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Supplemental information

Agonists and allosteric modulators

promote signaling from different

metabotropic glutamate receptor 5 conformations

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1	Supplementary information		
2	Agonists and allosteric modulators promote signalling from different		
3	metabotropic glutamate receptor 5 conformations [115]		
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Figure S1. Thermal stability measurements of mGlu₅-5M receptor solubilised in detergents, Related to Figure 1 and Table S2. (A) Thermostability of the mGlu₅-5M dimer was measured using $[^{3}H]$ -MPEP binding assay after extraction in different detergents including C₈E₄ (Black), NG (Orange), DM (Magenta), DDM (Green) and MNG3 (Blue), all supplemented with CHS. (B) Thermostability of the 7TM domain of mGlu₅-5M was measured for the same set of detergents. Experiments were performed in duplicates and each curve represents the average of three independent experiments with the exception of C₈E₄ for the mGlu₅-5M construct. T_m mean values are presented in Table S2.



Figure S2. Size exclusion chromatography and SDS-gel of detergent-purified thermostabilised
mGlu₅ receptor, Related to Figure 2 and Figure 4. Size exclusion chromatography was
performed using 24 ml S200 column for thermostabilised mGlu₅-5M dimer bound to inhibitors
(A), activators (C) and mGlu₅-StaR(569-836)-T4L bound to alloswitch-1 (E). 10% SDS-gel
electrophoresis of purified thermostabilised mGlu₅-5M dimer bound to antagonist and NAM
(B), agonist and PAM (D) and 15% SDS-gel electrophoresis of purified mGlu₅-StaR(569-836)T4L bound to alloswitch-1 (F).



- 57
- 58 Figure S3. CryoEM analysis of LY341495 and MPEP-bound mGlu₅ conformation, Related to 59 Figure 2. (A) A representative micrograph of antagonist and NAM bound mGlu₅ in detergent micelles on ice. Scale bar is 500Å. (B) A selection of reference-free 2D class averages of mGlu5 60 in antagonist bound conformation showing the distinct domains of the protein. The box size is 61 128 pixels and sampled at 3.56 Å. (C) Local resolution plot of antagonist mGlu₅ bound as 62 63 determined with Relion 3.1. Two different views of the maps are shown at different thresholds. 64 The molecule on the right has been rotated to show a different view of mGlu₅ and is at lower 65 threshold. The lower threshold where the 7TM is covered by the detergent/lipid belt and the 66 higher threshold showing the helices of the 7TM. Much of the VFT is between resolutions of 67 3.7 to 5 Å but the C-terminal part of CRD and the 7TM are of lower resolution. (D) The Fourier 68 shell correlation (FSC) curves of two half-maps with mask (blue), unmasked maps (black) and 69 the map and model (red) are shown. The estimated resolution by comparison of the two half-70 maps from refinement and postprocessing at FSC 0.143 is 4.0 Å and the one of map vs model 71 at FSC 0.5 is 4.3 Å. The FSC curves were estimated for the full molecule. (E) To verify 72 overfitting of the model refinement, one of the half-map used in refinement and the other half-73 map was used as test. (F) The CryoEM workflow showing the different steps used to obtain 74 the final map.
- 75



77 Figure S4. Cryo EM analysis of quisqualate and VU0424465-bound mGlu₅ conformation, 78 Related to Figure 2. (A) A representative micrograph of agonist and PAM bound mGlu₅ in 79 detergent micelles on ice. Scale bar is 500 Å. (B) A selection of reference-free 2D class averages of mGlu₅ in closed conformation showing the distinct domains of the protein. The 80 81 box size is 128 pixels and sampled at 3.56 Å. (C) Local resolution plot of mGlu₅ bound to 82 quisqualate as estimated with Relion 3.1. Two different views of the maps are shown at 83 different thresholds that hides detergent belt. The molecule on the right shows a rotated view of mGlu₅. Much of the VFTs are between resolutions of 3.6 to 5 Å but the C-terminal part of 84 85 CRDs and the 7TMs are of lower resolution. (D) The Fourier shell correlation (FSC) curves of 86 two half-maps with mask (blue), unmasked maps (black) and the map and model (red) are 87 shown. The estimated resolution by comparison of the two half-maps from refinement and 88 postprocessing at FSC 0.143 of half-maps is 3.8 Å and that of the map vs model at FSC 0.5 is 89 4.2 Å. The FSC curves were estimated for the full molecule. (E) Cross-validation of model 90 refinement. To verify overfitting of the model refinement, one of the half-map used in 91 refinement and the other half-map was used as test. (F) The CryoEM workflow showing the 92 different steps used to obtain the final map.



