

Supplementary Materials for

**Switchable immune modulator for tumor-specific activation
of anticancer immunity**

Yu Zhao, Yu-Qing Xie, Simon Van Herck, Sina Nassiri, Min Gao,
Yugang Guo, Li Tang*

*Corresponding author. Email: li.tang@epfl.ch

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Figs. S1 to S16
Table S1

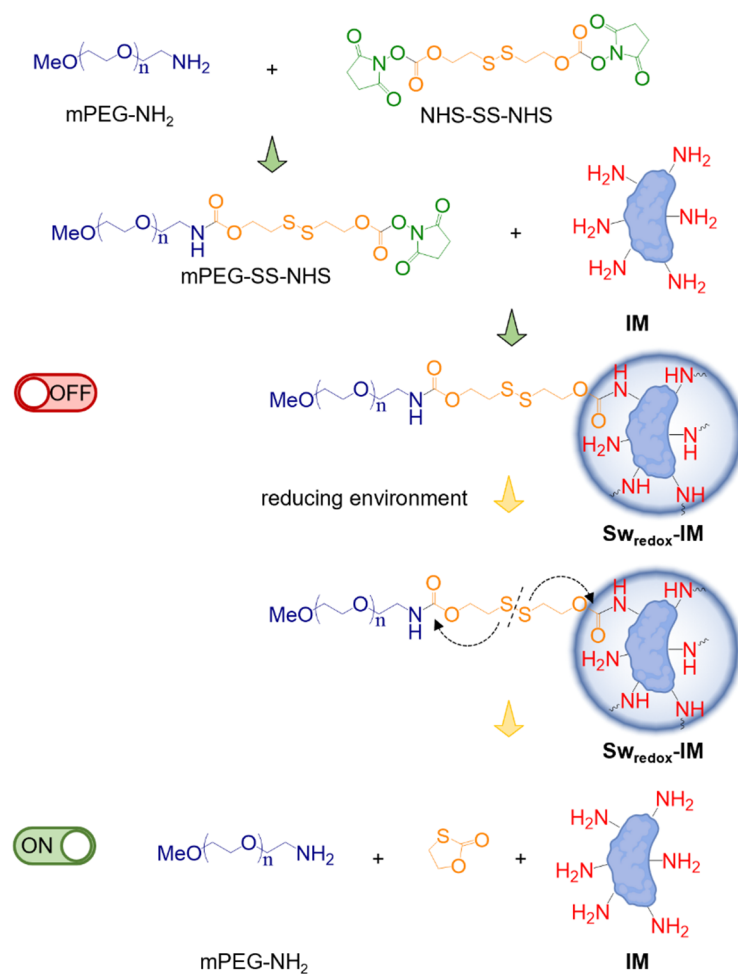


Fig. S1. Synthesis of redox-responsive switchable immune modulators (Sw_{redox}IMs) and selective switch on in the reducing environment. mPEG-NH₂, methoxy polyethylene glycol amine; NHS-SS-NHS, a redox-responsive linker bis(2,5-dioxopyrrolidin-1-yl) (disulfanediylbis(ethane-2,1-diyl) dicarbonate).

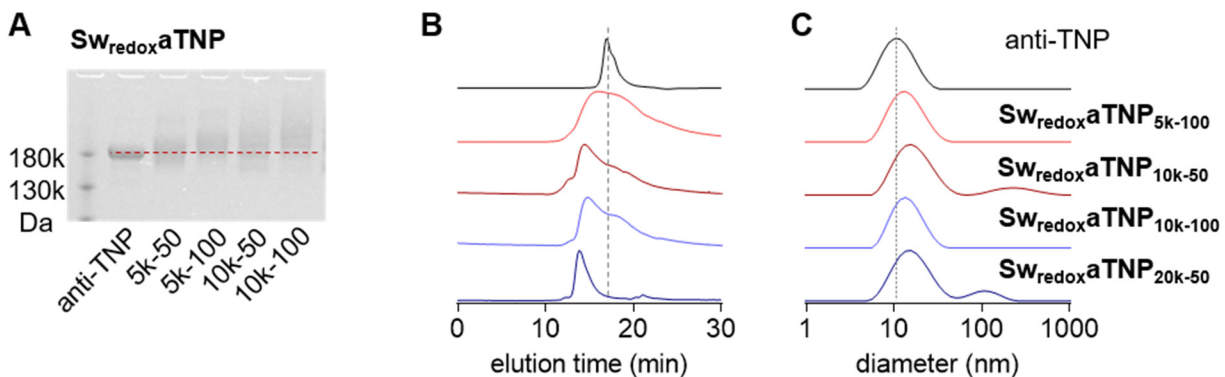


Fig. S2. Characterizations of $Sw_{redox}aTNPs$. (A) Sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) characterizations of $Sw_{redox}aTNPs$. (B) Ultra high-performance liquid chromatography (UHPLC) traces of $Sw_{redox}aTNPs$ with a size exclusion chromatography (SEC) column. (C) Hydrodynamic diameters of $Sw_{redox}aTNPs$ measured by dynamic light scattering (DLS). Dash lines indicate the molecular weight (A), elution time (B), and hydrodynamic diameter (C) of native anti-trinitrophenal (TNP) antibody.

