

1 **3D assessment of intervertebral disc degeneration in zebrafish identifies changes to**
2 **bone density that prime disc disease**

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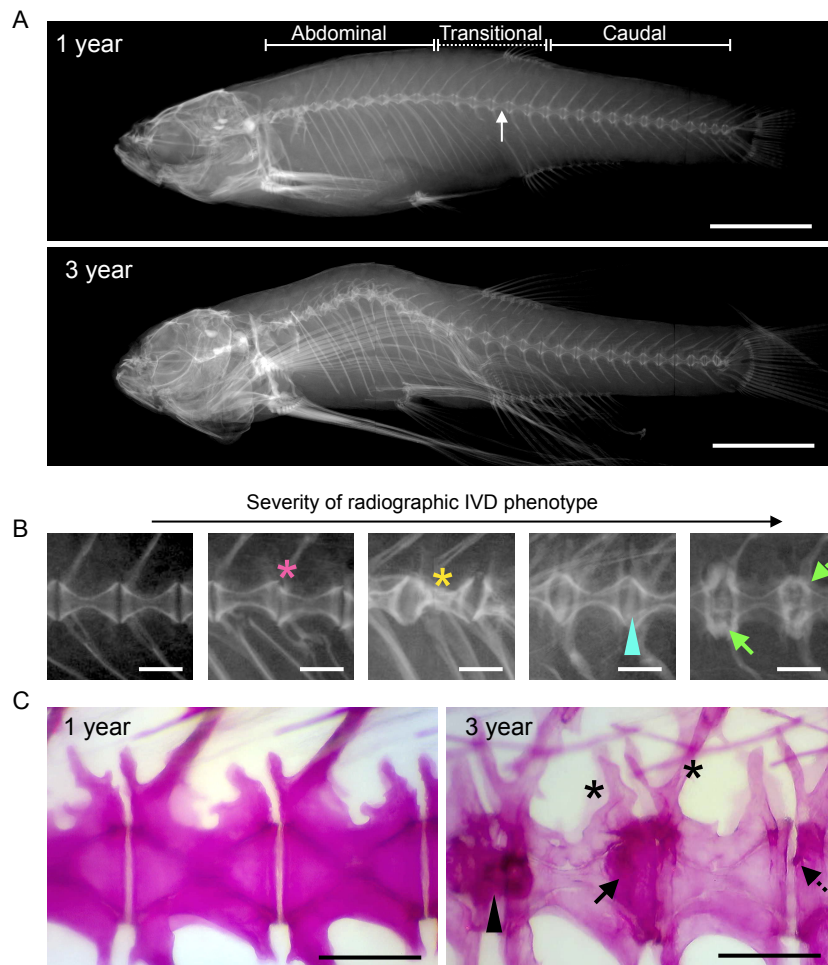
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9 **Supporting information listing**
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Item	Type	Title
Supplementary Fig. 1	Figure	IVD abnormalities detected through radiographs and Alizarin Red staining.
Supplementary Fig. 2	Figure	Shape deformities at the endplates during ageing.
Supplementary Fig. 3	Figure	Workflow for automated zebrafish vertebral centra segmentation.
Supplementary Fig. 4	Figure	Types of measurements retrieved from vertebral centra segmentation of young and aged zebrafish.
Supplementary Fig. 5	Figure	One year <i>sp7</i> ^{-/-} show lower bone density and dramatic abnormalities in the IVD.
Supplementary Fig. 6	Figure	Radiographs of the trunk of a 15 years old patient carrying a homozygous truncation in <i>SP7</i> (c.1052delA).
Supplementary Fig. 7	Figure	Workflow for segmentation and analyses of bone and osteocytes lacunae from SRCT images using a single U-Net model.
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Supplementary Fig. 9	Figure	Fibrotic and disorganised IVD characterize IVDD in <i>ctsk crp</i> and reduced bone elasticity modulus.