Figure S5. EM maps of select regions of full-length mGlu₅-5M bound to agonist (quisqualate) and antagonist (LY341495), Related to Figure 2. (A) α-helical and β-sheet regions from the VFT domain. The regions from each model include residues 26-42, 92-98 (β-sheet), 101-116,

153-165, 195-211 (α -helices). EM maps sharpened with B-factor of -122 Å² and -164 Å² for 97 98 agonist and antagonist respectively were used to represent the density using Pymol. (B) The 99 region comprising the CRD is shown. Only the backbone of the model is depicted due to lower resolution. EM maps sharpened with B-factor of -61 Å² and -82 Å² for agonist and antagonist 100 respectively was used for depiction of maps using Pymol. (C) Two stretches of residues (690-101 102 760, 775-818) in the 7TM for agonist and antagonist are shown. In the agonist map, the extra density observed between the two helices, which is most likely the ligand PAM used in the 103 104 preparation is marked with black arrow. The density for 7TMs of agonist is better than the 105 antagonist and is clearly visualised in the maps. The docking of the TMD into the EM maps was aided by the X-ray structure of the 7TM. EM maps sharpened with B-factor of -61 Å² and 106 -82 Å² for agonist and antagonist respectively was used for display of maps using Pymol. 107



Figure S6. Comparison of the LY341495 binding site in the VFT of mGlu receptors that belongs to three different groups, Related to Figure 3. (A) The LY341495 binding to mGlu₅ pocket observed from this work and belongs to the group I, (B) from the mGlu₁ structure (PDB

113 3KS9) that belongs to the same group, (C) mGlu₇ structure (PDB 3MQ4) that belongs to group

114 III and (D) mGlu₃ structure (PDB 3SM9) that belongs to group II.



Figure S7. Binding of LY341495 is likely to require a larger opening of the VFT, Related to Figure 3. (A-D) Distances between C α of the Y223 in lobe II and T173, A174 and S175 in lobe1 of protomer A (shown in panels A and B) and protomer B (shown in panels C and D). LY341494-bound mGlu₅ model is represented in blue (A and C) and apo inactive conformation (PDB 6N52) in cyan (B and D).

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Figure S8. Alloswitch-1 binding mode in the 7TM domain of the mGlu₅ 7TM, Related to Figure 4. (A) 2Fo-Fc map for alloswitch-1 bound to the thermostabilised 7TM domain of the human mGlu₅ receptor. Map was calculated using Refmac and figure made with pymol at 1.5 σ . (B) 2D representation of Alloswitch-1 ligand binding site. 2D plot was generated using LigPlot⁺. (C) Alloswitch-1 superposition with the NAM co-crystallised with the mGlu₅ Star

- 129 7TM domain. Alloswitch-1 superposition with Mavoglurant (Green; 4009), M-MPEP (Light
- 130 pink; 6FFI), Fenobam (Orange; 6FFH), HTL14242 (Cyan; 5CGD), HTL14242 derivative
- 131 (yellow; 5CGC), Alloswitch-1 (Purpleblue; 7P2L). NAMs are coloured as indicated in brackets
- 132 along with the PDB code. (D) Alloswitch-1 superposition with M-MPEP.
- 133



135 Figure S9. Probing the quisqualate and alloswitch-1 binding sites of mGlu₅ under dark conditions with IP1 accumulation assay, Related to Figure 4. (A-R) Quisqualate-induced IP1 136 137 accumulation (open circles) and alloswitch-1 inhibition of IP1 accumulation in the presence 138 (red circles) or absence (black circles) of a fixed concentration of quisqualate (300 nM; red 139 circle) for mGlu₅ mutants, under dark conditions. Single point mutations were tested for their 140 response to quisqualate and/or alloswitch-1 function of the mGlu₅. Data are expressed as a 141 percentage of the maximal quisqualate response and represent the mean \pm S.E.M. of at least 142 three independent experiments performed in duplicate.





