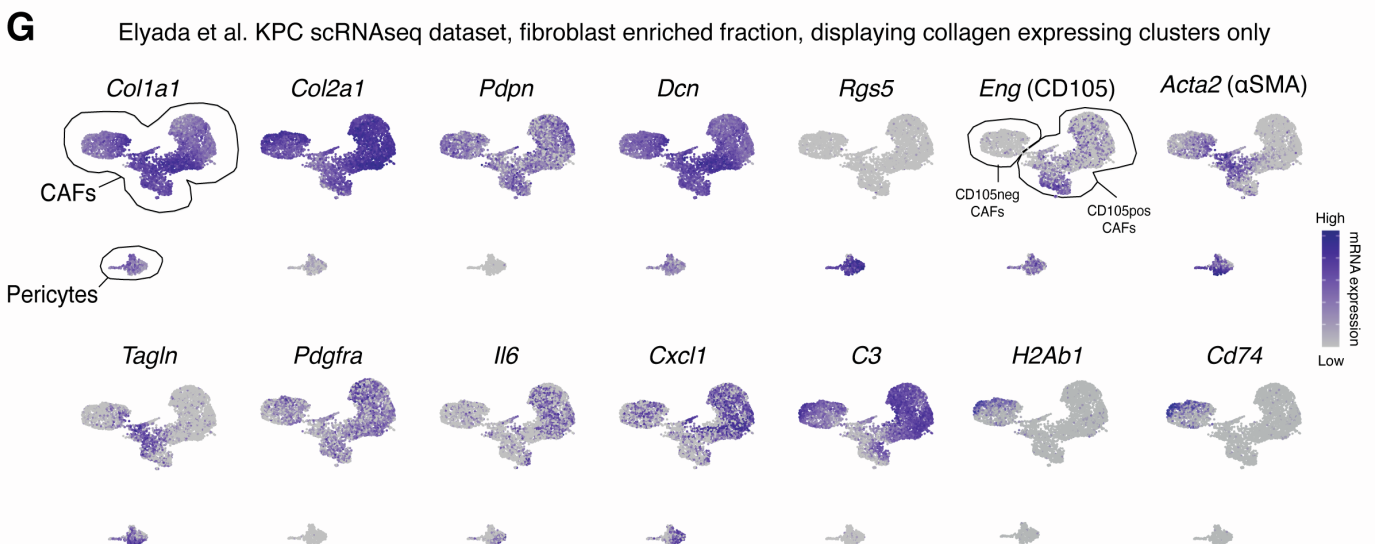
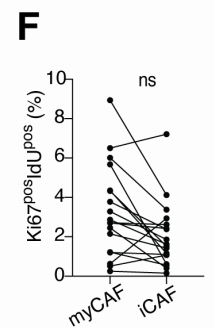
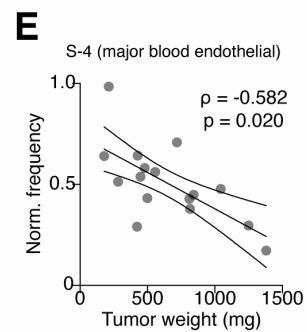
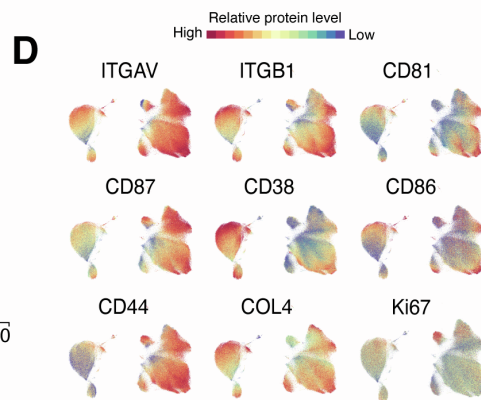
