

## A: Pre-Trycycler assembly

Assembly A:  
 contig\_1: TCGGCGTGTGGTCTAAAGACTCCGGATGGGGCGTCATGTTGATTCATCGATAATTTTC  
 contig\_2: AGCGTTGTACG

Assembly B:  
 contig\_1: GACGCCCCATCCGGAGTCTTTAGGACCCTCGCCGAGAAAAATTATCGATGAATCACCA  
 contig\_2: TTGTAGCGAGCG  
 contig\_3: AAAAAA

Assembly C:  
 contig\_1: GCCGAGAAAAATTATCGATGAATCAACCATGACGCCCATCCGGAGTCTTTAGACCACCGCC

Assembly D:  
 contig\_1: GATCCGGATGGGGCGTCATGTTGATTCATCGATAATTTTCTCGCGGGTGGTCTAAA  
 contig\_2: AACGCCGTACAAC

As input, Trycycler takes multiple different assemblies of the same genome. These can be generated using different assemblers and/or different read subsets.

## B: Clustering contigs

Cluster 1:  
 A\_contig\_1: TCGGCGTGTGGTCTAAAGACTCCGGATGGGGCGTCATGTTGATTCATCGATAATTTTC  
 B\_contig\_1: GACGCCCCATCCGGAGTCTTTAGGACCCTCGCCGAGAAAAATTATCGATGAATCACCA  
 C\_contig\_1: GCCGAGAAAAATTATCGATGAATCAACCATGACGCCCATCCGGAGTCTTTAGACCACCGCC  
 D\_contig\_1: GATCCGGATGGGGCGTCATGTTGATTCATCGATAATTTTCTCGCGGGTGGTCTAAA

Cluster 2:  
 A\_contig\_2: AGCGTTGTACG  
 B\_contig\_2: TTGTAGCGAGCG  
 D\_contig\_2: AACGCCGTACAAC

Cluster 3:  
 B\_contig\_3: AAAAAA

Contigs from all assemblies are clustered based on their *k*-mer content. Trycycler makes a tree of the contig relationships to help users distinguish good clusters (which represent completely assembled replicons) vs bad clusters (which contain spurious, fragmented or incorrectly assembled sequences).

## C: Reconciling contigs

Normalise strands and fix circularisation:

Cluster 1:  
 A\_contig\_1: GAAAATTATCGATGAATCAACCATGACGCCCATCCGGAGTCTTTAGACCACACGCCGA  
 B\_contig\_1: GACGCCCCATCCGGAGTCTTTAGGACCCTCGCCGAGAAAAATTATCGATGAATCACCA  
 C\_contig\_1: GCCGAGAAAAATTATCGATGAATCAACCATGACGCCCATCCGGAGTCTTTAGACCACCGCC  
 D\_contig\_1: TTTAGACCACCGCCGAGAAAAATTATCGATGAATCAACCATGACGCCCATCCGGATC

Cluster 2:  
 A\_contig\_2: CGTACAACGCT  
 B\_contig\_2: CGCTCGCTACAAC  
 D\_contig\_2: AACGCCGTACAAC

Contig sequences are flipped to their reverse complement as necessary to ensure that all sequences within each cluster are on the same strand. For circular clusters, sequences are aligned to each other to repair circularisation issues: trimming overlapping bases or adding missing bases.

Rotate to consistent start:

Cluster 1:  
 A\_contig\_1: ATGAATCAACCATGACGCCCATCCGGAGTCTTTAGACCACACGCCGAGAAAAATTATCG  
 B\_contig\_1: ATGAATCACCATGACGCCCATCCGGAGTCTTTAGGACCCTCGCCGAGAAAAATTATCG  
 C\_contig\_1: ATGAATCAACCATGACGCCCATCCGGAGTCTTTAGACCACCGCCGAGAAAAATTATCG  
 D\_contig\_1: ATGAATCAACCATGACGCCCATCCGGATCTTTAGACCACCGCCGAGAAAAATTATCG

Cluster 2:  
 A\_contig\_2: GCTCGTACAAC  
 B\_contig\_2: GCTCGTACAAC  
 D\_contig\_2: GCGCTACAAC

For each circular cluster, a starting sequence is identified (using a standard coding sequence, if possible) and the sequences are rotated to have a consistent start/end. Each cluster's sequences are now ready for global multiple sequence alignment.

## D: Multiple sequence alignment

Cluster 1:  
 A\_contig\_1: ATGAATCAACCATGACGCCCATCCGGAGTCTTTAG--ACCACACCGCAGAAAA--TTATCG  
 B\_contig\_1: ATGAATC--ACCATGACGCCCATCCGGAGTCTTTAGGACCCTCGCCGAGAAAAATTATCG  
 C\_contig\_1: ATGAATCAACCATGACGCCCATCCGGAGTCTTTAG--ACCAC--CGCCGAGAAAAATTATCG  
 D\_contig\_1: ATGAATCAACCATGACGCCCATCCGGAG--TCTTTAG--ACCACCGCCGAGAAAAATTATCG

Cluster 2:  
 A\_contig\_2: GCTCG--TACAAC  
 B\_contig\_2: GCTCGTACAAC  
 D\_contig\_2: GC--CGTACAAC

Trycycler uses MUSCLE to produce a global multiple sequence alignment for each of the clusters.

## E: Partitioning reads

All reads:  
 CTCGCC AATTAT AGAAAA CTCGCT  
 GAGAAA TTAGAC AACGCT TCGCTA  
 AGACCA CGAGAA CCGCGC GACCAC  
 TCTTTA CACTCG CGGAGT CGCTCG  
 ATCAAC GCTCGC GAAAAA AACCAT  
 GCTCTT CCGTA GTACAA CACCAT  
 ACCAA TACAAC TGACGC CCCATC  
 ATGACG CGCCGA CTACAA ACGCCG  
 TCCGGA AAAAAA GCTACA GGAGTC  
 CATGAC GCCCA ACAACG GATGAA

Cluster 1 reads:  
 CTCGCC AATTAT AGAAAA GAGAAA  
 TTAGAC AGACCA CGAGAA CCGCCG  
 GACCAC TCTTTA CACTCG CGGAGT  
 ATCAAC GAAAAA AACCAT GCTCTT  
 CACCAT ACCAA TGACGC CCCATC  
 ATGACG CGCCGA TCCGGA AAAAAA  
 GGAGTC CATGAC GCCCA GATGAA

Cluster 2 reads:  
 CTCGCT AACGCT TCGCTA CGCTCG  
 GTACAA GCTCG CCGCTA TACAAC  
 CTACAA ACGCCG GCTACA ACAACG

Reads are aligned to each contig sequence and assigned to the cluster to which they best align.

## F: Generating a consensus

Divide alignment into chunks:

Cluster 1:  
 ATGAATC ACCATGACGCCCATCCGGAGTCTTTAGGACCCTCGCCGAGAAAAATTATCG

Cluster 2:  
 GC TCG TACAAC

The multiple sequence alignment is divided into chunks: "same" chunks where the sequences agree and "different" chunks where there are multiple possible options.

Choose best option for each chunk:

Cluster 1:  
 ATGAATC ACCATGACGCCCATCCGGAGTCTTTAGGACCCTCGCCGAGAAAAATTATCG

Cluster 2:  
 GC TCG TACAAC

For each "different" chunk, the most popular option is chosen (as defined by the minimum total Hamming distance to other options). When there is a tie, reads are aligned to each alternative to decide which option to keep (the one with the best total read alignment score).

## G: Post-Trycycler polishing

Trycycler assembly: ATGAATCAACCATGACGCCCATCCGGAGTCTTTAGGACCCTCGCCGAGAAAAATTATCG

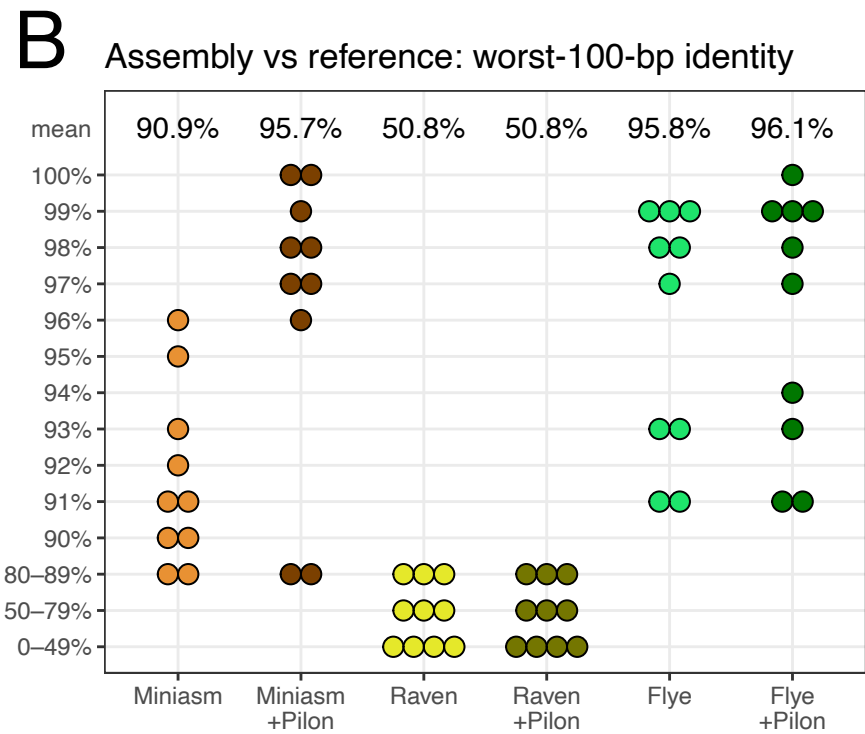
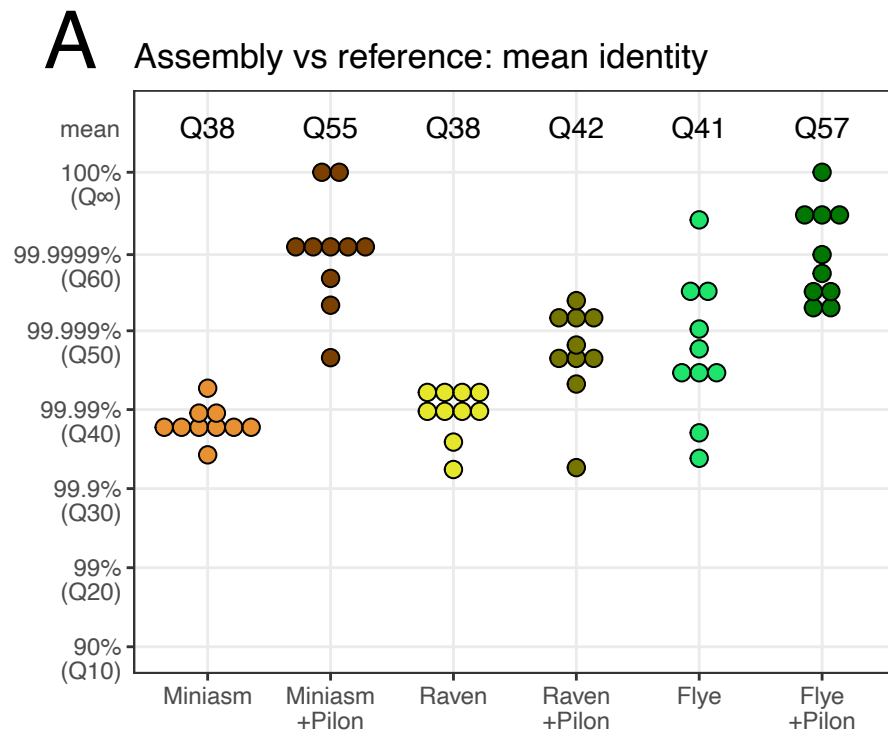
After long-read polishing: ATGAATCAACCATGACGCCCATCCGGAGTCTTTAGGACCCTCGCCGAGAAAAATTATCG

