1	Supplemental Data for
2	
3	The long noncoding RNA <i>Synage</i> regulates synapse stability and neuronal
4	function in the cerebellum
5	
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13	This PDF file includes:
14	Supplemental Figures S1-S11
15	Supplemental Tables S1-S3
16	

17 Supplemental Figure S1 (related to Figure 1)

18



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- Fig. S1 Distribution of *Synage* lncRNA in the C8-D1A cell line. RNA FISH of *Synage* (green)
- in the C8-D1A cerebellum cell line, lacZ probe (green) was used as a negative control. Nuclei were
- $_{23}$ stained with Hoechst 33342 (blue). Scale bar: 10 $\mu m.$

Supplemental Figure S2 (related to Figure 1)



29 Fig. S2 Synage IncRNA is conserved in its genomic location and in its distribution specificity

- 30 in the cerebellum. a Schematic view of the genomic loci of the *Synage* transcripts in the mouse
- 31 brains detected by ribo-minus RNA-Seq. **b** and **c** Schematic view of the genomic loci of the
- 32 homologous Synage, LOC106995009 and RP11-491F9.1, in the rhesus macaque (b) and human
- 33 genomes (c), respectively. The *Cbln1* gene is shown in the red box. The genomic loci of both *Synage*
- in the mouse and homologous *Synage* in the rhesus macaque and human are shown in the purple
- box. **d** Gene expression level for RP11-491F9.1 in human tissues screened from the GTEx project
- 36 database (dbGaP Accession phs000424.v8.p2).

37 38

Supplemental Figure S3 (related to Figure 1)



39 40

Fig. S3 The expression level of the mouse Synage transcripts in different brain regions. a 41 Schematic view of Synage transcripts detected by ribo-minus RNA-Seq. b FPKM (Fragments Per 42 Kilobase Million) value of the Synage transcripts in the whole brains of 2-month-old mice detected 43 by ribo-minus RNA-Seq (n=3). c-g The relative expression level of the Synage transcripts in the 44 adult mouse cerebellum (c), olfactory bulb (d), cortex (e), hypothalamus (f), and hippocampus (g), 45 detected by RT-qPCR (n=5). The sequence of n264625 overlaps with most of the sequence of 46 n285242, so that a RT-qPCR primer pair specifically targeting n285242 was unable to be designed, 47 48 but a RT-qPCR primer pair specifically targeting both n285242 and n264625 can be designed, so does a RT-qPCR primer pair specifically targeting n264625. The expression level of n285242 was 49 estimated indirectly by removing the expression level of n264625 transcript from the total 50 expression level of n285242 and n264625 transcripts detected by RT-qPCR. 51 52

Supplemental Figure S4 (related to Figure 1) 53

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Fig. S4 The characterization of the full length mouse Synage IncRNA. a The position of gene 57 specific primers (GSPs) targeting Synage in the 5'- and 3'-RACE experiments. b-d The 5'- (b) and 58 3'-RACE (c) PCR amplification of Synage fragments using respective GSPs. The 4.8 kb pAMP1 59 5'-RACE recombinant was a positive control, which was provided by the commercial 5'-RACE kit 60 (d). e cDNA sequencing and assembling of 5'- and 3'-RACE of Synage fragments. 61 62

63 Supplemental Figure S5 (related to Figure 2)



67 Fig. S5 Synage knockout causes cerebellar atrophy and neuronal loss. a Genotype identification of Synage wild-type (WT), heterozygous (HT), and homozygous (KO) mice by genomic PCR 68 amplification for Synage. b Genomic sequencing of Synage in WT and Synage KO mice. c The 69 distribution of Synage (green) in the brain sections of WT and KO adult mice detected by RNA 70 FISH. Nuclei were stained with Hoechst 33342 (blue). Left scale bar: 1 mm; right scale bar: 500 71 µm. d Representative images of 2-month-old WT and KO male mice. e and f Body weights of adult 72 WT and KO male (e) and female (f) mice. g and h Relative weights of whole brain in adult male 73 (g) and female (h) mice. i Relative weights of the cerebella in adult female mice, normalized to 74 body weight, each dot represents a mouse. **j and k** Representative images (**j**) and relative weights 75 (k) of the cerebella of 2-month-old WT, Synage HT, and KO male mice, normalized to body weight 76 (n=3). I and m Quantification of the number of Purkinje cells per cerebellar sections (I) and 77 representative immunofluorescence staining images of Purkinje cells (m) in the cerebella of adult 78 WT, Synage HT, and KO male mice (n=3). Nuclei were stained with Hoechst 33342 (blue). Left 79 80 scale bar: 500 µm; right scale bar: 50 µm. n and o Representative images of Western blots (n) and quantification (o) for both NeuN (a neuronal marker) and Gephyrin (an inhibitory postsynaptic 81 marker) in adult WT and KO mouse cerebella, normalized to GAPDH. 82