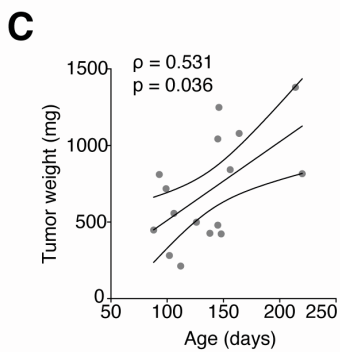
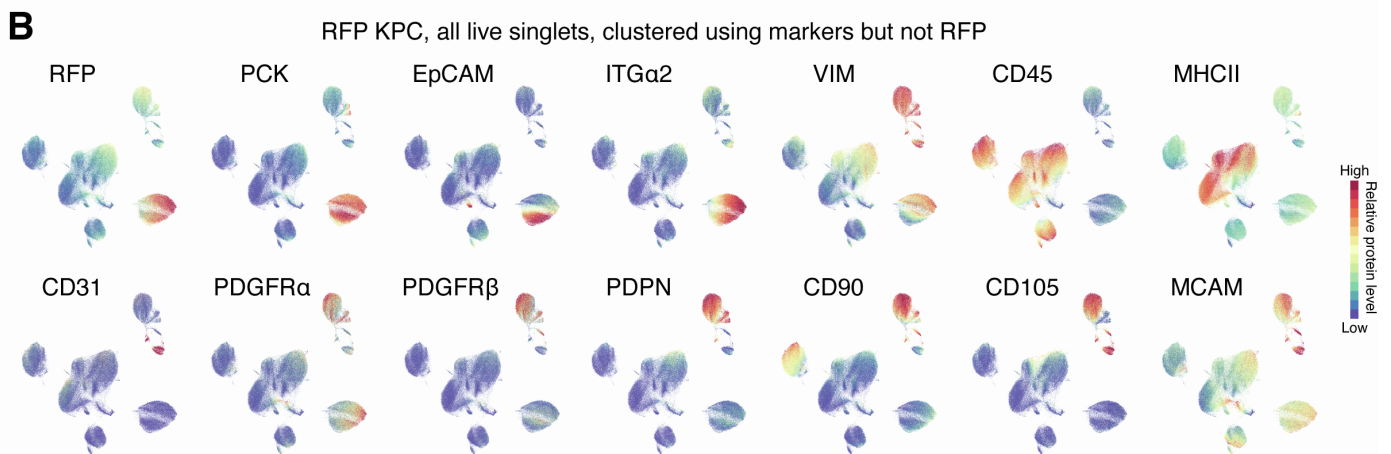
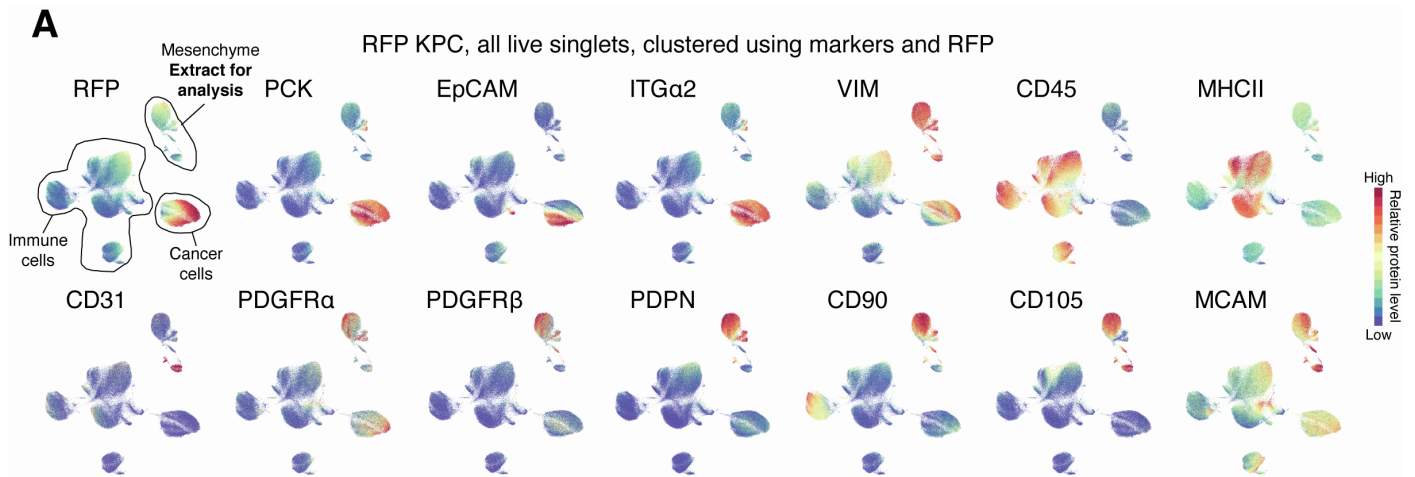