147 IP₁ accumulation (open circles) and alloswitch-1 inhibition of IP₁ accumulation in the presence 148 (red circles) or absence (black circles) of a fixed concentration of quisqualate (300 nM; red 149 circles) for mGlu₅ mutants, under illuminated conditions. Single point mutations were tested 150 for their response to quisqualate and/or alloswitch-1 function at the mGlu₅ under 380 nm 151 conditions. Data are expressed as a percentage of the maximal quisqualate response and 152 represent the mean \pm S.E.M. of at least three independent experiments performed in duplicate. 153



Figure S11. Quantification of cell surface expression of fluorescently labelled SNAP-tagged wildtype or mutant mGlu₅ receptors, Related to Figure 4. Data are expressed as the mean \pm S.E.M. of at least three separate experiments performed in either duplicate or triplicate. Data were analysed with a one-way ANOVA with post-hoc Dunnett's multiple comparison's test. **P<0.01 denotes significant difference in cell surface expression from the mGlu5 WT receptor expressing cells.



162 Figure S12. Interaction of the photoswitchable NAM, alloswitch-1, with the orthosteric 163 agonist, quisqualate, with mGlu₅ mutants in IP₁ accumulation assay, under dark or 380 nm conditions, Related to Figure 4. (A, B) The response of WT mGlu₅ to alloswitch-1 in dark or 164 380 nm conditions are shown. The effect of the single point mutations S809A^{7.39} (C, D) or 165 166 W785A^{6.50} (E, F) on the ability of alloswitch-1 to inhibit quisqualate-induced IP₁ accumulation 167 in a concentration dependent manner was determined and compared to mGlu₅ WT, under dark 168 and 380 nm conditions. Data are expressed as a percentage of the maximal quisqualate response 169 and represent the mean \pm S.E.M. of at least three independent experiments performed in duplicate. 170

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Table S1. Thermostability of human mGlu₅ mutants bound to the negative allosteric modulator MPEP, Related to Figure 1. The best single thermostable mutants A813L (mGlu₅-1M; TM7) was first selected and used as a platform for combining additional thermostable mutants in the following sequence order, T742A (TM5), S753A (TM5), T777A (TM6), and I799A (TM7). For each receptor mutants, mGlu₅-1M, mGlu₅-2M, mGlu₅-3M, mGlu₅-4M and the final thermostable mutant, mGlu₅-5M, T_m are expressed as the mean value from at least 3 independent experiments (\pm SEM), or \pm SD for the mean of 2 independent experiments.

Mutants name	mGlu ₅ mutations	Apparent T _m in
		MNG3 (°C)
mGlu₅-∆856	-	20.7 ± 0.5 , (n=12)
	R668A	22.9 ± 0.3 , (n=2)
	T742A	24.1 ± 1.3 , (n=6)
	G748A	22.6 ± 0.4 , (n=2)
	G794A	23.2 ± 0.1 , (n=2)
	S753A	21.5 ± 0.2 , (n=2)
	A787L	25.3 ± 0.2 , (n=3)
	T777A	24.1 ± 1.0 , (n=2)
	I799A	26.5 ± 0.9 , (n=3)
mGlu ₅ -1M	A813L	28.0 ± 1.2 , (n=4)
	A855L	22.4 (n=1)
	A856L	23.4 ± 0.9 , (n=2)
mGlu ₅ -2M	A813L/ T777A	32.5 ± 0.5 , (n=4)
mGlu ₅ -3M	A813L/T777A/ S753A	34.5 ± 1.3, (n=3)
mGlu ₅ -4M	A813L/T777A/S753A/ I799A	37.0 ± 1.0 , (n=2)
mGlu ₅ -5M	A813L/T777A/S753A/I799A/ T742A	39.5 ± 0.6 , (n=5)

Table S2. Detergent-thermostability comparison between the human mGlu₅-5M dimer and1837TM-mGlu₅-5M, both bound to the negative allosteric modulator MPEP, Related to Figure 1.184Apparent T_m values from each detergent are expressed as the mean \pm SEM from at least three185independent experiments performed in duplicates, with the exception of C₈E₄ for the mGlu₅-1865M construct. All detergents are supplemented with CHS.