84 **Supplemental Figure S6 (related to Figure 2)**





Fig. S6 Synage knockout mice show neuronal loss during cerebellar development. a-j 88 Representative immunofluorescence staining images (a) and quantification of the number of PCs 89 (Calbindin, green), the integrated intensity of granule cells (GCs), and the integrated intensity of 90 Bergmann glial cells (BGCs, Gdf10, red) in the cerebella from P10 (b-d), P23 (e-g), and 2-month-91 old (h-i) WT and Synage KO mice. Nuclei were stained with Hoechst 33342 (blue). Left scale bar: 92 500 µm; right scale bar: 50 µm. The signal intensity of Hoechst 33342 staining in the granule cell 93 laver was used to estimate GCs. 94





99 Fig. S7 *Synage* knockout impairs motor abilities and motor-dependent learning and memory

100 of mice. a The time(s) that adult mice stayed on the rotarod when tested at constant speeds between

4 and 40 rpm before falling in the rotarod test (WT: n = 10; KO: n = 8). **b** The average time(s) that

102 it took adult mice to cross an 80 cm beam with a flat surface of 10 mm or 8 mm width in the balance

103 beam test (WT: n = 12; KO: n = 12).

104 Supplemental Figure S8 (related to Figure 4)



108 Fig. S8 Knockdown of *Synage* results in substantial reduction in the expression of *Cbln1 in*

- 109 *vitro* and *in vivo*. a Relative expression levels of *Synage* transcripts in the HT-22 cell line detected
- 110 by RT-qPCR. b Relative expression levels of Synage lncRNA and Cbln1 mRNA after Synage
- shRNA knockdown in the HT-22 cell line detected by RT-qPCR. **c and d** Western blot analysis (c)
- and quantification (d) for CBLN1 protein in the HT-22 cell line following Synage knockdown,
- 113 normalized to β-Tubulin. e Relative expression levels of *Synage* lncRNA and *Cbln1* mRNA after
- 114 Synage shRNA knockdown in the mouse cerebella detected by RT-qPCR (OB, olfactory bulb;
- 115 Cereb, cerebellum). **f** and **g** Representative images of Western blot (**f**) and quantified expression
- 116 level (g) of CBLN1 protein, normalized to GAPDH, in 2-month-old WT and KO mice. h
- 117 Representative confocal images of CBLN1 with Calbindin by immunofluorescence staining in 3-
- 118 week-old WT and KO mouse cerebella after stereotaxic injection of AAV-control or AAV-Synage
- 119 into the neonatal mouse cerebella. Scale bar: $10 \ \mu m$.

- 121 Supplemental Figure S9 (related to Figure 5)
- 122





Fig. S9 Identification and verification of *Synage*-binding proteins. a Relative quantification of LRP1, HSP90AA1, and PSD-95 in the cerebellar cortex of 2-month-old WT and KO mice detected by immunofluorescence staining of the related proteins. **b-c** Western blots (**b**) and relative quantification (**c**) for LRP1, HSP90AA1, and PSD-95 proteins in the WT and KO mouse cerebella (n=3).

131 Supplemental Figure S10 (related to Figure 5)



- Fig. S10 Relative expression levels of *Synage* transcripts in the Neuro-2a cell line detected by
- **RT-qPCR**.