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12 **Supplementary Figures and legends**

13 **Supplementary Figure 1**



15 **Supplementary Fig 1. IVD abnormalities detected through radiographs and Alizarin Red**
16 **S staining.**

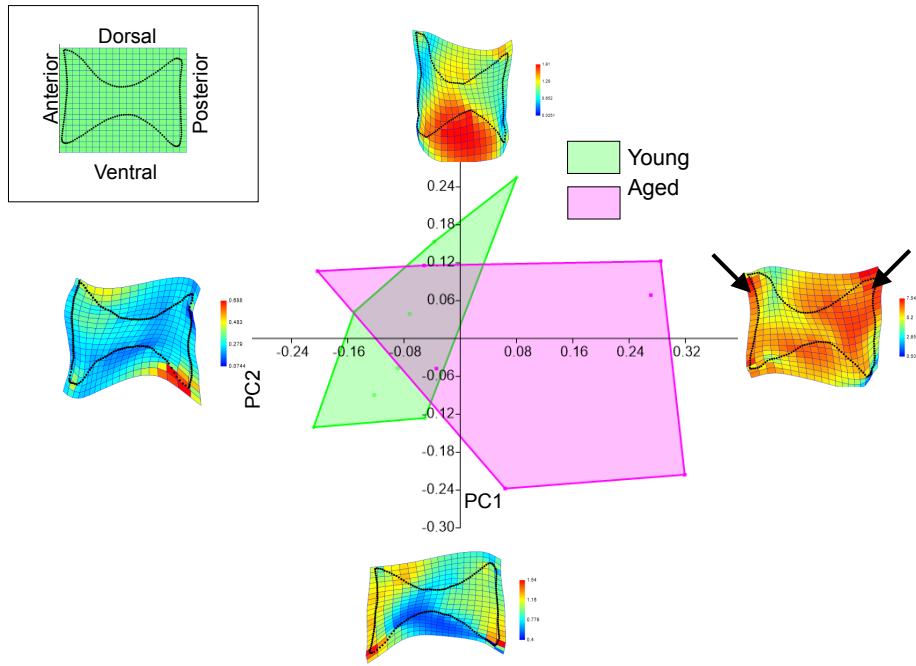
17 A) Radiographs of 1 and 3 year wt. The three areas of the axial skeleton are shown (abdominal,
18 transitional and caudal), in accordance with (De Clercq, Perrott et al. 2017). Transitional
19 vertebrae were chosen for TMD calculation from μ CT data. Note spinal curvature in aged
20 sample. Scale bars: 3 mm.

21 B) Examples of compromised radiographic IVDs, from normal (extreme left) to severely
22 affected (far most right). Misalignment (magenta asterisk), compromised centrum shape and
23 borders (yellow asterisk), IVD calcification (cyan arrowhead), sclerosis (green dashed arrow)
24 and fusions (green arrow). Scale bars: 500 μ m,

25 C) Alizarin Red S staining of 1 and 3 years old spines. Aged spines show uneven and lighter
26 centra staining, osteophytes (dashed arrow), sclerosis (arrow), IVD calcification (arrowhead)
27 and shape changes of arches and spinous processes (asterisks). Scale bars: 500 μ m.

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29 **Supplementary Figure 2**



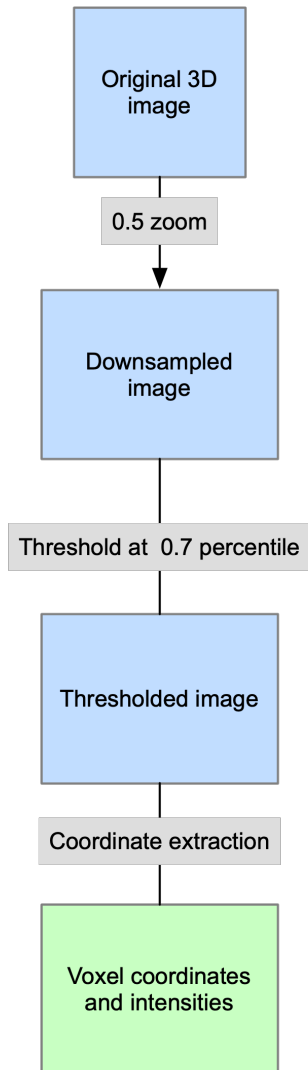
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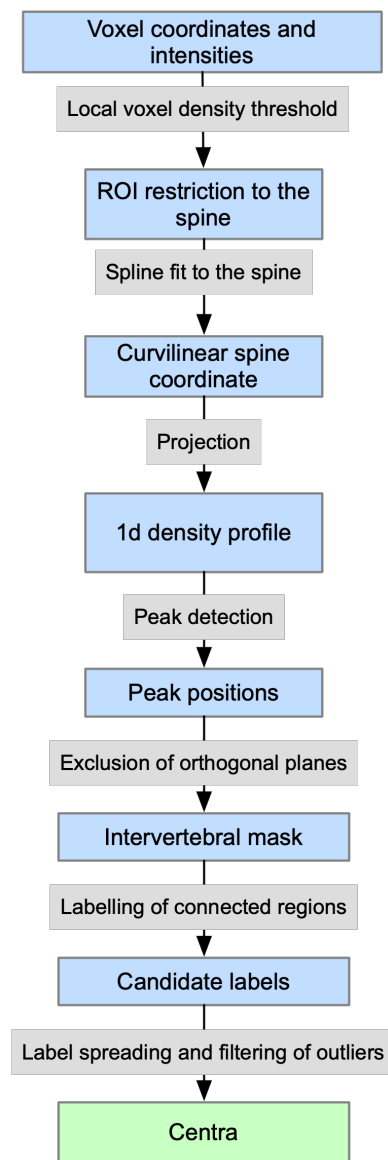
32 **Supplementary Fig. 2. Shape deformities at the endplates during ageing.**