After short-read polishing: ATGAATCAACCATGACGCCCATCCGGAGTCTTTAGGACCCTCGCCGAGAAAAATTATCG

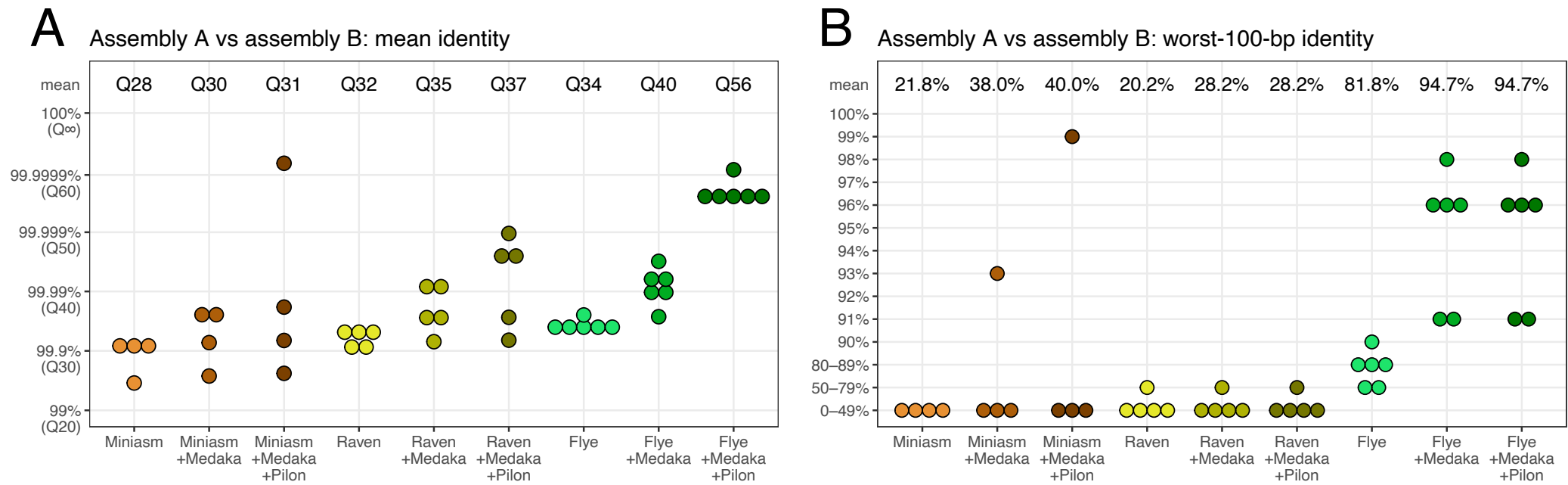
GCTCGTACAAC  
 GCTCGTACAAC  
 GCTCGTACAAC

After Trycycler is finished, platform-specific long-read polishing (e.g. Medaka for ONT sequencing) can reduce the number of small-scale errors in the assembly. If available, short-read polishing (e.g. with Pilon) can further reduce small-scale errors.

**Fig. S1:** steps in the Trycycler assembly pipeline.

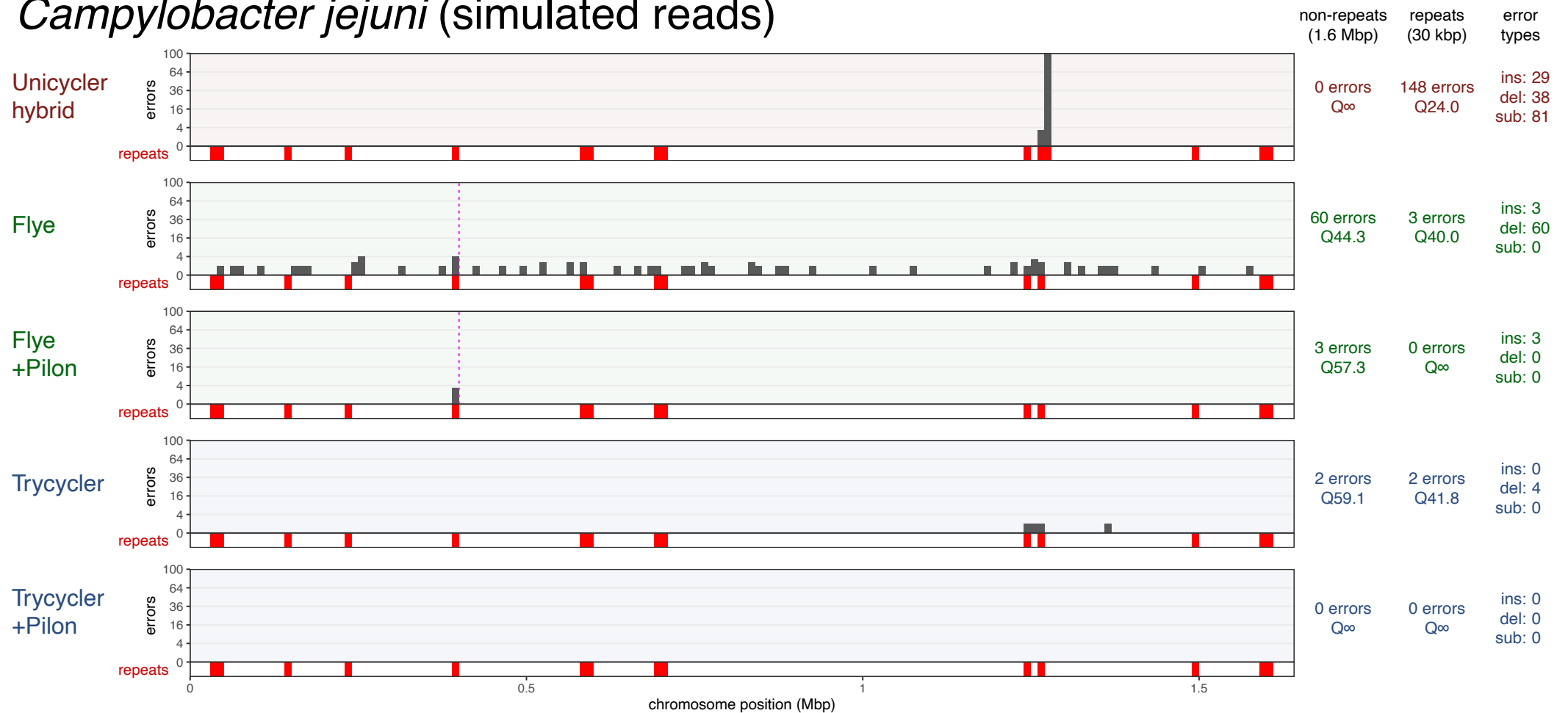


**Fig. S2:** results for the simulated read tests for long-read-only assemblers. This figure contains the same analyses as are shown in Figure 2, but it includes assemblies from all long-read-only assemblers (both before and after Pilon polishing).



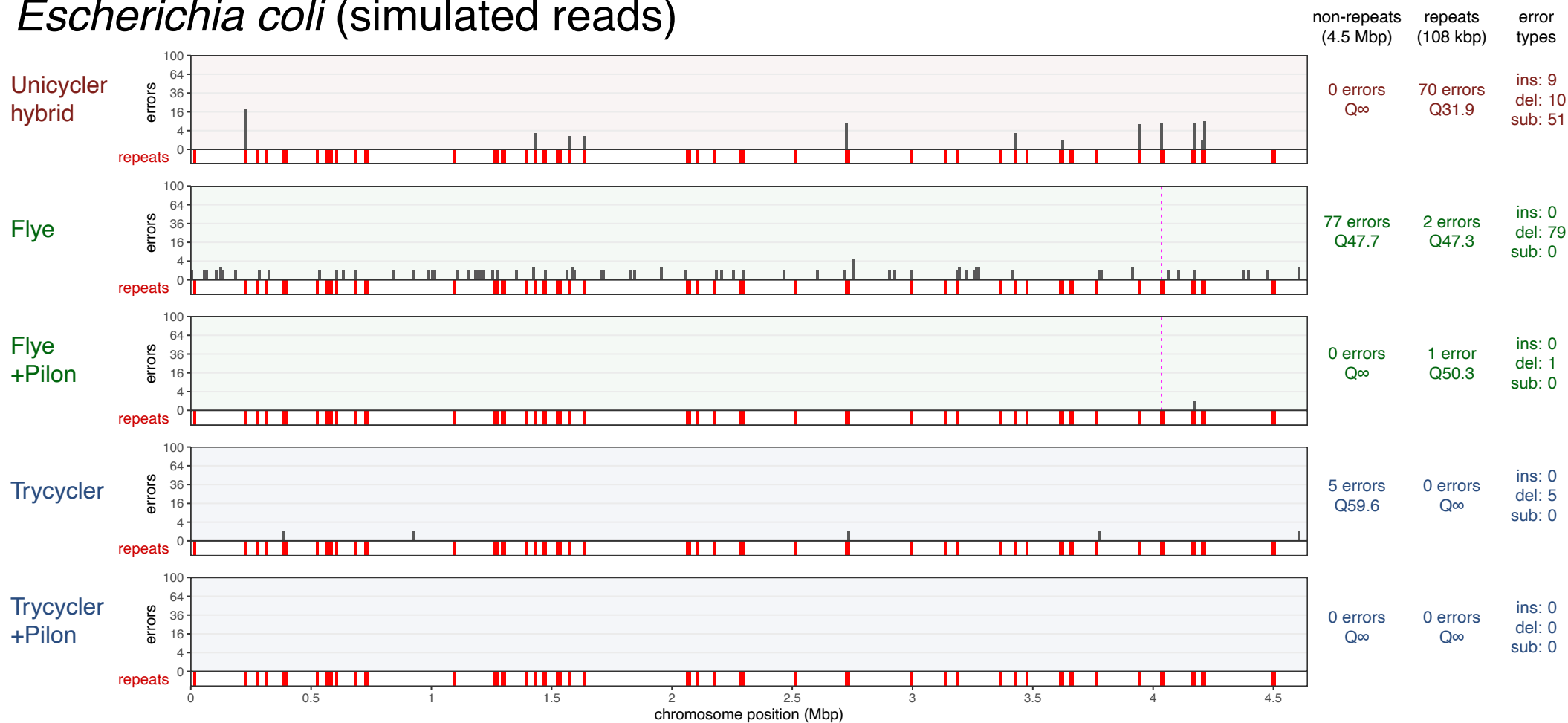
**Fig. S3:** results for the real read tests for long-read-only assemblers. This figure contains the same analyses as are shown in Figure 3, but it includes assemblies from all long-read-only assemblers (unpolished, Medaka-polished and Medaka+Pilon-polished for each). In two of the six genomes, Miniasm failed to produce a completed chromosome for both read sets, resulting in only four data points. In one of the six genomes, Raven failed to produce a completed chromosome for both read sets, resulting in only five data points.