Solubilisation	mGlu ₅ construct	Apparent T _m in
condition		detergents (°C)
MNG3-CHS	mGlu ₅ -5M	41.42 ± 0.18 , (n=3)
DDM-CHS	mGlu ₅ -5M	40.22 ± 0.24 , (n=4)
DM-CHS	mGlu ₅ -5M	32.75 ± 0.33 , (n=4)
NG-CHS	mGlu ₅ -5M	29.65 ± 0.32 , (n=4)
C ₈ E ₄ -CHS	mGlu ₅ -5M	28.76 ± 0.23 , (n=2)
MNG3-CHS	7TM-mGlu5-5M	33.81 ± 0.36, (n=5)
DDM-CHS	7TM-mGlu5-5M	34.26 ± 0.29 , (n=4)
DM-CHS	7TM-mGlu ₅ -5M	22.70 ± 0.26 , (n=4)
NG-CHS	7TM-mGlu5-5M	$19.42 \pm 0.30, (n=5)$
C ₈ E ₄ -CHS	7TM-mGlu5-5M	$18.16 \pm 0.67, (n=4)$
		, ()

-	PDB -7FD9, EMDB-31537	PDB-7FD8, EMDB-31536
	mGlu ₅ + LY341495 +MPEP	mGlu5 +quisqualate+VU0424465
Microscope	Titan Krios	Titan Krios
Operating voltage	300 kV	300 kV
Mode	EFTEM	EFTEM
Nominal magnification	130000x	130000x
Pixel size (Å)	0.89	0.89
Spot size	7	7
Beam size (µm)	0.9	1.05
C2 aperture (μm)	50	50
Objective aperture (µm)	100	100
Exposure time (sec)	5.2	9
Dose rate (e ⁻ /p/s)	7.6	5.894
Fractions (# of frames)	32	48
Dose/frame (e ⁻ /Å ² /s)	1.56	1.39
Total dose (e ⁻ /Å ²)	49.9	67
No. of particles in final map	142191	118016
Resolution nominal (Å), FSC 0.143	4.0	3.8
Auto B-factor sharpening (Å ²)*	-164	-122
Refinement		
Starting models	7P2L**, 6n52,	7P2L**, 6n50, 6n51,
Model composition		
Number of chains	2	2
Protein (No. of atoms)	5698	6014
Ligands (NAG, CHS,	40	62
Quisqualate, LY341495)		
Model Refinement		
FSC @ 0.5 (Å)	4.3	4.2
Average B factor (Å ²)		
Overall	147.5	113.4
Protein	148.0	113.2
NAG	93.7	84.5
Quisqualate/LY341495	61.6	38.8
CHS	-	185.1
RMS deviations		
Bonds (Å)	0.006	0.006
Angles (°)	1.1	1.2
Validation		
Ramachandran Plot (favoured/outliers)	93.6/0	91.3/0.13
Clash score	9.1	5.1
Molprobity score	1.9	1.8

201 **Table S3.** Cryo-EM Statistics for map and model refinement, Related to Figure 2.

* The auto B-factor sharpened map was used for model refinement. For map interpretation/model building, maps sharpened with multiple B-factors were used

** - The 7TM used as template is the alloswitch-1 model in this study

Table S4. Residues in the VFT binding sites for LY341495 and quisqualate in the mGlu5
structure compared with binding site residues in mGlu1, 3 and 7, Related to Figure 3. Residues
shown are within 4.0 Å of the ligand.

