

	Patient age	Tumor position	Tumor size (mm)	Histological type	Tumour Grade	Nodal Status	Metastases	Chemotherapy	Recurrence	Other	HLA type		
											A*03, A*29	B*07, B*51	C*02, C*07
CRC1	82	anterior resection	110	Adenocarcinoma of rectum	3	0	x	-	N/A		A*03, A*29	B*07, B*51	C*02, C*07
CRC2	84	left hemicolectomy	115 x 120 x 80	Adenocarcinoma	3	0	x	-	-		A*02, A*23	B*44	C*04, C*05
CRC3	57	right hemicolectomy	70 x 60	Adenocarcinoma of caecum	2B	0	x	neoadjuvant 5FU and cisplatin	-		A*02, A*31	B*40, B*45	C*03, C*06
CRC4	78	right hemicolectomy	45 x 30 x 20	Adenocarcinoma of caecum	4	1	x	adjuvant Capecitabine and Oxaliplatin	-		A*02, A*03	B*07, B*14:02	C*07, C*08
CRC5	73	right hemicolectomy	80 x 130 x 35	Adenocarcinoma of caecum	4	1	x	adjuvant Capecitabine and Oxaliplatin	-	K-RAS mutation	A*02	B*35, B*50:01	C*04, C*06
CRC6	61	left hemicolectomy	65 x 40 x 13	Adenocarcinoma of sigmoid	3	0	x	-	+		A*02, A*03	B*35, B*58	C*04, C*07

SUPPLEMENTARY TABLE S1 | Clinical data from patients of primary CRC samples used

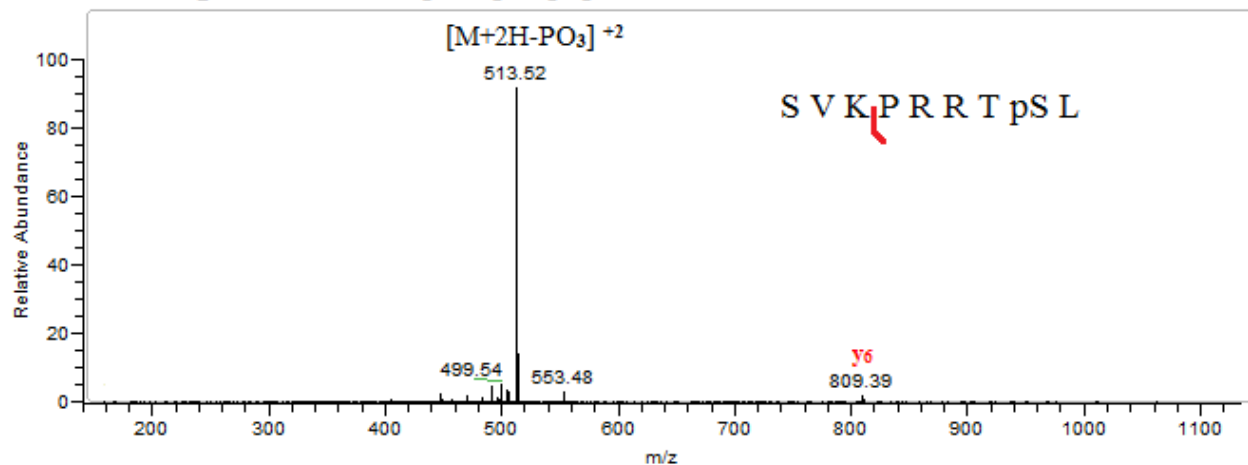
Patient samples were obtained at colorectal cancer resection. This table shows the clinical data for the three primary colorectal cancer specimens used in phosphopeptide discovery and the six used for extraction of tumor infiltrating lymphocytes for immunological assays.

	Tumor sample	Healthy tissue	Patient age	Primary tumor			Time to progression	Other malignancies	HLA type		
				Time since resection	Position	Dukes stage			A*02, A*03	B*07	C*07
CRCLM1	Liver metastasis	Liver	69	5 and 2 years	Rectal	B	6 weeks	CLL	A*02, A*03	B*07	C*07
CRCLM2	Liver metastasis	Liver	70	1 year	Rectal	B	N/A	-	A*02, A*24	B*08, B*27	C*07

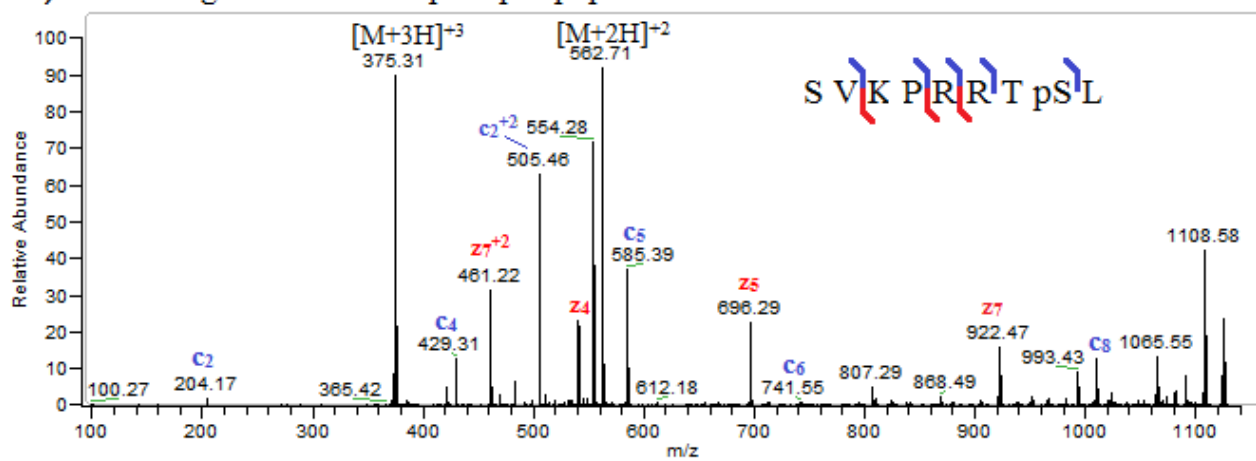
SUPPLEMENTARY TABLE S2 | Clinical data from patients of CRC liver metastasis samples used

Patient samples were obtained at resection of CRC liver metastases. This table shows the clinical data for the two colorectal cancer liver metastases used in phosphopeptide discovery and for extraction of tumor infiltrating lymphocytes for immunological assays.

A) CAD Fragmentation of a phosphopeptide



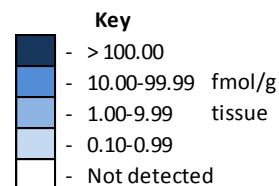
B) ETD Fragmentation of a phosphopeptide



SUPPLEMENTARY FIGURE S1 | Fragmentation of phosphopeptides

(A) CAD fragmentation results in the domination of a spectrum by the neutral loss of phosphoric acid, which can be used as a diagnostic in “CAD Neutral Loss Finder.” (B) ETD fragmentation preserves the post-translational modification, which allows us to sequence the phosphopeptide.

