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Supplemental information

**Tribbles pseudokinase NIPI-3 regulates
intestinal immunity in *Caenorhabditis elegans*
by controlling SKN-1/Nrf activity**

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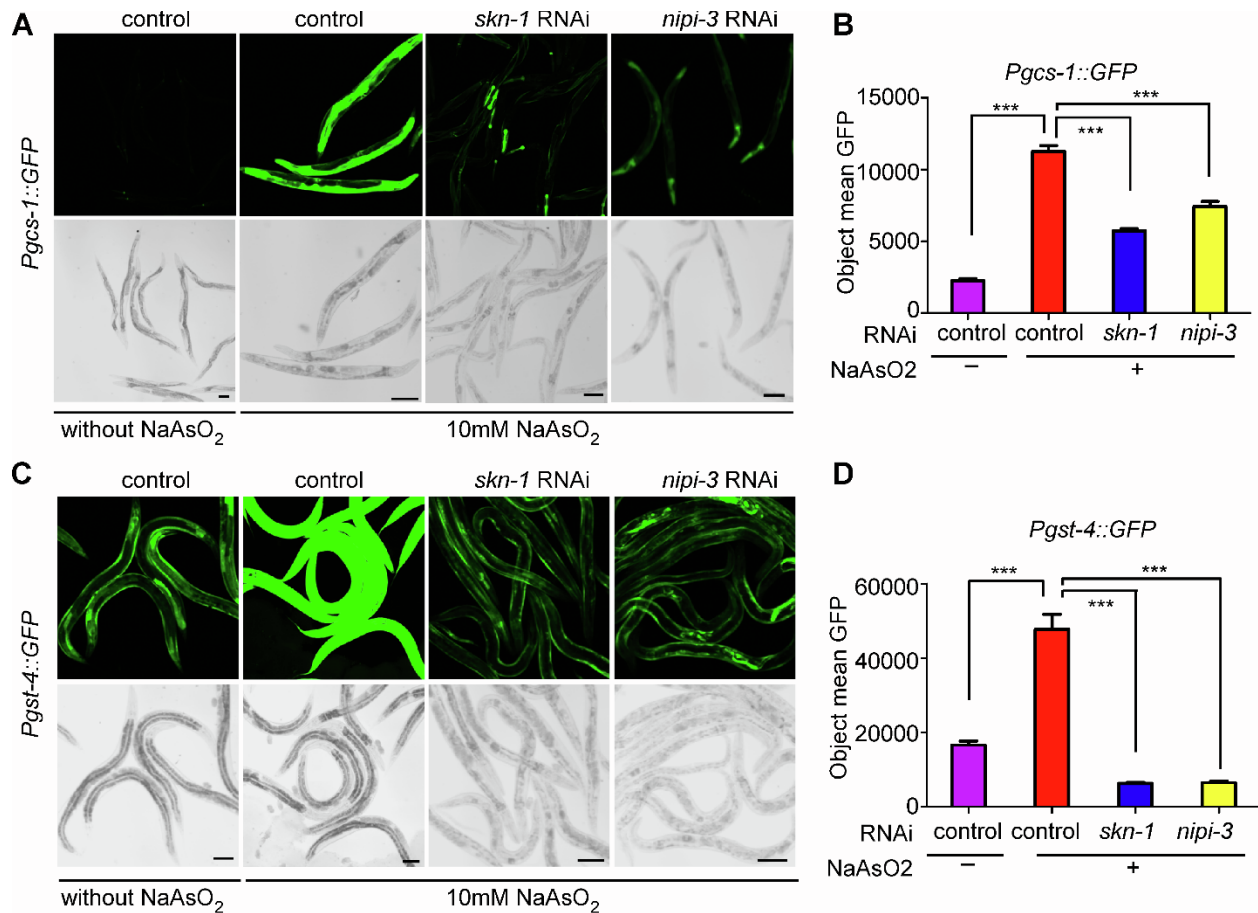


Figure S1: Loss of NIPI-3 reduces the expression of SKN-1 regulated genes in response to oxidant. Related to Figure 1. Fluorescence microscopy was used to analyze *Pgcs-1::gfp* (A) and *Pgst-4::gfp* (C) expression in wild type animals exposed to vector control, *skn-1* or *nipi-3* RNAi prior to a five hour exposure to 10 mM sodium arsenate in NGM plates (A) or M9 buffer (B). *Pgcs-1::gfp* and *Pgst-4::gfp* expression was less induced in the intestines of animals treated with *skn-1* and *nipi-3* RNAi as compared to vector control, and was comparable to the level of the control animals not exposed to oxidant. The scale bars are 100 μ m. The amount of GFP fluorescence produced by animals containing the *Pgcs-1::gfp* reporter (B) and the *Pgst-4::gfp* reporter (D) was quantified. The average gene expression of biological replicates is shown, and the error bars represent the SEM. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