33 Morphometric analysis showed distinct shape of vertebral centrum when comparing young and
34 aged samples (n= 8 for each group). Principal Component (PC) and PC2 displayed. Shape
35 deformation of the vertebral centrum, displayed with 20 grids colour-coded for points of stress
36 (deformation) in accord to the standard deviation of each PC. Note that in aged samples the
37 deformation of the centra is located towards the endplates (arrows, red colour grids). Top left,
38 vertebral centrum without any deformation, orientation is indicated. Non-parametric, ANOVA,
39 Bonferroni-corrected p-values. Euclidean distance measured. Graph generated in Past2.17.
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(A) Thresholding pipeline



(B) Detection pipeline



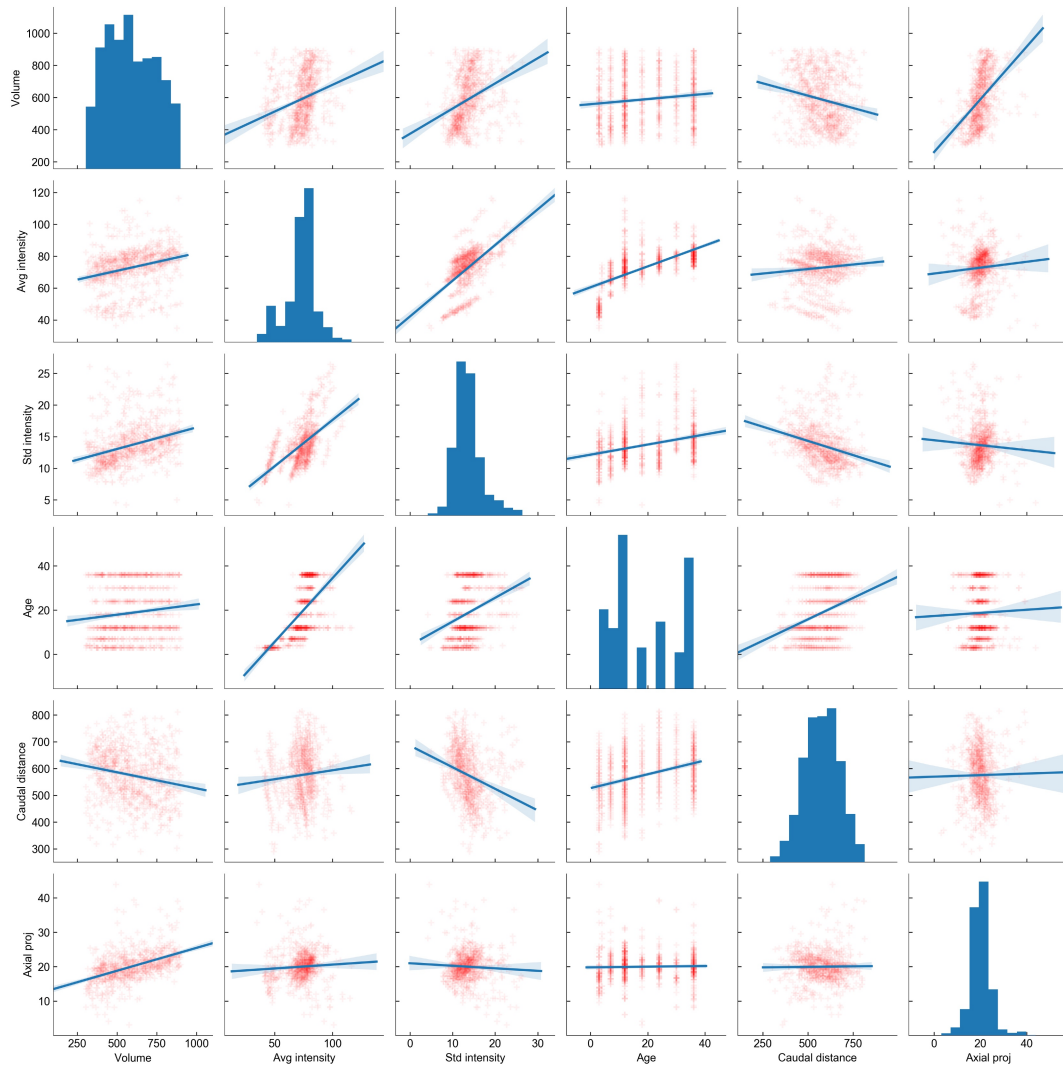
44 **Supplementary Fig. 3. Workflow for automated zebrafish vertebral centra segmentation.**