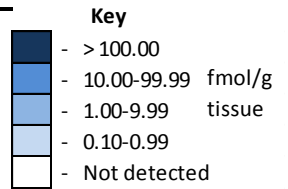
HLA binding	Phosphopeptide	Uniprot #	Protein Name	CRC1		CRC2		CRC3		LM1		LM2		colo205	sw620	hct116
				Healthy	Tumour	Healthy	Tumour	Healthy	Tumour	Healthy	Tumour	Healthy	Tumour			
A*01	ITQGtPLKY	Q9Y618	NCOR2													
	ISSsMHSly	P50616	TOB1													
	LTDPsPTISSY	Q8IX90	SKA3													
	NTDsPLRY	P08865	RPSAP19													
	TMAsgPKDNY	O60684	KPNA6													
A*02	RQIsQDVKL	Q01433	AMPD2													
	RVAsPTSGV	Q9Y4H2	IRS2													
	RQAsIELPSMAV	P33241	LSP1													
	ALDsGASLLHL	P57078	RIPK4													
	RLAsYLDRV	P05783	KRT18P19													
	KLIDRTEsL	P33241	LSP1													
	RIshELDS	P10451	SPP1													
	LLsEEVEL	Q8IY92	BTBD12													
	VMIGsPKKV	Q68CZ2	TNS3													
	SMTRsPPRV	Q9BRL6	SFRS2B													
	KLAsPELERL	P05412	JUN													
	KLIDIVsSQKV	O14757	CHEK1													
	FLDtPIAKV	Q969G9	NKD1													
	RTLsPEIITV	Q9H4A3	WNK1													
	RVLHsPPAV	Q9Y4B5	MTCL1													
	RLSsPLHFV	Q8NC44	FAM134A													
	VmIGsPKKV	Q68CZ2	TNS3													
	RTHsLLLLL	P34096	RNASE4													
RQAsIELPSM	P33241	LSP1														
A*03	RIYQyIQSR	Q9Y463	DYRK1B													
	RILsGVVTK	P62280	RPS11													
	GImsPLAKK	Q03989	ARID5A													
	RLSsPISKR	Q99728	BARD1													
	RVAsPTSGVK	Q9Y4H2	IRS2													
	RTRsLSSLREK	O94915	FRYL													
	RTMsEAALVRK	Q6ZTQ3	RASSF6													
	SVRRsVLmK	Q9H2J4	PDCL3													
	KLPsPAPARK	Q8IY33	MICALL2													
RVLsPLIIK	Q8NCN4	RNF169														
A*24	RFKtQPVTf	Q7Z7L8	AG2													
	RYQtQPVTl	O95425	SVIL													
A*29	RIYQyIQSRf	Q9Y463	DYRK1B													



SUPPLEMENTARY TABLE S3 | Phosphopeptides identified on CRC patient samples that are predicted to bind to HLA-A

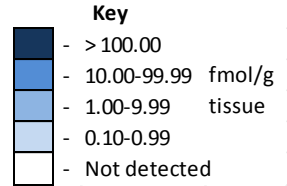
MHC class I-bound peptides were eluted from tumor tissue and its healthy counterpart, or cultured cell lines. Phosphopeptides were IMAC enriched and characterized using LC-MS/MS. MHC class-I binding was predicted, using known HLA-alleles. Here 3 primary CRC, 2 CRC liver metastases and 3 cell line samples were analyzed.

HLA binding	Phosphopeptide	Uniprot #	Protein Name	CRC1		CRC2		CRC3		LM1		LM2		colo205	sw620	hct116
				Healthy	Tumour	Healthy	Tumour	Healthy	Tumour	Healthy	Tumour	Healthy	Tumour			
B*07	LPFISRLsI	P47974	ZFP36L2													
	RPKtPPVVI	Q96A49	SYAP1													
	SPFKRQLsL	B7Z5W0	NUMBL													
	RPRsPNMQDL	Q6T310	RASL11A													
	LPVsPRLQL	P13688	CEACAM1													
	AVRPTRLsL	Q9Y4H2	IRS2													
	SPRsPDRTL	Q9UKN1	MUC12													
	QPQRRLsL	Q9ULW0	TPX2													
	VPRPERRsSL	Q6UWJ1	TMCO3													
	KPEsRRSLL	Q6WKZ4	RAB11FIP1													
	APRKGsFSAL	Q13619	CUL4A													
	RPTKIGRRsL	Q96HN2	AHCYL2													
	RPDVAKRLsL	O75815	BCAR3													
	RPFHGISTVsL	Q5VZ89	DENND4C													
	RPVtPVSDL	Q13118	KLF10													
	RPGsRQAGL	Q96JY6	PDLIM2													
	RPDsAHKML	Q8WX93	PALLD													
	RPRARsVDAL	Q86X29	LSR													
	RARGIsPIVF	Q96MU7	YTHDC1													
	APDsPRAFL	unknown	UNKNOWN													
	SPRSPsTTYL or SPR	Q13111	CHAF1A													
	RPFsPREAL	Q86V48	LUZP1													
	RPRPVsPSSL	P57059	SIK1													
KPAsPKFIVTL	Q6PJT7	ZC3H14														
TPRsPPLGL	Q16584	MAP3K11														
RPRsPTGPSNSF	Q96I25	RBM17														
RPVsPFQEL	unknown	UNKNOWN														
KPRsPVVEL	P25098	ADRBK1														
RPRGsQSL	P21860	ERBB3														
RPAAsPQRAQL	unknown	UNKNOWN														
B*08	DLKRRsMSI	Q96N67	DOCK7													
	SVKPRRTsL	P15822	HIVEP1													
	INKERRSsL	Q5JTZ5	C9ORF152													
B*15	RQDsTPGKVFL or F	P13056	NR2C1													
B*27	GRLsPAYSL	Q86UU1	PHLDB1													
	GRLsPVPVPR	Q9UKM9	RALY													
	RRFsRLENRY	O43293	DAPK3													
	RRDsLQKPGL	Q9NRM7	LATS2													
	RRAsQEANL	Q6PJG2	C14ORF43													
RRLsLFLNV	Q99836	MYD88														
B*27/C*07	KRFsFKKsF	P29966	MARCKS													
	RRLsDSPVF	P47974	ZFP36L2													
	KRYsGNMEY	O95835	LATS1													
	RRMsLLSVV	Q9ULI2	RIMKLB													
	RRSsFLQVF	Q15436	SEC23A													
	RRSsIQSTF or RRsS	Q92542	NCSTN													
	RRLsESSAL	Q96555	WRNIP1													
RRNsINRNF	O00160	MYO1F														
B*44	AENsPTRQQF	Q86XP3	DDX42													
	KEMsPTRQL	Q4G0N7	C6ORF225													
	AI sDLQQL	O15302	CAMK2													
B*45	EERsPPAP	P15408	FOSL2													
	EERsPSWISA	Q02952	AKAP12													
	EERs ^{ts} WISA	Q02952	AKAP12													
	EERsPSWISA	Q02952	AKAP12													
	REIsSSPTS	Q9UQ35	SRRM2													
	AEAPPSKsP	Q96D71	REPS1													
	AEKsYQNSP	Q15648	MED1													
SEAsPSREA	Q13111	CHAF1A														
B*51	FEDDDsNEKL	O43719	HTATSF1													