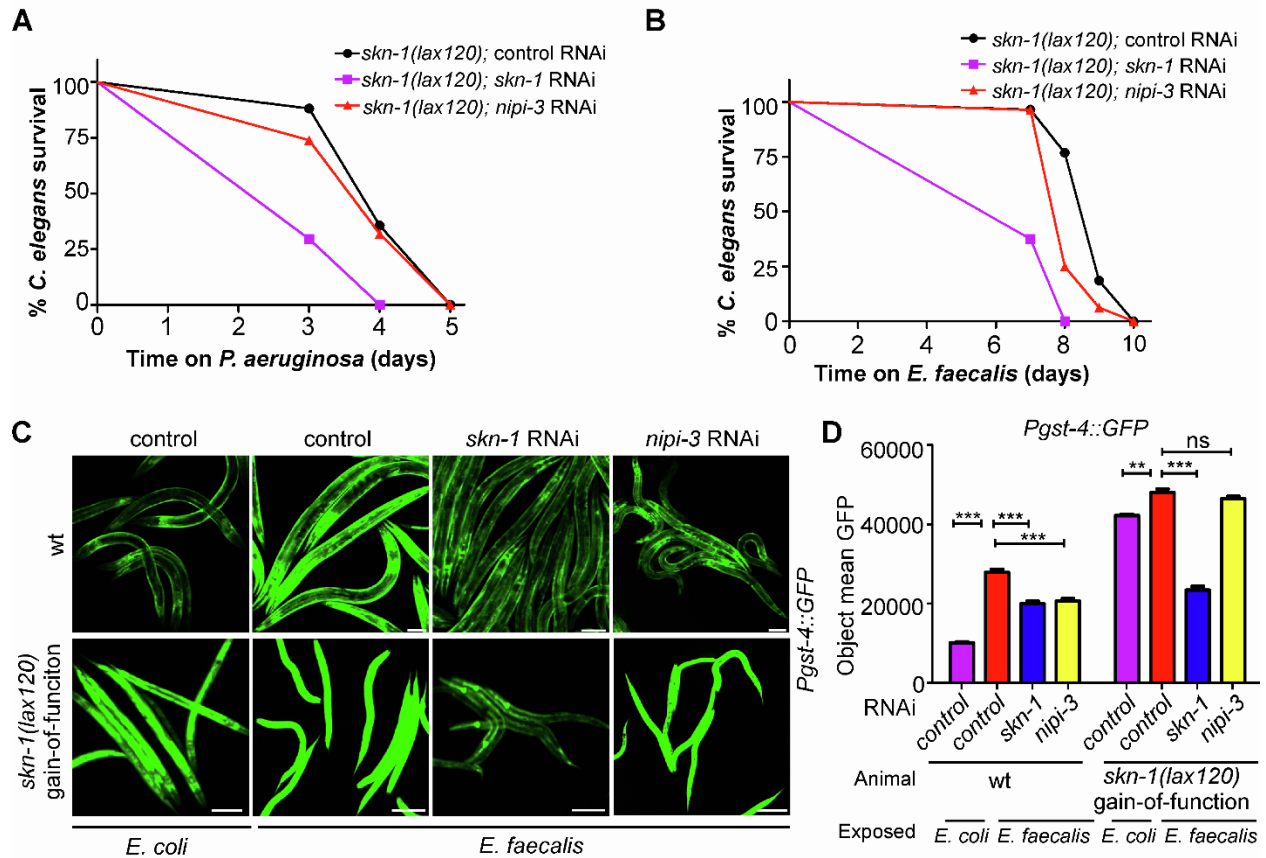


Figure S2: A *skn-1* gain-of-function mutant is unaffected by loss of NIPI-3. Related to Figure 2. *skn-1* gain-of-function animals (*skn-1(lax120)*) exposed to vector control, *skn-1*, or *nipi-3* RNAi prior to exposure to *P. aeruginosa* (A) or *E. faecalis* (B). $p < 0.0001$ for *skn-1* RNAi compared to *skn-1* or *nipi-3* RNAi. $p = ns$ and $p < 0.05$ for control vs. *nipi-3* RNAi on *P. aeruginosa* and *E. faecalis* respectively. Median survival and p values are listed in Table S1 along with replicates of all experiments. (C) Confocal microscopy of