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Supplemental information

**A molecular sensor determines
the ubiquitin substrate specificity
of SARS-CoV-2 papain-like protease**

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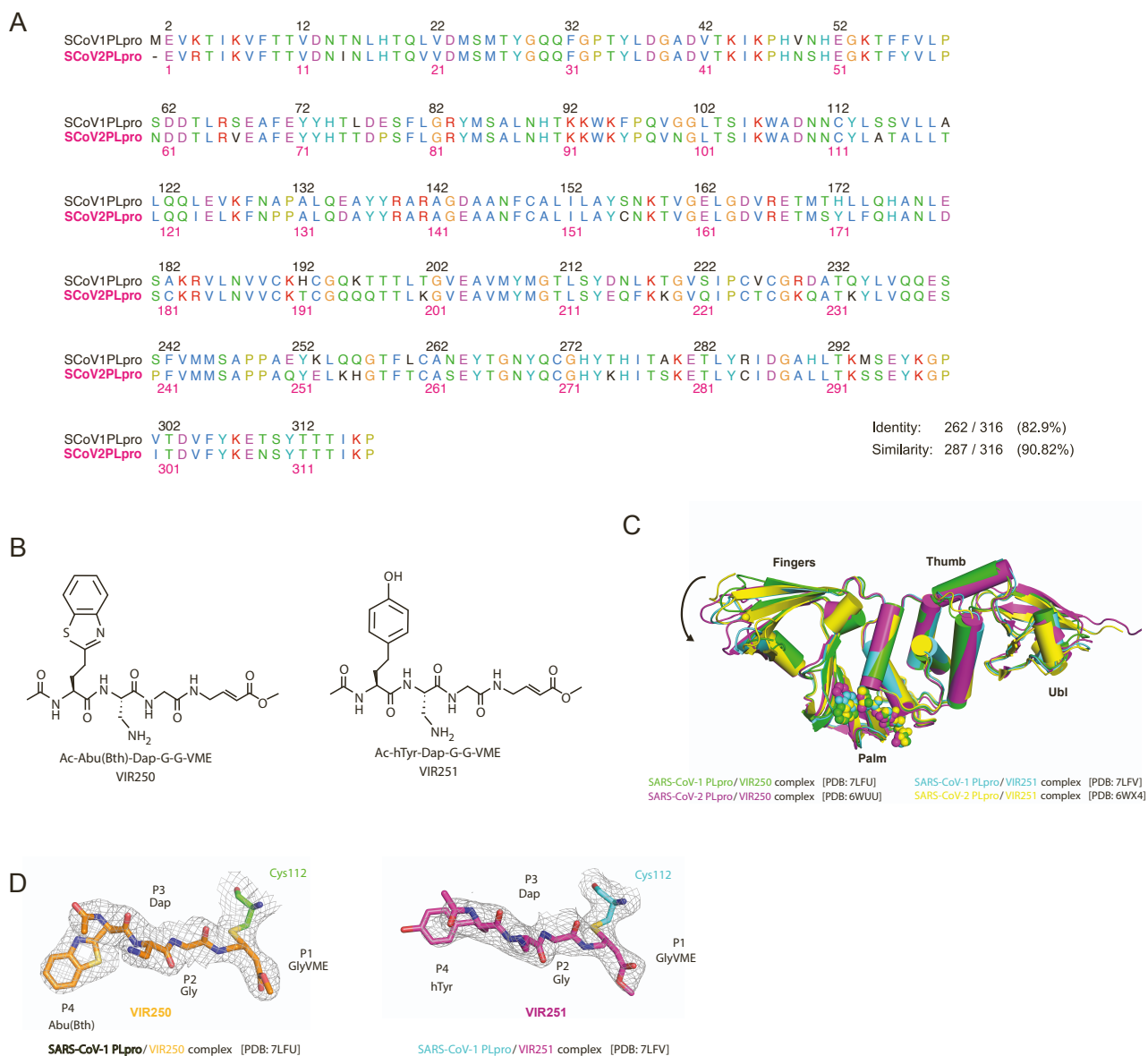


