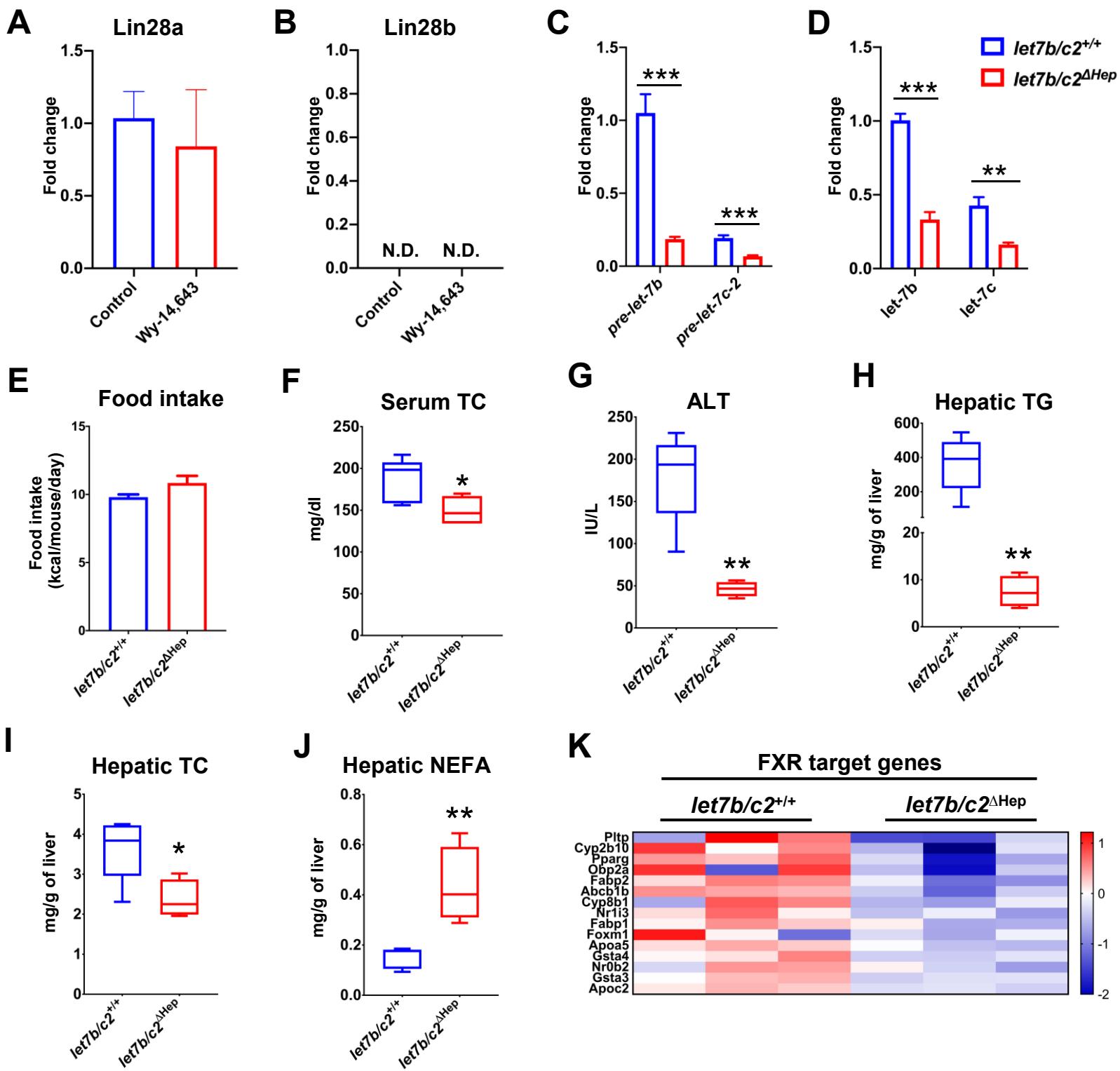


Supplemental information

Feedback repression of PPAR α signaling

by *Let-7* microRNA

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Supplemental Figure 1. Lin28 expressions in liver and phenotypic analyses of *let7b/c2^{+/+}* and *let7b/c2^{ΔHep}* mice. Related to Figure 1 and 2.