Supplemental information

**Single-cell analysis defines a pancreatic
fibroblast lineage that supports
anti-tumor immunity**

Colin Hutton, Felix Heider, Adrian Blanco-Gomez, Antonia Banyard, Alexander Kononov, Xiaohong Zhang, Saadia Karim, Viola Paulus-Hock, Dale Watt, Nina Steele, Samantha Kemp, Elizabeth K.J. Hogg, Joanna Kelly, Rene-Filip Jackstadt, Filipa Lopes, Matteo Menotti, Luke Chisholm, Angela Lamarca, Juan Valle, Owen J. Sansom, Caroline Springer, Angeliki Malliri, Richard Marais, Marina Pasca di Magliano, Santiago Zelenay, Jennifer P. Morton, and Claus Jørgensen



Supplemental Figure 1, related to Figure 1. Phenotypic and compositional heterogeneity of pancreatic cancer-associated mesenchymal cells

(A) UMAP projection of all live, single cells from an RFP KPC tumor sample using all phenotypic markers to drive clustering. PCK: pan-cytokeratin.

(B) UMAP projection of all live, single cells from an RFP KPC tumor sample, as for (A), but RFP is not used to drive clustering. PCK: pan-cytokeratin.

(C) Correlation between KPC mouse age and weight of tumors collected at humane endpoint. ρ =Spearman correlation coefficient, 90% confidence intervals displayed. Mean age of mice included in study was 142 days.

