## **Supporting Information**

## Wide-Range, Rapid, and Specific Identification of Pathogenic Bacteria by Surface-Enhanced Raman Spectroscopy

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Figure S1. Statistical diagrams of the length (A) and width (B) of AgNRs.



Figure S2. Mean average SERS spectra of blank and bacteria with different layers before (A-F) acetic acid treatment. (A) NL = 1, (B) NL = 2, (C) NL = 3, (D) NL = 4, (E) NL = 5, (F), NL = 6. Gray color shows the error bar of each spectra. The peak marked with \* indicates Si peak.



Figure S3. The integral of Raman shift: 1769 for (before acetic acid) and 526 for (after acetic acid) in the *E. Coli* O157 (red) and *B. thuringiensis* (grey), respectively.



**Figure S4.** SEM of SERS substrates with different number of layers (NL) of untreated (A-F) and acetic acid treated (A'-F') substrate and the diagram of the relationship between the NL and thickness of the substrates. (A), (A') NL = 1, (B), (B') NL = 2, (C), (C') NL = 3, (D), (D') NL = 4, (E), (E') NL = 5, (F), (F') NL = 6.



**Figure S5.** Relationship of NL and thickness of the substrates (the untreated marked in blue, the acetic treated marked in red). It can be seen that after the treatment of acetic acid, the thickness of SERS substrate is slightly thinner than the original one, indicating the partial removal of PVP on AgNRs.



**Figure S6.** (A) SEM image of AgNRs. The distance calculated between the two ends of AgNRs are marked in red, dashed box. (B) Statistical diagram of the gap distance between two ends of AgNRs calculated from (A). (C) Theoretical simulation of the electromagnetic field enhancement between two AgNRs. The length of AgNRs was 360 nm, and the gap between two

AgNRs was 3 nm. The excitation laser wavelength was 633 nm. The intensity of the electromagnetic field is represented by color scale. (D) Zoom of the hot spot between two AgNRs of (C).



Figure S7. SERS intensity of the monolayer AgNRs substrate, showing a relative standard deviation of 6%.



**Figure S8.** (A) SERS spectra of different region (selected) between the *B. melitensis* (red) and other 21 bacteria (grey). (B) Ratio of integral of Raman shift: 1165.49/1205.22 in the *B. melitensis* (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (ratio of integral of Raman shift: 1165.49/1205.22).



**Figure S9.** (A) SERS spectra of different region (selected) between the *F. tularensis* (red) and other 21 bacteria (grey). (B) Ratio of integral of Raman shift: 520.023/737.509 in the *F. tularensis* (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (ratio of integral of Raman shift: 520.023/737.509).



**Figure S10.** (A) SERS spectra of different region (selected) between the *Y. pestis* (red) and other 21 bacteria (grey). (B) Ratio of integral of Raman shift: 1525.96/1604.57 in the *Y. pestis* (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (ratio of integral of Raman shift: 1525.96/1604.57).



**Figure S11.** (A) SERS spectra of different region (selected) between the *E. coli* O157 (red) and other 21 bacteria (grey). (B) Ratio of integral of Raman shift: 516.381/518.808 in the *E. coli* O157 (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (ratio of integral of Raman shift: 516.381/518.808).



Figure S12. (A) SERS spectra of different region (selected) between the *S. paratyphi* A (red) and other 21 bacteria (grey).(B) Ratio of integral of Raman shift: 1397.16/1482.59 in the *S. paratyphi* A (red) and other 21 bacteria (grey). (C) ROC

curve with a 95% confidence interval (in brackets) generated from defined feature (ratio of integral of Raman shift: 1397.16/ 1482.59).



**Figure S13.** (A) SERS spectra of different region (selected) between the *S. paratyphi* B (red) and other 21 bacteria (grey). (B) Ratio of integral of Raman shift: 1034.78/ 1061.79 in the *S. paratyphi* B (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (ratio of integral of Raman shift: 1034.78/ 1061.79).



Figure S14. (A) SERS spectra of different region (selected) between the *S. paratyphi* C (red) and other 21 bacteria (grey).
(B) Integral of Raman shift: 1655.43 in the *S. paratyphi* C (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (integral of Raman shift: 1655.43).



**Figure S15.** (A) SERS spectra of different region (selected) between the *S. typhi* (red) and other 21 bacteria (grey). (B) Ratio of integral of Raman shift: 1184.28/1405.75 in the *S. typhi* (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (ratio of integral of Raman shift: 1184.28/1405.75).



**Figure S16.** (A) SERS spectra of different region (selected) between the *S. typhimurium* (red) and other 21 bacteria (grey). (B) Ratio of integral of Raman shift: 1096.53/1118.86 in the *S. typhimurium* (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (ratio of integral of Raman shift: 1096.53/1118.86).