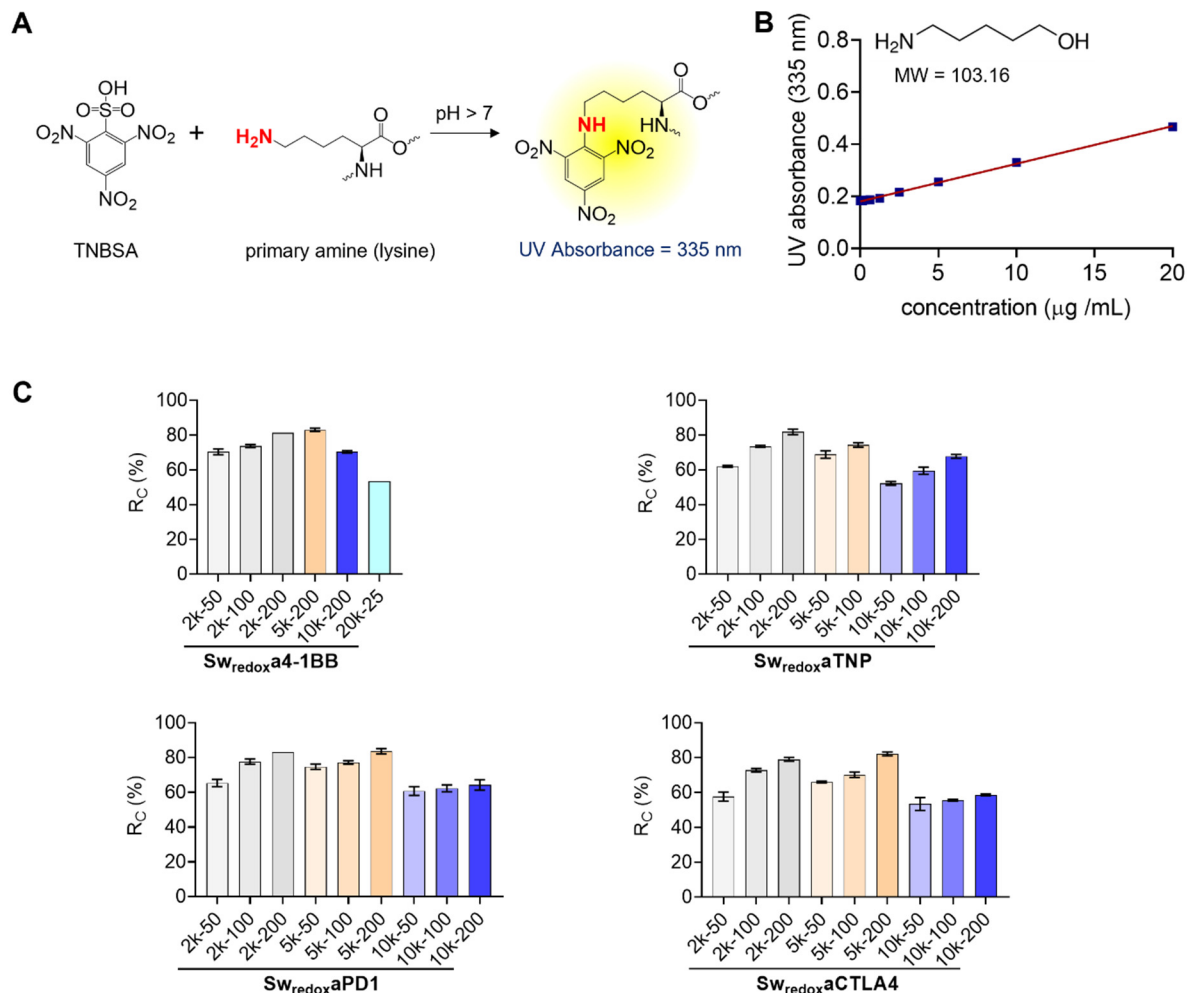


Fig. S3. Conjugation ratio (R_c) of Sw-IMs. (A) Schematic illustration of 2,4,6-trinitrobenzene sulfonic acid (TNBSA) assay for quantifying the number of primary amino groups. (B) Calibration curve using 5-amino-1-pentanol as a standard in TNBSA assay. (C) Conjugation ratio (R_c) of **Sw_{redox}a4-1BBs**, **Sw_{redox}aTNPs**, **Sw_{redox}aPD1s**, and **Sw_{redox}aCTLA4s**. R_c defined as the percentage of PEG conjugated amino groups among all the detectable amino groups in the molecule.

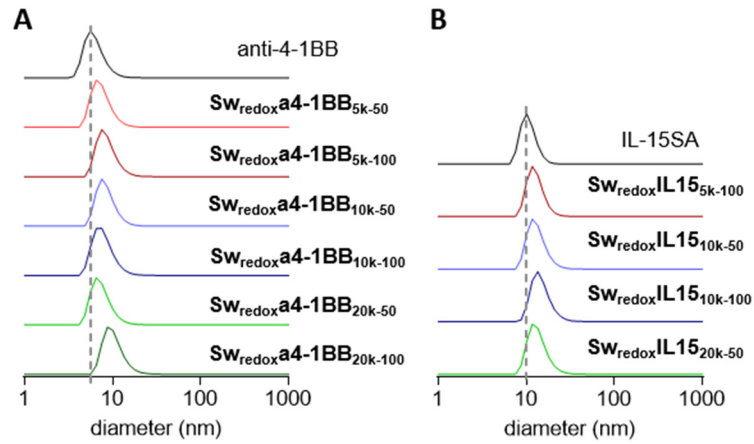


Fig. S4. DLS characterizations of Sw_{redox} IMs. (A) Hydrodynamic diameters of $Sw_{redox}a4-1BB$ s. (B) Hydrodynamic diameters of $Sw_{redox}IL15$ s. Dash lines indicate the size of the native antibody or cytokine.

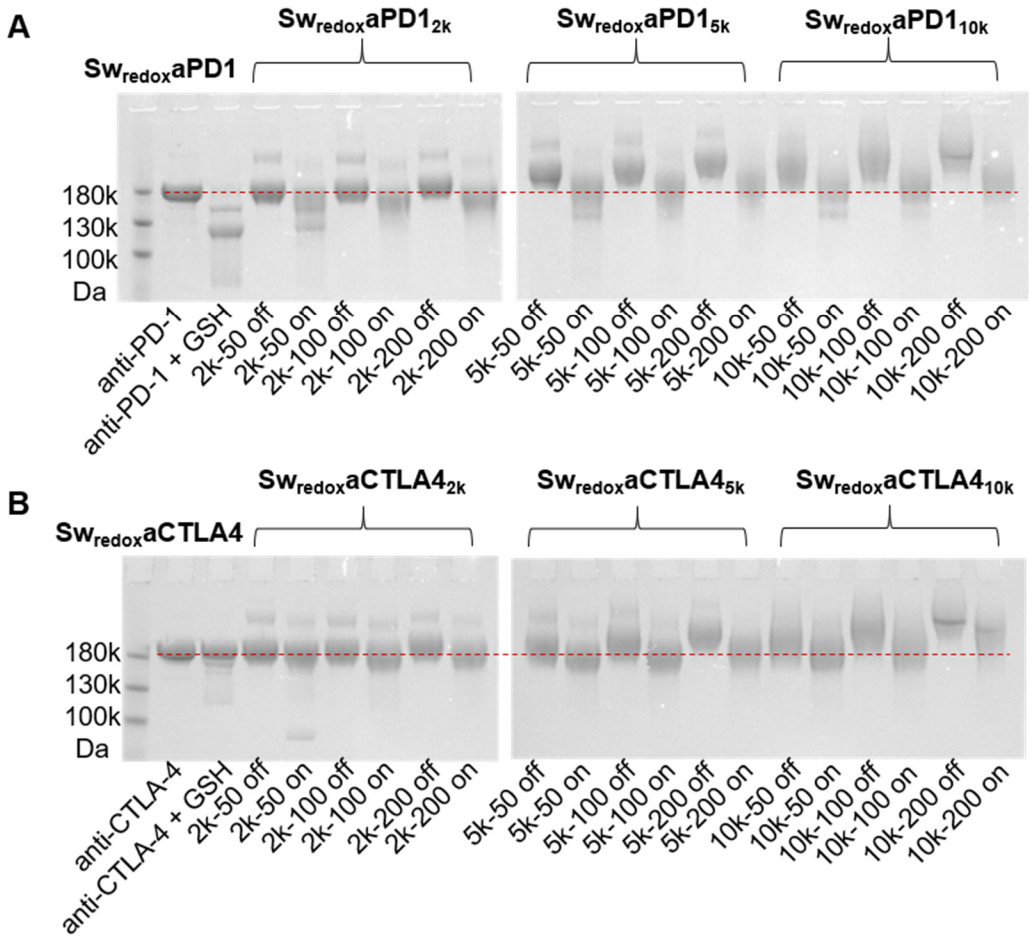


Fig. S5. SDS-PAGE characterizations of Sw_{redox}aPD1s (A) and Sw_{redox}aCTLA4s (B) at off and on status. Dash lines indicate the molecular weight of native antibodies.

