7.4 Hz, 2H), 7.87 (d, *J* = 7.3 Hz, 2H), 7.58 (q, *J* = 7.3 Hz,

2H), 7.51 (t, J = 7.4 Hz, 1H), 7.41 (ddt, J = 29.9, 15.5, 7.6 Hz, 7H), 7.30 (t, J = 7.8 Hz, 2H), 7.05 (s, 1H), 6.83 (d, J = 8.6 Hz, 1H), 6.21 (s, 1H), 6.11 (t, J = 9.5 Hz, 1H), 5.71 (dd, J = 9.6, 3.0 Hz, 1H), 5.68 (s, 1H), 5.48 (s, 1H), 5.24 (d, J = 7.5 Hz, 1H), 4.83 (s, 1H), 4.73 (dd, J = 12.2, 2.9 Hz, 1H), 4.57 (dd, J = 12.1, 5.0 Hz, 1H), 4.54–4.48 (m, 2H), 4.44 (br, 1H), 4.30 (ddd, J = 8.6, 4.8, 2.9 Hz, 1H), 3.70 (s, 3H), 2.84 (dd, J = 15.2, 5.0 Hz, 1H), 2.58 (dd, J = 15.2, 9.0 Hz, 1H), 2.13 (dq, J = 13.7, 6.8 Hz, 1H), 1.40 (s, 12H), 0.90 (d, J = 6.9 Hz, 3H), 0.87 (d, J = 6.9 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.30, 171.83, 171.60, 166.34, 166.09, 165.82, 165.51, 155.75, 139.18, 133.61, 133.59, 133.48, 133.23, 129.96, 129.93, 129.88, 129.85, 129.42, 129.05, 128.98, 128.65, 128.57, 128.51, 118.60, 80.47, 71.58, 70.97, 69.83, 67.41, 63.20, 57.27, 53.27, 52.24, 49.33, 31.31, 28.41, 28.30, 19.03, 17.87, 17.76; HRMS (ESI) calcd. for C₅₃H₅₉N₃O₁₅Na (M + Na)⁺ m/z 1000.3838, found 1000.3841.

N-(*Tert*-butoxycarbonyl)-3-(1-(2,3,4,6-tetra-O-benzoyl- α -D-mannopyranosyl)eth-1-en-1-yl)-L-alanyl-L-tertleucyl-L-alanine methyl ester (3as)



General Procedure A. White foam, 61.8 mg, 62%. $[\alpha]_D^{25}$ -54.7 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.10 (d, *J* = 7.2 Hz, 2H), 8.05 (d, *J* = 7.4 Hz, 2H), 7.98 (d, *J* = 7.2 Hz, 2H), 7.86 (d, *J* = 7.2 Hz, 2H), 7.58 (q, *J* = 7.6 Hz,

2H), 7.51 (t, J = 7.4 Hz, 1H), 7.48–7.33 (m, 7H), 7.29 (t, J = 7.8 Hz, 2H), 7.12 (d, J = 8.7 Hz, 1H), 6.59 (d, J = 7.3 Hz, 1H), 6.21 (s, 1H), 6.11 (t, J = 9.5 Hz, 1H), 5.72 (dd, J = 9.7, 3.0 Hz, H), 5.66 (s, 1H), 5.46 (d, J = 1.6 Hz, 1H), 5.26 (d, J = 8.1 Hz, H), 4.83 (s, 1H), 4.71 (dd, J = 12.2, 2.9 Hz, 1H), 4.61–4.53 (m, 2H), 4.47 (q, J = 7.6 Hz, 1H), 4.32–4.25 (m, 2H), 3.69 (s, 3H), 2.82 (dd, J = 15.3, 5.8 Hz, 1H), 2.61 (dd, J = 15.2, 8.6 Hz, 1H), 1.40 (s, 9H), 1.37 (d, J = 7.2 Hz, 3H), 1.02 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 173.30, 171.36, 169.68, 166.28, 166.09, 165.76, 165.50, 155.72, 139.39, 133.56, 133.53, 133.43, 133.16, 129.96, 129.93, 129.88, 129.85, 129.51, 129.06, 129.01, 128.63, 128.56, 128.48, 118.31, 80.28, 71.55, 71.00, 69.87, 67.36, 63.27, 61.00, 53.41, 52.45, 47.98, 34.71, 28.44, 28.33, 26.68, 18.28; HRMS (ESI) calcd. for C₅₄H₆₁N₃O₁₅Na (M + Na)⁺ m/z 1014.3998, found 1014.4005.

N-(*Tert*-butoxycarbonyl)-3-(1-(2,3,4,6-tetra-*O*-benzoyl-α-D-mannopyranosyl)eth-1-en-1-yl)-L-alanyl-L-alanyl-*O*-(tert-butyl)-L-threonine methyl ester (3at)



General Procedure A. White foam, 54.0 mg, 52%. $[\alpha]_D^{25}$ -52.2 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.11 (d, *J* = 7.8 Hz, 2H), 8.03 (d, *J* = 7.6 Hz, 2H), 7.98 (d, *J* = 8.0 Hz, 2H), 7.87 (d, *J* = 8.0 Hz, 2H), 7.58 (t, *J* = 7.2 Hz,

2H), 7.51 (t, J = 6.9 Hz, 2H), 7.47-7.35 (m, 7H), 7.29 (dd, J = 12.7, 5.7 Hz, 2H), 7.09 (d, J = 7.3 Hz, 1H), 6.55 (d, J = 9.4 Hz, 1H), 6.21 (s, 1H), 6.10 (t, J = 9.4 Hz, 1H), 5.73 (d, J = 9.5 Hz, 1H), 5.69 (s, 1H), 5.49 (s, 1H), 5.18 (d, J = 8.9 Hz, 1H), 4.82 (s, 1H), 4.73 (d, J = 12.2 Hz, 1H), 4.61–4.51 (m, 2H), 4.51–4.42 (m, 2H), 4.28 (ddt, J = 9.9, 5.0, 2.3 Hz, 1H), 4.21 (q, J = 6.0 Hz, 1H), 3.68 (s, 3H), 2.82 (d, J = 13.9 Hz, 1H), 2.60 (dd, J = 14.8, 8.9 Hz, 1H), 1.43 (d, J = 5.9 Hz, 3H), 1.40 (s, 9H), 1.15 (d, J = 4.8 Hz, 3H), 1.09 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.38, 171.11, 171.09, 166.32, 166.07, 165.68, 165.49, 155.62, 139.36, 133.55, 133.49, 133.41, 133.18, 129.95, 129.92, 129.89, 129.85, 129.51, 129.07, 129.03, 128.60, 128.56, 128.54, 128.47, 118.70, 80.28, 74.23, 71.54, 71.06, 69.80, 67.40, 63.20, 57.89, 53.34, 52.28, 49.14, 36.56, 28.39, 28.31, 20.96, 18.59; HRMS (ESI) calcd. for C₅₆H₆₅N₃O₁₆Na (M + Na)⁺ m/z 1058.4257, found 1058.4260.

N-(Tert-butoxycarbonyl)-3-(1-(3,4,6-tri-O-benzoyl-2-deoxy-2-phthalimido-β-D-



glucopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (3ba) General Procedure B. White foam, 70.8 mg, 85%.

 $[\alpha]_{\rm D}^{25}$ 28.1 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃)

δ 8.07 (d, J = 7.3 Hz, 2H), 7.90 (d, J = 7.4 Hz, 2H), 7.86 (d, J = 7.6 Hz, 1H), 7.74 (d, J = 7.4 Hz, 2H), 7.72–7.67 (m, 2H), 7.67–7.62 (m, 1H), 7.55 (t, J = 7.3 Hz, 1H), 7.47 (t, J = 7.8 Hz, 1H), 7.41 (q, J = 8.1 Hz, 3H), 7.32 (t, J = 7.6 Hz, 2H), 7.27–7.22 (m, 2H), 6.40 (dd, J = 10.4, 9.3 Hz, 1H), 5.69 (t, J = 9.7 Hz, 1H), 5.52 (d, J = 8.3 Hz, 1H), **5.10** (s, 1H, vinyl H), 5.06 (d, J = 10.6 Hz, 1H, anomeric H), 4.94 (s, 1H, vinyl H), 4.69–4.61 (m, 2H), 4.49 (ddt, J = 12.4, 8.5, 4.4 Hz, 2H), 4.27 (ddd, J = 10.3, 5.0, 3.0 Hz, 1H), 3.65 (s, 1H), 2.73 (dd, J = 15.5, 3.9 Hz, 1H), 2.53 (dd, J = 15.5, 8.6 Hz, 1H), 1.30 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.51, 167.99, 167.63, 166.24, 165.70, 165.31, 155.51, 139.29, 134.51, 134.24, 133.50, 133.36, 133.19, 131.63, 130.98, 129.95, 129.92, 129.89, 129.81, 129.72, 128.87, 128.68, 128.46, 128.37, 123.75, 119.43, 79.63, 79.49, 76.11, 71.80, 70.20, 63.18, 52.90, 52.57, 52.17, 32.68, 28.28; HRMS (ESI) calcd. for C₄₆H₄₄N₂O₁₃Na (M + Na)⁺ m/z 855.2736, found 855.741.

N-(*Tert*-butoxycarbonyl)-3-(1-(3,4,6-tri-O-benzoyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)eth-1-en-1-yl)-L-alanyl-L-leucine methyl ester (3bl)



General Procedure B (at 15–17 °C). White foam, 69.8 mg, 73%. [α]_D²⁵ 20.8 (*c* 1.1, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.04 (d, *J* = 7.7 Hz, 2H), 7.93 (d, *J* = 8.2 Hz, 2H), 7.86 (d, *J* = 7.3 Hz, 1H), 7.75 (d, *J* = 8.2 Hz, 2H), 7.72–7.67

(m, 2H), 7.66–7.62 (m, 1H), 7.54 (t, J = 7.4 Hz, 1H), 7.49 (t, J = 7.6 Hz, 1H), 7.41 (t, J = 7.7 Hz, 3H), 7.34 (t, J = 7.7 Hz, 2H), 7.29–7.20 (m, 2H), 6.84 (d, J = 8.2 Hz, 1H), 6.41 (t, J = 9.8 Hz, 1H), 5.68 (t, J = 9.7 Hz, 1H), 5.52 (d, J = 7.7 Hz, 1H), 5.13 (d, J = 10.5 Hz, 1H), 5.09 (s, 1H), 4.99 (s, 1H), 4.71 (t, J = 10.4 Hz, 1H), 4.64 (dd, J = 12.2, 2.9 Hz, 1H), 4.58 (dd, J = 12.5, 5.9 Hz, 1H), 4.54 (dd, J = 8.2, 4.9 Hz, 1H), 4.46–4.31 (m, 1H), 3.70 (s, 3H), 2.66 (dd, J = 14.9, 5.8 Hz, 1H), 2.65–2.56 (m, 1H), 1.59–1.49 (m, 2H), 1.46–1.36 (m, 10H), 0.96–0.80 (m, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 172.93, 171.06, 168.04, 167.59, 166.32, 165.73, 165.37, 155.58, 139.05, 134.49, 134.25, 133.55, 133.37, 133.27, 131.60, 131.01, 129.96, 129.91, 129.81, 129.58, 128.84, 128.68, 128.52, 128.50, 128.39, 123.78, 123.73, 119.44, 79.98, 79.31, 76.47, 71.92, 70.37, 63.70, 53.39, 52.78, 52.28, 50.76, 41.67, 34.49, 28.34, 24.69, 22.76, 22.00; HRMS (ESI) calcd. for C₅₂H₅₅N₃O₁₄Na (M + Na)⁺ m/z 968.3576, found 968.3581.

N-(*Tert*-butoxycarbonyl)-3-(1-(3,4,6-tri-*O*-benzoyl-2-deoxy-2-phthalimido-β-Dglucopyranosyl)eth-1-en-1-yl)-L-alanyl-L-alanyl-*O*-(tert-butyl)-L-threonine methyl ester (3bt)



General Procedure B (at 15–17 °C). White foam, 60.6 mg, 57%. $[\alpha]_D^{25}$ 20.0 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.03 (d, *J* = 7.4 Hz, 2H), 7.92 (d, *J* = 7.3 Hz, 2H), 7.86 (d, *J* = 7.2 Hz,

1H), 7.74 (d, *J* = 7.2 Hz, 2H), 7.71-7.67 (m, 2H), 7.68–7.61 (m, 1H), 7.53 (t, *J* = 7.4 Hz, 1H), 7.48 (t, *J* = 7.4 Hz, 1H), 7.41 (t, *J* = 7.7 Hz, 3H), 7.34 (t, *J* = 7.8 Hz, 2H), 7.25 (t, *J* = 7.8 Hz, 2H), 7.13 (d, *J* = 7.2 Hz, 1H), 6.51 (d, *J* = 8.6 Hz, 1H), 6.41 (t, *J* = 9.8

Hz, 2H), 5.69 (t, J = 9.7 Hz, 1H), 5.61 (d, J = 7.4 Hz, 1H), 5.11 (d, J = 10.9 Hz, 1H), 5.10 (s, 1H), 5.00 (s, 1H), 4.70 (t, J = 10.4 Hz, 1H), 4.62 (d, J = 4.3 Hz, 2H), 4.52–4.43 (m, 2H), 4.42–4.34 (m, 2H), 4.22 (q, J = 5.8 Hz, 1H), 3.72 (s, 3H), 2.70–2.61 (m, J = 5.9 Hz, 2H), 1.34 (s, 9H), 1.32 (d, J = 7.4 Hz, 3H), 1.16 (d, J = 6.3 Hz, 3H), 1.11 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.29, 171.15, 171.00, 168.01, 167.56, 166.32, 165.71, 165.33, 155.51, 139.03, 134.48, 134.24, 133.52, 133.35, 133.25, 131.61, 131.01, 129.96, 129.89, 129.81, 129.61, 128.87, 128.71, 128.53, 128.48, 128.38, 123.78, 123.73, 119.57, 79.96, 79.29, 76.44, 74.26, 71.90, 70.29, 67.41, 63.55, 57.91, 53.48, 52.76, 52.32, 49.11, 34.22, 28.41, 28.31, 20.95, 18.72; HRMS (ESI) calcd. for C₅₇H₆₄N₄O₁₆Na (M + Na)⁺ m/z 1083.4210, found 1083.4213.