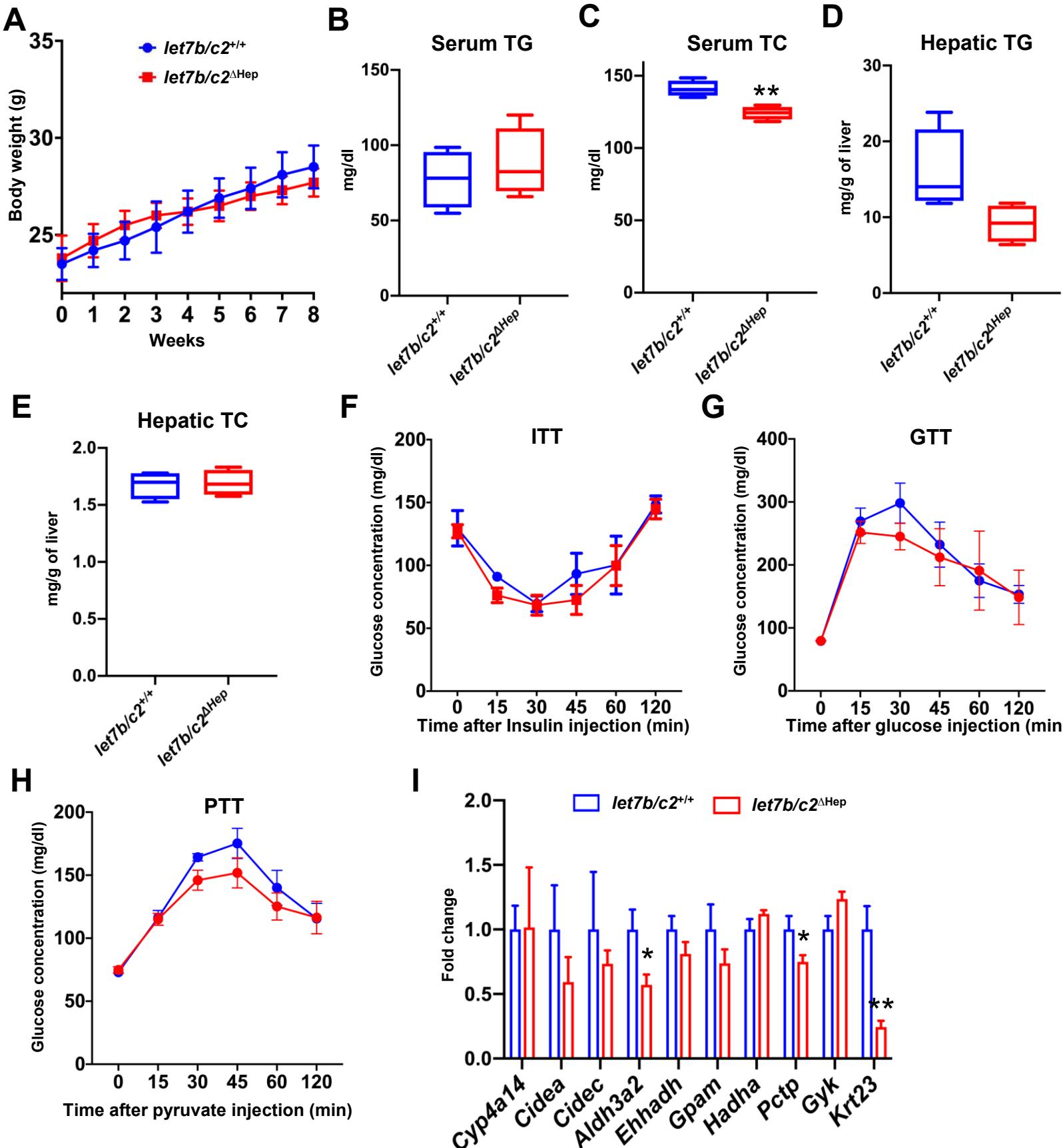
(A and B) Fold change of Lin28a (A) and Lin28b (B) mRNA abundance in Wy-14,643 treated liver. N.D.:Non-Detected.

(C and D) Fold change of pre-*let-7b* and pre-*let-7c-2* (C) and mature *let-7b* and *let-7c* (D) abundance in *let7b/c2^{+/+}* and *let7b/c2^{ΔHep}* livers.

(E) Average daily food intake (kcal/mouse/day) of *let7b/c2^{+/+}* and *let7b/c2^{ΔHep}* when HFD-feeding.

(F-J) Biochemical analyses for serum total cholesterol (TC) (F); alanine aminotransferase activity (ALT) (G); hepatic triglycerides (TG) (H); hepatic TC (I); and hepatic non-esterified fatty acid (NEFA) (J) in *let7b/c2^{+/+}* and *let7b/c2^{ΔHep}* mice after HFD-feeding.

(K) Heat map of FXR target genes identified by differential gene expression analysis of RNA-seq data from *let7b/c2^{+/+}* and *let7b/c2^{ΔHep}* livers after HFD-feeding. n = 5 mice/ group. Data are presented as mean ± SEM.