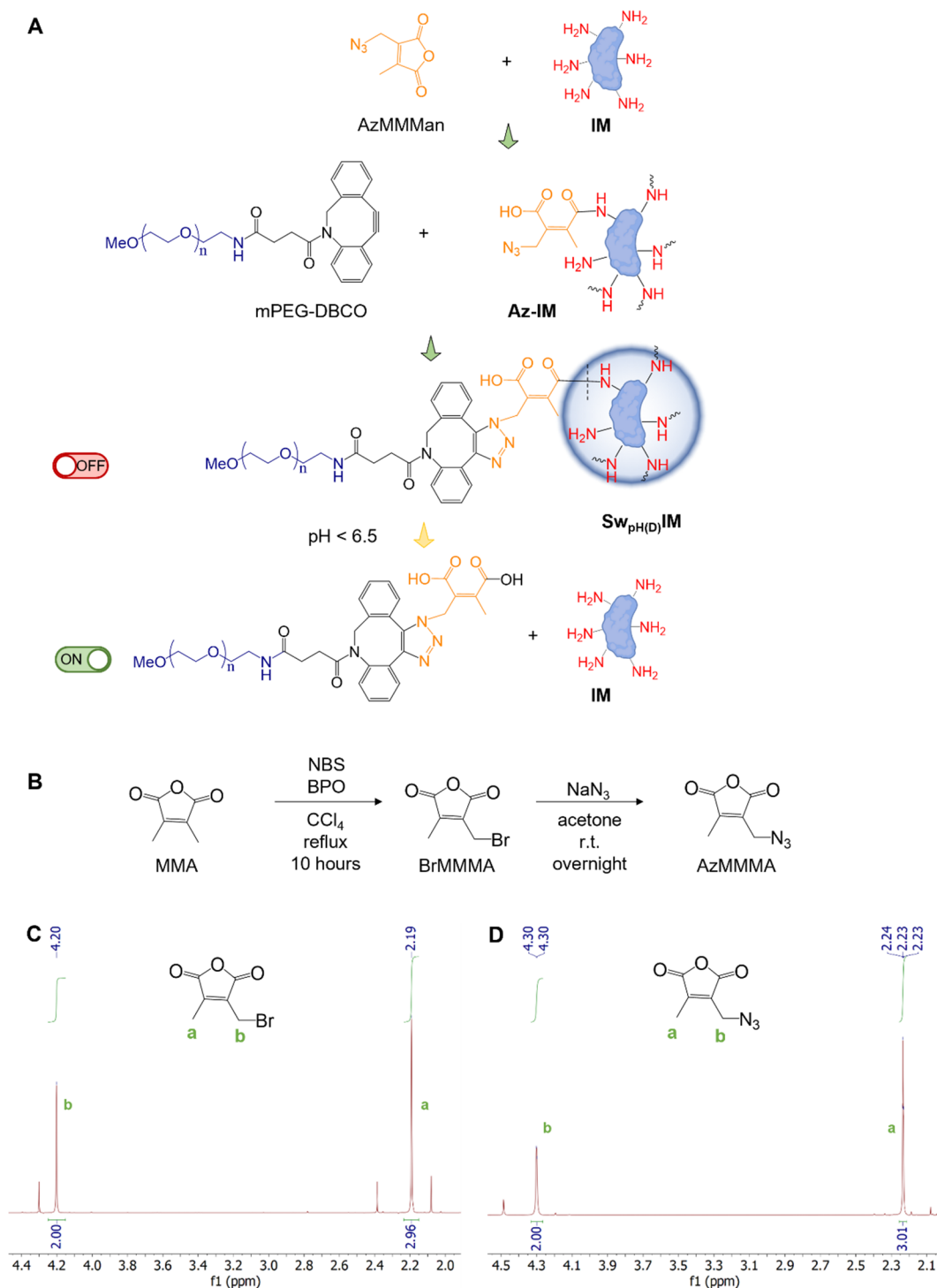


Fig. S6. Preparation of $Sw_{pH(D)}IMs$. (A) Schematic illustration of the preparation and switch on of $Sw_{pH(D)}IM$. mPEG-DBCO, dibenzocyclooctyne-functionalized mPEG; AzMMMan, (azidomethyl)methylmaleic anhydride; **Az-IM**, azido-functionalized **IM**. The dash line indicates the cleavable bond in acidic environment. (B) Synthesis of pH-responsive linker AzMMMan. (C) 1H NMR spectrum of the intermediate product (bromomethyl)methylmaleic anhydride (BrMMMan), (400 MHz, $CDCl_3$) 4.20 (s, 2H, CH_2Br), 2.19 (s, 3H, CH_3). (D) 1H NMR spectrum of AzMMMan, (400 MHz, $CDCl_3$) 4.29 (d, $J < 1.0$ Hz, 2H, CH_2N_3), 2.23 (t, $J = 1.0$ Hz, 3H, CH_3).

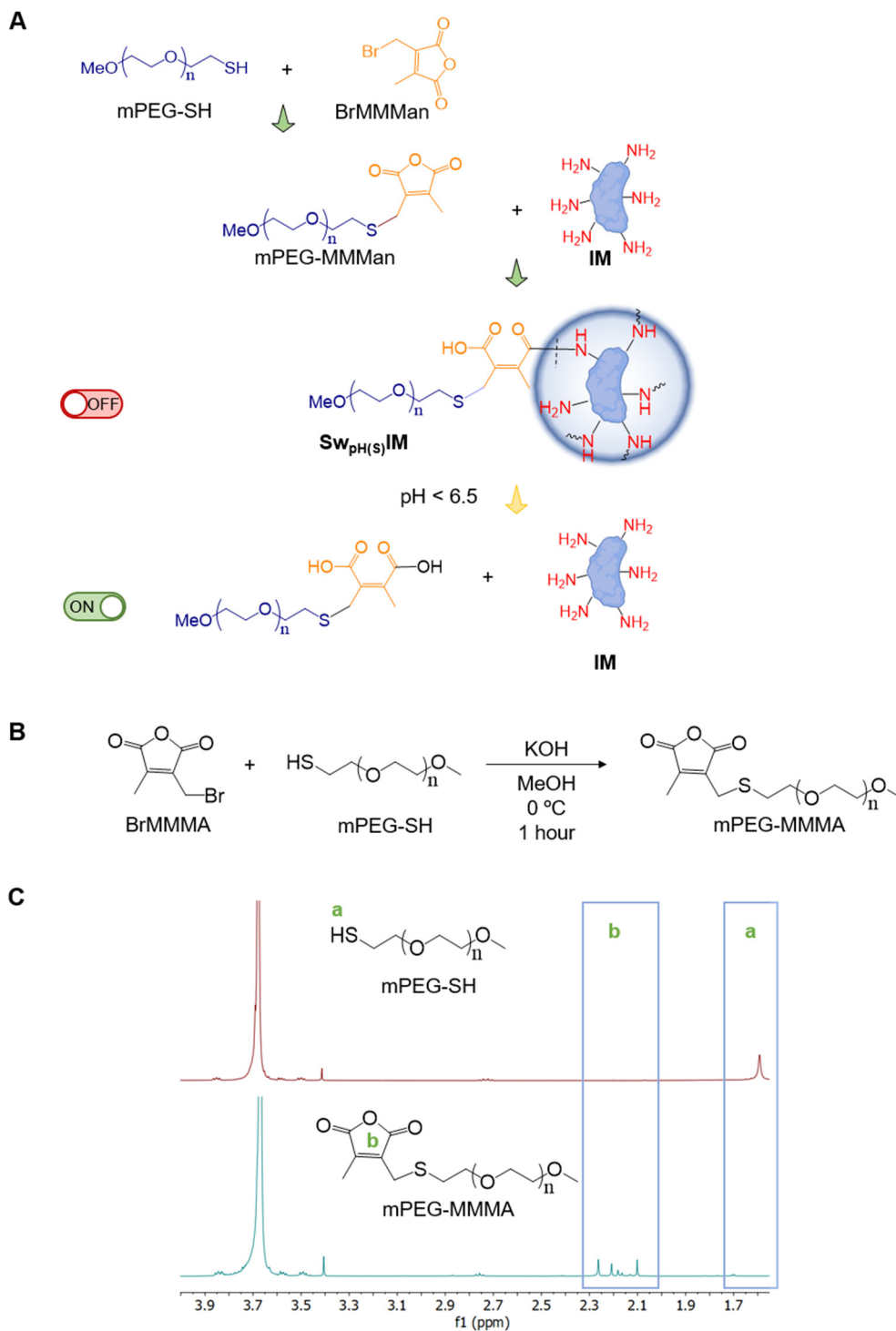


Fig. S7. Preparation of $Sw_{pH(s)}IMs$. (A) Schematic illustration of the preparation and switch on of $Sw_{pH(s)}IM$, mPEG-SH, mPEG thiol, the dash line indicates the cleavable bond in acidic environment. (B) Synthesis of MMA functionalized PEG, mPEG-MMMA. (C) 1H NMR spectra of mPEG-SH and mPEG-MMMA. Blue boxes highlight the characteristic peaks of MMA moieties and thiol groups.

