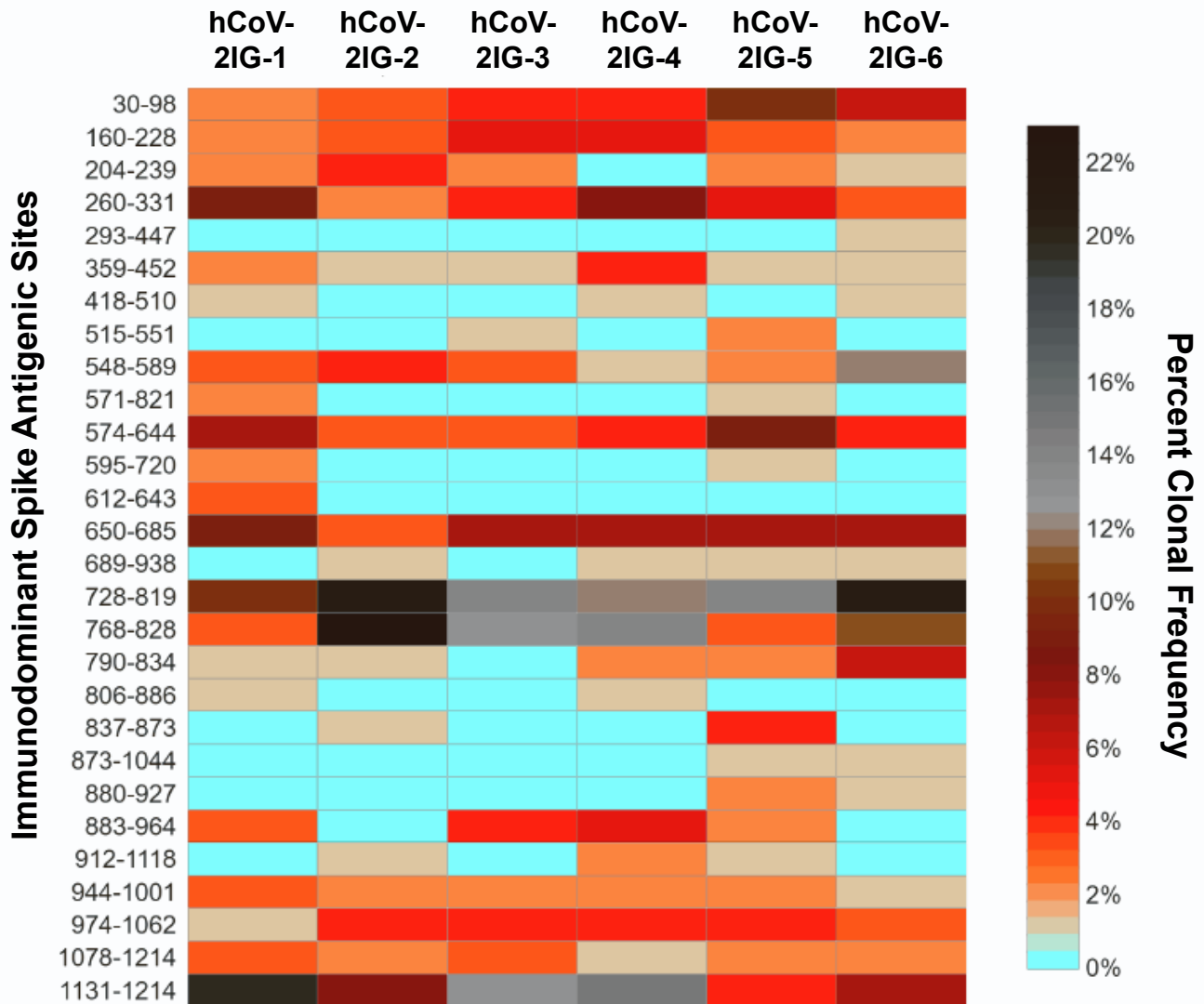


**iScience, Volume 24**

**Supplemental information**

**Epitope diversity of SARS-CoV-2 hyperimmune  
intravenous human immunoglobulins and  
neutralization of variants of concern**