vector control, *skn-1*, or *nipi-3* RNAi treated *skn-1(lax120)* animals carrying *Pgst-4::gfp* after exposure to *E. faecalis* or *E. coli* for 16 hours. The images are representative of three independent experiments ($n > 150$ animals). Scale bars indicate 100 μ m. (D) Quantification of mean GFP fluorescence for the animals in (C). Three biological replicates with at least 50 worms/replicate were analyzed. Error bars represent the SEM. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, ns not significant.

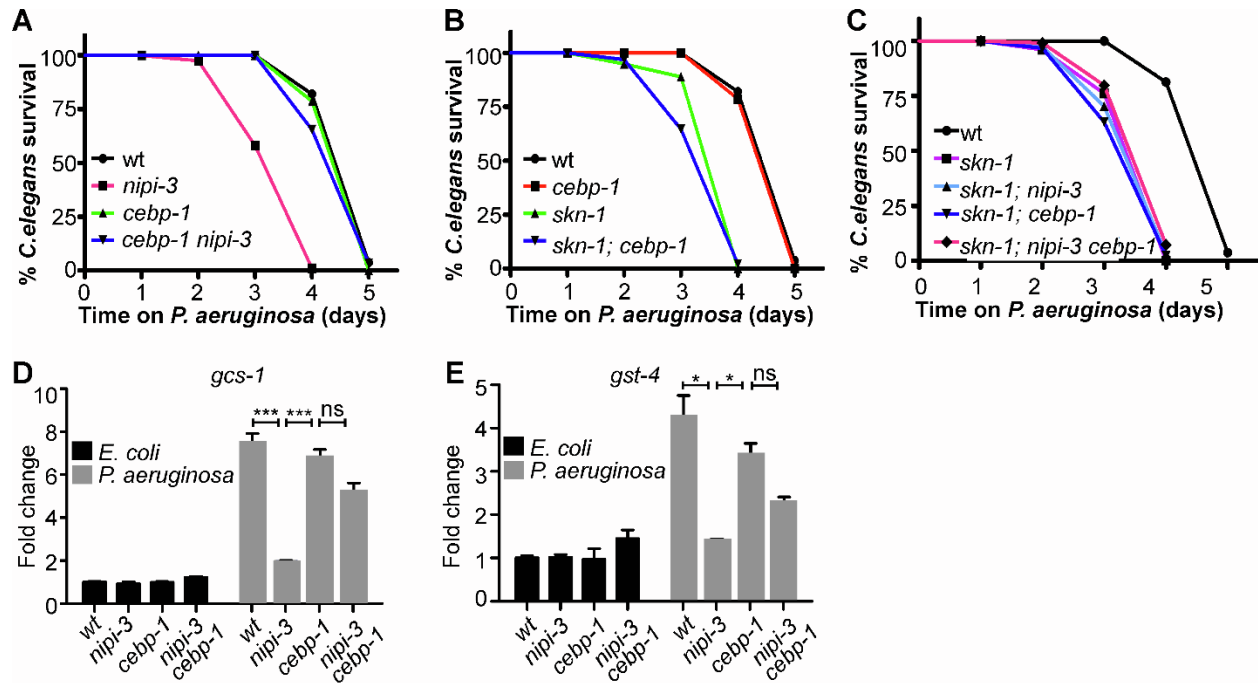
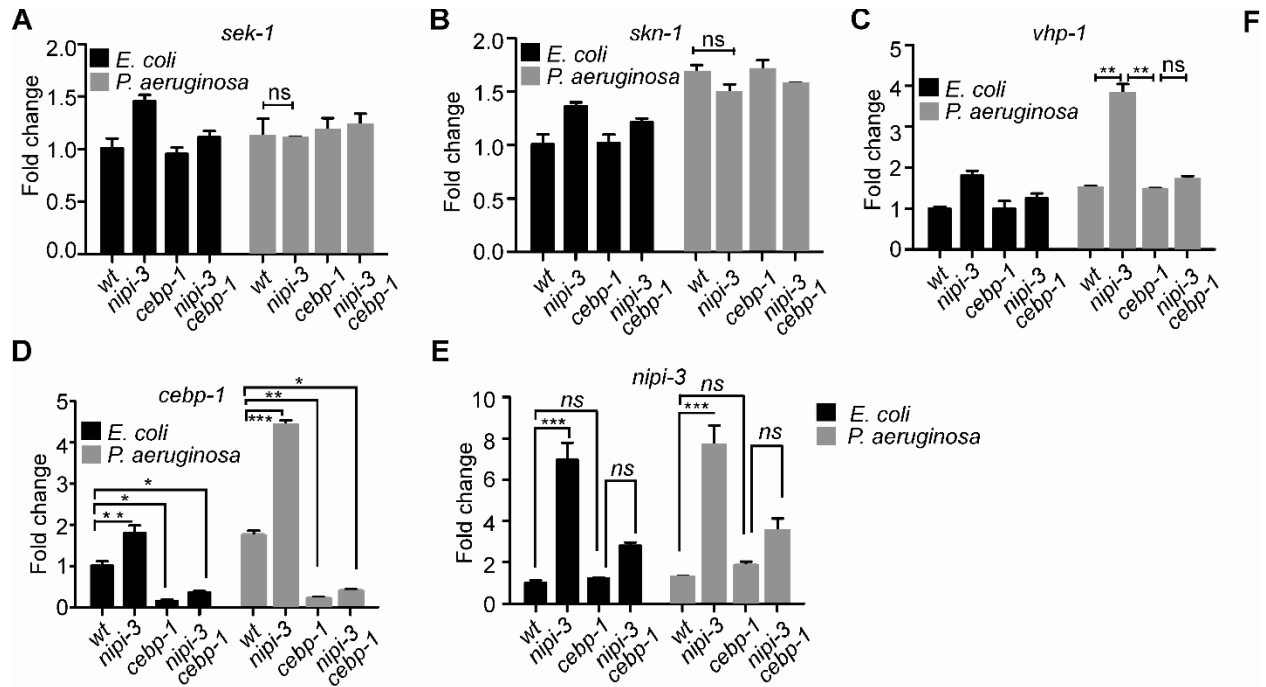