N-(9-Fluorenylmethoxycarbonyl)-*O*-(*tert*-butyl)-L-threonyl-3-(1-(3,4,6-tri-Obenzoyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)eth-1-en-1-yl)-L-alanyl-Lisoleucine methyl ester (3bu)



General Procedure B (at 25 °C). White foam, 95.5 mg, 78%. $[\alpha]_D^{25}$ 15.6 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.02 (d, *J* = 7.3 Hz, 2H), 7.92 (d, *J* = 7.3 Hz, 2H), 7.84 (d, *J* = 7.3 Hz, 1H), 7.76 (d, *J* = 7.6 Hz, 4H), 7.70 (d, *J* = 7.2 Hz, 1H), 7.63-7055 (m, 4H), 7.49 (dt, *J* = 14.6, 7.4 Hz, 2H), 7.41–7.38 (m, 5H), 7.34– 7.28 (m, 4H), 7.25 (t, *J* = 7.8 Hz, 2H), 6.96 (d,

J = 8.3 Hz, 1H), 6.45 (t, *J* = 9.8 Hz, 1H), 5.88 (d, *J* = 5.4 Hz, 1H), 5.69 (t, *J* = 9.7 Hz, 1H), 5.14 (d, *J* = 10.5 Hz, 1H), 5.06 (s, 1H), 4.95 (s, 1H), 4.83 (td, *J* = 8.5, 5.2 Hz, 1H), 4.69 (t, *J* = 10.4 Hz, 1H), 4.63–4.54 (m, 2H), 4.51 (dd, *J* = 8.4, 5.3 Hz, 1H), 4.43 (dd, *J* = 10.6, 7.3 Hz, 1H), 4.38–4.29 (m, 2H), 4.21 (t, *J* = 7.2 Hz, 1H), 4.18–4.09 (m, 2H), 3.71 (s, 3H), 2.84 (dd, *J* = 15.7, 5.2 Hz, 1H), 2.48 (dd, *J* = 15.6, 8.9 Hz, 1H), 1.90–1.78 (m, 1H), 1.51–1.33 (m, 1H), 1.29 (s, 3H), 1.24–1.14 (m, 1H), 1.02 (d, *J* = 6.2 Hz, 3H), 0.90-0.87 (m, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 171.72, 170.78, 169.10, 167.98, 167.71, 166.34, 165.71, 165.40, 155.86, 143.96, 143.78, 141.37, 141.36, 139.49, 134.44, 134.34, 133.50, 133.31, 133.11, 131.50, 130.90, 129.93, 129.89, 129.81, 129.67, 128.86, 128.73, 128.46, 128.42, 128.35, 127.80, 127.12, 125.24, 125.17, 123.86, 123.74, 120.07, 120.04, 117.51, 78.98, 76.37, 75.66, 71.93, 70.39, 66.96, 66.57, 63.70, 58.62, 56.83, 52.86, 52.04, 51.44, 47.23, 37.63, 35.27, 28.19, 25.34, 17.00, 15.51, 11.58; HRMS (ESI) calcd. for C₇₀H₇₂N₄O₁₆Na (M + Na)⁺ m/z 1247.4836, found 1247.4858

O-Benzyl-*N*-(*tert*-butoxycarbonyl)-L-threonyl-3-(1-(3,4,6-tri-*O*-benzoyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (3bp)



General Procedure B (at 25 °C). White foam, 64.5 mg, 63%. $[\alpha]_D^{25}$ 22.9 (*c* 1.2, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.04 (d, *J* = 7.2 Hz, 2H), 7.90 (d, *J* = 7.2 Hz, 2H), 7.86 (d, *J* = 7.3 Hz, 1H), 7.75 (t, *J* = 8.0 Hz, 3H), 7.69 (t, *J* = 7.2 Hz, 1H), 7.64 (t, *J* = 7.3 Hz, 1H), 7.54 (t, *J* = 7.4 3Hz, 1H), 7.47 (t, *J* = 6.9 Hz, 1H), 7.41 (t, *J* = 7.8 Hz, 3H), 7.35–7.28 (m, 6H), 7.25 (t, *J* = 7.8 Hz, 3H), 6.97 (d, *J* = 8.0 Hz, 1H), 6.38 (dd, *J* =

10.3, 9.3 Hz, 1H), 5.67 (t, J = 9.7 Hz, 1H), 5.53 (d, J = 6.7 Hz, 1H), 4.98 (d, J = 10.5 Hz, H), 4.87 (s, 1H), 4.77–4.70 (m, 1H), 4.68 (s, 1H), 4.63 (dd, J = 12.3, 2.8 Hz, 1H), 4.60–4.51 (m, 3H), 4.44 (dd, J = 12.2, 4.5 Hz, 1H), 4.27 (dd, J = 6.9, 3.3 Hz, 1H), 4.16 (dd, J = 9.8, 4.4 Hz, 1H), 4.05 (dq, J = 9.3, 5.7 Hz, 1H), 3.60 (s, 3H), 2.78 (dd, J = 15.9, 3.9 Hz, 1H), 2.36 (dd, J = 15.8, 10.2 Hz, 1H), 1.43 (s, 9H), 1.18 (d, J = 6.4 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.07, 169.81, 168.03, 167.69, 166.23, 165.74, 165.32, 155.55, 138.97, 138.27, 134.50, 134.41, 133.47, 133.35, 133.14, 131.57, 130.94, 129.93, 129.84, 129.78, 128.98, 128.76, 128.47, 128.44, 128.38, 127.87, 127.80, 123.90, 123.77, 117.95, 79.79, 79.11, 75.97, 75.13, 71.84, 71.43, 70.16, 63.10, 56.89, 52.97, 52.34, 50.75, 32.96, 28.44, 14.82; HRMS (ESI) calcd. for C₅₇H₅₇N₃O₁₅Na (M + Na)⁺ m/z 1046.3862, found 1046.3867.

N-(*Tert*-butoxycarbonyl)-3-(1-(3,4,6-tri-*O*-acetyl-2-deoxy-2-phthalimido-β-D-galactopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (3ca)

AcO NPhth 3ca General Procedure B (at 25–30 °C; using PPh₃ (20 mol%) or (*R*)-Tol-BINAP (10 mol%) resulted in similar yield). White foam, 51.6 mg, 79%. $[\alpha]_D^{25}$ 0.5 (*c* 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.87–7.80 (m, 2H), 7.76–7.74 (m, 2H), 5.96 (dd, *J* = 11.0, 3.3 Hz,

1H), 5.55 (d, J = 3.2 Hz, 1H), 5.44 (d, J = 8.3 Hz, 1H), 5.02 (s, 1H), 4.89 (s, 1H), 4.77 (d, J = 10.5 Hz, 1H), 4.59 (t, J = 10.7 Hz, 1H), 4.47 (td, J = 8.5, 3.7 Hz, 1H), 4.27–4.10 (m, 3H), 3.72 (s, 3H), 2.71 (dd, J = 15.9, 3.8 Hz, 1H), 2.50 (dd, J = 15.9, 8.8 Hz, 1H), 2.23 (s, 3H), 2.07 (s, 3H), 1.86 (s, 3H), 1.38 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.75, 170.49, 170.31, 169.82, 167.99, 167.92, 155.61, 139.55, 134.58, 134.35, 131.55, 131.06, 123.72, 118.98, 80.02, 79.63, 74.30, 68.51, 67.15, 61.52, 52.25, 52.21, 48.88, 32.46, 28.37, 20.84, 20.78, 20.65; HRMS (ESI) calcd. for C₃₁H₃₈N₂O₁₃Na (M + Na)⁺ m/z 669.2266, found 669.2267.

N-(*Tert*-butoxycarbonyl)-3-(1-(2,3,4-tri-O-(4-trifluoromethyl)benzoyl- β/α -D-xylopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (3da)



General Procedure B (at under 25–30 °C, using NiBr₂(DME) (10 mol%) as catalyst). ${}^{4}C_{1}(\beta)/{}^{1}C_{4}(\alpha)$ = 3:1, 57 mg, 65%. β anomer: $[\alpha]_{D}{}^{25}$ -2.5 (c 0.9,

CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.05 (t, J = 9.1 Hz, 4H), 8.00 (d, J = 8.2 Hz, 2H), 7.67 (t, J = 9.2 Hz, 4H), 7.60 (d, J = 8.2 Hz, 2H), 5.94 (t, J = 9.6 Hz, 1H), 5.47 (td, J = 9.7, 4.3 Hz, 2H), 5.36 (d, J = 8.0 Hz, 1H), 5.14 (s, 1H), 4.98 (s, 1H), 4.57–4.47 (m, 1H), 4.44 (dd, J = 11.3, 5.7 Hz, 1H), 4.13 (d, J = 9.7 Hz, 1H), 3.77 (s, 3H), 3.62 (t, J = 10.9 Hz, 1H), 2.77 (dd, J = 16.6, 3.7 Hz, 1H), 2.54 (dd, J = 16.5, 9.9 Hz, 1H), 1.42 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 173.10, 164.81, 164.68, 164.34, 155.68, 138.95, 135.27 (d, J = 33.18 Hz), 135.22 (d, J = 33.70 Hz), 135.18 (d, J = 33.73 Hz), 132.08, 131.98, 130.33, 130.31, 130.24, 125.73 (q, J = 4.1 Hz), 123.55 (q, J = 272.87 Hz), 123.51 (q, J = 273.15 Hz), 123.48 (q, J = 272.83 Hz), 118.23, 83.64, 79.89, 74.20, 70.72, 70.43, 66.97, 52.49, 51.81, 32.39, 28.45; HRMS (ESI) calcd. for C₄₀H₃₆F₉NO₁₁Na (M + Na)⁺ m/z 900.2037, found 900.2041. α anomer: ¹H NMR (600 MHz, CDCl₃) δ 8.29 (d, J = 8.0 Hz, 2H), 8.10 (d, J = 8.1 Hz, H), 8.01 (d, J = 8.1 Hz, 2H), 7.80 (d, J = 8.2 Hz)Hz, 2H), 7.59 (d, J = 8.2 Hz, 2H), 7.47 (d, J = 8.1 Hz, 2H), 5.69 (td, J = 3.0, 1.5 Hz, 1H, H3), 5.41 (d, J = 1.9 Hz, 1H, H2), 5.40 (s, 1H, vinyl-H), 5.14 (dd, J = 3.1, 1.7 Hz, 1H, H4), 5.13 (d, J = 8.4 Hz, 1H, NH), 5.10 (s, 1H, vinyl-H), 4.56 (s, 1H, H1), 4.54– 4.43 (m, 1H, α -H), 4.41 (d, J = 13.6 Hz, 1H, H5), 4.21 (dd, J = 13.6, 2.0 Hz, 1H, H5), 3.68 (s, 3H, OMe), 2.62 (dd, J = 14.8, 3.9 Hz, 1H, β -H), 2.50 (dd, J = 14.6, 10.1 Hz, 1H, β -H), 1.40 (s, 9H, Me); ¹³C NMR (150 MHz, CDCl₃) δ 172.87, 164.50, 164.43, 163.33, 155.52, 139.39, 135.57 (q, J = 32.7 Hz), 135.28 (q, J = 33.15 Hz), 135.19 (q, J = 33.32 Hz), 130.63, 130.53, 130.38, 125.98 (q, J = 3.6 Hz), 125.57 (q, J = 3.6 Hz), 125.41 (q, J = 3.6 Hz), 123.58 (q, J = 272.76 Hz), 123.46 (q, J = 272.77 Hz), 123.44 (q, J = 272.72 Hz), 116.09, 80.17, 75.70 (C1), 67.66 (C2), 67.60 (C4), 67.12 (C3), 66.87 (C5), 52.53, 52.07, 35.81, 28.36.

N-(*Tert*-butoxycarbonyl)-3-(1-(3,4,6-tri-O-acetyl-2-deoxy-2-phthalimido- α -D-mannopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (3ea)



General Procedure B (at 25 °C, using PPh₃ (20 mol%) as additive). White foam, 46.7 mg, 72%. $[\alpha]_D^{25}$ 12.6 (*c* 0.8, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.80 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.71 (dd, *J* = 5.5, 3.1 Hz, 2H), 5.43 (d, *J* = 10.5 Hz, 1H), 5.31 (t, *J* = 3.4 Hz, 1H), 5.28–5.21

(m, 2H), 5.06 (s, 1H), 4.97 (dd, J = 3.8, 1.7 Hz, 1H), 4.87–4.70 (m, 2H), 4.40 (dd, J = 11.7, 5.3 Hz, 1H), 4.38–4.29 (m, 1H), 4.24–4.15 (m, 1H), 3.67 (s, 3H), 2.56–2.45 (m, 2H), 2.23 (s, 3H), 2.13 (s, 3H), 2.07 (s, 3H), 1.40 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.86, 170.73, 169.88, 169.75, 167.90, 155.46, 139.58, 134.39, 131.48, 123.57, 119.42, 79.84, 74.09, 69.36, 69.04, 68.46, 60.59, 53.30, 52.28, 49.00, 33.01, 28.40, 21.17, 21.00, 20.91; HRMS (ESI) calcd. for C₃₁H₃₈N₂O₁₃Na (M + Na)⁺ *m*/*z* 669.2266, found 669.2257.

N-(*Tert*-butoxycarbonyl)-3-(1-(2,3,4-tri-*O*-benzoyl-α-L-rhamnopyranosyl)eth-1en-1-yl)-L-alanine methyl ester (3fa)

General Procedure B (at 25 °C, using PPh₃ (20 mol%) OBz as additive). White foam, 67 mg, 93%. $[\alpha]_D^{25}$ 114.8 (c NHBoc BzO 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.07 (d, J = CO₂Me 7.2 Hz, 2H), 7.98 (dd, J = 8.4, 1.2 Hz, 2H), 7.86 (d, J =BzO 3fa 7.2 Hz, 2H), 7.59 (t, J = 1 Hz, 1H), 7.52 (t, J = 7.4 Hz, 1H), 7.48–7.43 (m, 3H), 7.39 (t, J = 7.8 Hz, 2H), 7.29 (t, J = 7.8 Hz, 2H), 6.16 (s, 1H, H2), 5.69–5.66 (m, 2H, H3, H4), 5.64 (s, 1H, vinyl-H), 5.43 (s, 1H, vinyl-H), 5.16 (d, J = 8.2 Hz, 1H, NH), 4.74 (s, 1H, H1), 4.58 (td, J = 8.9, 4.1 Hz, 1H, α -H), 3.87 (m, 1H, H5), 3.76 (s, 3H), 2.85 (dd, J = 14.7, 4.3 Hz, 1H), 2.43 (dd, J = 14.7, 9.7 Hz, 1H), 1.42 (m, 12H); ¹³C NMR (125 MHz, CDCl₃) δ 172.85, 166.10, 165.77, 165.72, 155.40, 140.23, 133.46, 133.44, 133.34, 130.00, 129.80, 129.79, 129.66, 129.38, 129.18, 128.59, 128.54, 128.45, 118.48, 80.12, 75.92, 72.09, 70.98, 69.75, 69.71, 52.53, 52.08, 36.52, 28.35, 17.83; HRMS (ESI) calcd. for $C_{38}H_{41}NO_{11}Na (M + Na)^+ m/z 710.2572$, found 710.2574.