45 A) Pipeline for the first step of automation of centra segmentation, focused on thresholding the
46 image. To reduce computational costs, images are downsampled, and only the 0.7 highest
47 intensity percentile is retained. The thresholded image is then transformed into a set of
48 coordinates and intensities.

49 B) Pipeline for the second step of automation of centra segmentation, focused on detection of
50 vertebral centra (vertebrae). To detect the vertebrae we first automatically detect the region of
51 interest (ROI) excluding head and fins. We then focus on the spine region and establish a
52 reference curvilinear coordinate system over which we project the positions of the spine voxels.
53 This one-dimensional profile is used to identify intervertebral separations and a mask for the
54 labelling of connected regions. Through label spreading we recover the centra.

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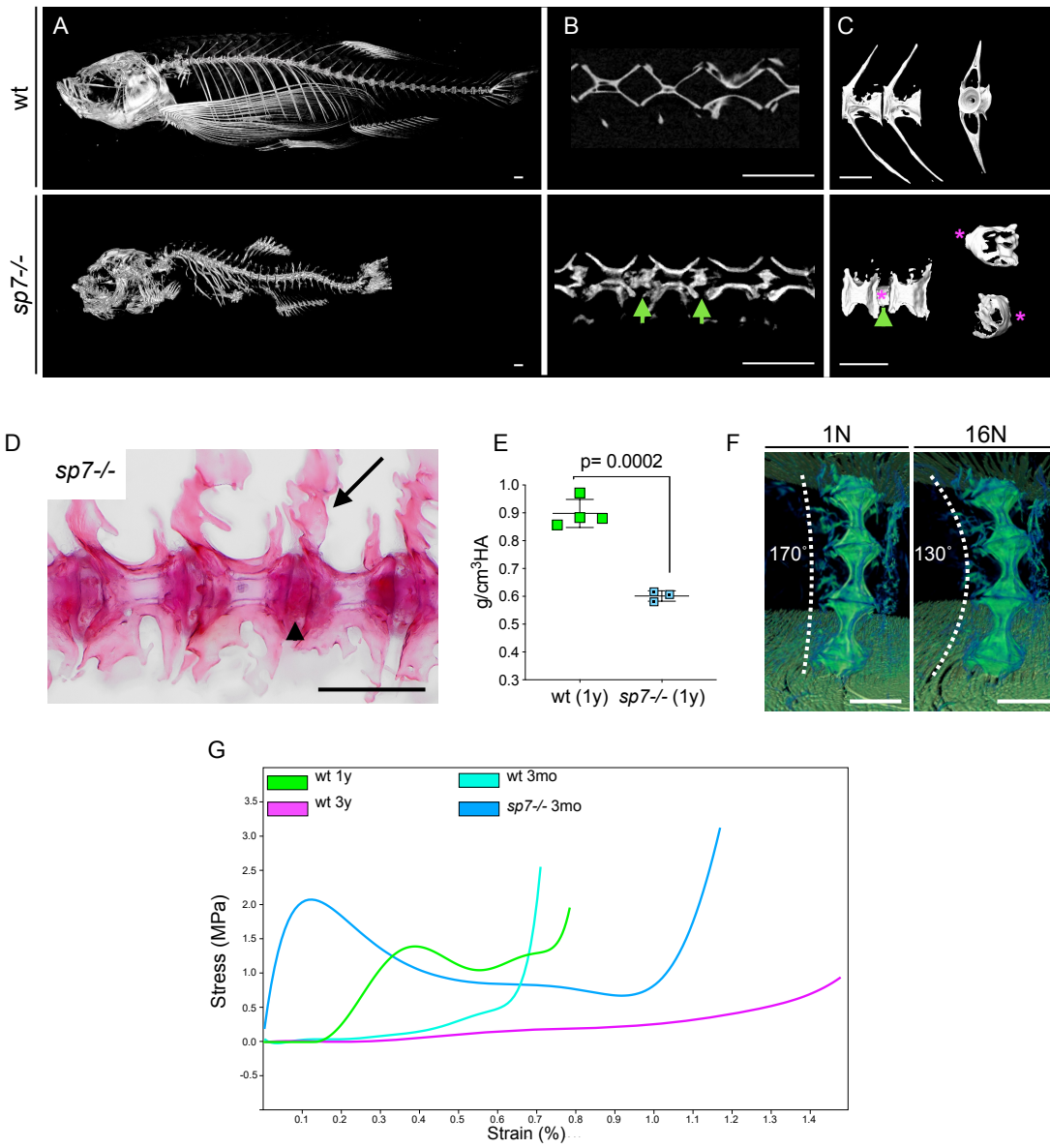
56 Supplementary Figure 4