Figure S1. PLpro alignments and inhibitor schematics, Related to Figure 1. **A)** Schematic of VIR250 and VIR251 inhibitors. **B)** Crystal structures of SCoV-1 and SCoV-2 PLpro with VIR250 and VIR251 aligned. VIR250 and VIR251 shown as spheres, arrow highlights slight rotation of the fingers domain. **C)** Sequence alignment of PLpro domain from SARS-CoV-1 and SARS-CoV-2, aligned using Local alignment (Smith-Waterman). SCoV-1 PLpro amino acid numbering is shown above the alignment (black) and SCoV-2 numbering is shown below the alignment (magenta). **D)** 2FoFc electron density maps of VIR250 (left) and VIR251 (right) contoured at 1.0σ and shown as mesh.

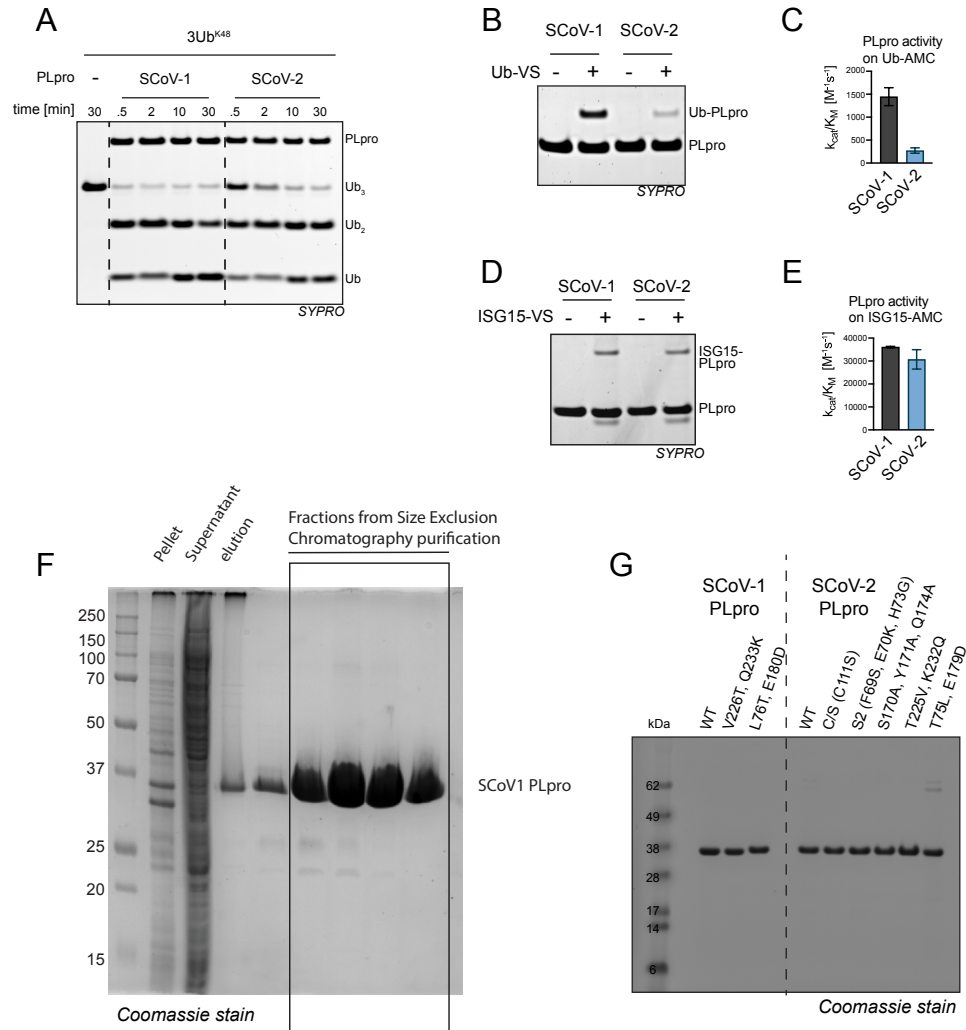


Figure S2. Comparison of SCoV-1 and SCoV-2 PLpro activity, Related to Figures 2, 3 and STAR Methods. **A)** Cleavage of K48-linked tetra-Ub chains (3UbK48) by SARS-CoV-1 vs. SARS-CoV-2 PLpro. **B)** Ub-Vinyl Sulfone (Ub-VS) labeling of SARS-CoV-1 vs. SARS-CoV-2 PLpro. **C)** Relative apparent k_{cat}/K_M values for hydrolysis of Ub-AMC by SARS-CoV-1 and SARS-CoV-2 PLpro. **D)** ISG15-VS labeling of SARS-CoV-1 vs. SARS-CoV-2 PLpro. Experiments in **A**, **B**, **D** were repeated at least three times with similar results. **E)** Relative apparent k_{cat}/K_M values for hydrolysis of ISG15-AMC by SARS-CoV-1 and SARS-CoV-2 PLpro. Error bars in **C**, **E** represent SD ($n = 3$ independent experiments). **F)** Last step of SCoV-1 PLpro purification on size exclusion chromatography, analyzed by SDS-PAGE and Coomassie staining. The pure fractions were pooled and concentrated. **G)** Purified SCoV-1 and SCoV-2 PLpro and mutants (2 μ M) analyzed by SDS-PAGE and Coomassie staining.