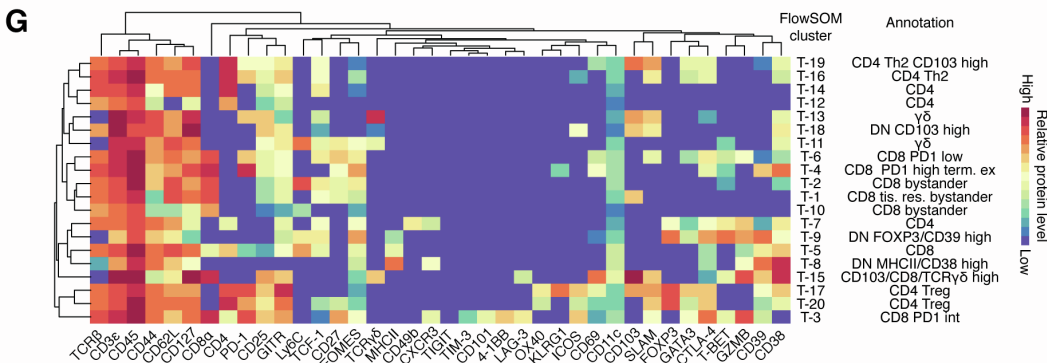
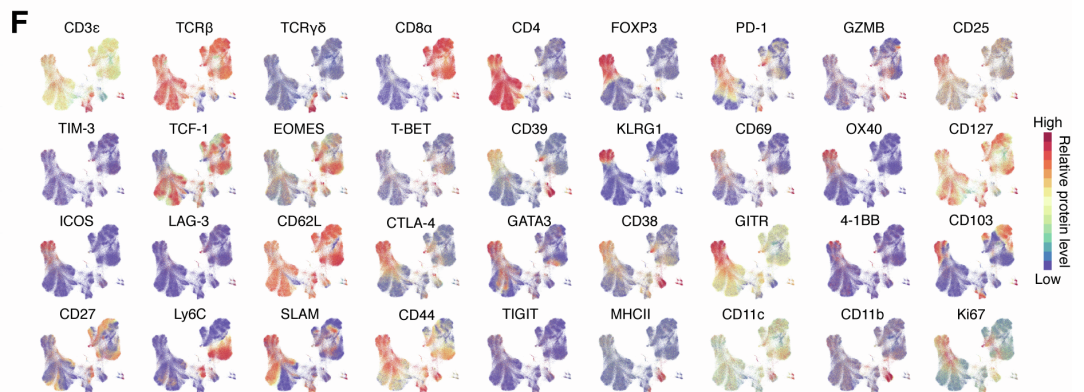
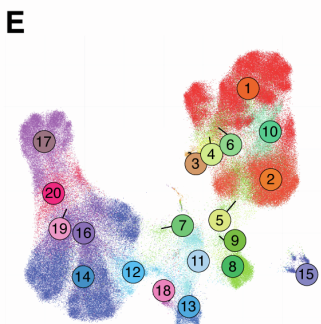
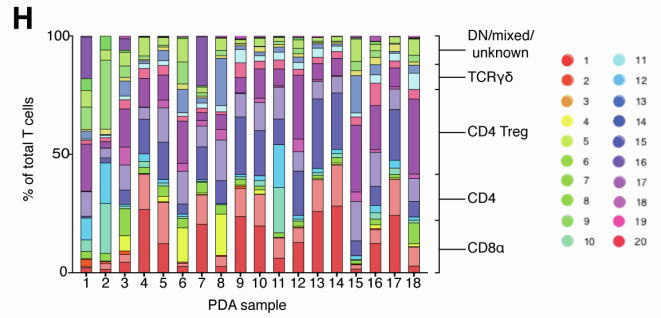
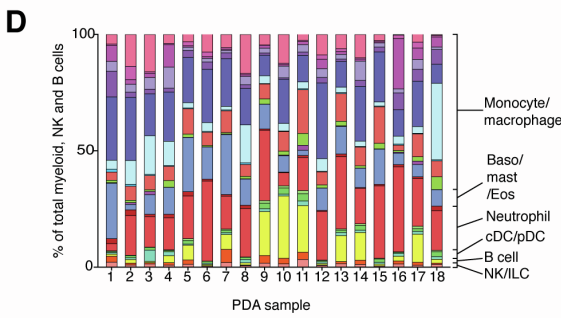
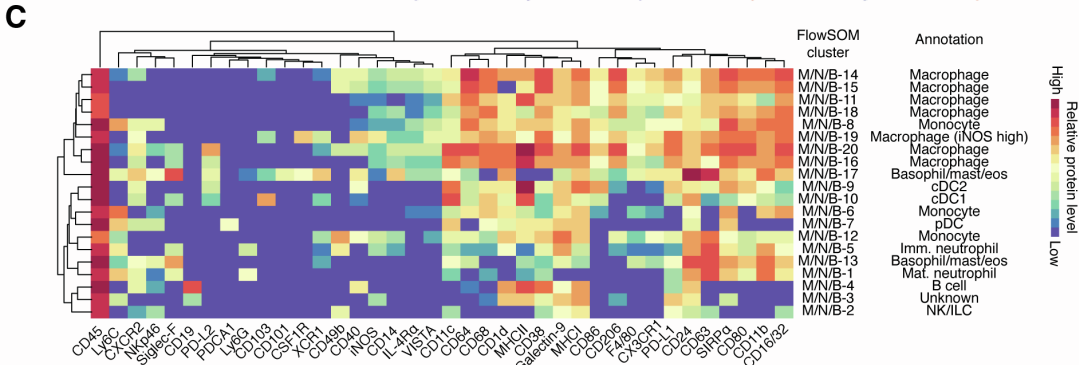
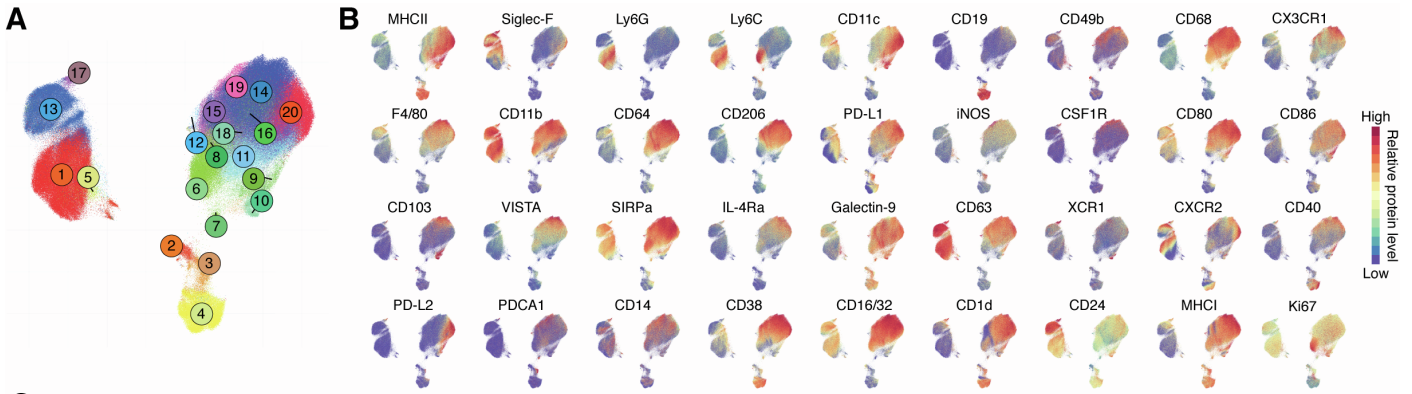
(D) UMAP projection from Figure 1A, displaying relative signal intensity of additional phenotypic markers not included in Figure 1D.