# *Campylobacter jejuni* (simulated reads)



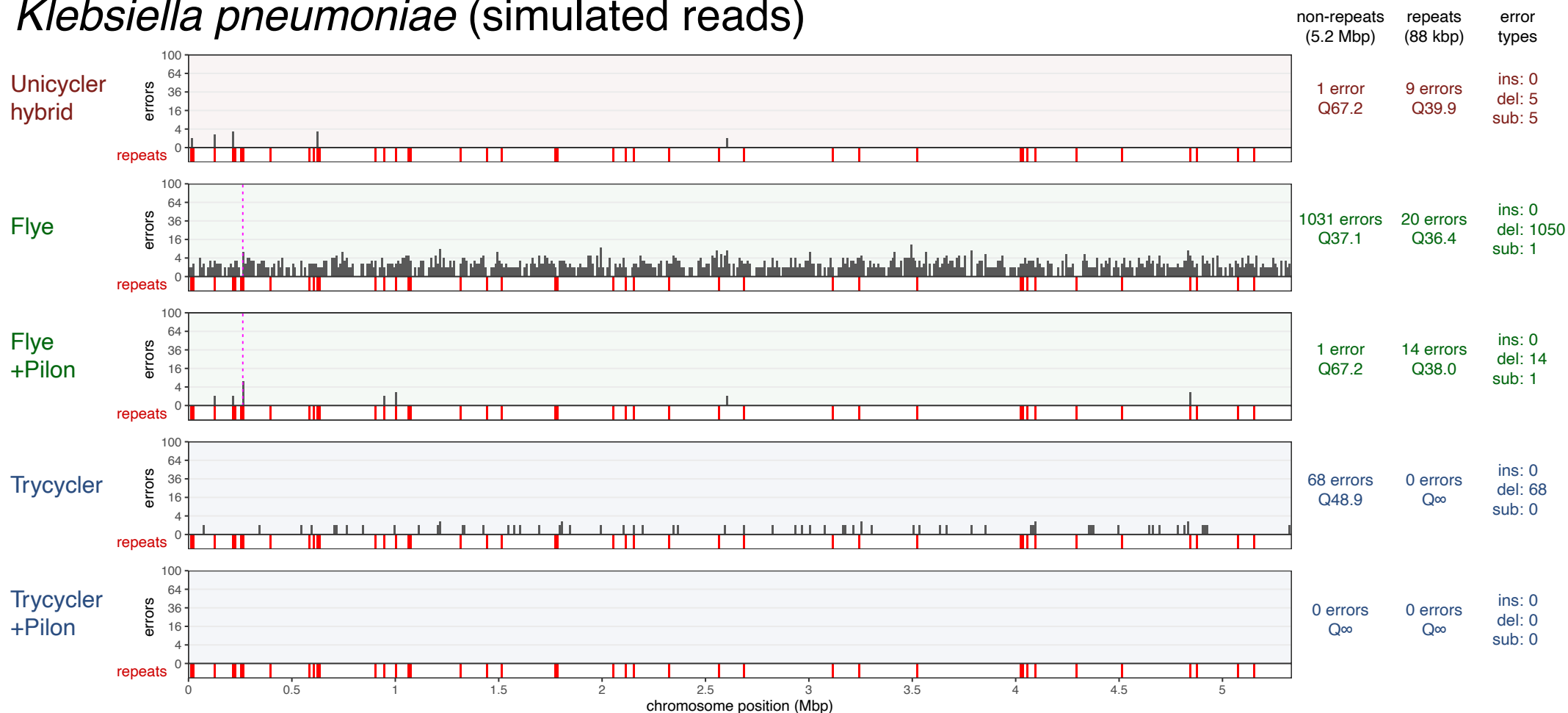
**Fig. S4-a:** error distributions (as defined by alignment to the reference sequence) for the simulated-read assemblies of the *Campylobacter jejuni* chromosome. Repetitive regions of the genome are shown as red blocks at the bottom of each plot. For Flye assemblies, the contig start/end position (where the sequence began before it was rotated to be consistent with the reference) is shown by the dashed line.

# *Escherichia coli* (simulated reads)



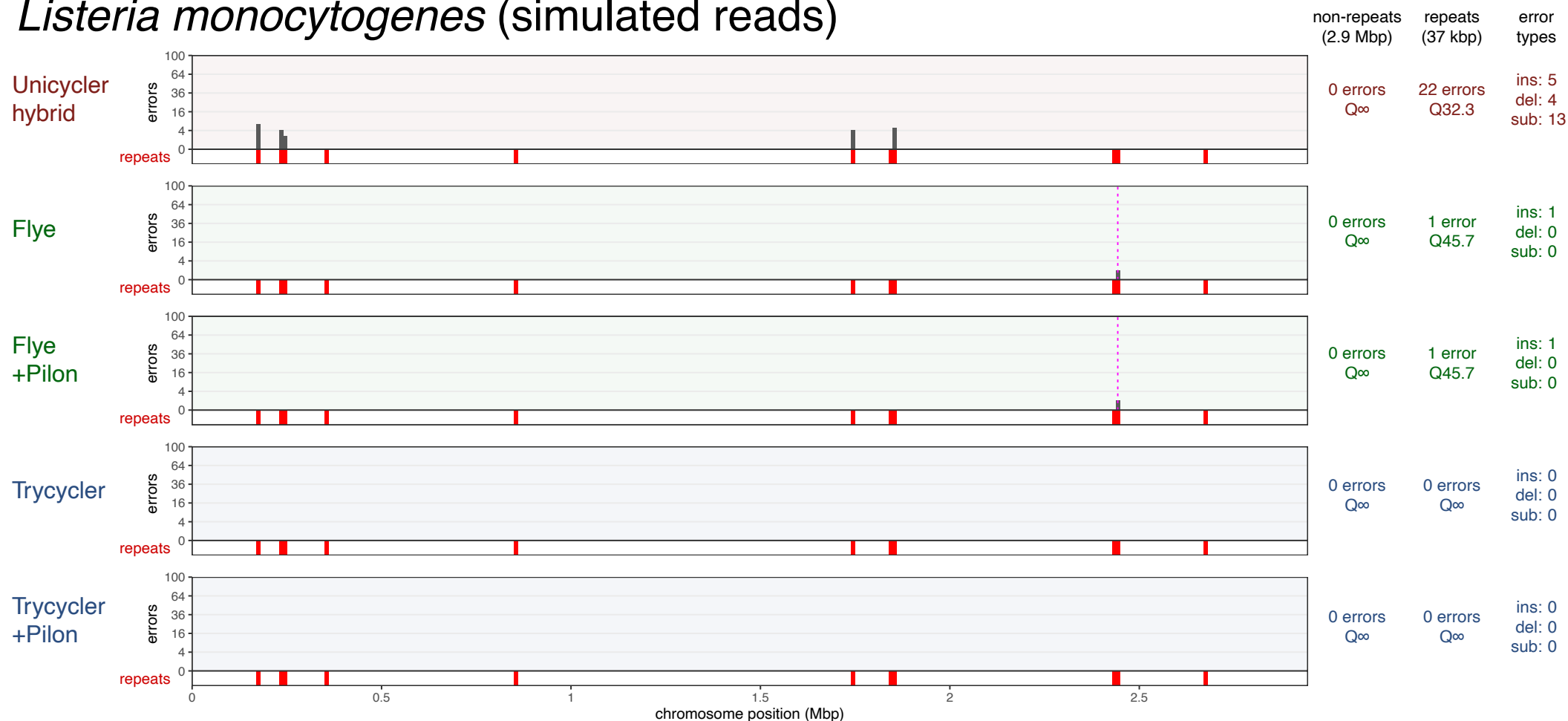
**Fig. S4-b:** error distributions (as defined by alignment to the reference sequence) for the simulated-read assemblies of the *Escherichia coli* chromosome. Repetitive regions of the genome are shown as red blocks at the bottom of each plot. For Flye assemblies, the contig start/end position (where the sequence began before it was rotated to be consistent with the reference) is shown by the dashed line.

# *Klebsiella pneumoniae* (simulated reads)



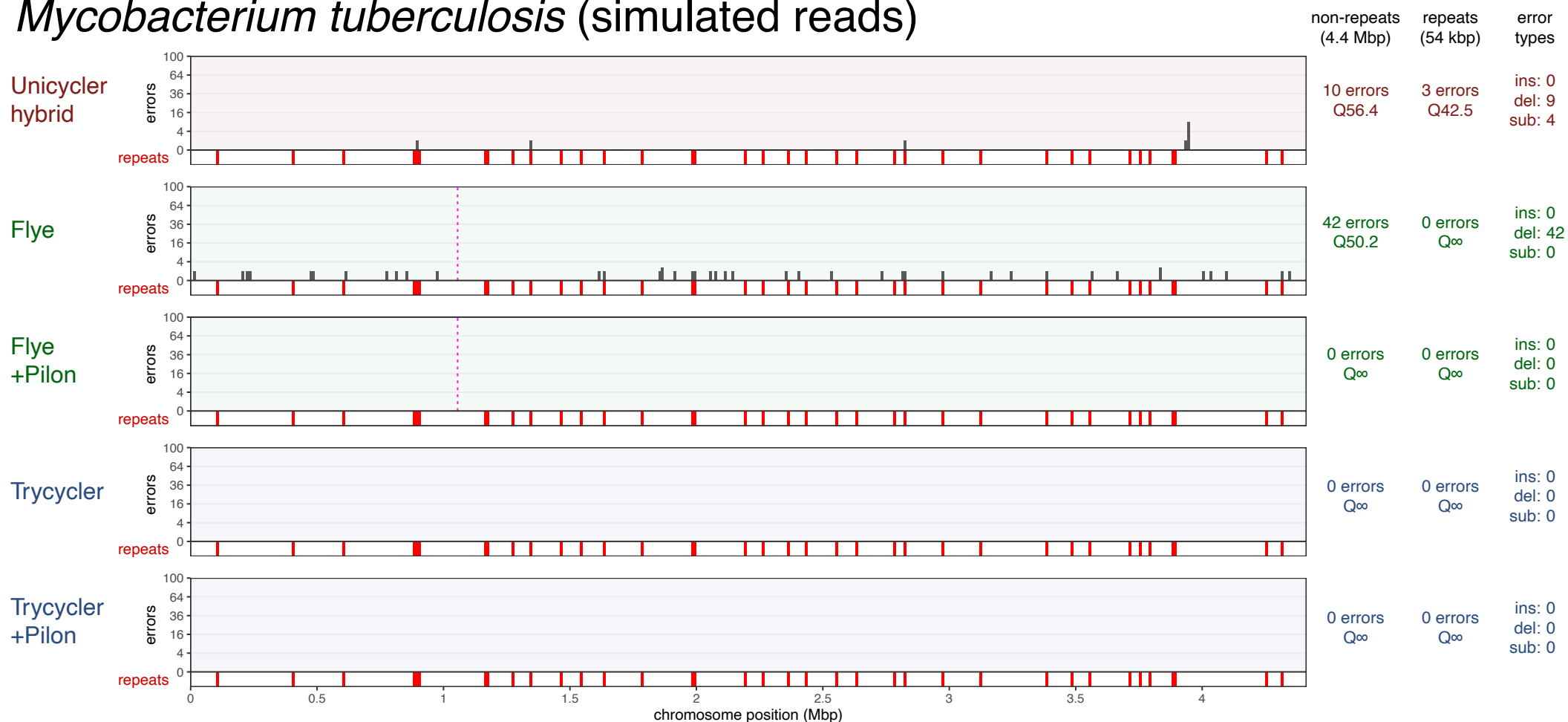
**Fig. S4-c:** error distributions (as defined by alignment to the reference sequence) for the simulated-read assemblies of the *Klebsiella pneumoniae* chromosome. Repetitive regions of the genome are shown as red blocks at the bottom of each plot. For Flye assemblies, the contig start/end position (where the sequence began before it was rotated to be consistent with the reference) is shown by the dashed line.

# *Listeria monocytogenes* (simulated reads)



**Fig. S4-d:** error distributions (as defined by alignment to the reference sequence) for the simulated-read assemblies of the *Listeria monocytogenes* chromosome. Repetitive regions of the genome are shown as red blocks at the bottom of each plot. For Flye assemblies, the contig start/end position (where the sequence began before it was rotated to be consistent with the reference) is shown by the dashed line.