mGlu ₁ PDB(3KS9)	mGlu5 LY341495 (7FD9)	mGlu ₃ (PDB 3SM9)	mGlu7 (PDB 3MQ4)	mGlu ₅ Quisqualate (7FD8)
Y74	Y64	R64	N74	Y64-
		R68	R78	
W110	W100			W100
G163	G150	S149	S157	
S164	S151	Y150	G158	S151
S165	S152	S151	S159	S152
S186	S173	A172	A180	S173
A187	A174	S173	S181	A174
T188	T175	T174	T182	T175
S189		S175		
D208		D194		
N235	N222	221	S229	
Y236	Y223	222	Y230	Y223
	E279			E279
				G280
D318	D305	D301		D305
G319		G302		G306
		K389	K407	K396

208	Table S5. Potency estimates for quisqualate-stimulated IP ₁ accumulation or alloswitch-1
209	inhibition of IP_1 accumulation in the presence or absence of a fixed concentration of
210	quisqualate (300 nM), under dark conditions at mGlu5 mutations, Related to Figure 4.
211	Estimated potency values represent the mean \pm S.E.M. of at least three independent
212	experiments performed in duplicate. Data were fitted to a standard logistic function and
213	analysed by one-way ANOVA, with Dunnett's post-test. *P<0.05, **P<0.01, ***P<0.001,
214	**** P <0.0001 denotes significant difference of ligand potency at an mGlu ₅ - Δ 856 as compared
215	to the mGlu ₅ WT receptor.

	Quisqualate pEC ₅₀	Alloswitch-1	Alloswitch-1 pIC ₅₀ values,
	values (n)	pIC ₅₀ values (n)	when interacted with 300
			nM quisqualate (n)
mGlu5	7.52±0.18(4)	6.79±0.09(4)	$6.66 \pm 0.03(4)$
mGlu₅-∆856	7.30±0.10(9)	6.94±0.13(9)	7.07±0.17(9)
I651A ^{3.36}	7.38±0.07(4)	6.90±0.13(4)	6.40±0.27(4)
P655A ^{3.40}	7.39±0.11(3)	7.24±0.11(4)	6.97±0.27(4)
S658A ^{3.43}	7.56±0.32(4)	7.27±0.26(4)	7.19±0.13(4)
Y659A ^{3.44}	7.13±0.28(4)	5.73±0.54(4)**	5.28±0.36(3)****
V740A ^{5.40}	6.90±0.09(4)	7.14±0.02(4)	7.08±0.16(4)
L744A ^{5.44}	7.41±0.06(4)	7.30±0.06(4)	7.47±0.12(4)
N747A ^{5.47}	7.31±0.16(4)	5.93±0.46(4)*	6.20±0.41(4)
T781A ^{6.46}	8.13±0.24(4)**	6.06±0.15(4)	5.84±0.19(4)**
W785A ^{6.50}	7.24±0.12(4)	5.50±0.09(4)***	5.72±0.43(3)**
F788A ^{6.53}	7.03±0.06(4)	7.62±0.12(4)	7.48±0.23(4)
Y792A ^{6.57}	7.55±0.06(4)	7.56±0.23(4)	$7.60 \pm 0.22(4)$
M802A ^{7.32}	7.28±0.05(4)	6.80±0.19(4)	6.74±0.12(4)
S805A ^{7.35}	7.39±0.08(4)	6.99±0.24(4)	6.75±0.23(3)
V806A ^{7.36}	7.10±0.15(4)	7.19±0.37(4)	6.94±0.42(4)
S809A ^{7.39}	7.39±0.23(4)	5.46±0.26(4)***	5.53±0.23(3)***
A810L ^{7.40}	7.39±0.20(3)	n.d.	5.50±0.02(3)***
A813L ^{7.43}	7.40±0.21(4)	6.93±0.42(4)	6.90±0.20

Table S6. Potency estimates for quisqualate-stimulated IP₁ accumulation or alloswitch-1 inhibition of IP₁ accumulation in the presence or absence of a fixed concentration of quisqualate (300 nM), under 380 nm conditions at mGlu₅ mutations, Related to Figure 4. Estimated potency values represent the mean \pm S.E.M. of at least three independent experiments performed in duplicate. Data were fitted to a standard logistic function and analysed by one-way ANOVA, potency values were compared to the mGlu₅- Δ 856 truncated in the C-terminus after Alanine 856.