142Fig. S11 LRP1 depletion inhibits the interaction between LRP1-HSP90AA1 and Synage. a-f143Western blots (a-c) and quantification (d-f) for knockdown analyses of LRP1 (a, d), HSP90AA1144(b, e), and PSD-95 (c, f) protein in the HT-22 cell line following each protein knockdown,145normalized to β-Tubulin. g The expression level of Synage lncRNA after LRP1 knockdown and146Synage overexpression in the HT-22 cell line detected by RT-qPCR. h Western blots assessing147LRP1 and HSP90AA1 immunoprecipitation (IP) by anti-LRP1 antibody after LRP1 knockdown148and Synage overexpression in the HT-22 cell line.149

Supplemental Table S1. AGO2 CLIP-Seq peak information in mouse cortex

AGO2 CLI	P-Seq pea	k information in	mouse cortex (scree	ned from GSE73058	8, Moore et al., 2015)
Chromosome	Strand	Start	End	Peak.ID	Gene symbol
	-	89,992,794	89,992,854	200144	Cbln1
	-	89,992,926	89,992,986	200161	Cbln1
	-	89,993,134	89,993,194	200186	Cbln1
	-	89,993,267	89,993,327	200198	Cbln1
	-	89,993,327	89,993,387	200212	Cbln1
	-	89,993,503	89,993,563	200243	Cbln1
	-	89,993,797	89,993,857	200292	Cbln1
	-	89,993,904	89,993,964	200314	Cbln1
chr8	-	89,994,005	89,994,065	200330	Cbln1
	-	89,994,208	89,994,268	200347	Cbln1
	-	89,995,646	89,995,706	200385	Cbln1
	-	89,995,883	89,995,943	200408	Cbln1
	-	89,996,015	89,996,075	200437	Cbln1
	+	90,016,211	90,016,271	142479	Synage (Gm2694
	+	90,022,615	90,022,675	142568	Synage (Gm2694
	+	90,028,588	90,028,648	142659	Synage (Gm2694
	+	90,038,866	90.038,926	142745	Svnage (Gm2694

Supplemental Table S2. Phenotypes of knockout mice

Phenotypes	<i>Synage-</i> /- mice	<i>Cbln1</i> ^{-/-} mice ⁽¹⁻³⁾	<i>LRP1-/-</i> mice ^(4, 5)
Decreased synaptic vesicles	+	-	NA
Impaired synaptic function	+	+	+
Synaptic loss	+	+	+
Neuronal loss	+	-	+
Abnormal electrophysiology	+	+	+
Motor dysfunction and memory deficits	+	+	+
Reduced fertility	+	-	+
Decreased cerebellar weight	+	-	NA