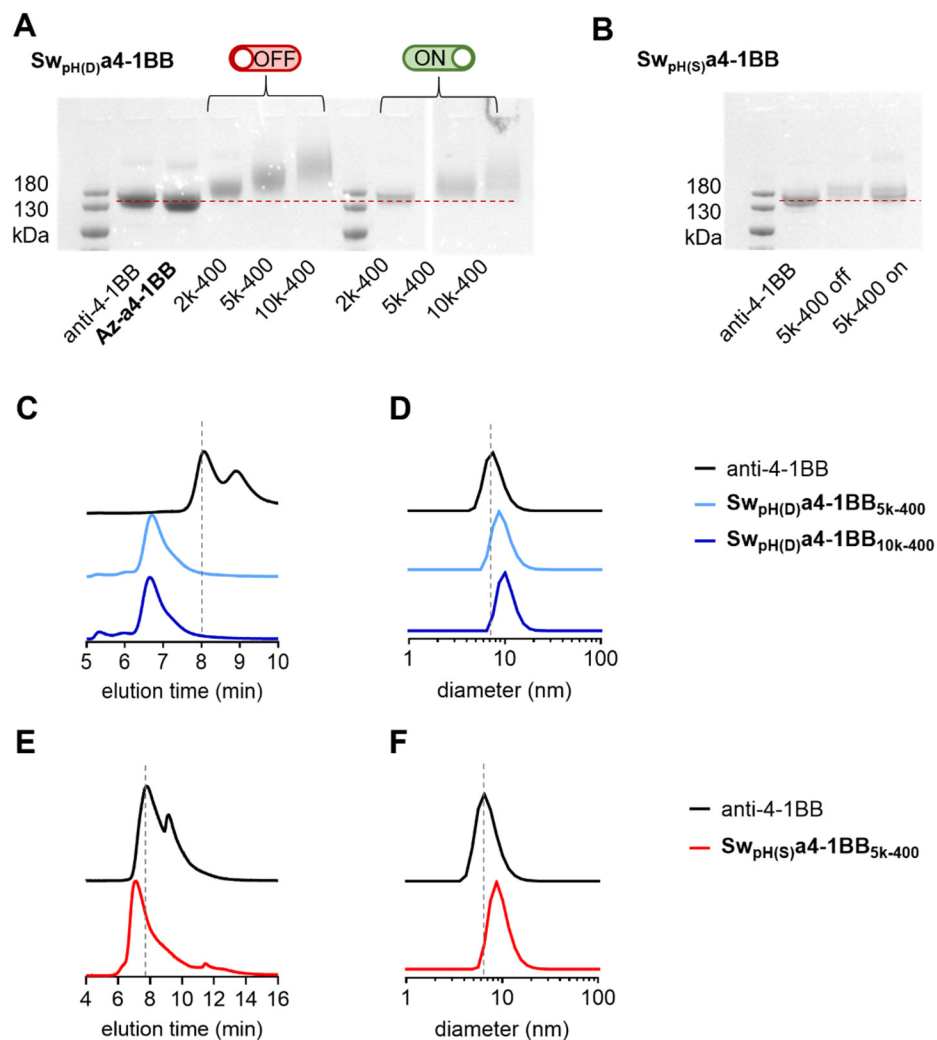


Fig. S8. Characterizations of acidic pH-responsive Sw-IMs. (A) SDS-PAGE characterization of $Sw_{pH(D)}a4-1BB$ s at off and on status. **Az-a4-1BB**, azido-functionalized anti-4-1BB. (B) SDS-PAGE characterization of $Sw_{pH(S)}a4-1BB$ at off and on status. (C) UHPLC-SEC traces of $Sw_{pH(D)}a4-1BB$ s. (D) Hydrodynamic diameters of $Sw_{pH(D)}a4-1BB$ s measured by DLS. (E) UHPLC-SEC traces of $Sw_{pH(S)}a4-1BB_{5k-400}$. (F) Hydrodynamic diameter of $Sw_{pH(S)}a4-1BB_{5k-400}$ measured by DLS. Dash lines indicate the molecular weight, elution time, and hydrodynamic diameter of native anti-4-1BB.

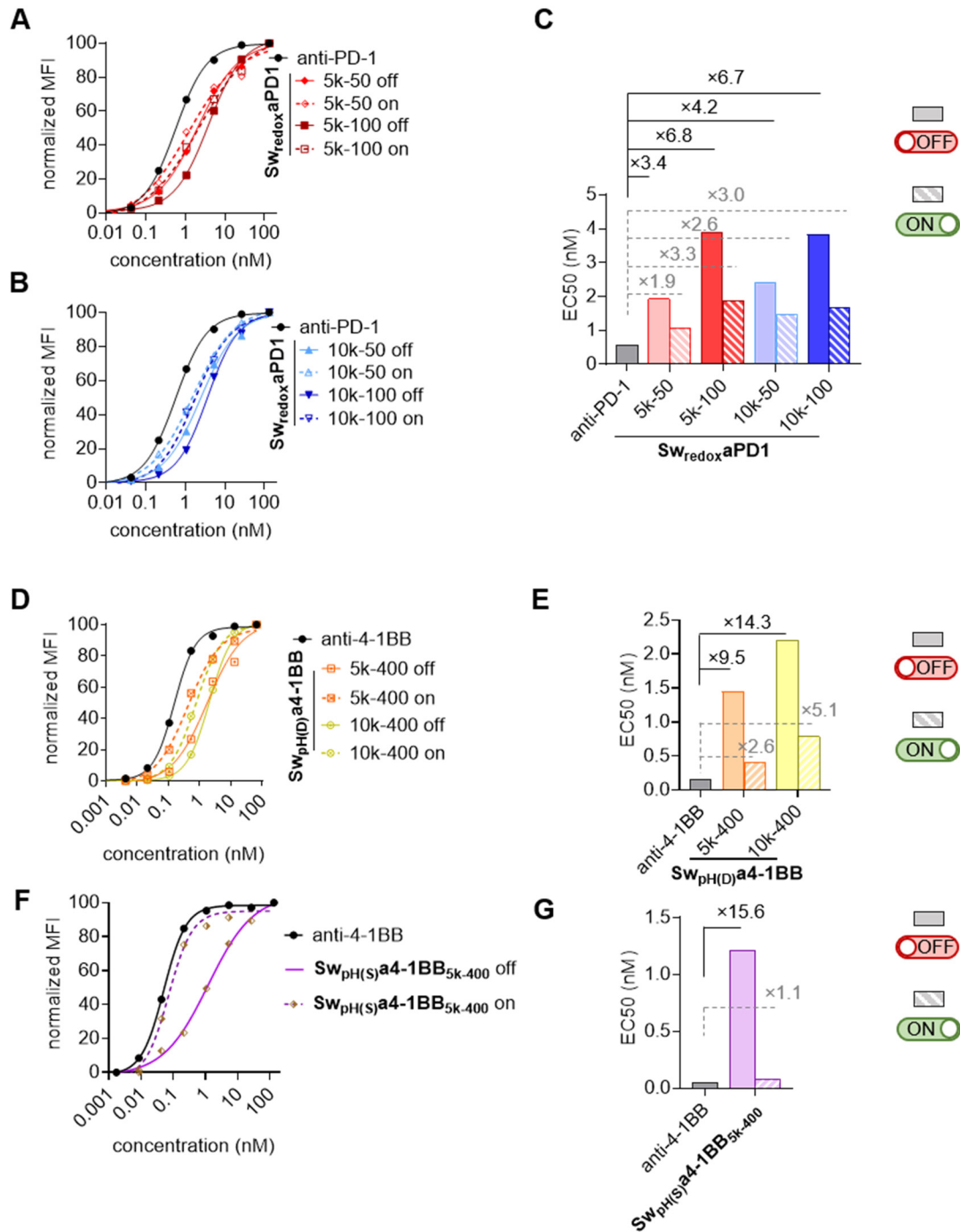


Fig. S9. Controlled switch off and on status of Sw-IMs characterized by a T cell binding assay. Activated CD8⁺ T cells (1×10^5) were incubated with Alexa FluorTM647 (Alx-647)-labelled Sw-IMs at off or on status of series diluted concentrations for 1 hour. Mean fluorescent intensity (MFI) of Alx-647 on CD8⁺ T cells was measured by flow cytometry to represent the binding capacity. (A, B, D, F) Binding capacity of Sw-IMs at off and on status with CD8⁺ T cells. The plot of each formulation is a representative of three independent experiments. (C, E, G) The half maximal effective concentrations (EC50s) of Sw-IMs at off and on status.