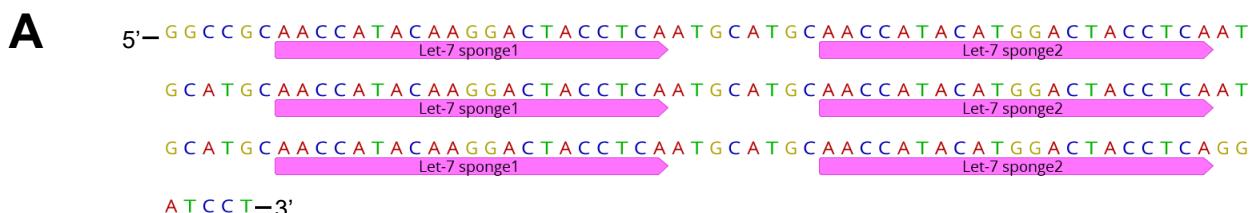
Supplemental Figure 2. Biochemical assays for normal chow diet-fed *let7b/c2^{+/+}* and *let7b/c2^{ΔHep}* mice. Related to Figure 2.

Body weight alterations of *let7b/c2^{+/+}* and *let7b/c2^{ΔHep}* mice during normal chow diet-feeding. Average daily food intake (kcal/mouse/day). N = 5 mice/ group. Data are presented as mean \pm SEM.

(B-E) Biochemical analyses for serum TG (B); serum TC (C); Hepatic TG (D); and Hepatic TC (E) in *let7b/c2^{+/+}* and *let7b/c2^{ΔHep}* mice after normal chow diet-feeding.

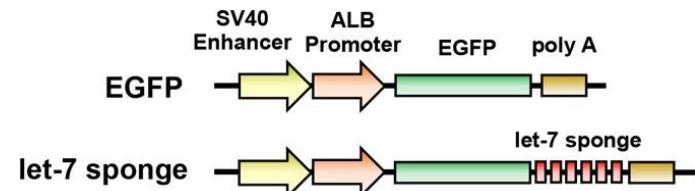
(F-H) ITT (F); GTT (G); and PTT (H) for normal chow diet-fed *let7b/c2^{+/+}* and *let7b/c2^{ΔHep}* mice.

(I) mRNA analysis by qRT-PCR of PPAR α target genes in *let7b/c2^{+/+}* and *let7b/c2^{ΔHep}* livers after 1-week HFD-feeding. n = 5 mice/ group. Data are presented as mean \pm SEM.