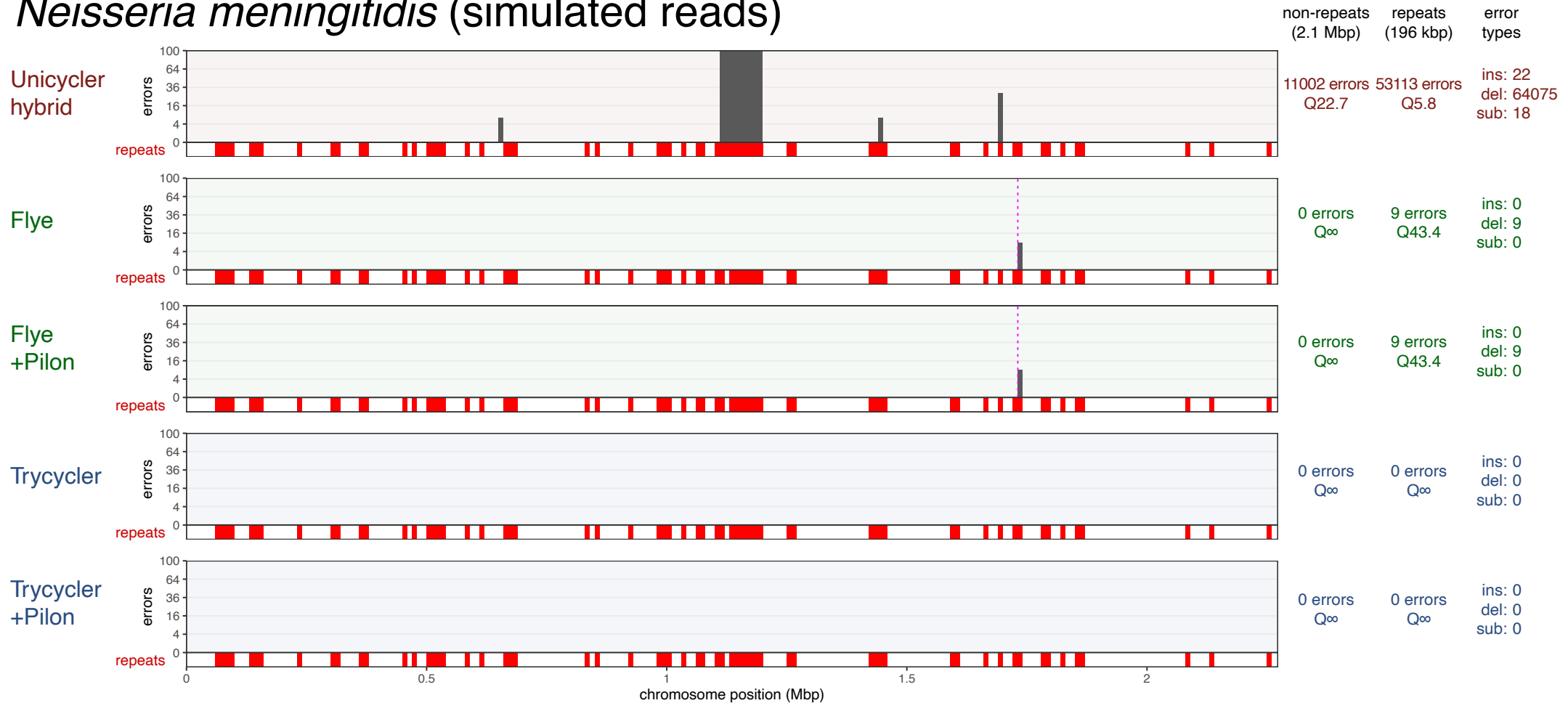
# *Mycobacterium tuberculosis* (simulated reads)



**Fig. S4-e:** error distributions (as defined by alignment to the reference sequence) for the simulated-read assemblies of the *Mycobacterium tuberculosis* chromosome. Repetitive regions of the genome are shown as red blocks at the bottom of each plot. For Flye assemblies, the contig start/end position (where the sequence began before it was rotated to be consistent with the reference) is shown by the dashed line.

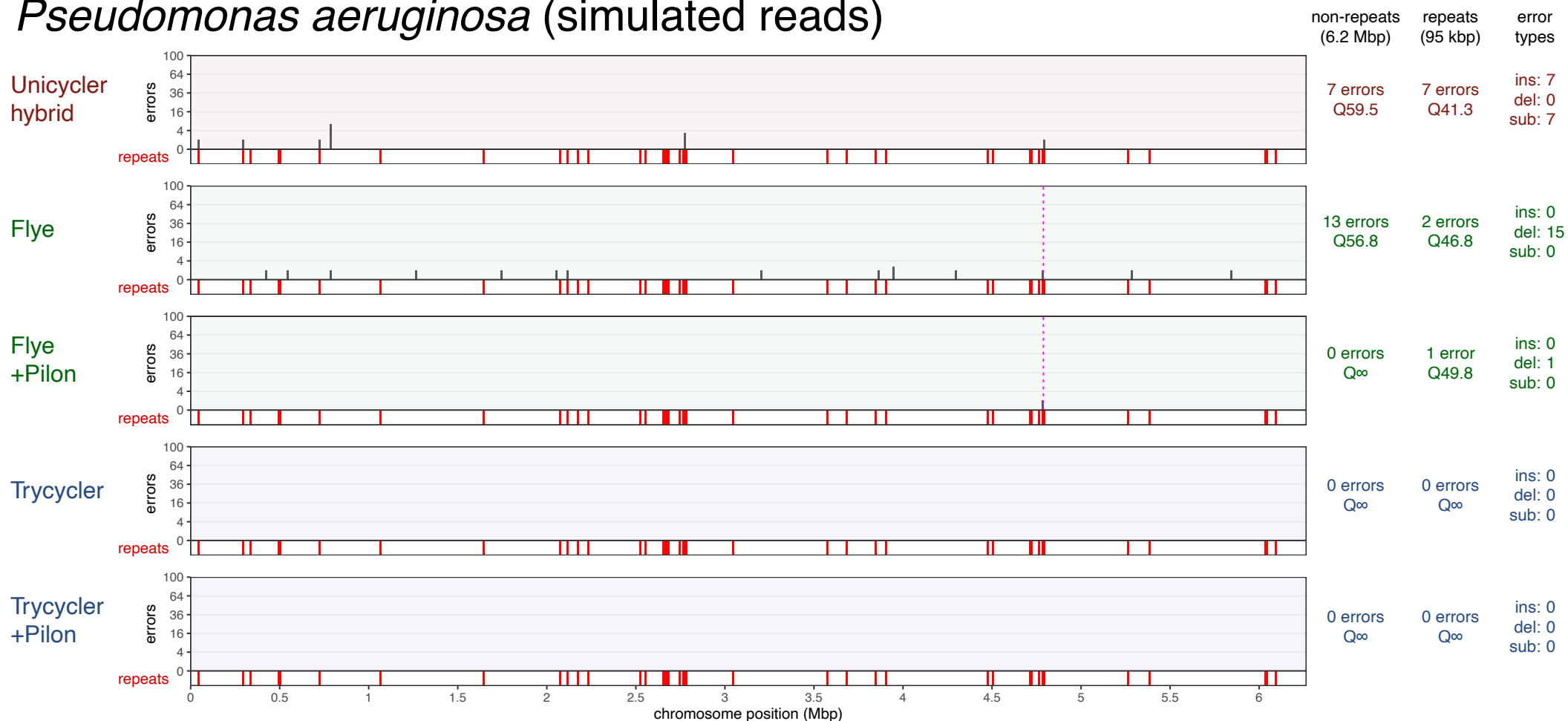


# *Neisseria meningitidis* (simulated reads)



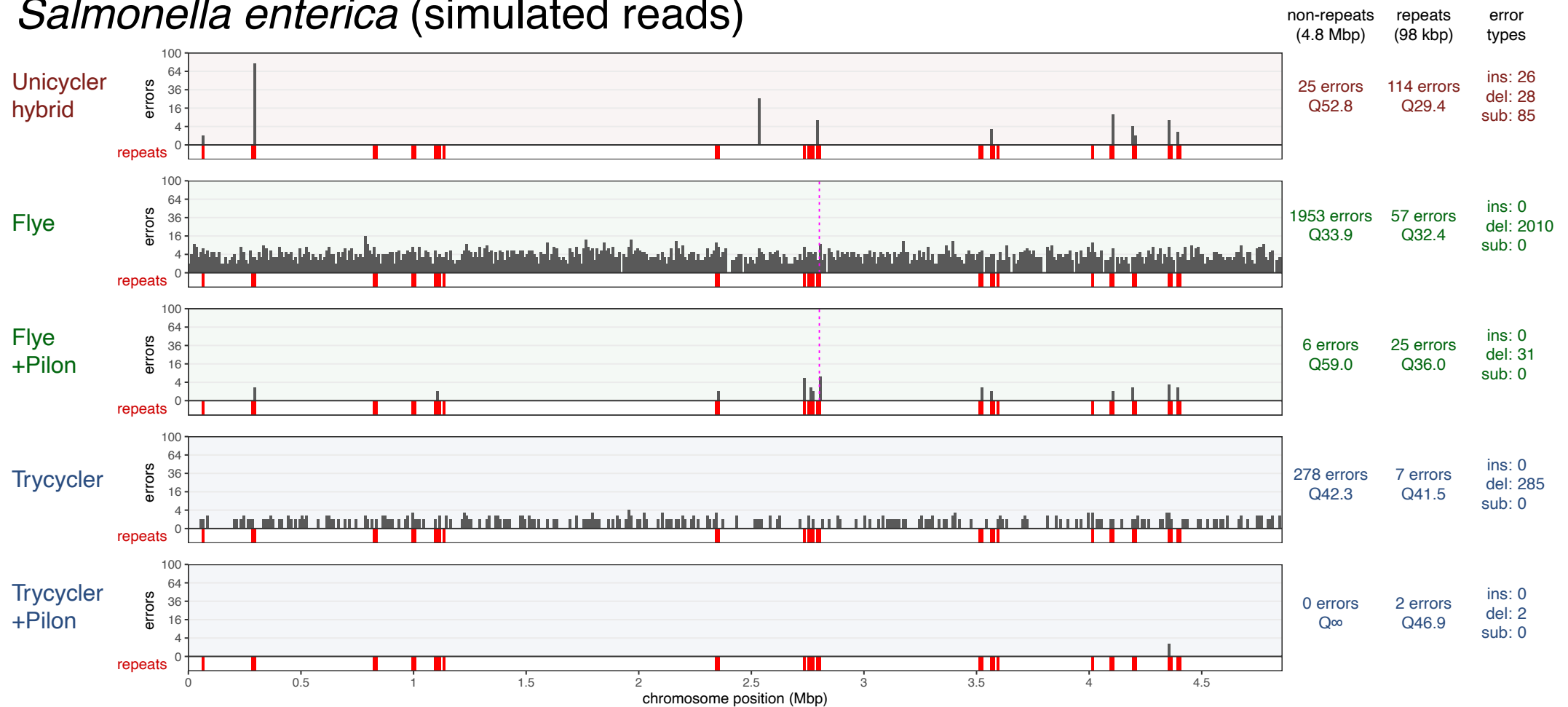
**Fig. S4-f:** error distributions (as defined by alignment to the reference sequence) for the simulated-read assemblies of the *Neisseria meningitidis* chromosome. Repetitive regions of the genome are shown as red blocks at the bottom of each plot. For Flye assemblies, the contig start/end position (where the sequence began before it was rotated to be consistent with the reference) is shown by the dashed line.

