Supporting Information

Synthesis and Antibacterial Activity of Propylamycin Derivatives Functionalized at the 5"- and Other Positions with a View to Overcoming Resistance due to Aminoglycoside Modifying Enzymes.

Dimitrijs Lubriks,^a Rimants Zogota,^a Vikram A Sarpe,^{b,c} Takahiko Matsushita,^d Girish C. Sati,^d Klara Haldimann,^e Marina Gysin,^e Erik C. Böttger,^e Andrea Vasella,^f Edgars Suna,^{a,*} Sven N. Hobbie,^{e,*} and David Crich.^{b,c,d,g,*}

- a) Latvian Institute of Organic Synthesis, Riga, Latvia, LV-1006
- b) Department of Pharmaceutical and Biomedical Sciences, University of Georgia, 250 West Green Street, Athens, GA 30602, USA
- c) Complex Carbohydrate Research Center, University of Georgia, 315 Riverbend Road, Athens, GA 30602, USA
- d) Department of Chemistry, Wayne State University, 5101 Cass Avenue, Detroit, MI 48202, USA
- e) Institute of Medical Microbiology, University of Zurich, Gloriastrasse 28, 8006 Zürich, Switzerland
- f) Organic Chemistry Laboratory, ETH Zürich, Vladimir-Prelog-Weg 1-5/10, 8093 Zürich, Switzerland
- g) Department of Chemistry, University of Georgia, 140 Cedar Street, Athens, GA 30602, USA
- * Edgars@osi.lv
- * Shobbie@imm.uzh.ch
- * David.crich@uga.edu

Table of Contents

	Expt	Spectra
General Experimental.	S5	
5"-O-tert-Butyldimethylsilyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-	S5	S48-S50
trifluoroacetyl paromomycin (17).		
6,3',2"',3"'',4"''-Penta-O-benzoyl-5"'-O-tert-butyldimethylsilyl-4',6'-O-	S6	S51-S53
benzylidene-1,3,2',2''',6'''-penta- <i>N</i> -trifluoroacetyl paromomycin (18)		
6,3',2'',3''',4'''-Penta-O-benzoyl-4',6'-O-benzylidene-1,3,2',2''',6'''-penta-N-	S7	S54-S56
trifluoroacetyl paromomycin (19).		
5"-Deoxy-5"-azido-6,3',2",3",4""-penta-O-benzoyl-4',6'-O-benzylidene-	S8	S57-S59
1,3,2',2''',6'''-penta-N-trifluoroacetyl paromomycin (20).		
5"-Deoxy-5"-amino-6,3',2",3",4"'-penta-O-benzoyl-4',6'-O-benzylidene-	S10	S60-S62
1,3,2',2''',6'''-penta-N-trifluoroacetyl paromomycin (21).		
5"-Deoxy-5"-(O-benzylcarbamoyl)-6,3',2",3",4"'-penta-O-benzoyl-4',6'-O-	S11	S63-S65
benzylidene-1,3,2',2"',6"'-penta- <i>N</i> -trifluoroacetyl paromomycin (22).		
5"-Deoxy-5"-(O-benzylcarbamoyl)-6,3',2",3",4"'-penta-O-benzoyl-	S12	S66-S68
1,3,2',2''',6'''-penta-N-trifluoroacetyl paromomycin (23).		
5"-Deoxy-5"-(O-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-O-benzoyl-	S13	S69-S76
1,3,2',2''',6'''-penta-N-trifluoroacetyl paromomycin (24).		
4',5''-Dideoxy-5''-(O-benzylcarbamoyl)-6,3',6',2'',3''',4'''-hexa-O-benzoyl-4'-	S14	S77-S84
iodo-1,3,2',2''',6'''-penta- <i>N</i> -trifluoroacetyl 4'- <i>epi</i> -paromomycin (25).		
4'-Allyl-4',5"-dideoxy-5"-(O-benzylcarbamoyl)-6,3',6',2",3"",4"'-hexa-O-	S15	S85-S89
benzoyl-1,3,2',2''',6'''-penta-N-trifluoroacetyl paromomycin (26) by allylation		
of 25.		
General Procedure A for the photochemically initiated radical allylation of iodo	S17	
paromomycin derivatives 25 and 29		
4',5"-Dideoxy-5"-(O-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-O-benzoyl-	S17	S90-S94
1,3,2',2''',6'''-penta-N-trifluoroacetyl paromomycin-4'-ene (27).		
5"-Deoxy-5"-(O-benzylcarbamoyl)-6,3',6',2",3"'',4"''-hexa-O-benzoyl-	S18	S95-S102
1,3,2',2''',6'''-penta-N-trifluoroacetyl 4'- <i>epi</i> -paromomycin (28).		

4',5"-Dideoxy-5"-(O-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-O-benzoyl-4'-	S19	S103-S110
iodo-1,3,2',2''',6'''-penta-N-trifluoroacetyl paromomycin (29).		
4'-Allyl-4',5"-dideoxy-5"-(O-benzylcarbamoyl)-6,3',6',2",3"",4"''-hexa-O-	S20	
benzoyl-1,3,2',2"',6"''-penta-N-trifluoroacetyl paromomycin (26) by allylation		
of 29.		
4',5"-Dideoxy-5"-(O-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-O-benzoyl-	S21	S111-S115
1,3,2',2''',6'''-penta-N-trifluoroacetyl paromomycin (30-H).		
4',5"-Dideoxy-4'-propyl-5"-amino-6,3',6',2",3"",4"'-hexa-O-benzoyl-	S21	S116-S118
1,3,2',2''',6'''-penta-N-trifluoroacetyl paromomycin (31).		
2,2,2-Trifluoro-N-(2-hydroxyethyl)acetamide (S1).	S22	S119
2,2,2-Trifluoro-N-(2-oxoethyl)acetamide (32).	S23	S120
2,2,2-Trifluoro-N-(3-oxopropyl)acetamide (33).	S23	S121
General Procedure B for reductive amination of amine 31 with aldehydes 32 and	S24	
33.		
4',5"-Dideoxy-4'-propyl-5"-(2-trifluoroacetamidoethylamino)-	S24	
6,3',6',2'',3''',4'''-hexa-O-benzoyl-1,3,2',2''',6'''-penta-N-trifluoroacetyl		
paromomycin (34).		
4',5"-Dideoxy-4'-propyl-5"-(3-trifluoroacetamidopropylamino)-	S24	
6,3',6',2'',3''',4'''-hexa-O-benzoyl-1,3,2',2''',6'''-penta-N-trifluoroacetyl		
paromomycin (35).		
General Procedure C for trifluoroacetylation of amines 34 and 35.	S25	
4',5"-Dideoxy-4'-propyl-5"-(2-aminoethylamino)- 6,3',6',2",3"',4"'-hexa-O-	S25	S122-S124
benzoyl-hepta-N-trifluoroacetyl paromomycin (36).		
4',5"-Dideoxy-4'-propyl-5"-(3-aminoethylamino)- 6,3',6',2",3"',4"'-hexa-O-	S26	S125-S127
benzoyl-hepta-N-trifluoroacetyl paromomycin (37).		
4',5"-Dideoxy-4'-propyl-5"-(O-benzylcarbamoyl)-6,3',6',2",3"",4"'-hexa-O-	S27	S128-S133
benzoyl-1,3,2',2''',6'''-penta-N-trifluoroacetyl paromomycin (38).		
4',5"-Dideoxy-4'-propyl-5"-formamido-6,3',6',2",3"',4"'-hexa-O-benzoyl-	S28	S134-S139
1,3,2',2''',6'''-penta-N-trifluoroacetyl paromomycin (39).		
4',5"-Dideoxy-4'-propyl-5"-(N-benzylureido)-6,3',6',2",3"',4"'-hexa-O-benzoyl-	S29	S140-S145
1,3,2',2''',6'''-penta-N-trifluoroacetyl paromomycin (40).		

4',5"-Dideoxy-4'-propyl-5"-(N-trifluoroacetylglycinamido)-6,3',6',2",3"',4"'-	S30	S146-S150
hexa-O-benzoyl-1,3,2',2''',6'''-penta-N-trifluoroacetyl paromomycin (41).		
General Procedure D for global deprotection of trifluoroacetamides 36, 37, 39	S31	
and 41.		
General Procedure E for global deprotection of trifluoroacetamides 38 and 40.	S32	
5"-Amino-4',5"-dideoxy-4'-propyl paromomycin pentaacetate (8).	S33	S151-S154
5"-(2-Aminoethylamino)-4',5"-dideoxy-4'-propyl paromomycin heptaacetate	S33	S155-S159
(9).		
5"-(3-Aminopropylamino)-4',5"-dideoxy-4'-propyl paromomycin heptaacetate	S34	S160-S164
(10).		
4',5"-Dideoxy-5"-formamido-4'-propyl paromomycin pentaacetate (11).	S35	S165-S168
4',5"-Dideoxy-4'-propyl-5"-ureido-paromomycin pentaacetate (12).	S36	S169-S172
4',5"-Dideoxy-5"-glycinamido-4'-propyl paromomycin hexacetate (13).	S36	S173-S176
Phenyl 2-azido-2,3,4-trideoxy-4-C-ethyl-1-thio-β-D-glucopyranoside (43).	S37	S177-S178
Phenyl 6- <i>O</i> -acetyl-2-azido-2,3,4-trideoxy-4- <i>C</i> -ethyl-1-thio-β-D-glucopyranoside	S38	S179-S181
(44).		
Phenyl 6-O-acetyl-2-azido-2,3,4-trideoxy-4-C-ethyl-1-thio-β-D-glucopyranosyl	S39	S182-S184
sulfoxide (45).		
1,3,2',3',4',2''',6'''-Heptadeoxy-4'-propyl-6'-acetyl-1,3,2',2''',6'''-pentaazido-	S40	S185-S190
6,2",5",3"',4"'-penta-O-benzyl paromomycin (47).		
3',4'-Dideoxy-4'-propyl paromomycin pentaacetate (14).	S41	S191-S196
1,3,2',2 ^{'''} ,6 ^{'''} -Penta- <i>N</i> -benzyloxycarbonyl paromomycin (48).	S41	
6',5"-Bis-O-(2,4,6-triisopropylbenzenesulfonyl)-1,3,2',2"', 6"'-penta-N-	S42	
benzyloxycarbonyl paromomycin (49).		
6',5"-Dideoxy-bis-N-(2-hydroxyethyl)-1,3,2',2"',6"'-penta-N-benzyloxycarbonyl	S43	
paromomycin (50).		
4',6',5"-Trideoxy-6',5"-[bis-N-(2-hydroxyethyl)]-4'-propyl paromomycin	S43	S197-S202
heptaacetate (15).		
Table S1. Bacterial strains used in this study	S44	
Cell-free translation inhibition graphs	S45	
References	S47	

General Experimental

All experiments were carried out under a dry argon atmosphere unless otherwise specified. All reagents and solvents were purchased from commercial suppliers and were used without further purification unless otherwise specified. Chromatographic purifications were carried over silica gel (230-400 mesh). Thin layer chromatography was performed with precoated glass backed plates (w/UV 254). TLC plates were visualized by UV irradiation (254 nm) and by charring with sulfuric acid in ethanol (20:80, v/v) or with ceric ammonium molybdate solution [Ce(SO₄)2: 4 g, (NH₄)₆Mo₇O₂₄: 10 g, H₂SO₄: 40 mL, H₂O: 360 mL]. Optical rotations were measured at 589 nm and 21 °C on a digital polarimeter with a path length of 10 cm. ¹H and ¹³C NMR spectra of all compounds were recorded using at 600 MHz unless otherwise specified and assignments made with the help of COSY, HMBC, and HSQC spectra. ESIHRMS were recorded using a time-of-flight mass spectrometer fitted with an electrospray source.



5"-O-tert-Butyldimethylsilyl-4',6'-O-benzylidene-1,3,2',2''',6'''penta-N-trifluoroacetyl paromomycin (17). An oven-dried 500 mL round-bottom flask was cooled under a stream of argon, equipped with a stirring bar and charged with 4',6'-O-benzylidene-penta-*N*trifluoroacetyl paromomycin **16** (52.6 g, 44.4 mmol, 1.0 equiv),

followed by anhydrous DMF (150 mL). Imidazole (7.56 g, 111 mmol, 2.5 equiv) was added to the stirred solution at ambient temperature, after which a solution of *tert*-butyldimethylsilyl chloride (10.4 g, 68.8 mmol, 1.5 equiv) in anhydrous DMF (50 mL) was added dropwise over a period of 15 min. After stirring at ambient temperature for 3 h (the reaction progress was monitored by UPLC-MS assay), water (800 mL) was added to the light yellowish solution and product was back-extracted with EtOAc (3×400 mL). The combined organic extracts were washed with aqueous HCl (0.1 M solution in water, 400 mL), water (400 mL), saturated aqueous NaHCO₃ solution (400 mL), then water (400 mL) and, finally brine (300 mL). After drying over anhydrous Na₂SO₄, all volatiles were removed under reduced pressure. The yellow residue was taken up in EtOAc (100 mL) and the solution was slowly poured over 20 min to the stirred hexane (2.0 L) to give a precipitate that was collected by suction filtration. The obtained white powder of **17** (54.9 g, 95 % yield) was dried under reduced pressure and used in subsequent step without further purification. [α]²³_D +9.2 (*c* 1.08, MeOH).

¹**H NMR** (400 MHz, CD₃CN) δ: 8.01 (d, *J* = 9.0 Hz, 1H), 7.81 (d, *J* = 9.2 Hz, 1H), 7.66 (d, *J* = 9.0 Hz, 2H), 7.52 – 7.43 (m, 3H), 7.41 – 7.35 (m, 3H), 5.55 (s, 1H), 5.47 (d, *J* = 3.8 Hz, 1H), 5.05 (d, *J* = 1.9 Hz, 1H), 5.00 (d, *J* = 5.4 Hz, 1H), 4.34 (d, *J* = 3.0 Hz, 1H), 4.24 – 4.11 (m, 4H), 4.08 – 3.97 (m, 5H), 3.92 – 3.81 (m, 3H), 3.76 – 3.62 (m, 8H), 3.59 – 3.39 (m, 6H), 2.01 (dt, *J* = 12.9, 4.6 Hz, 1H), 1.64 (q, *J* = 12.9 Hz, 1H), 0.90 (s, 9H), 0.08 (s, 3H), 0.07 (s, 3H) ppm.

¹³C{¹H} NMR (101 MHz, CD₃CN) δ: 158.1 – 157.4 (5×COCF₃), 138.9, 130.0, 129.2, 129.1, 127.4, 126.3, 115.8
– 115.5 (5×CF₃), 110.4, 102.5, 99.7, 98.2, 86.7, 84.5, 82.3, 79.4, 75.5, 74.1, 72.3, 69.7, 69.1, 68.6, 68.5, 64.6, 63.9, 55.8, 52.3, 50.7, 50.2, 41.3, 32.6, 26.3, 18.9, -5.3, -5.4 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+Na]⁺ Calcd for C₄₆H₅₈N₅O₁₉F₁₅NaSi 1320.3153; Found 1320.3129.



4',6'-O-benzylidene-1,3,2',2''',6'''-penta-N-trifluoroacetyl paromomycin (18). To a stirred solution of 5''-O-TBDMS paromomycin derivative **17** (54.9 g, 42.3 mmol, 1.0 equiv) in anhydrous pyridine (500 mL) at 0 °C (crushed ice bath) was added

6,3',2"',3"'',4"''-Penta-O-benzoyl-5"'-O-tert-butyldimethylsilyl-

DMAP (1.03 g, 8.46 mmol, 0.2 equiv), followed by benzoic anhydride (57.4 g, 254 mmol, 6.0 equiv). The resulting pale yellow solution was heated up to 70 °C and stirred for 40 h, and the reaction progress was monitored by UPLC-MS assay. The resulting dark red suspension was cooled to room temperature, concentrated under reduced pressure and additionally co-evaporated twice with toluene (2×200 mL). The brown oily residue was dissolved in EtOAc (600 mL) and washed with water (400 mL). The aqueous layer was back-extracted with EtOAc (2×200 mL). The combined organic extracts were washed with aqueous HCl (0.1 M solution in water, 200 mL), water (200 mL), saturated aqueous NaHCO₃ solution (400 mL), then water (400 mL) and, finally brine (400 mL). Drying over anhydrous Na₂SO₄, filtration and concentration to dryness under reduced pressure afforded brown oil that was filtered through a silica gel pad using gradient elution from 10 % EtOAc in petroleum ether to 50 % EtOAc in petroleum ether. Fractions containing the product (identified by TLC and ESI-MS), were combined and concentrated under reduced pressure. The yellow residue was taken up in DCM (100 mL) and the solution was slowly (within 20 min) poured into well-stirred hexane (2.0 L) to give a precipitate that was collected by suction filtration. The obtained white powder of **18** (70.4 g, 92 % yield) was dried under reduced pressure and used in subsequent step without further purification. An aliquot of **18** (200 mg) was purified by silica gel column chromatography using

gradient elution from 25 % EtOAc in hexane to 50 % EtOAc in hexane; analytical TLC on silica gel, 1:2 EtOAc/petroleum ether, R_f =0.40; $[\alpha]_D^{23}$ +20.9 (c 0.96, MeOH).

¹**H NMR** (600 MHz, CD₃OD) δ: 8.12-8.08 (m, 2H), 8.03-7.99 (m, 2H), 7.97-7.94 (m, 2H), 7.77-7.73 (m, 1H), 7.73 – 7.70 (m, 2H), 7.65 – 7.56 (m, 4H), 7.51 – 7.43 (m, 4H), 7.42 – 7.38 (m, 4H), 7.31 – 7.27 (m, 4H), 7.18 – 7.11 (m, 2H), 7.09-7.05 (m, 1H), 6.99 – 6.90 (m, 2H), 6.19 (d, J = 4.2 Hz, 1H), 5.60 – 5.53 (m, 2H), 5.34-5.31 (m, 1H), 5.27 (dd, J = 10.6, 9.3 Hz, 1H), 5.11 (dd, J = 4.4, 1.6 Hz, 1H), 5.03 (d, J = 1.6 Hz, 1H), 5.01 (t, J= 3.1 Hz, 1H), 4.65 (dd, J = 10.6, 4.2 Hz, 1H), 4.55 (dd, J = 6.9, 4.2 Hz, 1H), 4.38 – 4.28 (m, 4H), 4.19-4.15 (m, 1H), 4.04 (dd, J = 10.3, 8.3 Hz, 1H), 3.99 – 3.93 (m, 2H), 3.91 – 3.84 (m, 2H), 3.81 (d, J = 10.3 Hz, 1H), 3.79 – 3.74 (m, 2H), 3.65-3.61 (m, 2H), 3.58 (dd, J = 11.8, 2.3 Hz, 1H), 2.12 (q, J = 12.9 Hz, 1H), 1.99 (t, J = 4.4 Hz, 1H), 1.03 (s, 9H), 0.20 (s, 3H), 0.17 (s, 3H) ppm, 5H exchangeable protons merge with H₂O.

¹³C{¹H} NMR (151 MHz, CD₃OD) δ: 168.6, 167.4, 167.3, 166.3, 165.9, 159.6 – 157.4 (5×COCF₃), 138.8, 134.9, 134.4, 134.2, 134.1, 131.4, 131.1, 130.9, 130.8, 130.7, 130.7, 130.6, 130.5, 130.4, 130.0, 129.8, 129.6, 129.5, 129.3, 129.0, 128.9, 127.5, 117.7 – 116.5 (5×CF₃), 110.6, 103.2, 98.4, 97.7, 86.7, 83.7, 80.7, 77.8, 77.3, 77.1, 77.0, 76.0, 74.4, 73.8, 72.1, 71.8, 69.4, 67.7, 66.0, 64.6, 62.0, 61.5, 53.3, 50.5, 49.7, 49.5, 41.5, 32.2, 27.0, 19.7, -4.9, -5.3 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+Na]⁺ Calcd for C₈₁H₇₈N₅O₂₄F₁₅NaSi 1840.4464; Found 1840.4443.



6,3',2'',3''',4'''-Penta-O-benzoyl-4',6'-O-benzylidene-1,3,2',2''',6'''-penta-N-trifluoroacetyl paromomycin (19). To a stirred solution of penta-*O*-benzoyl paromomycin derivative **18** (70.4 g, 38.7 mmol, 1.0 equiv) in anhydrous THF (350 mL) was dropwise added TBAF (1.0 M solution in THF; 50.3 mL, 50.3 mmol,

1.3 equiv) over 20 min period. Upon completion of the addition, color of the solution changed from light yellow to light brownish. The solution was stirred at ambient temperature for 20 h, and the reaction progress was monitored by UPLC-MS assay. The resulting brown reaction mixture was quenched with saturated aqueous NH₄Cl solution (150 mL), whereupon color changed to orange. The obtained orange suspension was evaporated to dryness under reduced pressure. Water (300 mL) was added to the residue and the solution was back-extracted with EtOAc (3×400 mL). Combined organic layers were washed with brine (2×400 mL), dried over anhydrous Na₂SO₄, filtered and concentrated to dryness under reduced pressure. The yellow residue was taken up in EtOAc (120 mL) and the solution was slowly (over 20 min)

poured into well-stirred hexane (2.0 L) to give a precipitate that was collected by suction filtration. The obtained white powder of **19** (66.0 g, 100 % yield) was dried under reduced pressure and used in subsequent step without further purification. An aliquot of **7** (200 mg) was purified by reversed-phase preparative HPLC (XBridge[®] Prep C18 OBDTM column) using gradient elution from 5 % to 95 % MeCN in 0.1 % aqueous AcOH; analytical TLC on silica gel, 1:2 EtOAc/petroleum ether, R_f =0.21. [α]_D²³ +17.8 (*c* 1.07, MeOH).

¹**H NMR** (600 MHz, CD₃OD) δ : 8.27 (dd, *J* = 8.4, 1.4 Hz, 2H), 8.11 (dd, *J* = 8.4, 1.4 Hz, 2H), 7.97 (dd, *J* = 8.4, 1.4 Hz, 2H), 7.80 – 7.76 (m, 1H), 7.74 – 7.72 (m, 2H), 7.71 – 7.67 (m, 2H), 7.65 – 7.62 (m, 1H), 7.60 – 7.56 (m, 1H), 7.49 – 7.47 (m, 2H), 7.47 – 7.43 (m, 3H), 7.42 – 7.38 (m, 4H), 7.32 – 7.28 (m, 3H), 7.24 – 7.20 (m, 2H), 7.00 – 6.94 (m, 1H), 6.91 (dd, *J* = 8.4, 6.8 Hz, 2H), 6.21 (d, *J* = 4.1 Hz, 1H), 5.62 – 5.56 (m, 2H), 5.36 (s, 1H), 5.33 – 5.29 (m, 1H), 5.29 – 5.26 (m, 2H), 5.24 – 5.22 (m, 1H), 5.08 (d, *J* = 4.4 Hz, 1H), 4.73 (dd, *J* = 10.6, 4.1 Hz, 1H), 4.51 (dd, *J* = 8.4, 4.4 Hz, 1H), 4.45 (ddd, *J* = 8.9, 4.4, 2.0 Hz, 1H), 4.40 – 4.29 (m, 4H), 4.08 (dd, *J* = 10.3, 8.4 Hz, 1H), 4.01 – 3.93 (m, 3H), 3.91 – 3.84 (m, 2H), 3.83 – 3.79 (m, 1H), 3.75 – 3.67 (m, 2H), 3.54 (dd, *J* = 14.0, 4.4 Hz, 1H), 2.17 (q, *J* = 12.9 Hz, 1H), 2.03 (dt, *J* = 12.9, 4.4 Hz, 1H) ppm, 6H exchangeable protons merge with H₂O.

¹³C{¹H} NMR (151 MHz, CD₃OD) δ: 167.3, 167.2, 166.5, 166.1, 165.7, 160.0 – 158.3 (5×COCF₃), 138.9, 135.0, 135.0, 134.4, 134.4, 134.3, 131.3, 131.0, 130.9, 130.8, 130.7, 130.6, 130.5, 130.0, 129.9, 129.9, 129.7, 129.5, 129.3, 129.2, 129.1, 129.0, 127.5, 118.5 – 115.9 (5×CF₃), 110.3, 103.1, 98.4, 97.8, 86.8, 82.4, 80.5, 77.4, 77.2, 76.6, 75.7, 74.5, 71.7, 69.8, 69.4, 67.9, 64.7, 61.7, 53.3, 50.5, 50.1, 49.6, 49.5, 40.8, 32.1 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+Na]⁺ Calcd for C₇₅H₆₄N₅O₂₄F₁₅Na 1726.3599; Found 1726.3567.



5"-Deoxy-5"-azido-6,3',2",3",4""-penta-O-benzoyl-4',6'-Obenzylidene-1,3,2',2"",6""-penta-N-trifluoroacetyl paromomycin (20). A stirred solution of 5"-hydroxy paromomycin derivative 19 (40.0 g, 23.5 mmol, 1.0 equiv) in anhydrous DCM (500 mL) was cooled to -10 °C (crushed ice/NaCl bath) and treated with 2,6-

lutidine (13.6 mL, 117 mmol, 5.0 equiv). Trifluoromethanesulfonic anhydride (11.8 mL, 70.4 mmol, 3.0 equiv) was then added dropwise at -10 °C over a period of 20 min. Upon completion of the addition the yellow solution turned brown and was stirred at -10 °C for 1 h. The resulting mixture was washed with

water (300 mL) and organic layer was back-extracted with DCM (2×300 mL). Combined organic layers were dried over anhydrous Na₂SO₄, filtered and concentrated to dryness under reduced pressure. The residual brownish oil was dissolved in MeCN (200 mL) and sodium azide (7.79 g, 120 mmol, 5.0 equiv) was added, followed by 18-crown-6 (3.17 g, 12.0 mmol, 0.5 equiv). The resulting suspension was stirred at ambient temperature for 3 h, and the reaction progress was monitored by UPLC-MS assay. The obtained orange suspension was concentrated under the reduced pressure, and the residue was partitioned between EtOAc (300 mL) and water (400 mL). The aqueous layer was back-extracted with EtOAc (2×300 mL) and combined organic extracts were washed with brine (2×200 mL), dried over anhydrous Na₂SO₄, filtered and concentrated to dryness under reduced pressure. The crude product was purified by silica gel column chromatography using gradient elution from 20 % EtOAc in petroleum ether to 100 % EtOAc. Fractions containing the product (detected by TLC and ESI-MS) were combined and concentrated under reduced pressure. The yellow oily residue was taken up in DCM (30 mL) and the solution was slowly (over 20 min) poured into well-stirred hexane (1.0 L) to give a precipitate that was collected by suction filtration. The obtained pale yellow powder of 20 (31.7 g, 77 % yield) was dried under reduced pressure and used in subsequent step. An aliquot of **20** (200 mg) was purified by reversed-phase preparative HPLC (XBridge[®] Prep C18 OBD[™] column) using gradient elution from 5 % to 95 % MeCN in 0.1 % aqueous AcOH solution; analytical TLC on silica gel, 1:2 EtOAc/petroleum ether, $R_f=0.30$. $[\alpha]_D^{23}$ +41.9 (c 1.17, MeOH).

¹**H NMR** (600 MHz, CD₃OD) δ : 8.29 (dd, *J* = 8.4, 1.4 Hz, 2H), 8.11 (dd, *J* = 8.4, 1.4 Hz, 2H), 7.98 (dd, *J* = 8.4, 1.4 Hz, 2H), 7.81 – 7.78 (m, 1H), 7.74 – 7.68 (m, 4H), 7.66 – 7.62 (m, 1H), 7.60 – 7.57 (m, 1H), 7.50 – 7.47 (m, 2H), 7.47 – 7.42 (m, 3H), 7.41 – 7.37 (m, 4H), 7.31 – 7.27 (m, 3H), 7.24 – 7.19 (m, 2H), 6.97 – 6.92 (m, 1H), 6.91 – 6.86 (m, 2H), 6.30 (d, *J* = 4.1 Hz, 1H), 5.63 – 5.55 (m, 2H), 5.36 (s, 1H), 5.32 – 5.30 (m, 1H), 5.29 – 5.26 (m, 2H), 5.25 – 5.21 (m, 1H), 5.06 (d, *J* = 4.4 Hz, 1H), 4.69 (dd, *J* = 10.6, 4.1 Hz, 1H), 4.53 (dd, *J* = 8.6, 4.4 Hz, 1H), 4.49 (ddd, *J* = 9.3, 4.4, 1.9 Hz, 1H), 4.42 – 4.27 (m, 4H), 4.10 (dd, *J* = 10.3, 8.4 Hz, 1H), 3.98 (td, *J* = 9.7, 4.7 Hz, 1H), 3.95 (d, *J* = 9.3 Hz, 1H), 3.93 – 3.88 (m, 2H), 3.88 – 3.82 (m, 2H), 3.80 (dt, *J* = 8.6, 2.6 Hz, 1H), 3.67 (dd, *J* = 13.5, 2.6 Hz, 1H), 3.43 (dd, *J* = 14.1, 4.4 Hz, 1H), 2.18 (q, *J* = 12.9 Hz, 1H), 2.04 (dt, *J* = 12.9, 4.4 Hz, 1H) ppm, 5H exchangeable protons merge with H₂O.