N-(*Tert*-butoxycarbonyl)-3-(1-(6-*O*-*tert*-butyldiphenylsilyl-3,4-di-*O*-benzoyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (3ga)

TBDPSO BzO BzO NPhth 3ga General Procedure B (at 30 °C). White foam, 67 mg, 93%. [α]_D²⁵ 13.9 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.88 (dd, *J* = 13.0, 7.4 Hz, 3H), 7.79 (d, *J* = 7.4 Hz, 2H), 7.70 (q, *J* = 7.2, 6.5 Hz, 4H),

7.64 (t, J = 7.4 Hz, 1H), 7.56 (d, J = 7.1 Hz, 2H), 7.50 (t, J = 7.4 Hz, 1H), 7.43–7.32 (m, 6H), 7.26 (d, J = 9.6 Hz, 3H), 7.15 (t, J = 7.4 Hz, 2H), 6.37 (t, J = 9.8 Hz, 1H), 5.75 (t, J = 9.6 Hz, 1H), 5.08 (d, J = 7.2 Hz, 2H), 4.97 (t, J = 5.3 Hz, 2H), 4.65 (t, J = 10.4 Hz, 1H), 4.54 (td, J = 9.7, 9.2, 3.5 Hz, 1H), 3.96 (ddd, J = 10.0, 4.1, 2.3 Hz, 1H), 3.92–3.79 (m, 2H), 3.65 (s, 3H), 2.88–2.75 (m, 1H), 2.53 (dd, J = 16.9, 10.5 Hz, 1H), 1.34 (s, 9H), 1.04 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.91, 168.03, 167.99, 165.83, 165.08, 155.57, 139.81, 135.67, 135.55, 134.45, 134.11, 133.24, 133.22, 133.01, 132.99, 131.72, 131.02, 129.87, 129.85, 129.74, 129.64, 129.37, 128.89, 128.41, 128.33, 127.76, 127.64, 123.71, 116.80, 79.73, 79.59, 78.81, 72.28, 69.77, 62.78, 53.04, 52.30, 51.43, 32.35, 28.34, 26.69, 19.24; HRMS (ESI) calcd. for C₅₅H₅₈N₂O₁₂SiNa (M + Na)⁺ m/z 989.3651, found 986.3658.

N-(*Tert*-butoxycarbonyl)-3-(1-(4-*O*-benzyl-3,6-di-*O*-benzoyl-2-deoxy-2phthalimido-β-D-glucopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (3ha)

BzO BzO NPhth 3ha (m, 2H), 7.50 (t, J = 7.8 Hz, 3H), 7.55 (t, J = 7.8 Hz, 2H), 7.50 (t, J = 7.8 Hz, 3H), 7.55 (t, J = 7.8 Hz, 2H), 7.10 (q, J = 5.9, 5.3 Hz, 5H), 6.29 (dd, J = 10.4, 8.6 Hz, 1H), 5.61 (d, J = 8.0 Hz, 1H), 5.06 (s, 1H), 4.97 (d, J = 10.5 Hz, 1H), 4.88 (s, 1H), 4.66 (s, 2H), 4.56 (q, J = 10.8 Hz, 2H), 4.49 (t, J = 10.5 Hz, 1H), 4.43 (td, J = 8.1, 3.8 Hz, 1H), 4.02 (dt, J = 10.2, 3.0 Hz, 1H), 3.93 (t, J = 9.4 Hz, 1H), 3.61 (s, 3H), 2.67 (dd, J = 15.3, 4.0 Hz, 1H), 2.52 (dd, J = 15.4, 8.2 Hz, 1H), 1.26 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.51, 167.90, 166.35, 165.63, 155.57, 139.54, 136.94, 134.48, 134.16, 133.45, 133.34, 131.71, 131.06, 130.01, 129.99, 129.82, 129.26, 128.64, 128.57, 128.51, 128.33, 128.14, 123.80, 123.70, 119.44, 79.62, 79.22, 76.66, 75.00, 74.25, 63.25, 53.09, 52.81, 52.13, 32.43, 28.30; HRMS (ESI) calcd. for C₄₆H₄₆N₂O₁₂Na (M + Na)⁺ m/z 841.2943, found 841.2946.

N-(*Tert*-butoxycarbonyl)-3-(1-(2,3,4,6-*tetra*-O-benzoyl- β/α -D-glucopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (3ia)



General Procedure B. β **-Anomer** (white foam, 20 mg, 25%): $[\alpha]_D^{25}$ 16.8 (*c* 0.5, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.04 (dd, J = 7.8, 1.6 Hz, 2H), 7.93 (d, J = 7.7 Hz, 2H), 7.91 (d, J = 7.9 Hz, 2H), 7.83 (d, J = 7.3 Hz, 2H), 7.60–7.47 (m, 2H), 7.46–7.32 (m, 7H), 7.27 (t, J = 7.7 Hz, 2H), 5.97 (t, J = 9.6 Hz, 1H), 5.73–5.58 (m, 2H), 5.47 (t, J = 9.7 Hz, 1H), **5.12 (s, 1H, vinyl H)**, 4.93 (s, 1H, vinyl H), 4.65 (dd, J = 12.3, 3.0 Hz, 1H), 4.47 (ddt, J = 12.2, 8.0, 4.0 Hz, 2H), 4.23 (d, J = 9.8

Hz, 1H, anomeric H), 4.15 (dt, J = 8.7, 3.8 Hz, 1H), 3.69 (s, 2H), 2.76 (dd, J = 16.8, 3.4 Hz, 1H), 2.60 (dd, J = 16.6, 10.0 Hz, 1H), 1.37 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 173.13, 166.30, 165.99, 165.96, 165.38, 155.83, 138.88, 133.65, 133.62, 133.42, 133.28, 129.98, 129.95, 129.83, 129.73, 129.03, 128.96, 128.87, 128.57, 128.54, 128.46, 118.21, 83.26, 79.66, 76.22, 73.84, 70.39, 69.63, 63.25, 52.32, 51.93, 31.83, 28.41. **\alpha-Anomer** (white foam, 22 mg, 27%): $[\alpha]_D^{25}$ 6.7 (*c* 0.5, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.05 (d, J = 6.8 Hz, 2H), 7.98 (d, J = 6.8 Hz, 2H), 7.96 (d, J = 8.3 Hz, 2H), 7.98 (d, J = 8.3 Hz, 2H), 2H), 7.93 (d, J = 6.9 Hz, 2H), 7.58–7.52 (m, 2H), 7.50 (t, J = 7.5 Hz, 2H), 7.43 (t, J = 7.8 Hz, 2H), 7.37 (dt, J = 9.1, 7.7 Hz, 4H), 7.31 (t, J = 7.8 Hz, 2H), 6.11 (t, J = 7.9 Hz, 1H), **5.86** (s, 1H, vinyl H), 5.59 (t, J = 7.7 Hz, 1H), **5.38** (s, 1H, vinyl H), 5.13 (d, J = 8.1 Hz, 1H), 5.01 (d, J = 4.7 Hz, 1H, anomeric H), 4.65 (dd, J = 12.1, 6.6 Hz, 1H), 4.58 (dd, J = 12.1, 3.5 Hz, 1H), 4.46 (q, J = 7.1 Hz, 1H), 4.29 (td, J = 7.2, 3.5 Hz, 1H), 3.53 (s, 3H), 2.64 (dd, J = 14.8, 7.6 Hz, 1H), 2.56 (dd, J = 15.0, 6.4 Hz, 1H), 1.36 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.98, 166.30, 165.76, 165.42, 165.41, 155.32, 138.10, 133.60, 133.59, 133.30, 130.02, 129.93, 129.88, 129.71, 129.12, 129.00, 128.64, 128.63, 128.58, 128.52, 119.36, 80.07, 72.54, 71.48, 70.52, 69.92, 69.33, 62.97, 52.36, 52.25, 36.40, 28.33; HRMS (ESI) calcd. for $C_{45}H_{45}NO_{13}Na$ (M + Na)⁺ m/z 830.2783, found 830.2780.

N-(*Tert*-butoxycarbonyl)-3-(1-(4,6-di-*O*-benzoyl-2-deoxy-2-phthalimido-3-*O*-(2,3,4,6-tetra-*O*-acetyl-β-D-galactopyranosyl)-β-D-glucopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (3ja)



General Procedure B (at 25 °C). White foam, 61.4 mg, 58%. [α]_D²⁵ -23.1 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.06 (t, *J* = 7.1 Hz, 4H), 7.90 (d, *J* = 6.6 Hz, 1H), 7.91–7.78 (m, 3H), 7.57 (t, *J* =

7.4 Hz, 1H), 7.54 (t, J = 7.5 Hz, 1H), 7.46–7.40 (m, 4H), 5.51 (d, J = 8.4 Hz, 1H), 5.44 (t, J = 9.5 Hz, 1H), 5.04–4.97 (m, 2H), 4.95 (s, 1H), 4.88 (dd, J = 10.4, 7.8 Hz, 1H), 4.85 (s, 1H), 4.66 (td, J = 9.5, 8.9, 3.9 Hz, 2H), 4.56 (dd, J = 10.4, 3.5 Hz, 1H), 4.49–4.41 (m, 3H), 4.22 (d, J = 7.9 Hz, 1H), 4.18–4.13 (m, 1H), 3.64 (s, 3H), 3.56–3.42 (m, 2H), 3.33 (dd, J = 9.8, 6.8 Hz, 1H), 2.68 (dd, J = 15.7, 3.6 Hz, 1H), 2.47 (dd, J = 15.8, 8.9 Hz, 1H), 1.99 (s, 3H), 1.86 (s, 3H), 1.82 (s, 3H), 1.81 (s, 3H), 1.31 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 172.51, 170.23, 170.15, 170.11, 169.08, 168.79, 167.69, 166.32, 164.75, 155.53, 139.74, 134.85, 133.41, 133.16, 131.26, 131.20, 129.97, 129.91, 129.88, 129.72, 128.43, 128.41, 123.86, 123.77, 118.85, 100.22, 79.94, 79.62, 76.05, 75.73, 70.94, 70.32, 69.12, 66.32, 63.43, 60.21, 53.92, 52.43, 52.16, 32.27, 28.29, 20.72, 20.52, 20.47, 20.41; HRMS (ESI) calcd. for C₅₃H₅₈N₂O₂₁Na (M + Na)⁺ *m/z* 1081.3424, found 1081.3427.

N-(*Tert*-butoxycarbonyl)-3-(1-(3,6-di-*O*-benzoyl-2-deoxy-2-phthalimido-4-*O*-(3,4,6-tri-*O*-acetyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)-β-D-glucopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (3ka)

AcO OAc OBz NHBoc AcO BzO NPhth CO₂Me **3ka** General Procedure B (at 25 °C). White foam, 61.8 mg, 58% (66% yield based on recovered starting material). $[\alpha]_D^{25}$ 78.8 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.94 (d, *J* = 7.8 Hz, 2H), 7.89–

7.79 (m, 3H), 7.70 (tt, J = 7.8, 3.6 Hz, 1H), 7.64–7.37 (m, 12H), 6.23 (dd, J = 10.2, 8.5 Hz, 1H), 5.70 (d, J = 8.4 Hz, 1H), 5.55 (dd, J = 10.7, 9.2 Hz, 1H), 5.23 (d, J = 8.3 Hz, 1H), 5.00 (t, J = 9.6 Hz, 1H), 4.99 (s, 1H), 4.92 (d, J = 10.5 Hz, 1H), 4.84 (s, 1H), 4.61 (d, J = 11.5 Hz, 1H), 4.39 (t, J = 10.4 Hz, 1H), 4.26 (dd, J = 10.7, 8.4 Hz, 1H), 4.26 (m, 1H), 4.20 (t, J = 9.2 Hz, 1H), 4.06 (dd, J = 12.1, 3.5 Hz, 1H), 3.93 (d, J = 9.9 Hz, 1H), 3.85 (dd, J = 12.3, 3.4 Hz, 1H), 3.51 (dd, J = 12.6, 2.2 Hz, 1H), 3.47 (s, 3H), 3.19 (dt, J = 10.0, 2.9 Hz, 1H), 2.53 (dd, J = 15.4, 4.0 Hz, 1H), 2.35 (dd, J = 15.4, 8.9 Hz, 1H), 2.00 (s, 3H), 1.87 (s, 3H), 1.76 (s, 3H), 1.24 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.44, 170.66, 170.16, 169.26, 167.90, 167.76, 165.48, 165.01, 155.35, 139.12, 134.48, 134.28, 134.09, 133.73, 133.14, 131.73, 130.97, 129.83, 129.72, 129.56, 129.05, 128.85, 128.41, 123.74, 123.61, 118.93, 97.78, 79.56, 79.02, 76.40, 76.20, 73.12, 71.83, 70.53, 67.95, 62.59, 60.90, 55.06, 53.04, 52.26, 52.02, 32.44, 28.24, 20.82, 20.57, 20.40; HRMS (ESI) calcd. for C₅₉H₅₉N₃O₂₁Na (M + Na)⁺ *m*/*z* 1168.3533, found 1168.3532.

 $\label{eq:linear} N-(Tert-butoxycarbonyl)-3-(1-(3,4-di-{\it O}-benzoyl-(2,3,4-tri-{\it O}-benzoyl-\alpha-L-rhamnopyranosyl)-\alpha-L-rhamnopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (3la)$



The reaction conditions were modified basing on **General procdure A**: 0.1 mmol scale, alkyne (3.0 equiv.), NiBr₂(DME) (20 mol%) instead of NiCl₂(DME) as catalyst, dtbbpy (20 mol%) and PPh₃ (30 mol%) as additive, THF (1.5 mL), 25 °C, 36 h. White foam, 70.8 mg, 68%. [α]_D²⁵ 141.8 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.04–8.01 (m, 4H), 8.00–7.96 (m, 4H), 7.83 (dd, *J* = 8.3, 1.2 Hz, 2H), 7.55 (t, *J* = 7.5 Hz, 1H), 7.51 (q, *J* = 7.7 Hz, 2H), 7.45–7.35 (m, 8H), 7.32 (t, *J* = 7.6 Hz, 2H), 7.25 (t, *J* = 5.9 Hz, 2H), 5.98 (dd, *J* = 10.1, 3.4 Hz, 1H), 5.85 (dd, *J* = 3.6, 1.8 Hz,

1H), 5.72 (q, J = 8.8 Hz, 1H), 5.66 (t, J = 10.0 Hz, 2H), 5.48 (s, 1H), 5.41 (s, 1H), 5.21 (d, J = 8.3 Hz, 1H), 5.16 (d, J = 1.7 Hz, 1H), 4.89 (s, 1H), 4.75 (t, J = 2.7 Hz, 1H), 4.62 (td, J = 9.1, 3.7 Hz, 1H), 4.40 (dq, J = 12.7, 6.3 Hz, 1H), 3.85–3.69 (m, 5H), 2.88 (dd, J = 14.4, 3.9 Hz, 1H), 2.36 (dd, J = 14.1, 9.7 Hz, 1H), 1.45 (d, J = 6.3 Hz, 3H), 1.39 (d, J = 6.6 Hz, 3H), 1.38 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.73, 166.44, 166.03, 165.44, 165.29, 165.09, 155.34, 141.04, 133.42, 133.37, 133.36, 133.24, 133.03, 130.06, 129.98, 129.90, 129.84, 129.81, 129.65, 129.52, 129.50, 129.38, 128.90, 128.62, 128.54, 128.49, 128.46, 128.30, 118.45, 99.36, 79.91, 76.46, 75.16, 72.23, 72.00, 71.89, 70.70, 69.89, 69.83, 67.79, 52.54, 52.16, 37.78, 28.31, 17.92, 17.70; HRMS (ESI) calcd. for C₅₈H₅₉NO₁₇Na (M + Na)⁺ m/z 1064.3675, found 1064.3679.