Table S1. Crystallographic Data and Refinement Statistics, Related to Figure 1.

	SARS-CoV-1 PLpro/VIR250 complex	SARS-CoV-1 PLpro/VIR251 complex
PDB ID	7LFU	7LFV
Source	APS 24 IDE	APS 24 IDE
Wavelength (Å)	1.00	1.00
Resolution Limits (Å)	70.7-2.29 (2.37-2.29)	132.7-2.23 (2.30-2.23)
Space Group	I222	P6 ₁ 22
Unit Cell (Å) <i>a</i> , <i>b</i> , <i>c</i>	70.8, 90.4, 113.6	103.6, 103.6, 265.4
Unit Cell (°) α , β , γ	90, 90, 90	90, 90, 120
Number of observations	108176	1632487
Number of reflections	16767 (1626)	42208 (3796)
Completeness (%)	100 (100)	99.9 (99.3)
Mean <i>I</i> / σ	16.0 (0.7)	30.9 (2.0)
CC _{1/2}	0.9986 (0.408)	1.00 (0.756)
R _{merge} ^a	0.074 (2.15)	0.099 (2.04)
R _{pim}	0.032 (0.962)	0.016 (0.341)
Refinement Statistics		
Resolution Limits (Å)	70.7-2.29 (2.36-2.29)	53.3-2.23 (2.28-2.23)
# of reflections (work/free)	16663 (1668)	42096 (2000)
Completeness (%)	99.3 (93.0)	99.9 (99.0)
Protein/solvent/ligand atoms	2414/0/36	5009/156/35
R _{cryst} ^b	0.224 (0.360)	0.197 (0.297)
R _{free}	0.265 (0.380)	0.235 (0.352)
Bonds (Å)/ Angles (°)	0.003/0.499	0.009/1.078
B-factors: protein/solvent/ligand (Å ²)	91.5/--/84.2	64.8/62.9/79.4
Ramachandran plot statistics (%)		
favored	96.3	96.8
allowed	3.7	3.2
outliers	0	0
MolProbity score	1.81- 96 th percentile (N=8848, 2.29 Å ± 0.25Å)	1.64- 97 th percentile (N=10112, 2.23 Å ± 0.25Å)

Parentheses indicate statistics for the high-resolution data bin for x-ray data.

a. $R_{merge} = \frac{\sum hkl \sum i |I(hkl)_i - \langle I(hkl) \rangle|}{\sum hkl \sum i \langle I(hkl)_i \rangle}$.

b. $R_{cryst} = \frac{\sum hkl |F_o(hkl) - F_c(hkl)|}{\sum hkl |F_o(hkl)|}$, where *F_o* and *F_c* are observed and calculated structure factors, respectively.