# *Pseudomonas aeruginosa* (simulated reads)



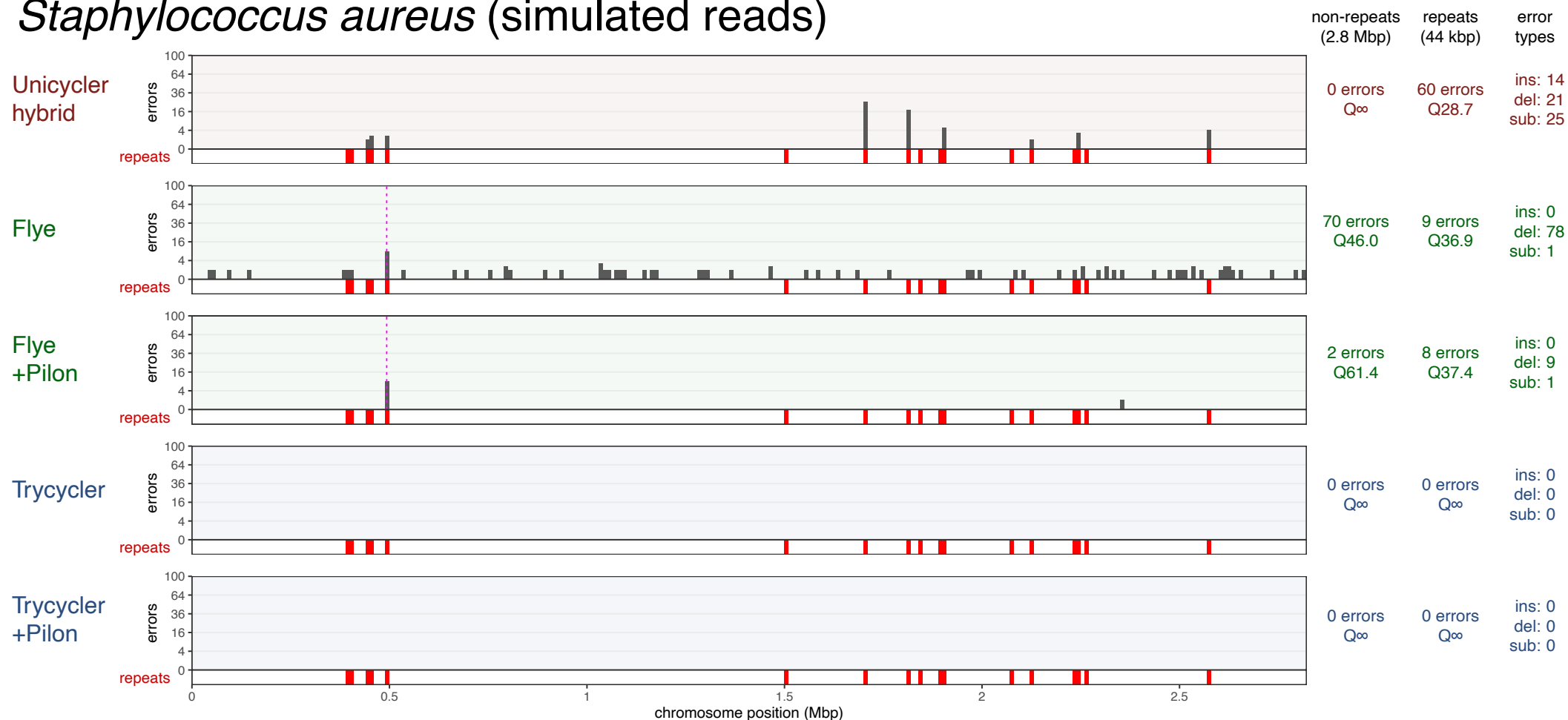
**Fig. S4-g:** error distributions (as defined by alignment to the reference sequence) for the simulated-read assemblies of the *Pseudomonas aeruginosa* chromosome. Repetitive regions of the genome are shown as red blocks at the bottom of each plot. For Flye assemblies, the contig start/end position (where the sequence began before it was rotated to be consistent with the reference) is shown by the dashed line.

# *Salmonella enterica* (simulated reads)



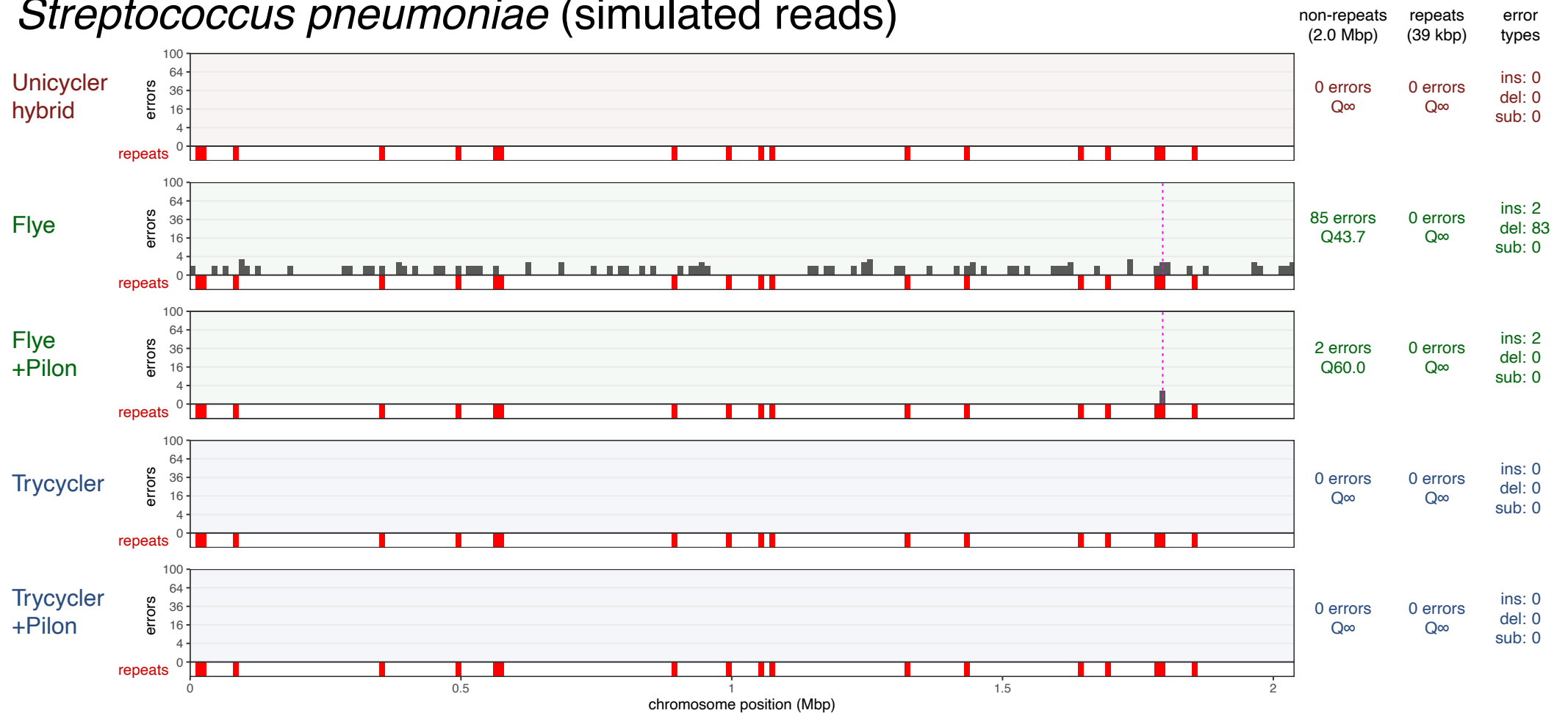
**Fig. S4-h:** error distributions (as defined by alignment to the reference sequence) for the simulated-read assemblies of the *Salmonella enterica* chromosome. Repetitive regions of the genome are shown as red blocks at the bottom of each plot. For Flye assemblies, the contig start/end position (where the sequence began before it was rotated to be consistent with the reference) is shown by the dashed line.

# *Staphylococcus aureus* (simulated reads)



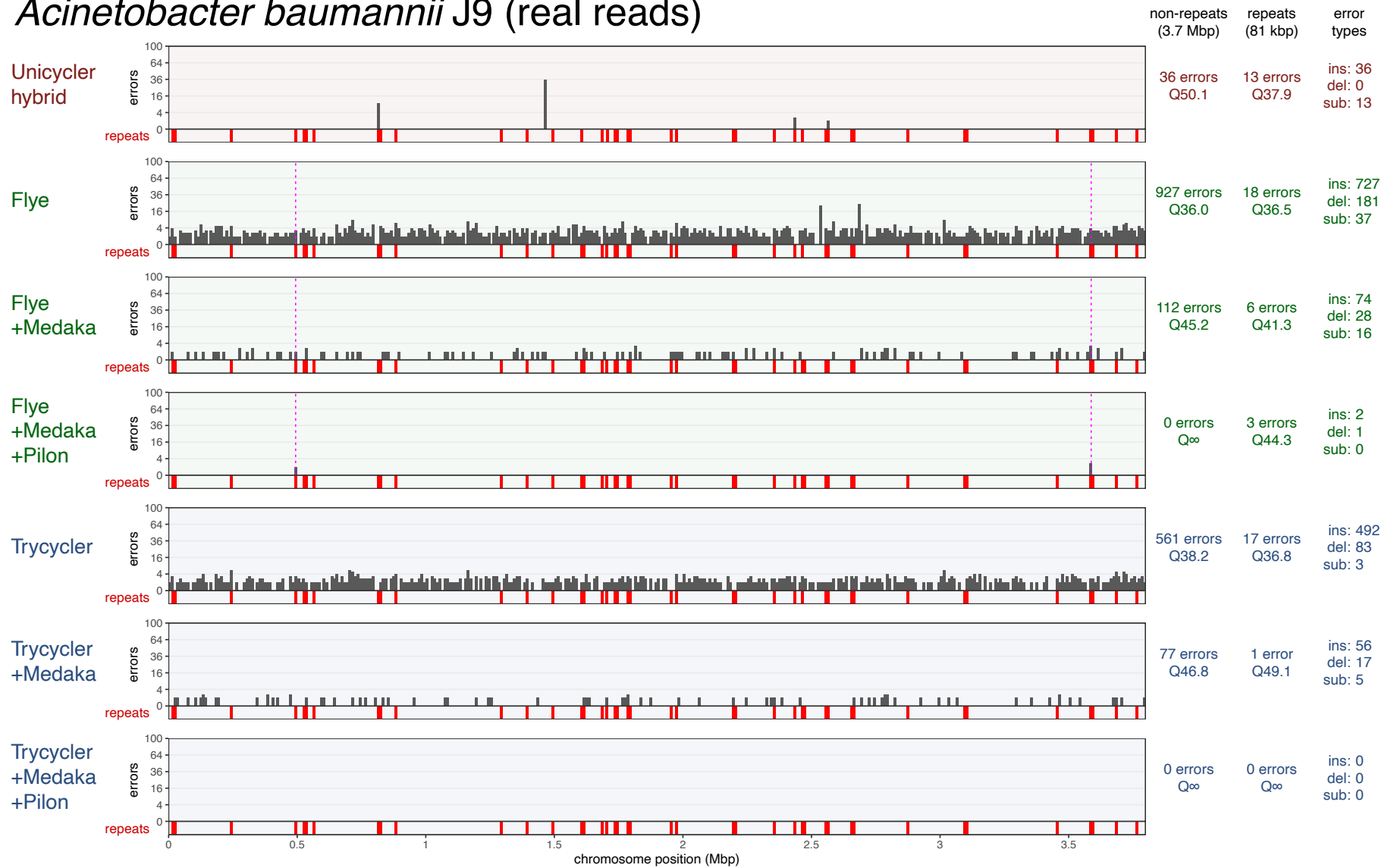
**Fig. S4-i:** error distributions (as defined by alignment to the reference sequence) for the simulated-read assemblies of the *Staphylococcus aureus* chromosome. Repetitive regions of the genome are shown as red blocks at the bottom of each plot. For Flye assemblies, the contig start/end position (where the sequence began before it was rotated to be consistent with the reference) is shown by the dashed line.