**Juanjie Tang, Yuri Lee, Supriya Ravichandran, Gabrielle Grubbs, Chang Huang, Charles B. Staft, Tony Wang, Basil Golding, Hana Golding, and Surender Khurana**



**Figure S1. SARS-CoV-2 epitope profile of six hCoV-2IG batches by GFPDL analyses. Related to Figure 1.** Heat map of immunodominant sites ( $\geq 3$  clonal frequency in at least one hCoV-2IG lot) on the SARS-CoV-2 spike recognized by IgG antibodies in six hCoV-2IG lots identified using GFPDL analyses. The immunodominant sites on the left indicate amino acid residue of the antigenic sites in the spike protein. Color scale on the right represents range of percentage of clonal occurrences (frequency) of each site. Heat map was generated using R package.



**Table S1: Sequence conservation of GFPDL-identified antigenic regions/sites among different SARS-CoV-2 VOCs\* Related to Figure 1 and Figure 2.**

AA	Sequence	WA-1	CA (B.1.429)	UK (B.1.1.7)	JP (P.1)	SA (B.1.351)
30-98	NSFTRGVVYPDKVFRSSVLHSTQDLFLPFFSNVTFWFAIHVSGTNGTKRFDNPLVFNDFGVYFASTEKS	100	100	97.1	100	98.5
160-228	YSSANNCTFEYVYQPFMLDLEGGKQGNFKNLREFVFKNIDGYFKIYKSHKHTPINLVRDLPGGFSALEPLVD	100	100	100	98.5	98.5
204-239	YSKHTPINLVRDLPGGFSALEPLVDLPIGINITRFQ	100	100	100	100	97.2
260-331	AGAAAYVGYLQPRTFLLKYNENGTITDAVDCALDPLSEKCTLKSFTVEKGIYQTSNFRVQPTESIVRFPN LDPLSEKCTLKSFTVEKGIYQTSNFRVQPTESIVRFPNITLCPGGEVFNATRFASVYAWNRKRISNCVADYSVLY	100	100	100	100	100
293-447	NSASFSTFKCYGVSPKLNLDLCTFNVYADSFVIRGDEVRIAPGQTKGIADYNYKLPDDFTGCVIWNSNLDLSDK VGG	100	100	100	99.3	99.3
359-452	SNCVADYSVLYNSASFSTFKCYGVSPKLNLDLCTFNVYADSFVIRGDEVRIAPGQTKGIADYNYKLPDDFTGCVI AWNSNLDLSDKVGNNYLY	100	98.9	100	98.9	98.9
418-510	IADYNYKLPDDFTGCVIWNSNLDLSDKVGNNYLYRFLFRKSNLKPFFERDISTEIQAGSTPCNGVEGFNCFYPL QSYGFQPTNGVGYQPYRV	100	98.9	98.9	97.8	97.8
515-551	FELLHAPATVCGPKKSTNLVKNKCNVFNFLGTGTGV	100	100	100	100	100
548-589	GTGVLTESNKKFLPFQQRDIADTTDAVRDPQTEILDITP DTTDAVRDPQTEILDITPCSFVGGVSVITPGTNTSNQVAVLYQDVNCTEVPVAIHADQLTPTWRVYSTGNSVFQTR AGCLIGAEHVNNNSYECDIPIGAGICASYQTQNSPRRARSVASQSIIAYTMSLGAENSVAYSNNIAIPTNFTISVTT EILPVSMTKTSVDCTMYICGDSTECSNLLQYGSFCTQLNRALTGIAVEQDKNTQEVFAVQKQIYKTPPIKDFGGF NFSQILPDPSPKSKRSFIEDL	100	99.6	98.8	99.2	99.2
574-644	DAVRDPQTEILDITPCSFVGGVSVITPGTNTSNQVAVLYQDVNCTEVPVAIHADQLTPTWRVYSTGNSVFQ VSVITPGTNTSNQVAVLYQDVNCTEVPVAIHADQLTPTWRVYSTGNSVFQTRAGCLIGAEHVNNNSYECDIPIGAGI CASYQTQNSPRRARSVASQSIIAYTMSLGAENSVAYSNNIAIPTNFTI	100	98.5	98.5	98.5	98.5