**Figure S17.** (A) SERS spectra of different region (selected) between the *S. enteritidis* (red) and other 21 bacteria (grey). (B) Ratio of integral of Raman shift: 1542.8/1554.36 in the *S. enteritidis* (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (ratio of integral of Raman shift: 1542.8/1554.36).



Figure S18. (A) SERS spectra of different region (selected) between the *S. choleraesuis* (red) and other 21 bacteria (grey). (B) Ratio of integral of Raman shift: 860.018/1020.1 in the *S. choleraesuis* (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (ratio of integral of Raman shift: 860.018/1020.1).



Figure S19. (A) SERS spectra of different region (selected) between the *V. parahaemolyticus* (red) and other 21 bacteria (grey). (B) Ratio of integral of Raman shift: 1097.65/1147.76 in the *V. parahaemolyticus* (red) and other 21 bacteria (grey).
(C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (ratio of integral of Raman shift: 1097.65/1147.76).



Figure S20. (A) SERS spectra of different region (selected) between the *L. monocytogenes* (red) and other 21 bacteria (grey). (B) Ratio of integral of Raman shift: 876.19/ 927.929 in the *L. monocytogenes* (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (ratio of integral of Raman shift: 876.19/ 927.929).



**Figure S21.** (A) SERS spectra of different region (selected) between the *L. innocua* (red) and other 21 bacteria (grey). (B) Integral of Raman shift: 1551.21 in the *L. innocua* (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (integral of Raman shift: 1551.21).



**Figure S22.** (A) SERS spectra of different region (selected) between the *B. anthracis* (red) and other 21 bacteria (grey). (B) Ratio of integral of Raman shift: 929.074/1034.78 in the *B. anthracis* (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (ratio of integral of Raman shift: 929.074/1034.78).



**Figure S23.** (A) SERS spectra of different region (selected) between the *B. subtilis var.niger* (red) and other 21 bacteria (grey). (B) Integral of Raman shift: 1266.09 in the *B. subtilis var.niger* (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (integral of Raman shift: 1266.09).



Figure S24. (A) SERS spectra of different region (selected) between the *B. thuringiensis* (red) and other 21 bacteria (grey). (B) Ratio of integral of Raman shift: 1155.53/1209.62 in the *B. thuringiensis* (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (ratio of integral of Raman shift: 1155.53/1209.62).



**Figure S25.** (A) SERS spectra of different region (selected) between the *B. cereus* (red) and other 21 bacteria (grey). (B) Ratio of integral of Raman shift: 575.619/ 637.952 in the *B. cereus* (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (ratio of integral of Raman shift: 575.619/ 637.952).



**Figure S26.** (A) SERS spectra of different region (selected) between the *B. subtilis* (red) and other 21 bacteria (grey). (B) Ratio of integral of Raman shift: 1289.41/1442.14 in the *B. subtilis* (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (ratio of integral of Raman shift: 1289.41/1442.14).



**Figure S27.** (A) SERS spectra of different region (selected) between the *S. aureus* (red) and other 21 bacteria (grey). (B) Ratio of integral of Raman shift: 965.628/1000.85 in the *S. aureus* (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (ratio of integral of Raman shift: 965.628/1000.85).



Figure S28. (A) SERS spectra of different region (selected) between the *C. neoformans* (red) and other 21 bacteria (grey).
(B) Integral of Raman shift: 575.619 in the *C. neoformans* (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (integral of Raman shift: 575.619).



**Figure S29.** (A) SERS spectra of different region (selected) between the *M. smegmatis* (red) and other 21 bacteria (grey). (B) Ratio of integral of Raman shift: 517.595/ 520.023 in the *M. smegmatis* (red) and other 21 bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (ratio of integral of Raman shift: 517.595/ 520.023).



Figure S30. (A) Microscopic image of bacteria mixture (contains of *M. smegmatis* (Mycobacterium, marked in red circle), *S. aureus* (Coccus, marked in yellow circle), *E, coli O157, S. paratyphi* A, *V. parahaemolyticus* and *L. monocytogenes* (Bacterium, marked in blue circle)) in Raman system. (B) Integral of Raman shift: 1031 in the *M. smegmatis* (red) and other bacteria (grey). (C) ROC curve with a 95% confidence interval (in brackets) generated from defined feature (integral of Raman shift: 1031).