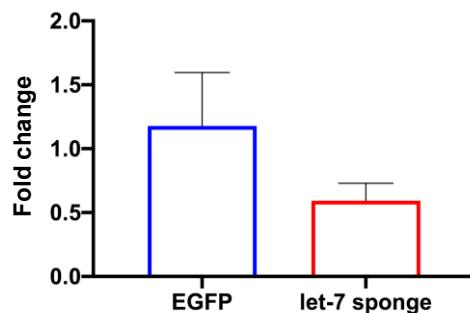


B

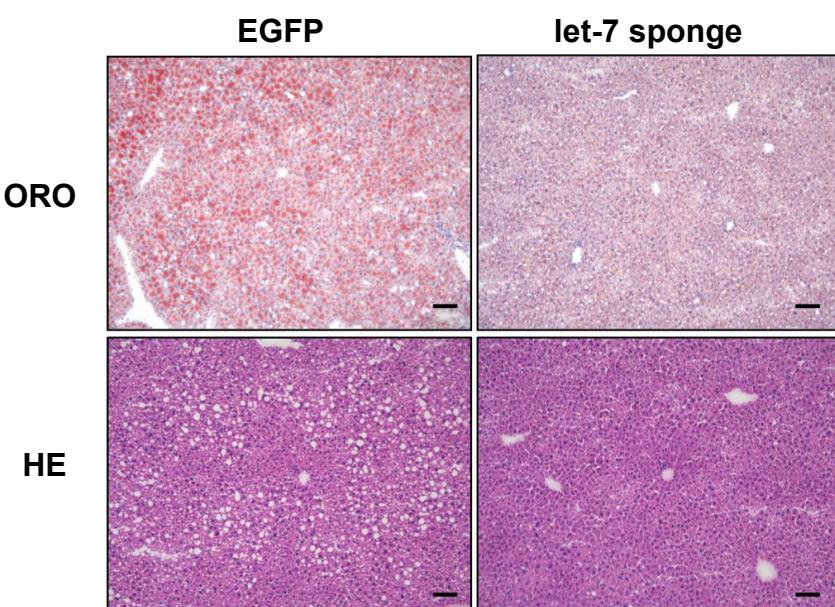
Self-complementary AAV-8



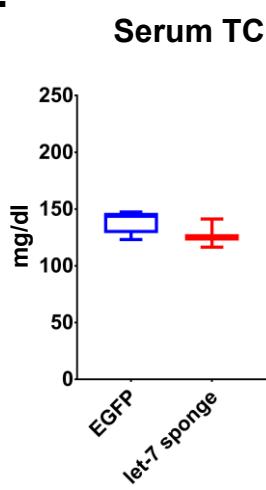
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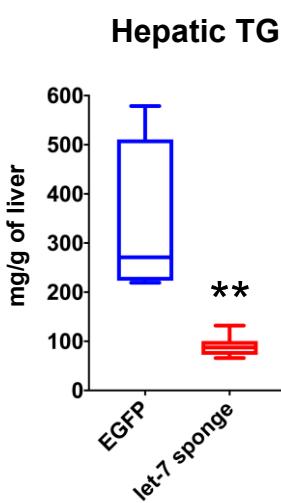
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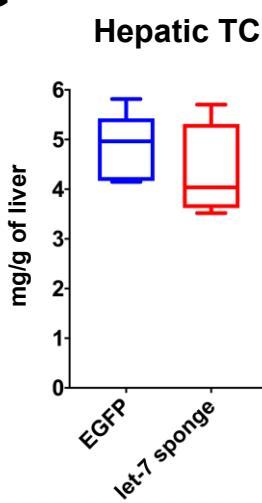
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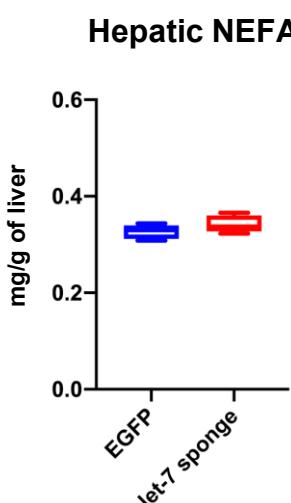
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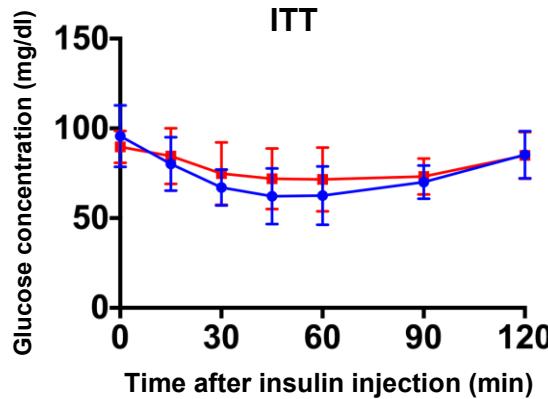
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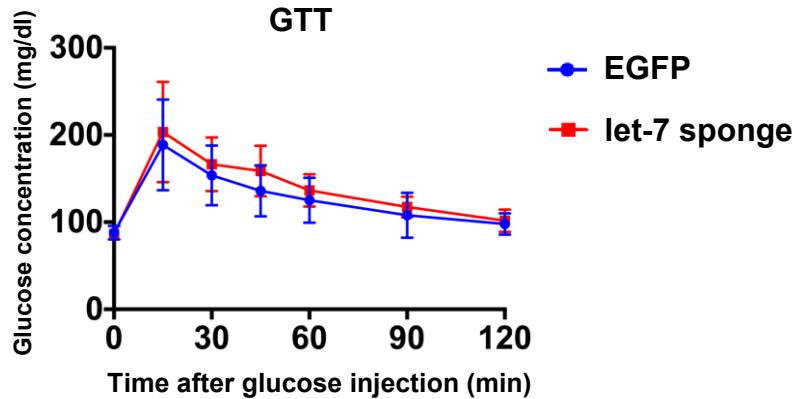
H



I



J



Supplemental Figure 3. let-7 sponge AAV construction and phenotypes of let-7 sponge AAV-transduced mice related to Figure 2.

(A) Sequence of let-7 sponge inserted in let-7 sponge AAV vector.

(B) Scheme of let-7 sponge expression AAV vector.