58 **Supplementary Fig. 4. Measurements retrieved from vertebral centra segmentation of**
59 **young and aged zebrafish.**

60 Graphs showing age (wt, in months) of volume, average intensity (bone density), standard (std)
61 deviation of intensity per vertebra, caudal distance (distance to the caudal fin), axial projection
62 (length). Each point represents one centrum. Taking ageing as a comparison, small increase in
63 volume, average intensity, std intensity, and length is observed in wt spines. Higher densities
64 are observed in centra farther from the tail. Fish numbers: 1y (n= 36), 2y (n= 16) and 3y (n=
65 34). Graphs generated in Python.

66



69 **Supplementary Fig. 5. One year *sp7-/-* show lower bone density and dramatic**
70 **abnormalities in the IVD.**

71 A) 3D volumetric render from μ CT of 1y old wt and *sp7-/-* 1y. Scale bars= 500 μ m.

72 B) Sagittal orthoslice across the spine. Arrows point to IVD calcification in *sp7-/-*. Scale bars=
73 500 μ m.

74 C) Segmentation of two consecutive vertebrae from wt 1y and *sp7-/-* 1y. Note abnormal shape
75 in vertebrae centra in *sp7-/-*. IVD calcification was segmented (asterisk and arrowhead),
76 revealing not a solid mineralized rod, but calcified sheet layers. Asterisks positioned at the
77 same region to help with orientation of IVD calcification. Scale bars not placed for segmented
78 IVD calcified rods. Scale bars= 500 μ m.

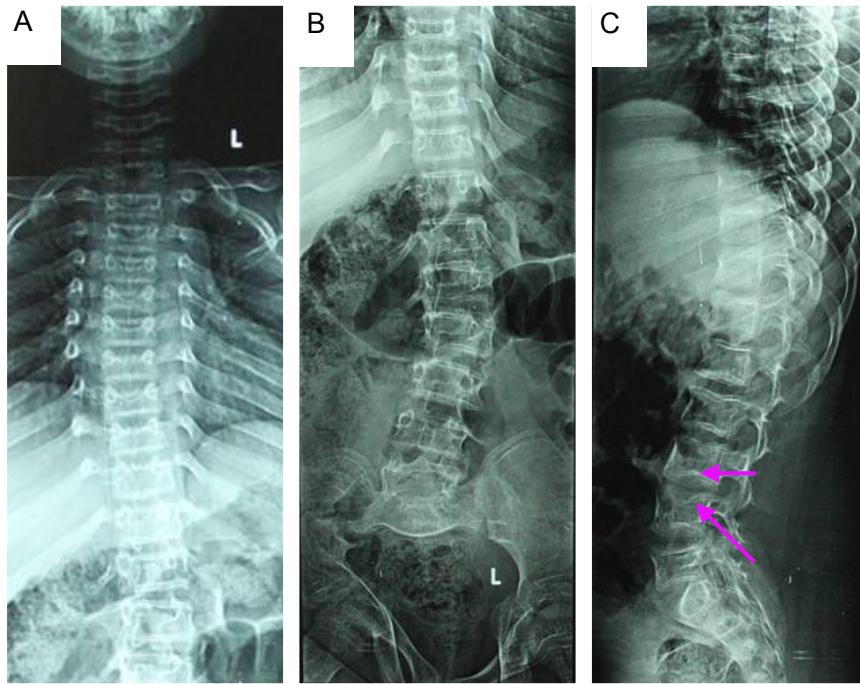
79 D) Alizarin Red showing a region of the *sp7-/-* spine. Note abnormal shape of centra and arches
80 (arrow) and IVD calcification (arrowhead). Scale bars= 500 μ m.