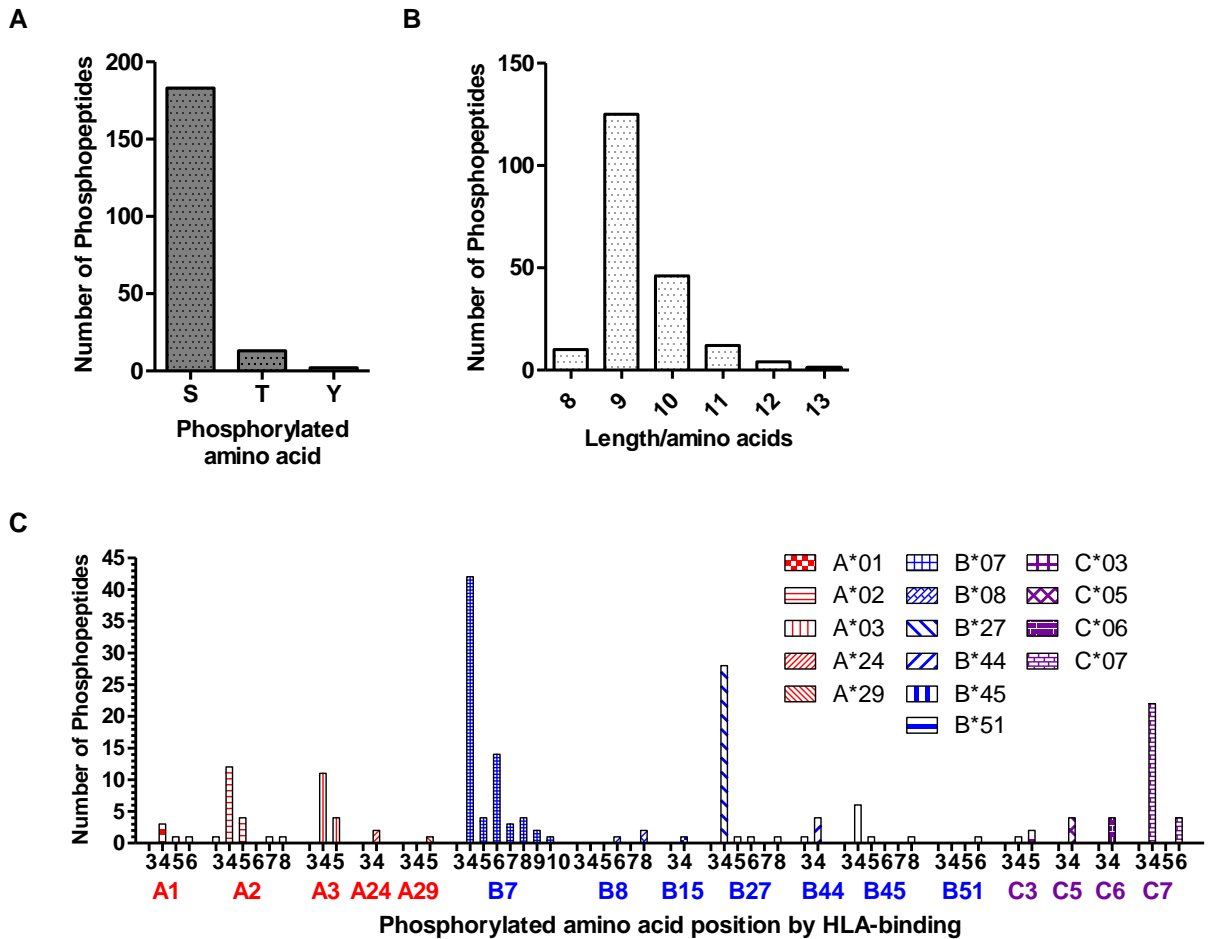


SUPPLEMENTARY TABLE S4 | Phosphopeptides identified on CRC patient samples that are predicted to bind to HLA-B

HLA binding	Phosphopeptide	Uniprot #	Protein Name	CRC1		CRC2		CRC3		LM1		LM2		colo205	sw620	hct116
				Healthy	Tumour	Healthy	Tumour	Healthy	Tumour	Healthy	Tumour	Healthy	Tumour			
C*03	KAFsPVRSV	Q02363	ID2													
	RAHSsPASL	P46937	YAP1													
	RSHsSPASL	Q9GZV5	WWTR1													
C*05	RADsPVHVM	O95402	MED26													
	RRDsIVAEL	O14579	COPE													
	SIDsPQKL	Q12888	TP53BP1													
	RSDsYVEL	P52298	NCBP2													
C*06	RRSsSVAQV	O15205	UBD													
	RRPsLLSEF	O75376	NCOR1													
	RRNsAPVSV	Q2M123	ARHGAP31													
C*07	RRGsFEVTL	Q8IZQ5	SELH													
	RRLsFLVSY	P47897	QARS													
	RKLSVILIL	Q13433	SLC39A6													
	KRFsGTVRL	P62906	RPL10AP9													
	RRSsQSWSL	Q9Y4E1	FAM21C													
	HRNsMKVFL	Q9NPR2	SEMA4B													
	RRKsQVAEL	Q9BYG3	MKI67IP													
	KRLsVERIY	P11388	TOP2A													
	RRLsGPLHTL	Q86Y91	KIF18B													
	KLFPDtPLAL	Q12906	ILF3													
	KYIsGPHEL	P49454	CENPF													
	RRFsGTAVY	Q6AHZ1	ZNF518A													
	MPRQPsATRL	Q6NZ67	FAM128B													
C*07/B*27	HRLsPVKGEF	Q9Y2L9	LRCH1													
	RRIDIsPSTLR	Q9NYF8	BCLAF1													
	RRFsPPRRM	Q15287	LOC643446													
	RRIsGVDRYY	O15239	NDUFA1													
	KRMSPKPEL	P41208	CETN2													
C*06/C*07	RRIsDPQVF	Q4L180	FILIP1L													