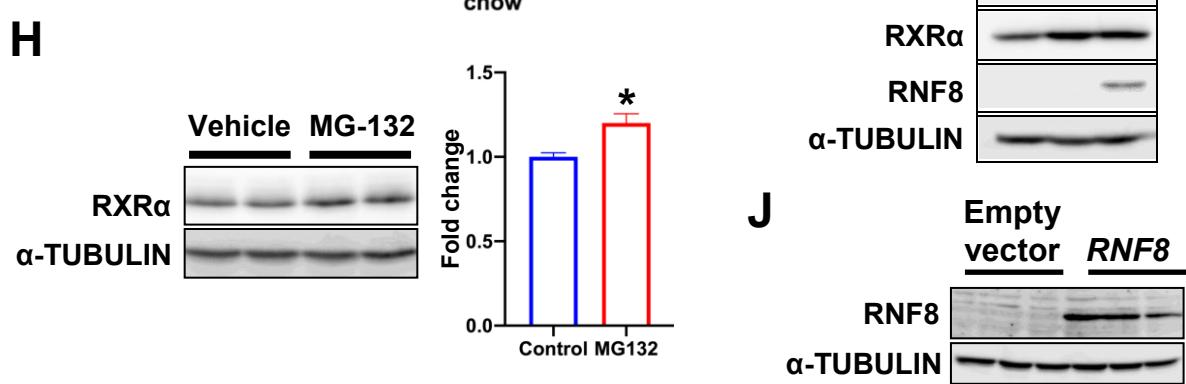
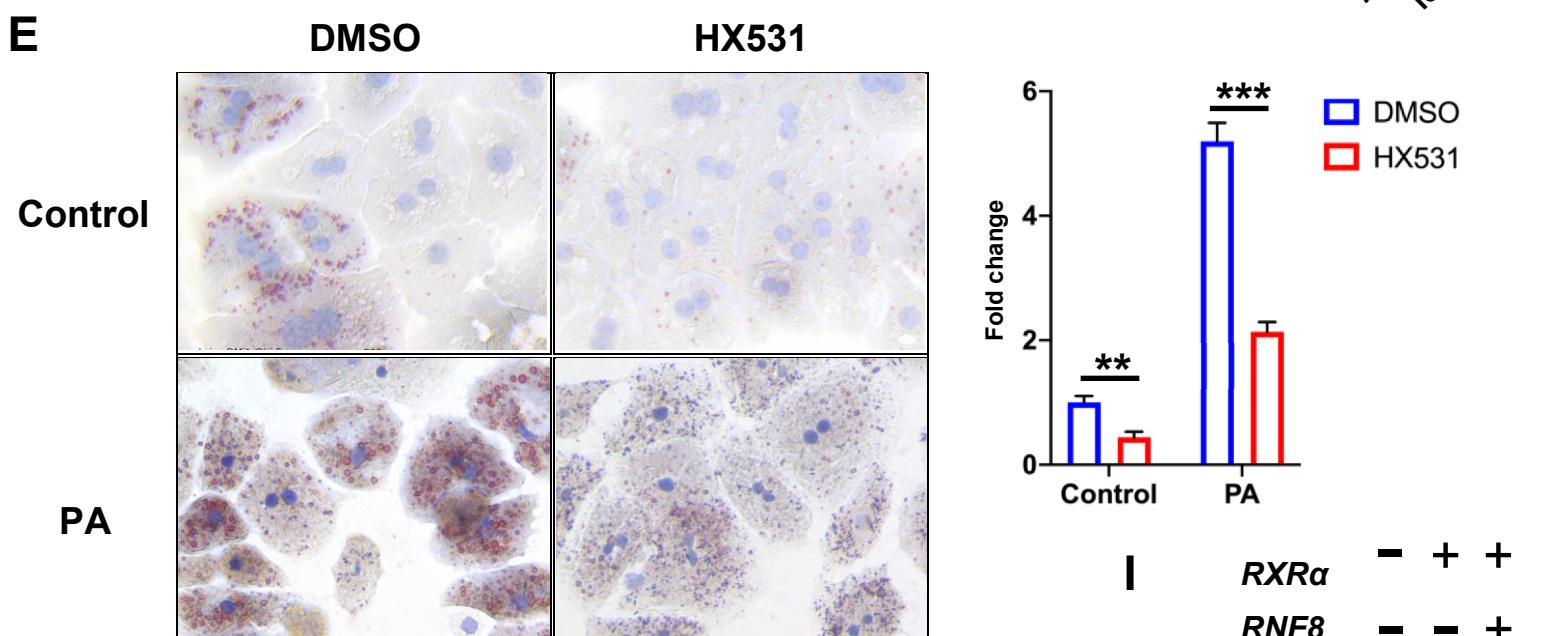
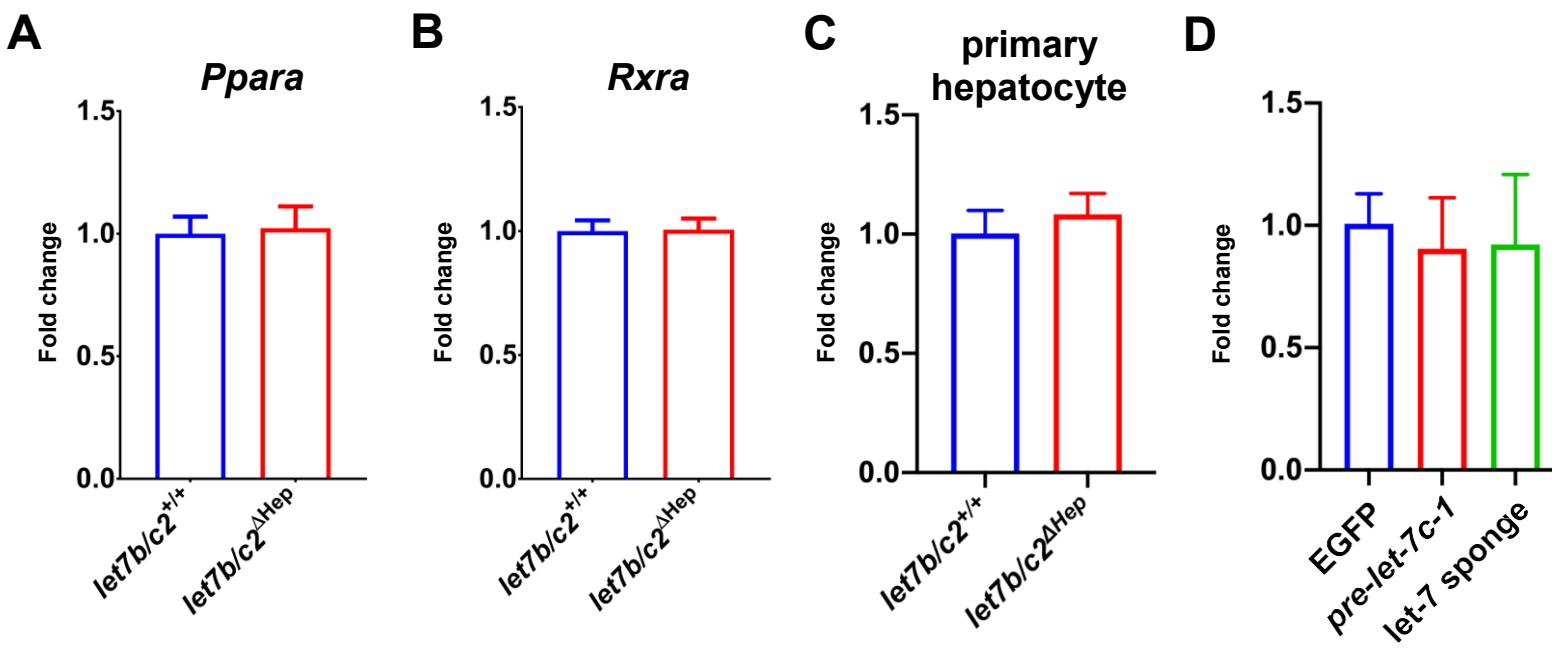
(C) qPCR analysis of GFP DNA in primary hepatocytes transduced by EGFP and let-7 sponge expressing AAV.