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	Quisqualate pEC ₅₀ values (n)	Alloswitch-1 pIC ₅₀ values (n)	Alloswitch-1 pIC ₅₀ values, when interacted with 300 nM quisqualate (n)
mGlu5	7.68±0.15(3)	4.48±0.12(3)	5.31±0.29(4)
mGlu₅-∆856	7.56±0.12(8)	5.44±0.23(8)	5.58±0.24(8)
I651A ^{3.36}	7.67±0.25(4)	5.39±0.20(4)	5.16±0.13(4)
P655A ^{3.40}	7.17±0.10(4)	n.d.	n.d.
S658A ^{3.43}	7.91±0.14(4)	5.51±0.35(4)	6.20±0.29(4)
Y659A ^{3.44}	7.37±0.13(4)	6.21±0.32(4)	5.61±0.31(3)
V740A ^{5.40}	7.32±0.17(4)	5.54±0.22(3)	6.07±0.75(3)
L744A ^{5.44}	7.54±0.20(4)	5.68±0.15(4)	5.70±0.09(4)
N747A ^{5.47}	7.73±0.24(4)	5.63±0.44(3)	5.70±0.31(4)
T781A ^{6.46}	7.81±0.09(4)	n.d.	n.d.
W785A ^{6.50}	7.30±0.17(4)	n.d.	n.d.
F788A ^{6.53}	7.12±0.22(4)	5.93±0.20(4)	5.89±0.15(4)
Y792A ^{6.57}	7.85±0.15(4)	6.16±0.59(4)	6.44±0.48(4)
M802A ^{7.32}	7.42±0.10(4)	5.58±0.32(4)	4.92±0.43(4)
S805A ^{7.35}	7.69±0.40(4)	5.84±0.17(4)	5.27±0.24(4)
V806A ^{7.36}	7.25±0.22(4)	5.43±0.22(4)	4.92±0.19(2)
S809A ^{7.39}	7.53±0.15(4)	n.d.	n.d.
A810L ^{7.40}	7.22±0.20(4)	n.d.	n.d.
A813L ^{7.43}	7.26±0.17(3)	5.76±0.38(3)	5.60±0.20(4)

Table S7. X-Ray Data collection and refinement statistics, Related to Figure 4.

PDB ID	- 7P2L*
Space group	C2
a, b, c (Å)	141.69, 43.40, 82.12, 90, 99.384, 90
α, β. γ (°)	90, 90, 90
Beamline	SLS-X06SA
Wavelength (Å)	1.0
Resolution (Å)	41.45-2.54 (2.61–2.54)**
Rmeas	0.25 (2.68)
Ι/σ (Ι)	6.35 (1.05)
Completeness (%)	99.9 (100)
Multiplicity	10.66 (9.30)
CC1/2 (%)	0.99 (0.51)
Refinement	
Resolution (Å)	49.11-2.54
No. of unique reflections	16594
Rwork/Rfree ***	0.23/0.28
R.m.s. deviations	
Bond lengths (Å)	0.01
Bond angles (°)	1.73
B-factor	
Total	75.0
Protein	75.2
Ligand	57.2
Water	60.1
Ramachandran Plot	
Favored (%)	95.5
Outliter (%)	0.25
Clash score	2.6
MolProbity score	1.4

* Data processing and refinement statistics are reported with Friedel pairs merged.

** Values in parentheses are for the highest resolution shell.

*** R_{free} was calculated using 5% of randomly selected subset of reflections and the remaining 95% of reflections was used for calculation of R_{work} .

- Table S8. Potency measurement for agonist quisqualate-stimulated and ago-PAM VU0424465
- 235 in IP_1 accumulation in the presence or absence of a fixed concentration of LY341495 (100
- μ M), Related to Figure 5. Values are the average +/- S.E.M from at least 3 different experiments.

	mGlu ₅ -Δ856 pEC ₅₀
LY341495	5.51 ± 0.63 (n=3)
Quisqualate	6.83 ±0.16 (n=6)
Quisqualate + LY341495 (10 ⁻⁴ M)	5.27 ±0.05 (n=3)
VU0424465	7.47 ±0.08 (n=6)
VU0424465+ LY341495 (10 ⁻⁴ M)	6.82 ±0.05 (n=6)