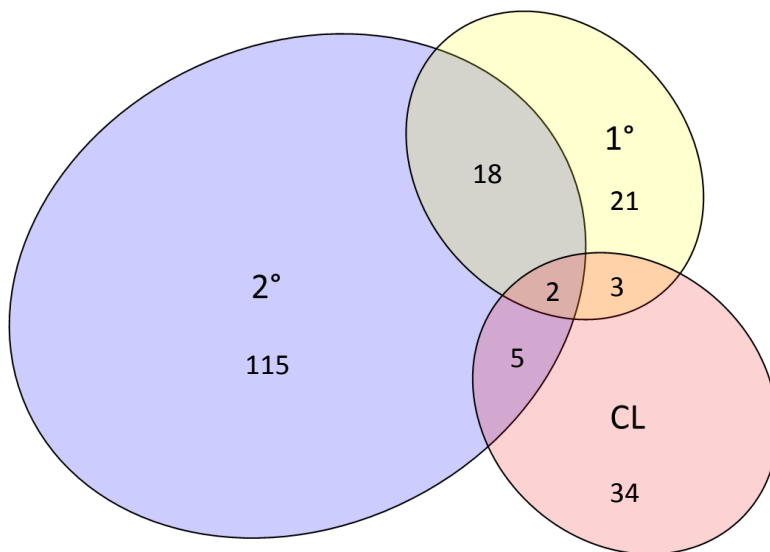


SUPPLEMENTARY TABLE S5 | Phosphopeptides identified on CRC patient samples that are predicted to bind to HLA-C



SUPPLEMENTARY FIGURE S2 | Phosphopeptide characteristics

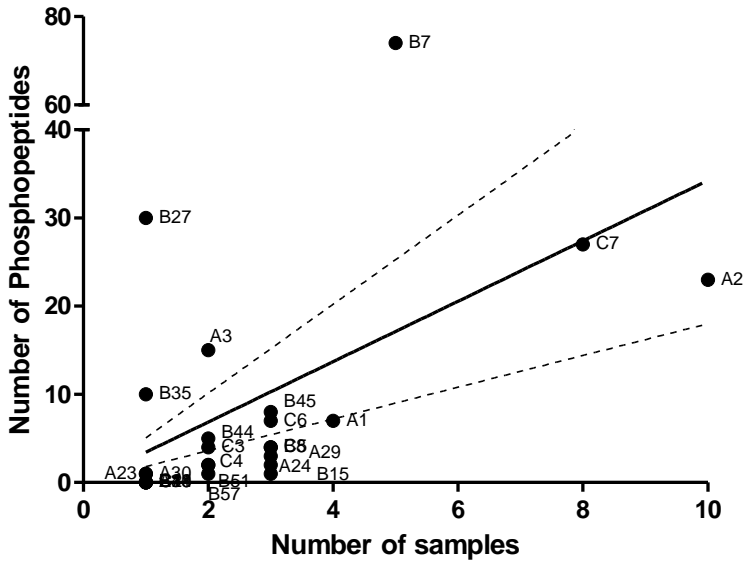
- (A) The number of CRC phosphopeptides identified containing phospho-serine (S), phospho-threonine (T) and phospho-tyrosine (Y).
- (B) The lengths of the CRC phosphopeptides identified.
- (C) The position of the phosphorylated amino acid in the phosphopeptides, according to HLA-binding.



SUPPLEMENTARY FIGURE S3 | How the phosphopeptides identified are shared between samples

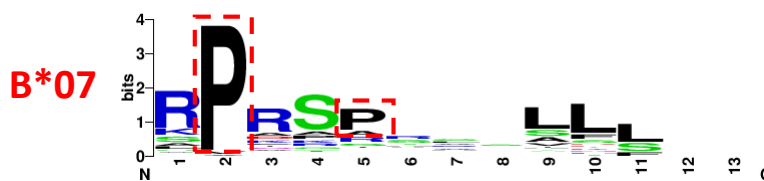
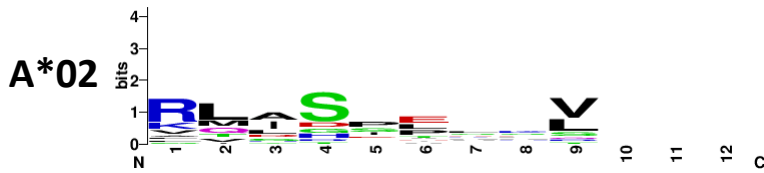
A Euler plot showing the overlap of presentation of phosphopeptides between differing types of CRC samples: - primary CRC – 1°; secondary CRC liver metastases – 2°; and cell lines –CL.

A

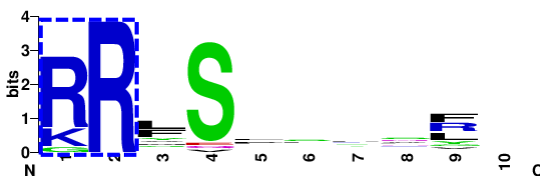


B

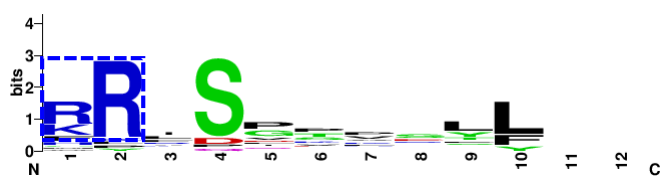
Kinase name	Consensus sequence	# phosph. sites
PKACa (PRKACA)	xxrxRRlSlxxxxxx	734
PKCa (PRKCA)	xxxxRRxSfKrkxxx	523
CK2a1 (CSNK2A1)	xxxeedSDdEeeee	483
ERK2 (MAPK1)	xpxpPlSPtppxxx	410
CDK1 (CDC2)	xxxxlpXSPxkkxxx	393
SRC	xxeedvYgxvxxxx	385
ERK1	xpppPlSPtptxxx	292
CDK2	xxxxpxSPgKkxlx	201
PDK1 (PDPK1)	xxgxtTxTFCGTpeY	43



B*27



C*07



SUPPLEMENTARY FIGURE S4 | Phosphopeptides binding motifs vs. kinase binding motifs