¹³C{¹H} NMR (151 MHz, CD₃OD) δ: 167.3, 167.2, 166.5, 166.0, 165.7, 159.8 – 158.4 (5×COCF₃), 138.8, 135.0, 135.0, 134.4, 134.4, 134.3, 131.3, 131.0, 130.9, 130.8, 130.7, 130.6, 130.5, 130.0, 129.9, 129.9, 129.7, 129.5, 129.4, 129.2, 129.1, 129.0, 127.5, 118.7 – 115.7 (5×CF₃), 110.6, 103.1, 98.3, 98.0, 87.1, 80.8, 79.5, 77.3, 77.1, 76.4, 75.1, 74.3, 71.6, 69.8, 69.5, 67.8, 64.8, 53.3, 50.6, 50.4, 50.1, 49.6, 49.5, 40.7, 32.1 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+Na]⁺ Calcd for C₇₅H₆₃N₈O₂₃F₁₅Na 1751.3664; Found 1751.3624.



5"-Deoxy-5"-amino-6,3',2",3",4"'-penta-O-benzoyl-4',6'-Obenzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (21). To a stirred solution of 5"-deoxy-5"-azido paromomycin derivative 20 (31.7 g, 18.3 mmol, 1.0 equiv) in a mixture of degassed THF (300 mL, bubbling a vigorously stirred THF with

argon for 15 min) and water (206 mL) was dropwise added PMe₃ (1.0 M solution in THF; 18.3 mL, 18.3 mmol, 1.0 equiv) over 20 min at 0 °C (crushed ice bath). After stirring at ambient temperature for 10 h (the reaction progress was monitored by UPLC-MS assay), the yellowish mixture was concentrated to dryness under reduced pressure. The crude product was purified by silica gel column chromatography using gradient elution from 25 % EtOAc in hexane to 100 % EtOAc. Fractions containing the product (detected by TLC and ESI-MS) were combined and concentrated under reduced pressure to give product **21** (12.6 g, 40 % yield) as a white powder. An aliquot of **21** (100 mg) was purified by reversed-phase preparative HPLC (XBridge[®] Prep C18 OBDTM column) using gradient elution from 10 % to 90 % MeCN in 0.1 % aqueous AcOH solution; analytical TLC on silica gel, 1:1 EtOAc/petroleum ether, R_f =0.22; $[\alpha]_D^{23}$ +18.0 (*c* 1.00, MeOH).

¹**H NMR** (600 MHz, CD₃OD) δ: 8.35 – 8.30 (m, 2H), 8.14 (dd, *J* = 8.4, 1.4 Hz, 2H), 7.97 (dd, *J* = 8.4, 1.4 Hz, 2H), 7.83 – 7.78 (m, 1H), 7.75 – 7.72 (m, 2H), 7.72 – 7.69 (m, 2H), 7.67 – 7.65 (m, 1H), 7.62 – 7.58 (m, 1H), 7.52 – 7.48 (m, 2H), 7.48 – 7.42 (m, 4H), 7.42 – 7.38 (m, 3H), 7.32 – 7.28 (m, 3H), 7.23 (dd, *J* = 8.4, 7.4 Hz, 2H), 6.97 – 6.92 (m, 1H), 6.89 (dd, *J* = 8.4, 6.7 Hz, 2H), 5.99 (d, *J* = 4.2 Hz, 1H), 5.64 – 5.58 (m, 2H), 5.40 (s, 1H), 5.35 – 5.32 (m, 1H), 5.30 (t, *J* = 2.9 Hz, 1H), 5.28 – 5.26 (m, 1H), 5.13 (d, *J* = 4.2 Hz, 1H), 4.74 (dd, *J* = 10.6, 4.2 Hz, 1H), 4.46 – 4.42 (m, 1H), 4.41 (d, *J* = 8.8 Hz, 1H), 4.37 (d, *J* = 4.4 Hz, 1H), 4.36 – 4.34 (m, 1H), 4.34 – 4.28 (m, 2H), 4.28 – 4.25 (m, 1H), 4.25 – 4.22 (m, 1H), 4.15 – 4.11 (m, 1H), 4.01 – 3.96 (m, 3H), 3.93 – 3.89 (m, 1H), 3.86 (t, *J* = 9.9 Hz, 1H), 3.34 (dd, *J* = 14.1, 3.4 Hz, 1H), 3.27 (dd, *J* = 13.7, 2.8 Hz, 1H), 2.92 (dd, *J* = 13.7, 8.4 Hz, 1H), 2.25 (q, *J* = 12.9 Hz, 1H), 2.05 (dt, *J* = 12.9, 4.4 Hz, 1H) ppm, 7H exchangeable protons merge with H₂O.

¹³C{¹H} NMR (151 MHz, CD₃OD) δ: 167.3, 167.2, 166.4, 165.8, 165.6, 160.2 – 158.4 (5×COCF₃), 138.7, 135.0, 135.0, 134.5, 134.5, 134.5, 131.3, 131.0, 131.0, 130.7, 130.6, 130.6, 130.5, 130.0, 130.0, 129.9,

129.8, 129.7, 129.6, 129.2, 129.1, 129.0, 127.5, 117.8 − 116.5 (5×CF₃), 110.5, 103.1, 98.0, 97.6, 87.3, 80.5, 77.5, 77.2, 77.1, 75.8, 75.6, 71.2, 69.9, 69.4, 67.7, 64.8, 53.6, 50.5, 49.9, 49.6, 49.4, 44.4, 40.7, 31.8 ppm. **HRMS** (ESI/Q-TOF): *m/z* [M+H]⁺ Calcd for C₇₅H₆₆N₆O₂₃F₁₅ 1703.3940; Found 1703.3945.



5"-Deoxy-5"-(O-benzylcarbamoyl)-6,3',2",3"',4"'-penta-O-benzoyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (22). A stirred solution of 5"-deoxy-5"-amino paromomycin derivative 21 (3.59 g, 2.11 mmol, 1.0 equiv) in anhydrous DCM (30 mL) was cooled to 0 °C (crushed

ice bath) and Cbz-Cl (1.2 mL, 8.43 mmol, 4.0 equiv) was added dropwise over 5 min. The resulting yellowish solution was stirred at 0 °C for 1 h followed by dropwise addition (over 5 min) of pyridine (0.34 mL, 4.21 mmol, 2.0 equiv). The resulting yellow solution was stirred at 0 °C for 1 h, then warmed up to room temperature and stirring was continued for additional 2 h. The reaction progress was monitored by UPLC-MS assay. The yellowish solution was diluted with water (50 mL) and back-extracted with DCM (2×30 mL). Combined organic extracts were washed with brine (30 mL), dried over anhydrous Na₂SO₄ and evaporated under reduced pressure. The crude yellow oil was purified by silica gel column chromatography using gradient elution from 25 % EtOAc in hexane to 50 % EtOAc in hexane. Fractions containing the product (detected with TLC and ESI-MS) were combined and concentrated under reduced pressure. The residue was dissolved in DCM (10 mL) and hexane (100 mL) was added. The formed white precipitate was collected by suction filtration and dried under reduced pressure to give product **22** (3.45 g, 89 % yield) as a white powder. An aliquot of **22** (100 mg) was purified by reversed-phase preparative HPLC (XBridge® Prep C18 OBDTM column) using gradient elution from 10 % to 90 % MeCN in 0.1 % aqueous AcOH solution; analytical TLC on silica gel, 1:2 EtOAc/petroleum ether, R_r =0.24. [α]²³_D +6.5 (c 1.74, MeOH).

¹**H NMR** (600 MHz, CDCl₃) δ: 8.66 (t, J = 5.9 Hz, 1H), 8.12 – 8.02 (m, 2H), 7.97 – 7.93 (m, 4H), 7.93 – 7.87 (m, 1H), 7.87 – 7.83 (m, 2H), 7.81 – 7.74 (m, 2H), 7.69 – 7.65 (m, 2H), 7.63 – 7.60 (m, 1H), 7.59 – 7.57 (m, 1H), 7.53 – 7.46 (m, 5H), 7.43 – 7.40 (m, 2H), 7.38 – 7.36 (m, 2H), 7.33 – 7.28 (m, 11H), 7.21 – 7.15 (m, 2H), 7.09 – 7.03 (m, 2H), 7.01 – 6.89 (m, 1H), 6.08 – 5.95 (m, 1H), 5.70 – 5.61 (m, 1H), 5.59 – 5.53 (m, 1H), 5.46 – 5.40 (m, 2H), 5.34 – 5.24 (m, 3H), 5.14 (d, J = 5.1 Hz, 1H), 5.11 (t, J = 4.5 Hz, 1H), 5.08 – 5.02 (m, 2H), 4.95 – 4.82 (m, 1H), 4.57 – 4.50 (m, 1H), 4.33 – 4.27 (m, 3H), 4.25 – 4.18 (m, 2H), 4.15 – 4.07 (m, 2H), 4.00

- 3.97 (m, 2H), 3.90 - 3.81 (m, 2H), 3.71 - 3.63 (m, 2H), 3.45 - 3.28 (m, 1H), 3.27 - 3.13 (m, 1H), 2.39 - 2.29 (m, 1H) ppm.

¹³C{¹H} NMR (151 MHz, CDCl₃) δ: 167.3, 167.2, 165.8, 165.1, 164.2, 157.7, 158.4 – 157.0 (5×COCF₃), 136.8, 136.3, 134.4, 134.3, 134.2, 133.8, 133.7, 130.3, 130.0, 129.9, 129.8, 129.8, 129.4, 128.9, 128.9, 128.8, 128.7, 128.7, 128.6, 128.4, 128.3, 128.2, 128.1, 127.9, 127.8, 127.4, 126.4, 126.3, 117.1 – 114.3 (5×CF₃), 107.0, 102.0, 98.8, 97.6, 81.1, 78.7, 77.9, 75.5, 74.2, 72.7, 69.9, 68.9, 68.4, 67.1, 64.1, 53.9, 49.9, 49.4, 49.2, 48.7, 42.3, 40.9, 31.7 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+Na]⁺ Calcd for C₈₃H₇₁N₆O₂₅F₁₅Na 1859.4127; Found 1859.4125.



5"-Deoxy-5"-(*O*-benzylcarbamoyl)-6,3',2",3"",4""-penta-*O*benzoyl-1,3,2',2"",6""-penta-*N*-trifluoroacetyl paromomycin (23). A solution of 5"-deoxy-5"-carboxybenzylamino paromomycin derivative 22 (2.64 g, 1.43 mmol, 1.0 equiv) in 80 % aqueous acetic acid (12.4 mL, prepared by mixing 10.1 mL of

glacial acetic acid and 2.3 mL of water) was heated to 65 °C for 18 h, and the reaction progress was monitored by UPLC-MS assay. After cooling to room temperature, the colorless solution was concentrated under reduced pressure and the residue was co-evaporated with toluene (50 mL). The crude colorless oil was purified by silica gel column chromatography using gradient elution from 25 % EtOAc in hexane to 100 % EtOAc. Fractions containing the product (detected with TLC and ESI-MS) were combined and concentrated under reduced pressure. The residue was dissolved in DCM (10 mL) and hexane (100 mL) was added. The formed white precipitate was collected by suction filtration and dried under reduced pressure to give product **23** (2.16 g, 86 % yield) as a white powder. An aliquot of **23** (100 mg) was purified by reversed-phase preparative HPLC (XBridge® Prep C18 OBDTM column) using gradient elution from 5 % to 95 % MeCN in 0.1 % aqueous AcOH solution; analytical TLC on silica gel, 1:1 EtOAc/petroleum ether, R_f =0.32. [α] $_D^{23}$ +27.4 (c 1.02, MeOH).

¹**H NMR** (600 MHz, CD₃OD) δ : 8.26 – 8.23 (m, 2H), 8.10 – 8.06 (m, 2H), 8.00 – 7.97 (m, 2H), 7.85 – 7.80 (m, 2H), 7.76 – 7.72 (m, 1H), 7.64 – 7.58 (m, 6H), 7.48 – 7.42 (m, 7H), 7.37 – 7.33 (m, 2H), 7.32 – 7.29 (m, 1H), 7.29 – 7.24 (m, 2H), 7.15 (dd, J = 7.7, 7.5 Hz, 1H), 7.01 – 6.97 (m, 2H), 5.88 (d, J = 4.0 Hz, 1H), 5.38 (dd, J = 11.0, 8.8 Hz, 1H), 5.34 (s, 1H), 5.33 – 5.30 (m, 1H), 5.29 (t, J = 3.0 Hz, 1H), 5.25 (d, J = 12.6 Hz, 1H), 5.22 (d, J = 4.6 Hz, 1H), 5.20 – 5.17 (m, 2H), 5.17 – 5.13 (m, 1H), 4.54 (dd, J = 11.0, 4.0 Hz, 1H), 4.51 – 4.47 (m, 1H),

4.36 – 4.31 (m, 2H), 4.25 (dd, J = 8.1, 3.6 Hz, 1H), 4.12 (dd, J = 10.4, 8.6 Hz, 1H), 3.95 (ddd, J = 8.4, 6.1, 3.0 Hz, 1H), 3.90 (d, J = 10.5 Hz, 1H), 3.81 – 3.79 (m, 2H), 3.78 – 3.75 (m, 1H), 3.62 (dd, J = 14.2, 3.2 Hz, 1H), 3.57 (d, J = 6.5 Hz, 2H), 3.40 (dd, J = 14.2, 6.2 Hz, 1H), 2.11 (q, J = 12.6 Hz, 1H), 2.03 (dt, J = 13.1, 4.6 Hz, 1H) ppm, 8H exchangeable protons merge with H₂O.

¹³C{¹H} NMR (151 MHz, CD₃OD) δ: 167.8, 167.2, 166.5, 166.4, 165.5, 159.2, 159.9 – 158.2 (5×COCF₃), 138.3, 135.1, 134.9, 134.7, 134.4, 134.3, 134.2, 131.3, 131.1, 131.0, 130.9, 130.8, 130.7, 130.5, 129.9, 129.8, 129.6, 129.5, 129.5, 129.4, 129.4, 129.2, 129.0, 128.8, 118.8 – 115.5 (5×CF₃), 109.7, 98.9, 97.5, 85.1, 81.9, 78.5, 77.7, 76.8, 76.5, 74.9, 74.2, 73.8, 69.6, 69.4, 67.8, 61.9, 53.4, 50.4, 50.3, 49.7, 49.6, 43.8, 41.0, 32.3 ppm.

HRMS (ESI/Q-TOF): *m/z* [M+Na]⁺ Calcd for C₇₆H₆₅N₆O₂₅F₁₅Na 1771.3814; Found 1771.3860



5"-Deoxy-5"-(*O*-benzylcarbamoyl)-6,3',6',2",3"",4""-hexa-*O*benzoyl-1,3,2',2"",6""-penta-*N*-trifluoroacetyl paromomycin (24). A stirred solution of paromomycin derivative 23 (2.06 g, 1.18 mmol, 1.0 equiv) in anhydrous MeCN (25 mL) was cooled to 0 °C (crushed ice bath). NEt₃ (0.25 mL, 1.76 mmol, 1.5 equiv)

was added, followed by dropwise addition of a benzoyl cyanide (170 mg, 1.29 mmol, 1.1 equiv) solution in anhydrous MeCN (5 mL) over 30 min period. After stirring for additional 2 h at 0 °C, the clear colorless solution was quenched by addition of MeOH (0.5 mL) and concentrated under reduced pressure. The crude product was purified by silica gel column chromatography using gradient elution from 25 % EtOAc in hexane to 50 % EtOAc in hexane. Fractions containing the product (detected with TLC and ESI-MS), were combined and concentrated under reduced pressure. The residue was dissolved in DCM (10 mL) and hexane (100 mL) was added to the well-stirred solution. The formed white precipitate was collected by suction filtration and dried under reduced pressure to give product **24** (1.98 g, 91 % yield) as a white powder. An aliquot of **24** (100 mg) was purified by reversed-phase preparative HPLC (XBridge[®] Prep C18 OBDTM column) using gradient elution from 5 % to 95 % MeCN in 0.1 % aqueous AcOH solution; analytical TLC on silica gel, 1:1 EtOAc/petroleum ether, R_r =0.44. [α]^{2,3}_D +39.3 (*c* 1.09, MeOH).

¹**H NMR** (600 MHz, CD₃OD) δ: 8.30 – 8.17 (m, 2H), 8.14 – 8.03 (m, 4H), 8.04 – 7.98 (m, 2H), 7.84 – 7.76 (m, 2H), 7.74 (dd, J = 7.7, 7.5 Hz, 1H), 7.68 – 7.56 (m, 5H), 7.56 – 7.48 (m, 2H), 7.48 – 7.40 (m, 7H), 7.36 – 7.27 (m, 4H), 7.27 – 7.18 (m, 3H), 7.11 (dd, J = 7.7, 7.5 Hz, 1H), 7.01 – 6.93 (m, 2H), 5.93 (d, J = 4.0 Hz, 1H), 5.49

-5.44 (m, 1H), 5.37 (s, 1H), 5.33 (t, J = 9.8 Hz, 1H), 5.27 (t, J = 3.0 Hz, 1H), 5.23-5.12 (m, 3H), 5.11 (d, J = 12.6 Hz, 1H), 5.06 (d, J = 12.6 Hz, 1H), 4.67 (d, J = 12.6 Hz, 1H), 4.64 - 4.55 (m, 2H), 4.47 - 4.42 (m, 1H), 4.39 (t, J = 9.0 Hz, 1H), 4.36 - 4.28 (m, 2H), 4.26 (dd, J = 8.4, 4.7 Hz, 1H), 4.12 (dd, J = 10.3, 8.4 Hz, 1H), 4.09 - 4.02 (m, 2H), 3.96 - 3.88 (m, 1H), 3.85 - 3.80 (m, 1H), 3.62 (dd, J = 14.6, 3.2 Hz, 1H), 3.55 - 3.44 (m, 2H), 3.37 (dd, J = 14.6, 5.7 Hz, 1H), 2.15 (q, J = 12.9 Hz, 1H), 2.05 (dt, J = 12.9, 4.4 Hz, 1H) ppm, 7H exchangeable protons merge with H₂O.

¹³C{¹H} NMR (151 MHz, CD₃OD) δ: 168.1, 167.8, 167.2, 166.4, 166.3, 165.5, 159.0, 160.1 – 157.7 (5×COCF₃), 138.3, 135.0, 134.9, 134.7, 134.4, 134.3, 131.3, 131.2, 131.0, 130.9, 130.8, 130.7, 130.6, 130.5, 129.9, 129.7, 129.6, 129.6, 129.5, 129.4, 129.4, 129.2, 128.9, 128.5, 118.5 – 116.0 (5×CF₃), 109.6, 98.8, 97.6, 85.3, 81.5, 78.1, 78.1, 76.7, 76.6, 74.9, 73.7, 72.0, 69.6, 69.3, 67.7, 67.7, 63.7, 53.5, 50.6, 50.3, 49.8, 43.4, 40.8, 32.2 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+Na]⁺ Calcd for C₈₃H₇₁N₆O₂₆F₁₅Na 1875.4076; Found 1875.4088.



4',5"-Dideoxy-5"-(*O*-benzylcarbamoyl)-6,3',6',2",3"',4" -hexa-*O*-benzoyl-4'-iodo-1,3,2',2"',6"'-penta-*N*-trifluoroacetyl 4'-*epi*paromomycin (25). A stirred solution of hexa-*O*-benzoyl paromomycin derivative 24 (6.39 g, 3.45 mmol, 1.0 equiv) in anhydrous DCM (70 mL) was cooled to 0 °C (crushed ice bath) and treated with anhydrous pyridine (2.1 mL, 25.9 mmol, 7.5 equiv).

Trifluoromethanesulfonic anhydride (1.7 mL, 10.3 mmol, 3.0 equiv) was then added dropwise over a period of 15 min. Upon completion of the addition, the yellow solution turned light brown. After stirring at 0 °C for 1 h, the solution was washed with water (120 mL) and organic layer was back-extracted with DCM (2×60 mL). Combined organic layers were dried over anhydrous Na₂SO₄, filtered and concentrated to dryness under reduced pressure. The obtained yellow oil was dissolved in anhydrous acetone (70 mL) and potassium iodide (5.73 g, 34.5 mmol, 10.0 equiv) was added. The resulting well-stirred suspension was heated at 55 °C for 6 h, and the reaction progress was monitored by UPLC-MS assay. After cooling to room temperature, the orange suspension was filtered through the pad of *Celite*[®]. The filter plug was washed with DCM, the combined filtrates were dried over anhydrous Na₂SO₄, filtered and evaporated under reduced pressure. The crude product was purified by silica gel column chromatography using gradient elution from 25 % EtOAc in hexane to 50 % EtOAc in hexane. Fractions containing the product (detected with TLC and ESI-MS) were combined and concentrated under reduced pressure. The residue was dissolved in DCM (20 mL) and hexane (200 mL) was added to the well-stirred solution. The formed

white precipitate was collected by suction filtration and dried under reduced pressure to give product **25** (5.64 g, 83 % yield) as a white powder. An aliquot of **25** (100 mg) was purified by reversed-phase preparative HPLC (XBridge[®] Prep C18 OBDTM column) using gradient elution from 5 % to 95 % MeCN in 0.1 % aqueous AcOH solution; analytical TLC on silica gel, 1:1 EtOAc/petroleum ether, R_{f} =0.56; $[\alpha]_{D}^{23}$ +28.8 (*c* 1.04, MeOH).

¹**H NMR** (600 MHz, CD₃OD) δ : 8.31-8.23 (m, 2H), 8.13-8.06 (m, 2H), 8.06 – 7.98 (m, 4H), 7.82-7.73 (m, 3H), 7.70 – 7.57 (m, 5H), 7.56 – 7.37 (m, 11H), 7.35 – 7.30 (m, 2H), 7.29 – 7.20 (m, 3H), 7.05 (dd, *J* = 7.6, 7.4 Hz, 1H), 7.00-6.90 (m, 2H), 6.11 (d, *J* = 3.8 Hz, 1H), 5.40 (s, 1H), 5.35 – 5.29 (m, 2H), 5.27 – 5.25 (m, 1H), 5.24 – 5.11 (m, 5H), 5.03 (dd, *J* = 11.0, 3.8 Hz, 1H), 4.66 (dd, *J* = 11.0, 3.8 Hz, 1H), 4.61 (dd, *J* = 10.7, 5.4 Hz, 1H), 4.52 – 4.47 (m, 1H), 4.41 – 4.28 (m, 5H), 4.07 (dd, *J* = 10.3, 8.5 Hz, 1H), 3.94 (ddd, *J* = 8.9, 5.8, 3.5 Hz, 1H), 3.90 – 3.86 (m, 1H), 3.75 – 3.64 (m, 2H), 3.61 – 3.49 (m, 2H), 3.42 (dd, *J* = 14.7, 5.8 Hz, 1H), 2.14 (q, *J* = 12.8 Hz, 1H), 2.04 (dt, *J* = 12.8, 4.4 Hz, 1H) ppm, 6H exchangeable protons merge with H₂O.

¹³C{¹H} NMR (151 MHz, CD₃OD) δ: 167.1, 167.0, 166.6, 166.5, 166.2, 165.6, 159.2, 160.2 – 158.1 (5×COCF₃), 138.4, 135.0, 135.0, 134.9, 134.7, 134.6, 134.3, 131.3, 131.1, 130.9, 130.9, 130.7, 130.6, 130.5, 130.2, 129.9, 129.8, 129.6, 129.5, 129.4, 129.3, 129.1, 128.9, 128.8, 118.5 – 115.7 (5×CF₃), 109.8, 98.8, 97.3, 85.4, 81.7, 77.9, 77.6, 76.7, 76.7, 73.7, 70.2, 69.7, 67.8, 67.7, 67.6, 67.5, 52.0, 50.3, 50.3, 49.7, 43.4, 40.9, 37.8, 32.0 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+Na]⁺ Calcd for C₈₃H₇₀N₆O₂₅F₁₅NaI 1985.3093; Found 1985.3112.



4'-Allyl-4',5"-dideoxy-5"-(O-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-O-benzoyl-1,3,2',2"',6"'-penta-Ntrifluoroacetyl paromomycin (26). To a stirred solution of 4'deoxy-4'-iodo-4'-*epi*-paromomycin 25 (1.01 g, 0.51 mmol, 1.0 equiv) in anhydrous 1,2-dichlorobenzene (35 mL) under argon atmosphere was added allyl phenyl sulfone (0.79 mL,

5.13 mmol, 10.0 equiv). After stirring at ambient temperature for 5 min, the resulting colorless clear solution was cooled to 0 °C (crushed ice bath) and BEt₃ (1.0 M solution in hexanes; 1.54 mL, 1.54 mmol, 3.0 equiv) was then added over a period of 1 min with gentle stirring keeping the needle under the surface of the solution. The septa was removed and the colorless reaction solution was stirred under air at 0 °C for 1 h, whereupon another portion of BEt₃ (1.0 M solution in hexanes; 1.54 mL, 1.54 mmol, 3.0 equiv)

was added by the same technique and stirring continued for another 1 hour. Altogether, five portions of BEt₃ (5×1.54 mL) were added after every 1-2 h to achieve full conversion (the reaction progress was monitored by UPLC-MS assay). Them, the reaction mixture was concentrated under reduced pressure, the residue was dissolved in DCM (20 mL) and filtered through a silica gel pad (45×45 mm). The filter plug was first washed with DCM (300 mL) to separate the excess of allyl phenyl sulfone, and then washed with EtOAc (250 mL) to collect the product). The product-containing filtrate was evaporated under reduced pressure and the residue was purified by reversed-phase preparative HPLC (XBridge[®] Prep C18 OBDTM column) using gradient elution from 70 % to 95 % MeCN in 0.1 % aqueous AcOH solution. Fractions containing the product (detected by TLC and ESI-MS), were combined and concentrated under reduced pressure. The residue was dissolved in DCM (5 mL) and hexane (50 mL) was added to the solution. The formed white precipitate was dried under reduced pressure to give product **26** (498 mg, 40 % yield) as a white powder; analytical TLC on silica gel, 1:1 EtOAc/petroleum ether, R_f =0.60; [α]^{2.3} +28.7 (*c* 1.21, MeOH).

¹**H NMR** (400 MHz, CD₃OD) δ: 8.29 – 8.21 (m, 2H), 8.10-8.02 (m, 4H), 8.01-7.95 (m, 2H), 7.83 – 7.74 (m, 3H), 7.70 – 7.58 (m, 5H), 7.55 – 7.42 (m, 9H), 7.36 – 7.22 (m, 7H), 7.08 (dd, J = 7.7, 7.5 Hz, 1H), 7.00-6.93 (m, 2H), 6.02 (d, J = 4.0 Hz, 1H), 5.88 – 5.72 (m, 1H), 5.50 (t, J = 10.7 Hz, 1H), 5.38 (s, 1H), 5.33 (dd, J = 10.5, 9.3 Hz, 1H), 5.28 (t, J = 2.9 Hz, 1H), 5.24 – 5.17 (m, 3H), 5.16 – 5.07 (m, 2H), 5.06 – 4.97 (m, 1H), 4.91 – 4.88 (m, 1H), 4.67 – 4.58 (m, 3H), 4.49 – 4.45 (m, 1H), 4.41 (t, J = 8.8 Hz, 1H), 4.38 – 4.26 (m, 3H), 4.15 – 4.07 (m, 2H), 3.98 – 3.89 (m, 1H), 3.88 – 3.85 (m, 1H), 3.72 – 3.63 (m, 1H), 3.58 – 3.48 (m, 2H), 3.42 – 3.34 (m, 1H), 2.50 – 2.40 (m, 1H), 2.31 – 2.24 (m, 2H), 2.14 (q, J = 12.7 Hz, 1H), 2.07-2.00 (m, 1H) ppm, 6H exchangeable protons merge with H₂O.