Figure S3: NIPI-3's effect on SKN-1 activation requires CEBP-1. Related to Figure 3. (A) Survival curves of wild type, *nipi-3(fr4)*, *cebp-1(tm2807)* and *nipi-3(fr4) cebp-1(tm2807)* on *P. aeruginosa* PA14. $p < 0.0001$ for *nipi-3* compared to all other strains. $p = ns$ for wild type, *cebp-1* and *cebp-1 nipi-3* compared to each other. **(B)** Survival curves of wild type, *cebp-1(tm2807)*, *skn-1(zu135)* and *skn-1(zu135); cebp-1(tm2807)* on *P. aeruginosa* PA14. $p < 0.0001$ for wild type compared to *skn-1* and *skn-1; cebp-1* and for *cebp-1* compared to *skn-1* and *skn-1; cebp-1*. $p = ns$ for wild type compared to *cebp-1* and for *skn-1* compared to *skn-1; cebp-1*. **(C)** Survival curves of wild type, *skn-1*, *skn-1; nipi-3*, *skn-1; cebp-1*, *skn-1; nipi-3 cebp-1* on *P. aeruginosa* PA14. $p < 0.0001$ for wild type compared to all strains. $p = ns$ for comparisons between all strains except wild type. For the survival experiments, animals were exposed to *P. aeruginosa* after treatment with *cdc-25.1* RNAi. Killing assays were performed in triplicate and repeated independently three times. Sample sizes, median survival, and p values of all trials are shown in Tables S1. **(D & E)** The defect in *gcs-1* **(D)** and *gst-4* **(E)** expression levels in the *nipi-3* background are suppressed by loss of *cebp-1* as revealed by qRT-PCR. Wild type (N2), *nipi-3(fr4)*, *cebp-1(tm2807)* and *nipi-3(fr4) cebp-1(tm2807)* animals were collected following exposure to *P. aeruginosa* or *E. coli* for 8 hours. Gene expression values are relative to the *act-1* housekeeping gene, with the N2 animals exposed to *E. coli* set to 1. The data shown are the mean \pm SEM of the three independent experiments, each of which was performed in triplicate. * $p < 0.05$, *** $p < 0.001$, ns not significant.



Figures S4: CEBP-1 affects *vhp-1*, *cebp-1* and *nipi-3* transcript levels. Related to Figure 4. qRT-PCR analysis of *sek-1* (A), *skn-1* (B), *vhp-1* (C), *cebp-1* (D), *nipi-3* (E) expression levels in animals of the indicated genotypes (wild type (N2), *nipi-3(fr4)*, *cebp-1(tm2807)*, *nipi-3(fr4) cebp-1(tm2807)*) exposed to *P. aeruginosa* or *E. coli* for 8 hours. The data shown are the mean \pm SEM of three independent experiments, each of which was performed triplicate. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, ns not significant.