(E) Spearman correlation analysis between the relative frequency of the endothelial cell subset, S-4 and PDA tumor weight. ρ =Spearman correlation coefficient, 90% confidence intervals displayed.

(F) Frequency of myCAFs (α SMA^{high} PDGFR α ^{low}) and iCAFs (α SMA^{low} PDGFR α ^{high}) in S-phase (Ki67^{pos} IdU^{pos}).

(G) Reanalysis of single cell transcriptomic data from Elyada et al. The pre-defined fibroblast enriched fraction was used and only collagen expressing cells analyzed. Data is displayed as UMAP projections with overlaid relative expression levels of example genes, including canonical fibroblast genes (*Col1a1/2*, *Pdpr*, *Dcn*), the canonical pericyte gene (*Rgs5*), *Eng* (the gene that encodes CD105) and myCAF-, iCAF- and apCAF-associated genes (*Acta2*, *Tagln* and *Pdgfra*, *Ilf6*, *Cxcl1*, *C3*, and *H2Ab1*, *Cd74* respectively).

Samples are compared using Spearman correlation adjusted for multiple testing using Benjamini-Hochberg correction (C, E) or paired t-test (F). ns. non-significant



Supplemental Figure 2, related to Figure 2. Phenotypic and compositional heterogeneity of pancreatic cancer-associated immune subsets

(A) UMAP projection of single CD45^{pos} CD3^{neg} cells (MNB: Myeloid, NK, B cell) from 18 of the 19 tumors analyzed for the mesenchymal stroma with color-coded FlowSOM clusters (1-20). Each sample contributes an equal number of cells to the dataset. Total of 5×10^5 cells displayed.

(B) UMAP projection (from (A)) displaying overlaid relative signal intensity of example phenotypic markers.

(C) Heatmap of marker median mass intensities (MMIs) displayed as z-scores. Each FlowSOM MNB cluster is grouped by unsupervised hierarchical clustering based on marker MMIs. Cell-type annotations based on canonical phenotypic markers are listed.