¹³C{¹H} NMR (101 MHz, CD₃OD) δ: 67.8, 167.6, 167.2, 166.4, 166.2, 165.6, 160.2 – 158.3 (5×COCF₃), 159.0, 138.3, 135.4, 135.0, 134.9, 134.6, 134.5, 134.5, 134.3, 131.3, 131.1, 131.0, 130.9, 130.8, 130.6, 130.5, 129.9, 129.7, 129.6, 129.5, 129.4, 129.4, 129.1, 128.9, 128.5, 117.9, 119.2 – 115.5 (5×CF₃), 109.8, 98.8, 97.8, 85.7, 81.4, 78.2, 78.1, 76.8, 76.7, 73.7, 72.4, 71.3, 69.7, 67.7, 67.7, 64.7, 54.4, 50.7, 50.3, 49.7, 43.5, 42.7, 40.8, 32.6, 32.3 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+Na]⁺ Calcd for C₈₆H₇₅N₆O₂₅F₁₅Na 1899.4440; Found 1899.4436.

General Procedure A for the photochemically initiated radical allylation of iodo paromomycin derivatives 25 and 29

An oven-dried Biotage microwave pressure vial (10 mL) was cooled under a stream of argon, equipped with a stirring bar and charged with iodide (1.0 equiv) and 2,4,5,6-tetra(9*H*-carbazol-9-yl)isophthalonitrile (4CzIPN) (0.05 equiv). The vial was sealed and flushed with argon. Anhydrous degassed DMSO was added, followed by NEt₃ (3.0 equiv) and allyl phenyl sulfone (2.0-5.0 equiv). The well-stirred yellow reaction mixture was irradiated at ambient temperature with blue LED light for 1-18 h. After dilution with water (15 mL) and addition of EtOAc (15 mL), layers were separated, and the aqueous layer was back-extracted with EtOAc (3×15 mL). The combined organic layers were washed with brine (10 mL), dried over anhydrous Na₂SO₄ and evaporated under reduced pressure.



4',5"-Dideoxy-5"-(*O*-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-*O*benzoyl-1,3,2',2"',6"'-penta-*N*-trifluoroacetyl paromomycin-4'ene (27). The title compound was obtained from 4'-*epi*-iodoparomomycin 25 (50 mg, 0.026 mmol, 1.0 equiv), 4CzIPN (1 mg, 0.0013 mmol, 0.05 equiv), allyl phenyl sulfone (8 μL, 0.051 mmol,

2.0 equiv) and NEt₃ (11 µL, 0.076 mmol, 3.0 equiv) in degassed DMSO (1 mL) by following General Procedure A. Pure product **27** was obtained by silica gel column chromatography using gradient elution from 25 % EtOAc in heptane to 35 % EtOAc in heptane; white powder (30 mg, 64 % yield), analytical TLC on silica gel, 1:2 EtOAc/petroleum ether, R_f =0.26. [α]_D²³ +55.7 (*c* 0.93, MeOH).

¹**H NMR** (400 MHz, CD₃OD) δ : 8.28 – 8.21 (m, 2H), 8.13 – 8.08 (m, 2H), 8.08 – 8.03 (m, 2H), 8.01 – 7.96 (m, 2H), 7.93 – 7.86 (m, 2H), 7.78 – 7.72 (m, 1H), 7.70 – 7.56 (m, 7H), 7.53 – 7.44 (m, 7H) 7.42 – 7.37 (m, 2H), 7.36 – 7.22 (m, 6H), 7.11 – 7.01 (m, 1H), 6.07 – 5.98 (m, 1H) 5.73 – 5.64 (m, 1H), 5.41 – 5.33 (m, 2H), 5.31 – 5.27 (m, 2H), 5.25 (dd, *J* = 9.0, 3.6 Hz, 2H), 5.19 – 5.15 (m, 1H), 5.14 – 5.09 (m, 1H), 5.08 – 5.04 (m, 1H), 4.97 – 4.90 (m, 1H), 4.83 – 4.74 (m, 2H), 4.47 (t, *J* = 6.6 Hz, 1H), 4.39 – 4.24 (m, 3H), 4.17 – 4.05 (m, 2H), 4.04 – 3.96 (m, 1H), 3.72 – 3.66 (m, 1H), 3.60 – 3.49 (m, 4H), 2.16 (q, *J* = 12.7 Hz, 1H), 2.09 – 2.01 (m, 1H) ppm, 6H exchangeable protons merge with H₂O.

¹³C{¹H} NMR (101 MHz, CD₃OD) δ: 167.4, 167.2, 167.1, 166.6, 166.4, 165.5, 160.2 - 158.2 (5×COCF₃), 149.5, 138.3, 135.1, 134.9, 134.9, 134.6, 134.4, 131.3, 131.1, 131.0, 130.9, 130.8, 130.7, 130.7, 130.6, 130.5, 129.9, 129.9, 129.7, 129.6, 129.5, 129.5, 129.3, 128.9, 128.7, 119.0 - 115.2 (5×CF₃), 109.3, 100.0,

99.0, 97.8, 83.3, 82.2, 78.8, 78.4, 76.7, 75.9, 73.7, 69.5, 68.0, 67.8, 67.7, 63.8, 51.3, 50.4, 49.9, 49.5, 43.1, 41.1, 31.9 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+Na]⁺ Calcd for C₈₃H₆₉N₆O₂₅F₁₅Na 1857.3971; Found 1857.4004.



5"-Deoxy-5"-(O-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-Obenzoyl-1,3,2',2"',6"'-penta-N-trifluoroacetyl 4'-epiparomomycin (28). A stirred solution of 4'-hydroxy paromomycin derivative 24 (600 mg, 0.32 mmol, 1.0 equiv) in anhydrous DCM (12 mL) was cooled to 0 °C (crushed ice bath)

and treated with anhydrous pyridine (0.2 mL, 2.4 mmol, 7.5 equiv). Trifluoromethanesulfonic anhydride (0.16 mL, 0.97 mmol, 3.0 equiv) was then added dropwise over a period of 5 min, whereupon the light yellow solution turned brownish. After stirring at 0 °C for 1 h, the reaction mixture was washed with aqueous 5 % KHSO₄ solution (15 mL), the organic layer was washed with saturated aqueous NaHCO₃ solution (15 mL) and water (20 mL). Organic layer was dried over anhydrous Na₂SO₄, filtered and concentrated to dryness under reduced pressure. The brownish viscous oily residue was dissolved in anhydrous DMF (8 mL) and sodium nitrite (134 mg, 1.94 mmol, 6.0 equiv) was added, whereupon the color turned dark red. The reaction mixture was stirred at ambient temperature for 18 h, and the reaction color gradually changed back to the light brown. Water (20 mL) was then added to the brownish reaction mixture and the solution was back-extracted with EtOAc (3×20 mL). Combined organic layers were washed with water (20 mL) and brine (20 mL), dried over anhydrous Na₂SO₄ filtered and concentrated to dryness under reduced pressure. The crude product was purified by silica gel column chromatography using gradient elution from 25 % EtOAc in hexane to 50 % EtOAc in hexane. Fractions containing the product (detected by TLC and ESI-MS), were combined and concentrated under reduced pressure. The residue was dissolved in DCM (5 mL) and hexane (50 mL) was poured to the solution. The formed white precipitate was collected by suction filtration and dried under reduced pressure to give product 28 (427 mg, 71 % yield) as a white powder. An aliquot of 28 (60 mg) was purified by reversed-phase preparative HPLC (XBridge[®] Prep C18 OBD[™] column) using gradient elution from 50 % to 95 % MeCN in 0.1 % aqueous AcOH solution; analytical TLC on silica gel, 1:1 EtOAc/petroleum ether, R_{f} =0.56. $[\alpha]_{D}^{23}$ +22.0 (c 1.08, MeOH).

¹**H NMR** (600 MHz, CD₃OD) δ: 8.30-8.22 (m, 2H), 8.12-8.07 (m, 2H), 8.07 – 8.00 (m, 5H), 7.82 – 7.78 (m, 2H), 7.78-7.72 (m, 1H), 7.69 – 7.54 (m, 7H), 7.53 – 7.40 (m, 9H), 7.37 – 7.31 (m, 2H), 7.31 – 7.24 (m, 3H), 7.14 – 7.08 (m, 1H), 7.01 – 6.94 (m, 2H), 6.05 (d, *J* = 3.8 Hz, 1H), 5.44 – 4.98 (m, 10H), 4.57 – 4.52 (m, 1H), 4.51 – 4.46 (m, 2H), 4.39 – 4.26 (m, 5H), 4.10 – 4.01 (m, 1H), 3.95 (ddd, *J* = 8.4, 5.4, 3.4 Hz, 1H), 3.86 – 3.79 (m, 1H), 3.69 – 3.62 (m, 1H), 3.60 (dd, *J* = 14.0, 5.4 Hz, 1H), 3.53 (dd, *J* = 14.3, 7.2 Hz, 1H), 3.44 (dd, *J* = 14.3, 5.4 Hz, 1H), 2.14 (q, *J* = 12.8 Hz, 1H), 2.03 (dt, *J* = 12.8, 4.3 Hz, 1H) ppm, 7H exchangeable protons merge with H₂O.

¹³C{¹H} NMR (151 MHz, CD₃OD) δ: 167.5, 167.4, 167.2, 166.5, 166.3, 165.5, 160.0 – 158.1 (5×COCF₃),
138.4, 135.0, 134.9, 134.7, 134.5, 134.4, 134.3, 134.2, 131.3, 131.1, 131.0, 130.9, 130.9, 130.8, 130.7,
130.6, 130.5, 130.5, 129.9, 129.7, 129.6, 129.6, 129.5, 129.5, 129.4, 129.2, 128.9, 128.7, 118.7 – 115.7
(5×CF₃), 109.7, 98.9, 97.7, 85.3, 82.0, 78.1, 77.4, 76.8, 76.5, 73.7, 72.6, 69.7, 69.6, 67.8, 67.8, 66.4, 62.7,
50.4, 50.3, 49.8, 49.6, 43.3, 40.9, 32.1 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+Na]⁺ Calcd for C₈₃H₇₁N₆O₂₆F₁₅Na 1875.4076; Found 1875.4097.



4',5"-Dideoxy-5"-(O-benzylcarbamoyl)-6,3',6',2",3"',4" -hexa-O-benzoyl-4'-iodo-1,3,2',2"',6" -penta-N-trifluoroacetyl
paromomycin (29). A stirred solution of 4'-epi-hydroxy
paromomycin derivative 28 (368 mg, 0.20 mmol, 1.0 equiv) in
anhydrous DCM (10 mL) was cooled to 0 °C (crushed ice bath) and
treated with anhydrous pyridine (0.12 mL, 1.5 mmol, 7.5 equiv).

Trifluoromethanesulfonic anhydride (0.10 mL, 0.60 mmol, 3.0 equiv) was then added dropwise over a period of 5 min. Upon completion of the addition the yellow solution turned brownish. After stirring at 0 °C for 1 h, the resulting mixture was washed with aqueous 5 % KHSO₄ solution (15 mL). Organic layer was washed with saturated aqueous NaHCO₃ solution (15 mL) and water (20 mL). Organic layer was dried over anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. The brownish oily residue was dissolved in anhydrous acetone (10 mL) and potassium iodide (330 mg, 2.0 mmol, 10.0 equiv) was added. The resulting well-stirred suspension was heated at at 55 °C for 6 h, and the reaction progress was monitored by UPLC-MS assay. After cooling to room temperature, the brown suspension was filtered through the pad of *Celite*[®]. The filter plug was washed with DCM, the filtrate was dried over anhydrous Na₂SO₄ and evaporated under reduced pressure. The crude product was purified by silica gel column

chromatography using gradient elution from 25 % EtOAc in hexane to 50 % EtOAc in hexane. Productcontaining fractions (detected with TLC and ESI-MS) were combined and concentrated under reduced pressure. The residue was dissolved in DCM (5 mL) and hexane (50 mL) was added. The formed white precipitate was collected by suction filtration and dried under reduced pressure to give product **29** (297 mg, 76 % yield) as a white powder. An aliquot of **29** (60 mg) was purified by reversed-phase preparative HPLC (XBridge[®] Prep C18 OBDTM column) using gradient elution from 50 % to 95 % MeCN in 0.1 % aqueous AcOH solution; analytical TLC on silica gel, 1:1 EtOAc/petroleum ether, R_f =0.62. [α]²³_D+21.5 (c 1.21, MeOH).

¹**H NMR** (600 MHz, CD₃OD) δ: 8.28-8.21 (m, 2H), 8.11-8.05 (m, 4H), 8.03-7.99 (m, 2H), 7.80 – 7.73 (m, 3H), 7.66 – 7.58 (m, 5H), 7.52 – 7.41 (m, 9H), 7.32 – 7.22 (m, 7H), 7.12-7.05 (m, 1H), 6.99 – 6.93 (m, 2H), 6.14 – 6.06 (m, 1H), 5.72 – 5.64 (m, 1H), 5.36 – 5.29 (m, 2H), 5.26 (t, *J* = 3.0 Hz, 1H), 5.23 – 5.12 (m, 3H), 5.12 – 5.06 (m, 2H), 4.84 – 4.80 (m, 2H), 4.69 (dd, *J* = 10.5, 4.0 Hz, 1H), 4.51 – 4.39 (m, 4H), 4.37 – 4.29 (m, 2H), 4.25 (dd, *J* = 8.4, 4.7 Hz, 1H), 4.15 – 4.07 (m, 1H), 3.92 – 3.85 (m, 1H), 3.84 – 3.79 (m, 1H), 3.66 – 3.57 (m, 1H), 3.55 – 3.45 (m, 2H), 3.36 (dd, *J* = 14.4, 5.7 Hz, 1H), 2.17 (q, *J* = 12.8 Hz, 1H), 2.05 (dt, *J* = 12.8, 4.3 Hz, 1H) ppm, 6H exchangeable protons merge with H₂O.

¹³C{¹H} NMR (151 MHz, CD₃OD) δ: 167.6, 167.2, 166.8, 166.4, 166.2, 165.5, 159.8 – 158.4 (5×COCF₃),
138.2, 135.0, 134.9, 134.7, 134.6, 134.4, 134.2, 131.3, 131.0, 131.0, 130.9, 130.9, 130.7, 130.6, 130.5,
130.5, 129.9, 129.9, 129.8, 129.6, 129.6, 129.6, 129.5, 129.4, 129.1, 129.0, 128.9, 128.6, 118.6 – 115.9
(5×CF₃), 110.0, 98.7, 97.7, 85.6, 81.4, 78.2, 78.0, 76.7, 76.7, 74.6, 73.8, 72.6, 69.7, 67.7, 67.7, 66.7, 54.2,
50.6, 50.3, 49.7, 49.6, 43.3, 40.8, 32.2, 25.0 ppm.

HRMS (ESI/Q-TOF): m/z [M+Na]⁺ Calcd for C₈₃H₇₀N₆O₂₅F₁₅NaI 1985.3093; Found 1985.3099.



4'-Allyl-4',5"-dideoxy-5"-(*O*-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-*O*-benzoyl-1,3,2',2"',6"'-penta-*N*trifluoroacetyl paromomycin (26). The title compound was obtained from 4'-iodo-paromomycin 29 (100 mg, 0.050 mmol, 1.0 equiv), 4CzIPN (2 mg, 0.0025mmol, 0.05 equiv), allylphenyl sulfone (40 μL, 0.255 mmol, 5.0 equiv) and NEt₃ (25 μL,

0.155 mmol, 3.0 equiv) in degassed DMSO (2 mL) by following General Procedure A. Pure product 26 was

obtained as a white powder (54 mg, 58 % yield) after purification by reversed-phase preparative HPLC. Compound **26** had spectra data identical to the above isolated sample.



4',5"-Dideoxy-5"-(O-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-O-benzoyl-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (30-H). The title compound was formed as a side-product in the photochemical allylation of 4'-iodo-paromomycin **29** (see above). Purification of crude product mixture by reversed-phase preparative HPLC afforded **30-H** as a white powder (17 mg, 19 %

yield); analytical TLC on silica gel, 1:1 EtOAc/petroleum ether, $R_f=0.30$; $[\alpha]_D^{23}$ +38.3 (c 0.34, MeOH).

¹**H NMR** (400 MHz, CD₃CN) δ: 8.35 – 8.14 (m, 3H), 8.09 – 7.92 (m, 8H), 7.88 – 7.71 (m, 4H), 7.71 – 7.56 (m, 7H), 7.54 – 7.39 (m, 10H), 7.36 – 7.24 (m, 7H), 7.10 – 7.06 (m, 1H), 6.99 – 6.95 (m, 1H), 6.25 – 6.06 (m, 1H), 6.01 – 5.84 (m, 1H), 5.39 – 5.22 (m, 4H), 5.21 – 5.17 (m, 1H), 5.15 – 5.04 (m, 3H), 5.02 – 4.67 (m, 2H), 4.54 – 4.38 (m, 3H), 4.09 – 3.84 (m, 3H), 3.74 – 3.59 (m, 2H), 3.49 – 3.35 (m, 1H), 3.27 – 3.11 (m, 1H), 2.46 – 2.37 (m, 1H), 2.14 – 2.04 (m, 2H).

¹³C{¹H} NMR (101 MHz, CD₃CN) δ: 166.9, 166.6, 166.6, 166.5, 165.8, 165.6, 165.0, 157.9-157.1 (5xCOCF₃), 137.9, 134.9, 134.8, 134.7, 134.4, 134.4, 134.3, 134.2, 134.1, 134.0, 130.9, 130.8, 130.8, 130.6, 130.5, 130.4, 130.3, 130.3, 130.2, 129.7, 129.7, 129.5, 129.5, 129.3, 129.1, 129.0, 128.9, 128.8, 128.8, 128.5, 128.4, 118.5 – 116.0 (5×CF₃, overlapped with MeCN), 115.5, 109.4, 98.2, 97.9, 97.0, 84.7, 82.7, 81.5, 81.0, 78.3, 78.0, 76.9, 76.6, 76.3, 75.7, 72.9, 69.5, 69.2, 69.1, 67.6, 67.2, 67.1, 66.3, 63.3, 53.5, 50.8, 49.9, 49.7, 49.6, 49.4, 49.1, 44.1, 40.7, 33.5, 31.6.

HRMS (ESI/Q-TOF): *m*/*z* [M+Na]⁺ Calcd for C₈₃H₇₁N₆O₂₅F₁₅Na 1859.4127; Found 1859.4147.

CF₃COHN OBz NHCOCF₃ NHCOCF₃ OBz OBz NHCOCF₃ 4',5"-Dideoxy-4'-propyl-5"-amino-6,3',6',2",3"",4"''-hexa-Obenzoyl-1,3,2',2"',6"''-penta-N-trifluoroacetyl paromomycin (31). 5 % Pd on carbon (1.69 g, 0.79 mmol, 3.0 equiv) was added at ambient temperature to a stirred solution of alkene 26 (498 mg, 0.26 mmol, 1.0 equiv) in 80 % aqueous acetic acid (12.4 mL, prepared by mixing 10.1 mL of glacial acetic acid and 2.3 mL

of water) and the mixture was stirred under 5 atm of hydrogen at ambient temperature for 18 h. The reaction progress was monitored by UPLC-MS assay. The black suspension was filtered through a silica gel pad, the filter plug was washed with 80 % aqueous acetic acid and the filtrate was evaporated under reduced pressure. The yellowish residue was dissolved in DCM (5 mL) and hexane (50 mL) was added to the solution. The formed beige precipitate was collected by suction filtration and dried under reduced pressure to give product **31** (463 mg, 100 % yield) as a white powder. An aliquot of **31** (100 mg) was purified by reversed-phase preparative HPLC (XBridge[®] Prep C18 OBDTM column) using gradient elution from 5 % to 95 % MeCN in 0.1 % aqueous AcOH solution. Product-containing fractions were detected by TLC and ESI-MS (visualization of TLC spots was done by immersing the TLC plate into a solution of sulfuric acid in ethanol (1:15 v:v) and subsequent drying the TLC plate with heat-gun). Combined fractions were concentrated under reduced pressure to afford pure material; analytical TLC on silica gel, 1:1 EtOAc/petroleum ether, R_f =0.16. $[\alpha]_D^{23}$ +34.9 (c 1.26, MeOH).

¹**H NMR** (400 MHz, CD₃OD) δ: 8.32 – 8.28 (m, 2H), 8.16-8.06 (m, 4H), 8.02 – 7.99 (m, 2H), 7.80 – 7.75 (m, 1H), 7.74 – 7.68 (m, 4H), 7.67 – 7.60 (m, 3H), 7.57 – 7.50 (m, 3H), 7.50 – 7.43 (m, 4H), 7.43 – 7.38 (m, 2H), 7.25-7.18 (m, 2H), 6.99 – 6.88 (m, 3H), 6.02 (d, J = 4.0 Hz, 1H), 5.50 (t, J = 10.7 Hz, 1H), 5.37 (s, 1H), 5.34 – 5.26 (m, 3H), 5.24 – 5.21 (m, 1H), 5.11 (d, J = 4.0 Hz, 1H), 4.68 (dd, J = 12.5, 2.4 Hz, 1H), 4.56 – 4.49 (m, 2H), 4.47 – 4.39 (m, 2H), 4.37 – 4.25 (m, 2H), 4.25 – 4.20 (m, 1H), 4.14 – 4.06 (m, 2H), 3.96 (t, J = 2.4 Hz, 1H), 3.86 – 3.75 (m, 2H), 3.36 (dd, J = 14.1, 3.9 Hz, 1H), 3.06 (dd, J = 13.7, 3.6 Hz, 1H), 2.83 (dd, J = 13.7, 7.4 Hz, 1H), 2.32 (tt, J = 10.9, 3.6 Hz, 1H), 2.19 (q, J = 12.9 Hz, 1H), 2.03 (dt, J = 12.9, 4.4 Hz, 1H), 1.54 – 1.37 (m, 3H), 1.33 – 1.24 (m, 1H), 0.79 (t, J = 6.8 Hz, 3H) ppm, 7H exchangeable protons merge with H₂O.

¹³C{¹H} NMR (101 MHz, CD₃OD) δ: 167.8, 167.6, 167.3, 166.4, 165.9, 165.6, 160.2 – 157.9 (5×COCF₃), 158.8, 135.0, 134.6, 134.5, 134.5, 134.4, 131.3, 131.1, 131.1, 131.0, 130.7, 130.7, 130.6, 130.6, 130.5, 130.5, 130.0, 129.8, 129.7, 129.7, 129.6, 129.3, 129.2, 129.2, 129.0, 119.0 – 115.2 (5×CF₃), 110.0, 98.2, 97.5, 86.8, 81.3, 77.7, 77.7, 77.1, 76.1, 75.0, 73.0, 71.2, 69.9, 67.6, 65.0, 54.4, 50.7, 50.0, 49.8, 49.5, 49.3, 44.8, 42.8, 40.6, 32.1, 30.4, 20.5, 14.7 ppm.

HRMS (ESI/Q-TOF): *m/z* [M+H]⁺ Calcd for C₇₈H₇₂N₆O₂₃F₁₅ 1745.4409; Found 1745.4438.

2,2,2-Trifluoro-*N*-(**2-hydroxyethyl**)**acetamide (S1)**. The title compound was prepared according to the literature procedure.² Thus, to a stirred solution of ethyl-2,2,2-trifluoroacetate (15.7 g, 111 mmol, 1.35 equiv) in anhydrous DCM (10 mL) at 0 °C (crushed ice bath) was

added 2-amino-1-ethanol (5.0 mL, 82 mmol, 1.0 equiv). After warming up to room temperature, the colorless solution was stirred at ambient temperature for 2 h and then concentrated to dryness under reduced pressure to give product S1 as a colorless viscous oil (12.3 g, 96 % yield). The oil solidified as a white solid after standing for several days. ¹H NMR (300 MHz, CDCl₃) δ : 6.83 (br s, 1H), 3.81 (t, J = 5.2 Hz, 2H), 3.55 (dt, J = 5.5, 5.2 Hz, 2H), 1.94 (br s, 1H) ppm, which was identical to that from the literature.¹

2,2,2-Trifluoro-N-(2-oxoethyl)acetamide (32). An oven-dried pressure tube CF_3 N H V (100 mL) was cooled under a stream of argon and charged with 2,2,2-trifluoro-*N*-(2 budgestamide **\$1**, 1.50 g. 9.55 mmol, 1.0 equiv), IBX (2.94 g, (2-hydroxyethyl)acetamide **S1**, 1.50 g, 9.55 mmol, 1.0 equiv), IBX (2.94 g,

10.5 mmol, 1.1 equiv) and anhydrous THF (25 mL). After heating at 60 °C for 18 h, the orange suspension was cooled to room temperature and filtered through the pad of Celite®. The filter plug was washed with THF and filtrate was evaporated to dryness under reduced pressure to give product 32 as a brownish solid (1.48 g, 100 % yield) which was used further without additional purification. ¹H NMR (400 MHz, CDCl₃) δ : 9.72 (s, 1H), 7.02 (br s, 1H), 4.35 (dd, J = 4.8, 0.6 Hz, 2H) ppm.



 $\begin{array}{c} O \\ CF_{3} \end{array} \begin{pmatrix} O \\ N \end{pmatrix} \begin{pmatrix} O \\ H \end{pmatrix} \\ H \end{pmatrix} = \begin{array}{c} \textbf{2,2,2-Trifluoro-N-(3-oxopropyl)acetamide (33)}. \ \text{To a cooled (0 °C, crushed ice bath) neat ethyl-2,2,2-trifluoroacetate (1.78 g, 12.6 mmol, 1.35 equiv) was } \end{array}$ added neat 3,3-diethoxypropan-1-amine (1.5 mL, 9.3 mmol, 1.0 equiv). The

yellowish solution was warmed up to room temperature, stirred at ambient temperature for 3 h and then concentrated to dryness under reduced pressure to give N-(3,3-diethoxypropyl)-2,2,2-trifluoroacetamide as a yellow oil (2.26 g, 100 % yield).²

The obtained N-(3,3-diethoxypropyl)-2,2,2-trifluoroacetamide (2.26 g, 9.3 mmol, 1.0 equiv) was added dropwise to aqueous HCI (1.0 M solution in water, 30 mL) over a period of 15 min at 0 °C (crushed ice bath). After warming to ambient temperature and stirring for 1 h, the yellowish solution was extracted with Et₂O (3×40 mL). Combined organic extracts were dried over anhydrous Na₂SO₄, filtered and concentrated to dryness under reduced pressure to give product 33 as a colorless oil (1.27 g, 81 % yield) which was used in further steps without additional purification. ¹H NMR (400 MHz, CDCl₃) δ : 9.83 (s, 1H), 6.84 (br s, 1H), 3.66 (dt, J = 5.9, 5.6 Hz, 2H), 2.85 (t, J = 5.6 Hz, 2H) ppm, which was identical to that from the literature.³



General Procedure B for reductive amination of amine 31 with aldehydes 32 and 33. An oven-dried 50 mL round-bottom flask was cooled under an argon atmosphere and charged with amine 31

(1.0 equiv) and aldehydes **32** or **33** (2.0 equiv), followed by anhydrous THF (5 mL/0.1 mmol of the amine). The clear solution was stirred at ambient temperature for 10 min, then Na(OAc)₃BH (4.0 equiv) was added. The resulting pale yellow suspension was stirred at ambient temperature for 18 h and the reaction progress was monitoring by UPLC-MS assay. Upon complete conversion, the yellowish suspension was evaporated under reduced pressure. The crude residue was diluted with water (20 mL) and extracted with EtOAc (3×20 mL). Combined organic extracts were washed with brine (15 mL), dried over anhydrous Na₂SO₄, filtered and evaporated under reduced pressure. The residue pressure. The residue was dissolved in DCM (5 mL) and hexane (50 mL) was added. The formed pale yellow precipitate was filtered off, washed with hexane (30 mL), dried under reduced pressure and used in the next step without further purification.