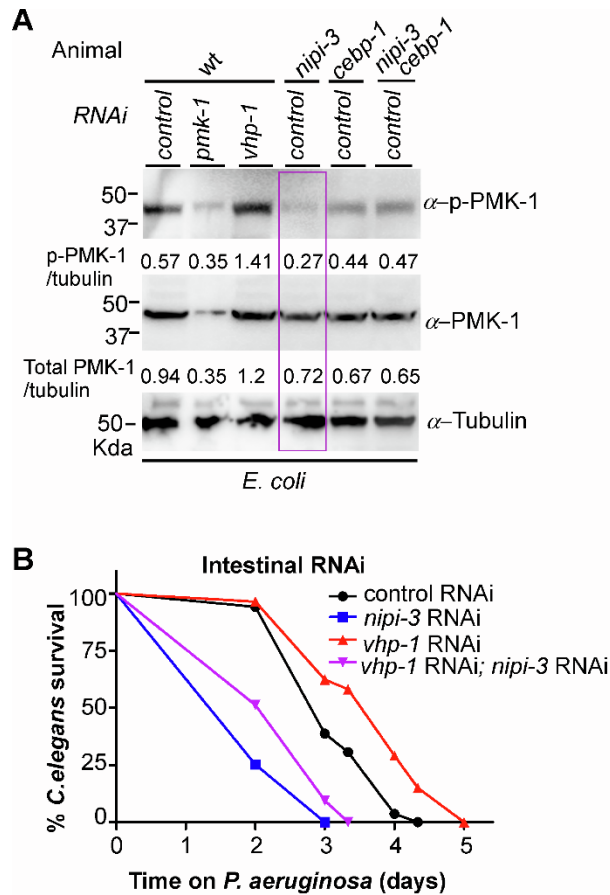


Figure S5: NIPI-3 regulates the PMK-1 p38 MAPK pathway via the phosphatase VHP-1. Related to Figure 5. (A) Loss of *nipi-3* results in lower levels of PMK-1 phosphorylation. Immunoblot analysis using an α -phospho-p38 antibody, an α -p38 antibody and an α -tubulin antibody (loading control) from lysates of the indicated animals exposed to *E. coli* for 16 hours. Band intensities relative to α -tubulin are indicated below each blot with the control set to 1. The lane containing the *nipi-3* lysate is boxed. **(B)** Epistasis analysis of intestine-only RNAi animals, either wild type or also containing the *pmk-1(ku25)* allele, exposed to *nipi-3* and *vhp-1* RNAi. $p < 0.0001$ for all strain comparisons. **(C)** Epistasis analysis of intestine-only RNAi animals exposed to *nipi-3* RNAi in both wild type and *pmk-1(km25)* mutant backgrounds. $p < 0.0001$ for all strain comparisons except for *pmk-1*; control RNAi to *pmk-1*; *nipi-3* RNAi, which is not significant. The VP303 background was used for all strains to enable intestinal specific RNAi, and *C. elegans* survival on *P. aeruginosa* was tracked over time. Data are representative of three trials. Sample sizes, median survival, and p values of all trials are shown in Table S1.

TABLE S2. Oligonucleotides related to STAR methods and Key Resources Table.