595-720	YQDVNCTEVPVAIHADQLTPTWRVYSTGNSVF	100	99.2	97.6	98.4	98.4
612-643	YQDVNCTEVPVAIHADQLTPTWRVYSTGNSVF	100	96.8	96.8	96.8	96.8
650-685	LIGAEHVNNNSYECDIPIGAGICASYQTQNSPRRAR SOSIIAYTMSLGAENSVAYSNNIAIPTNFTISVTTTELIPVSMTKTSVDCTMYICGDSTECSNLLQYGSFCTQLNR LTGIAVEQDKNTQEVFAVQKQIYKTPPIKDFGGFNFSQILPDPSPKSKRSFIEDLLFNKVTLDAGFIKQYGDCLGDI AARDLCAQKFNGLTVLPPLLTDEMIAYTSALLAGTITSGWTFGAGAALQIPFAMQMAYRFGIGVTVQNVLYENQ KLIANQFNLSAIGKIQDLSL	100	100	97.2	97.2	100
689-938	LTGIAVEQDKNTQEVFAVQKQIYKTPPIKDFGGFNFSQILPDPSPKSKRSFIEDLLFNKVTLDAGFIKQYGDCLGDI AARDLCAQKFNGLTVLPPLLTDEMIAYTSALLAGTITSGWTFGAGAALQIPFAMQMAYRFGIGVTVQNVLYENQ KLIANQFNLSAIGKIQDLSL	100	100	99.6	100	99.6
728-819	PVSMTKTSVDCTMYICGDSTECSNLLQYGSFCTQLNRALTGIAVEQDKNTQEVFAVQKQIYKTPPIKDFGGFNFS QILPDPSPKSKRSFIE	100	100	100	100	100
768-828	TGIAVEQDKNTQEVFAVQKQIYKTPPIKDFGGFNFSQILPDPSPKSKRSFIEDLLFNKVTL	100	100	100	100	100
790-834	KTPPIKDFGGFNFSQILPDPSPKSKRSFIEDLLFNKVTLDAGFI	100	100	100	100	100
806-886	KRSFIEDLLFNKVTLDAGFIKQYGDCLGDIARDLCAQKFNGLTVLPPLLTDEMIAYTSALLAGTITSGW	100	100	100	100	100
837-873	YGDCLGDIARDLCAQKFNGLTVLPPLLTDEMIAY YTSALLAGTITSGWTFGAGAALQIPFAMQMAYRFGIGVTVQNVLYENQKLIANQFNLSAIGKIQDLSLSTASALGKLQ DVVNQNAQALNTLVKQLSSNFGAISVNLNLDLRLDKVEAEVQIDRLITGRQLSQTYYTQQLIRAAEIRASANLAAT KMSECVLGQSKRVDFCG	100	100	100	100	100
873-1044	DVVNQNAQALNTLVKQLSSNFGAISVNLNLDLRLDKVEAEVQIDRLITGRQLSQTYYTQQLIRAAEIRASANLAAT KMSECVLGQSKRVDFCG	100	100	100	100	100
880-927	GTITSGWTFGAGAALQIPFAMQMAYRFGIGVTVQNVLYENQKLIANQF	100	100	100	100	100
883-964	TSGWTFGAGAALQIPFAMQMAYRFGIGVTVQNVLYENQKLIANQFNLSAIGKIQDLSLSTASALGKLQDVVNQNAQ ALNTLVK	100	100	100	100	100
912-1118	TQNVLYENQKLIANQFNLSAIGKIQDLSLSTASALGKLQDVVNQNAQALNTLVKQLSSNFGAISVNLNLDLRLDKVE AEVQIDRLITGRQLSQTYYTQQLIRAAEIRASANLAATKMSECVLGQSKRVDFCGKGYHLSFPQSAPHGVVFLH VTYVPAQEKNFITTAICHGDKAHFPREGVVFVSNNGTHWVFTQRNFYEPQIITD	100	100	99	99.5	100
944-1001	ALGKLDVVNQNAQALNTLVKQLSSNFGAISVNLNLDLRLDKVEAEVQIDRLITGRQLSQTYYTQQLIRAAEIRASANLAAT KMSECVLGQSKRVDFCGKGYHLSFPQSAPHGVVFLH	100	100	98.2	100	100
974-1062	SSVNLNLDLRLDKVEAEVQIDRLITGRQLSQTYYTQQLIRAAEIRASANLAATKMSECVLGQSKRVDFCGKGYHLSFPQSAPHGVVFLH MSFPQSAPHGVVFLH	100	100	98.8	98.8	100
1078-1214	APAICHGDKAHFPREGVVFVSNNGTHWVFTQRNFYEPQIITDNTFVSGNCDVIGVNNTVYDPLQPELDSFKEELD KYFKNHTSPDVLGDISGINASVNIQKEIDRLNEVAKNLSLIDLQELGKYEYKWPW	100	100	99.2	99.2	100
1131-1214	GIVNNTVYDPLQPELDSFKEELDKYFKNHTSPDVLGDISGINASVNIQKEIDRLNEVAKNLSLIDLQELGKYEYKWPW QYIKWPW	100	100	100	98.8	100