Source	bacteria	culture medium			
Anti-plague Institute Hebei Province	B. melitensis	Luria-Bertani			
Beijing Institute of Microbiology and Epidemiology	F. tularensis	Alkaline Peptone Broth			
Anti-plague Institute Hebei Province	Y. pestis	Luria-Bertani			
Beijing Institute of Microbiology and Epidemiology	E. coli O157 Luria-Bertani				
BNCC337823	S. paratyphi A	Luria-Bertani			
BNCC103389	S. paratyphi B	Luria-Bertani			
BNCC339948	S. paratyphi C	Luria-Bertani			
BNCC108331	S. typhi	Luria-Bertani			
BNCC108207	S. typhimurium	Luria-Bertani			
BNCC353759	S. enteritidis	Luria-Bertani			
BNCC126547	S. choleraesuis	Luria-Bertani			
BNCC105263	V. parahaemolyticus	Luria-Bertani			
BNCC338021	L. monocytogenes	Brain Heart Infusion Agar			
BNCC139467	L. innocua	Luria-Bertani			
Beijing Institute of Microbiology and Epidemiology	B. anthracis Luria-Bertani				
Beijing Institute of Microbiology and Epidemiology	B. subtilis var.niger Luria-Bertani				

BNCC136456	B. thuringiensis	Luria-Bertani
BNCC103930	B. cereus	Luria-Bertani
BNCC188062	B. subtilis	Luria-Bertani
BNCC310011	S. aureus	Luria-Bertani
BNCC337347	C. neoformans	YM Medium
BNCC104116	M. smegmatis	Brain Heart Infusion Agar

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classification				Region	Region	Region	Region	Region			p-value
			1	2	3	4	5		confidence		
Classification of	Morphological		– bacteria						AUC	interval	
dyeing	classification	species									
		brucella	B. melitensis	518	1000	1101	1208	1761	0.926	0.783-1.0	0.0053
		francisella	F. tularensis	523	886	1495	1509	1622	0.947	0.857-1.0	0.0092
		yersinia	Y. pestis	971	1092	1099	1584	1613	0.974	0.915-1.0	0.0039
		escherichia	E, coli O157	518	888	1167	1190	1799	0.974	0.921-1.0	< 0.0001
_	Bacterium		S. paratyphi A	1177	1446	1461	1485	1518	1	1.0	< 0.0001
G-			S. paratyphi B	752	868	939	1007	1039	0.926	0.794-1.0	0.0090
		salmonella	S. paratyphi C	1530	1625	1668	1731	1964	1	0.979-1.0	0.0001
			S. typhi	1189	1410	1511	1673	1759	1	1.0	0.0006
			S. typhimurium	1122	1231	1270	1385	1634	0.947	0.87-0.989	0.0168
			S. enteritidis	524	692	841	1108	1496	0.804	0.5-1.0	0.0140
			S. choleraesuis	526	742	855	945	1759	0.942	0.825-1.0	0.0010
	Vibrio	Vibrio	V. parahaemolyticus	863	1154	1165	1339	1636	0.926	0.778-1.0	0.0019
		listeria	L. monocytogenes	522	1617	1774	1869	1910	0.968	0.915-1.0	0.0067
			L. innocua	521	642	1237	1545	1559	0.738	0.54-0.926	0.2157
	Bacterium bacillus	B. anthracis	521	1024	932	1566	1596	0.968	0.899-1.0	0.0001	
G+		bacillus	B. subtilis var.niger	1132	1234	1313	1340	1598	1	1.0	0.0010
			B. thuringiensis	660	878	1214	1551	1755	0.979	0.937-1.0	0.003
			B. cereus	898	1402	1605	1621	1657	0.984	0.952-1.0	0.0017
			B. subtilis	521	1103	1167	1197	1294	0.995	0.955-1.0	< 0.0001
	Coccus	staphylococcu	S. aureus	854	1006	1145	1638	1842	0.966	0.862-1.0	0.0049
		e	C. neoformans	580	777	911	963	1053	1	1.0	< 0.0001
acid-fast staining	Mycobacterium	mycobacteriu	M. smegmatis	521	999	1066	1233	1701	0.984	0.937-1.0	< 0.0001

Table S3. Major spectral band assignments for bacteria.<sup>1-3</sup>

Raman Shift /cm <sup>-1</sup>	Band assignments	
~520	Si wafer	
580	C-O-C glycosidic ring deformation	
640-660	Nucleic acids	
742-777	Adenine, FAD, NAG	
841	Cytosine, uracil, tyrosine	
~855	ring breathing Tyr protein	
911-932	C-C stretching modes in proteins	
939-971	C-N stretching modes	
~1000	Phenylalanine ring breathing	
1024-1066	C-C ring breathing	
~1100	C-O stretching (amides), proteins	
1122	NH <sup>3+</sup> formation vibration	
1130-1180	Amide, DNA, C-N stretching vibration of adenine	
1216-1295	Amide III	
1313-1340	Adenine breathing ring	
1385	Thymine breathing ring	
1498-1410	C-O vibration, COO- symmetrical vibration	
1430-1461	CH2 deformation lipids, proteins, carbohydrates	
1494-1509	Phe	
1516-1553	C-H bend or C=C stretching	
1559-1605	C-N stretching proteins, DNA, Amides	
1617-1621	C=C stretching olefinic	

## REFERENCES

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