$N-(Tert-butoxycarbonyl)-3-(1-(4-O-acetyl-3-O-(2,3,4-tri-O-acetyl-\alpha-L-fucopyranosyl)-6-O-benzoyl-2-deoxy-2-phthalimido-\beta-D-glucopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (3ma)$



General Procedure B (at 30 °C). White foam, 56.7 mg, 60%. $[\alpha]_D^{25}$ –15.6 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.09 (d, *J* = 7.8 Hz, 2H), 7.88 (d, *J* = 7.0 Hz, 1H), 7.77 (d, *J* = 7.1 Hz, 1H), 7.71 (p, *J* = 7.2 Hz, 2H), 7.56 (t, *J* = 7.4 Hz, 1H), 7.45 (t, *J* = 7.6 Hz, 2H), 5.53 (d, *J* = 8.3 Hz, 1H), 5.16 (d, *J* = 3.2 Hz, 1H), 5.13 (t, *J* = 9.4 Hz,

1H), 5.05 (dd, J = 11.2, 3.4 Hz, 1H), 5.04 (d, J = 3.4 Hz, 1H), 4.96 (s, 1H), 4.95 (dd, J = 10.6, 3.3 Hz, 1H), 4.84 (s, 1H), 4.72 (d, J = 10.6 Hz, 1H), 4.67 (t, J = 9.6 Hz, 1H), 4.54 (dd, J = 12.3, 2.8 Hz, 1H), 4.41 (q, J = 10.5, 9.0 Hz, 2H), 4.31 (dd, J = 12.2, 5.6 Hz, 1H), 4.21 (q, J = 6.4 Hz, 1H), 3.88–3.83 (m, 1H), 3.61 (s, 3H), 2.63 (dd, J = 15.5, 3.7 Hz, 1H), 2.46 (dd, J = 15.6, 8.6 Hz, 1H), 2.08 (s, 3H), 2.06 (s, 3H), 1.78 (s, 3H), 1.58 (s, 3H), 1.27 (s, 9H), 1.04 (d, J = 6.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.48, 170.49, 170.47, 170.01, 169.28, 168.40, 167.62, 166.36, 155.55, 139.61, 134.40, 134.17, 133.25, 131.63, 131.57, 129.99, 129.89, 129.82, 128.53, 124.45, 123.26, 119.15, 97.20, 79.59, 79.40, 76.25, 71.84, 71.36, 68.46, 67.10, 66.00, 63.23, 53.34, 52.62, 52.13, 32.41, 28.29, 21.19, 20.69, 20.54, 20.28, 15.42; HRMS (ESI) calcd. for C₄₈H₅₃N₂O₁₉Na (M + Na)⁺ m/z 961.3237, found 961.3225.

N-(*Tert*-butoxycarbonyl)-3-(*E*-(1-(2,3,4,6-tetra-*O*-benzoyl-α-D-mannopyranosyl)-2-phenyl)eth-1-en-1-yl)-L-alanine methyl ester (3av1)



General Procedure B. White foam, 34.4 mg, 39%. $[\alpha]_D^{25}$ –55.3 (*c* 0.5, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.09 (d, *J* = 4.2 Hz, 2H), 8.08 (d, *J* = 4.1 Hz, 2H), 7.99 (d, *J* = 6.8 Hz, 2H), 7.91 (d, *J* = 6.8 Hz, 2H), 7.59 (dt, *J* = 14.9, 7.5 Hz, 2H), 7.52 (t, *J* = 7.4 Hz, 1H), 7.48 (t, *J* =

7.4 Hz, 1H), 7.45–7.40 (m, 5H), 7.38 (t, J = 7.0 Hz, 5H), 7.35–7.28 (m, 3H), **7.21** (s, **1H, vinyl H**), 6.34 (s, 1H), 6.10 (t, J = 9.1 Hz, 1H), 5.85 (dd, J = 9.4, 2.9 Hz, 1H), 5.13–5.04 (m, 2H), 4.71 (dd, J = 12.0, 3.0 Hz, 1H), 4.64 (dd, J = 12.1, 5.8 Hz, 1H), 4.59 (q, J = 7.7 Hz, 1H), 4.30–4.20 (m, 1H), 3.65 (s, 3H), 3.09 (dd, J = 14.3, 6.4 Hz, 1H), 2.79 (dd, J = 14.2, 8.0 Hz, 1H), 1.36 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.84, 166.33, 166.12, 165.69, 165.56, 155.28, 135.99, 133.64, 133.55, 133.51, 133.23, 132.53, 130.03, 129.97, 129.90, 129.88, 129.63, 129.09, 129.01, 128.71, 128.68, 128.61, 128.57, 127.81, 80.06, 71.88, 70.95, 69.75, 67.65, 63.42, 52.48, 30.66, 28.35; HRMS (ESI) calcd. for C₅₁H₄₉NO₁₃Na (M + Na)⁺ m/z 906.3096, found 906.3093.

N-(*Tert*-butoxycarbonyl)-3-(E-(2-(2,3,4,6-tetra-O-benzoyl- α -D-mannopyranosyl)-2-phenyl)eth-1-en-1-yl)-L-alanine methyl ester (3av2)



General Procedure B. Colorless oil, 8.6 mg, 13%. $[\alpha]_D^{25}$ –49.6 (*c* 0.3, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.04 (d, *J* = 9.7 Hz, 2H), 7.97 (t, *J* = 7.7 Hz, 4H), 7.86 (d, *J* = 7.0 Hz, 2H), 7.61–7.54 (m, 2H), 7.54–7.49 (m, 1H), 7.48–7.35 (m, 8H), 7.31 (dt, *J* = 13.5, 7.6 Hz, 4H), 7.23 (t, *J* = 6.1 Hz, 2H), **6.20 (t,** *J*)

= 7.4 Hz, 1H, vinyl H), 6.03 (t, J = 9.2 Hz, 1H), 5.95 (s, 1H), 5.80 (dd, J = 9.4, 3.0 Hz, 1H), 5.23 (d, J = 8.0 Hz, 1H), 5.13 (s, 1H), 4.60–4.40 (m, 3H), 4.40–4.35 (m, 1H), 3.75 (s, 3H), 2.67–2.55 (m, 1H), 2.49 (dt, J = 14.5, 7.3 Hz, 1H), 1.42 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 166.38, 165.66, 165.61, 136.56, 133.58, 133.50, 133.38, 133.15, 130.01, 129.96, 129.90, 129.69, 129.22, 129.18, 128.80, 128.68, 128.62, 128.59, 128.49, 128.46, 127.97, 126.57, 79.92, 78.05, 70.54, 69.86, 67.88, 63.51, 53.45, 52.61, 32.80, 28.45; HRMS (ESI) calcd. for C₅₁H₄₉NO₁₃Na (M + Na)⁺ m/z 906.3096, found 906.3093.

N-(*Tert*-butoxycarbonyl)-3-(*E*-(1-(3,4,6-tri-*O*-benzoyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)-2-phenyl)eth-1-en-1-yl)-L-alanine methyl ester (3bv1)



General Procedure B. White foam, 38 mg, 42%. $[\alpha]_D^{25}$ -25.0 (*c* 0.5, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.11 (d, *J* = 7.5 Hz, 2H), 7.92 (d, *J* = 6.9 Hz, 2H), 7.83 (d, *J* = 7.3 Hz, 1H), 7.77 (d, *J* = 7.8 Hz, 2H), 7.69 (d, *J* = 7.2 Hz, 1H), 7.64 (t, *J* = 7.2 Hz, 1H), 7.60

(d, J = 7.3 Hz, 1H), 7.54 (t, J = 7.7 Hz, 1H), 7.48 (t, J = 7.4 Hz, 1H), 7.46–7.39 (m, 3H), 7.33 (t, J = 7.8 Hz, 2H), 7.26 (t, J = 7.8 Hz, 2H), 7.19 (t, J = 7.2 Hz, 2H), 7.14 (t, J = 7.4 Hz, 1H), 6.96 (d, J = 7.4 Hz, 2H), **6.65 (s, 1H, vinyl H**), 6.48 (t, J = 9.8 Hz, 1H), 5.77 (t, J = 9.7 Hz, 1H), 5.16 (d, J = 10.5 Hz, 1H), 5.13 (d, J = 8.8 Hz, 1H), 4.82 (t, J = 10.4 Hz, 1H), 4.69 (d, J = 12.1 Hz, 1H), 4.58 (dd, J = 12.1, 4.7 Hz, 2H), 4.32 (dt, J = 8.1, 3.6 Hz, 1H), 3.52 (s, 3H), 2.89 (dd, J = 14.3, 8.9 Hz, 1H), 2.78 (dd, J = 14.3, 4.2

Hz, 1H), 1.34 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.77, 168.26, 167.64, 166.39, 165.81, 165.34, 155.15, 135.89, 134.51, 134.29, 133.48, 133.36, 133.11, 132.30, 131.62, 131.03, 130.08, 129.98, 129.89, 129.81, 129.05, 128.83, 128.64, 128.53, 128.48, 128.46, 128.39, 128.35, 128.30, 127.49, 123.80, 123.65, 80.68, 79.52, 76.19, 72.07, 70.21, 63.34, 53.38, 53.01, 52.21, 29.56, 28.40; HRMS (ESI) calcd. for C₅₂H₄₈N₂O₁₃Na (M + Na)⁺ m/z 931.3049, found 931.3044.

N-(*Tert*-butoxycarbonyl)-3-(*E*-(2-(3,4,6-tri-*O*-benzoyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)-2-phenyl)eth-1-en-1-yl)-L-alanine methyl ester (3bv2)



General Procedure B. Colorless oil, 4.7 mg, 5%. $[\alpha]_D^{25}$ 57.2 (*c* 0.2, CHCl₃); ¹H NMR (500 MHz, CDCl₃ δ 8.06 (d, *J* = 8.1 Hz, 2H), 7.89 (dd, *J* = 8.4, 1.3 Hz, 2H), 7.75 (d, *J* = 7.3 Hz, 1H), 7.71–7.53 (m, 6H), 7.50–7.41 (m, 3H), 7.38 (t, *J* = 7.4 Hz, 1H),

7.32 (dd, J = 8.3, 7.4 Hz, 2H), 7.21 (dd, J = 8.3, 7.4 Hz, 2H), 7.10–7.04 (m, 3H), 6.97 (dd, J = 7.4, 2.0 Hz, 2H), 6.24 (dd, J = 10.4, 9.3 Hz, 1H), **5.84 (t, J = 7.4 Hz, 1H, vinyl H**), 5.62 (t, J = 9.7 Hz, 1H), 5.36 (d, J = 10.3 Hz, 1H), 4.96 (d, J = 8.2 Hz, 1H), 4.72 (dd, J = 12.1, 3.0 Hz, 1H), 4.46 (dd, J = 11.7, 3.9 Hz, 1H), 4.43 (t, J = 9.7 Hz, 1H), 4.27 (ddd, J = 10.1, 4.3, 3.0 Hz, 1H), 4.21 (q, J = 6.3 Hz, 1H), 3.53 (s, 3H), 2.50–2.40 (m, 1H), 2.40–2.20 (m, 1H), 1.40 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 168.18, 166.32, 165.95, 165.35, 135.71, 134.18, 134.02, 133.46, 133.27, 133.20, 130.00, 129.98, 129.94, 129.87, 129.40, 129.11, 128.81, 128.55, 128.49, 128.34, 128.13, 127.51, 123.53, 123.46, 79.42, 78.94, 75.93, 72.50, 70.16, 63.11, 54.26, 52.83, 52.19, 31.72, 28.45; HRMS (ESI) calcd. for C₅₂H₄₈N₂O₁₃Na (M + Na)⁺ m/z 931.3049, found 931.3046.

 $N-(Tert-butoxycarbonyl)-3-(1-(2,4-di-O-benzoyl-3-O-(4-O-benzoyl-3-O-(3,4-di-O-benzoyl-2-O-(2,3,4-tri-O-benzoyl-\alpha-L-rhamnopyranosyl)-\alpha-L-rhamnopyranosyl)-\alpha-L-rhamnopyranosyl)-\alpha-L-rhamnopyranosyl)-\alpha-L-rhamnopyranosyl)-theorem (14)$



Supplementary Figure 36. Preparation of pentasaccharide amino acid 14.

The amount of **S29** was small (about 0.0122 mmol, Supplementary Figure 11), therefore accurately weighing other reagents was difficult. As a compromise, we conducted the synthesis with 1a as the reference substrate. Thus, to an oven-dried 10 mL Schlenk tube (Titan, TF891910) containing a Teflon coated magnetic stirring bar was added glycosyl bromide 1a (66 mg, 0.1 mmol), NiCl₂(DME) (10 mol%), dtbbpy (12 mol%), (R)-Tol-BINAP (10 mol%), and Na₂CO₃ (2.5 equiv). The tube was sealed with a rubber cap and wrapped with parafilm, and was then evacuated then refilled with Ar for at least five cycles. Alkyne 2a (56 mg, 0.25 mmol) was dissolved in THF (1.0 mL) and injected into the reaction tube. When stirring, silane PMHS (32 µL, 2.5 equiv.) was injected via microliter syringe. The tube was kept stirring under 30 $\,^{\circ}$ C for 1.5 h. Then **S29** (which was used directly after preparation, about 0.0122 mmol, see Supplementary Figure 11) was dissolved in THF (0.3 mL) and was injected into the reaction tube via microliter syringe; the residue glycosyl bromide was dissolved in another portion of THF (0.2 mL) and added. The tube was evacuated then refilled with Ar for three cycles again, then the mixture was stirred at 30 $\,^{\circ}$ C for 36 h. The reaction mixture was diluted with CH₂Cl₂ (20 mL) and filtered. The filtrate was concentrated in The residue was purified by preparative TLC (petroleum ether/ vacuo. dichloromethane/ethyl acetate = 2:1:1, $R_f = 0.3$) to afford the desired product **3aa** (59.4) mg, 73%) and 14 (10 mg, 39% over two steps).