# *Streptococcus pneumoniae* (simulated reads)



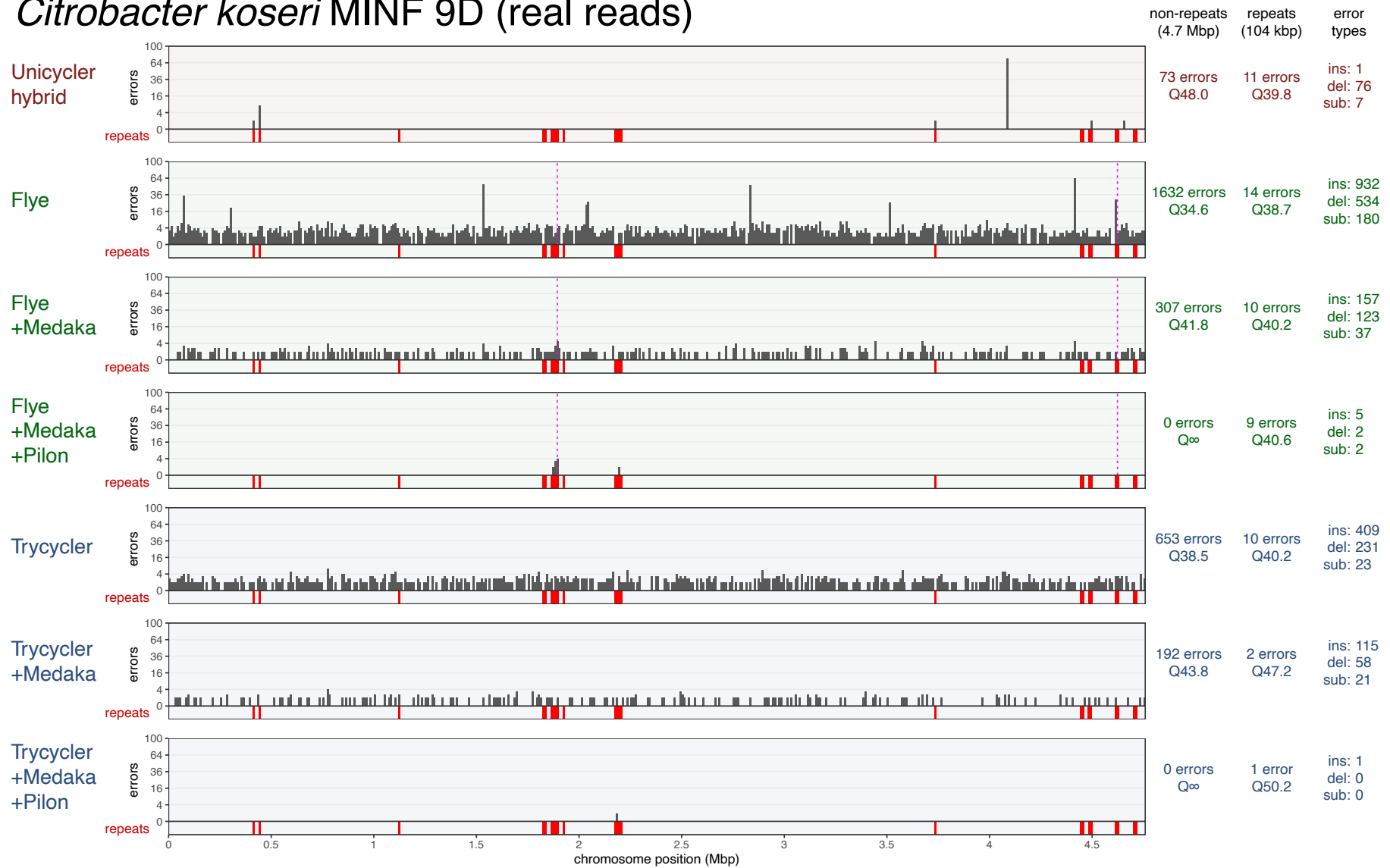
**Fig. S4-j:** error distributions (as defined by alignment to the reference sequence) for the simulated-read assemblies of the *Streptococcus pneumoniae* chromosome. Repetitive regions of the genome are shown as red blocks at the bottom of each plot. For Flye assemblies, the contig start/end position (where the sequence began before it was rotated to be consistent with the reference) is shown by the dashed line.

# *Acinetobacter baumannii* J9 (real reads)



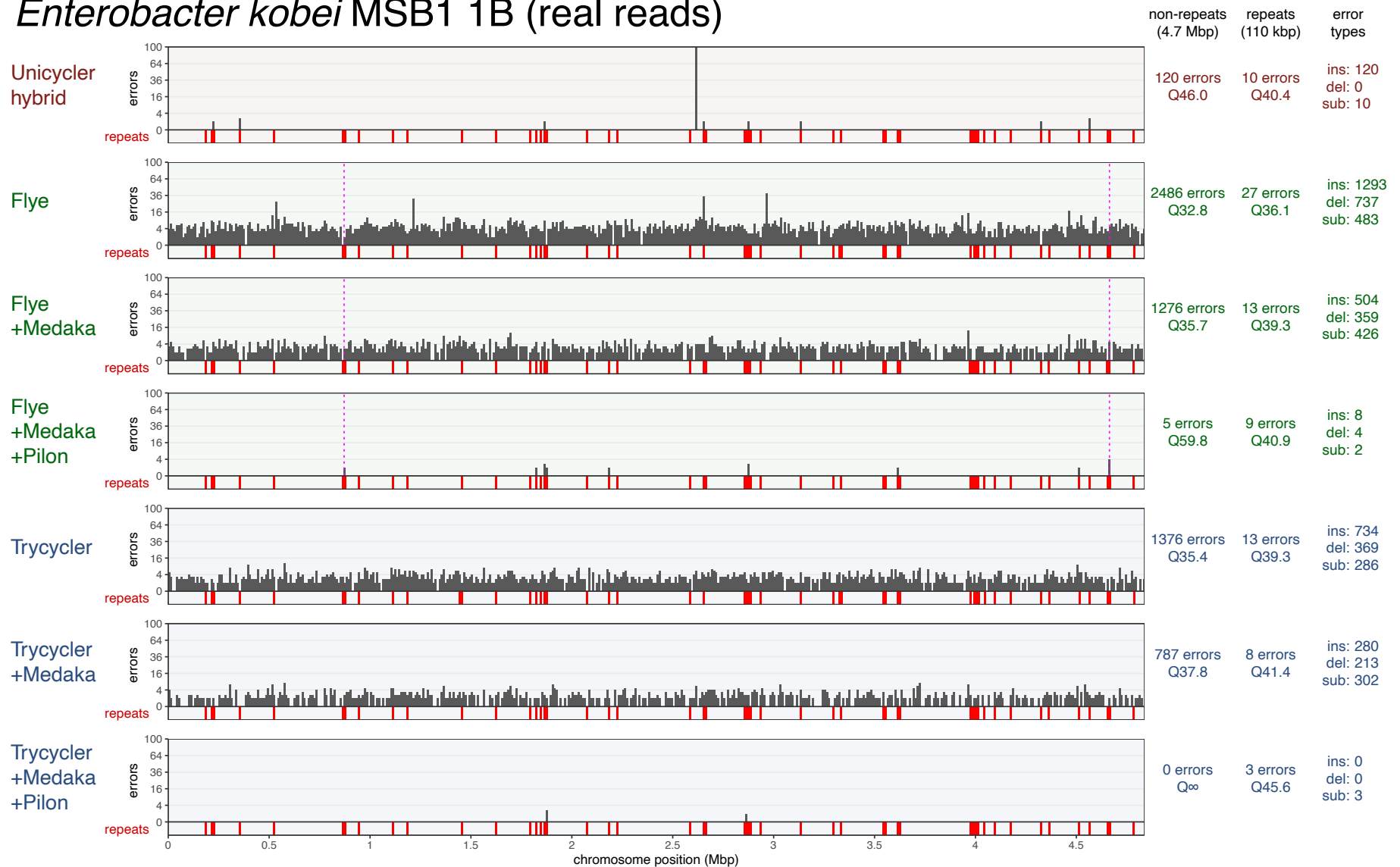
**Fig. S4-k:** error distributions (as defined by alignment between two independent assemblies) for the real-read assemblies of the *Acinetobacter baumannii* chromosome. Repetitive regions of the genome are shown as red blocks at the bottom of each plot. For Flye assemblies, the contig start/end positions (where the sequences began before they were rotated to be consistent with the reference) are shown by the dashed lines.

# Citrobacter koseri MINF 9D (real reads)



**Fig. S4-1:** error distributions (as defined by alignment between two independent assemblies) for the real-read assemblies of the *Citrobacter koseri* chromosome. Repetitive regions of the genome are shown as red blocks at the bottom of each plot. For Flye assemblies, the contig start/end positions (where the sequences began before they were rotated to be consistent with the reference) are shown by the dashed lines.

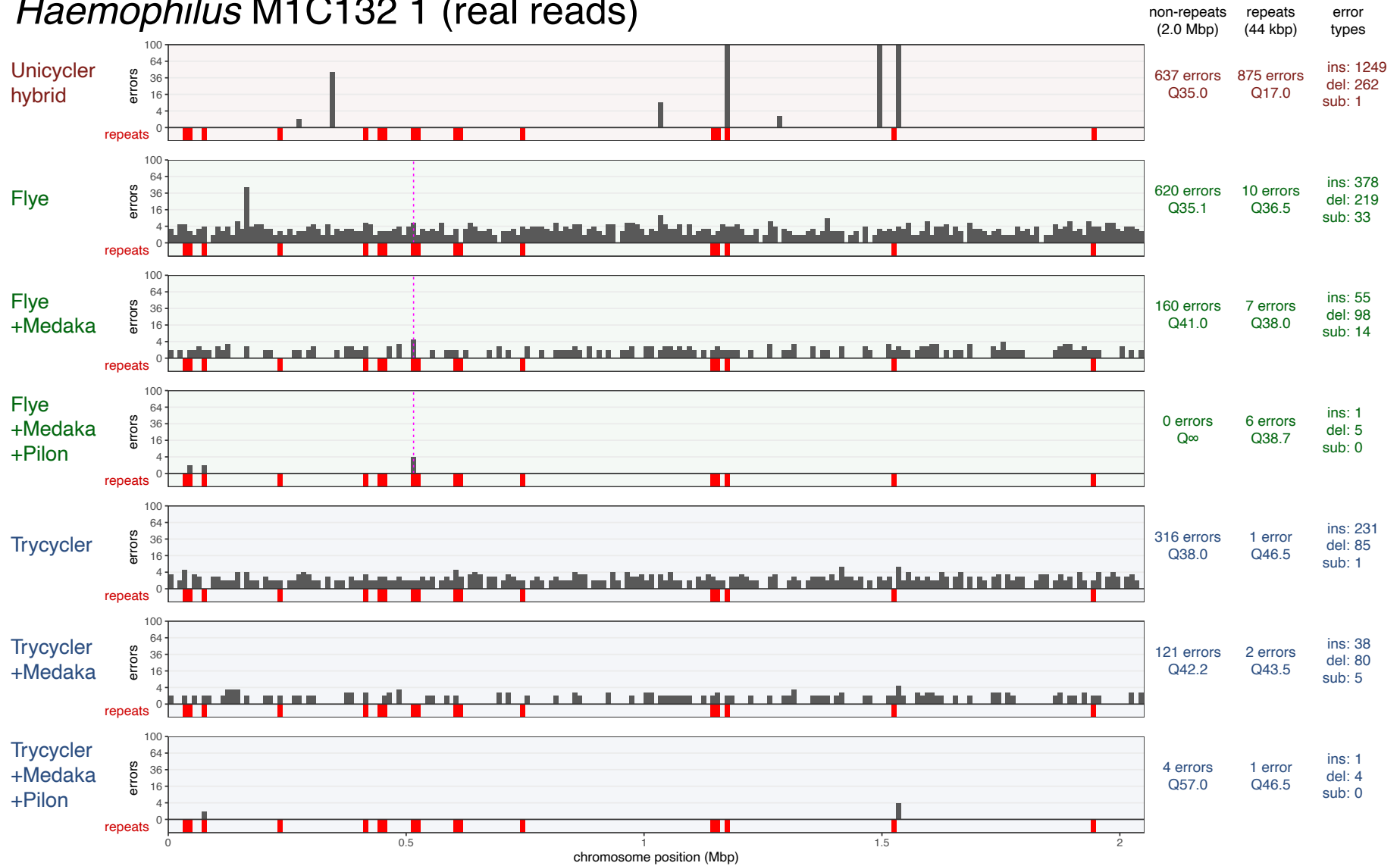
# Enterobacter kobei MSB1 1B (real reads)



**Fig. S4-m:** error distributions (as defined by alignment between two independent assemblies) for the real-read assemblies of the *Enterobacter kobei* chromosome. Repetitive regions of the genome are shown as red blocks at the bottom of each plot. For Flye assemblies, the contig start/end positions (where the sequences began before they were rotated to be consistent with the reference) are shown by the dashed lines.