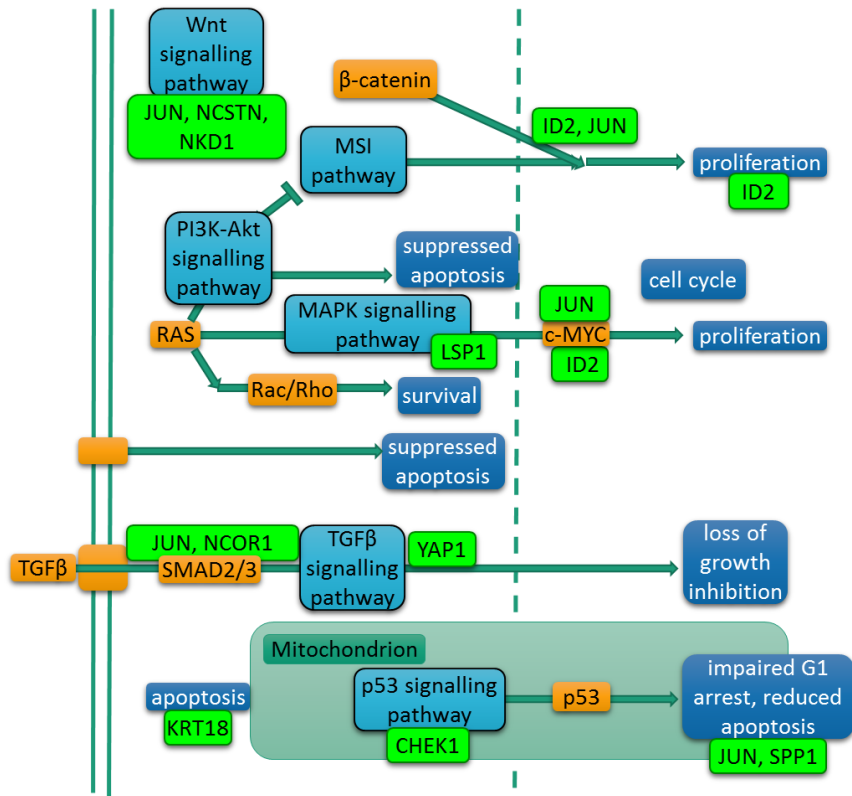
- (A) The number of phosphopeptides identified, plotted against the number of samples of each HLA-type. HLA-B*07 and HLA-B*27 are shown to be outliers, with many more phosphopeptides per sample than the general trend.
- (B) Published kinase binding motifs are compared to logoplots of the phosphopeptide sequences identified on CRC, showing key consensus sequences around the phosphoserine are shared for HLA-B*07 with ERK1 and ERK2; and for HLA-B*27 and HLA-C*07 with PKACa and PKCa.

	Phospho-peptide	Uniprot #	Source protein	Malignant Samples		
				CRC	Leukemia	Melanoma
A*02	RVAsPTSGV	Q9Y4H2	IRS2	colo205, hct116	CLL1, CLL2, MCL	DM331, SLM2, COV413
	VMIGsPKKV	Q68CZ2	TNS3	sw620	MCL	DM331, SLM2, COV413
	VmIGsPKKV	Q68CZ2	TNS3	sw620	MCL	DM331, SLM2, COV413
B*07	GPRsAsLLSL	Q9Y4H4	GPSM3	CRCLM1	AML1	
	RPFsPREAL	Q86V48	LUZP1	CRC1, CRCLM1	AML1, ALL1, CLL2, CLL4, HCL1, B-LCL	
	RPRPVsPSSL	P57059	SIK1	CRC1, CRCLM1	AML1	
	RPRsPRQNSI	Q99700	ATXN2	CRCLM1	AML1, B-LCL	
	RPVsPFQEL	unknown	unknown	CRC1, CRCLM1	AML1, ALL1, CLL2, CLL4, HCL1, B-LCL	
	TPRsPPLGL	Q16584	MAP3K11	CRC1, CRCLM1	AML1, CLL2, CLL4, HCL1, B-LCL	

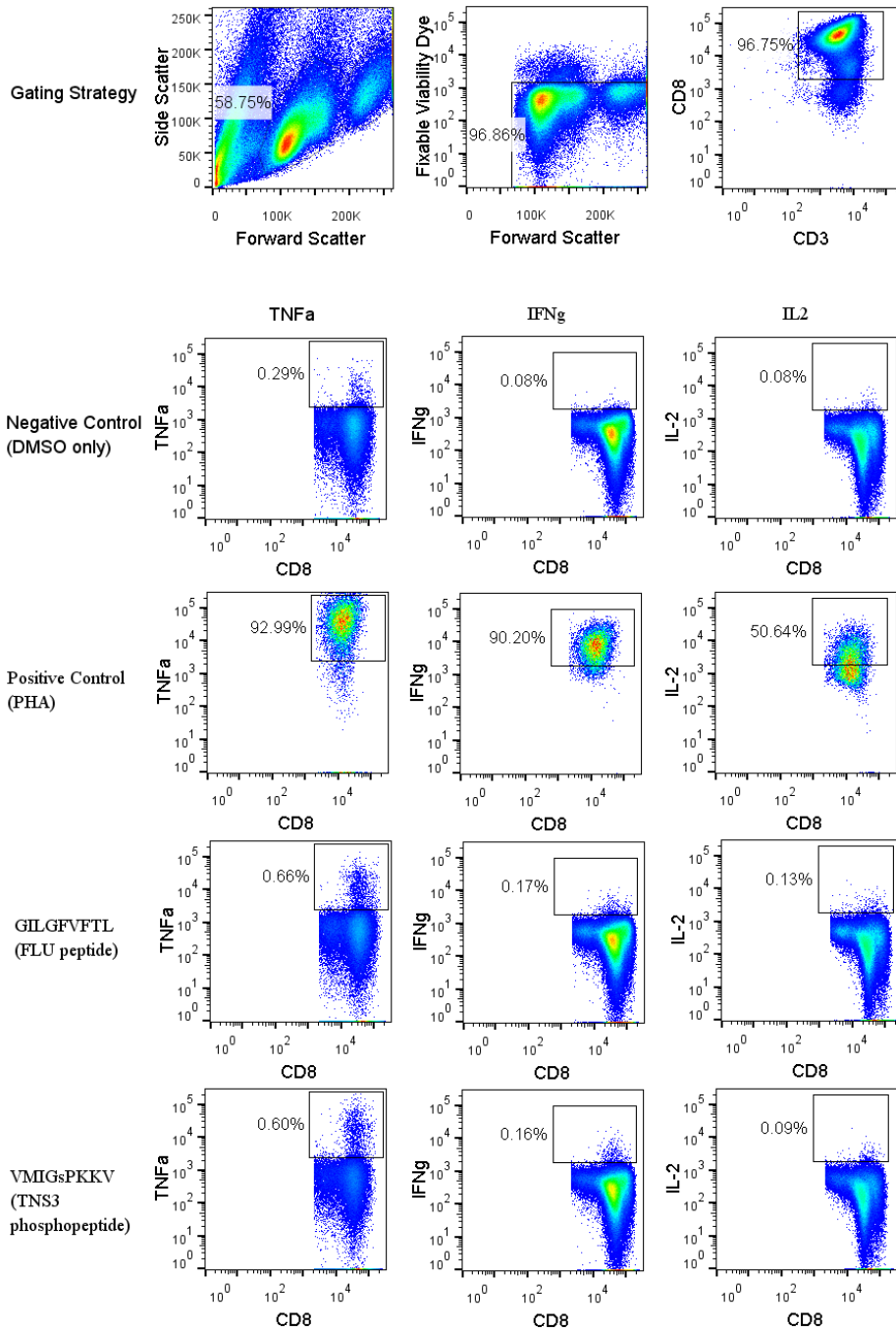
SUPPLEMENTARY TABLE S6 | Phosphopeptides shared across different malignancies have been identified in multiple samples