[α]_D²⁵ 83.9 (*c* 0.3, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 8.13 (d, *J* = 7.7 Hz, 2H), 8.06 (d, *J* = 7.9 Hz, 2H), 8.02 (d, *J* = 7.8 Hz, 2H), 8.00 (d, *J* = 7.6 Hz, 2H), 7.96 (t, *J* = 8.2 Hz, 4H), 7.85 (s, 2H), 7.80 (d, *J* = 7.7 Hz, 2H), 7.67–7.54 (m, 5H), 7.53–7.45 (m, 7H), 7.43–7.35(m, 5H), 7.30–7.23 (m, 5H), 7.06 (t, *J* = 7.7 Hz, 1H), 6.85 (t, *J* = 7.5 Hz, 1H), 6.01 (dd, *J* = 10.7, 9.0 Hz, 1H), 5.83 (dd, *J* = 10.1, 3.0 Hz, 2H), 5.80–5.72 (m, 3H), 5.50–5.45 (m, 2H), 5.43 (t, *J* = 10.0 Hz, 1H), 5.28 (s, 1H), 5.27 (s, 1H), 5.15 (d, *J* = 6.7 Hz, 1H), 5.08–5.01 (m, 2H), 4.89 (s, 1H), 4.87 (s, 1H), 4.73 (t, *J* = 9.8 Hz, 1H), 4.32–4.26 (m, 2H), 4.17 (s, 1H), 4.09 (d, *J* = 9.0 Hz, 1H), 4.02 (s, 1H), 3.94–3.88 (m, 2H), 3.83–3.73 (m, 1H), 3.79–3.74

(m, 1H), 3.72 (s, 1H), 3.62 (dq, J = 11.8, 6.1 Hz, 1H), 2.75 (d, J = 15.3 Hz, 1H), 2.39 (dd, J = 15.1, 9.6 Hz, 1H), 2.04 (s, 3H), 1.99 (s, 3H), 1.84 (s, 3H), 1.38 (s, 9H), 1.34 (d, J = 6.4 Hz, 3H), 1.28 (d, J = 6.1 Hz, 3H), 0.84 (d, J = 6.2 Hz, 3H), 0.71 (d, J = 6.2 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 172.86, 170.78, 169.97, 169.86, 167.78, 166.15, 165.91, 165.71, 165.59, 165.38, 165.15, 165.08, 164.29, 155.43, 140.74, 134.15, 133.67, 133.58, 133.55, 133.21, 133.17, 133.10, 133.01, 131.57, 130.10, 130.01, 130.00, 129.99, 129.93, 129.82, 129.71, 129.63, 129.57, 129.52, 129.45, 129.44, 129.08, 128.80, 128.72, 128.54, 128.48, 128.42, 128.39, 124.05, 117.60, 100.51, 99.17, 99.08, 98.23, 80.05, 76.52, 75.40, 75.11, 73.86, 73.18, 72.70, 71.92, 71.57, 71.18, 70.54, 70.36, 70.31, 69.98, 69.54, 69.37, 68.37, 68.16, 67.47, 61.82, 54.69, 53.93, 53.58, 52.53, 52.23, 35.92, 28.38, 20.90, 20.85, 20.71, 17.67, 17.58, 17.50, 17.41; HRMS (ESI) calcd. for C₁₁₁H₁₁₀N₂O₃₇Na (M + Na)⁺ m/z 2085.6680, found 2085.6686.

N-(*Tert*-butoxycarbonyl)-3-(1-(3-O-(2,3,4-tri-O-acetyl- α -L-fucopyranosyl)-4-O-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)-6-O-benzoyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (15)



General Procedure B (at 30 °C). White foam, 71 mg, 58%. $[\alpha]_D^{25}$ –48.8 (*c* 1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 8.11 (d, *J* = 7.8 Hz, 2H), 7.84 (dd, *J* = 5.7, 2.8 Hz, 1H), 7.79 (dd, *J* = 5.4, 3.3 Hz, 1H), 7.78–7.75 (m, 2H), 7.64 (t, *J* = 7.2 Hz, 1H), 7.55 (t, *J* = 7.5 Hz, 2H), 5.41–5.32 (m, 3H,

NH, H3', H4', H4''), 5.15 (t, J = 7.81 Hz, 1H, H2''), 5.14 (dd, J = 9.7, 4.3 Hz, 1H, H2'), 5.01 (q, J = 6.7 Hz, 1H, H5'), 4.95 (d, J = 4.0 Hz, 1H, H1'), 4.94 (dd, J = 12.18, 2.9 Hz, 1H), 4.88 (t, J = 9.46 Hz, 1H, H3), **4.85 (s, 1H, vinyl-H**), 4.78 (dd, J = 10.9, 4.1 Hz, 1H, H3''), **4.76 (s, 1H, vinyl-H**), 4.69 (d, J = 8.2 Hz, 1H, H1''), 4.60 (d, J = 10.5 Hz, 1H, H1), 4.55 (dd, J = 11.4, 5.9 Hz, 1H, H6'), 4.44 (dd, J = 12.2, 3.9 Hz, 1H, H6), 4.38 (dt, J = 8.0, 3.9 Hz, 1H, α -H), 4.34 (t, J = 10.4 Hz, 1H, H2), 4.30 (dd, J = 11.4, 8.2 Hz, 1H, H6''), 4.07 (t, J = 9.4 Hz, 1H, H4), 3.84–3.78 (m, 1H, H5), 3.76–3.73

(m, 1H, H5^{''}), 3.56 (s, 3H, OMe), 2.61 (dd, J = 16.0, 3.6 Hz, 1H, β-H), 2.40 (dd, J = 15.9, 9.3 Hz, 1H, β-H), 2.18 (s, 3H), 2.15 (s, 3H), 2.07 (s, 3H), 2.07 (s, 3H), 2.01 (s, 3H), 1.97 (s, 3H), 1.92 (s, 3H), 1.30 (s, 9H), 1.24 (d, J = 6.6 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) δ carbonyl (172.50, 170.69, 170.54, 170.42, 170.36, 169.98, 169.70, 169.09, 168.63, 168.07, 165.81, 155.39), arene and alkene (139.57, 134.51, 134.45, 133.58, 131.47, 131.07, 129.74, 128.79, 123.79, 123.53, 118.43), 100.63 (C1''), 95.08 (C1'), 80.01 (C1), 79.49 (C-Boc), 77.28 (C5), 75.38 (C4), 71.96 (C3), 71.38 (C4' or C4''), 71.14 (C5''), 71.08 (C4' or C4''), 68.88 (C2' or C2''), 68.01 (C3''), 67.95 (C2' or C2''), 66.59 (C3'), 64.30 (C5'), 62.16 (C6), 60.44 (C6''), 54.51(C2), 52.16 (α-C), 52.00 (OMe), 31.80 (β-C), 28.20 (Me-Boc), Me of acetyl (20.82, 20.71, 20.70, 20.65, 20.55, 20.53), 15.88 (C6'); HRMS (ESI) calcd. for C₅₈H₇₀N₂O₂₇Na (M + Na)⁺ m/z 1249.4058, found 1249.4046.

2.6 The Scale-up Reaction







12 h

24 h

48 h



Supplementary Figure 37. A gram scale preparation of 3ba.

To a 250 mL three neck round-bottom flask containing a Teflon coated magnetic stirring bar was added glycosyl bromide 1b (2.465 g, 3.6 mmol), NiCl₂(DME) (79 mg, 10 mol%), dtbbpy (145 mg, 15 mol%), (*R*)-Tol-BINAP (146 mg, 6 mol%), and Na₂CO₃ (954 mg, 2.5 equiv). The flask was equipped with a balloon filled with Ar and sealed with two rubber caps then with parafilm and evacuated then refilled with Ar for at least five cycles. Compound 2a (2.04 g, 2.5 equiv.) was dissolved in THF (36 mL) and injected into the flask. The flask was evacuated and refilled with Ar for another three cycles. When stirring, silane PMHS (1.116 mL, 2.5 equiv.) was injected via microliter syringe. After which a light green solution was obtained and 10 minutes later the reaction mixture began to turn red slowly. 30 minutes later the reaction mixture became dark red. The mixture was kept stirring vigorously under indicated temperature (25–28 °C) for 48 h until the completion of the reaction as determined by TLC. The mixture was then diluted with CH_2Cl_2 (50 mL) and filtered. The filtrate was concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 2:1 or petroleum ether/dichloromethane/ethyl acetate = 2:1:0.5) to afford the desired product **3ba** (1.96 g, 65%) as a white solid.

2.7 Transformations of vinyl C-glycosyl amino acids

Protecting group transformation



To a stirred solution of **3ba** (416.5 mg, 0.5 mmol) in MeOH (5 mL) was added 80% hydrazine hydrate (160 μ L, about 5.0 equiv.) via microliter syringe at 0 °C. The mixture was stirred at 0 °C for 9 h to open the Phth ring. After the disappearance of **3ba** as determined by TLC (petroleum ether/ethyl acetate = 1:1), the mixture was diluted and azeotroped with toluene (3×30 mL) to remove the residue hydrazine. The residue was dissolved in MeOH/HOAc (10 mL, 4:1, v/v), and the solution was heated under 70 °C for 1.5 h to remove the residue Phth. The mixture was concentrated *in vacuo* and dried over oil pump. The residue was dissolved in dry CH₂Cl₂ (7 mL), to which Et₃N (0.8 mL) and Ac₂O (0.5 mL) were added. The mixture was stirred at room temperature for 6 h, after which MeOH (0.5 mL) was added and stirred for 0.5 h. The solvent was removed *in vacuo*. The residue was purified by column chromatography (petroleum ether/ethyl acetate = 1:1) to provide **8** (223 mg, 60% yield).

N-(*Tert*-butoxycarbonyl)-3-(1-(3,4,6-tri-O-benzoyl-2-deoxy-2-acetylamido- β -D-glucopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (8)

 $\begin{array}{c} H NM \\ H NM \\ BzO \\ BzO \\ NHAc \\ \mathbf{8} \\ \mathbf{8} \\ \mathbf{8} \\ \mathbf{8} \\ \mathbf{8} \\ \mathbf{8} \\ \mathbf{10} \\ \mathbf{10$

¹H NMR (500 MHz, CDCl₃) δ 8.02 (d, J = 7.7 Hz, 2H), 7.91 (d, J = 7.7 Hz, 2H), 7.87 (d, J = 8.0 Hz, 2H), 7.53– 7.45 (m, 3H), 7.40–7.29 (m, 6H), 6.11 (d, J = 10.8 Hz, 1H), 5.85 (d, J = 7.4 Hz, 1H), 5.66 (t, J = 9.7 Hz, 1H), 5.62–5.49 (m, 1H), 5.10 (s, 1H), 5.01 (s, 1), 4.63 (dd,

J = 12.3, 2.8 Hz, 1), 4.51–4.42 (m, 3H), 4.17–4.07 (m, 1H), 4.00 (d, J = 9.7 Hz, 1H), 3.68 (s, 3H), 2.74 (d, J = 15.5 Hz, 1H), 2.60 (dd, J = 16.1, 10.7 Hz, 1H), 1.81 (s, 3H), 1.39 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 173.14, 170.95, 167.42, 166.31, 165.24, 156.03, 139.70, 133.83, 133.57, 133.20, 129.94, 129.86, 129.74, 129.66, 128.88, 128.64, 128.56, 128.46, 117.23, 84.64, 79.45, 76.06, 74.64, 69.49, 63.32, 52.25, 51.93, 51.78, 32.05, 28.41, 23.14; HRMS (ESI) calcd. for C₄₀H₄₄N₂O₁₂Na (M + Na)⁺ m/z 767.2786, found 767.2792.



The **3ba-epimer** was prepared according to **General Procedure B** and was used for analysing the dr value of **3ba**. To an oven-dried 10 mL Schlenk tube (Titan, TF891910) containing a Teflon coated magnetic stirring bar were added glycosyl bromide **1b** (0.1 mmol), NiCl₂(DME) (10 mol%), dtbbpy (12 mol%), (*R*)-Tol-BINAP (10 mol%), and Na₂CO₃ (2.5 equiv). The tube was sealed with a rubber cap then with parafilm and evacuated then refilled with Ar for at least five cycles. *N*-Boc-Pra-OMe (**2a**-*rac*, prepared by mixing the *D*- and *L*-enantiomer in 1:1 ratio, 2.0 equiv) was dissolved in THF (1.0 mL) and injected into the reaction tube. When stirring, PMHS (32 μ L, 2.5 equiv.) was injected via microliter syringe. The tube was kept stirring under 30 °C for 36 h. The reaction mixture was diluted with CH₂Cl₂ (20 mL) and filtered. After concentration, the residue was purified by flash column chromatography to afford the desired product (69 mg, 83%) as a syrup. The two diastereomers could not be seperated by flash column chromatography and were obtained as a mixtue in a 1:1 dr ratio.

N-(*Tert*-butoxycarbonyl)-3-(1-(3,4,6-tri-*O*-benzoyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl) eth-1-en-1-yl)- D/L-alanine methyl ester (3ba-epimer)



S-Diastereomer: ¹H NMR (500 MHz, CDCl₃) δ 8.07 (d, J = 7.3 Hz, 2H), 7.90 (d, J = 7.4 Hz, 2H), 7.86 (d, J = 7.6 Hz, 1H), 7.74 (d, J = 7.4 Hz, 2H), 7.72–7.67 (m, 2H), 7.67–7.62 (m, 1H), 7.55 (t, J = 7.3 Hz, 1H), 7.47 (t, J = 7.8 Hz, 1H), 7.41 (q, J = 8.1 Hz, 3H), 7.32

(t, J = 7.6 Hz, 2H), 7.27–7.22 (m, 2H), 6.40 (dd, J = 10.4, 9.3 Hz, 1H), 5.69 (t, J = 9.7 Hz, 1H), 5.52 (d, J = 8.3 Hz, 1H), 5.10 (s, 1H), 5.06 (d, J = 10.6 Hz, 1H), 4.94 (s, 1H), 4.69–4.61 (m, 2H), 4.49 (ddt, J = 12.4, 8.5, 4.4 Hz, 2H), 4.27 (ddd, J = 10.3, 5.0, 3.0 Hz, 1H), 3.65 (s, 1H), 2.73 (dd, J = 15.5, 3.9 Hz, 1H), 2.53 (dd, J = 15.5, 8.6 Hz, 1H), 1.30 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.51, 167.99, 167.63, 166.24, 165.70, 165.31, 155.51, 139.29, 134.51, 134.24, 133.50, 133.36, 133.19, 131.63, 130.98, 129.95, 129.92, 129.89, 129.81, 129.72, 128.87, 128.68, 128.46, 128.37, 123.75, 119.43, 79.63, 79.49, 76.11, 71.80, 70.20, 63.18, 52.90, 52.57, 52.17, 32.68, 28.28. *R***-Diastereomer:** ¹H NMR (500 MHz, CDCl₃) δ 8.11 (d, J = 7.0 Hz, 2H), 7.91 (d, J = 7.0 Hz, 2H), 7.88 (d, J = 7.3 Hz, 1H), 7.77–7.62 (m, 5H), 7.57 (q, J = 6.3, 5.4 Hz, 1H), 7.51–7.44 (m, 3H), 7.41 (t, J = 7.5 Hz, 1H), 7.33 (t, J = 7.9 Hz, 2H), 7.25 (t, J = 7.4 Hz, 2H), 6.38 (t, J = 9.8 Hz, 1H), 5.72 (t, J = 9.7 Hz, 1H), 5.68 (d, J = 7.2 Hz, 1H), 5.19 (s, 1H), 5.11 (s, 1H), 5.09 (d, J = 10.5 Hz, 1H), 4.62 (t, J = 10.4 Hz, 1H), 4.60 (s, 2H), 4.33–4.25 (m, 2H), 3.71 (s, 3H), 2.57 (d, J = 7.3 Hz, 2H), 1.31 (s, 9H); ¹³C NMR (125

 $\begin{array}{l} MHz, CDCl_3)\,\delta\,172.76,\,168.09,\,167.36,\,166.29,\,165.75,\,165.24,\,155.60,\,139.57,\,134.53,\\ 134.32,\,\,133.51,\,\,133.38,\,\,133.22,\,\,131.59,\,\,130.98,\,\,130.04,\,\,129.96,\,\,129.93,\,\,129.81,\\ 129.65,\,128.87,\,128.64,\,128.55,128.46,\,128.37,\,123.77,\,119.96,\,79.71,\,79.05,\,75.92,\\ 71.78,\,69.79,\,62.68,54.26,\,52.89,\,52.33,\,31.91,\,28.28 \end{array}$



Supplementary Figure 38. ¹H NMR spectrum of the 1:1 mixture of 3ba epimers.



Supplementary Figure 39. ¹³C NMR spectrum of the 1:1 mixture of 3ba epimers.