81 E) TMD values of 1 year wt and *sp7-/-*, calculated from μ CT data acquired under the same
82 conditions. Non-parametric, two-tail, Mann-Whitney test; data are mean SD. P-values are
83 indicated.

84 F) 3D volumetric render of the same *sp7-/-* sample during the compression (1N and 16N). Note
85 the bent spine at 16N. Scale bars= 500 μ m.

86 G) Stress x Strain from the vertebral compression experiment to show Young's modulus. Lines
87 represent average of three samples. Graph generated in MatLab.

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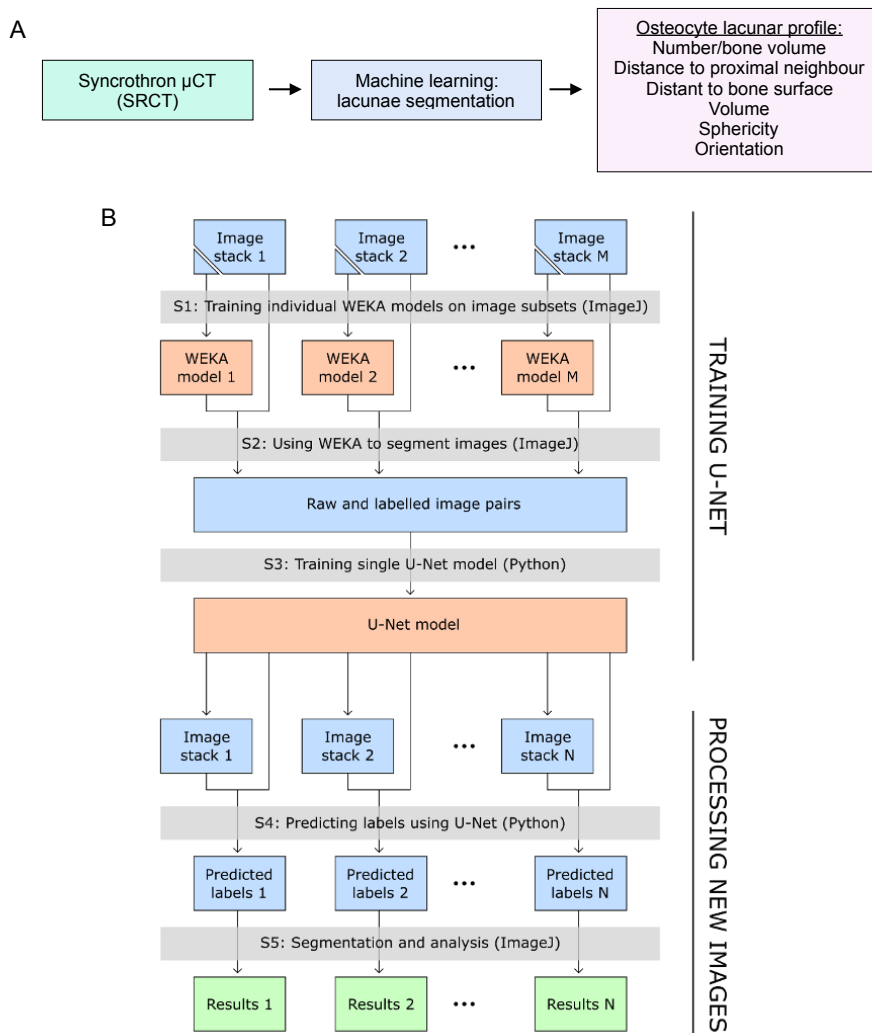


91 **Supplementary Fig. 6. Radiographs of the trunk of a 15 years old patient carrying a**
92 **homozygous truncation in *SP7* (c.1052delA).**

93 A and B) Anterior posterior x-ray view of dorsolumbar spine showing mild platyspondyly and
94 scoliosis with mild deviation to the right in the thoracic region and convex to the left in the
95 lumbar region. Posterior aspects of ribs are deformed with smooth kinking.