4',5"-Dideoxy-4'-propyl-5"-(2-trifluoroacetamidoethylamino)-6,3',6',2",3"',4"'-hexa-O-benzoyl-1,3,2',2"',6"'-penta-Ntrifluoroacetyl paromomycin (34). The title compound was obtained as a yellowish solid (648 mg, 100 % yield) from amine 31 (600 mg, 0.34 mmol, 1.0 equiv), aldehyde 32 (107 mg, 0.69 mmol, 2.0 equiv) and Na(OAc)₃BH (292 mg, 1.38 mmol, 4.0 equiv) by

following General Procedure B. The crude product 34 was used in further without additional purification.



4',5"-Dideoxy-4'-propyl-5"-(3-trifluoroacetamidopropylamino)-6,3',6',2",3",4" hexa-O-benzoyl-1,3,2',2",6" -penta-N-trifluoroacetyl paromomycin (35). The title compound was obtained as a yellowish solid (653 mg, 100 % yield) from amine 31 (600 mg, 0.34 mmol, 1.0 equiv), aldehyde 33 (116 mg, 0.69 mmol, 2.0 equiv) and Na(OAc)₃BH (292 mg, 1.38 mmol,

4.0 equiv) by following General Procedure B. The crude product **35** was used in further steps without additional purification.

General Procedure C for trifluoroacetylation of amines 34 and 35.

An oven-dried 50 mL round-bottom flask was cooled under an argon atmosphere and charged with amine **34** and **35** (1.0 equiv), followed by anhydrous DCM (5 mL/0.1 mmol of the amine). The clear solution was cooled to 0 °C (crushed ice) and trifluoroacetic anhydride (4.0–5.0 equiv) was added dropwise within 5 min. The resulting yellowish solution was stirred at 0 °C for 30 min, then pyridine (2.0-2.5 equiv) was added dropwise within 5 min at 0 °C temperature. The yellowish solution was stirred at ambient temperature for 2 h and the reaction progress was monitored by UPLC-MS assay. Upon complete conversion water (1 mL) was added to the dark brownish solution and all volatiles were removed under reduced pressure. The crude product was diluted with aqueous HCl (0.5 M solution in water, 50 mL) and back-extracted with EtOAc (3×30 mL). Combined organic extracts were washed with brine (20 mL), dried over anhydrous Na₂SO₄, filtered through a silica gel pad and evaporated under reduced pressure. The residue was purified by reversed-phase preparative HPLC (XBridge[®] Prep C18 OBDTM column) using gradient elution from 70 % to 95 % MeCN in 0.1 % aqueous AcOH solution. The obtained product after chromatography and evaporation was dissolved in DCM (5 mL) and hexane (50 mL) was added. The formed pale yellow precipitate was filtered off, washed with hexane (30 mL) and dried under reduced pressure.



4',5"-Dideoxy-4'-propyl-5"-(2-aminoethylamino)-

6,3',6',2",3"',4"'-hexa-O-benzoyl-hepta-N-trifluoroacetyl paromomycin (36). The title compound was obtained from amine 34 (648 mg, 0.34 mmol, 1.0 equiv), trifluoroacetic anhydride (0.19 mL, 1.38 mmol, 4.0 equiv) and pyridine (55 μL, 0.69 mmol, 2.0 equiv) by following General Procedure C. Purification by

reversed-phase preparative HPLC afforded **36** as a white powder (224 mg, 33 % yield); analytical TLC on silica gel, 1:1 EtOAc/petroleum ether, R_{f} =0.31.

¹**H NMR** (600 MHz, CD₃OD) δ : 8.29 – 8.24 (m, 2H), 8.13 – 8.10 (m, 2H), 8.08 – 7.98 (m, 4H), 7.80 – 7.77 (m, 1H), 7.74 – 7.59 (m, 7H), 7.55 – 7.44 (m, 9H), 7.28 – 7.21 (m, 2H), 7.00-6.95 (m, 1H), 6.94-6.90 (m, 1H), 6.90-6.86 (m, 1H), 6.01-5.97 (m, 1H), 5.47 – 5.40 (m, 1H), 5.29 (dd, *J* = 10.5, 9.3 Hz, 1H), 5.25-5.23 (m, 1H), 5.22 – 5.17 (m, 1H), 5.16 (d, *J* = 4.4 Hz, 1H), 5.10 – 5.07 (m, 1H), 4.69 – 4.63 (m, 1H), 4.61 – 4.54 (m, 2H), 4.47 – 4.43 (m, 1H), 4.42 – 4.36 (m, 2H), 4.36 – 4.29 (m, 2H), 4.18 – 4.03 (m, 4H), 4.01 – 3.93 (m, 1H), 3.79 (dt, *J* = 13.9, 5.2 Hz, 2H), 3.75 – 3.68 (m, 1H), 3.66 – 3.58 (m, 2H), 3.56 – 3.47 (m, 2H), 3.44 (dd, *J* = 13.9, 8.4 Hz, 1H), 2.38 – 2.26 (m, 1H), 2.12 (q, *J* = 12.8 Hz, 1H), 2.05 (dt, *J* = 12.8, 4.4 Hz, 1H), 1.48 – 1.35 (m, 3H),

1.31 – 1.23 (m, 1H), 0.78 (dt, *J* = 10.5, 6.9 Hz, 3H) ppm, 6H exchangeable protons merge with H₂O. $[\alpha]_D^{23}$ +34.0 (*c* 1.21, MeOH).

¹³C{¹H} NMR (151 MHz, CD₃OD) δ: 167.9, 167.6, 167.2, 166.2, 166.1, 165.7, 159.8 – 157.6 (7×COCF₃), 135.1, 135.0, 134.6, 134.5, 134.4, 131.3, 131.1, 131.0, 130.8, 130.8, 130.7, 130.7, 130.6, 130.0, 129.9, 129.8, 129.7, 129.7, 129.6, 129.5, 129.3, 129.2, 129.0, 118.6 – 115.7 (7×CF₃), 110.2, 98.4, 97.8, 85.9, 79.5, 78.4, 78.0, 77.8, 77.6, 76.8, 76.2, 75.7, 74.2, 73.5, 73.1, 71.4, 69.9, 68.0, 65.0, 54.1, 50.6, 49.6, 48.3, 46.9, 42.7, 40.5, 37.7, 32.3, 30.4, 20.8, 14.6 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+Na]⁺ Calcd for C₈₄H₇₄N₇O₂₅F₂₁Na 2002.4297; Found 2002.4412.



4',5"-Dideoxy-4'-propyl-5"-(3-aminoethylamino)-1,3diaminopropane)-6,3',6',2",3",4"'-hexa-O-benzoyl-hepta-*N*-trifluoroacetyl paromomycin (37). The title compound was obtained from amine 35 (653 mg, 0.34 mmol, 1.0 equiv), trifluoroacetic anhydride (0.19 mL, 1.38 mmol, 4.0 equiv) and pyridine (55 μL, 0.69 mmol, 2.0 equiv) by following General

Procedure C. Purification by reversed-phase preparative HPLC afforded **37** as a white powder (232 mg, 34 % yield); analytical TLC on silica gel, 1:1 EtOAc/petroleum ether, $R_f=0.31$. $[\alpha]_D^{23}$ +29.0 (*c* 1.14, MeOH).

¹**H NMR** (600 MHz, CDCl₃) δ : 8.63 (dd, *J* = 9.0, 3.6 Hz, 1H), 8.27 – 8.18 (m, 2H), 8.14 – 8.07 (m, 2H), 7.96 – 7.91 (m, 4H), 7.77 – 7.70 (m, 2H), 7.68 – 7.61 (m, 4H), 7.60 – 7.49 (m, 6H), 7.49 – 7.40 (m, 8H), 7.32 (t, *J* = 6.2 Hz, 1H), 7.25 – 7.22 (m, 1H, overlapped with CHCl₃), 6.84 (d, *J* = 7.6 Hz, 1H), 6.79 (t, 1H, J=7.0 Hz), 6.74 (t, *J* = 7.6 Hz, 2H), 6.60 (d, *J* = 9.7 Hz, 1H), 5.94 (d, *J* = 4.0 Hz, 1H), 5.42 (s, 1H), 5.37 – 5.30 (m, 2H), 5.20 (t, *J* = 9.9 Hz, 1H), 5.15 (t, *J* = 2.7 Hz, 1H), 5.04 – 4.97 (m, 2H), 4.66 (dd, *J* = 12.1, 3.6 Hz, 1H), 4.53 (dd, *J* = 12.1, 5.1 Hz, 1H), 4.46 (dt, *J* = 10.2, 4.0 Hz, 1H), 4.39 (dd, *J* = 8.0, 4.3 Hz, 1H), 4.33 – 4.24 (m, 2H), 4.24 – 4.15 (m, 4H), 4.13-4.07 (m, 2H), 3.99 – 3.91 (m, 2H), 3.62 (p, *J* = 7.6 Hz, 1H), 3.57 – 3.50 (m, 1H), 3.36 (dd, *J* = 13.9, 3.6 Hz, 1H), 1.67 (q, *J* = 12.7 Hz, 1H), 1.45 – 1.37 (m, 3H), 1.35-1.27 (m, 1H), 0.80 (t, *J* = 6.8 Hz, 3H) ppm, 6H exchangeable protons merge with H₂O.

¹³C{¹H} NMR (151 MHz, CDCl₃) δ: 167.8, 166.8, 166.7, 164.7, 164.6, 164.4, 158.8 – 155.8 (7×COCF₃), 134.4, 134.1, 134.0, 133.8, 133.7, 133.6, 130.4, 130.0, 129.9, 129.7, 129.7, 129.6, 129.6, 129.1, 129.0, 128.9, 128.9, 128.8, 128.7, 128.4, 128.1, 128.0, 127.8, 126.9, 117.4 – 114.2 (7×CF₃), 109.0, 97.6, 96.0, 85.2, 79.0,

75.1, 74.9, 74.7, 73.7, 71.8, 70.2, 69.6, 66.6, 65.6, 53.4, 49.5, 48.9, 47.4, 45.2, 43.7, 42.7, 41.5, 39.5, 36.4, 32.1, 29.2, 29.0, 26.1, 20.4, 14.4 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+Na]⁺ Calcd for C₈₅H₇₆N₇O₂₅F₂₁Na 2016.4453; Found 2016.4429.



4',5"-Dideoxy-4'-propyl-5"-(*O*-benzylcarbamoyl)-6,3',6',2",3"',4"''-hexa-*O*-benzoyl-1,3,2',2"',6"''-penta-*N*trifluoroacetyl paromomycin (38). A stirred solution of 5"-deoxy-5"-amino paromomycin derivative **31** (515 mg, 0.30 mmol, 1.0 equiv) in anhydrous DCM (20 mL) was cooled to 0 °C (crushed ice bath) and Cbz-Cl (126 μL, 0.89 mmol, 3.0 equiv) was added

dropwise over 5 min. The resulting yellowish solution was stirred at 0 °C for 1 h followed by dropwise addition (over 5 min) of pyridine (71 µL, 0.89 mmol, 3.0 equiv). The resulting yellow solution was stirred at 0 °C for 1 h, then warmed up to room temperature and stirring was continued for additional 2 h. The reaction progress was monitored by UPLC-MS assay. The yellowish solution was diluted with water (50 mL) and back-extracted with DCM (2×30 mL). Combined organic extracts were washed with brine (50 mL), dried over anhydrous Na₂SO₄ and evaporated under reduced pressure. The crude yellow oil was dissolved in DCM (5 mL) and hexane (50 mL) was added to the solution. The formed beige precipitate was collected by suction filtration and dried under reduced pressure to give product **38** (520 mg, 94 % yield) as a white powder. An aliquot of **38** (50 mg) was purified by reversed-phase preparative HPLC (XBridge[®] Prep C18 OBDTM column) using gradient elution from 60 % to 95 % MeCN in 0.1 % aqueous AcOH solution. Product-containing fractions were detected by TLC and ESI-MS. Combined fractions were concentrated under reduced pressure to afford pure material; analytical TLC on silica gel, 1:1 EtOAc/petroleum ether, R = 0.62. [α] $_{D^3}^{23}$ +75.7 (c 0.53, CHCl₃).

¹**H NMR** (600 MHz, CDCl₃) δ: 8.66 (t, J = 5.6 Hz, 1H), 8.13 – 8.03 (m, 4H), 7.99 – 7.95 (m, 2H), 7.94 – 7.91 (m, 2H), 7.91 – 7.83 (m, 2H), 7.74 – 7.67 (m, 2H), 7.65 – 7.60 (m, 2H), 7.60 – 7.56 (m, 2H), 7.51 – 7.45 (m, 5H), 7.44 – 7.39 (m, 4H), 7.37 – 7.29 (m, 5H), 7.28 – 7.27 (m, 1H), 7.26 – 7.20 (m, 3H), 7.14 – 7.04 (m, 2H), 6.87 – 6.81 (m, 1H), 6.81 – 6.73 (m, 1H), 6.02 – 5.91 (m, 1H), 5.67 (d, J = 3.7 Hz, 1H), 5.39 – 5.31 (m, 2H), 5.29 – 5.21 (m, 2H), 5.15 – 5.08 (m, 3H), 5.01 (t, J = 10.0 Hz, 1H), 4.81 (s, 1H), 4.59 – 4.50 (m, 2H), 4.42 – 4.37 (m, 1H), 4.33 – 4.28 (m, 1H), 4.19 – 4.15 (m, 1H), 4.14 – 4.09 (m, 2H), 4.09 – 4.05 (m, 2H), 4.01 – 3.94 (m, 2H), 3.78 – 3.71 (m, 1H), 3.62 (ddd, J = 14.2, 6.1, 2.9 Hz, 1H), 3.37 (ddd, J = 14.5, 8.8, 5.4 Hz, 1H), 3.33

– 3.24 (m, 1H), 2.30 – 2.22 (m, 1H), 2.09 – 2.02 (m, 1H), 1.58 – 1.51 (m, 1H), 1.44 – 1.38 (m, 2H), 0.78 (t, J = 7.0 Hz, 3H).

¹³C{¹H} NMR (151 MHz, CDCl₃) δ: 167.2, 166.9, 165.7, 164.9, 164.1, 163.1, 157.8, 157.7-156.5 (5×COCF₃),
136.4, 134.4, 134.2, 134.2, 134.0, 133.9, 133.7, 130.3, 129.9, 129.9, 129.8, 129.6, 129.0, 128.9, 128.8,
128.8, 128.7, 128.7, 128.4, 128.3, 128.3, 128.2, 127.9, 127.4, 117.0-114.8 (5×CF₃), 99.1, 96.2, 81.3, 78.2,
77.4, 77.2, 76.9, 75.5, 74.2, 72.7, 70.8, 70.7, 69.0, 67.3, 67.1, 64.8, 53.8, 49.7, 49.5, 48.7, 42.0, 41.0, 31.5,
29.4, 19.8, 19.6, 14.4 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+H]⁺ Calcd for C₈₆H₇₇N₆O₂₅F₁₅Na 1901.4597; Found 1901.4622.



4',5"-Dideoxy-4'-propyl-5"-formamido-6,3',6',2",3"',4"'-hexa-O-benzoyl-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (39). A stirred solution of 5"-deoxy-5"-amino paromomycin derivative **31** (500 mg, 0.29 mmol, 1.0 equiv) in anhydrous DCM (20 mL) was treated with formyl acetate (113 μ L, 1.43 mmol, 5 equiv) at ambient temperature. The resulting yellowish solution

was stirred at ambient temperature overnight. The reaction progress was monitored by UPLC-MS assay. The yellowish solution was evaporated under reduced pressure. The crude yellow oil was purified by reversed-phase preparative HPLC (XBridge[®] Prep C18 OBDTM column) using gradient elution from 60 % to 95 % MeCN in 0.1 % aqueous AcOH solution. Product-containing fractions were detected by TLC and ESI-MS. Combined fractions were concentrated under reduced pressure to give product **39** (290 mg, 57 % yield) as a white powder; analytical TLC on silica gel, 1:1 EtOAc/petroleum ether, R_f =0.56. [α]²³_D +60.4 (c 0.49, CHCl₃).

¹**H NMR** (600 MHz, CDCl₃) δ : 9.02 (s, 1H), 8.19 (s, 1H), 8.15 – 8.05 (m, 4H), 8.02 – 7.96 (m, 2H), 7.94 – 7.90 (m, 2H), 7.88 – 7.81 (m, 2H), 7.74 – 7.68 (m, 2H), 7.68 – 7.64 (m, 2H), 7.64 – 7.57 (m, 3H), 7.57 – 7.39 (m, 9H), 7.35 – 7.30 (m, 1H), 7.29 – 7.26 (m, 2H), 7.16 (t, J = 7.4 Hz, 1H), 7.13 – 7.04 (m, 2H), 6.91 – 6.75 (m, 3H), 5.72 (d, J = 3.6 Hz, 1H), 5.42 – 5.33 (m, 2H), 5.30 – 5.25 (m, 1H), 5.23 (t, J = 3.0 Hz, 1H), 5.10 (d, J = 4.6 Hz, 1H), 5.02 (t, J = 9.9 Hz, 1H), 4.78 – 4.71 (m, 1H), 4.64 (dd, J = 12.2, 6.1 Hz, 1H), 4.58 – 4.52 (m, 1H), 4.42 – 4.34 (m, 1H), 4.23 – 3.95 (m, 9H), 3.77 – 3.67 (m, 2H), 3.53 – 3.46 (m, 1H), 3.41 – 3.34 (m, 1H), 2.30 – 2.22 (m, 1H), 2.15 – 2.09 (m, 1H), 1.64 – 1.56 (m, 1H), 0.80 (t, J = 7.0 Hz, 3H).

¹³C{¹H} NMR (151 MHz, CDCl₃) δ: 167.1, 166.8, 166.8, 165.5, 164.8, 164.1, 162.5, 158.2-156.6 (5×COCF₃),
134.3, 134.1, 134.0, 133.9, 133.6, 130.2, 129.7, 129.7, 129.7, 129.4, 129.0, 128.8, 128.7, 128.7, 128.6, 128.6, 128.2, 128.0, 128.0, 127.3, 116.9-114.4 (5×CF₃), 107.3, 98.0, 95.8, 80.6, 77.2, 77.0, 76.8, 74.8, 74.0,
72.9, 70.9, 70.6, 69.0, 67.0, 64.6, 53.7, 49.3, 48.3, 41.9, 40.8, 38.2, 31.4, 29.4, 19.8, 14.3 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+H]⁺ Calcd for C₇₉H₇₂N₆O₂₄F₁₅ 1773.4358; Found 1773.4353.



4',5"-Dideoxy-4'-propyl-5"-(N-benzylureido)-6,3',6',2",3"',4"'hexa-O-benzoyl-1,3,2',2"',6"'-penta-N-trifluoroacetyl
paromomycin (40). A stirred solution of 5"-deoxy-5"-amino
paromomycin derivative 31 (720 mg, 0.41 mmol, 1.0 equiv) in
anhydrous DCM (25 mL) was treated with benzyl isocyanate
(130 μL, 1.03 mmol, 2.5 equiv) at ambient temperature. The

resulting yellowish solution was stirred at ambient temperature overnight. The reaction progress was monitored by UPLC-MS assay. The yellowish solution was diluted with water (50 mL) and back-extracted with DCM (3×30 mL). Combined organic extracts were washed with brine (30 mL), dried over anhydrous Na₂SO₄ and evaporated under reduced pressure. The crude yellow oil was purified by reversed-phase preparative HPLC (XBridge[®] Prep C18 OBDTM column) using gradient elution from 60 % to 95 % MeCN in 0.1 % aqueous AcOH solution. Product-containing fractions were detected by TLC and ESI-MS. Combined fractions were concentrated under reduced pressure to give product **40** (350 mg, 45 % yield) as a white powder; analytical TLC on silica gel, 1:1 EtOAc/petroleum ether, R_f =0.59. [α]_D²³ +44.5 (*c* 0.31, CHCl₃).

¹**H NMR** (600 MHz, CDCl₃) δ : 9.78 (s, 1H), 8.12 – 8.08 (m, 2H), 8.06 – 8.03 (m, 2H), 7.96 – 7.92 (m, 2H), 7.92 – 7.89 (m, 2H), 7.88 – 7.82 (m, 2H), 7.71 – 7.65 (m, 3H), 7.63 (t, J = 7.5 Hz, 1H), 7.60 – 7.54 (m, 4H), 7.50 – 7.46 (m, 3H), 7.44 – 7.35 (m, 6H), 7.32 – 7.26 (m, 7H), 7.26 – 7.21 (m, 3H), 7.10 – 7.03 (m, 2H), 6.81 – 6.61 (m, 2H), 5.84 (d, J = 3.7 Hz, 1H), 5.62 (dd, J = 8.5, 3.8 Hz, 1H), 5.44 (t, J = 6.0 Hz, 1H), 5.33 – 5.27 (m, 2H), 5.26 – 5.22 (m, 2H), 5.11 (d, J = 5.0 Hz, 1H), 4.86 (t, J = 9.9 Hz, 1H), 4.78 – 4.73 (m, 1H), 4.69 – 4.63 (m, 1H), 4.55 – 4.48 (m, 2H), 4.41 – 4.33 (m, 3H), 4.20 – 4.14 (m, 2H), 4.09 – 4.03 (m, 3H), 3.99 – 3.93 (m, 2H), 3.85 (t, J = 9.5 Hz, 1H), 3.68 (dd, J = 13.9, 5.7 Hz, 1H), 3.39 – 3.32 (m, 1H), 3.13 – 3.01 (m, 1H), 2.29 (dt, J = 13.6, 4.6 Hz, 1H), 2.03 – 1.99 (m, 1H), 0.87 – 0.83 (m, 3H).

¹³C{¹H} NMR (151 MHz, CDCl₃) δ: 167.6, 167.2, 166.7, 165.3, 164.9, 164.0, 159.1, 158.7-157.0 (5×COCF₃), 139.5, 134.5, 134.3, 134.1, 134.0, 133.6, 130.2, 129.9, 129.9, 129.8, 129.8, 129.6, 129.2, 128.9, 128.8,

128.8, 128.8, 128.7, 128.7, 128.3, 128.2, 128.2, 127.4, 127.3, 127.2, 117.1-114.4 (5×CF₃), 107.2, 99.7, 95.1, 81.9, 81.4, 79.5, 77.4, 77.2, 76.9, 75.8, 74.4, 73.0, 70.9, 70.8, 68.8, 65.1, 53.6, 49.7, 48.9, 48.7, 44.3, 42.4, 42.1, 41.5, 31.7, 29.5, 29.2, 20.5, 19.6, 14.4 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+H]⁺ Calcd for C₈₆H₇₉N₇O₂₄F₁₅ 1878.4937; Found 1878.4929.



4',5"-Dideoxy-4'-propyl-5"-(N-trifluoroacetylglycinamido)6,3',6',2",3"',4"'-hexa-O-benzoyl-1,3,2',2"',6"'-penta-Ntrifluoroacetyl paromomycin (41). A stirred solution of (2,2,2trifluoroacetyl)glycine (34 mg, 0.20 mmol, 1.4 equiv), HATU (82
mg, 0.22 mmol, 1.5 equiv) in anhydrous DMF (3 mL) at ambient
temperature was treated with 4-methylmorpholine (40 μL,

0.36 mmol, 2.5 equiv). The resulting yellowish solution was stirred at ambient temperature for 1 h followed by addition of 5"-deoxy-5"-amino paromomycin **31** (250 mg, 0.14 mmol, 1.0 equiv) solution in anhydrous DMF (2 mL). The resulting yellowish solution was stirred at ambient temperature overnight. The reaction progress was monitored by UPLC-MS assay. The yellowish solution was diluted with saturated aqueous NaHCO₃ solution (50 mL) and back-extracted with DCM (3×30 mL). Combined organic extracts were washed with brine (30 mL), dried over anhydrous Na₂SO₄ and evaporated under reduced pressure. The crude yellow oil was purified by reversed-phase preparative HPLC (XBridge[®] Prep C18 OBDTM column) using gradient elution from 60 % to 95 % MeCN in 0.1 % aqueous AcOH solution. Product-containing fractions were detected by TLC and ESI-MS. Combined fractions were concentrated under reduced pressure to give product **41** (145 mg, 53 % yield) as a white powder; analytical TLC on silica gel, 1:1 EtOAc/petroleum ether, R_f =0.54. [α]²³ +49.9 (*c* 0.33, MeOH).

¹**H NMR** (600 MHz, CD₃OD) δ : 8.30 – 8.23 (m, 2H), 8.13 – 8.05 (m, 4H), 8.05 – 7.99 (m, 2H), 7.80 – 7.73 (m, 3H), 7.71 – 7.66 (m, 2H), 7.66 – 7.59 (m, 3H), 7.56 – 7.51 (m, 2H), 7.51 – 7.42 (m, 7H), 7.27 – 7.21 (m, 2H), 7.06 – 7.01 (m, 1H), 6.98 – 6.91 (m, 2H), 6.03 (d, J = 4.0 Hz, 1H), 5.48 (t, J = 10.7 Hz, 1H), 5.34 (s, 1H), 5.33 – 5.26 (m, 2H), 5.20 – 5.12 (m, 3H), 4.66 – 4.53 (m, 3H), 4.47 – 4.43 (m, 1H), 4.39 – 4.28 (m, 3H), 4.23 (dd, J = 8.5, 4.5 Hz, 1H), 4.16 – 4.03 (m, 4H), 3.98 (dd, J = 14.2, 2.8 Hz, 1H), 3.91 – 3.83 (m, 2H), 3.57 (d, J = 6.5 Hz, 2H), 3.19 (dd, J = 14.2, 7.6 Hz, 1H), 2.44 – 2.36 (m, 1H), 2.20 – 2.12 (m, 1H), 2.07 – 2.01 (m, 1H), 1.47 – 1.40 (m, 2H), 1.35 – 1.22 (m, 2H), 0.78 (t, J = 6.9 Hz, 3H) ppm, 7H exchangeable protons merge with H₂O.

¹³C{¹H} NMR (151 MHz, CD₃OD) δ: 170.4, 167.9, 167.6, 167.3, 166.5, 166.1, 165.6, 160.2-158.0 (6xCOCF₃), 135.0, 134.9, 134.6, 134.6, 134.5, 134.3, 131.3, 131.1, 131.1, 131.0, 130.8, 130.7, 130.7, 130.5, 130.0, 129.9, 129.8, 129.6, 129.6, 129.6, 129.3, 129.3, 129.1, 118.9-115.6 (6×CF₃), 110.0, 98.9, 97.6, 85.9, 81.3, 78.8, 77.8, 76.9, 76.5, 73.9, 72.9, 71.4, 69.8, 67.8, 64.9, 54.5, 50.6, 50.2, 49.6, 49.4, 49.2, 49.0, 48.8, 48.6, 48.4, 43.6, 43.0, 42.6, 40.9, 32.3, 30.5, 20.8, 14.7 ppm.

HRMS (ESI/Q-TOF): *m/z* [M+Na]⁺ Calcd for C₈₂H₇₃N₇O₂₅F₁₈Na 1920.4266; Found 1920.4257.

General Procedure D for global deprotection of trifluoroacetamides 36, 37, 39 and 41.