(D) Oil Red O staining of liver sections from wild-type mice infected with EGFP or let-7 sponge expressing AAV after being on HFD for eight weeks. Bars = 100 μ m

(E-H) Biochemical analyses for serum TC (D); hepatic TG (E); hepatic TC (F); and hepatic NEFA (G) concentration in EGFP and let-7 sponge AAV-infected mice after HFD-feeding.

(I and J) ITT (H); and GTT (I) for normal chow diet-fed EGFP and let-7 sponge AAV-infected mice.

Data are presented as mean \pm SEM. (n = 4-5 mice per group; *p < 0.05)



Supplemental Figure 4. RXR α expressions, functions and protein ubiquitination by let-7 miRNA related to Figure 3 and 4.

- (A-B) Fold change of PPAR α (A); and RXR α (B) mRNA in *let7b/c2^{+/+}* and *let7b/c2^{ΔHep}* livers fed HFD.
(C) Fold change of RXR α mRNA in *let7b/c2^{+/+}* and *let7b/c2^{ΔHep}* primary hepatocytes.
(D) Fold change of RXR α mRNA in EGFP, pre-let-7c-1 and let-7 sponge AAV-transduced liver after HFD-feeding.
(E) Oil red O staining and the morphometric analysis for lipid accumulation in mouse primary hepatocytes treated with Palmitic acid (800 μ M) and/or HX-531 (5 μ M).
(F) Western blot analysis and the densitometric quantification of RXR α in 60% HFD and/or Wy-14,643-treated mouse livers.
(G) Fold change of Rnf8 mRNA in EGFP and pre-let-7c-1 AAV-transduced liver.
(H) Western blot analysis and the densitometric quantification of RXR α in *Rxra* transfected Hepa-1 cells treated with the proteasome inhibitor MG-132.
(I) Western blot analysis of K48 polyubiquitin chain in *Rxra* and *Rnf8* transfected Hepa-1 cells.
(J) Western blot analysis of RNF8 protein in *Rnf8* transfected Hepa-1 cells.
Data are presented as mean \pm SEM. (n = 4-5 blots or fields per group; *p < 0.05, **p < 0.01, ***p < 0.001)