Supplementary Figure 40. HPLC chromatogram of the 1:1 mixture of **3ba** epimers (Chiralpak® ID3, *n*-hexane/*i*-PrOH 70:30, 1.0 mL/min, detection at 214 nm)



Supplementary Figure 41. HPLC chromatogram of **3ba** prepared by the present method (Chiralpak® ID3, *n*-hexane/*i*-PrOH 70:30, 1.0 mL/min, detection at 214 nm).


In addition, the synthesized **3ba-**R was subjected to the deprotection procudure mentioned above, and the resulting **8-**R was obtained as a standard to check the retention of chirality in compund **8**.

N-(*Tert*-butoxycarbonyl)-3-(1-(3,4,6-tri-*O*-benzoyl-2-deoxy-2-acetylamido- β -D-glucopyranosyl)eth-1-en-1-yl)-*D*-alanine methyl ester (8-*R*)



White powder, 18 mg, 55%. $[\alpha]_D^{25}$ -20.6 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.07 (d, *J* = 7.0 Hz, 2H), 7.90 (d, *J* = 8.1 Hz, 2H), 7.86 (d, *J* = 7.8 Hz, 2H), 7.55 (t, *J* = 7.5 Hz, 1H), 7.52–7.40 (m, 4H), 7.34 (t, *J* = 7.7 Hz, 2H), 7.31 (t, *J* = 7.7 Hz, 2H), 5.92 (d, *J*

= 9.2 Hz, 1H), 5.70 (d, J = 6.7 Hz, 1H), 5.66 (d, J = 9.6 Hz, 1H), 5.63 (d, J = 9.8 Hz, 1H), 5.22 (s, 2H), 4.56 (d, J = 3.3 Hz, 2H), 4.36 (q, J = 9.8 Hz, 1H), 4.31–4.24 (m, 1H), 4.11 (dt, J = 8.4, 3.2 Hz, 1H), 4.05 (d, J = 10.2 Hz, 1H), 3.71 (s, 3H), 2.72 (dd, J = 15.1, 4.5 Hz, 1H), 2.53 (dd, J = 14.9, 10.1 Hz, 1H), 1.80 (s, 3H), 1.33 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.93, 170.30, 167.22, 166.35, 165.18, 155.76, 140.14, 133.76, 133.55, 133.28, 130.02, 129.97, 129.81, 129.60, 128.93, 128.71, 128.63, 128.58, 128.55, 119.44, 83.09, 79.79, 75.92, 74.52, 69.09, 62.69, 54.43, 52.41, 52.38, 32.61, 28.35, 23.27.

The **8-epimer** was prepared via mixing compound **8** and compound **8-***R* in 53.3:47.7 ratio (as determined by chiral HPLC). The retention of chirality in compound **8** was proven by chiral HPLC analysis (Supplementary Figures 42 and 43).



Supplementary Figure 42. HPLC chromatogram of the mixture of **8-***R* and **8**(*S*) prepared by the present method (Chiralpak® ID3, *n*-hexane/*i*-PrOH 70:30, 1.0 mL/min, detection at 214 nm).



Supplementary Figure 43. HPLC chromatogram of **8** prepared by the present method (Chiralpak® ID3, *n*-hexane/*i*-PrOH 70:30, 1.0 mL/min, detection at 214 nm).

Saccharide chain elongation

HO

BzO[^] BzO



Supplementary Figure 44. Preparation of vinyl C-disaccharide amino acid 9.

To a stirred solution of **3ga** (596.7 g, 0.62 mmol) in pyridine (20 mL) was added HFpyridine complex (2 mL) at 0 °C. The mixture was gradually warmed up to room temperature and stirred for 2 h. The reaction mixture was diluted with EtOAc, washed with sat. aq. NaHCO₃ and brine. After dried over Na₂SO₄ and concentrated, the residue was purified by silica gel flash column chromatography (petroleum ether/ethyl acetate = 2:1) to afford **S39** (382 mg, 85%).

N-(*Tert*-butoxycarbonyl)-3-(1-(3,4-di-*O*-benzoyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (S39)



1H), 5.11 (s, 1H), 5.05 (d, J = 10.5 Hz, 1H), 4.90 (s, 1H), 4.78 (td, J = 9.6, 3.9 Hz, 1H), 4.65 (t, J = 10.4 Hz, 1H), 3.98–3.91 (m, 1H), 3.78 (d, J = 8.7 Hz, 4H), 3.73–3.67 (m, 1H), 2.83 (dd, J = 14.7, 3.9 Hz, 1H), 2.32 (dd, J = 14.5, 9.7 Hz, 1H), 1.43 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.86, 167.99, 167.52, 165.80, 165.72, 155.58, 139.05, 134.50, 134.28, 133.58, 133.36, 131.69, 131.05, 130.02, 129.85, 128.98, 128.85, 128.53, 128.42, 123.76, 121.55, 80.45, 79.81, 79.05, 71.93, 70.20, 61.86, 53.06, 52.78, 52.47, 34.36, 28.44; HRMS (ESI) calcd. for C₃₉H₄₀N₂O₁₂Na (M + Na)⁺ m/z 751.2473, found 751.2476.

Compound **S39** (36.4 mg, 0.05 mmol), 2,3,4-tri-*O*-acetyl- β -L-fucopyranosyl 2-(cyclopropylethynyl)benzoate **S40** (22.9 mg, 1.0 equiv.), and 4Å MS (50 mg) were suspended in extra dry CH₂Cl₂ (1.0 mL). The mixture was stirred at room temperature for 0.5 h, then cooled to 0 °C. Au(PPh₃)NTf₂ (3.6 mg, 10 mol%) was added in one portion. The reaction mixture was stirred for 0.5 h at room temperature, and was quenched with Et₃N (10 µL). The mixture was diluted with CH₂Cl₂ (20 mL) and filtrated. After concentration, the residue was purified by preparative TLC (petroleum ether/dichloromethane/ethyl acetate = 2:1:1) to afford **9** (40 mg, 81%) as a white foam. N-(*Tert*-butoxycarbonyl)-3-(1-(3,4-di-O-benzoyl-6-O-(2,3,4-tri-O-acetyl- α -L-fucopyranosyl)-2-deoxy-2-phthalimido- β -D-glucopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (9)



[α]_D²⁵ 2.5 (*c* 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.91 (d, J = 7.9 Hz, 2H), 7.86 (d, J = 7.2 Hz, 1H), 7.69 (dt, J = 19.2, 7.5 Hz, 5H), 7.49 (t, J = 7.4 Hz, 1H), 7.42 (t, J = 7.4 Hz, 1H), 7.35 (t, J = 7.6 Hz, 2H), 7.30– 7.23 (m, 2H), 6.30 (t, J = 9.8 Hz, 1H), 5.62 (t, J = 9.7Hz, 1H), 5.46 (d, J = 8.7 Hz, 1H), 5.27–5.19 (m, 2H), 5.17–5.13 (m, 1H), 5.11 (s, 1H), 5.00 (d, J = 10.8 Hz, 1H), 4.85 (d, J = 7.9 Hz, 1H), 4.62–4.52 (m, 2H), 4.10

(d, J = 10.0 Hz, 1H), 4.04–3.96 (m, 2H), 3.96–3.88 (m, 1H), 3.74 (s, 3H), 2.85 (dd, J = 16.1, 5.5 Hz, 1H), 2.45 (dd, J = 16.1, 8.6 Hz, 1H), 2.24 (s, 3H), 2.15 (s, 3H), 1.99 (s, 3H), 1.40 (s, 9H), 1.11 (d, J = 6.4 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.89, 170.87, 170.32, 169.63, 167.98, 167.66, 165.75, 164.92, 155.56, 139.55, 134.51, 134.24, 133.36, 133.32, 131.63, 130.98, 129.90, 129.76, 129.19, 128.79, 128.39, 123.78, 123.72, 118.56, 100.75, 79.95, 79.79, 78.24, 72.17, 71.52, 70.74, 69.44, 69.38, 69.05, 66.87, 52.85, 52.46, 51.91, 32.07, 28.39, 21.04, 20.81, 20.79, 16.08; HRMS (ESI) calcd. for C₅₁H₅₆N₂O₁₉Na (M + Na)⁺ m/z 1023.3369, found 1023.3375.

Peptide chain elongation



Supplementary Figure 45. Preparation of vinyl C-glycosyl dipeptide 10.

To a stirred solution of **3ba** (83.3 mg, 0.1 mmol) in dry CH₂Cl₂ (1 mL) was added TFA (0.5 mL) at 0 $^{\circ}$ C. The mixture was stirred for 1.5 h at room temperature to remove the Boc group. The solution was then concentrated and dried *in vacuo*. The residue was dissolved in dry DMF (1 mL), and cooled to -10 $^{\circ}$ C. *N*-((Benzyloxy)carbonyl)-*O*-(*tert*-butyl)-L-serine (45 mg, 1.5 equiv.) and HOBt (1.5 equiv.) were dissolved in dry DMF (1 mL), and this solution was transfered into the above solution slowly under -10 $^{\circ}$ C. DIPEA (70 µL, 4.0 equiv.) was injected followed by addition of EDCI (1.5 equiv.). The reaction mixture was stirred under -10 $^{\circ}$ C for 0.5 h, and was then warmed up to room temperature and stirred for 6 h. After removal of solvent, the residue was purified by column chromatography (petroleum ether/ethyl acetate = 1:1) to provide **10** (93.3 mg, 92%) as a white solid.

O-Tert-butyl-*N*-(benzyloxycarbonyl-)-L-serinyl-3-(1-(3,4,6-tri-*O*-benzoyl-2deoxy-2-phthalimido-β-D-glucopyranosyl)eth-1-en-1-yl)-L-alanine methyl ester (10)



[α]_D²⁵ 28.5 (*c* 0.9, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 8.06 (d, J = 7.3 Hz, 2H), 7.91 (d, J = 7.4 Hz, 2H), 7.85 (d, J = 7.4 Hz, 1H), 7.76 (d, J = 7.5 Hz, 2H), 7.70 (d, J = 7.0 Hz, 1H), 7.64 (t, J = 7.3 Hz, 1H), 7.59–7.51 (m, 2H), 7.46 (t, J = 7.4 Hz, 1H), 7.45–7.38 (m, 3H), 7.37– 7.28 (m, 7H), 7.28–7.22 (m, 2H), 7.09 (d, J = 8.2 Hz, 1H), 6.42 (t, J = 9.8 Hz, 1H), 5.72 (t, J = 9.7 Hz, 1H),

5.17–5.00 (m, 4H), 4.91 (s, 1H), 4.78 (td, J = 9.4, 3.9 Hz, 1H), 4.69 (dd, J = 12.1, 2.9 Hz, 1H), 4.65 (t, J = 10.4 Hz, 1H), 4.46 (dd, J = 12.2, 4.5 Hz, 1H), 4.29–4.13 (m, 2H), 3.76 (dd, J = 8.9, 3.9 Hz, 1H), 3.61 (s, 3H), 3.37 (t, J = 8.1 Hz, 1H), 2.84 (dd, J = 15.8, 3.9 Hz, 1H), 2.46 (dd, J = 15.7, 10.1 Hz, 1H), 1.15 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 171.91, 170.18, 168.01, 167.71, 166.21, 165.69, 165.26, 155.89, 139.10, 136.37, 134.46, 134.36, 133.43, 133.31, 133.11, 131.53, 129.87, 129.86, 129.77, 129.69, 128.89, 128.55, 128.41, 128.33, 128.18, 128.14, 123.76, 123.72, 118.13, 79.24, 75.98, 74.08, 71.83, 70.13, 66.91, 63.06, 61.76, 54.32, 52.96, 52.32, 50.84, 33.14, 27.35; HRMS (ESI) calcd. for C₅₆H₅₅N₃O₁₅Na (M + Na)⁺ m/z 1032.3525, found 1032.3527.

Deprotection of ester groups

N-(Tert-butoxycarbonyl)-3-(1-(α-D-mannopyranosyl)eth-1-en-1-yl)-L-alanine (11)



To a Schlenk flasks (10 mL) equipped with a stirr bar were added **3aa** (41.0 mg, 0.05 mmol), LiOH (9 mg, 7.5 equiv.), MeOH (4.0 mL) and deionized water (1.0 mL). The mixture was stirred at room temperature for 10 h, after which, HOAc (24 μ L) was added and the mixture was stirred for 20 mins. After dilution with MeOH (5.0 mL) and concentration, the mixture was purified by column chromatography (CH₂Cl₂/MeOH using a gradient ratio of 10/1, 5/1, 2/1, 1/1) to provide **11** as a white solid (19 mg, 100%): [α]_D²⁵ 31.8 (*c* 0.9, MeOH); ¹H NMR (500 MHz, D₂O) δ 5.22 (d, *J* = 9.9 Hz, 1H), 5.12 (s, 1H), 4.40 (d, *J* = 12.1 Hz, 1H), 4.29 (t, *J* = 2.8 Hz, 1H), 4.02–3.91 (m, 1H), 3.77 (ddd, *J* = 10.9, 9.4, 2.7 Hz, 1H), 3.68 (dd, *J* = 12.3, 6.7 Hz, 1H), 3.59 (t, *J* = 8.9 Hz, 1H), 3.34 (t, *J* = 6.9 Hz, 1H), 2.49–2.33 (m, 2H), 1.35 (s, 9H); ¹³C NMR (126 MHz, D₂O) δ 179.18, 157.29, 140.17, 116.58, 80.90, 78.07, 74.96, 70.66, 68.90, 67.49, 60.95, 54.90, 35.57, 27.51; HRMS (ESI) calcd. for C₁₆H₂₇NO₉Na (M + Na)⁺ m/z 400.1587, found 400.1579.

N-(*Tert*-butoxycarbonyl)-3-(1-(2-deoxy-2-acetylamido-β-D-glucopyranosyl)eth-1en-1-yl)-L-alanine (12)



To a Schlenk flasks (10 mL) equipped with a stirr bar were added **8** (37.5 mg, 0.05 mmol), LiOH (9 mg, 7.5 equiv.), MeOH (4.0 mL) and deionized water (1.0 mL). The mixture was stirred at room temperature for 10 h, after which, HOAc (24 μ L) was added and the mixture was stirred for 20 mins. After dilution with MeOH (5.0 mL) and concentration, the mixture was purified by column chromatography (CH₂Cl₂/MeOH using a gradient ratio of 5/1, 2/1, 1/1, 1/2, pure MeOH) to provide **12** as a white solid (22 mg, 100%): [α]_D²⁵ 10.3 (*c* 1.0, MeOH); ¹H NMR (500 MHz, CD₃OD) δ 5.06 (s, 1H), 4.99 (s, 1H), 4.30 (dd, *J* = 10.1, 3.6 Hz, 1H), 3.88 (t, *J* = 10.0 Hz, 1H), 3.85 (dd, *J* = 12.7, 2.5 Hz, 1H), 3.77 (d, *J* = 10.2 Hz, 1H), 3.70 (dd, *J* = 12.1, 6.0 Hz, 2H), 3.44 (t, *J* = 9.2 Hz, 2H), 3.35 (t, *J* = 10.9 Hz, 1H), 3.26 (ddd, *J* = 9.2, 6.3, 2.1 Hz, 1H), 2.75 (dd, *J* = 15.5, 3.5 Hz, 1H), 2.34 (dd, *J* = 15.4, 10.1 Hz, 1H), 1.91 (s, 3H), 1.42 (s, 9H); ¹³C NMR (125 MHz, CD₃OD) δ 173.53, 157.99, 143.51, 117.88, 84.87, 82.03, 80.02, 77.36, 72.27, 63.21, 55.68, 54.70, 49.03, 35.60, 28.87, 22.95; HRMS (ESI) calcd. for C₁₈H₃₀N₂O₉Na (M + Na)⁺ m/z 441.1844, found 441.1841.