Key

- signalling pathway
- phosphopeptide source protein
- result
- key protein

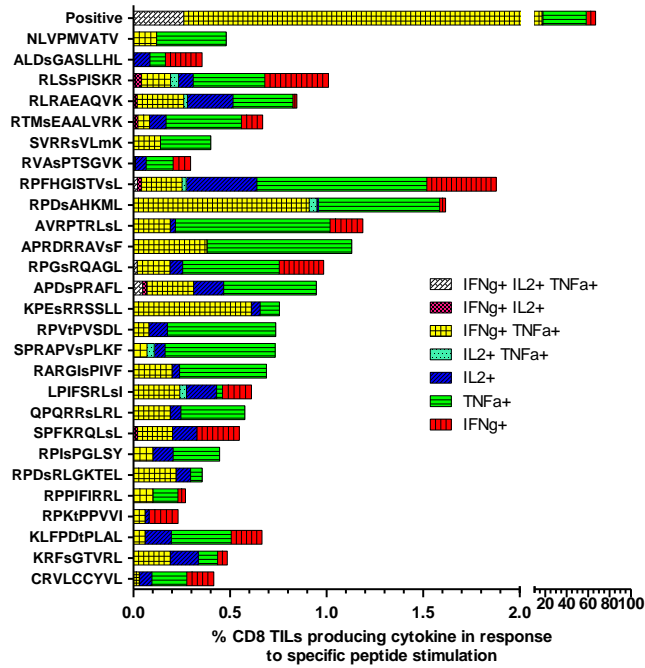


SUPPLEMENTARY FIGURE S5 | The source proteins of phosphopeptides identified superimposed onto the KEGG CRC signaling pathways

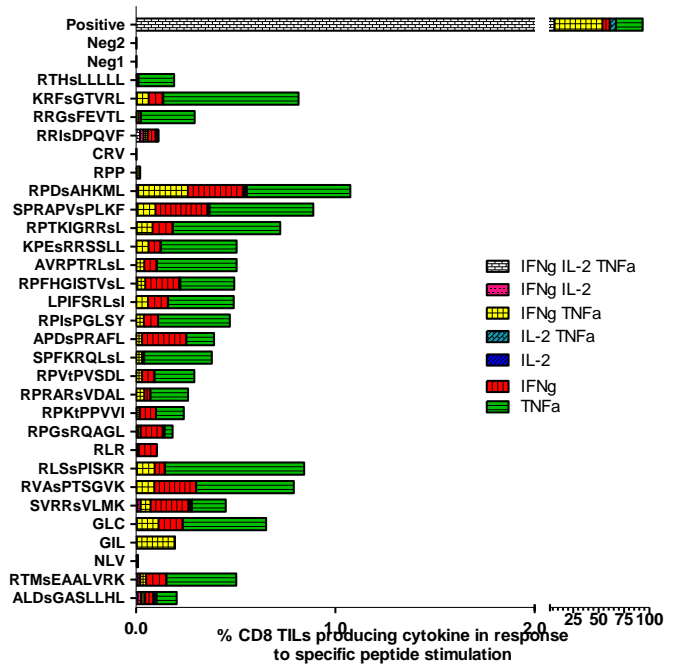


SUPPLEMENTARY FIGURE S6 | Batch analysis of Intracellular Cytokine Staining (ICS) used to assess TIL targeting of phosphopeptides

A

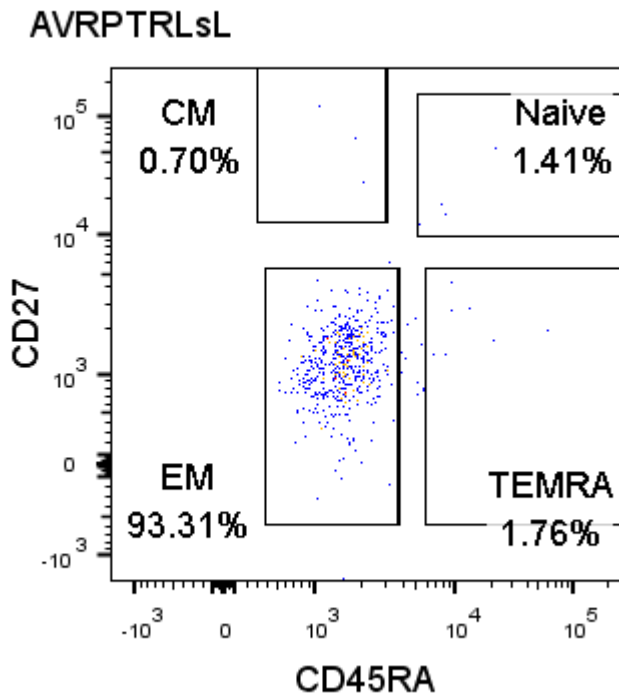


B



SUPPLEMENTARY FIGURE S7 | Raw data for the two different CRCLM1 ICS assay repeats

(A) After one REP cycle of TILs and (B) after a second REP cycle.



SUPPLEMENTARY FIGURE S8 | An example of memory phenotype of TIL cultures targeting CRC-associated phosphopeptides

CD45RA and CD27 staining was used to gate central memory (CM), effector memory (EM), naïve and terminal effector memory (TEMRA) phenotypes.