Supplemental Table 1. *let-7* microRNA family in mice related to Figure 1

Mature <i>let-7</i> microRNA	Precursor	Primary transcript	Clustered <i>let-7</i> precursor	Chromosome	miRBase ID
<i>let-7a</i>	<i>pre-let-7a-1</i>	let-7adf cluster	<i>pre-let-7d, pre-let-7f-1</i>	Chr.13	MI0000556
	<i>pre-let-7a-2</i>	3110039I08Rik		Chr.9	MI0000557
<i>let-7b</i>	<i>pre-let-7b</i>	Lincppara	<i>pre-let-7c-1</i>	Chr.15	MI0000558
<i>let-7c</i>	<i>pre-let-7c-1</i>	2810055G20Rik		Chr.16	MI0000559
	<i>pre-let-7c-2</i>	Lincppara	<i>pre-let-7b</i>	Chr.15	MI0000560
<i>let-7d</i>	<i>pre-let-7d</i>	let-7adf cluster	<i>pre-let-7a-1, pre-let-7f-1</i>	Chr.13	MI0000405
<i>let-7e</i>	<i>pre-let-7e</i>	Spaca6		Chr.17	MI0000561
<i>let-7f</i>	<i>pre-let-7f-1</i>	let-7adf cluster	<i>pre-let-7a-1, pre-let-7d</i>	Chr.13	MI0000562
	<i>pre-let-7f-2</i>	Huve1	<i>pre-miR-98</i>	Chr.X	MI0000563
<i>let-7g</i>	<i>pre-let-7g</i>	Wdr82		Chr.9	MI0000137
<i>let-7i</i>	<i>pre-let-7i</i>	mmu-let-7i		Chr.10	MI0000138
<i>miR-98</i>	<i>pre-miR-98</i>	Huve1	<i>pre-let-7f-2</i>	Chr.X	MI0000586

Summary of mature *let-7* miRNA, the precursors and the primary transcripts. Mature *let-7a*, *c* and *f* are processed from two precursors. *Pre-let-7a-1*, *pre-let-7d* and *pre-let-7f-1* are processed from an identical primary transcript. *Pre-let-7b* and *pre-let-7c-1* are also processed from an identical primary transcript

Supplemental Table 2. Downregulated pathways in *let7b/c2^{ΔHep}* mouse liver compared with *let7b/c2^{+/+}* related to Figure 3

Upstream Regulator	Expr Log Ratio	Molecule Type	Predicted Activation State	Activation z-score
PPARA	0.032	ligand-dependent nuclear receptor	Inhibited	-6.751
INSIG1	0.13	other	Inhibited	-3.564
Alpha catenin		group	Inhibited	-3.389
HOXA3		transcription regulator	Inhibited	-3.357
CFTR		ion channel	Inhibited	-3.194
PPARG	-1.333	ligand-dependent nuclear receptor	Inhibited	-3.171
NR1H4	0.035	ligand-dependent nuclear receptor	Inhibited	-3.134
PPARD	0.348	ligand-dependent nuclear receptor	Inhibited	-3.119
KLF15	0.058	transcription regulator	Inhibited	-2.721
LPL	1.748	enzyme	Inhibited	-2.619
TP73		transcription regulator	Inhibited	-2.601
PNPLA2	0.618	enzyme	Inhibited	-2.498
LCAT	-0.135	enzyme	Inhibited	-2.449
ABCG1	0.414	transporter	Inhibited	-2.433
INSIG2	-0.922	other	Inhibited	-2.415
let-7a-5p (and other miRNAs w/seed GAGGUAG)		mature microRNA	Inhibited	-2.414
ZFP36	-0.182	transcription regulator	Inhibited	-2.412
let-7		microRNA	Inhibited	-2.401
ZBTB20	0.568	transcription regulator	Inhibited	-2.4

Pathway analysis for RNA-seq data from 60% HFD-fed *let7b/c2^{+/+}* and *let7b/c2^{ΔHep}* mouse livers. PPAR and FXR pathways (blue labeled) were significantly repressed by *let7b/c2* deletion in liver.

Supplemental Table 3. Primers and Oligos used in this paper related to STAR Methods