An oven-dried 20 mL pressure vial was cooled under an argon atmosphere and charged with trifluoroacetamides 36, 37, 39, or 41 (1.0 equiv). Anhydrous MeOH (4 mL/0.1 mmol of the starting material) was added and the resulting clear solution was treated under argon atmosphere with $Mg(OMe)_2$ (7-8 % solution in anhydrous methanol; 23.0-30.0 equiv). After stirring for 24 h at 65 °C temperature (the reaction progress was monitored by UPLC-MS assay), the resulting white slurry was concentrated to dryness under reduced pressure. Anhydrous Et₂O (10 mL) was added to the white residue and the resulting white suspension was sonicated for 2 min in an ultrasonic bath, whereupon the resulting white suspension was centrifuged for 10 min at 2150 rpm. The colorless clear supernatant was decanted and fresh anhydrous Et₂O (10 mL) was added to the white residue. The sonication/centrifugation sequence was repeated two more times. The resulting white precipitate was dried under reduced pressure. The residue was taken up in water (10 mL/0.1 mmol of the starting material), stirred and treated with Ba(OH)₂ (20.0 equiv) at ambient temperature. The resulting white slurry was stirred and heated to 80 °C temperature for 18 h and the reaction progress was monitored by UPLC-MS assay. The white reaction suspension was cooled to 0 °C (crushed ice bath) and dry ice was added portionwise until pH 7–8. The resulting mixture was centrifuged for 10 min at 2150 rpm. The clear supernatant was decanted and fresh MeOH (10 mL) was added to the white precipitate. The mixture was sonicated for 2 min in an ultrasonic bath. Fresh MeOH (10 mL) was added once again and the sonication/centrifugation sequence was repeated two more times. The combined supernatant solutions were concentrated under reduced pressure. The crude product was diluted with MeOH (15 mL), the resulting suspension was filtered, and the filter cake was washed with MeOH. The combined filtrates were concentrated under reduced pressure. The residue was purified by reversed-phase preparative HPLC (XBridge® BEH Prep OBD[™] Amide column) using gradient elution from 95:5 A:B to 10:90 A:B (eluent A - 0.05 % solution of AcOH in MeCN;

eluent B - 0.05 % solution of AcOH in water). Product-containing fractions (identified by ninhydrin stain test and ESI-MS) were combined and concentrated. The residue was treated with glacial acetic acid (0.5 mL), followed by addition of MeCN (5 mL). The formed white solid was dried under reduced pressure.

General Procedure E for global deprotection of trifluoroacetamides 38 and 40.

An oven-dried 20 mL pressure vial was cooled under an argon atmosphere and charged with trifluoroacetamides **38** or ,**40** (1.0 equiv). Anhydrous MeOH (4 mL/0.1 mmol of the starting material) was added and the resulting clear solution was treated under argon atmosphere with Mg(OMe)₂ (7-8 % solution in anhydrous methanol; 23.0-30.0 equiv). After stirring for 24 h at 65 °C temperature (the reaction progress was monitored by UPLC-MS assay), the resulting white slurry was concentrated to dryness under reduced pressure. Anhydrous Et₂O (10 mL) was added to the white residue and the resulting white suspension was sonicated for 2 min in an ultrasonic bath, whereupon the resulting white suspension was centrifuged for 10 min at 2150 rpm. The colorless clear supernatant was decanted and fresh anhydrous Et₂O (10 mL) was added to the white residue. The sonication/centrifugation sequence was repeated two more times. The resulting white precipitate was dried under reduced pressure. The residue was taken up in water (10 mL/0.1 mmol of the starting material), stirred and treated with Ba(OH)₂ (20.0 equiv) at ambient temperature. The resulting white slurry was stirred and heated to 80 °C temperature for 18 h and the reaction progress was monitored by UPLC-MS assay. The white reaction suspension was cooled to 0 °C (crushed ice bath) and dry ice was added portionwise until pH 7–8. The resulting mixture was centrifuged for 10 min at 2150 rpm. The clear supernatant was decanted and fresh MeOH (10 mL) was added to the white precipitate. The mixture was sonicated for 2 min in an ultrasonic bath. Fresh MeOH (10 mL) was added once again and the sonication/centrifugation sequence was repeated two more times. The combined supernatant solutions were concentrated under reduced pressure. The crude product was diluted with MeOH (15 mL), the resulting suspension was filtered, and the filter cake was washed with MeOH. The combined filtrates were concentrated under reduced pressure. The residue was diluted with AcOH (16 mL) and water (4 mL), and to the well-stirred solution was added 10 % Pd on carbon (1.0 equiv) at ambient temperature to a stirred solution. The reaction mixture was stirred under 5 atm of hydrogen at ambient temperature for 18 h, and progress of the reaction was monitored by UPLC-MS assay. The black suspension was filtered through the pad of *Celite*[®], the filter plug was washed with 50% AcOH in water and the filtrate was evaporated under reduced pressure. The residue was purified by reversed-phase preparative HPLC (XBridge[®] BEH Prep OBD[™] Amide

column) using gradient elution from 95:5 A:B to 10:90 A:B (eluent A - 0.05 % solution of AcOH in MeCN; eluent B - 0.05 % solution of AcOH in water). Product-containing fractions (identified by ninhydrin stain test and ESI-MS) were combined and concentrated. The residue was treated with glacial acetic acid (0.5 mL), followed by addition of MeCN (5 mL). The formed white solid was dried under reduced pressure.



5"-Amino-4',5"-dideoxy-4'-propyl paromomycin pentaacetate (8). The title compound was obtained from paromomycin derivative **38** (550 mg, 0.29 mmol, 1.0 equiv), $Mg(OMe)_2$ (7-8 % solution in anhydrous methanol; 10.0 mL, 7.02 mmol, 24.0 equiv), barium hydroxide (1.0 g, 5.85 mmol, 20.0 equiv) and Pd/C (10 mol% on carbon; 247 mg, 0,23 mmol, 1 equiv) by following General Procedure E.

Purification by reversed-phase preparative HPLC, followed by treatment with glacial acetic acid afforded the hexaacetate salt of **8** as a white amorphous solid (75.0 mg, 32 % yield). $[\alpha]_D^{23}$ +39.1 (*c* 0.81, D₂O).

¹**H NMR** (600 MHz, D₂O) δ: 5.78 (1H, d, J=3.8 Hz) 5.47 (1H, s) 5.33 (1H, s) 4.56 (1H, t, J=5.9 Hz) 4.48 (1H, d, J=5.2 Hz) 4.35 (1H, t, J=5.2 Hz) 4.34-4.30 (1H, m) 4.26 (1H, t, J=3.1 Hz) 4.04-3.93 (3H, m) 3.93-3.84 (3H, m) 3.78-3.72 (2H, m) 3.64-3.60 (1H, m) 3.52-3.41 (4H, m) 3.41-3.27 (3H, m) 2.44 (1H, dt, J=12.1, 3.4 Hz) 1.97 (15H, s) 1.82 (1H, q, J=12.7 Hz) 1.73-1.67 (1H, m) 1.56-1.47 (2H, m) 1.41-1.27 (2H, m) 0.91 (3H, t, J=7.2 Hz) ppm, 24H exchangeable protons merge with H₂O.

¹³C{¹H} NMR (151 MHz, D₂O) δ: 108.8, 95.3, 95.1, 83.0, 77.4, 77.0, 76.8, 74.0, 73.0, 72.1, 70.2, 67.6, 67.4, 67.1, 61.1, 55.2, 50.8, 50.2, 50.0, 49.2, 42.1, 41.7, 40.4, 28.5, 27.9, 22.5, 18.6, 13.7 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+H]⁺ Calcd for C₂₆H₅₃N₆O₁₂ 641.3721; Found 641.3732.

Elemental analysis: Anal. Calcd for C₂₈H₅₇N₇O₁₂•5AcOH•4H₂O•4H₂CO₃: C, 38.10; H, 7,03; N, 6.66; Found C, 38.25; H, 6.80; N, 6.67.



5"-(2-Aminoethylamino)-4',5"-dideoxy-4'-propyl paromomycin **heptaacetate (9)**. The title compound was obtained from paromomycin derivative **36** (223 mg, 0.11 mmol, 1.0 equiv), Mg(OMe)₂ (7-8 % solution in anhydrous methanol; 3.7 mL, 2.6 mmol, 23.0 equiv) and barium hydroxide (386 mg, 2.25 mmol, 20.0 equiv) by following General Procedure D.

Purification by reversed-phase preparative HPLC, followed by treatment with glacial acetic acid afforded the heptaacetate salt of **9** as a white amorphous solid (25.0 mg, 20 % yield). $[\alpha]_D^{23}$ +38.5 (*c* 1.50, D₂O).

¹**H NMR** (600 MHz, D_2O) δ : 5.71 (d, J = 3.9 Hz, 1H), 5.39 (d, J = 3.3 Hz, 1H), 5.32 (d, J = 1.8 Hz, 1H), 4.45 (t, J = 5.3 Hz, 1H), 4.38 (dd, J = 5.3, 3.3 Hz, 1H), 4.34 – 4.30 (m, 1H), 4.27 – 4.21 (m, 2H), 3.99 – 3.86 (m, 5H), 3.85 – 3.83 (m, 1H), 3.76 – 3.68 (m, 2H), 3.62 – 3.59 (m, 1H), 3.49 – 3.38 (m, 3H), 3.37 – 3.29 (m, 2H), 3.18 (t, J = 6.9 Hz, 2H), 3.10 (dd, J = 12.9, 3.8 Hz, 1H), 3.05 – 3.00 (m, 2H), 2.94 (dd, J = 12.9, 8.5 Hz, 1H), 2.43 (dt, J = 12.7, 4.2 Hz, 1H), 1.80 (q, J = 12.7 Hz, 1H), 1.71 – 1.64 (m, 1H), 1.55 – 1.45 (m, 2H), 1.41 – 1.26 (m, 2H), 0.90 (t, J = 7.2 Hz, 3H) ppm, 26H exchangeable protons merge with H₂O.

¹³C¹H NMR (151 MHz, D₂O) δ: 109.3, 96.1, 95.4, 83.9, 79.7, 78.0, 77.4, 74.0, 73.2, 72.1, 70.3, 67.6, 67.3, 67.0, 61.2, 55.3, 50.9, 50.8, 49.7, 49.2, 45.5, 42.3, 40.4, 37.9, 28.7, 28.0, 18.6, 13.7 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+H]⁺ Calcd for C₂₈H₅₈N₇O₁₂ 684,4143; Found 684,4145.

Elemental analysis: Anal. Calcd for C₂₈H₅₇N₇O₁₂•7AcOH•2H₂O: C, 44,24; H, 7,87; N, 8,60; Found C, 44,24; H, 7,68; N, 8,36.

x 7 AcOH Åн.

(10). The title compound was obtained from paromomycin derivative 37 (231 mg, 0.12 mmol, 1.0 equiv), Mg(OMe)₂ (7-8 % solution in anhydrous methanol; 4.4 mL, 3.1 mmol, 27.0 equiv) and barium hydroxide (397 mg, 2.31 mmol, 20.0 equiv) by following General Procedure D. Purification by reversed-phase preparative HPLC was followed by treatment with glacial acetic acid to afford the heptaacetate salt of 10 as a white amorphous solid (28.9 mg, 22 % yield). $[\alpha]_{D}^{23}$ +49.8 (c 1.52, D₂O).

5"-(3-Aminopropylamino)-4',5"-dideoxy-4'-propyl paromomycin heptaacetate

¹**H NMR** (600 MHz, D_2O) δ : 5.67 (d, J = 3.8 Hz, 1H), 5.44 (d, J = 2.6 Hz, 1H), 5.30 (d, J = 1.9 Hz, 1H), 4.51 (t, J = 5.6 Hz, 1H), 4.43 (dd, J = 5.1, 2.6 Hz, 1H), 4.35 – 4.30 (m, 2H), 4.24 (t, J = 3.1 Hz, 1H), 3.94 – 3.82 (m, 6H), 3.75 – 3.69 (m, 2H), 3.61 – 3.57 (m, 1H), 3.47 – 3.35 (m, 4H), 3.34 – 3.28 (m, 1H), 3.26 – 3.19 (m, 2H), 3.13 – 3.05 (m, 4H), 2.38 (dt, J = 12.7, 4.2 Hz, 1H), 2.12 – 2.01 (m, 2H), 1.74 (q, J = 12.7 Hz, 1H), 1.69 – 1.63 (m, 1H), 1.57 – 1.44 (m, 2H), 1.41 – 1.25 (m, 2H), 0.90 (t, J = 7.2 Hz, 3H) ppm, 26H exchangeable protons merge with H_2O .

¹³C¹H} NMR (151 MHz, D₂O) δ: 108.8, 96.2, 95.3, 83.2, 77.8, 77.4, 77.2, 73.8, 72.9, 72.2, 70.4, 67.7, 67.4, 67.3, 61.2, 55.5, 50.8, 50.4, 50.1, 49.3, 45.2, 42.3, 40.4, 36.8, 29.2, 27.9, 24.5, 18.4, 13.8 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+H]⁺ Calcd for C₂₉H₆₀N₇O₁₂ 698,4300; Found 698,4333.

Elemental analysis: Anal. Calcd for C₂₉H₅₉N₇O₁₂•7AcOH•2H₂O: C, 44,75; H, 7,95; N, 8,49; Found C, 44,79; H, 8,07; N, 8,40.



4',5''-Dideoxy-5''-formamido-4'-propyl paromomycin pentaacetate (11). The title compound was obtained from paromomycin derivative **39** (330 mg, 0.19 mmol, 1.0 equiv), Mg(OMe)₂ (7-8 % solution in anhydrous methanol; 6.5 mL, 4.6 mmol, 24.0 equiv) and barium hydroxide (651 mg, 3.8 mmol, 20.0 equiv) by following General Procedure D. Purification by reversed-phase preparative HPLC was followed by treatment with glacial

acetic acid to afforded the pentaacetate salt of **11** as a white amorphous solid (45 mg, 27 % yield). $[\alpha]_D^{23}$ +36.5 (*c* 0.33, D₂O).

¹**H NMR** (600 MHz, D₂O) δ: 8.19 (1H, s) 5.69 (1H, d, J=3.9 Hz) 5.34 (1H, d, J=3.5 Hz) 5.31-5.29 (1H, m) 4.44 (1H, t, J=5.6 Hz) 4.36-4.31 (2H, m) 4.28-4.24 (2H, m) 3.95-3.84 (6H, m) 3.73-3.67 (3H, m) 3.63-3.60 (1H, m) 3.56 (1H, dd, J=14.6, 6.0 Hz) 3.49-3.39 (4H, m) 3.37-3.31 (1H, m) 2.44 (1H, dt, J=12.3, 3.9 Hz) 1.80 (1H, q, J=12.6 Hz) 1.72-1.66 (1H, m) 1.56-1.48 (2H, m) 1.41-1.29 (2H, m) 0.91 (3H, t, J=7.2 Hz) ppm, 23H exchangeable protons merge with H₂O.

¹³C{¹H} NMR (151 MHz, D₂O) δ: 164.9 (folded signal), 109.7, 96.2, 95.6, 84.1, 79.9, 79.4, 79.1, 76.8, 74.3, 73.0, 72.2, 70.3, 70.2, 67.6, 67.4, 61.2, 55.4, 50.8, 49.7, 49.1, 42.3, 40.5, 39.4, 28.8, 27.9, 22.6, 18.5, 13.7 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+H]⁺ Calcd for C₂₇H₅₃N₆O₁₃ 669.3671; Found 669.3654.

Elemental analysis: Anal. Calcd for C₂₇H₅₂N₆O₁₃•6AcOH•4H₂O: C, 42,54; H, 7,69; N, 7,63; Found C, 42,44; H, 7,27; N, 7,56.



4',5''-Dideoxy-4'-propyl-5''-ureido-paromomycin pentaacetate (12). The title compound was obtained from paromomycin derivative **40** (350 mg, 0.19 mmol, 1.0 equiv), Mg(OMe)₂ (7-8 % solution in anhydrous methanol; 6.3 mL, 4.4 mmol, 24 equiv), barium hydroxide (638 mg, 3.7 mmol, 20.0 equiv) and Pd/C (10 mol% on carbon; 193 mg, 0.18 mmol, 1 equiv) by following General Procedure E. Purification by reversed-phase preparative

HPLC was followed by treatment with glacial acetic acid to afforded the pentaacetate salt of **12** as a white amorphous solid (55 mg, 31 % yield). $[\alpha]_D^{23}$ +227.8 (*c* 0.48, D₂O).

¹**H NMR** (600 MHz, D₂O) δ: 5.67 (1H, d, J=4.0 Hz) 5.34 (1H, d, J=3.9 Hz) 5.31 (1H, d, J=1.8 Hz) 4.45 (1H, t, J=5.3 Hz) 4.36-4.31 (2H, m) 4.26-4.22 (2H, m) 3.97-3.89 (4H, m) 3.88 (1H, d, J=8.7 Hz) 3.85 (1H, t, J=1.8 Hz) 3.75-3.68 (2H, m) 3.63-3.61 (1H, m) 3.53 (1H, dd, J=14.6, 4.9 Hz) 3.50-3.44 (2H, m) 3.44-3.32 (4H, m) 2.45 (1H, dt, J=12.8, 4.3 Hz) 1.81 (1H, q, J=12.8 Hz) 1.73-1.66 (1H, m) 1.56-1.47 (2H, m) 1.42-1.26 (2H, m) 0.91 (3H, t, J=7.2 Hz) ppm, 24H exchangeable protons merge with H₂O.

¹³C{¹H} NMR (151 MHz, D₂O) δ: 151.4, 109.7, 96.5, 95.7, 84.2, 80.6, 79.7, 77.1, 74.4, 72.8, 72.1, 67.6, 67.4, 67.2, 67.0, 61.2, 55.5, 50.8, 49.6, 49.2, 42.3, 41.5, 40.5, 28.7, 27.9, 27.7, 22.5, 18.5, 13.8 ppm.

HRMS (ESI/Q-TOF): m/z [M+H]⁺ Calcd for C₂₇H₅₄N₇O₁₃ 684.3780; Found 684.3764.

Elemental analysis: Anal. Calcd for C₂₇H₅₃N₇O₁₃•6AcOH•5H₂O: C, 41,04; H, 7.73; N, 8.65; Found C, 41.26; H, 7.07; N, 8,44.



4',5"-Dideoxy-5"-glycinamido-4'-propyl paromomycin hexacetate (13). The title compound was obtained from paromomycin derivative **41** (295 mg, 0.16 mmol, 1.0 equiv), Mg(OMe)₂ (7-8 % solution in anhydrous methanol; 5.2 mL, 3.8 mmol, 24.0 equiv) and barium hydroxide (532 mg, 3.11 mmol, 20.0 equiv) by following General Procedure D. Purification by reversed-phase preparative HPLC was followed by treatment with glacial

acetic acid to afford the hexaacetate salt of **13** as a white amorphous solid (75 mg, 43 % yield). $[\alpha]_D^{23}$ +42.6 (*c* 0.50, D₂O).

¹**H NMR** (600 MHz, D₂O) δ: 5.77 (0.40H, d, J=3.8 Hz) 5.67 (0.6H, d, J=3.8 Hz) 5.46 (0.40H, d, J=2.0 Hz) 5.36 (0.60H, d, J=3.8 Hz) 5.32 (1H, dd, J=5.8, 2.0 Hz) 4.56 (0.40H, dd, J=6.8, 5.1 Hz) 4.47 (0.40H, dd, J=5.0, 2.0 Hz) 4.39 (0.60H, t, J=5.6 Hz) 4.36-4.33 (1H, m) 4.33-4.27 (1H, m) 4.27-4.22 (1.60H, m) 3.97-3.94 (1H, m) 3.93-3.89 (3H, m) 3.88-3.84 (3H, m) 3.79-3.69 (3H, m) 3.63-3.60 (1H, m) 3.55-3.48 (1H, m) 3.48-3.39 (4H, m) 3.37-3.26 (2H, m) 2.42 (1H, ddt, J=12.4, 8.2, 4.2 Hz) 1.96 (20H, s) 1.77 (1H, q, J=12.4 Hz) 1.73-1.66 (1H, m) 1.57-1.46 (2H, m) 1.41-1.29 (2H, m) 0.91 (3H, t, J=7.1 Hz) ppm, 25H exchangeable protons merge with H₂O.
¹³C{¹H} NMR (151 MHz, D₂O) δ: 156.7, 109.4, 108.9, 96.5, 95.8, 95.6, 95.2, 84.1, 83.1, 80.4, 79.1, 77.4, 77.0, 74.1, 73.9, 73.1, 72.3, 70.2, 67.6, 67.5, 67.2, 61.2, 55.4, 55.3, 50.8, 49.9, 49.2, 42.2, 41.7, 41.5, 40.4, 29.0, 27.9, 22.7, 18.5, 13.7 ppm.

HRMS (ESI/Q-TOF): *m*/*z* [M+H]⁺ Calcd for C₂₈H₅₆N₇O₁₃ 698.3936; Found found 698.3942.

Elemental analysis: Anal. Calcd for C₂₈H₅₅N₇O₁₃•7AcOH•3H₂O: C, 43.04; H, 7.65; N, 8.36; Found C, 43.16; H, 7.37; N, 8.17.

Phenyl 2-azido-2,3,4-trideoxy-4-C-ethyl-1-thio-β-D-glucopyranoside (43). To a solution of 42 (218 mg, 1.10 mmol) in 1,2-dichloroethane (6.5 ml) were added trimethyl(phenylthio)silane (630 μ l, 3.31 mmol) and Znl₂ (1.06 g, 3.31 mmol) and the mixture was stirred for 45 h at room temperature. After diluting with 1,2-dichloroethane (11 ml), the mixuture was passed through a Celite pad and the cake was washed with 1,2-dichloroethane (6.5 ml). The combined filtrate was washed with saturated NaHCO₃ solution and saturated NaCl solution (1×20 ml, each), dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was taken in a solution of methanol/water (10:1, 6 ml) and added K₂CO₃ (305 mg, 2.21 mmol). The mixture was stirred for 1 h at room temperature and then concentrated *in vacuo*. The residue was taken in chloroform (10 ml) and washed with water (1×10 ml). The aqueous phase was extracted with chloroform (2×5 ml). The organic phase was combined, washed with saturated NaCl solution (1×20 ml, dried over MgSO₄, filtered, and concentrated NaCl solution (1×20 ml, dried over MgSO₄, filtered, and the cake was phase was extracted with chloroform (2×5 ml). The organic phase was combined, washed with saturated NaCl solution (1×20 ml), dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was taken in chloroform (2×5 ml). The organic phase was combined, washed with saturated NaCl solution (1×20 ml), dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was purified by Flash column chromatography on silica gel (hexane:ethyl acetate = 12:1 to 6:1) to afford **43** (295 mg, 87%) as an anomeric mixture (α : β = 2.8:1); analytical TLC on silica gel, 3:7 EtOAc/hexanes, *R*_f=0.60.

Analytical data for **43** (α anomer): ¹H NMR (500 MHz, CDCl₃) δ: 7.58 – 7.48 (m, 2H), 7.37 – 7.24 (m, 3H), 5.55 (dd, *J* = 4.9, 1.3 Hz, 1H), 4.04 (ddd, *J* = 10.5, 5.8, 2.5 Hz, 1H), 3.84 (dt, *J* = 12.5, 4.6 Hz, 1H), 3.81 – 3.73 (m, 1H), 3.67 – 3.61 (m, 1H), 2.07 (dtd, *J* = 13.0, 4.3, 1.5 Hz, 1H), 1.82 (t, *J* = 6.2 Hz, 1H), 1.78 – 1.68 (m, 1H), 1.54 (q, *J* = 12.5 Hz, 1H), 1.50 – 1.15 (m, 4H), 0.94 (t, *J* = 7.1 Hz, 3H).

¹³C{¹H} NMR (125 MHz, CDCl₃) δ: 133.6, 132.6, 129.1, 127.7, 88.6, 73.8, 62.8, 59.3, 35.2, 33.1, 29.8, 19.1, 14.2.

S37

Analytical data for **43** (β anomer): ¹H NMR (500 MHz, CDCl₃) δ : 7.58 – 7.48 (m, 2H), 7.37 – 7.24 (m, 3H), 4.50 (d, *J* = 9.9 Hz, 1H), 3.81 – 3.73 (m, 1H), 3.61 – 3.55 (m, 1H), 3.31 (ddd, *J* = 11.5, 9.9, 4.8 Hz, 1H), 3.24 (ddd, *J* = 9.6, 6.6, 2.6 Hz, 1H), 2.33 (dt, *J* = 13.0, 4.4 Hz, 1H), 2.16 (t, *J* = 6.6 Hz, 1H), 1.68 – 1.58 (m, 1H), 1.50 – 1.15 (m, 5H), 1.12 – 1.00 (m, 1H), 0.90 (t, *J* = 7.1 Hz, 1H).

¹³C{¹H} NMR (125 MHz, CDCl₃) δ: 132.7, 132.2, 129.0, 128.0, 88.2, 83.3, 63.1, 59.7, 35.6, 35.3, 32.7, 19.2, 14.2.

HRMS (ESI/Q-TOF): *m*/*z* [M+Na]⁺ Calcd for C₁₅H₂₁N₃O₂SNa [M+Na]⁺ 330.1252, found 330.1252.

Phenyl 6-O-acetyl-2-azido-2,3,4-trideoxy-4-C-ethyl-1-thio-β-D-glucopyranoside (44). To a solution of 43 (203 mg, 660 μmol) in pyridine (2.1 ml) was added acetic anhydride (187 μl, 1.98 mmol) and the mixture was stirred for 8 h at room temperature. After completion, the reaction mixture was concentration *in vacuo*, co-evaporated with

toluene, and purified by Flash column chromatography on silica gel (hexane:ethyl acetate = 7:1 to 4:1) to afford **44** (223 mg, 97%) as an anomeric mixture (α : β = 2.7:1); analytical TLC on silica gel, 1:4 EtOAc/hexane, R_f =0.85.

Analytical data for **44** (α anomer): ¹H NMR (500 MHz, CDCl₃) δ : 7.57 – 7.51 (m, 2H), 7.35 – 7.23 (m, 3H), 5.58 (dd, *J* = 4.9, 1.4 Hz, 1H), 4.31 – 4.17 (m, 3H), 3.90 (dt, *J* = 12.5, 4.6 Hz, 1H), 2.13 – 2.06 (m, 1H), 2.04 (s, 3H), 1.78 – 1.70 (m, 1H), 1.55 (q, *J* = 12.5 Hz, 1H), 1.51 – 1.18 (m, 4H), 0.93 (t, *J* = 6.9 Hz, 3H).

¹³C{¹H} NMR (125 MHz, CDCl₃) δ: 170.8, 133.8, 132.7, 132.1, 129.0, 127.5, 88.4, 71.3, 64.2, 59.0, 35.7, 33.2, 30.0, 20.8, 19.0, 14.1.

Analytical data for **44** (β anomer): ¹H NMR (500 MHz, CDCl₃) δ : 7.63 – 7.57 (m, 2H), 7.35 – 7.23 (m, 3H), 4.47 (d, *J* = 9.9 Hz, 1H), 4.35 (dd, *J* = 11.9, 2.3 Hz, 1H), 4.13 (dd, *J* = 12.0, 6.6 Hz, 1H), 3.39 (ddd, *J* = 10.2, 6.6, 2.3 Hz, 1H), 3.34 (ddd, *J* = 11.6, 9.9, 4.9 Hz, 1H), 2.34 (dt, *J* = 13.1, 4.4 Hz, 1H), 2.09 (s, 3H), 1.78 – 1.61 (m, 2H, H4), 1.48 – 1.37 (m, 2H), 1.32 – 1.20 (m, 1H), 1.17 – 1.07 (m, 1H), 0.90 (t, *J* = 7.0 Hz, 3H).

¹³C{¹H} NMR (125 MHz, CDCl₃) δ: 170.9, 133.8, 132.7, 132.6, 128.8, 127.8, 88.4, 80.5, 64.4, 59.4, 35.7, 35.7, 32.9, 20.9, 19.1, 14.1.

HRMS (ESI/Q-TOF): *m/z* [M+Na]⁺ Calcd for C₁₇H₂₃N₃O₃SNa 372.1358, found 372.1352.

^{OAc} N₃ $\stackrel{\circ}{,}$ S $\stackrel{\circ}{,}$ Phenyl 6-O-acetyl-2-azido-2,3,4-trideoxy-4-C-ethyl-1-thio-β-D-glucopyranosyl sulfoxide (45). To a solution of 44 (173 mg, 495 μmol) in mixture of acetonitrile/water (10:1, 4.4 ml) were added, NaHCO₃ (208 mg, 2.47 mmol) and a

solution of Selectfluor (190 mg, 594 µmol) in acetonitrile/water (10:1, 4.4 ml). The reaction mixture was stirred at room temperature for 1 h. After completion, solvents were evaporated under reduced pressure, the residue was taken up in dicloromethane (120 ml) and washed with saturated NaHCO₃ solution:water (1:9), brine solution (1×120 ml, each), dried over MgSO₄, filtered, and concentrated under reduced pressure to give **45** (177 mg, 98%) as an anomeric mixtures (α : β = 2.7:1) which are also a mixture of diastereomer on sulfur. The sulfur of α anomer is most likely with configuration *S*_s. These compounds were used to the next reaction without purification; analytical TLC on silica gel, 1:4 EtOAc/hexane, *R*_f=0.20.