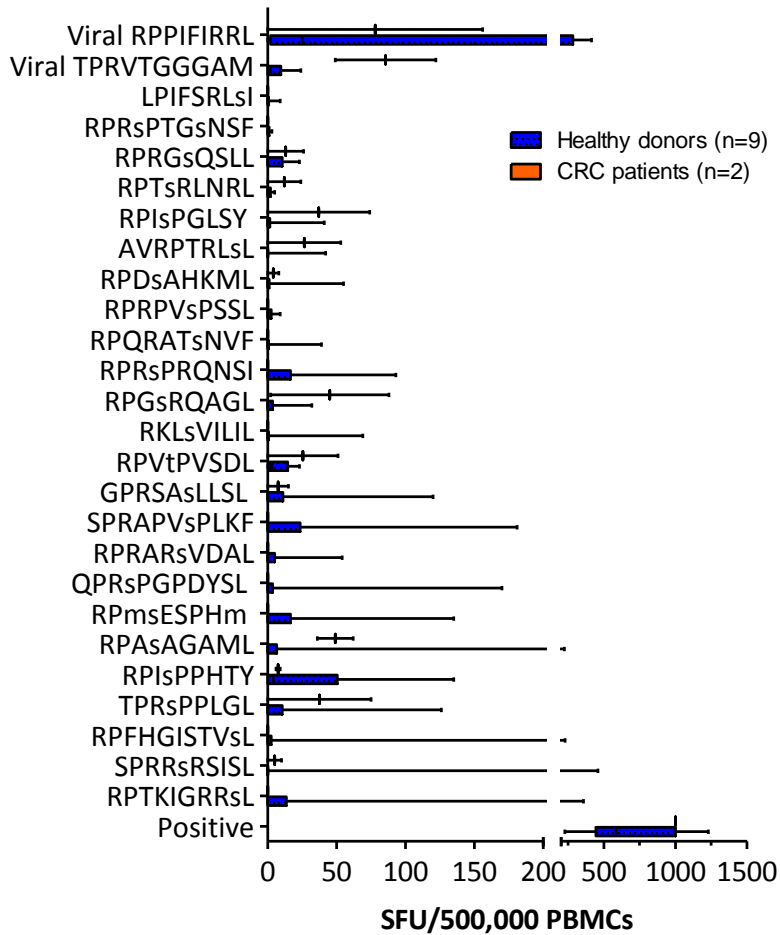
CRC #	Patient age (at time of sample)	Histology	Tumour size (mm)	Surgery type	Tumour Grade	Nodal Status	Metastases	Neo-Adjuvant RT
22	68	Adenocarcinoma	20 x 25 x 10	Right hemicolectomy	1	0	X	NIL
23	74	Adenocarcinoma	40 x 83 x 15	Right hemicolectomy	3	0	x	NIL
25	48	Mucinous adenocarcinoma	65 x 85 x 12	Subtotal colectomy	3	0	x	NIL
26	74	Adenocarcinoma of rectum	30 x 20 x 6	Anastamotic doughnut	3	0	x	25 Gray in 5 fractions
27	66	Adenocarcinoma	60 x 50	Left colon distal doughnut	2	0	x	NIL

SUPPLEMENTARY TABLE S7 | Clinical data of CRC patients whose PBMCs were used in the ELISpot assay



SUPPLEMENTARY FIGURE S10 | IFN γ ELISpot was used to assess *ex vivo* T cell responses to phosphopeptides from healthy donor and CRC patient PBMCs

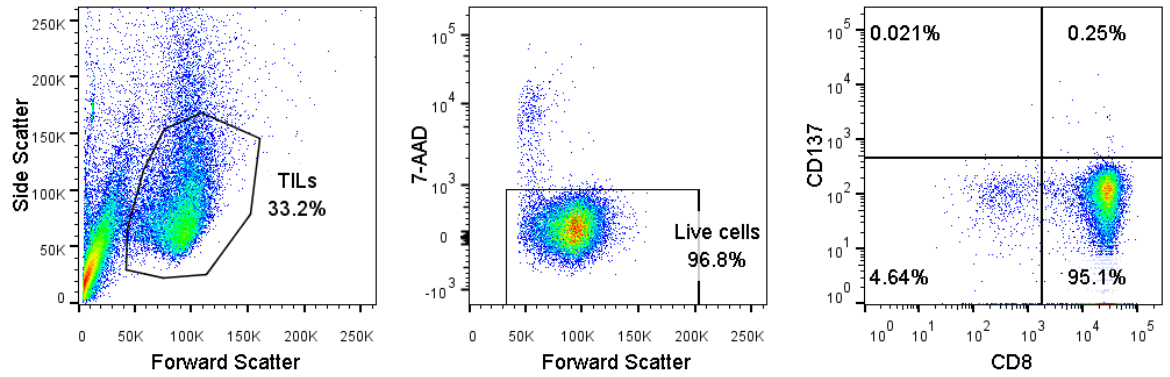
Blood was collected from healthy donors and CRC patients, the PBMCs extracted and cultured for 6-days in the presence of phosphopeptide. The cells were then transferred to a prepared IFN γ ELISpot plate and restimulated with phosphopeptide overnight.



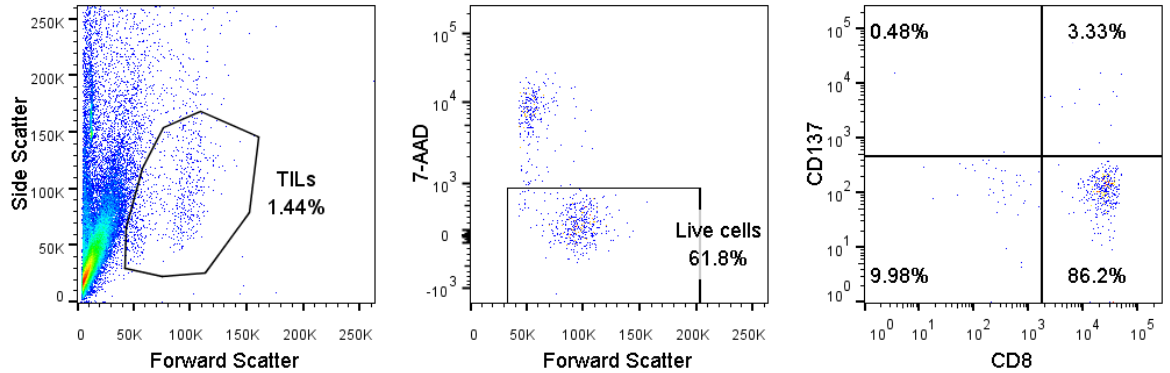
SUPPLEMENTARY FIGURE S11 | T cells targeting the tumor-associated phosphopeptides are found in a number of patients

Comparison of HD and CRC patient PBMC IFN γ production targeting HLA-B*07-associated phosphopeptides. Too few HLA-B*07+ patient samples were obtained for meaningful results, as it is a lower frequency allele, yet there were some responses that were higher in patients than healthy donors.