\*Percent sequence conservation of GFPDL identified antigenic sites in the spike protein of SARS-CoV-2 variants namely WA-1 strain (QI187782.1), CA variant (B.1.429, EPI\_ISL\_648527), UK variant (B.1.1.7, QQQ47833.1), JP variant (P.1, QRX39425.1), and SA variant (B.1.351, EPI\_ISL\_678597) was calculated using Sequence Identity Matrix function in BioEdit.

**Table S2: SARS-CoV-2 variants mutations introduced in the spike plasmid for production of SARS-CoV-2 pseudovirions to test them in PsVNA.  
Related to Figure 2.**

<b>Mutations constructed in the spike plasmids</b>	
<b>CA</b>	S13I, W152C, L452R, D614G
<b>UK</b>	Del69-70, Del145,N501Y, A570D, D614G, P681H, T716I, S982A, D1118H
<b>JP</b>	L18F, T20N, P26S, D138Y, R190S, K417T, E484K, N501Y, H655Y, T1027I, D614G, V1176F
<b>SA</b>	L18F, D80A, D215G, Del242-244, K417N, E484K, N501Y, D614G, A701V

**Table S3: Neutralization titers of convalescent plasma, I2019-VIG and hCoV-2IG against SARS-CoV-2 variants\* Related to Figure 2.**