Analytical data for **45** (α anomer with *S*₅ diastereomer): ¹H NMR (500 MHz, CDCl₃) δ: 7.77 – 7.46 (m, 5H), 4.76 (ddd, *J* = 10.7, 5.3, 2.3 Hz, 1H), 4.30 (d, *J* = 5.9 Hz, 1H), 4.15 – 4.08 (m, 1H), 4.06 (dd, *J* = 12.3, 2.4 Hz, 1H), 4.02 (dd, *J* = 12.3, 5.3 Hz, 1H), 2.34 (q, *J* = 12.3 Hz, 1H), 2.22 (dt, *J* = 12.6, 4.4 Hz, 1H), 1.99 (s, 3H), 1.79 – 1.60 (m, 1H), 1.51 – 1.43 (m, 1H), 1.43 – 1.33 (m, 3H), 1.33 – 1.26 (m, 2H), 0.91 (t, *J* = 6.9 Hz, 3H).

¹³C{¹H} NMR (125 MHz, CDCl₃) δ: 170.7, 141.3 130.9, 129.1, 125.5, 91.6, 77.1, 64.4, 58.8, 35.1, 33.1, 29.5, 20.8, 18.9, 14.1.

HRMS (ESI/Q-TOF): *m*/*z* [M+Na]⁺ Calcd for C₁₇H₂₃N₃O₄SNa 388.1307, found 388.1311.



1,3,2',3',4',2''',6'''-Heptadeoxy-4'-propyl-6'-acetyl-1,3,2',2''',6'''-pentaazido-6,2'',5'',3''',4'''-penta-O-benzyl paromomycin (47). Donor **45** (100 mg, 0.27 mmol), acceptor **46** (248 mg, 0.25 mmol), and TTBP (203 mg, 0.82 mmol) were charged to a round-bottom flask, co-evaporated with toluene three times, and dried in *vacuo* overnight. The flask was purged with argon, mixed with activated 4 Å MS and the mixture was dissolved in dry dichloromethane

(4 mL) and stirred at room temperature for 1 h. The reaction mixture was cooled to -70 °C, treated with Tf₂O (48 μ L, 0.29 mmol) for 30 min, warmed slowly to -50 °C and continued stirring for 3 h, at -50 °C before the reaction was quenched with triethylamine (0.2 mL). The reaction mixture was diluted with

EtOAc (15 mL), filtered through Celite[®], and the filtrate was washed with saturated aqueous solution of NaHCO₃ followed by brine. Solvents were evaporated under reduced pressure and the crude product was purified by column chromatography on silica gel (hexanes: ethyl acetate 20:1 to 4:1) to give **47** (147 mg, α : β , 12:1, 48%) as a white foam; analytical TLC on silica gel, 1:4 EtOAc/hexanes, R_f =0.55; $[\alpha]_D^{23}$ +37.5 (c 1, CHCl₃).

Analytical data for **47** (α anomer): ¹**H NMR** (600 MHz, CDCl₃) δ : 7.64 – 6.91 (m, 25H), 6.09 (d, *J* = 3.5 Hz, 1H), 5.69 (d, *J* = 5.6 Hz, 1H), 5.01 (d, *J* = 10.7 Hz, 1H), 4.90 (d, *J* = 1.9 Hz, 1H), 4.74 (d, *J* = 10.7 Hz, 1H), 4.64 (dd, *J* = 21.6, 11.9 Hz, 2H), 4.56 (d, *J* = 11.8 Hz, 1H), 4.53 – 4.43 (m, 3H), 4.39 – 4.26 (m, 5H), 4.21 – 4.15 (m, 1H), 4.01 (t, *J* = 5.2 Hz, 1H), 3.99 – 3.94 (m, 2H), 3.87 (dd, *J* = 10.3, 2.3 Hz, 1H), 3.80 (dq, *J* = 4.9, 2.9, 2.0 Hz, 2H), 3.75 (dd, *J* = 9.7, 8.8 Hz, 1H), 3.67 (dd, *J* = 13.0, 8.5 Hz, 1H), 3.63 – 3.58 (m, 1H), 3.52 – 3.41 (m, 2H), 3.40 – 3.36 (m, 1H), 3.32 (t, *J* = 9.3 Hz, 1H), 3.18 – 3.14 (m, 1H), 2.94 (dd, *J* = 12.9, 4.0 Hz, 1H), 2.87 (ddd, *J* = 12.6, 4.6, 3.5 Hz, 1H), 2.27 (dt, *J* = 13.2, 4.6 Hz, 1H), 2.11 (s, 3H), 1.92 (dt, *J* = 12.4, 4.3 Hz, 1H), 1.83 (q, *J* = 12.3 Hz, 1H), 1.59 (qd, *J* = 14.7, 14.1, 5.2 Hz, 1H), 1.51 – 1.35 (m, 3H), 1.33 – 1.16 (m, 2H), 0.93 (t, *J* = 7.1 Hz, 3H).

¹³C{¹H} NMR (151 MHz, CDCl₃) δ: 171.0, 138.5, 138.0, 137.7, 137.1, 137.0, 128.7, 128.7, 128.5, 128.5, 128.5, 128.5, 128.4, 128.4, 128.4, 128.3, 128.3, 128.2, 128.1, 127.9, 127.9, 127.9, 127.8, 127.7, 127.5, 127.5, 127.4, 106.0, 98.6, 95.4, 84.4, 82.4, 82.0, 81.9, 75.6, 75.0, 74.8, 74.3, 73.3, 73.2, 72.9, 72.4, 71.8, 71.6, 70.9, 70.4, 64.7, 60.6, 60.5, 57.3, 57.2, 51.1, 35.4, 33.3, 32.8, 27.4, 21.0, 19.0, 14.2.

HRMS (ESI/Q-TOF): m/z [M+Na]⁺ Calcd for C₆₃H₇₃N₁₅O₁₃Na 1270.5404, found 1270.5409.



3',4'-Dideoxy-4'-propyl paromomycin pentaacetate (14). A solution of compound **47** (90 mg, 72.1 μ mol) in dioxane: 10% aq. acetic acid in deionized water (1:1, 3 mL) was treated with Pd(OH)₂/C (150 mg) at room temperature under 48 atm of hydrogen for 24 h. After completion as indicated by LRMS, the reaction mixture was filtered through Celite^{*},

filtrate was concentrated to dryness. The crude reaction mixture was taken up in 2.5 mL saturated aqueous barium hydroxide solution and heated to 65 °C for 3 hr. After completion as indicated by LRMS, barium hydroxide was quenched by addition of dry ice (pH =7), the white precipitate was filtered off and filtrate was concentrated to dryness. The crude product was taken up in 10% aqueous acetic acid and

loaded on a CM Sephadex C-25 column, eluted with water followed by a gradient of 0.1% - 1.0% ammonium hydroxide in deionized water. The fractions containing the product were combined and lyophilized with glacial acetic acid to afford **14** (6.6 mg, 41%) as pentaacetate salt in the form of off white solid. $[\alpha]_{D}^{23}$ +50.8 (*c* 0.26, H₂O).

¹**H NMR** (600 MHz, D_2O) δ : 5.38 (d, J = 3.7 Hz, 1H), 5.17 (d, J = 3.0 Hz, 1H), 5.12 (d, J = 1.8 Hz, 1H), 4.32 (t, J = 5.8 Hz, 1H), 4.15 (dd, J = 5.1, 3.1 Hz, 1H), 4.13 (t, J = 4.7 Hz, 1H), 4.05 (t, J = 3.1 Hz, 1H), 4.03 – 3.99 (m, 1H), 3.80 (t, J = 9.6 Hz, 1H), 3.75 – 3.65 (m, 3H), 3.64 (dt, J = 2.8, 1.3 Hz, 1H), 3.58 (dd, J = 12.3, 5.0 Hz, 1H), 3.55 – 3.49 (m, 2H), 3.46 (dd, J = 12.1, 6.8 Hz, 1H), 3.42 – 3.40 (m, 1H), 3.39 – 3.32 (m, 1H), 3.25 (dd, J = 13.7, 6.6 Hz, 1H), 3.21 – 3.14 (m, 2H), 2.30 (dt, J = 12.6, 4.3 Hz, 1H), 1.93 – 1.86 (m, 1H) 1.79 (s, 15H), 1.66 (q, J = 12.7 Hz, 1H), 1.54 (q, J = 11.2, 10.4 Hz, 1H), 1.42 (q, J = 12.3 Hz, 1H), 1.24 – 1.19 (m, 2H), 1.09 – 1.04 (m, 1H), 0.99 (q, J = 9.6 Hz, 1H), 0.68 (t, J = 6.9 Hz, 3H).

¹³C{¹H} NMR (151 MHz, D₂O) δ: 179.3, 109.9, 95.4, 95.3, 84.0, 81.3, 78.0, 75.2, 73.2, 72.1, 70.1, 67.5, 67.2, 61.1, 60.2, 50.7, 49.6, 49.2, 49.0, 40.3, 33.8, 32.0, 28.1, 26.3, 22.0, 18.3, 13.2.

HRMS (ESI/Q-TOF): m/z [M+H]⁺ Calcd for C₂₆H₅₂N₅O₁₂ 626.3612, found 626.3618.



1,3,2',2''',6'''-Penta-N-benzyloxycarbonyl paromomycin (48). A solution of Propylamycin pentaacetate **1** (500 mg, 0.53 mmol) and *N*-(benzyloxycarbonyloxy)succinimide (1.16 g, 4.68 mmol) in DMF (2 mL) was cooled to 0 °C in an ice bath, added DIPEA (1.63 mL, 9.35 mmol) and stirred at 0 °C for 3 h. After completion, the reaction mixture was quenched by addition of 0.5 mL butylamine, diluted

with ethyl acetate (50 mL) and washed sequentially with 0.5 N aq. NaOH solution (10 mL), 1 N HCl solution (10 mL) followed by saturated aqueous NaHCO₃ solution. The aqueous layer was re-extracted in ethyl acetate and the combined organic layer was washed with brine, dried on Na₂SO₄, evaporated, and purified by column chromatography on silica gel (1:3 hexane: ethyl acetate to 20:1 ethyl acetate: methanol) to obtain compound **48** as a white foam (596 mg, 86%); analytical TLC on silica gel, 3:17 Methanol/ethyl acetate, R_f =0.70; $[\alpha]_D^{23}$ +29.4 (*c* 1, MeOH).

HRMS (ESI/Q-TOF): *m*/*z* [M+H]⁺ Calcd for C₆₆H₈₁N₅O₂₃Na 1334.5214; Found 1334.5215.

6',5"-**Bis**-*O*-(2,4,6-triisopropylbenzenesulfonyl)-1,3,2',2"', 6"'-penta-*N*-benzyloxycarbonyl paromomycin (**49**). A solution of compound **48** (530 mg, 0.40 mmol), in pyridine (4 mL) was treated with 2,4,6triisopropylbenzenesulfonyl chloride (367 mg, 1.21 mmol) at room temperature for 18 h. After complete consumption f starting material, Ac₂O (315µL, 3.33 mmol) was added to the reaction mixture and continued stirring at room temperature for another 5 h. The reaction mixture was diluted with ethyl acetate (100 mL), washed with 1 N HCl solution (2x10 mL), saturated aqueous NaHCO₃ and brine solution (1 x15 ml, each), dried over Na₂SO₄, filtered, concentrated under reduced pressure and purified by column chromatography on silica gel (hexnes: ethyl acetate 3:2 to 1:3) to obtain acetylated intermediate (525 mg, 63%) as off white foam; analytical TLC on silica gel, 1:1 EtOAc/Hexane, R_f =0.55; [α]₂²³ +19.7 (*c* 1, CHCl₃).

HRMS (ESI/Q-TOF): *m*/*z* [M+Na]⁺ Calcd for C₁₀₆H₁₃₅N₅O₃₂S₂Na 2076.8429, found 2076.8469.



The acetylated ditrisyl intermediate obtained in previous step (190 mg, 92.4 μ mol) was treated with 0.2 M solution of NaOMe in methanol (5 mL) at room temperature for 8 h. After complete deacetylation, the reaction mixture was neutralized with amberlyst[®] 15, filtered and purified by column chromatography on silica gel (methanol: dichloromethane 0:1 to 1:4) to obtain

compound **49** in 45% yield (136 mg) over 3 steps; analytical TLC on silica gel, 1:19 Methanol/EtOAc, $R_{f}=0.75$; $[\alpha]_{D}^{23} = +30.2$ (*c* 1, CHCl₃).

HRMS (ESI/Q-TOF): *m*/*z* [M+Na]⁺ Calcd for C₉₆H₁₂₅N₅O₂₇S₂Na 1866.7896, found 1866.7882.



6',5"-Dideoxy-bis-N-(2-hydroxyethyl)-1,3,2',2"',6"'-penta-Nbenzyloxycarbonyl paromomycin (50). A solution of compound 49 (70 mg, 37.9 μmol) in 8 mL of ethanolamine was stirred at room temperature for 12 h. After complete conversion as indicated by LRMS, the reaction mixture was cooled diluted with dichloromethane (25 mL) and washed with brine (2x 4 mL). The aqueous layer was

extracted in dichloromethane (4x10 mL), and the combined organic layer was dried over Na₂SO₄, concentrated under reduced pressure and purified by column chromatography on silica gel (methanol: dichloromethane 1:25 to 1:5) to afford compound **49** (19 mg, 40%); analytical TLC on silica gel, 3:17 Methanol/CHCl₃, R_f =0.8; $[\alpha]_D^{23}$ +24.1 (*c* 0.9, MeOH)

HRMS (ESI/Q-TOF): *m*/*z* [M+H]⁺ Calcd for C₇₀H₉₂N₇O₂₃ 1398.6245, found 1398.6238.



6',5"-Dideoxy-bis-*N***-(2-hydroxyethyl)** paromomycin (15). A solution of compound **50** (18 mg, 12.9 μ mol) in dioxane: deionized water (1:1, 1.5 mL) was treated with Pd/C (30 mg) at room temperature under 1 atm of hydrogen (balloon) for 18 h. After completion, as indicated by LRMS, the reaction mixture was filtered through Celite[®], concentrated to dryness and purified by passing through a CM Sephadex C-25 column, loading in 10% aqueous

acetic acid and eluting with deionized water followed by a gradient of 0.1% - 1.0% ammonium hydroxide in deionized water. The fractions containing the product were combined and lyophilized with glacial acetic acid to afford **15** (6.6 mg, 44%) as heptaacetate salt in the form of a white solid. $[\alpha]_D^{23}$ +26.7 (*c* 0.44, H₂O).

¹**H NMR** (600 MHz, D_2O) δ : 5.79 (d, *J* = 3.8 Hz, 1H), 5.34 (d, *J* = 2.2 Hz, 1H), 5.14 (d, *J* = 1.8 Hz, 1H), 4.36 (t, *J* = 5.8 Hz, 1H), 4.27 (dd, *J* = 5.2, 2.3 Hz, 1H), 4.21 (ddd, *J* = 9.6, 6.6, 3.1 Hz, 1H), 4.16 (d, *J* = 5.4 Hz, 1H), 4.07 (t, *J* = 3.1 Hz, 1H), 3.97 (t, *J* = 10.5 Hz, 1H), 3.89 (t, *J* = 9.4 Hz, 1H), 3.83 (td, *J* = 9.8, 9.2, 4.0 Hz, 2H), 3.71 (q, *J* = 6.1, 5.3 Hz, 4H), 3.67 (d, *J* = 3.2 Hz, 1H), 3.66 – 3.56 (m, 1H), 3.43 (d, *J* = 3.0 Hz, 1H), 3.38 (dd, *J* = 13.1, 3.0 Hz, 1H), 3.32 (dd, *J* = 13.1, 2.6 Hz, 1H), 3.29 – 3.12 (m, 6H), 3.09 (dt, *J* = 10.3, 4.6 Hz, 4H), 2.24 – 2.19 (m, 1H), 1.61 (q, *J* = 12.5 Hz, 1H), 1.47 (td, *J* = 10.4, 5.0 Hz, 1H), 1.34 (td, *J* = 11.1, 10.7, 5.5 Hz, 2H), 1.26 – 1.05 (m, 2H), 0.73 (t, *J* = 7.2 Hz, 3H).

¹³C{¹H} NMR (151 MHz, D₂O) δ: 106.9, 94.0, 93.2, 82.1, 75.9, 75.6, 71.7, 70.6, 68.8, 67.7, 66.2, 66.0, 64.8, 60.0, 56.1, 55.2, 54.9, 53.5, 49.4, 48.9, 48.3, 48.2, 48.0, 47.5, 42.6, 39.8, 39.0, 26.5, 21.4, 17.2, 12.3.

HRMS (ESI/Q-TOF): m/z [M+Na]+ Calcd for C₃₀H₆₁N₇O₁₃Na 750.4203, found 750.4225.

No	Bacterial species	Aminoglycoside	Source	Reference
		Resistance Genes		
AG001	Escherichia coli	None	Clinical isolate, IMM	[4]
AG003	Escherichia coli	aac(3)-II	Clinical isolate, IMM	[4]
AG038	Staphylococcus aureus	None	Clinical isolate, IMM	[4]
AG163	Escherichia coli	aph(3')-I	Clinical isolate, IMM	[5]
AG166	Escherichia coli	aph(3')-IIa	Clinical isolate, IMM	[6]
AG173	Escherichia coli	aac(3)-IV	Clinical isolate, IMM	[6]
AG175	Escherichia coli	aac(6')-I	Clinical isolate, IMM	[7]
AG215	Klebsiella pneumoniae	None	Clinical isolate, IMM	[8]
AG220	Pseudomonas aeruginosa	aph(3')-IIb	ATCC 27853	
AG225	Acinetobacter baumannii	None	Clinical isolate, IMM	[4]
AG290	Enterobacter cloacae	None	Clinical isolate, IMM	[4]
EC026	Escherichia coli	None	Laboratory strain DH5 α	[5]
EC102	Escherichia coli	armA	Engineered strain, IMM	[5]
EC103	Escherichia coli	rmtB	Engineered strain, IMM	[5]
EC118	Escherichia coli	aac(3)-IV	Engineered strain, IMM	[5]
EC125	Escherichia coli	aph(3')-IIb	Engineered strain, IMM	[5]
EC141	Escherichia coli	aph(3')-VI	Engineered strain, IMM	[5]
EC189	Escherichia coli	aph(3')-Ia	Engineered strain, IMM	This study
EC191	Escherichia coli	aph(3')-IIa	Engineered strain, IMM	This study
Strains u	sed as source of hybrid riboso	omes for in-vitro trai	nslation assays	
SZ380	Mycobacterium smegmatis	None	Engineered strain, IMM	[9]
SZ480	M. smegmatis Cyt14	None	Engineered strain, IMM	[9]
SZ485	M. smegmatis Mit13	None	Engineered strain, IMM	[10]
SZ496	<i>M. smegmatis</i> Mit A1555G	None	Engineered strain, IMM	[10]

Table S1. Bacterial strains use	ed in this study
---------------------------------	------------------

IMM, Institute of Medical Microbiology, University of Zurich

Cell-free translation inhibition graphs





References

[1] Bobkov, G. V.; Mikhailov, S. N.; Van Aerschot, A.; Herdewijn, P. Phosphoramidite building blocks for efficient incorporation of 2'-*O*-aminoethoxy(and propoxy)methyl nucleosides into oligonucleotides. *Tetrahedron.* **2008**, *64*, 6238–6251.

[2] Durrwachter, J. R.; Wong, C. H. Fructose 1,6-diphosphate aldolase-catalyzed stereoselective synthesis of *C*-alkyl and *N*-containing sugars: thermodynamically controlled C-C bond formations. *J. Org. Chem.* **1988**, *53*, 4175–4181.

[3] Wei, M.; Li, Z.; Li, T.; Wu, B.; Liu, Y.; Qu, J.; Li, X.; Li, L.; Cai, L.; Wang, P. G. Transforming Flask Reaction into Cell-Based Synthesis: Production of Polyhydroxylated Molecules via Engineered *Escherichia coli*. *ACS Catal*. **2015**, *5*, 4060–4065.

[4] Sati, G. C., Shcherbakov, D., Hobbie, S., Vasella, A., Böttger, E. C., and Crich, D. N6', N6'''and O4'-Modifications to Neomycin Affect Ribosomal Selectivity without Compromising Antibacterial Activity. *ACS Infect. Dis.* **2017**, *3*, 368-377.

[5] Sonousi, A.; Quirke, J.C.K.; Waduge, P.; Janusic, T.; Gysin, M.; Haldimann, K.; Xu, S.; Hobbie, S.N.; Sha, S.H.; Schacht, J.; Chow, C.S.; Vasella, A.; Böttger, E.C.; Crich, D. An Advanced Apralog with Increased in vitro and in vivo Activity toward Gram-negative Pathogens and Reduced ex vivo Cochleotoxicity. *Chem. Med. Chem.* **2021**, *16*, 335-339

[6] Quirke, J.C.K.; Rajasekaran, P.; Sarpe, V.A.; Sonousi, A.; Osinnii, I.; Gysin, M.; Haldimann, K.; Fang, Q. J.; Shcherbakov, D.; Hobbie, S.N.; Sha, S.H.; Schacht, J.; Vasella, A.; Böttger, E.C.; Crich, D. Apralogs: Apramycin 5-O-Glycosides and Ethers with Improved Antibacterial Activity and Ribosomal Selectivity and Reduced Susceptibility to the Aminoacyltransferase (3)-IV Resistance Determinant. *J. Am. Chem. Soc.* 2020, *142*, 530-544

[7] Sonousi, A.; Sarpe, V.A.; Brilkova, M.; Schacht, J.; Vasella, A.; Böttger, E.C.; Crich, D. Effects of the 1-*N*-(4-Amino-2*S*-hydroxybutyryl) and 6'-*N*-(2-Hydroxyethyl) Substituents on Ribosomal Selectivity, Cochleotoxicity and Antibacterial Activity in the Sisomicin Class of Aminoglycoside Antibiotics. *ACS Infect. Dis.* **2018**, *4*, 1114-1120.

[8] Mandhapati, A. R.; Yang, G.; Kato, T.; Shcherbakov, D.; Hobbie, S. N.; Vasella, A.; Böttger, E. C.; Crich, D. Structure-Based Design and Synthesis of Apramycin-Paromomycin Analogues. Importance of the Configuration at the 6'-Position and Differences between the 6'-Amino and Hydroxy Series. *J. Am. Chem. Soc.* **2017**, *139*, 14611-14619

[9] Hobbie, S.N.; Akshay, S.; Kalapala, S.; Bruell, C., Shcherbakov, D.; Böttger, EC. Genetic analysis of interactions with eukaryotic rRNA identify the mitoribosome as target in aminoglycoside ototoxicity. *Proc. Natl. Acad. Sci. USA.* **2008**, *105*, 20888-20893

[10] Hobbie, S.N.; Kalapala, S.; Akshay, S.; Bruell, C., Schmidt, S.; Dabow, S.; Vasella, A.; Sander, P.; Böttger, E.C. Engineering the rRNA decoding site of eukaryotic cytosolic ribosomes in bacteria. *Nucleic Acids Res.* **2007**, *35*, 6086-6093

5"-O-tert-Butyldimethylsilyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (17)

[¹H-NMR, 400 MHz, CD₃CN]



5"-O-tert-Butyldimethylsilyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (17)

[¹³C-NMR, 100.6 MHz, CD₃CN]

OSPT2-RZE-5.11.fid NHCOCF3 NHCOCF3 TBDMSC ЪΗ CF₃COHN OH NHCOCF₃ ÓН والفرير فبالموجدة فيسائدون والمراجع والمارج والمرافق والمارك a a Marina da Marina 200 -2(90 f1 (ppm) 30 190 110 100 80 70 60 50 40 20 10 180 170 160 150 140 130 120 ò -10

5"-O-tert-Butyldimethylsilyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (17)

Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Sy ESI+	napt G2-S	i Capillary, k Cone, V:	V: 0. 40	7	LC: Aco	quity UP	LC H-Cla	ss Column:	Acquity UPLC BEH C18 2.1x50mm, 1.7μm
<i>Sample:</i> HRMS_2019_08 MS_POS_2300_	3_406 1 _RES_7mir	000 Zogota R n ACN_F	ZE-6 form_5-9	8_040_	_4min	1:C,4	1.0000	00 MS_T	une Col#43
Elemental Com	position R	Report:							
Tolerance = 3.0 PP Element prediction Number of isotope	M / DBE: : Off peaks used :	: min = -1.5, max for i-FIT = 3	x = 50.0						
Monoisotopic Mass 526 formula(e) eva Elements Used: C: 0-50 H: 1-110	s, Even Elec luated with N: 1-10	etron Ions 2 results within l O: 0-20 F: 15-	imits (all r -15 Na: 1	esults (uj -1 Si: 1	o to 1000) -1) for each 1	nass)		
Mass 1320.3129	RA C 100.00 1	Calc. Mass 320.3153	mDa -2.4	PPM -1.8	DBE 13.5	i-FIT 521.8	Norm 0.079	Conf(%) 92.45	Formula C46 H58 N5 O19 F15 Na



6,3',2",3"',4"'-Penta-O-benzoyl-5"-O-tert-butyldimethylsilyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (18)

[¹H-NMR, 600 MHz, CD₃OD]



6,3',2",3"',4"'-Penta-O-benzoyl-5"-O-tert-butyldimethylsilyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (18)

[¹³C-NMR, 150.9 MHz, CD₃OD]



6,3',2",3"',4"'-Penta-O-benzoyl-5"-O-tert-butyldimethylsilyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (18)

Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Synapt G2-Si	Capillary, kV:	0.7	LC: Acquity UPLC H-Class	Column:	Acquity UPLC BEH C18
ESI+	Cone, V:	40			2.1x50mm, 1.7μm

Sample:

HRMS_2019_08_431 1001 Zogota RZE-7 MS_POS_500-2000_RES_7min_ACN_Form_5-98_040_7min_1:C,5_10.000000_MS_Tune____Col#43

Elemental Composition Report:

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions 349 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass) Elements Used: C: 0-81 H: 1-110 N: 1-10 O: 0-25 F: 15-15 Na: 1-1 Si: 1-1

Mass	RA	Calc. Mass	mDa	PPM	DBE	i-FIT	Norm	Conf(%)	Formula
1840.4443	100.00	1840.4464	-2.1	-1.1	38.5	439.4	n/a	n/a	C81 H78 N5 O24 F15 Na Si

1001 Zogota RZE-7



6,3',2",3"',4"'-Penta-O-benzoyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (19)

[¹H-NMR, 600 MHz, CD₃OD]



6,3',2",3"',4"'-Penta-O-benzoyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (19)

[¹³C-NMR, 150.9 MHz, CD₃OD]



6,3',2",3"',4"'-Penta-O-benzoyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (19)

Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Syn ESI+	napt G2-S	i Capillary, l Cone, V:	<v: 0<br="">4</v:>).7 IO	LC: Ac	quity UP	PLC H-Cla	ass Column:	Acquity 2.1x50r	UPLC BEH C18 nm, 1.7μm
<i>Sample:</i> HRMS_2019_08 MS_POS_500-20	8_433 1 000_RES_	.002 Zogota R _7min ACN_I	ZE-9 Form_5-	98_040	_7min	1:C,6	10.000	0000 MS_T	une	Col#43
Elemental Com	position F	Report:								
Tolerance = 5.0 PP Element prediction Number of isotope	M / DBE : Off peaks used	: min = -1.5, ma for i-FIT = 3	x = 50.0							
Monoisotopic Mass 402 formula(e) eva Elements Used: C: 0-75 H: 1-110	s, Even Elec luated with N: 1-10	ctron Ions 1 results within O: 0-25 F: 15	limits (all -15 Na:	results (u 1-1	p to 1000)) for each 1	mass)			
Mass 1726.3567	RA C 100.00 1	Calc. Mass 726.3599	mDa -3.2	PPM -1.9	DBE 38.5	i-FIT 420.6	Norm n/a	Conf(%) n/a	Formul C75 He	a 54 N5 O24 F15 Na
1002 Zogota RZ HRMS_2019_08_4 100	'E-9 433 1510 (4 771	.224) Cm (1509	:1517-(14	80:1489+	1550:1556	6))				1: TOF MS ES+ 3.33e6