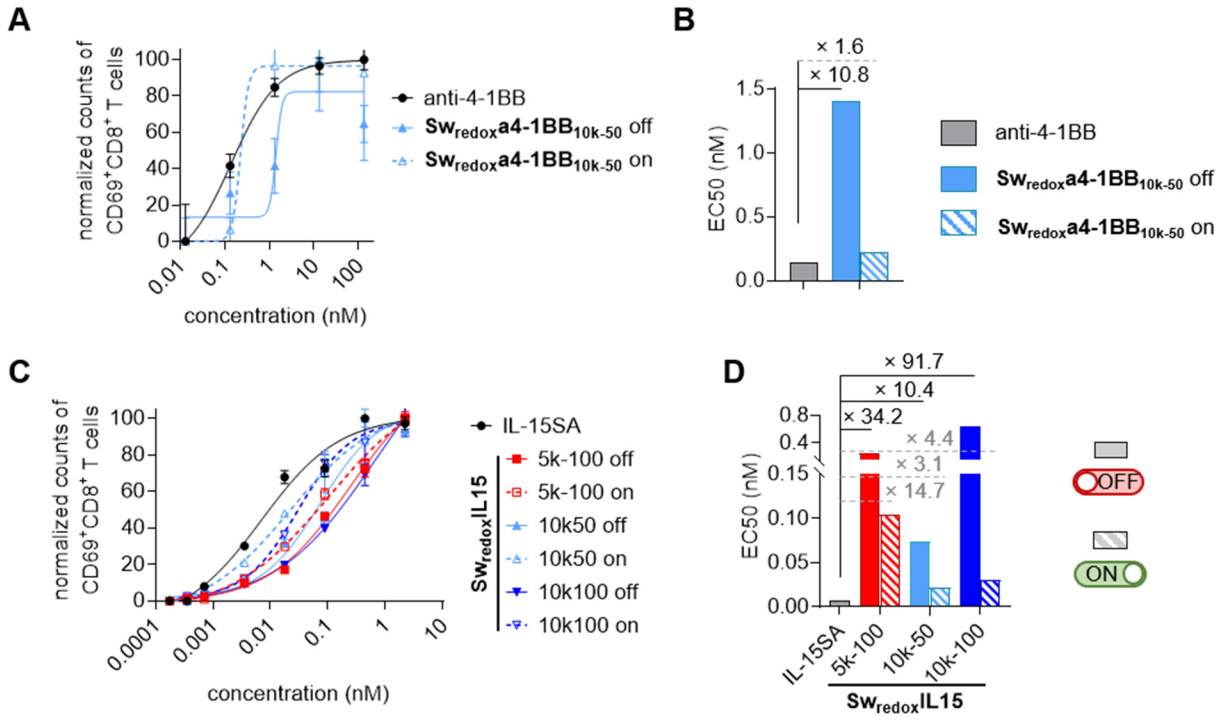


Fig. S10. Controlled switch off and on status of Sw-IMs characterized by a functional activity assay (A) Activities of Sw_{redox}a4-1BB_{10k-50} and native anti-4-1BB at off and on status in stimulating naïve CD8⁺ T cells. (B) EC50s of Sw_{redox}a4-1BB_{10k-50} at off and on status. (C) Activities of Sw_{redox}IL15s and native IL-15SA at off and on status in stimulating CD8⁺ T cells. (D) EC50s of Sw_{redox}IL15s at off and on status.

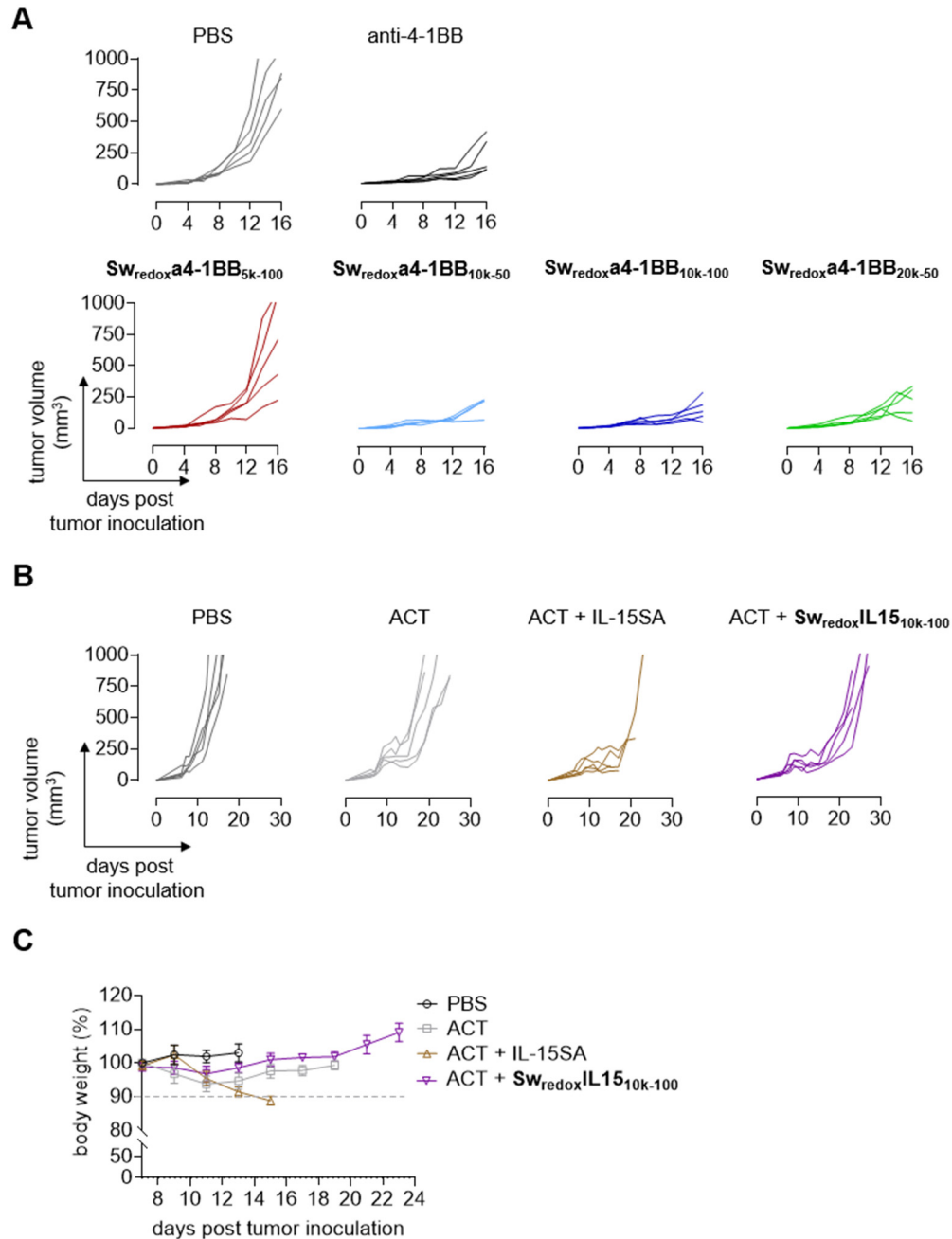


Fig. S11. Selective switch on of Sw-IMs in the tumor microenvironment leading to antitumor immunity. (A) C57BL/6 mice were inoculated subcutaneously (s.c.) with MC38 murine colon adenocarcinoma cells (2×10^5) and received intraperitoneal (i.p.) administration of native anti-4-1BB (100 μ g), Sw_{redox}a4-1BBs (equivalent dose of anti-4-1BB), or PBS on day 4, 7, and 10 ($n = 5$ mice). Shown are individual tumor growth curves. (B, C) C57BL/6 mice were inoculated s.c. with B16F10 melanoma cells (5×10^5) and received adoptive transfer of activated Pmel Thy1.1⁺CD8⁺ T cells (1×10^7) on day 7 followed by intravenous (i.v.) administration of native IL-15SA (5 μ g), Sw_{redox}IL15_{10k-100} (equivalent dose of IL-15SA), or PBS every other day from day 7 to day 21 ($n = 5$ mice). (B) Individual tumor growth curves. (C) Average body weight of the treated mice.

