



Fig. S4: Role of endogenous RTase and RNase H1 in antiviral responses in primary mESCs.

a-c, The derivation and characterization of primary mESCs. **a**, Phase contract images of the two primary mESCs: ESC-18 and ESC-22 (left panel) and alkaline phosphatase staining (AP staining) of the primary ESCs (right panel). **b**, The ESC-18 and ESC-22 cells were fixed and stained with pluripotent markers: SSEA-1 (red) and Oct4 (green). Nuclei were stained with DAPI (blue). Scale bar, 10 μ m. **c**, Spontaneous differentiation of the primary mESCs through EB formation. The expression levels of the pluripotent markers (*nanog* and *oct4*) and the germ layer markers (*brachyury*, mesendoderm; *foxa2*, endoderm; *mixl*, mesoderm; *nestin*, ectoderm) were measured by qRT-PCR. **d, e**, Inhibition of endogenous RTase activity by AZT promoted virus infection in the primary ESCs. ESC-18 (**d**) and ESC-22 (**e**) cells were pre-treated with 100 μ M AZT for 6 h and then were infected with EMCV (moi=1) for another 24 h. The RNA level of EMCV was determined by qRT-PCR (left panel) and the viral titers were examined by plaque assay (right panel). **f, g**, RNase H1 restricted virus infection in the primary mESCs. ESC-18 (**f**) and ESC-22 (**g**) cells were infected with EMCV (moi = 1) after transfected with siRNAs for 36 h. Twenty-four hours later, the RNA level of RNaseH1 and EMCV were determined by qRT-PCR (left panel) and viral titers were valued by plaque assay (right panel). Data in **a, b** are representative of two independent experiments. The graphs represent means \pm SD from three (**c**) or four (**d-g**) independent replicates measured in triplicate. Statistics were calculated by the two-tailed unpaired Student's *t*-test.