N-(*Tert*-butoxycarbonyl)-3-(1-(α-D-mannopyranosyl)eth-1-en-1-yl)-L-alanyl-Lalanyl-*O*-(*tert*-butyl)-L-threonine (13)



To a Schlenk flasks (10 mL) equipped with a stirr bar were added **3at** (52.0 mg, 0.05 mmol), LiOH (9 mg, 7.5 equiv.), MeOH (4.0 mL) and deionized water (1.0 mL). The mixture was stirred at room temperature for 10 h, after which, HOAc (24 μ L) was added and the mixture was stirred for 20 mins. After dilution with MeOH (5.0 mL) and concentration, the mixture was purified by column chromatography (CH₂Cl₂/MeOH using a gradient ratio of 10/1, 5/1, 2/1, 1/1, 1/2) to provide **13** as a white solid (29 mg, 96%): [α]_D²⁵ -0.7 (*c* 1.0, MeOH); ¹H NMR (500 MHz, CD₃OD) δ 5.21 (s, 1H), 5.17 (s, 1H), 4.49 (q, *J* = 7.0 Hz, 1H), 4.45 (d, *J* = 3.5 Hz, 1H), 4.33 (t, *J* = 7.5 Hz, 1H), 4.28–4.22 (m, 2H), 4.18 (t, *J* = 3.6 Hz, 1H), 3.83 (dd, *J* = 11.9, 6.9 Hz, 1H), 3.77 (dd, *J* = 12.0, 2.9 Hz, 1H), 3.73 (dd, *J* = 7.9, 3.2 Hz, 1H), 3.66 (t, *J* = 7.7 Hz, 1H), 3.42 (td, *J* = 7.3, 2.9 Hz, 1H), 2.53 (d, *J* = 7.4 Hz, 2H), 1.43 (s, 9H), 1.39 (d, *J* = 7.0 Hz, 3H), 1.19 (s, 9H), 1.14 (d, *J* = 6.1 Hz, 3H); ¹³C NMR (12 MHz, CD₃OD) δ 174.38, 174.09, 157.73, 143.19, 117.52, 80.72, 78.46, 78.02, 75.03, 72.72, 70.16, 69.76, 69.40, 62.57, 60.88, 55.01, 50.48, 37.33, 28.94, 28.74, 21.55, 18.37; HRMS (ESI) calcd. for C₂₇H₄₇N₃O₁₂Na (M + Na)⁺ m/z 628.3052, found 628.3054.

2.8 Radical clock experiment



Supplementary Figure 46. Preparation of glycosyl bromide substrate 6.

To a solution of 2-O-allyl-3.4,6-tri-O-benzoyl- α , β -D-glucopyranose²⁶ (533 mg, 1.0 mmol) in pyridine (4 mL) was added Ac₂O (282 µL, 3.0 equiv.) at 0 °C. The mixture was stirred at room temperature for 3 h. After completion, MeOH (1.0 mL) was added and the mixture was kept stirring for 1 h. After concentration, the crude product was purifed by column chromatography (petroleum ether/ethyl acetate = 4:1) to provide 2-*O*-allyl-3,4,6-tri-*O*-benzoyl- α , β -D-glucopyranosyl acetate **S41** (470 mg, 82%) as a white solid: ¹H NMR (500 MHz, CDCl₃) δ 8.01 (d, J = 8.4 Hz, 2H), 7.96 (d, J = 8.2 Hz, 2H), 7.90 (dd, J = 14.5, 8.2 Hz, 2H), 7.57–7.47 (m, 3H), 7.43–7.29 (m, 6H), 6.48 (d, J = 3.6 Hz, 0.6H), 5.89 (t, J = 9.8 Hz, 1H), 5.83 (d, J = 7.9 Hz, 0.4H), 5.76–5.65 (m, 1.4H), 5.59 (dt, J = 19.5, 9.8 Hz, 1H), 5.18 (ddq, J = 18.9, 17.4, 1.6 Hz, 1H), 5.12 (dt, J = 10.3, 1.3 Hz, 0.6H), 5.06 (dq, J = 10.3, 1.3 Hz, 0.4H), 4.57 (ddt, J = 10.3, 7.5, 3.0Hz, 1H), 4.47–4.34 (m, 1H), 4.22–4.16 (m, 1H), 4.15–4.05 (m, 1.4H), 4.02 (ddt, J = 13.0, 6.0, 1.3 Hz, 0.6H), 3.88 (dd, J = 10.0, 3.6 Hz, 0.6H), 3.76 (dd, J = 9.3, 8.0 Hz, 0.4H), 2.26 (s, 1.8H), 2.17 (s, 1.2H); ¹³C NMR (125 MHz,CDCl₃) δ 169.32, 166.23, 165.93, 165.66, 165.44, 134.14, 133.92, 133.56, 133.39, 133.32, 133.23, 133.19, 129.99, 129.98, 129.91, 129.88, 129.85, 129.53, 129.41, 128.95, 128.89, 128.54, 128.53, 128.51, 128.49, 128.45, 118.54, 117.87, 93.95, 89.69, 77.97, 75.97, 74.31, 73.73, 72.91, 72.48, 71.83, 70.28, 69.29, 69.18, 63.04, 62.89, 21.23, 21.16; HRMS (ESI) calcd. for $C_{32}H_{30}O_{10}Na (M + Na)^+ m/z 597.1731$, found 597.1737.

To a stirred solution of **S41** (173 mg, 0.3 mmol) in CH₂Cl₂ (2.0 mL) was added 33% HBr in HOAc (0.5 mL) dropwise under 0 °C. The mixture was kept stirring for 0.5 h at 0 °C, and was then diluted with CH₂Cl₂ (50 mL). The mixture was washed with ice-water (3×20 mL), sat. aq. NaHCO₃ (3×10 mL), and brine (2×20 mL) successively. The organic layer was dried over Na₂SO₄, filtered, and concentrated to give **6** as a white foam, which was in high purity and used directly in the coupling reaction. **6**: ¹H NMR (500 MHz, CDCl₃) δ 8.04 (dd, *J* = 8.4, 1.4 Hz, 2H), 7.94 (ddd, *J* = 8.0, 7.5, 1.4 Hz, 4H), 7.56 (ddt, *J* = 8.8, 7.2, 1.3 Hz, 1H), 7.53–7.47 (m, 2H), 7.43 (dd, *J* = 8.2, 7.5 Hz, 2H), 7.37 (dd, *J* = 16.2, 8.5 Hz, 4H), 6.61 (d, *J* = 3.8 Hz, 1H), 5.99 (t, *J* = 9.6 Hz, 1H), 5.78 (ddt, *J* = 17.3, 10.3, 5.8 Hz, 1H), 5.65 (t, *J* = 10.0 Hz, 1H), 5.25 (dq, *J* = 17.3, 1.5 Hz, 1H), 5.16 (dq, *J* = 10.3, 1.2 Hz, 1H), 4.68–4.60 (m, 2H), 4.49–4.43 (m, 1H), 4.14 (ddt, *J* = 13.0, 5.9, 1.3 Hz, 1H), 4.07 (ddt, *J* = 13.0, 5.8, 1.4 Hz, 1H), 3.78 (dd, *J* = 9.6, 3.8 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 166.14, 165.61, 165.41, 133.74, 133.67, 133.32, 130.05, 129.92, 129.84, 129.66, 129.46, 128.75, 128.57, 128.54, 128.50, 118.84, 89.35, 77.06, 72.68, 72.31, 72.21, 68.28, 62.26.



Supplementary Figure 47. The reaction of bromide 6 with alkyne 2a.

To an oven-dried 10 mL Schlenk tube (Titan, TF891910) containing a Teflon coated magnetic stirring bar were added glycosyl bromide 6 (89.3 mg, 0.15 mmol), NiCl₂(DME) (10 mol%), dtbbpy (12 mol%), (R)-Tol-BINAP (10 mol%), and Na₂CO₃ (2.5 equiv). The tube was sealed with a rubber cap then with parafilm and evacuated then refilled with Ar for at least five cycles. Compound 2a was dissolved in THF (1.5 mL) and injected into the reaction tube. When stirring, PMHS (48 µL, 2.5 equiv.) was injected via microliter syringe. The tube was kept stirring at 30 °C for 36 h. The mixture was diluted with CH₂Cl₂ (20 mL) and filtered. After concentration, the residue was purified by preparative TLC (toluene/ethyl acetate/ $CH_2Cl_2 = 5:0.5:1$) to afford the 7 (36.8 mg, d.r. = 3:2) as a syrup: ¹H NMR (500 MHz, CDCl₃) δ 8.02 (d, J = 6.9 Hz, 2H), 8.01–7.90 (m, 4H), 7.58–7.47 (m, 3H), 7.45–7.32 (m, 6H), 5.75 (dd, J = 8.6, 6.2 Hz, 0.4 H), 5.68 (dd, J = 7.2, 4.6 Hz, 0.6 H), 5.46–5.38 (m, 1H), 5.10 (d, J = 8.1 Hz, 0.4 H), 4.95 (d, J = 8.3 Hz, 0.6 H), 4.91 (s, 0.4 H), 4.89 (s, 0.6 H), 4.87 (dd, J = 11.6, 4.3 Hz, 0.4 H), 4.84 (s, 0.4 H), 4.76 (s, 0.6 H), 4.70 (dd, J = 12.1, 6.9 Hz, 0.4 H), 4.56 (t, J = 4.2 Hz, 1H), 4.50 (dd, J = 12.4, 3.7 Hz, 0.6 H), 4.46 (dd, J = 11.4, 4.1 Hz, 0.6 H), 4.46 (m, 0.4 H), 4.43–4.33 (m, 1.6 H), 4.31–4.27 (m, 1 H), 4.24 (t, J = 5.8 Hz, 0.4 H), 4.03 (t, J = 7.8 Hz, 0.6 H), 3.77 (dd, J = 10.8, 7.9 Hz, 0.6 H), 3.54 (dd, J = 9.0, 5.5 Hz, 0.4 H), 2.69–2.42 (m, 2H), 2.40–2.30 (m, 1H), 2.30–2.20 (m, 1H), 2.19–2.09 (m, 1H), 1.42 (s, 3.6 H), 1.41 (s, 5.4 H); ¹³C NMR (125 MHz, CDCl₃) δ 172.99, 166.31, 166.29, 165.77, 165.74, 165.43, 154.11, 142.33, 141.83, 133.53, 133.45, 133.35, 133.31, 133.29, 129.99, 129.95, 129.91, 129.87, 129.84, 129.79, 129.76, 129.74, 129.46, 129.38, 129.24, 129.08, 128.57, 128.54, 128.52, 128.51, 128.46, 115.39, 114.82, 80.33, 80.08, 79.04, 78.82, 77.16, 73.63, 73.07, 72.60, 72.08, 71.81, 71.57, 68.60, 67.78, 63.06, 62.48, 52.40, 52.35, 52.06, 51.99, 43.28, 42.01, 39.44, 38.97, 37.25, 30.33, 28.40; HRMS (ESI) calcd. for $C_{41}H_{45}NO_{12}Na (M + Na)^+ m/z$ 766.2834, found 766.2835.

3. NMR Spectra

Supplementary Figure 48. ¹H NMR spectrum of compound 1c (500 MHz, CDCl₃, 25 °C)





Supplementary Figure 49. ¹H NMR spectrum of compound 1d (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 50. ¹³C NMR spectrum of compound 1d (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 51. ¹H NMR spectrum of compound 1e (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 52. ¹³C NMR spectrum of compound **1e** (125 MHz, CDCl₃, 25 °C) **1e**





Supplementary Figure 53. ¹H NMR spectrum of compound S3 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 54. ¹³C NMR spectrum of compound S3 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 55. ¹H NMR spectrum of compound 1g (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 56. ¹³C NMR spectrum of compound 1g (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 57. ¹H NMR spectrum of compound **S6** (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 58. ¹³C NMR spectrum of compound S6 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 59. ¹H NMR spectrum of compound 1h (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 60. ¹³C NMR spectrum of compound 1h (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 61. ¹H NMR spectrum of compound **S9** (500 MHz, CDCl₃, 25 °C) **S9**

Supplementary Figure 62. ¹³C NMR spectrum of compound S9 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 64. ¹³C NMR spectrum of compound S11 (125 MHz, CDCl₃, 25 °C) S11



Supplementary Figure 63. ¹H NMR spectrum of compound S11 (500 MHz, CDCl₃, 25 °C)



Supplementary Figure 65. ¹H NMR spectrum of compound 1j (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 66. ¹³C NMR spectrum of compound 1j (125 MHz, CDCl₃, 25 °C)



Supplementary Figure 67. ¹H NMR spectrum of compound S14 (500 MHz, CDCl₃, 25 °C) S14



Supplementary Figure 68. ¹³C NMR spectrum of compound S14 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 69. ¹H NMR spectrum of compound S16 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 70. ¹³C NMR spectrum of compound S16 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 71. ¹H NMR spectrum of compound S17 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 72. ¹³C NMR spectrum of compound S17 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 73. ¹H NMR spectrum of compound 1k (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 74. ¹³C NMR spectrum of compound 1k (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 75. ¹H NMR spectrum of compound S19 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 76. ¹³C NMR spectrum of compound S19 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 77. ¹H NMR spectrum of compound 11 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 78. ¹³C NMR spectrum of compound 11 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 79. ¹H NMR spectrum of compound S23 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 80. ¹³C NMR spectrum of compound S23 (125 MHz, CDCl₃, 25 °C) S23





Supplementary Figure 81. ¹H NMR spectrum of compound S24 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 82. ¹³C NMR spectrum of compound S24 (125 MHz, CDCl₃, 25 °C) S24





Supplementary Figure 83. ¹H NMR spectrum of compound S25 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 84. ¹³C NMR spectrum of compound S25 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 85. ¹H NMR spectrum of compound S26 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 86. ¹³C NMR spectrum of compound S26 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 87. ¹H NMR spectrum of compound 1m (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 88. ¹³C NMR spectrum of compound 1m (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 89. ¹H NMR spectrum of compound S27 (500 MHz, CDCl₃, 25 °C)





S28 170.80 169.83 165.90 65.38 65.07 131.51 130.08 130.04 129.99 129.97 129.93 129.80 129.65 129.54 129.53 129.47 129.42 129.41 128.91 128.85 128.71 128.71 128.48 128.48 128.37 128.34 128.34 167.75 65.71 34.17 77.16 70.52 70.28 21.12 20.92 20.87 20.68 17.78 17.65 17.49 17.28 BZC óвz BZO NPhth BzO BzÖ ∖ ÓAc OAc BZO BzÓ ÓBz 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0