96 C) X-ray lateral view of dorsolumbar spine showing generalised osteopenia, biconcave thoracic
97 and lumbar vertebrae with irregular transverse lines of sclerosis at the upper and lower vertebral
98 cortical endplates and increased intervertebral disc space (arrows).

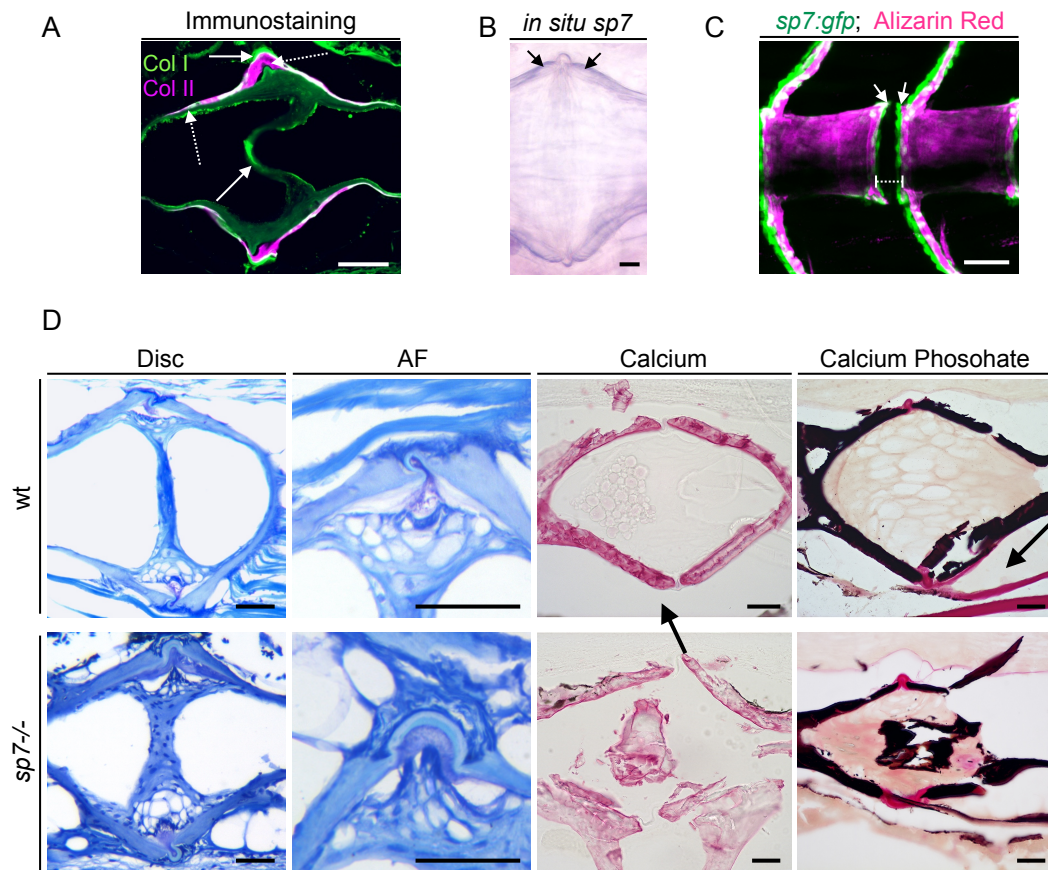
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102 **Supplementary Fig. 7. Workflow for segmentation and analyses of bone and osteocytes**
103 **lacunae from SRCT images using a single U-Net model.**