	WA-1		CA		UK		JP		SA	
	PsVNA50	PsVNA80	PsVNA50	PsVNA80	PsVNA50	PsVNA80	PsVNA50	PsVNA80	PsVNA50	PsVNA80
<b>IVIG batches produced in 2019 prior to COVID-19</b>										
2019-IVIG-1	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20
2019-IVIG-2	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20
2019-IVIG-3	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20
2019-IVIG-4	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20
2019-IVIG-5	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20
2019-IVIG-6	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20
2019-IVIG-7	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20
2019-IVIG-8	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20
2019-IVIG-9	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20
2019-IVIG-10	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20
2019-IVIG-11	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20
2019-IVIG-12	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20
2019-IVIG-13	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20
2019-IVIG-14	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20
2019-IVIG-15	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20
2019-IVIG-16	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20
<b>Convalescent plasma batches produced from COVID-19 survivors</b>										
CP-1	243.1	28.0	24.7	<20	53.4	<20	287.0	61.1	107.4	30.6
CP-2	369.2	56.8	122.9	30.0	128.3	23.9	126.0	24.7	25.5	<20
CP-3	204.9	30.1	146.9	37.3	168.4	24.0	112.0	27.3	47.2	<20
CP-4	268.0	31.3	92.8	<20	59.8	<20	53.8	<20	<20	<20
CP-5	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20
CP-6	211.1	47.2	123.3	26.1	64.8	21.6	<20	<20	<20	<20
CP-7	67.0	<20	98.5	22.2	36.6	<20	24.1	<20	<20	<20
CP-8	119.6	30.2	156.8	33.0	35.4	<20	<20	<20	<20	<20
CP-9	852.7	74.9	191.0	55.6	100.7	35.4	122.6	25.4	51.7	<20
<b>hCoV-2IG batches produced from COVID-19 convalescent plasma donors</b>										
hCoV-2IG-1	2878.8	515.0	2009.3	613.7	1799.4	401.1	672.2	269.1	382.0	154.5
hCoV-2IG-2	2919.7	435.0	1301.5	305.8	1224.9	394.0	681.7	235.8	259.0	119.5
hCoV-2IG-3	3309.1	593.2	2081.3	495.7	1957.1	499.1	833.4	302.2	278.0	124.1
hCoV-2IG-4	1514.5	292.9	1257.8	261.8	935.6	286.8	653.3	176.7	184.5	80.5
hCoV-2IG-5	1590.1	323.0	1295.7	341.2	1288.5	402.4	832.3	280.8	333.6	39.7
hCoV-2IG-6	1239.5	168.3	440.8	144.4	473.4	126.8	298.5	65.2	105.6	33.5

\*PsVNA titer Cut-off value: 1:20.

**Table S4: PRNT50 and PRNT90 of the six hCoV-2IG batches against SARS-CoV-2 WA-1, UK and SA strains in classical wild-type SARS-CoV-2 virus neutralization assay. Related to Figure 2.**

	WA-1		UK		SA	
	PRNT50	PRNT90	PRNT50	PRNT90	PRNT50	PRNT90
<b>hCoV-2IG-1</b>	5184.7	1723.8	2324.7	730.1	1111.4	296.1
<b>hCoV-2IG-2</b>	2260.6	264.4	1826.4	405.6	553.7	200.3
<b>hCoV-2IG-3</b>	4030	1504	1556.8	463.5	486.4	130.5
<b>hCoV-2IG-4</b>	3580.7	1153.8	1336.9	298.6	361	79.3
<b>hCoV-2IG-5</b>	1929.1	803.8	1674.6	370.8	716	194.3
<b>hCoV-2IG-6</b>	1988.8	663.7	708.1	50.9	96.8	4.3

**Table S5: Antibody concentration (in mg/mL) required for each of the six hCoV-2IG batches to achieve 50% (ID50) or 80% (ID80) neutralization of SARS-CoV-2 variants in PsVNA. Related to Figure 2.**

	WA-1		CA		UK		JP		SA	
	ID50 (mg/mL)	ID80 (mg/mL)	ID50 (mg/mL)	ID80 (mg/mL)	ID50 (mg/mL)	ID80 (mg/mL)	ID50 (mg/mL)	ID80 (mg/mL)	ID50 (mg/mL)	ID80 (mg/mL)
hCoV-2IG-1	0.035	0.194	0.050	0.163	0.056	0.249	0.149	0.372	0.262	0.647
hCoV-2IG-2	0.034	0.230	0.077	0.327	0.082	0.254	0.147	0.424	0.386	0.837
hCoV-2IG-3	0.030	0.169	0.048	0.202	0.051	0.200	0.120	0.331	0.360	0.806
hCoV-2IG-4	0.066	0.341	0.080	0.382	0.107	0.349	0.153	0.566	0.542	1.242
hCoV-2IG-5	0.063	0.310	0.077	0.293	0.078	0.249	0.120	0.356	0.300	2.519
hCoV-2IG-6	0.081	0.594	0.227	0.692	0.211	0.789	0.335	1.534	0.947	2.988