Supplementary Figure 91. ¹³C NMR spectrum of compound S28 (125 MHz, CDCl₃, 25 °C)



Supplementary Figure 92. ¹H NMR spectrum of compound S31 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 93. ¹³C NMR spectrum of compound S31 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 94. ¹H NMR spectrum of compound S32 (500 MHz, CDCl₃, 25 °C)



Supplementary Figure 95. ¹³C NMR spectrum of compound S32 (125 MHz, CDCl₃, 25 °C)



Supplementary Figure 96. ¹H NMR spectrum of compound S33 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 97. ¹³C NMR spectrum of compound S33 (500 MHz, CDCl₃, 25 °C)





Supplementary Figure 98. ¹H NMR spectrum of compound 2a (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 99. ¹H NMR spectrum of compound 2b (500 MHz, CDCl₃, 25 °C)





Supplementary Figure 100. ¹H NMR spectrum of compound 2c (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 101. ¹H NMR spectrum of compound 2d (500 MHz, CDCl₃, 25 °C)




Supplementary Figure 102. ¹H NMR spectrum of compound 2e (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 103. ¹H NMR spectrum of compound 2f (500 MHz, CDCl₃, 25 °C) 2f





Supplementary Figure 104. ¹H NMR spectrum of compound 2g (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 105. ¹H NMR spectrum of compound **2h** (500 MHz, CDCl₃, 25 °C) **2h**





Supplementary Figure 106. ¹H NMR spectrum of compound 2i (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 107. ¹³C NMR spectrum of compound **2i** (125 MHz, CDCl₃, 25 °C) **2i**





Supplementary Figure 108. ¹H NMR spectrum of compound 2j (500 MHz, CDCl₃, 25 °C)



Supplementary Figure 109. ¹H NMR spectrum of compound 2k (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 110. ¹³C NMR spectrum of compound 2k (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 111. ¹H NMR spectrum of compound 2l (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 112. ¹³C NMR spectrum of compound **2l** (125 MHz, CDCl₃, 25 °C) **2l**





Supplementary Figure 113. ¹H NMR spectrum of compound 2m (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 114. ¹³C NMR spectrum of compound 2m (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 115. ¹H NMR spectrum of compound 2n (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 116. ¹³C NMR spectrum of compound **2n** (125 MHz, CDCl₃, 25 °C) **2n**





Supplementary Figure 117. ¹H NMR spectrum of compound 20 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 118. ¹³C NMR spectrum of compound **20** (125 MHz, CDCl₃, 25 °C) **20**





Supplementary Figure 119. ¹H NMR spectrum of compound 2p (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 120. ¹³C NMR spectrum of compound **2p** (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 121. ¹H NMR spectrum of compound 2q (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 122. ¹³C NMR spectrum of compound 2q (125 MHz, CDCl₃, 25 °C) 2q





Supplementary Figure 123. ¹H NMR spectrum of compound 2r (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 124. ¹³C NMR spectrum of compound 2r (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 125. ¹H NMR spectrum of compound 2s (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 126. ¹³C NMR spectrum of compound **2s** (125 MHz, CDCl₃, 25 °C) **2s**





Supplementary Figure 127. ¹H NMR spectrum of compound 2t (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 128. ¹³C NMR spectrum of compound **2t** (125 MHz, CDCl₃, 25 °C) **2t**





Supplementary Figure 129. ¹H NMR spectrum of compound 2u (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 130. ¹³C NMR spectrum of compound 2u (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 131. ¹H NMR spectrum of compound 2v (500 MHz, CDCl₃, 25 °C)



Supplementary Figure 132. ¹H NMR spectrum of compound 3a-1 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 133. ¹³C NMR spectrum of compound 3a-1 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 134. ¹H NMR spectrum of compound 4a (500 MHz, CDCl₃, 25 °C)



Supplementary Figure 135. ¹H NMR spectrum of compound 3b-1 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 136. ¹³C NMR spectrum of compound 3b-1 (125 MHz, CDCl₃, 25 °C) 3b-1





Supplementary Figure 137. ¹H NMR spectrum of compound 5a (500 MHz, CDCl₃, 25 °C)



Supplementary Figure 138. ¹H NMR spectrum of compound 3b-2 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 139. ¹³C NMR spectrum of compound 3b-2 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 140. ¹H NMR spectrum of compound 4b (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 141. ¹³C NMR spectrum of compound 4b (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 142. ¹H NMR spectrum of compound 5b (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 143. ¹³C NMR spectrum of compound 5b (125 MHz, CDCl₃, 25 °C)



Supplementary Figure 144. ¹H NMR spectrum of (*R*)-Tol-BINAP(O)₂ (500 MHz, CDCl₃, 25 °C) **di-oxidized (R)-Tol-BINAP(O)2**



Supplementary Figure 145. ³¹P NMR spectrum of (*R*)-Tol-BINAP(O)₂ (162 MHz, CDCl₃, 25 °C) **di-oxidized (R)-Tol-BINAP(O)2**







Supplementary Figure 147. ¹³C NMR spectrum of compound 3aa (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 148. ¹H NMR spectrum of compound 3ab (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 149. ¹³C NMR spectrum of compound 3ab (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 150. ¹H NMR spectrum of compound 3ac (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 151. ¹³C NMR spectrum of compound 3ac (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 152. ¹H NMR spectrum of compound 3ad (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 153. ¹³C NMR spectrum of compound 3ad (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 154. ¹H NMR spectrum of compound **3ae** (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 155. ¹³C NMR spectrum of compound 3ae (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 156. ¹H NMR spectrum of compound 3af (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 157. ¹³C NMR spectrum of compound 3af (125 MHz, CDCl₃, 25 °C)







Supplementary Figure 159. ¹³C NMR spectrum of compound **3ag** (125 MHz, CDCl₃, 25 °C) **3ag**







Supplementary Figure 161. ¹³C NMR spectrum of compound **3ah** (125 MHz, CDCl₃, 25 °C) **3ah**





Supplementary Figure 162. ¹H NMR spectrum of compound 3ai (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 163. ¹³C NMR spectrum of compound 3ai (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 164. ¹H NMR spectrum of compound 3aj (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 165. ¹³C NMR spectrum of compound 3aj (125 MHz, CDCl₃, 25 °C) 3aj







Supplementary Figure 167. ¹³C NMR spectrum of compound 3ak (500 MHz, CDCl₃, 25 °C)







Supplementary Figure 169. ¹³C NMR spectrum of compound 3al (125 MHz, CDCl₃, 25 °C)






Supplementary Figure 171. ¹³C NMR spectrum of compound 3am (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 172. ¹H NMR spectrum of compound 3an (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 173. ¹³C NMR spectrum of compound 3an (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 174. ¹H NMR spectrum of compound 3ao (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 175. ¹³C NMR spectrum of compound 3ao (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 176. ¹H NMR spectrum of compound 3ap (500 MHz, CDCl₃, 25 °C)



Supplementary Figure 177. ¹³C NMR spectrum of compound 3ap (125 MHz, CDCl₃, 25 °C)



Supplementary Figure 178. ¹H NMR spectrum of compound (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 179. ¹³C NMR spectrum of compound 3aq (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 180. ¹H NMR spectrum of compound 3ar (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 181. ¹³C NMR spectrum of compound 3ar (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 182. ¹H NMR spectrum of compound 3as (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 183. ¹³C NMR spectrum of compound 3as (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 184. ¹H NMR spectrum of compound 3at (500 MHz, CDCl₃, 25 °C)



Supplementary Figure 185. ¹³C NMR spectrum of compound 3at (125 MHz, CDCl₃, 25 °C)



Supplementary Figure 186. ¹H NMR spectrum of compound 3ba (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 187. ¹³C NMR spectrum of compound 3ba (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 188. ¹H NMR spectrum of compound 3bl (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 189. ¹³C NMR spectrum of compound 3bl (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 190. ¹H NMR spectrum of compound 3bt (500 MHz, CDCl₃, 25 °C)







Supplementary Figure 192. ¹H NMR spectrum of compound 3bu (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 193. ¹³C NMR spectrum of compound 3bu (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 194. ¹H NMR spectrum of compound 3bp (500 MHz, CDCl₃, 25 °C)



Supplementary Figure 195. ¹³C NMR spectrum of compound 3bp (125 MHz, CDCl₃, 25 °C)



Supplementary Figure 196. ¹H NMR spectrum of compound **3ca** (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 197. ¹³C NMR spectrum of compound 3ca (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 198. ¹H NMR spectrum of compound **3da-**β (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 199. ¹³C NMR spectrum of compound **3da-**β (125 MHz, CDCl₃, 25 °C)



Supplementary Figure 200. ¹H-COSY spectrum of compound **3da-**β (500 MHz, CDCl₃, 25 °C) Key COSY correlations







3da-β-COSY

Supplementary Figure 201. HSQC NMR spectrum of compound $3da-\beta$ (CDCl₃) Key HSQC correlations



3da-
$$\beta$$
, ⁴C₁(β), R = 4-CF₃-PhCO



3da-β-HSQC







Supplementary Figure 203. ¹³C NMR spectrum of compound **3da-***α* (150 MHz, CDCl₃, 25 °C)

Supplementary Figure 204. ¹H-COSY spectrum of compound **3da-***α* (500 MHz, CDCl₃, 25 °C) Key COSY correlations







Supplementary Figure 205. HSQC NMR spectrum of compound $3da-\alpha$ (CDCl₃)



Supplementary Figure 206. NOE NMR spectrum of compound **3da-***α* (CDCl₃) Key NOE correlations







Supplementary Figure 207. ¹H NMR spectrum of compound **3ea** (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 208. ¹³C NMR spectrum of compound **3ea** (125 MHz, CDCl₃, 25 °C) **3ea**





Supplementary Figure 209. ¹H NMR spectrum of compound 3fa (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 210. ¹³C NMR spectrum of compound 3fa (125 MHz, CDCl₃, 25 °C)



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Supplementary Figure 211. ¹H COSY NMR spectrum of compound 3fa (500 MHz, CDCl₃, 25 °C) Key COSY correlations





Supplementary Figure 212. HSQC NMR spectrum of compound 3fa (CDCl₃)







Supplementary Figure 213. ¹H NMR spectrum of compound 3ga (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 214. ¹³C NMR spectrum of compound 3ga (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 215. ¹H NMR spectrum of compound 3ha (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 216. ¹³C NMR spectrum of compound **3ha** (125 MHz, CDCl₃, 25 °C) **3ha**







Supplementary Figure 218. ¹³C NMR spectrum of compound **3ia**β (125 MHz, CDCl₃, 25 °C) **3ia-b**





Supplementary Figure 219. ¹H NMR spectrum of compound **3ia**α (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 220. ¹³C NMR spectrum of compound 3iaα (125 MHz, CDCl₃, 25 °C)

1.12

4.5

8 8

5.5

93 66

5.0

88

6.0

3

2.15

8.0

 Ξ

2 8.5 2.30 2.30 2.23 2.30 2.39

7.0

6.5

7.5

۲

2.61

3.5

4.0

۶H

1.11

2.5

2.0

3.0

0.73H

1.5

1.0

0.5

0.0

-0.5





Supplementary Figure 221. ¹H NMR spectrum of compound 3ja (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 222. ¹³C NMR spectrum of compound 3ja (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 223. ¹H NMR spectrum of compound 3ka (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 224. ¹³C NMR spectrum of compound 3ka (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 225. ¹H NMR spectrum of compound 3la (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 226. ¹³C NMR spectrum of compound 3la (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 227. ¹H NMR spectrum of compound 3ma (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 228. ¹³C NMR spectrum of compound 3ma (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 229. ¹H NMR spectrum of compound 3av1 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 230. ¹³C NMR spectrum of compound 3av1 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 231. ¹H NMR spectrum of compound 3av2 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 232. ¹³C NMR spectrum of compound **3av2** (125 MHz, CDCl₃, 25 °C) **3av2**







Supplementary Figure 234. ¹³C NMR spectrum of compound 3bv1 (125 MHz, CDCl₃, 25 °C)




Supplementary Figure 235. ¹H NMR spectrum of compound **3bv2** (500 MHz, CDCl₃, 25 °C) **3bv2**

Supplementary Figure 236. ¹³C NMR spectrum of compound **3bv2** (125 MHz, CDCl₃, 25 °C) **3bv2**





Supplementary Figure 238. ¹³C NMR spectrum of compound **S41** (125 MHz, CDCl₃, 25 °C) **S41**



Supplementary Figure 237. ¹H NMR spectrum of compound **S41** (500 MHz, CDCl₃, 25 °C) **S41**



Supplementary Figure 239. ¹H NMR spectrum of compound **6** (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 240. ¹³C NMR spectrum of compound 6 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 242. ¹³C NMR spectrum of compound 7 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 243. ¹H NMR spectrum of compound **8** (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 244. ¹³C NMR spectrum of compound 8 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 245. ¹H NMR spectrum of compound **8-***R* (500 MHz, CDCl₃, 25 °C) **8-***R*

Supplementary Figure 246. ¹³C NMR spectrum of compound 8-*R* (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 247. ¹H NMR spectrum of compound S39 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 248. ¹³C NMR spectrum of compound S39 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 249. ¹H NMR spectrum of compound 9 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 250. ¹³C NMR spectrum of compound 9 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 251. ¹H NMR spectrum of compound 10 (500 MHz, CDCl₃, 25 °C)

Supplementary Figure 252. ¹³C NMR spectrum of compound 10 (125 MHz, CDCl₃, 25 °C)





Supplementary Figure 253. ¹H NMR spectrum of compound 11 (500 MHz, D₂O, 25 °C)

Supplementary Figure 254. ¹³C NMR spectrum of compound 11 (125 MHz, D₂O, 25 °C)





Supplementary Figure 255. ¹H NMR spectrum of compound 12 (500 MHz, CD₃OD, 25 °C)

Supplementary Figure 256. ¹³C NMR spectrum of compound 12 (125 MHz, CD₃OD, 25 °C)





Supplementary Figure 257. ¹H NMR spectrum of compound 13 (500 MHz, CD₃OD, 25 °C)

Supplementary Figure 258. ¹³C NMR spectrum of compound **13** (125 MHz, CD₃OD, 25 °C) 13





14 170.78 169.97 167.78 165.91 165.71 165.38 165.38 $\begin{array}{c} 134.15\\ 133.58\\ 133.55\\ 133.55\\ 133.57\\ 133.157\\ 131.57\\ 131.57\\ 131.57\\ 133.00\\ 133.00\\ 133.00\\ 133.00\\ 133.00\\ 129.93\\ 129.95\\ 129.45\\ 129.55\\ 129.63\\ 129.54\\ 129.57\\ 129.55\\ 129.55\\ 129.56$ MeO₂C B7O BzO BzÓ ∖ ÓAc OAc BZO 7 BzO ÓBz Acetone 0 180 170 160 150 140 130 120 110 100 **9**0 80 70 60 50 40 30 20 10

Supplementary Figure 260. ¹³C NMR spectrum of compound 14 (150 MHz, CDCl₃, 25 °C)



Supplementary Figure 261. ¹H NMR spectrum of compound 15 (600 MHz, CDCl₃, 25 °C)



Supplementary Figure 262. ¹³C NMR spectrum of compound 15 (150 MHz, CDCl₃, 25 °C)

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