104 A) Machine learning was used to retrieve osteocyte lacunae information from SRCT data. B)
105 The workflow was split into two parts: (1) Creation of the single U-Net model for pixel
106 classification and (2) use of the U-Net model on previously-unseen images to facilitate
107 downstream segmentation and analysis. The two parts of the workflow were split into five
108 discrete steps (labelled S1-S5) and use either Python or ImageJ/Fiji. S1: Example 2D regions
109 from a subset (M) of the images to be ultimately processed (N ; where $M \leq N$) were extracted
110 and used to train WEKA pixel classification models (one classifier created per image stack).
111 S2: The WEKA classifiers were used to create probability maps for all slices in the
112 corresponding example image stacks. These probability maps were binarised and converted to
113 pairs of raw and class-labelled images using a MIA workflow (v0.14.13). The image pairs were
114 then converted to 512×512 px² overlapping tiles and combined into a single dataset. S3: A
115 single Tensorflow/Keras U-Net model was trained on the tiled image pairs, which can
116 subsequently be applied to all SRCT images acquired in a similar manner. S4: The U-Net
117 model was applied to image stacks (both previously seen and unseen) and outputs probability
118 maps for classes of interest. S5: Probability maps were segmented and analysed using MIA to
119 yield measurements of lacunae counts, morphology and bone morphology.

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124 **Supplementary Fig. 8. IVD changes in *sp7*^{-/-} are part of premature degeneration.**

125 A) Histological section of the intervertebral disc of wt, immune-stained for collagen type II
126 and collagen type I. Collagen type I localises to the notochord strand and layer in AF (arrows),
127 collagen type II in AF and notochord sheath (dashed arrows). Scale bars= 50 µm.

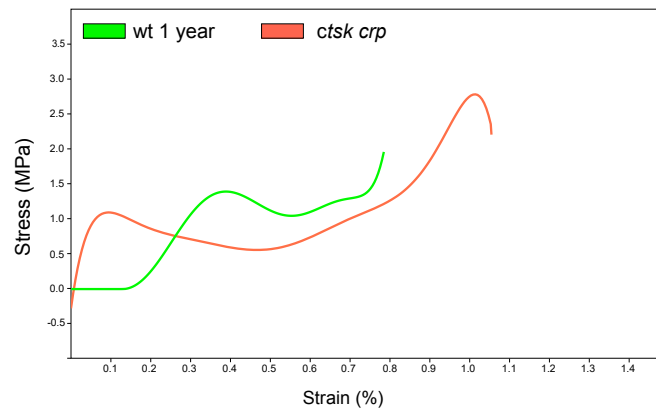
128 B) Whole- mount *in situ* hybridisation for *sp7* showing the IVD region. *sp7* is expressed at the
129 endplates, no signal was observed in the NP. *In situ* hybridization was carried out as previously
130 described (Kague, Roy et al. 2016). Scale bars= 50 µm.

131 C) Maximum projection from confocal stacks of *Tg(Ola.Sp7:nlsGFP)^{zfl132}* live stained with
132 Alizarin Red S. *Sp7*⁺ in osteoblasts at the endplate (arrows). IVD (dashed line). Scale bar = 50
133 µm.

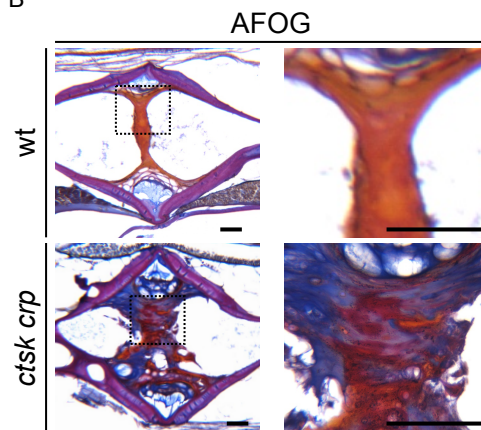
134 D) Histological sections of wt and *sp7*^{-/-} (3 months old) to show the first signs abnormalities
135 in the AF (Toluidine Blue). Cryosections of 6 months old fish stained for Alizarin Red S to
136 show that calcium is the component of disc calcification. Von-Kossa show calcium phosphate
137 in calcified IVD (arrows) and bones. Scale bars= 50 µm.

138

A



B



141 **Supplementary Fig. 9. Fibrotic and disorganised IVD characterise IVDD in *ctsk crp* and**
142 **reduced bone elasticity modulus.**

143 A) Stress x Strain graph to represent Young's modulus of wt and *ctsk crp* (1 year) during
144 vertebral compression. Lines are average of each group (n= 3). The exponential ramp of each
145 line correspond to Young's modulus (elasticity). The peak of each curve shows the failure
146 point. Graph generated in MatLab.

147 B) Histological sections of wt and *ctsk crp* (1 year) stained with AFOG (Acid Fuchsin Orange
148 G) to show fibrosis (orange) and disorganization of the NP. The region within the dashed box
149 is shown in higher magnification. Scale bars = 50 μ m.

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