Primers for mouse Quantitative RT-PCR		
	Forward (5' to 3')	Reverse (5' to 3')
<i>pre-Let-7a-1</i>	TTAGGGTCACACCCACCAAC	ATCACCTTAGGAAAGACAGTAGATTG
<i>pre-Let-7a-2</i>	TAGAGTTACATCAAGGGAGATAACTG	GTGCAAGTCCAAGGAAAG
<i>pre-Let-7b</i>	TCAGGGCAGTGATGTTGCC	TCAGGGAAAGGCAGTAGGTTG
<i>pre-Let-7c-1</i>	GAGTTACACCCCTGGGAGTTAAGT	AGTGTGCTCCAAGGAAAGCTAG
<i>pre-Let-7c-2</i>	TTTTGGGCTCTGCCCGC	GCGGCCACTTCAGGAAAGACAG
<i>pre-Let-7d</i>	TTAGGGCAGAGATTTGCC	GCCCTAAGAAAGGCAGCAGG
<i>pre-Let-7e</i>	TGAGGAAGACACCCGGAGGAG	GCAGCCTGGGAAAGCTAG
<i>pre-Let-7f-1</i>	GTGGGGTAGTGATTTACCCCTG	CTCAGGAAAGCAATAGATTGTATAG
<i>pre-Let-7f-2</i>	TTAGGGTCATACCCCATCTTGG	CGTGGGAAAGACAGTAGACTGTATAG
<i>pre-Let-7g</i>	TGAGGGCTATGATACCACCCCG	GCAAGGCAGTGGCTGTAC
<i>pre-Let-7i</i>	GGTCGGGTTGTGACATTGC	CTAGCAAGGCAGTAGCTTGC
<i>pre-miR98</i>	GTGGGGTAGGGATTTAGGCC	ATGCCACACACCAAGGAAAGTAG
<i>Cyp4a14</i>	CCTGACTTTCTTCGCGCTGC	TGATCACTCCATCTGTGTGCT
<i>Ehhad</i>	CCGGTCAATGCCATCAGT	CCGGTCAATGCCATCAGT
<i>Hadha</i>	AGGATCCTACGCTCCAGAGG	AAGCAGGATCCACTGATGGC
<i>Krt23</i>	CTGATTCACTCCGCCAAGA	TCCATCGTCCCTTCAGTGT
<i>Aldh3a2</i>	GGTCAAACCTGCATTGCTCC	TCCCCATAAAAGCTTCAACCG
<i>Cidea</i>	GACAGAAATGGACACCGGGTA	TGACATTGAGACAGCCGAGG
<i>Cidec</i>	TGGCACAATCGTGGAGACAG	CTCTCTTGCCTGTTCTG
<i>Gpam</i>	TGTGTACCCGAAGGTCTCCA	TGTTTACATCGGCCAAGGCT
<i>Pctp</i>	GGCTCTCTCGCAGATGTTT	TGAGCCAGGACGGAATTGG
<i>Gyk</i>	TGGCCTAAATGAAAGCTGGGG	TGCACTGGCTCCAAATAAG
<i>Gapdh</i>	AGGTGGTGTGAACGGATTG	TGTAGACCATGTAGTTGAGGTCA
<i>Ppara</i>	CACGCATGTGAAGGCTGTAA	CACGCATGTGAAGGCTGTAA
<i>Rxra</i>	CTCAATGGCGTCCTCAAGGT	ACCCCATAGTGTGCTGAG
<i>Rnf8</i>	GACTTCCAGGAGGAGGCAC	CCAGGGAGTTGGTCTGGAC
<i>Egfp</i>	AAGCTGACCCCTGAAGTTCATCTGC	CTTGTAGTTGCCGTGTCCTTGAA
Oligos for Let-7 sponge AAV construction (5' to 3')		
<i>Let-7 sponge</i>	GGCCGCAACCATAACAGGACTACCTCAATGCATGCAACCATACTGGACTACCTCAATGCATGCAACCATAACAGG ACTACCTCAATGCATGCAACCATACTGGACTACCTCAATGCATGCAACCATAACAGGACTACCTCAATGCATGCAA CCATACATGGACTACCTCAGGATCCT	
<i>Let-7 sponge complement</i>	CTAGAGGATCCTGAGGTAGTCCATGTATGGTGCATGCATTGAGGTAGTCCTGTATGGTGCATGCATTGAGGTAG TCCATGTATGGTGCATGCATTGAGGTAGTCCTGTATGGTGCATGCATTGAGGTAGTCATGTATGGTGCATGC ATTGAGGTAGTCCTGTATGGTGC	
Primers for <i>pre-Let-7c-1</i> AAV construction (5' to 3')		
<i>Let-7c-1 NotI EcoRV</i>	AAAGCGGCCGCTAACGGTGGTCAAGG	AAAGATATCGCGTTGTGTCATTGCTTACAAG
Oligos for <i>Rnf8</i> 3'UTR luciferase reporter vectors		
<i>Rnf8</i> 3'UTR Wild-type	AAAGCTAGCGACTGAGTTAGGAAGTCTGTGCCACTGGCCTCCGTATGCAGGCCCTGCTGCTACCACCCCCAA CGTGTGCTGGGACCCCTGCTCGGACTGCCTGACTGTGCATTAATGAGTTGAGTTCTGTGTTCTTAATTGTTGATT GTTTTGATACCTCATGCACCCCTTGTGACATCATTGGACAGCTGGGTGTGAGTCCTGTGGCTGTGGAAGGT GGCCTGCATTTGAGAGCCTGGTTACCTTTATGAAAACATGATGTCATTCTGGGAATAAAATGTATAAAACTC TCAGTTGCTCAGCTGGCTCGAGAAA	
<i>Rnf8</i> 3'UTR Mutant	AAAGCTAGCGACTGAGTTAGGAAGTCTGTGCCACTGGCCTCCGTATGCAGGCCCTGCTGCTACCACCCCCAA CGTGTGCTGGGACCCCTGCTCGGACTGCCTGACTGTGCATTAATGAGTTGAGTTCTGTGTTCTTAATTGTTGATT GTTTTGAGCGGCCCTGACCCCTTGTGACATCATTGGACAGCTGGGTGTGAGTCCTGTGGCTGTGGAAG GTGGCCTGCATTTGAGAGCCTGGTTACCTTTATGAAAACATGATGTCATTCTGGGAATAAAATGTATAAAAC TCTCAGTTGCTCAGCTGGCTCGAGAAA	

Primers and oligos used for qRT-PCR. AAV constructs and *Rnf8* 3'UTR reporter vectors.