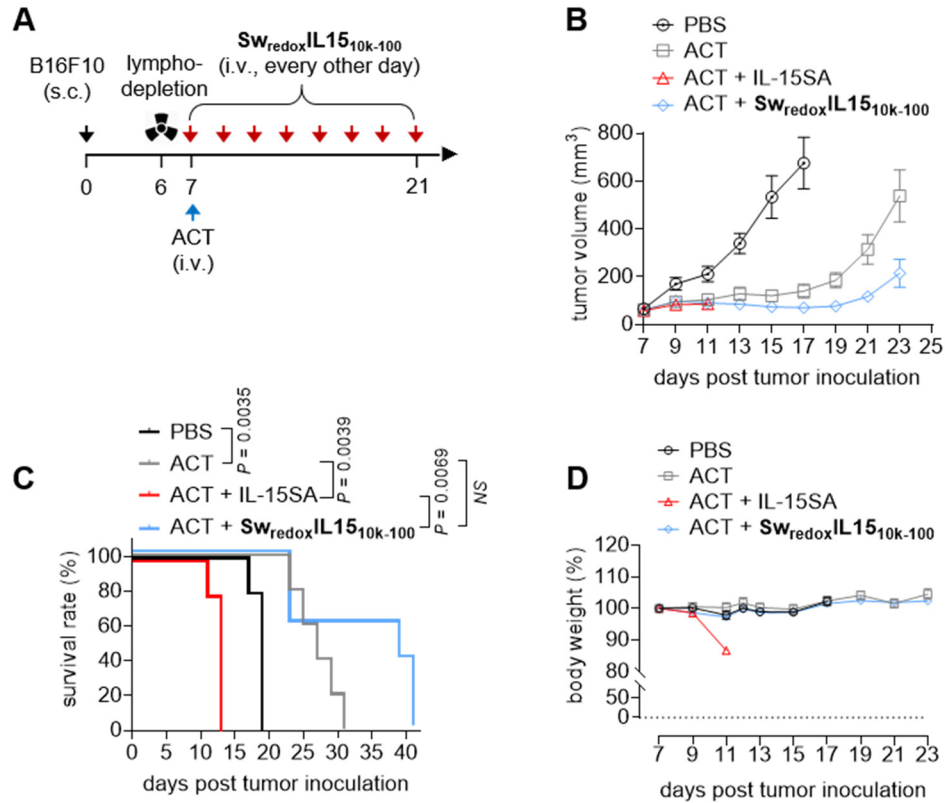
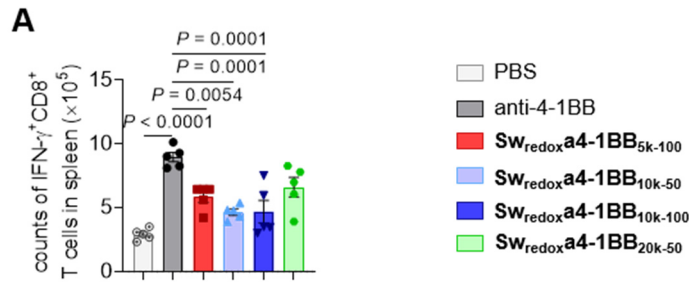


Fig. S12. $Sw_{redox}IL15_{10k-100}$ led to equivalent antitumor immunity but much enhanced safety compared to the native IL-15SA in adoptive T cell transfer (ACT) therapy with prior lymphodepletion. (A) C57BL/6 mice were inoculated s.c. with B16F10 cells (5×10^5) and received lymphodepletion (4 Gy) one day prior to the adoptive transfer of activated Pmel Thy1.1⁺CD8⁺ T cells (1×10^7) on day 7, which was followed by i.v. administration of native IL-15SA (10 μ g), $Sw_{redox}IL15_{10k-100}$ (equivalent dose of IL-15SA), or PBS every other day from day 7 to day 21 ($n = 5$ mice). (B) Average tumor growth curves. (C) Survival curves. (D) Average body weights. All data represent the mean \pm s.e.m. and are analyzed by Log-rank test; NS, not significant ($P > 0.05$).

spleen



liver

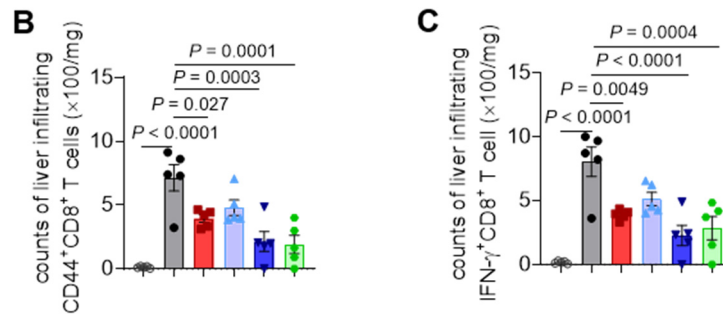


Fig. S13. Sw_{redox}a4-1BBs significantly reduced the toxicity of anti-4-1BB agonistic antibody therapy. C57BL/6 mice bearing MC38 colon adenocarcinoma were treated as shown in Fig. 3A. Mice were euthanatized on day 16 and the tissues were processed for histological and flow cytometry analyses (n = 5 mice). **(A)** Counts of IFN- γ -secreting CD8⁺ T cells in spleen. **(B)** Counts of liver infiltrating CD44⁺CD8⁺ T cells. **(C)** Counts of liver infiltrating IFN- γ -secreting CD8⁺ T cells. All data represent the mean \pm s.e.m. and are analyzed by one-way ANOVA.