CRC14 VMI line pre-selection

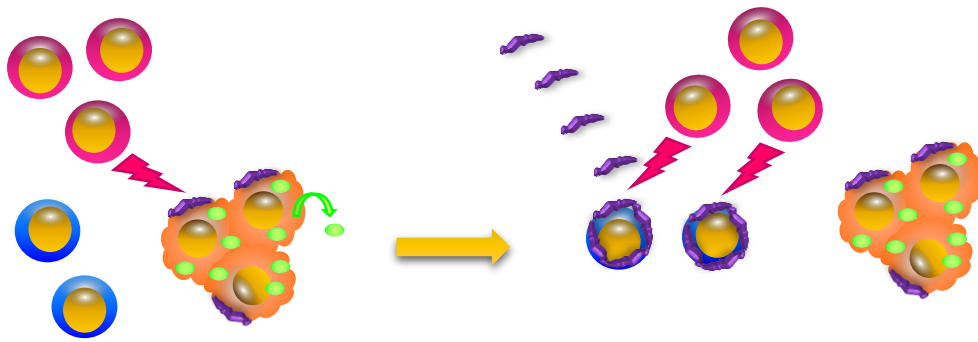


CRC14 VMI line post CD137 MACS selection



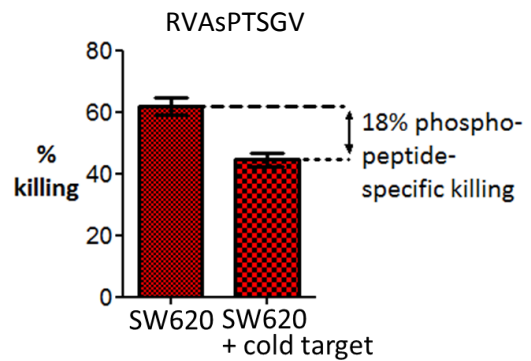
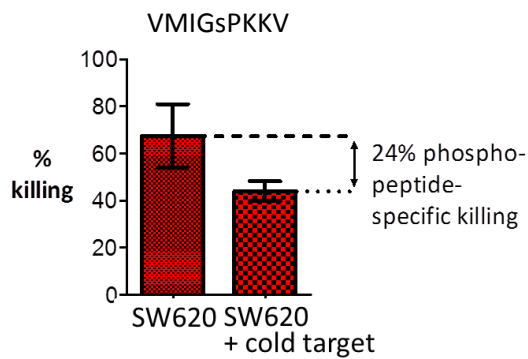
SUPPLEMENTARY FIGURE S12 | CD137 MACS sorting of CRC14

CD8⁺ TILs were plated at 1E6/ml in TIL medium and stimulated with 10 µg/ml of the VMI (TNS3) phosphopeptide. On days 7 and 10 TILs were adjusted to 5E5/ml and half of the medium exchanged. On day 14 phosphopeptide-specific TILs were selected, using CD137 MACS (Miltenyi biotec). Prior to selection, half of the cells were peptide pulsed for 2 hours, washed and added to the other half overnight. These were then rapidly expanded, as described previously.



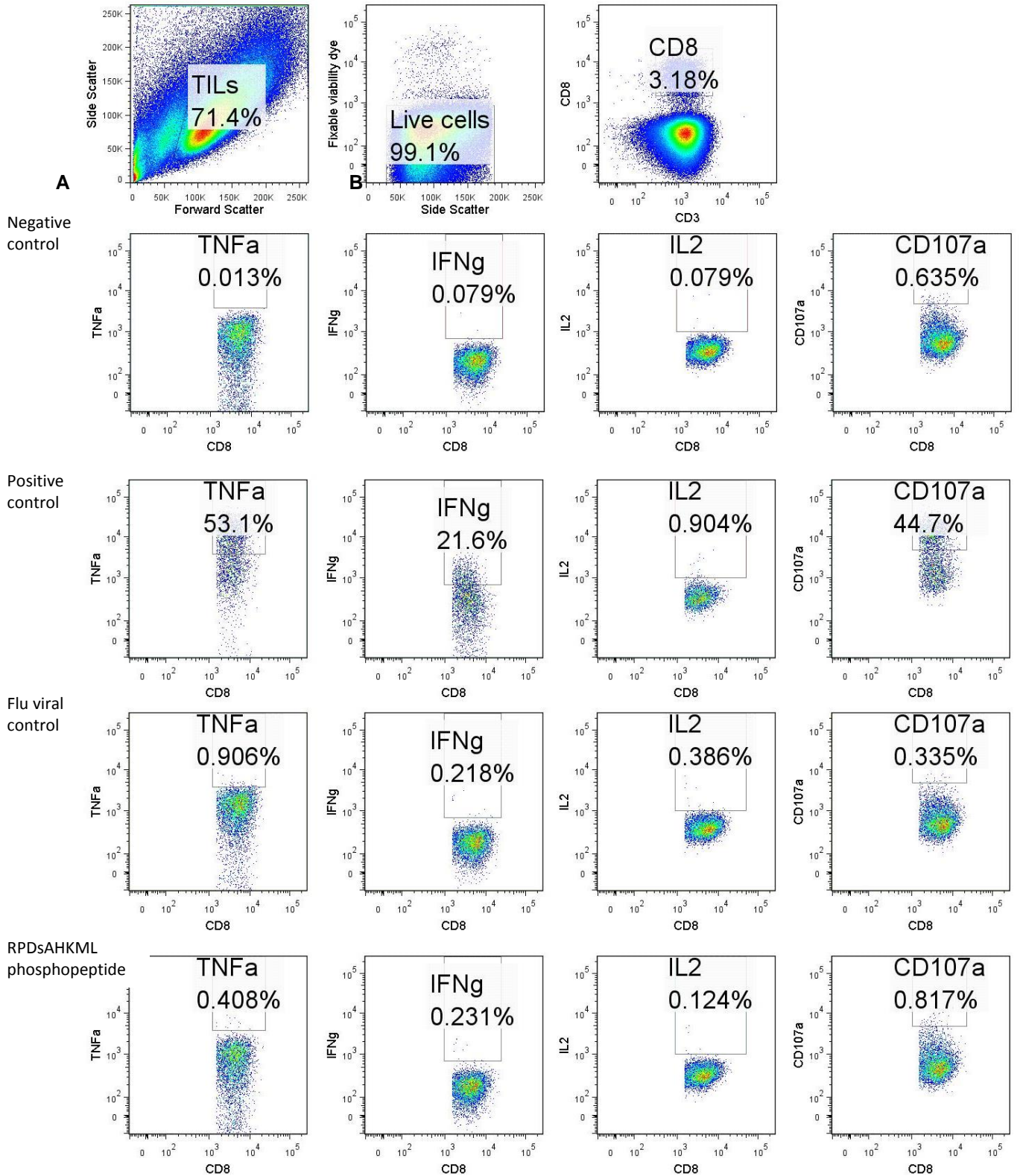
T cells recognise phosphopeptide and kill “hot target” CRC cells, releasing ligand. Autologous B cells (self) are not targeted.

“Cold target” - autologous B cells - pulsed with excess phosphopeptide - act as a decoy and **inhibit** T cell recognition of CRC cells. Ligand is not released.



SUPPLEMENTARY FIGURE S13 | Cold-target inhibition to quantify phosphopeptide-specific killing of CRC cell lines

Healthy donor PBMCs were used to establish T cell lines targeting two phosphopeptides – VMIGsPKKV and RVAsPTSGV. After selection and rapid expansion, the lines were used in Europium release killing assays to assess killing of Sw620 cells, which natively express those phosphopeptides. Cold-target inhibition was used to quantify the killing that was targeting the phosphopeptides, as depicted.



SUPPLEMENTARY FIGURE S14 | Degranulation marker CD107a is upregulated in response to phosphopeptide RPDsAHKML in young CRCLM1 TILs, but not bystander T cells targeting a peptide from influenza.