155 Abbreviations: +, present; -, absent; NA, not applicable

Name	Application	Sequence (5'-3')
Synage_sgRNA 1	CRISPR/Cas9	TCCGGCCGGGAGTCAGACGA
Synage_sgRNA 2	CRISPR/Cas9	AAGGGCATGGTGGGTTGGCG
Synage_genotype_F1	genotype	GAGCCAATGGTTGCCCTGTC
Synage_genotype_F2	genotype	ACAAAGGCGCGGATCAAGC
Synage_genotype_F3	genotype	GGATACCCACCCTGCATTATC
Synage_genotype_R1	genotype	AACTCCACTGTTACTGCTAATACA
Synage_genotype_R2	genotype	CTCCTACCTGAAAGCCCACGAA
GAPDH_F	RT_qPCR	ACATCATCCCTGCATCCACTG
GAPDH_R	RT_qPCR	CCTGCTTCACCACCTTCTTG
Synage_p1_F	RT_qPCR	ACCAAGCATGTCCTGAAGATG
Synage p1 R	RT qPCR	CTGGGGATACTGCTGAATCAA
Synage p2 F	RT qPCR	ACTCTCAGCCTAACGTCTCCAA
Synage p2 R	RT qPCR	GATGCTCCAGCTTCTCAGACAG
Cbln1 F	RT qPCR	CTGGCTGTATTCCGTATT
<i>Cbln1</i> R	RT qPCR	ACAAGCATCAGAGAACAA
	RT qPCR	TTAGCGTTAGCGTTTATTGAGCAC
mmu-mir-325-3p R	RT aPCR	TATGGTTGTTCACGAGTCCTTGTC
<i>U</i> /6 F	RT aPCR	ATTGGAACGATACAGAGAAGATT
U6 R	RT aPCR	GGAACGCTTCACGAATTTG
n424059 F	RT aPCR	TCGCCCGAGCCTTCTACTTG
n424059 R	RT aPCR	CAGGGGCTTCTATCCACTCTAC
n264625 (Svnage-P1) F	RT aPCR	GATTCAGCTCGCTCACACCT
n264625 (Synage-P1) R	RT aPCR	CATTGTGCATGTAGCAGCGAA
n412835 F	RT aPCR	TGCTGATAACCCACCTTGGC
n412835 R	RT_qPCR	AAGACCGCACCACAATCCAT
n264625 & n285242 (Synage-P2) F	RT_qPCR	AGAGAAACAACTGCCCTTCTCC
n264625 & n285242 (Synage-P2) R	RT aPCR	TTCCTGAGCAGGCAGTATCC
n274385 F	RT aPCR	GTGAAACTACATCCACGCCCA
n274385 B	RT_qPCR	CTGACCGTTGTTCTAACGCA
n271779 F	RT aPCR	ACGTACCCTGCACACATA
n271779 R	RT_qPCR	GCCAAAGCCCCCAGCTAATA
Chln1 NRO F1	NRO	GTCTACAACAGACAGACCATCC
Chln1 NRO R1	NRO	CACTGATTTCTGACGCTTG
Chln1 NRO F2	NRO	ATCAGGAGCACCAACCATGA
Chln1 NRO R2	NRO	CAATCCATTCTGGGTGCTGC
$Gm^{2}694$ NRO F1	NRO	AACACAGCTGCCTCTACCAC
$Gm^{2}694$ NRO R1	NRO	TCGGCGCGATGCTGCACT
$G_m 2694$ NRO F2	NRO	
$Gm^{2}694$ NRO R2	NRO	TCTTCATCTTCAGGACATGC
$Gm^{2}694$ NRO F3	NRO	TGTCTGAGAAGCTGGAGCATCA
$G_m 2694$ NRO R3	NRO	GAACGGGAGTAGATCTGAC
HPRT NDO EL	NRO	GCTTCCTCACACCGCTT
HIRI_NKO_II HDDT_NDO_D1	NRO	TCTGCTGGAGTCCCCTTG
HPRT NPO E2	NRO	GGAATGTGTCCTGTAAAAGT
$\frac{HPRT}{P} NPO P2$	NRO	
HPRT NDO E2	NRO	
$\frac{HPRT}{P} NDO D2$	NRO	
Surgeo 2/DACE CSD1	DACE	
Synage_SKACE_GSP1	RACE	
Synage 2'D ACE_CSD2	DACE	
Synage 5 DACE CSP1	KAUE	
Synuge_SKACE_USP1	RACE	
Synage_JKACE_GSP2	KACE	
Synage_SKACE_GSP3	KACE	
Synage_SIKINAI	Knockdown	
Synage_shRNA2	Knockdown	AGCGCGGTGCGTCGCGATT

LRP1_shRNA1	Knockdown	TTACCAAAGAATGTGTTCAGC
LRP1_shRNA2	Knockdown	TTAGCCAGTGTATTTGTTCGC
LRP1_shRNA3	Knockdown	TTCATACATCTTGTAGGTAGG
HSP90AA1_shRNA1	Knockdown	AAACTTAGGGTTGTTCTCGGG
HSP90AA1_shRNA2	Knockdown	ATCATCCTCATCAATACCTAG
HSP90AA1_shRNA3	Knockdown	AATGAAATTCAGATACTCAGG
PSD-95_shRNA1	Knockdown	TATACTGAGCGATGATCGTG
PSD-95_shRNA2	Knockdown	ACACTGTTGACCGCCAGGATCTTGTCTCC
PSD-95_shRNA3	Knockdown	GAGTCTCTCTCGGGGCTGGGACCCAGATGT

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