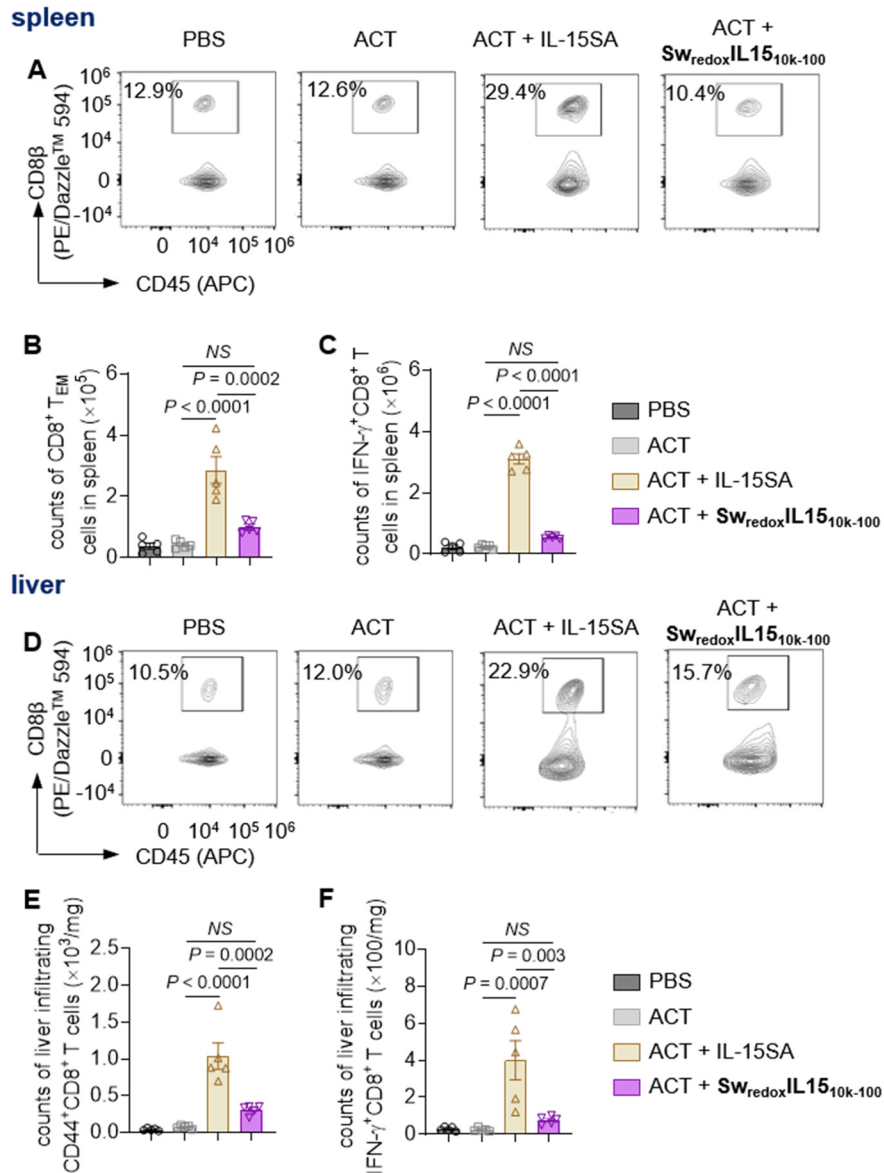


Fig. S14. $Sw_{redox}IL15$ markedly reduced the toxicity of the combination immunotherapy of IL-15SA and ACT. C57BL/6 mice were inoculated s.c. with B16F10 melanoma (5×10^5) and received adoptive transfer of Thy1.1⁺CD8⁺ T cells (1×10^7) on day 7 followed by i.v. administration of native IL-15SA (10 μ g), $Sw_{redox}IL15_{10k-100}$ (equivalent dose of IL-15SA), or PBS every other day from day 7 to day 13. Mice were euthanatized on day 14 and the tissues were processed for flow cytometry analyses (n = 5 mice). (A) Representative flow cytometry plots of CD8⁺ T cells among all lymphocytes (CD45⁺) in spleen. (B) Counts of effector memory CD8⁺ T cells (T_{EM}, CD44^{high}CD62L^{low}) in spleen. (C) Counts of IFN- γ -secreting CD8⁺ T cells in spleen. (D) Representative flow cytometry plots of CD8⁺ T cells among all liver infiltrating lymphocytes (CD45⁺). (E) Counts of liver infiltrating CD44⁺CD8⁺ T cells. (F) Counts of liver-infiltrating IFN- γ -secreting T cells. All data represent the mean \pm s.e.m. and are analyzed by one-way ANOVA; NS, not significant (P > 0.05).

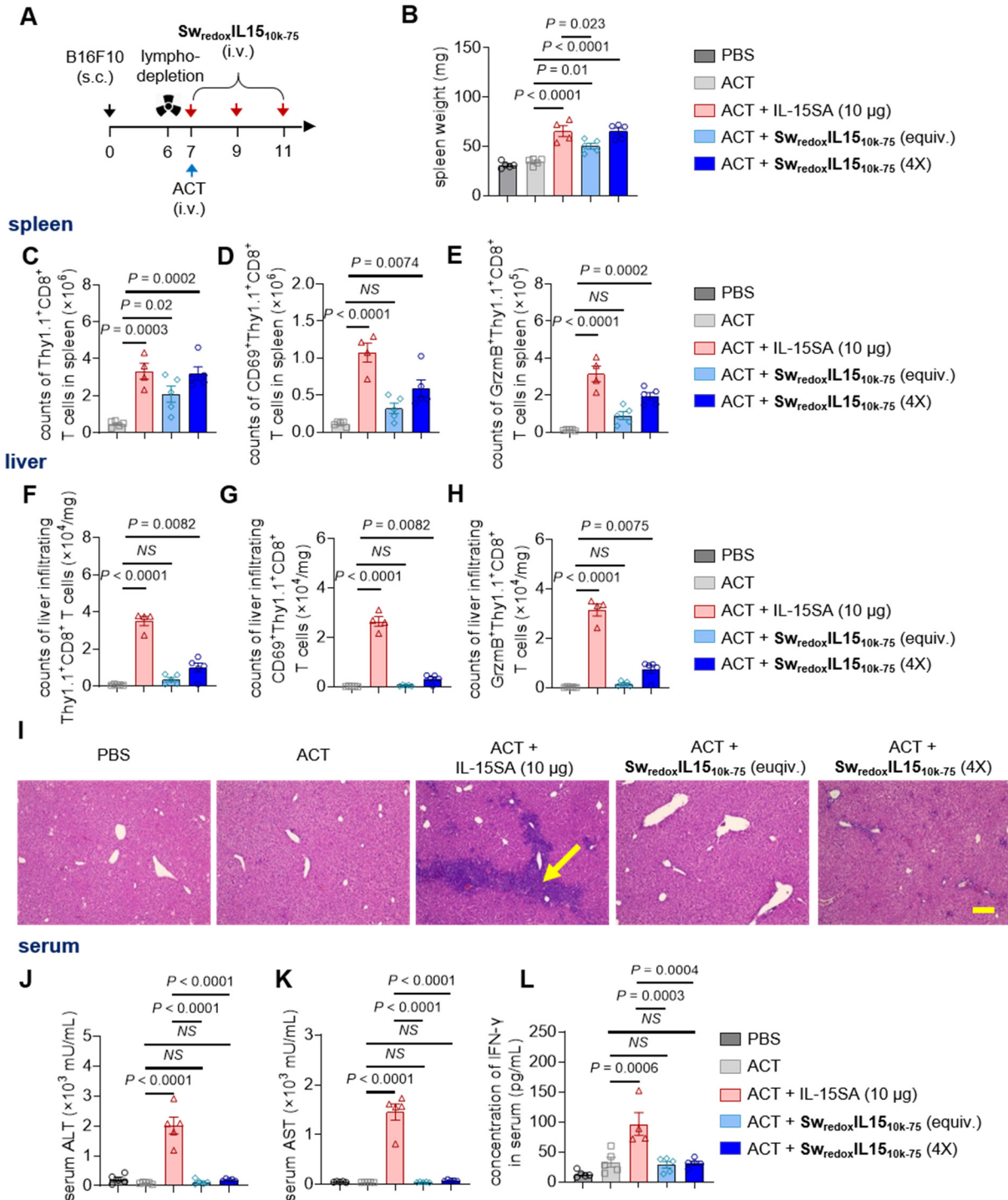


Fig. S15. $Sw_{redox}IL15$ markedly reduced the toxicity of the combination immunotherapy of IL-15SA and ACT with prior lymphodepletion. (A) C57BL/6 mice were inoculated s.c. with B16F10 melanoma (5×10^5) and received lymphodepletion (4 Gy) one day prior to the adoptive transfer of activated Pmel Thy1.1⁺CD8⁺ T cells (1×10^7) on day 7, which was followed by i.v. administration of native IL-15SA (10 μ g), $Sw_{redox}IL15_{10k-75}$ (equivalent dose of IL-15SA), $Sw_{redox}IL15_{10k-75}$ (4 \times dose of IL-15SA), or PBS on day 7, 9, and 11. Mice were euthanized on

day 12 and the tissues were processed for flow cytometry and histological analyses (n = 5 mice). **(B)** Average spleen weight. **(C)** Counts of Thy1.1⁺CD8⁺ T cells in spleen. **(D)** Counts of CD69⁺Thy1.1⁺CD8⁺ T cells in spleen. **(E)** Counts of GrzmB-secreting Thy1.1⁺CD8⁺ T cells in spleen. **(F)** Counts of liver infiltrating Thy1.1⁺CD8⁺ T cells. **(G)** Counts of liver infiltrating CD69⁺Thy1.1⁺CD8⁺ T cells. **(H)** Counts of liver infiltrating GrzmB-secreting Thy1.1⁺CD8⁺ T cells. **(I)** Histopathological analyses of liver tissues. Yellow arrows indicate tissue damage. Scale bar, 200 μm. **(J)** Serum activity of alanine aminotransferase (ALT). **(K)** Serum activity of aspartate transaminase (AST). **(L)** Serum concentration of IFN-γ. All data represent the mean ± s.e.m. and are analyzed by one-way ANOVA; *NS*, not significant (P > 0.05).