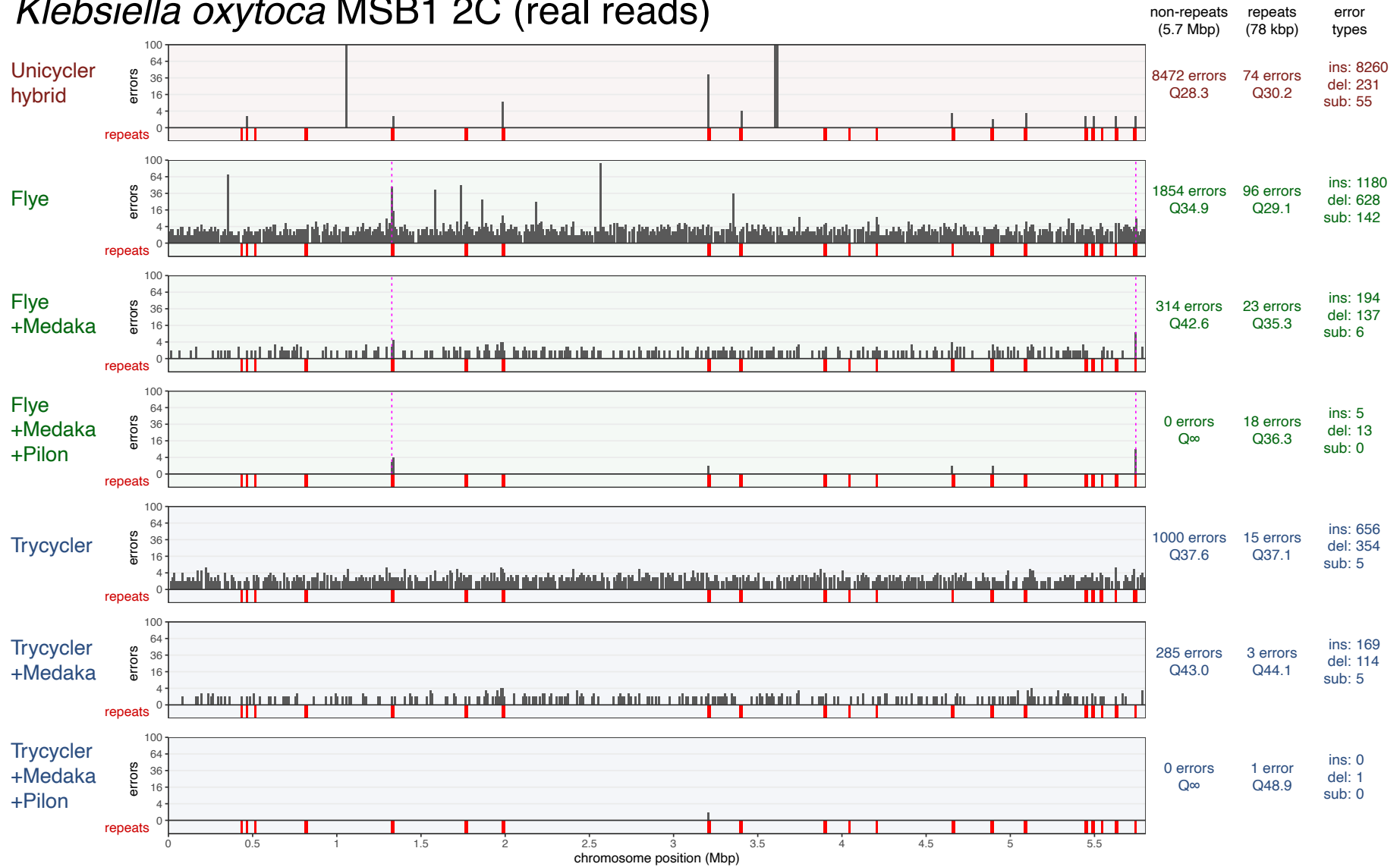


# Haemophilus M1C132 1 (real reads)



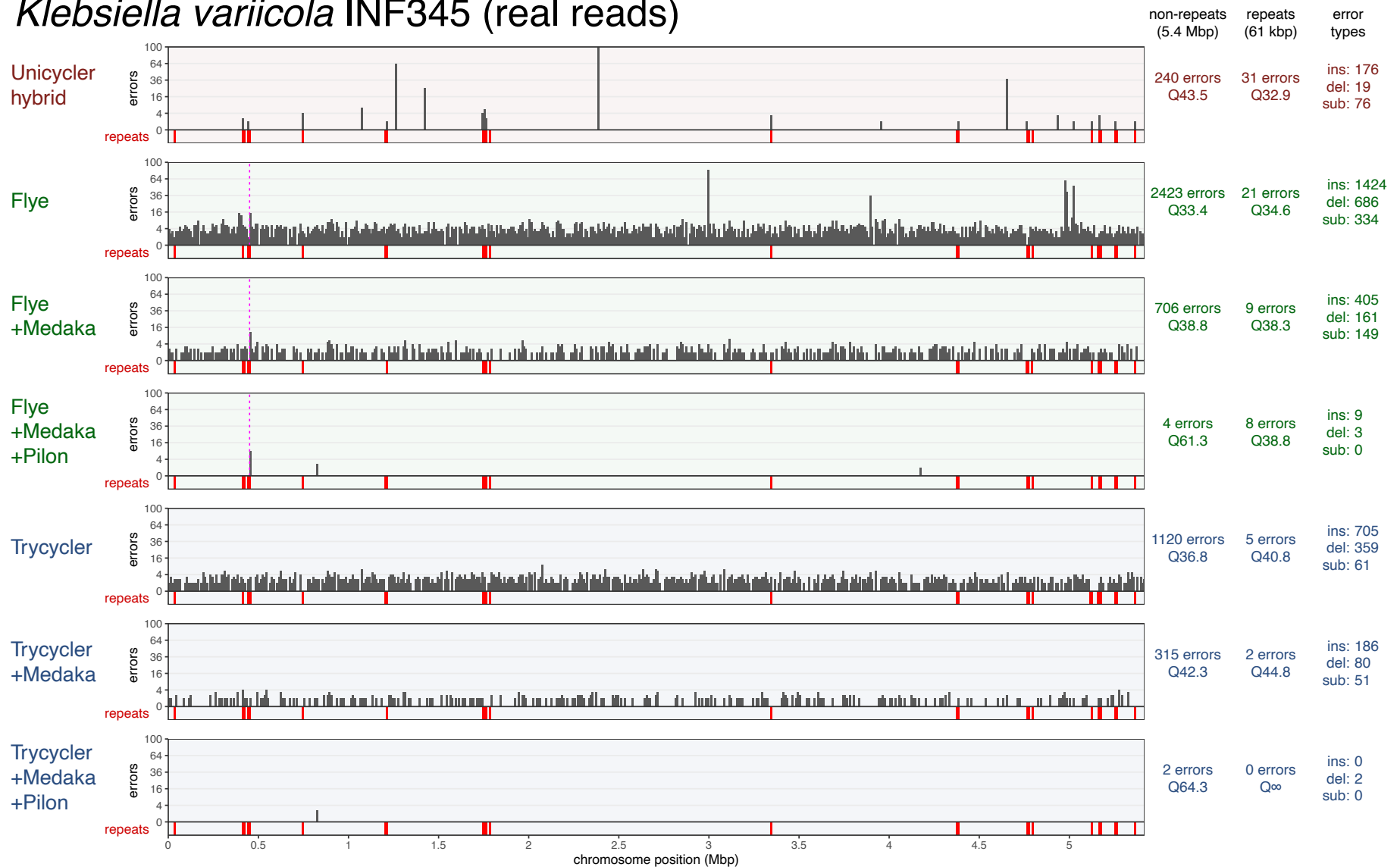
**Fig. S4-n:** error distributions (as defined by alignment between two independent assemblies) for the real-read assemblies of the *Haemophilus* chromosome. Repetitive regions of the genome are shown as red blocks at the bottom of each plot. For Flye assemblies, the contig start/end positions (where the sequences began before they were rotated to be consistent with the reference) are shown by the dashed lines.

# *Klebsiella oxytoca* MSB1 2C (real reads)

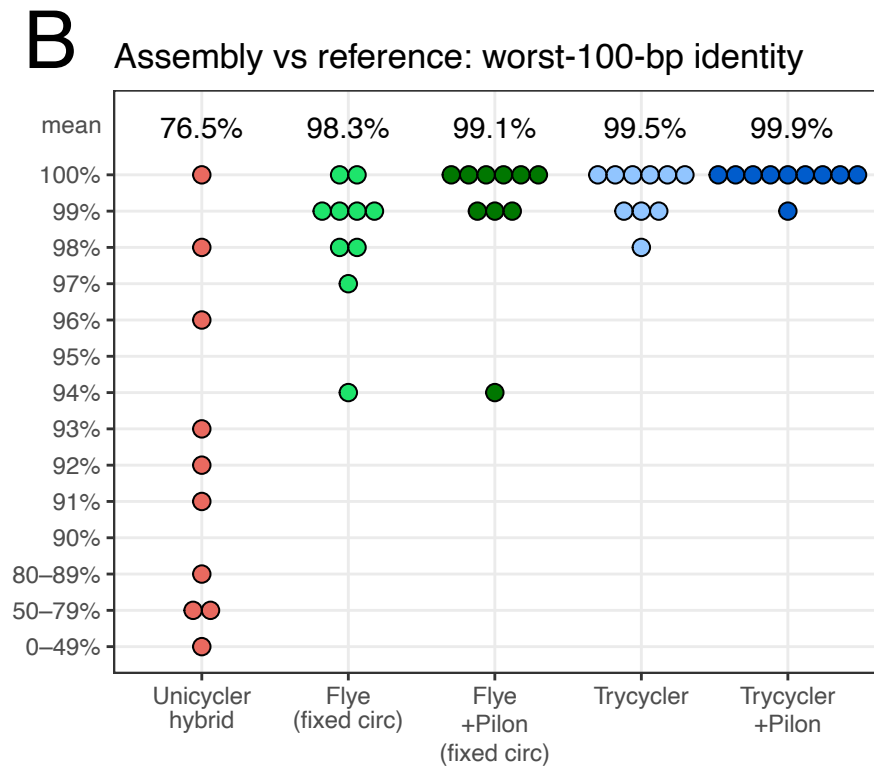
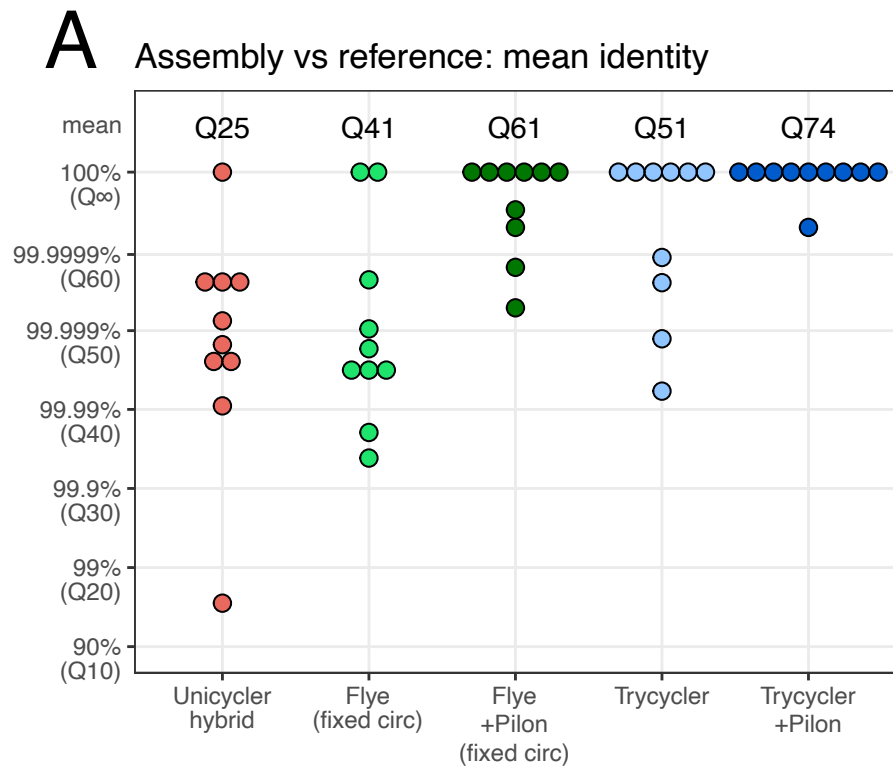


**Fig. S4-o:** error distributions (as defined by alignment between two independent assemblies) for the real-read assemblies of the *Klebsiella oxytoca* chromosome. Repetitive regions of the genome are shown as red blocks at the bottom of each plot. For Flye assemblies, the contig start/end positions (where the sequences began before they were rotated to be consistent with the reference) are shown by the dashed lines.