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Oligonucleotides		
confirm <i>skn-1(zu135)</i> genotype FWD primer zu135-seq-F: TTGTGGGGATTGAAGTGGGT	Sigma-Aldrich	N/A
confirm <i>skn-1(zu135)</i> genotype REV primer zu135-seq-R: CTCAAATAGGGCAAGTGGGT	Sigma-Aldrich	N/A
confirm <i>nipi-3(fr4)</i> genotype FWD primer <i>nipi-3</i> seq-F: CATCTCATCCATGACCGCTG	Sigma-Aldrich	N/A
confirm <i>nipi-3(fr4)</i> genotype REV primer <i>nipi-3</i> -seq-R: TTGAACTGCAAACCTCCG	Sigma-Aldrich	N/A
confirm <i>pmk-1(km25)</i> genotype FWD primer <i>pmk-1</i> -F: ACTTTCTCGTTAATCCTATAAGTTG	Sigma-Aldrich	N/A
confirm <i>pmk-1(km25)</i> genotype REV primer <i>pmk-1</i> -R: TTTGATAGATCAGCTCCCATCAACAT TG	Sigma-Aldrich	N/A
confirm <i>skn-1(lax120)</i> genotype FWD primer <i>lax120</i> -F: TGAAAAATAAAAAATGTTGAATG	Sigma-Aldrich	N/A
confirm <i>skn-1(lax120)</i> genotype REV primer <i>lax120</i> -R: ATTGATAGTTGGAATGAGAAGGTAT AGG	Sigma-Aldrich	N/A
confirm <i>cebp-1(tm2807)</i> genotype FWD primer <i>cebp-1</i> -F: GTTTGTACTTTCTTGCTCTGGTCTGTG G	Sigma-Aldrich	N/A
Confirm <i>cebp-1(tm2807)</i> genotype REV primer <i>cebp-1</i> -R: ATATCTATTTTTTATCCATTTTCATAG TGTTGG	Sigma-Aldrich	N/A
confirm intestinal special RNAi animals <i>rde-1(ne219)</i> -F: TGTTCAATTACACTATTCACAAGCAT TGG	Sigma-Aldrich	N/A
confirm intestinal special RNAi animals <i>rde-1(ne219)</i> -R: TTCAGGTCGAGATTGACAGAACG	Sigma-Aldrich	N/A
confirm <i>vhp-1(sa366)</i> genotype FWD primer <i>vhp-1</i> -F: ATGAAAATGGGAAGCGATGATGCAT ACAG	Sigma-Aldrich	N/A
confirm <i>vhp-1(sa366)</i> genotype REV primer <i>vhp-1</i> -R: ATTCTTGATGGAAGTCCAAGAGCTTT TGG	Sigma-Aldrich	N/A

qRT-PCR for <i>act-1</i> FWD primer RT-actin F: CCATCATGAAGTGCACATTG	Sigma-Aldrich	N/A
qRT-PCR for <i>act-1</i> REV primer RT-actin R: CATGGTTGATGGGGCAAGAG	Sigma-Aldrich	N/A
qRT-PCR for <i>cebp-1</i> FWD primer RT- cebp-1 F: GCAAGACAAGACTCTCTTAC	Sigma-Aldrich	N/A
qRT-PCR for <i>cebp-1</i> REV primer RT- cebp-1 R: CCAAGGTCCAGCTCAGTTTC	Sigma-Aldrich	N/A
qRT-PCR for <i>nipi-3</i> FWD primer RT-nipi- 3-F: GATGAAATCCGTTTAAAGCGAAC	Sigma-Aldrich	N/A
qRT-PCR for <i>nipi-3</i> REV primer RT-nipi-3- R: GGCGTAGGAGTGAATTGTAG	Sigma-Aldrich	N/A
qRT-PCR for <i>gcs-1</i> FWD primer RT-gcs- 1F: AATGCCTTACGGAGGTCTC	Sigma-Aldrich	N/A
qRT-PCR for <i>gcs-1</i> REV primer RT-gcs- 1R: AAGAGATGGGAACGATATCG	Sigma-Aldrich	N/A
qRT-PCR for <i>gst-4</i> FWD primer RT-gst-4 F: AAAGCTGAAGCCAACGACTC	Sigma-Aldrich	N/A
qRT-PCR for <i>gst-4</i> REV primer RT-gst-4 R: TCTGCAGTTTTTCCAGCG	Sigma-Aldrich	N/A
qRT-PCR for <i>vhp-1</i> FWD primer RT-vhp- 1-F: CCTCTACAACCATGTCTATCC	Sigma-Aldrich	N/A
qRT-PCR for <i>vhp-1</i> REV primer RT-vhp-1- R: CATGGTCTCATCCAAGCTATCA	Sigma-Aldrich	N/A
qRT-PCR for <i>skn-1</i> FWD primer RT-skn1- 1F: CTGGCATCCTCTACCACCAC	Sigma-Aldrich	N/A
qRT-PCR for <i>skn-1</i> REV primer RT-skn1- 1R: TTGGTGATGATGGCCGTGTT	Sigma-Aldrich	N/A
qRT-PCR for <i>sek-1</i> FWD primer RT-sek- 1F: CACTGTTTGGCGACGATGAG	Sigma-Aldrich	N/A
qRT-PCR for <i>sek-1</i> REV primer RT-sek- 1R: ATTCCGTCCACGTTGCTGAT	Sigma-Aldrich	N/A