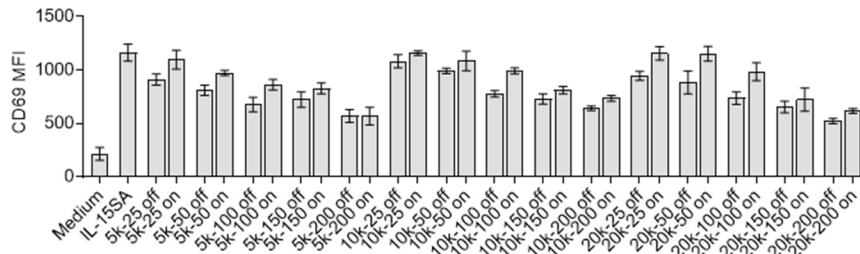
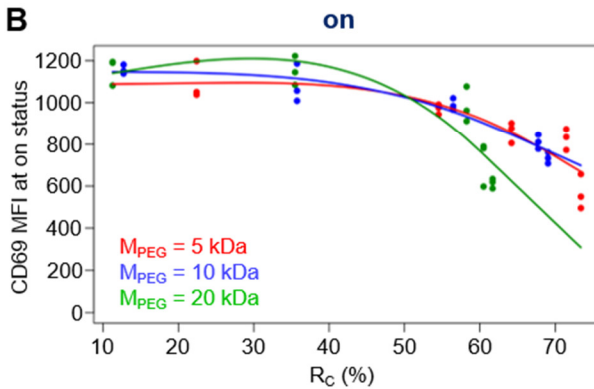
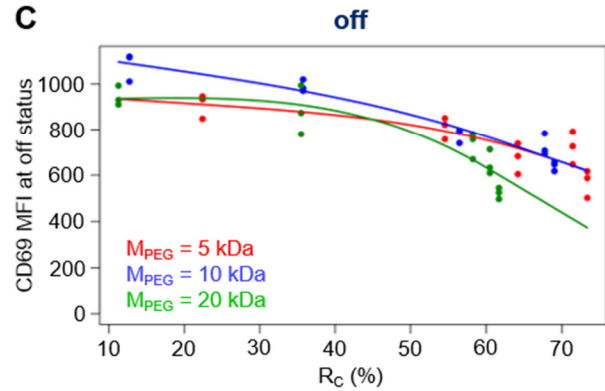
A $Sw_{redox}IL15$ **B****C**

Fig. S16. In vitro T cell activation by $Sw_{redox}IL15$ s of different formulations. (A) MFI of CD69 expressed by $CD8^+$ T cells cultured with $Sw_{redox}IL15$ s at off and on status or native IL-15SA. **(B, C)** Plots of CD69 MFI of $Sw_{redox}IL15$ s with different M_{PEG} at off and on status as functions of R_C . Smooth curves were obtained using penalized cubic regression splines.

Table S1. Preparation of Sw-IMs of various formulations.

Entry	Sw-IM*	IM	linker [†]	M _{PEG} [‡] (Da)	R _F [§]	R _C (%)
1	Sw _{redox} aTNP _{2k-50}	anti-TNP	SS	2k	50	62.1
2	Sw _{redox} aTNP _{2k-100}	anti-TNP	SS	2k	100	73.6
3	Sw _{redox} aTNP _{2k-200}	anti-TNP	SS	2k	200	81.9
4	Sw _{redox} aTNP _{5k-50}	anti-TNP	SS	5k	50	68.9
5	Sw _{redox} aTNP _{5k-100}	anti-TNP	SS	5k	100	74.3
6	Sw _{redox} aTNP _{10k-50}	anti-TNP	SS	10k	50	52.3
7	Sw _{redox} aTNP _{10k-200}	anti-TNP	SS	10k	200	67.8
8	Sw _{redox} a4-1BB _{2k-50}	anti-4-1BB	SS	2k	50	47.7
9	Sw _{redox} a4-1BB _{2k-100}	anti-4-1BB	SS	2k	100	52.8
10	Sw _{redox} a4-1BB _{2k-200}	anti-4-1BB	SS	2k	200	56.6
11	Sw _{redox} a4-1BB _{5k-200}	anti-4-1BB	SS	5k	200	83.1
12	Sw _{redox} a4-1BB _{10k-200}	anti-4-1BB	SS	10k	200	80.4
13	Sw _{redox} a4-1BB _{20k-25}	anti-4-1BB	SS	20k	25	53.5
14	Sw _{redox} aPD1 _{2k-50}	anti-PD-1	SS	2k	50	65.4
15	Sw _{redox} aPD1 _{2k-100}	anti-PD-1	SS	2k	100	77.7
16	Sw _{redox} aPD1 _{2k-200}	anti-PD-1	SS	2k	200	83.2
17	Sw _{redox} aPD1 _{5k-50}	anti-PD-1	SS	5k	50	74.7
18	Sw _{redox} aPD1 _{5k-100}	anti-PD-1	SS	5k	100	77.2
19	Sw _{redox} aPD1 _{5k-200}	anti-PD-1	SS	5k	200	83.6
20	Sw _{redox} aPD1 _{10k-50}	anti-PD-1	SS	10k	50	60.8
21	Sw _{redox} aPD1 _{10k-200}	anti-PD-1	SS	10k	200	64.3
22	Sw _{redox} aCTLA4 _{2k-50}	anti-CTLA-4	SS	2k	50	57.7
23	Sw _{redox} aCTLA4 _{2k-100}	anti-CTLA-4	SS	2k	100	72.9
24	Sw _{redox} aCTLA4 _{2k-200}	anti-CTLA-4	SS	2k	200	79.1
25	Sw _{redox} aCTLA4 _{5k-50}	anti-CTLA-4	SS	5k	50	66.0
26	Sw _{redox} aCTLA4 _{5k-100}	anti-CTLA-4	SS	5k	100	70.2
27	Sw _{redox} aCTLA4 _{5k-200}	anti-CTLA-4	SS	5k	200	82.2
28	Sw _{redox} aCTLA4 _{10k-50}	anti-CTLA-4	SS	10k	50	53.5
29	Sw _{redox} aCTLA4 _{10k-200}	anti-CTLA-4	SS	10k	200	58.7
30	Sw _{redox} IL15 _{5k-25}	IL-15SA	SS	5k	25	22.4
31	Sw _{redox} IL15 _{5k-50}	IL-15SA	SS	5k	50	54.5
32	Sw _{redox} IL15 _{5k-150}	IL-15SA	SS	5k	150	71.5
33	Sw _{redox} IL15 _{5k-200}	IL-15SA	SS	5k	200	73.4
34	Sw _{redox} IL15 _{10k-25}	IL-15SA	SS	10k	25	12.7
35	Sw _{redox} IL15 _{10k-75}	IL-15SA	SS	10k	75	49.5
36	Sw _{redox} IL15 _{10k-150}	IL-15SA	SS	10k	150	67.7
37	Sw _{redox} IL15 _{10k-200}	IL-15SA	SS	10k	200	69.0
38	Sw _{redox} IL15 _{20k-25}	IL-15SA	SS	20k	25	11.3
39	Sw _{redox} IL15 _{20k-100}	IL-15SA	SS	20k	100	58.2
40	Sw _{redox} IL15 _{20k-150}	IL-15SA	SS	20k	150	60.5

41	Sw_{redox}IL15_{20k-200}	IL-15SA	SS	20k	200	61.7
42	Sw_{pH(D)}a4-1BB_{2k-400}	anti-4-1BB	DBCO	2k	400	
43	Sw_{pH(D)}a4-1BB_{10k-400}	anti-4-1BB	DBCO	10k	400	

***Sw-IM**, switchable immune modulator. **Sw-IM** is denoted as **Sw_{linker}protein_{MPEG-RF}**. **redox**, redox-responsive linker; **pH(D)**, acidic pH-responsive linker with a DBCO-triazole spacer; **aTNP**, anti-TNP antibody; **a4-1BB**, anti-4-1BB antibody; **aPD1**, anti-PD-1 antibody; **aCTLA4**, anti-CTLA-4 antibody; **IL15**, IL-15SA.

†The chemical structure of the linkers is shown in Fig. 1. SS, disulphate linker; DBCO, dibenzocyclooctyne.

‡M_{PEG}, molecular weight of polyethylene glycol (PEG).

§R_F, feeding mole ratio of PEG to **IM**.

¶R_C, conjugation ratio, defined as (number of conjugated NH₂ groups / total number of NH₂ groups of **IM**) × 100%.