5"-Deoxy-5"-azido-6,3',2",3",4"'-penta-O-benzoyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (20)

[¹H-NMR, 600 MHz, CD₃OD]



5"-Deoxy-5"-azido-6,3',2",3",4"'-penta-O-benzoyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (20)

[¹³C-NMR, 150.9 MHz, CD₃OD]

OSPT2-RZE-38-prep.4.fid



5"-Deoxy-5"-azido-6,3',2",3"',4"'-penta-O-benzoyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (20)

Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Synapt G2-Si ESI+	Capillary, kV: Cone, V:	0.7 40	LC: Acquity UP	PLC H-Class	Column:	Acquity UPLC BEH C18 2.1x50mm, 1.7μm
<i>Sample:</i> HRMS_2019_08_435 100 MS_POS_500-2000_RES_7	03 Zogota RZE-38 min ACN_Form_	5-98_040_	_7min 1:C,7	10.000000	MS_T	une Col#43
Elemental Composition Re	port:					
Tolerance = 5.0 PPM / DBE: n Element prediction: Off Number of isotope peaks used fo	nin = -1.5, max = 50.0 r i-FIT = 3	0				
Monoisotopic Mass, Even Electrr 252 formula(e) evaluated with 1 Elements Used: C: 0-75 H: 1-110 N: 1-10 C	on Ions results within limits (): 0-23 F: 15-15 N	all results (uj Va: 1-1	p to 1000) for each 1	mass)		
Mass RA Cal	le Mass mD	DDM	DBE LEIT	Norm Cor	f(0/a)	Formula

1751.3624	100.00	1751.3664	-4.0	-2.3	40.5	463.4	n/a	n/a	C75 H63 N8 O23 F15 Na
Mass	RA	Calc. Mass	mDa	PPM	DBE	1-1-11	Norm	Conf(%)	Formula

1003 Zogota RZE-38



5"-Deoxy-5"-amino-6,3',2",3"',4"'-penta-O-benzoyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (21)

[¹H-NMR, 600 MHz, CD₃OD]



5"-Deoxy-5"-amino-6,3',2",3"',4"'-penta-O-benzoyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (21)

[¹³C-NMR, 150.9 MHz, CD₃OD]

OSPT2-RZE-21.10.fid summa 5+6 NHCOCF3 NHCOCF3 ЪВz CF3COHN OBz ÓBz. ÓBz NHCOCF3 whitehilter 200 120 100 90 f1 (ppm) 80 20 -10 190 180 170 160 150 140 130 110 70 60 50 40 30 10 0

5"-Deoxy-5"-amino-6,3',2",3"',4"'-penta-O-benzoyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (21)

Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija											
MS: Waters Synapt G2-S ESI+	i Capillary, kV: Cone, V:	0.7 40	LC: Acquity U	PLC H-Clas	s Column:	Acquity UPLC BEH C18 2.1x50mm, 1.7μm					
Sample: HRMS_2019_08_437 1004 Zogota RZE-16 MS_POS_500-2000_RES_7min ACN_Form_5-98_040_7min 1:C,8 10.000000 MS_Tune Col#43 Elemental Composition Report: Col#43 Col#43 Col#43											
Elemental Composition Report: Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3											
Monoisotopic Mass, Even Electron Ions 293 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass) Elements Used: C: 0-75 H: 1-110 N: 1-10 O: 0-23 F: 15-15											
Mass RA C	alc. Mass mD	a PPM	DBE i-FIT	Norm	Conf(%)	Formula C75 H66 N6 O23 E15					



5"-Deoxy-5"-(O-benzylcarbamoyl)-6,3',2",3"',4"'-penta-O-benzoyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (22)

[¹H-NMR, 600 MHz, CDCl₃]

OSPT2-RZE-31-prep2.1.fid NHCOCF3 NHCOCF3 Cbz ÒВz CF3COHN OBz ЭВz ÓBz ŃHCOCF₃ water H grease ,H96.0 S C 2.03-1.92-0.81-К 6 õ 2.0 11.5 11.0 10.5 10.0 9.5 9.0 8.5 8.0 7.5 5.5 f1 (ppm) 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5 -1 7.0 6.5 6.0 5.0 4.5 4.0

5"-Deoxy-5"-(O-benzylcarbamoyl)-6,3',2",3"',4"'-penta-O-benzoyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (22)

[¹³C-NMR, 150.9 MHz, CDCl₃]

OSPT2-RZE-31-prep2.4.fid



5"-Deoxy-5"-(O-benzylcarbamoyl)-6,3',2",3"',4"'-penta-O-benzoyl-4',6'-O-benzylidene-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (22)

Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Synapt G2- ESI+	Si Capillary, kV: Cone, V:	0.7 40	LC: Acc	quity UP	LC H-Cla	ss Column:	Acquity UPLC BEH C18 2.1x50mm, 1.7μm
<i>Sample:</i> HRMS_2019_08_439 MS_POS_500-2000_RES	1005 Zogota RZE-3 _7min ACN_Form	1 _5-98_040_	_7min	1:D,1	10.000	000 MS_1	Fune Col#43
Elemental Composition	Report:						
Tolerance = 5.0 PPM / DBI Element prediction: Off Number of isotope peaks used	$E: \min = -1.5, \max = 50$ 1 for i-FIT = 3	.0					
Monoisotopic Mass, Even Ele 290 formula(e) evaluated with Elements Used: C: 0-83 H: 1-110 N: 1-10	ectron Ions 1 results within limits O: 0-25 F: 15-15	(all results (u Na: 1-1	p to 1000)	for each	mass)		
Mass RA 1859.4125 100.00	Calc. Mass mD 1859.4127 -0.2	a PPM 2 -0.1	DBE 43.5	i-FIT 485.1	Norm n/a	Conf(%) n/a	Formula C83 H71 N6 O25 F15 Na
1005 Zogota RZE-31							



[¹H-NMR, 600 MHz, CD₃OD]



[¹³C-NMR, 150.9 MHz, CD₃OD]

OSPT2-RZE-42-prep.4.fid



Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Synapt G2 ESI+	2- Si Capillary, kV Cone, V:	7: 0.7 40	LC: Acquit	y UPLC H-Cla	iss Column:	Acquity U 2.1x50mr	IPLC BEH C18 n, 1.7μm			
Sample: HRMS_2019_08_441 1006 Zogota RZE-42 MS_POS_500-2000_RES_7min ACN_Form_5-98_040_7min 1:D,2 10.000000 MS_Tune Col#43										
Elemental Composition Report:										
Tolerance = 5.0 PPM / DI Element prediction: Off Number of isotope peaks use	BE: $\min = -1.5$, $\max =$ ed for i-FIT = 3	= 50.0								
Monoisotopic Mass, Even E 307 formula(e) evaluated wi Elements Used: C: 0-76 H: 1-110 N: 1-10	lectron Ions th 2 results within lin 0 O:0-25 F:15-1	nits (all results († 5 Na: 1-1	up to 1000) for a	each mass)						
Mass RA 1771.3860 100.00	Calc. Mass 1771.3814 1771.3926	mDa PPM 4.6 2.6 -6.6 -3.7	DBE i-F 38.5 53 38.5 53	FIT Norm 33.3 0.427 33.9 1.057	Conf(%) 65.26 34.74	Formula C76 H67 C75 H67	N6 O25 F15 Na N8 O24 F15 Na			
1006 Zogota RZE-42										



[¹H-NMR, 600 MHz, CD₃OD]



[¹³C-NMR, 150.9 MHz, CD₃OD]

OSPT2-RZE-43-prep.4.fid





[¹H, ¹³C HSQC, 600 MHz, 150.9 MHz, CD₃OD]



[¹H, ¹³C HMBC, 600 MHz, 150.9 MHz, CD₃OD]




[¹H, ¹H ROESY, 600 MHz, CD₃OD]

BZO-HO-NHTFA TFAHI NHTFA Cbz ÒBZ TFAHN OBz OBz hum ÓBZ NHTFA OSPT2-RZE-43-prep.13.ser 0 (1)0 00 . 1 0 - 2 Ð 69 .3 00 month f1 (ppm) 0 • 0 0 % **(3**) - 5 **WIN** ഞ 0 0 0 6 JUMMANJA .7 8 . 9 4.5 4.0 f2 (ppm) 9.0 7.5 7.0 6.5 6.0 5.5 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 8.5 8.0 5.0

5"-Deoxy-5"-(O-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-O-benzoyl-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (24)

[¹H, ¹H COSY, 600 MHz, CD₃OD]

BzO---HO--NHTFA -NHTFA Cbz ÒBz TFAHN AMMA MAMANAN OBz ЭВz OBZ NHTFA OSPT2-RZE-43-prep.15.ser 0 1 ŧ 2 . . 5 - 3 ana hala walk f1 (ppm) í ¥ I . 1 ^{1 2 1} - 5 ::: ŧ. 4 - 6 1 .7 . . 1 1 H ł \$ - 8 4 ŝ 3 - 9 10.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5 f2 (ppm)

5"-Deoxy-5"-(O-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-O-benzoyl-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (24)

[¹H, ¹H DQF-COSY, 600 MHz, CD₃OD]

Latvijas Organiskās sintēzes institūts

Fizikāli organiskās ķīmijas laboratorija

MS: Waters Synapt G2-S ESI+	i Capillary, kV: Cone, V:	0.7 40	LC: Ac	quity UP	PLC H-Cla	i ss Colu	umn:	Acquity UPLC BEF 2.1x50mm, 1.7μn	l C18 n
<i>Sample:</i> HRMS_2019_08_443 1 MS_POS_500-2000_RES_	.007 Zogota RZE- _7min ACN_Forr	43 n_5-98_040	_7min	1:D,3	10.000	000	MS_T	une Col#43	
Elemental Composition Report:									
Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3 Monoisotopic Mass, Even Electron Ions 300 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass) Elements Used: C: 0-83 H: 1-110 N: 1-10 O: 0-26 F: 15-15 Na: 1-1									
Mass RA (ale Mass m	D ₂ PPM	DBE	i-FIT	Norm	Conf(%)		Formula	
1875.4088 100.00 1	875.4076 1.	2 0.6	43.5	525.9	n/a	n/a		C83 H71 N6 O26	F15 Na
1007 Zogota RZE-43 HRMS_2019_08_443 1546 (4.322) Cm (1544:1564-(1504:1521+1595:1615)) 100 100									



[¹H-NMR, 600 MHz, CD₃OD]



[¹³C-NMR, 150.9 MHz, CD₃OD]

OSPT2-RZE-44-prep.4.fid





[¹H, ¹³C HSQC, 600 MHz, 150.9 MHz, CD₃OD]

4',5"-Dideoxy-5"-(*O*-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-*O*-benzoyl-4'-iodo-1,3,2',2"',6"'-penta-*N*-trifluoroacetyl 4'-*epi*-paromomycin (25) [¹H, ¹³C HMBC, 600 MHz, 150.9 MHz, CD₃OD]





[¹H, ¹H ROESY, 600 MHz, CD₃OD]



[¹H, ¹H COSY, 600 MHz, CD₃OD]



[¹H, ¹H DQF-COSY, 600 MHz, CD₃OD]

Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Synapt G2-Si	Capillary, kV:	0.7	LC: Acquity UPLC H-Class	Column:	Acquity UPLC BEH C18
ESI+	Cone, V:	40			2.1x50mm, 1.7μm

Sample:

HRMS_2019_08_445 1008 Zogota RZE-44 MS_POS_500-2000_RES_7min ACN_Form_5-98_040_7min 1:D,4 10.000000 MS_Tune Col#43

Elemental Composition Report:

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions 294 formula(e) evaluated with 2 results within limits (all results (up to 1000) for each mass) Elements Used: C: 0-83 H: 1-110 N: 1-10 O: 0-25 F: 15-15 Na: 1-1 I: 1-1

Mass	RA	Calc. Mass	mDa	PPM	DBE	i-FIT	Norm	Conf(%)	Formula
1985.3112	100.00	1985.3093	1.9	1.0	43.5	398.8	0.082	92.11	C83 H70 N6 O25 F15 Na I
		1985.3206	-9.4	-4.7	43.5	401.2	2.539	7.89	C82 H70 N8 O24 F15 Na I

1008 Zogota RZE-44



[¹H-NMR, 400 MHz, CD₃OD]



[¹³C-NMR, 100.6 MHz, CD₃OD]

OSPT2-DIMI-1134.11.fid





[¹H, ¹³C HSQC, 400 MHz, 100.6 MHz, CD₃OD]

OB; BzO-CF₃OCHN NHCOCF₃ NHCOCF3 Cbz OBz CF3COHN OBz ÓB₂ 1 An n.M.M.M. . hund OBz NHCOCF3 OSPT2-DIMI-1134.12.ser 0 1 20 . . 3 . 000 0 0000 00 DÓ 80 0 40 . I. e- Ø:d 222 60 - 60 ١, 0 8000 80 0 0 0 ' 0 - 80 00 f1 (ppm) . . 0 0 0 0 100 . . Ó . 0 00 00 0 120 . • @@@@@@ ; 0 0 20 0 ٥ 140 ٥ Ð ٥ .0 0 0 160 9 00 0° 00 ° **m** Ø Ø 0 \mathbf{x}_{i} 180

5.5

6.0

5.0 4.5 f2 (ppm)

.

6.5

٩,

7.0

7.5

8.0

4'-Allyl-4',5"-dideoxy-5"-(O-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-O-benzoyl-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (26)

[¹H, ¹³C HMBC, 400 MHz, 100.6 MHz, CD₃OD]

٥

9.0

8.5

 ~ 10

4.0

3.5

3.0

.

1.5

2.5

2.0

200

0.5

1.0

Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Synap ESI+	t G2-Si Capillar Cone, V	y, kV: 0.7 : 40	LC: Acc	quity UP	LC H-Clas	s Column:	Acquity UPLC BEH C18 2.1x50mm, 1.7μm	
<i>Sample:</i> HRMS_2019_08_44 MS_POS_500-2000	7 1009 Zogota _RES_7min ACN	a RZE-47 N_Form_5-98_04	10_7min	1:D,5	10.0000	000 MS_T	ūne Col#43	
Elemental Composi	tion Report:							
Tolerance = 5.0 PPM / Element prediction: Off Number of isotope peak	DBE: $\min = -1.5$, s used for i-FIT = 3	max = 50.0						
Monoisotopic Mass, Even Electron Ions 279 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass) Elements Used: C: 0-86 H: 1-110 N: 1-10 O: 0-25 F: 15-15 Na: 1-1								
Mass RA 1899.4436 100	Calc. Mass .00 1899.4440	mDa PPN -0.4 -0.2	1 DBE 44.5	i-FIT 484.0	Norm n/a	Conf(%) n/a	Formula C86 H75 N6 O25 F15 Na	

1009 Zogota RZE-47



[¹H-NMR, 400 MHz, CD₃OD]



[¹³C-NMR, 100.6 MHz, CD₃OD]

OSPT2-RZE-115.22.fid





[¹H, ¹³C HSQC, 400 MHz, 100.6 MHz, CD₃OD]



[¹H, ¹H COSY, 400 MHz, CD₃OD]

f1 (ppm)

Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Synapt G2-Si	Capillary, kV:	0.7	LC: Acquity UPLC H-Class	Column:	Acquity UPLC BEH C18
ESI+	Cone, V:	40			2.1x50mm, 1.7µm

Sample:

HRMS_2020_05_212 2374 Zogota RZE-115 MS_POS_2300_RES_7min ACN_Form_5-98_040_7min 1:D,8 5.000000 MS_Tune Col#66

Elemental Composition Report:

Single Mass Analysis Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 5

Monoisotopic Mass, Even Electron Ions 1127 formula(e) evaluated with 4 results within limits (up to 5 closest results for each mass) Elements Used: C: 0-100 H: 0-100 N: 0-10 O: 0-25 F: 15-15 Na: 1-1

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	Norm	Conf(%)	Formula
1857.4004	1857.4011	-0.7	-0.4	48.5	947.8	2.200	11.08	C88 H69 N4 O23 F15 Na
	1857.3984	2.0	1.1	49.5	946.6	1.000	36.80	C84 H65 N10 O21 F15 Na
	1857.3971	3.3	1.8	44.5	946.9	1.290	27.52	C83 H69 N6 O25 F15 Na
	1857.4083	-7.9	-4.3	44.5	947.0	1.403	24.59	C82 H69 N8 O24 F15 Na

2374 Zogota RZE-115



[¹H-NMR, 600 MHz, CD₃OD]



[¹³C-NMR, 150.9 MHz, CD₃OD]

OSPT2-RZE-107.4.fid



OB; ΗΟ BzO----CF₃OCH NHCOCF3 NHCOCF3 Cbz. ÒBz CF3COHN OBz 0B2 . M. WAMAL Luh. OBz NHCOCF3 OSPT2-RZE-107.2.ser •0 0 0. - 0 - 10 - 20 - 30 OX. • 28 -40 -0 🔘 0 - 50 - 60 • 0 JUNIT f1 (ppm) • - 70 - 80 1011 - 90 000 6 0 100 110 120 0 even (1990) - 130 -. . . 140 150 0 0 0 0 10.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 f2 (ppm)

5"-Deoxy-5"-(O-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-O-benzoyl-1,3,2',2"',6"'-penta-N-trifluoroacetyl 4'-epi-paromomycin (28)

[¹H, ¹³C HSQC, 600 MHz, 150.9 MHz, CD₃OD]



[¹H, ¹³C HMBC, 600 MHz, 150.9 MHz, CD₃OD]

[¹H, ¹H ROESY, 600 MHz, CD₃OD]







[¹H, ¹H COSY, 600 MHz, CD₃OD]

f1 (ppm)

OBz ΗΟ BzC CF NHCOCF3 -NHCOCF3 Cbz ÒBz CF3COHN QBz OBz NHCOCF3 OSPT2-RZE-107.14.ser 8 P 0 . - 1 -- 2 5 - 3 MANULUM 81 f1 (ppm) *! -- 5 - 6 **LAUNIMUL** . 7 . - 8 - 9 - 10 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 f2 (ppm)

5"-Deoxy-5"-(O-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-O-benzoyl-1,3,2',2"',6"'-penta-N-trifluoroacetyl 4'-epi-paromomycin (28)

[¹H, ¹H DQF-COSY, 600 MHz, CD₃OD]

Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Synapt G2-Si	Capillary, kV:	0.7	LC: Acquity UPLC H-Class	Column:	Acquity UPLC BEH C18
ESI+	Cone, V:	40			2.1x50mm, 1.7μm

Sample:

HRMS_2020_04_322 2182 Zogota RZE-107

MS_POS_500-2300_RES_7min ACN_Form_5-98_040_7min 2:A,1 1.000000 MS_Tune Col#66

Elemental Composition Report:

Single Mass Analysis Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 5

Monoisotopic Mass, Even Electron Ions 1508 formula(e) evaluated with 10 results within limits (up to 5 closest results for each mass) Elements Used: C: 0-100 H: 0-100 N: 0-10 O: 0-30 Na: 1-1 F: 15-15

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	Norm	Conf(%)	Formula
1875.4097	1875.4103	-0.6	-0.3	42.5	771.1	2.217	10.90	C87 H75 O28 Na F15
	1875.4090	0.7	0.4	48.5	770.1	1.261	28.35	C84 H67 N10 O22 Na F15
	1875.4116	-1.9	-1.0	47.5	771.2	2.265	10.38	C88 H71 N4 O24 Na F15
	1875.4076	2.1	1.1	43.5	770.2	1.343	26.10	C83 H71 N6 O26 Na F15
	1875.4063	3.4	1.8	38.5	770.3	1.416	24.27	C82 H75 N2 O30 Na F15





[¹H-NMR, 600 MHz, CD₃OD]







[¹H, ¹³C HSQC, 600 MHz, 150.9 MHz, CD₃OD]

OBz NHCOCF3 NHCOCF3 Cbz OBz CF₃COHN OBz mm ÒВz WW OBz NHCOCF3 OSPT2-RZE-108.3.ser 0 20 • 1 . 40 . Ó0 0 - 60 • . 0 IL LI 3 8 38 - 80 f1 (ppm) 00 -. . 0 100 ò . 120 ٠ ° (****** 140 0 160 b (g o ' 4. . 0 6⁰ 0 0 180 200 11 10 -1

8

9

7

6

4',5"-Dideoxy-5"-(O-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-O-benzoyl-4'-iodo-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (29)

[¹H, ¹³C HMBC, 600 MHz, 150.9 MHz, CD₃OD]

f2 (ppm)

4

3

2

1

Ö

5



[¹H, ¹H ROESY, 600 MHz, CD₃OD]



[¹H, ¹H COSY, 600 MHz, CD₃OD]
OBz NHCOCF3 -NHCOCF3 Cbz ЪBz CF3COHN OBz ÒBz ÓBz NHCOCF3 OSPT2-RZE-108.14.ser 1 - 0 -0-- 1 4 ... £ - 2 - 3 JANNAN 4. f1 (ppm) 4 -. - 5 . - 6 . - 7 WWW 5 - 8 - 9 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5 f2 (ppm)

4',5"-Dideoxy-5"-(O-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-O-benzoyl-4'-iodo-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (29)

[¹H, ¹H DQF-COSY, 600 MHz, CD₃OD]

Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Synapt G2-Si	Capillary, kV:	0.7	LC: Acquity UPLC H-Class	Column:	Acquity UPLC BEH C18
ESI+	Cone, V:	40			2.1x50mm, 1.7μm

Sample:

HRMS_2020_04_324 2183 Zogota RZE-108

MS_POS_500-2300_RES_7min_ACN_Form_5-98_040_7min_2:A,2_1.000000 MS_Tune Col#66

Elemental Composition Report:

Single Mass Analysis Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 5

Monoisotopic Mass, Even Electron Ions 1619 formula(e) evaluated with 11 results within limits (up to 5 closest results for each mass) Elements Used: C: 0-100 H: 0-100 N: 0-10 O: 0-30 F: 15-15 Na: 1-1 I: 1-1

Mass	Calc Mass	mDa	PPM	DBE	i-FIT	Norm	Conf(%)	Formula
1085 2000	1025 2002	0.6	0.2	12 5	0027	1 1 4 5	21.94	C82 H70 N6 O25 E15 No I
1905.5099	1985.5095	0.0	0.5	45.5	992.1	1.145	51.04	Cos H/0 NO 025 F15 Na1
	1985.3107	-0.8	-0.4	48.5	992.8	1.231	29.19	C84 H66 N10 O21 F15 Na I
	1985.3080	1.9	1.0	38.5	992.8	1.161	31.33	C82 H74 N2 O29 F15 Na I
	1985.3120	-2.1	-1.1	42.5	994.7	3.093	4.54	C87 H74 O27 F15 Na I
	1985.3134	-3.5	-1.8	47.5	995.1	3.472	3.11	C88 H70 N4 O23 F15 Na I





[¹H-NMR, 400 MHz, CD₃CN]



[¹³C-NMR, 100.6 MHz, CD₃CN]





[¹H, ¹³C HSQC, 400 MHz, 100.6 MHz, CD₃CN]



[¹H, ¹³C HMBC, 400 MHz, 100.6 MHz, CD₃CN]

Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Synapt G2-Si	Capillary, kV:	0.7	LC: Acquity UPLC H-Class	Column:	Acquity UPLC BEH C18
ESI+	Cone, V:	40			2.1x50mm, 1.7μm

Sample:

Elemental Composition Report:

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 5

Monoisotopic Mass, Even Electron Ions 3951 formula(e) evaluated with 15 results within limits (up to 5 closest results for each mass) Elements Used: C: 0-100 H: 0-100 N: 0-25 O: 0-25 Na: 1-1 F: 15-15

Mass	RA	Calc. Mass	mDa	PPM	DBE	i-FIT	Norm	Conf(%)	Formula
1859.4147	100.00	1859.4145	0.2	0.1	41.5	852.6	0.258	77.26	C69 H63 N22 O22 Na F15
		1859.4140	0.7	0.4	48.5	857.2	4.817	0.81	C84 H67 N10 O21 Na F15
		1859.4167	-2.0	-1.1	47.5	857.3	4.952	0.71	C88 H71 N4 O23 Na F15
		1859.4127	2.0	1.1	43.5	857.1	4.743	0.87	C83 H71 N6 O25 Na F15
		1859.4172	-2.5	-1.3	40.5	854.0	1.592	20.35	C73 H67 N16 O24 Na F15

484 Lubriks DLP-PR22-H



[¹H-NMR, 400 MHz, CD₃OD]



[¹³C-NMR, 100.6 MHz, CD₃OD]

OSPT2-DIMI-1168.11.fid OBz BzO CE NHCOCF3 **OCHN** NHCOCF3 ÒBz H_2N CF3COHN OBz 0 ÒΒz OBZ NHCOCF3



Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

1200 1300 1400

1500

0 |---500

600

700

800

900

1000 1100

MS: Waters Syn ESI+	napt G2-Si	Capillary, kV: Cone, V:	0.7 40		LC: Ac	quity UP	PLC H-Cla	ss Co	olumn:	Acquity U 2.1x50mn	PLC BEH C18 n, 1.7μm
Sample: HRMS_2019_08 MS_POS_500-20	_449 10 000_RES_7	10 Zogota RZE-4 'min ACN_Forn	48 n_5-98	_040_	_7min	1:D,6	10.000	000	MS_T	ine (Col#43
Elemental Comp	position Re	eport:									
Tolerance = 5.0 PPN Element prediction: Number of isotope p	M / DBE:1 Off peaks used fo	min = -1.5, max = 5 or i-FIT = 3	50.0								
Monoisotopic Mass, 276 formula(e) evalu Elements Used: C: 0-78 H: 1-110	, Even Electr luated with 2 N: 1-10 C	on Ions results within limit D: 0-23 F: 15-15	ts (all res	sults (up	o to 1000)) for each 1	mass)				
Mass 1	RA Ca	lc. Mass mi	Da F	PPM	DBE	i-FIT	Norm	Conf(%	6)	Formula	N6 022 E15
1/43.4430	100.00 17-	45.4522 -8	.4 -	4.8	38.5	271.4	2.087	12.41		C77 H72	N8 O22 F15
1010 Zogota RZ HRMS_2019_08_4	ZE-48 449 1240 (3	.474)							1	745.4438 1746.44	1: TOF MS E 2.87 64
- - -										1747.44	98

1700

1600

1767.4252

1800

1769.4312

1900

CF₃COHN

---- m/z

OBz

0-

ÓBz NHCOCF3

OBz

O

2,2,2-Trifluoro-*N*-(2-hydroxyethyl)acetamide (S1)



2,2,2-Trifluoro-*N*-(2-oxoethyl)acetamide (32)

[¹H-NMR, 400 MHz, CDCl₃]

OSPT2-DIMI-1150pr.10.fid



2,2,2-Trifluoro-N-(3-oxopropyl)acetamide (33)

[¹H-NMR, 400 MHz, CDCl₃]



4',5"-Dideoxy-4'-propyl-5"-(2-aminoethylamino)-1,2-ethanediamine)-6,3',6',2",3"',4"'-hexa-*O*-benzoyl-hepta-*N*-trifluoroacetyl paromomycin (36)

[¹H-NMR, 600 MHz, CD₃OD]



4',5"-Dideoxy-4'-propyl-5"-(2-aminoethylamino)-1,2-ethanediamine)-6,3',6',2",3"',4"'-hexa-O-benzoyl-hepta-N-trifluoroacetyl paromomycin (36)

[¹³C-NMR, 150.9 MHz, CD₃OD]

OSPT2-RZE-53.4.fid



4',5"-Dideoxy-4'-propyl-5"-(2-aminoethylamino)-1,2-ethanediamine)-6,3',6',2",3"',4"'-hexa-O-benzoyl-hepta-N-trifluoroacetyl paromomycin (36)

Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Synapt G2-Si	Capillary, kV:	0.7	LC: Acquity UPLC H-Class	Column:	Acquity UPLC BEH C18
ESI+	Cone, V:	40			2.1x50mm, 1.7µm

Sample:

HRMS_2019_08_460 1011 Zogota RZE-53 MS_POS_500-2300_RES_7min ACN_Form_5-98_040_7min 1:F,1 10.000000 MS_Tune Col#43

Elemental Composition Report:

Tolerance = 15.0 PPM / DBE: min = -1.5, max = 50.0Element prediction: Off Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions 264 formula(e) evaluated with 2 results within limits (all results (up to 1000) for each mass) Elements Used: C: 0-84 H: 1-110 N: 1-10 O: 0-25 F: 21-21 Na: 1-1

Mass	RA	Calc. Mass	mDa	PPM	DBE	i-FIT	Norm	Conf(%)	Formula
2002.4412	100.00	2002.4409	0.3	0.1	40.5	486.5	0.273	76.11	C83 H74 N9 O24 F21 Na
		2002.4297	11.5	5.7	40.5	487.6	1.432	23.89	C84 H74 N7 O25 F21 Na

1011 Zogota RZE-53



4',5"-Dideoxy-4'-propyl-5"-(3-aminoethylamino)-1,3-diaminopropane)-6,3',6',2",3"',4"'-hexa-O-benzoyl-hepta-N-trifluoroacetyl paromomycin (37)

[¹H-NMR, 600 MHz, CDCl₃]



4',5"-Dideoxy-4'-propyl-5"-(3-aminoethylamino)-1,3-diaminopropane)-6,3',6',2",3"',4"'-hexa-O-benzoyl-hepta-N-trifluoroacetyl paromomycin (37)

[¹³C-NMR, 150.9 MHz, CDCl₃]

OSPT2-RZE-55.4.fid



4',5"-Dideoxy-4'-propyl-5"-(3-aminoethylamino)-1,3-diaminopropane)-6,3',6',2",3",4"'-hexa-O-benzoyl-hepta-N-trifluoroacetyl paromomycin (37)

Latvijas Organiskās sintēzes institūts Fizikāli organiskās kīmijas laboratorija

MS: Waters Synapt G2-Si	Capillary, kV:	0.7	LC: Acquity UPLC H-Class	Column:	Acquity UPLC BEH C18
ESI+	Cone, V:	40			2.1x50mm, 1.7μm

Sample:

HRMS_2020_02_160 1864 Zogota RZE-55 MS_POS_500-2300_RES_7min ACN_Form_5-98_040_7min 2:C,7 5.000000 MS_Tune Col#43

Elemental Composition Report:

Multiple Mass Analysis: 2 mass(es) processed Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions 17545 formula(e) evaluated with 8 results within limits (up to 3 best isotopic matches for each mass) Elements Used: C: 0-90 H: 0-80 N: 0-7 O: 0-25 F: 0-21 Na: 0-1

Mass	RA	Calc. Mass	mDa	PPM	DBE	i-FIT	Norm	Conf(%)	Formula
2016.4429	100.00	2016.4453	-2.4	-1.2	40.5	586.5	0.376	68.65	C85 H76 N7 O25 F21 Na
		2016.4442	-1.3	-0.6	44.5	587.7	1.559	21.03	C88 H75 N7 O24 F20 Na
		2016.4477	-4.8	-2.4	43.5	588.4	2.271	10.32	C87 H75 N7 O25 F21
2017.4457	93.39	2017.4446	1.1	0.5	44.5	607.3	0.103	90.22	C89 H75 N6 O23 F21 Na
		2017.4533	-7.6	-3.8	43.5	609.5	2.325	9.78	C90 H78 N4 O25 F20 Na

1864 Zogota RZE-55



[¹H-NMR, 600 MHz, CDCl₃]



[¹³C-NMR, 150.9 MHz, CDCl₃]



OBz BzO-NHTFA -NHTFA Cbz. OBz M TFAHN OBz ÒΒz OSPT2-DIMI-1179.2.ser ÓBz NHTFA -0 60 0 - 10 000 - 20 0 - 30 00 00 0 00 -40 0 - 50 0 00 - 60 00 f1 (ppm) 60 0 A. - 70 باللمب - 80 0 0 - 90 0 - 100 -110 00-0-00-0 -120 - 130 - 140 9.0 8.5 4.5 4.0 3.5 3.0 2.5 2.0 0.5 0.0 8.0 7.5 7.0 6.5 6.0 5.5 5.0 1.5 1.0

4',5"-Dideoxy-4'-propyl-5"-(O-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-O-benzoyl-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (38)

[¹H, ¹³C HSQC, 600 MHz, 150.9 MHz, CDCl₃]

4',5"-Dideoxy-4'-propyl-5"-(*O*-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-*O*-benzoyl-1,3,2',2"',6"'-penta-*N*-trifluoroacetyl paromomycin (38) [¹H, ¹³C HMBC, 600 MHz, 150.9 MHz, CDCl₃]



[¹H, ¹H ROESY, 600 MHz, CDCl₃] OBz BzO-NHTFA TFAHN NHTFA Cbz OBz TFAHN OBz AMA ANA MI ÒB: ÓBZ NHTFA OSPT2-DIMI-1179.5.ser . 1 595 *"* - 2 . - 3 o 0⁰ 0⁰ ° 0 f1 (ppm) 0 - 5 . ° % 3 0 - 6 . 7 MMMM - 8 - 9 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0

4',5"-Dideoxy-4'-propyl-5"-(O-benzylcarbamoyl)-6,3',6',2",3"',4"'-hexa-O-benzoyl-1,3,2',2"',6"'-penta-N-trifluoroacetyl paromomycin (38)

Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Synapt G2-Si	Capillary, kV:	0.7	LC: Acquity UPLC H-Class	Column:	Acquity UPLC BEH C18
ESI+	Cone, V:	40			2.1x50mm, 1.7μm

Sample:

HRMS_2020_10_384 3166 Lubriks DIMI-1388 MS_POS_500-2000_RES_7min_ACN_Form_5-98_040_7min Col#66 1:E,6 1.000000 MS_Tune

Elemental Composition Report:

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0Element prediction: Off Number of isotope peaks used for i-FIT = 7

Monoisotopic Mass, Even Electron Ions 2256 formula(e) evaluated with 15 results within limits (up to 5 closest results for each mass) Elements Used: C: 0-100 H: 0-100 N: 0-15 O: 0-30 F: 15-15 Na: 1-1

Mass	RA	Calc. Mass	mDa	PPM	DBE	i-FIT	Norm	Conf(%)	Formula
1901.4022	100.00	1901.4628	-0.1	-0.1	35.5	1618.1	0.031	96.92	C75 H77 N12 O28 F15 Na
		1901.4610	1.2	0.6	48.5	1622.4	4.359	1.28	C87 H73 N10 O21 F15 Na
		1901.4637	-1.5	-0.8	47.5	1624.3	6.260	0.19	C91 H77 N4 O23 F15 Na
		1901.4597	2.5	1.3	43.5	1622.3	4.274	1.39	C86 H77 N6 O25 F15 Na





[¹H-NMR, 600 MHz, CDCl₃]



[¹³C-NMR, 150.9 MHz, CDCl₃]





[¹H, ¹³C HSQC, 600 MHz, 150.9 MHz, CDCl₃]



[¹H, ¹³C HMBC, 600 MHz, 150.9 MHz, CDCl₃]



Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Synapt G2-Si ESI+	Capillary, kV: Cone, V:	0.7 40	LC: Acquity UP	LC H-Class	Column:	Acquity UPLC BEH C18 2.1x50mm, 1.7μm
<i>Sample:</i> HRMS_2020_10_386 31 MS_POS_500-2000_RES_7	67 Lubriks DIMI-: min ACN_Form_	1389 _5-98_040	_7min 1:E,7	1.000000	MS_T	une Col#66
Elemental Composition	Report:					
Tolerance = 5.0 PPM / DBE: r Element prediction: Off Number of isotope peaks used fo	nin = -1.5, max = 50 r i-FIT = 7	.0				
Monoisotopic Mass, Even Electr 3130 formula(e) evaluated with 2 Elements Used: C: 0-100 H: 0-100 N: 0-15	on Ions 21 results within limi O: 0-30 F: 15-15	ts (up to 5 cl	osest results for each	mass)		

Mass	RA	Calc. Mass	mDa	PPM	DBE	i-FIT	Norm	Conf(%)	Formula
1773.4353	100.00	1773.4350	0.3	0.2	27.5	1282.4	0.317	72.80	C63 H72 N14 O29 F15
		1773.4358	-0.5	-0.3	39.5	1285.1	2.957	5.20	C79 H72 N6 O24 F15
		1773.4345	0.8	0.5	34.5	1284.9	2.823	5.94	C78 H76 N2 O28 F15
		1773.4372	-1.9	-1.1	44.5	1285.2	3.107	4.48	C80 H68 N10 O20 F15
		1773.4332	2.1	1.2	40.5	1284.3	2.155	11.59	C75 H68 N12 O22 F15

3167 Lubriks DIMI-1389



[¹H-NMR, 600 MHz, CDCl₃]



[¹³C-NMR, 150.9 MHz, CDCl₃]





[¹H, ¹³C HSQC, 600 MHz, 150.9 MHz, CDCl₃]



[¹H, ¹³C HMBC, 600 MHz, 150.9 MHz, CDCl₃]


Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Synapt G2-Si ESI+	Capillary, kV: Cone, V:	0.7 40	LC: Acquity UPLC H-Class	Column:	Acquity UPLC BEH C18 2.1x50mm, 1.7μm

Sample:

HRMS_2020_10_382 3165 Lubriks DIMI-1387 MS_POS_500-2000_RES_7min ACN_Form_5-98_040_7min 1:E,5 1.000000 MS_Tune Col#66

Elemental Composition Report:

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0Element prediction: Off Number of isotope peaks used for i-FIT = 7

Monoisotopic Mass, Even Electron Ions 2255 formula(e) evaluated with 17 results within limits (up to 5 closest results for each mass) Elements Used: C: 0-100 H: 0-100 N: 0-15 O: 0-30 F: 15-15

Mass	RA	Calc. Mass	mDa	PPM	DBE	i-FIT	Norm	Conf(%)	Formula
1878.4929	100.00	1878.4929	0.0	0.0	31.5	1339.8	0.151	85.98	C70 H79 N15 O29 F15
		1878.4924	0.5	0.3	38.5	1344.1	4.476	1.14	C85 H83 N3 O28 F15
		1878.4937	-0.8	-0.4	43.5	1344.1	4.452	1.17	C86 H79 N7 O24 F15
		1878.4910	1.9	1.0	44.5	1341.8	2.244	10.60	C82 H75 N13 O22 F15
		1878.4950	-2.1	-1.1	48.5	1344.1	4.496	1.12	C87 H75 N11 O20 F15

3165 Lubriks DIMI-1387



[¹H-NMR, 400 MHz, CD₃OD]



[¹³C-NMR, 100.6 MHz, CD₃OD]





[¹H, ¹³C HSQC, 400 MHz, 100.6 MHz, CD₃OD]



[¹H, ¹³C HMBC, 400 MHz, 100.6 MHz, CD₃OD]

Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Synapt G2-Si ESI+	Capillary, kV: Cone, V:	0.7 40	LC: Acquity UP	LC H-Class	Column: Acquint 2.1x	uity UPLC BEH C18 50mm, 1.7μm			
Sample: HRMS_2021_05_164 902 MS_POS_500-2000_RES_7	2 Lubriks DIMI-11 min ACN_Form_	53-atk 5-98_040_	_7min 1:B,1	1.000000	MS_Tune	Col#66			
Elemental Composition Report:									
Tolerance = 5.0 PPM / DBE: m Element prediction: Off Number of isotope peaks used for	nin = -1.5, max = 50.0 r i-FIT = 5	0							
	-								

Monoisotopic Mass, Even Electron Ions 2004 formula(e) evaluated with 12 results within limits (up to 5 closest results for each mass) Elements Used: C: 0-100 H: 0-110 N: 0-15 O: 0-25 F: 18-18 Na: 1-1

Mass	RA	Calc. Mass	mDa	PPM	DBE	i-FIT	Norm	Conf(%)	Formula
1920.4257	100.00	1920.4266	-0.9	-0.5	40.5	990.4	1.666	18.91	C82 H73 N7 O25 F18 Na
		1920.4240	1.7	0.9	41.5	989.4	0.625	53.52	C78 H69 N13 O23 F18 Na
		1920.4235	2.2	1.1	48.5	992.3	3.541	2.90	C93 H73 N O22 F18 Na
		1920.4280	-2.3	-1.2	45.5	990.4	1.693	18.40	C83 H69 N11 O21 F18 Na
		1920.4208	4.9	2.6	49.5	991.5	2.769	6.27	C89 H69 N7 O20 F18 Na

902 Lubriks DIMI-1153-atk



5"-Amino-4',5"-dideoxy-4'-propyl paromomycin pentaacetate (8)

[¹H-NMR, 600 MHz, D₂O]





5"-Amino-4',5"-dideoxy-4'-propyl paromomycin pentaacetate (8)

5"-Amino-4',5"-dideoxy-4'-propyl paromomycin pentaacetate (8)



[¹H, ¹³C HSQC, 600 MHz, 150.9 MHz, D₂O]

5"-Amino-4',5"-dideoxy-4'-propyl paromomycin pentaacetate (8)

Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Sy ESI+	ynapt G2	2-Si Capillary Cone, V:	, kV: 0	0.7 40	LC: Ac	quity UF	PLC H-Cla	ss Column:	Acquity UPLC BEH C18 2.1x50mm, 1.7μm
<i>Sample:</i> HRMS_2019_0 MS_POS_RES_	06_031 4min	787 Lubriks I ACN_Form_5	DIMI-118 5-98_040	0 _4min	1:C,5	5.0000	000	MS_Tune	Col#43
Elemental Co	mpositi	on Report:							
Tolerance = 5.0 PI Element prediction Number of isotope	PM / DF n: Off e peaks use	3E: min = -1.5, n ed for i-FIT = 3	max = 50.0						
Monoisotopic Mas 1165 formula(e) er Elements Used: C: 1-100 H: 1-11	ss, Even E valuated w 10 N: 0-1	lectron Ions vith 7 results with 10 O: 0-15	nin limits (u	ip to 3 clos	sest results	for each	mass)		
Mass 641.3732	RA 100.00	Calc. Mass 641.3721 641.3735 641.3743	mDa 1.1 -0.3 -1.1	PPM 1.7 -0.5 -1.7	DBE 3.5 8.5 20.5	i-FIT 143.3 152.5 162.8	Norm 0.000 9.206 19.429	Conf(%) 99.99 0.01 0.00	Formula C26 H53 N6 O12 C27 H49 N10 O8 C43 H49 N2 O3
787 Lubriks D HRMS_2019_06	D IMI-118 6_031 414	<mark>0</mark> 4 (1.195) Cm (3	99:436-(5	52:590+2	93:325))	454.25	525		1: TOF MS ES+ 8.53e6





[¹H-NMR, 600 MHz, D₂O]



[¹³C-NMR, 150.9 MHz, D₂O]

OSPT2-RZE-77-salt.4.fid



[¹H, ¹³C HSQC, 600 MHz, 150.9 MHz, D₂O]



[¹H, ¹³C HMBC, 600 MHz, 150.9 MHz, D₂O]



Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Syna ESI+	apt G2·	- Si Capillary, Cone, V:	, kV:	0.7 40	LC: Ac	quity UP	LC H-Cla	ass (Column:	Acquity UPLC BEH C18 2.1x50mm, 1.7μm
<i>Sample:</i> HRMS_2020_02_ MS_POS_RES_7m	_181 nin	1867 Zogota ACN_Form_5	RZE-77 5-98_04	0_7min	2:D,7	10.000	0000	MS_1	Гune	Col#43
Elemental Comp	positic	on Report:								
Tolerance = 5.0 PPM Element prediction: C Number of isotope pe	[/ DB] Off eaks used	E: min = -1.5, n 1 for i-FIT = 3	hax = 50.0)						
Monoisotopic Mass, 1 1114 formula(e) evan Elements Used: C: 0-50 H: 0-80 N	Even Ek uated wi N: 0-10	ectron Ions th 6 results with O: 0-15	in limits	(up to 3 best	isotopic	matches fo	r each ma	ass)		
Mass R 684.4145 14	CA 00.00	Calc. Mass 684.4143 684.4170 684.4138	mDa 0.2 -2.5 0.7	PPM 0.3 -3.7 1.0	DBE 3.5 2.5 21.5	i-FIT 623.3 628.4 630.1	Norm 0.007 5.148 6.870	Conf(99.32 0.58 0.10	%)	Formula C28 H58 N7 O12 C32 H62 N O14 C41 H50 N9 O
1867 Zogota RZ HRMS_2020_02_1 100-	E-77 181 138	(0.406) Cm (1	33:140-(74:83+233	:240))	684.4145				1: TOF MS ES+ 8.51e





[¹H-NMR, 600 MHz, D₂O]



[¹³C-NMR, 150.9 MHz, D₂O]

OSPT2-RZE-83-salt.4.fid



[¹H, ¹³C HSQC, 600 MHz, 150.9 MHz, D₂O]



[¹H, ¹³C HMBC, 600 MHz, 150.9 MHz, D₂O]



Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Synapt G ESI+	2-Si Capillary, kV Cone, V:	/: 0.7 40	LC: Acquity UF	PLC H-Class	Column:	Acquity UPLC BEH C18 2.1x50mm, 1.7μm
<i>Sample:</i> HRMS_2020_02_463 MS_POS_RES_4min	1967 Zogota RZ ACN_Form_5-98	E-83 8_040_4min	1:C,3 5.000	000 M:	S_Tune	Col#66
Elemental Composit	ion Report:					
Tolerance = 5.0 PPM / D Element prediction: Off Number of isotope peaks us	BE: min = -1.5, max = eed for i-FIT = 3	= 50.0				
Monoisotopic Mass, Even H 1515 formula(e) evaluated v Elements Used: C: 0-50 H: 0-100 N: 0-J	Electron Ions with 5 results within 1 0 O: 0-20	imits (all results (up to 1000) for each	ı mass)		
Mass RA 698 4333 100 00	Calc. Mass 698 4300	mDa PPM	DBE i-FIT	Norm Co 0.475 62	onf(%) 20	Formula C29 H60 N7 O12
100.00	698.4327 698.4340	0.6 0.9 -0.7 -1.0	2.5 466.2 7.5 467.1	1.413 24 2.271 10	.35 .32	C33 H64 N O14 C34 H60 N5 O10
1967 Zogota RZE-83 HRMS_2020_02_463 89) (0.272) Cm (88:93	-(66:73+127:13) 351 2267	7))			1: TOF MS ES+ 8.35e6
100						0.0000
-						
~			54	1 2110		698.4333
161.0936 188.1	297 341.2	352.2291 2069 405.	51 1445454.2528	512.3143		699.4370 752.3517
0- h	250 300	350 400	0 450 50	0 550	600	650 700 750 m/z

[¹H-NMR, 600 MHz, D₂O]





S166



[¹H, ¹³C HSQC, 600 MHz, 150.9 MHz, D₂O]

Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Synapt G2-Si	Capillary, kV:	0.7	LC: Acquity UPLC H-Class	Column:	Acquity UPLC BEH C18
ESI+	Cone, V:	40			2.1x50mm, 1.7μm

Sample:

 HRMS_2019_06_033
 788 Lubriks DIMI-1181

 MS_POS_RES_4min
 ACN_Form 5-98_040_4min
 1:C,6
 1.000000
 MS_Tune
 Col#43

Elemental Composition Report:

Tolerance = 3.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions 1224 formula(e) evaluated with 4 results within limits (all results (up to 1000) for each mass) Elements Used: C: 1-100 H: 1-110 N: 0-10 O: 0-15

Mass 669.3654	RA 100.00	Calc. Mass 669.3671	mDa -1.7	PPM -2.5	DBE 4.5	i-FIT 271.3	Norm 0.001	Conf(%) 99.93	Formula C27 H53 N6 O13
		669.3639	1.5	2.2	12.5	279.3	8.003	0.03	C38 H53 O10
		669.3652	0.2	0.3	17.5	279.7	8.324	0.02	C39 H49 N4 O6
		669.3665	-1.1	-1.6	22.5	280.1	8.744	0.02	C40 H45 N8 O2

788 Lubriks DIMI-1181



[¹H-NMR, 600 MHz, D₂O]







[¹H, ¹³C HSQC, 600 MHz, 150.9 MHz, D₂O]

Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters S ESI+	Synapt G2-Si	Capillary, kV: Cone, V:	0.7 40	LC: Acq	uity UP	LC H-Cla	ss Colur	nn: Acquity 2.1x50r	ν UPLC BEH C18 mm, 1.7μm	
<i>Sample:</i> HRMS_2019_ MS_POS_RES	06_017 78 _4min A0	36 Lubriks DIM CN_Form_5-98	l-1171 _040_4min	2:B,7	5.0000	00	MS_Tune	Col#43	3	
Elemental Composition Report:										
Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3										
Monoisotopic M 1240 formula(e) Elements Used: C: 1-100 H: 1-	ass, Even Elect evaluated with 110 N: 0-10	ron Ions 5 results within lin O: 0-15	mits (up to 3 c	losest results	for each r	nass)				
Mass	RA C	alc. Mass 1	nDa PPN	A DBE	i-FIT	Norm	Conf(%)	Formu	ıla 154 NZ O13	
004.3704	68 68	34.3761 (34.3774 -	0.3 0.4 -1.5	17.5 22.5	535.5 536.4	7.116 8.000	0.08 0.03	C39 H C40 H	154 N7 O15 150 N5 O6 146 N9 O2	
786 Lubriks HRMS_2019_ 100	DIMI-1171 _06_017 86 (i	0.264) Cm (84:	93-69:74)	49	97.2569				1: TOF MS E 3.0;	S+ 2e6
-										
~	188.12	77	342.6924				684	.3764		
- - - - - - - - - - - - - - - - - - -	170.1171	249.1323 337.17 334.179	17 343.193 92 349.6	7 998 00 450	498.2	592 1.2719	00 650	698.3918 706.358 722.3	0 326 800 850	n/z



4',5"-Dideoxy-5"-glycinamido-4'-propyl paromomycin hexacetate (13)

[¹H-NMR, 600 MHz, D₂O]









4',5"-Dideoxy-5"-glycinamido-4'-propyl paromomycin hexacetate (13)



[¹H, ¹³C HSQC, 600 MHz, 150.9 MHz, D₂O]

4',5"-Dideoxy-5"-glycinamido-4'-propyl paromomycin hexacetate (13)

Latvijas Organiskās sintēzes institūts Fizikāli organiskās ķīmijas laboratorija

MS: Waters Synapt G2-Si ESI+	Capillary, kV: Cone, V:	0.7 40	LC: Acquity UPLC H-Class	Column:	Acquity UPLC BEH C18 2.1x50mm, 1.7μm

Sample:

HRMS_2019_04_226 623 DIMI-1154 MS_POS_RES_4min ACN_Form_5-98_040_4min MS_Tune 1:E,6 0.300000 Col#43

Elemental Composition Report:

Tolerance = 2.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions 5251 formula(e) evaluated with 7 results within limits (all results (up to 1000) for each mass) Elements Used: C: 1-100 H: 1-110 N: 0-50 O: 0-50

Mass	RA	Calc. Mass	mDa	PPM	DBE	i-FIT	Norm	Conf(%)	Formula
698.3942	100.00	698.3936	0.6	0.9	4.5	745.2	0.194	82.40	C28 H56 N7 O13
		698.3949	-0.7	-1.0	9.5	747.1	2.128	11.91	C29 H52 N11 O9
		698.3936	0.6	0.9	15.5	747.9	2.918	5.40	C26 H44 N21 O3
		698.3954	-1.2	-1.7	2.5	751.0	6.036	0.24	C14 H48 N23 O10
		698.3931	1.1	1.6	22.5	752.7	7.779	0.04	C41 H48 N9 O2

623 DIMI-1154







Phenyl 2-azido-2,3,4-trideoxy-4-*C*-ethyl-1-thio-β-D-glucopyranoside (43)

Phenyl 2-azido-2,3,4-trideoxy-4-*C*-ethyl-1-thio-β-D-glucopyranoside (43)







Phenyl 6-*O*-acetyl-2-azido-2,3,4-trideoxy-4-*C*-ethyl-1-thio-β-D-glucopyranoside (44)

Phenyl 6-*O*-acetyl-2-azido-2,3,4-trideoxy-4-*C*-ethyl-1-thio-β-D-glucopyranoside (44)

[¹³C-NMR, 125 MHz, CDCl₃]




Phenyl 6-*O*-acetyl-2-azido-2,3,4-trideoxy-4-*C*-ethyl-1-thio-β-D-glucopyranoside (44)

Phenyl 6-*O*-acetyl-2-azido-2,3,4-trideoxy-4-*C*-ethyl-1-thio-β-D-glucopyranosyl sulfoxide (45)









Phenyl 6-O-acetyl-2-azido-2,3,4-trideoxy-4-C-ethyl-1-thio-β-D-glucopyranosyl sulfoxide (45)



1,3,2',3',4',2''',6'''-Heptadeoxy-4'-propyl-6'-acetyl-1,3,2',2''',6'''-pentaazido-6,2'',5'',3''',4'''-penta-O-benzyl paromomycin (47)



1,3,2',3',4',2''',6'''-Heptadeoxy-4'-propyl-6'-acetyl-1,3,2',2''',6'''-pentaazido-6,2'',5'',3''',4'''-penta-O-benzyl paromomycin (47)



1,3,2',3',4',2"',6"'-Heptadeoxy-4'-propyl-6'-acetyl-1,3,2',2"',6"'-pentaazido-6,2",5",3"',4"'-penta-O-benzyl paromomycin (47)



1,3,2',3',4',2''',6'''-Heptadeoxy-4'-propyl-6'-acetyl-1,3,2',2''',6'''-pentaazido-6,2'',5'',3''',4'''-penta-O-benzyl paromomycin (47)



1,3,2',3',4',2"',6"'-Heptadeoxy-4'-propyl-6'-acetyl-1,3,2',2"',6"'-pentaazido-6,2",5",3"',4"'-penta-O-benzyl paromomycin (47)



1,3,2',3',4',2"',6"'-Heptadeoxy-4'-propyl-6'-acetyl-1,3,2',2"',6"'-pentaazido-6,2",5",3"',4"'-penta-O-benzyl paromomycin (47)

OAc

C

Ó ÓBn

ÒBn



[¹³C-NMR, 151 MHz, D₂O]



[¹H, ¹H COSY, 600 MHz, CDCl₃]





[¹H, ¹³C HSQC, 600 MHz, 151 MHz, CDCl₃]





 H_2

ЮH

x 5 AcOH

4',6',5"-Trideoxy-6',5"-[bis-N-(2-hydroxyethyl)]-4'-propyl paromomycin heptaacetate (15)

[¹H-NMR, 600 MHz, D₂O]





4',6',5"-Trideoxy-6',5"-[bis-N-(2-hydroxyethyl)]-4'-propyl paromomycin heptaacetate (15)



4',6',5"-Trideoxy-6',5"-[bis-*N***-(2-hydroxyethyl)]-4'-propyl paromomycin heptaacetate (15)** [¹H, ¹H COSY, 600 MHz, CDCl₃]



4',6',5"-Trideoxy-6',5"-[bis-*N*-(2-hydroxyethyl)]-4'-propyl paromomycin heptaacetate (15)

[¹H, ¹³C HSQC, 600 MHz, 151 MHz, CDCl₃]



4',6',5"-Trideoxy-6',5"-[bis-N-(2-hydroxyethyl)]-4'-propyl paromomycin heptaacetate (15)

4',6',5"-Trideoxy-6',5"-[bis-N-(2-hydroxyethyl)]-4'-propyl paromomycin heptaacetate (15)

Elemental Composition Report

Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

4 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass) Elements Used: C: 29-31 H: 59-62 N: 5-8 O: 12-14 Na: 0-1 vas-diethanolamin-final

2017_0422_24 11 (0.213)

LCT Premier 1: TOF MS ES+ 1.27e+003

Page 1

100-		750.4203												
% 689.1597 707.3863		7.3863	728.4359 734.4423		4423	751.4251 766.3975		778.4514 792.45		42 806.4567		822.4	252 835.39	<u>98</u>
690	700	710	720	730	740 750	760	770	780	790	800	810	820	830	11/2
Minimum: Maximum:			5.0	5.0	-1.5 50.0	5								
Mass	Calc.	Mass	mDa	PPM	DBE	i-F	ΊT	i-FIT	(Norm)	Formul	a			
750.4203	750.4	225	-2.2	-2.	9 3.5	26.	2	0.0		C30 H	161 N7	013	Na	



S202