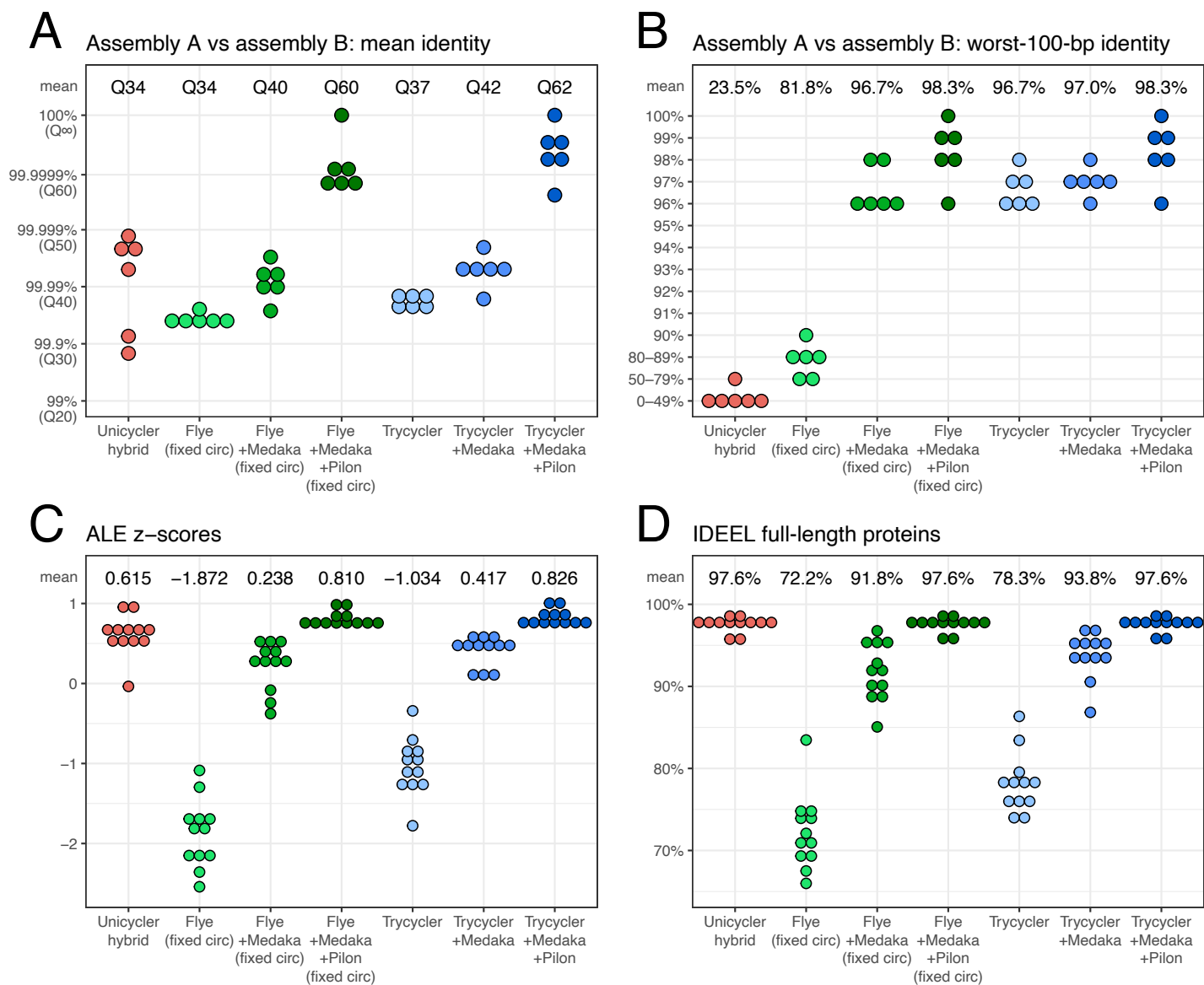
# *Klebsiella variicola* INF345 (real reads)



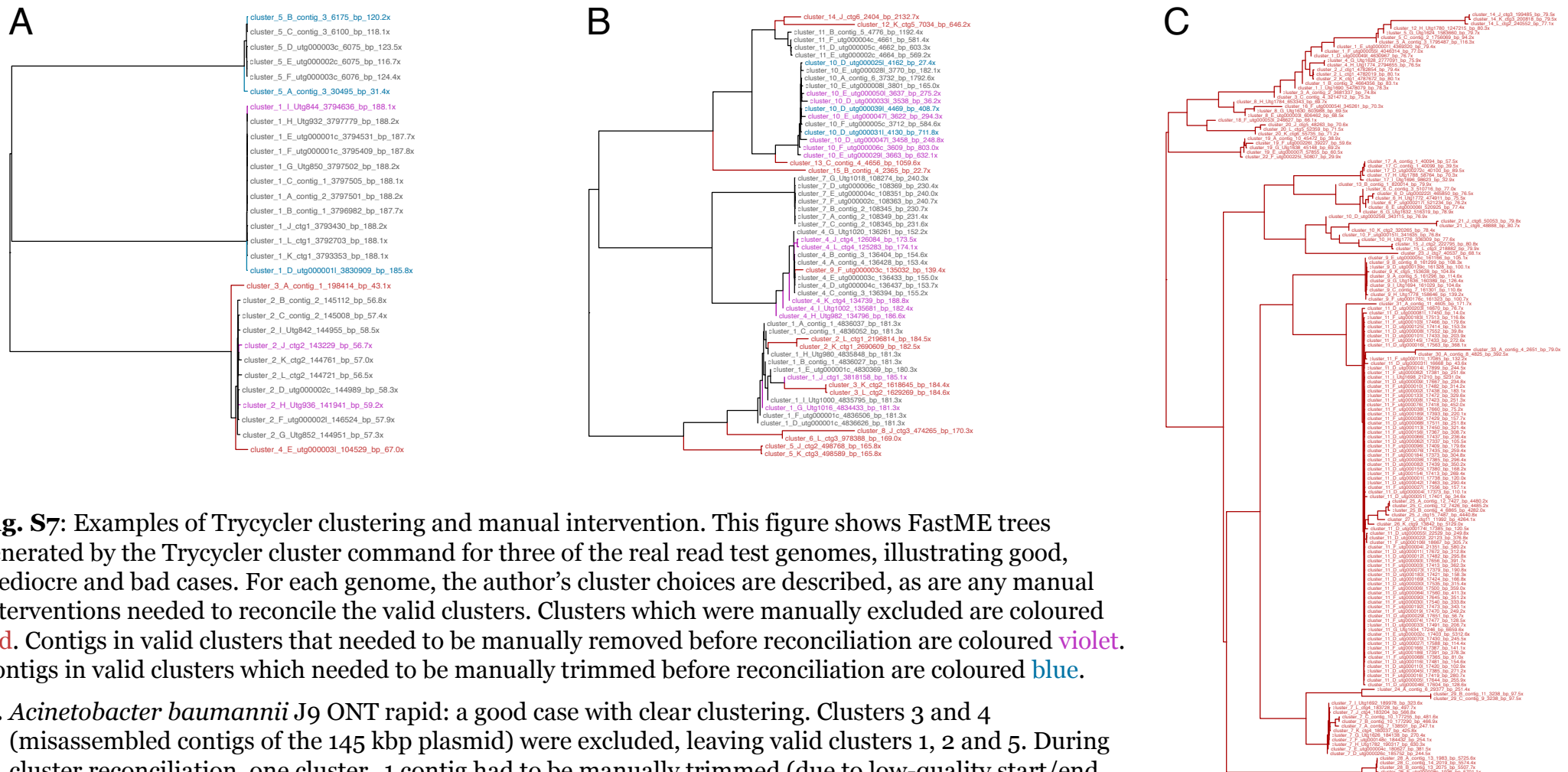
**Fig. S4-p:** error distributions (as defined by alignment between two independent assemblies) for the real-read assemblies of the *Klebsiella variicola* chromosome. Repetitive regions of the genome are shown as red blocks at the bottom of each plot. For Flye assemblies, the contig start/end positions (where the sequences began before they were rotated to be consistent with the reference) are shown by the dashed lines.



**Fig. S5:** results for the simulated read tests. This figure contains the same analyses as are shown in Figure 2, but the Flye assemblies have had their circularisation manually repaired.



**Fig. S6:** results for the real read tests. This figure contains the same analyses as are shown in Figure 3, but the Flye assemblies have had their circularisation manually repaired.



**Fig. S7:** Examples of Trycycler clustering and manual intervention. This figure shows FastME trees generated by the Trycycler cluster command for three of the real read test genomes, illustrating good, mediocre and bad cases. For each genome, the author's cluster choices are described, as are any manual interventions needed to reconcile the valid clusters. Clusters which were manually excluded are coloured red. Contigs in valid clusters that needed to be manually removed before reconciliation are coloured violet. Contigs in valid clusters which needed to be manually trimmed before reconciliation are coloured blue.

- A. *Acinetobacter baumannii* J9** ONT rapid: a good case with clear clustering. Clusters 3 and 4 (misassembled contigs of the 145 kbp plasmid) were excluded, leaving valid clusters 1, 2 and 5. During cluster reconciliation, one cluster\_1 contig had to be manually trimmed (due to low-quality start/end sequence), one cluster\_1 contig had to be manually removed (due to poor pairwise alignment), two cluster\_2 contigs had to be manually removed (due to being incomplete) and two cluster\_5 contigs had to be manually trimmed (due to excessive length).
- B. *Enterobacter kobei* MSB1\_1B** ONT rapid: a mediocre case with more complex clusters. Clusters 2, 3, 5, 6, 8 (misassembled contigs of the chromosome), 9 (misassembled contig of the 136 kbp plasmid), 12, 13, 14, and 15 (misassembled contigs of the small plasmids) were excluded, leaving valid clusters 1, 4, 7, 10 and 11. During cluster reconciliation, two cluster\_1 contigs had to be manually removed (due to being incomplete or poor pairwise alignment), five cluster\_4 contigs had to be manually removed (due to being incomplete), three cluster\_10 contigs had to be manually trimmed (due to excessive length) and six cluster\_10 contigs had to be manually removed (due to being incomplete or unable to circularise).
- C. *Serratia marcescens* 17-147-1671** ONT rapid: a bad case where valid clusters were unclear. Insufficient read length and genome heterogeneity both contributed to the poor results. Without good clusters, it was not possible to proceed with Trycycler assembly.