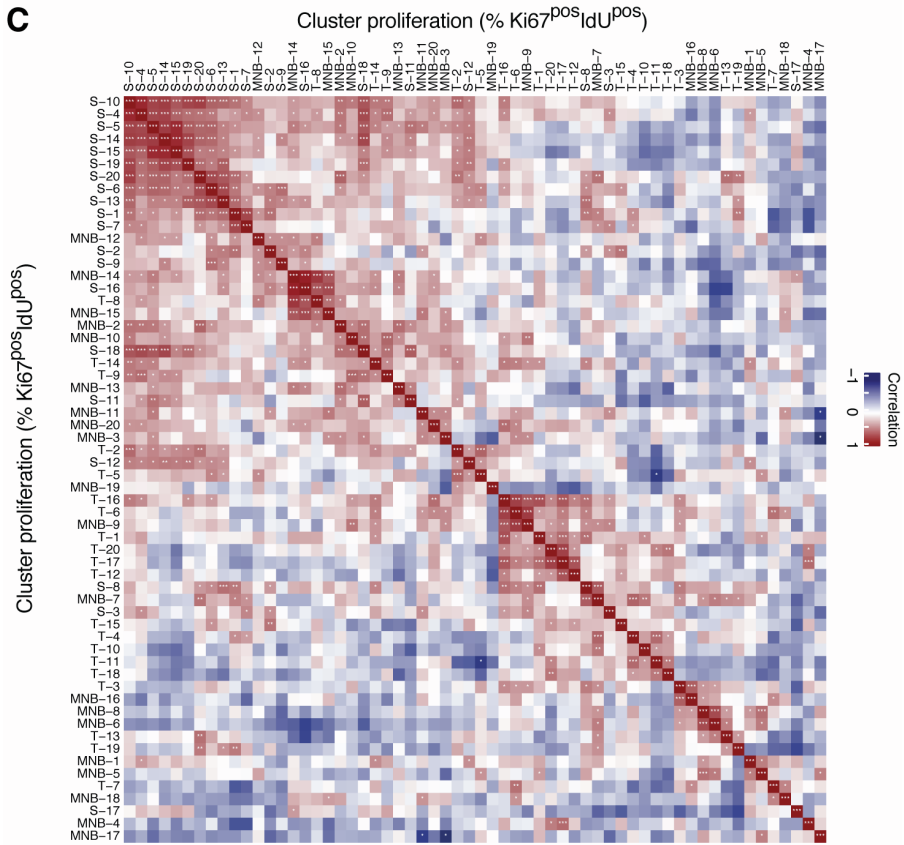
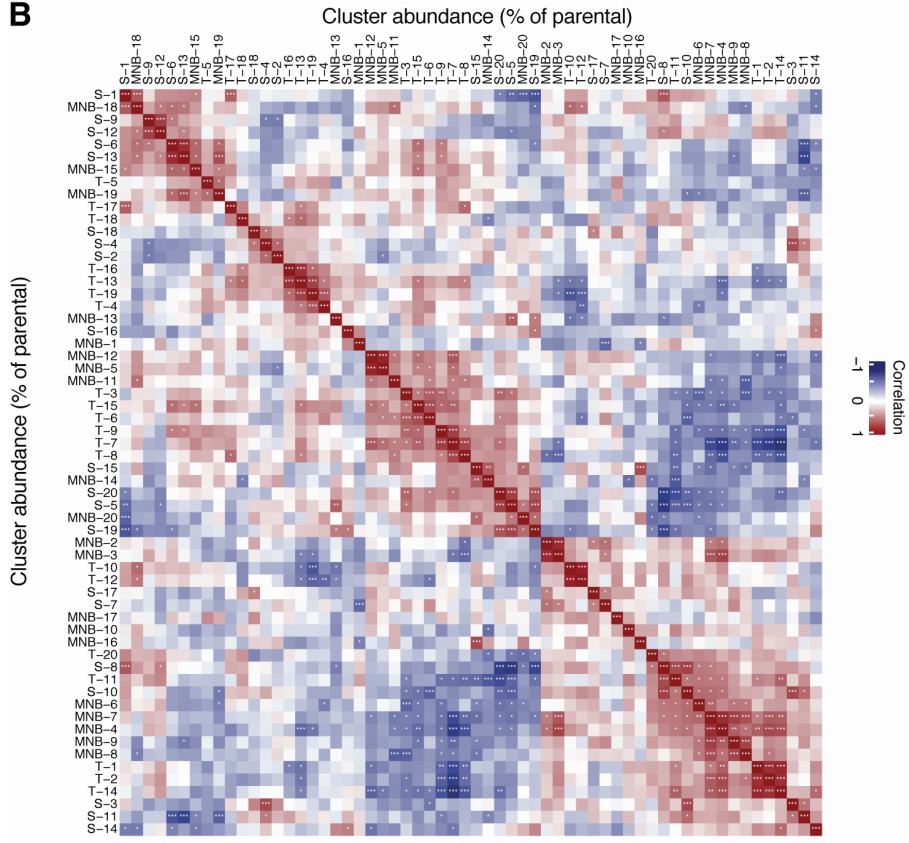
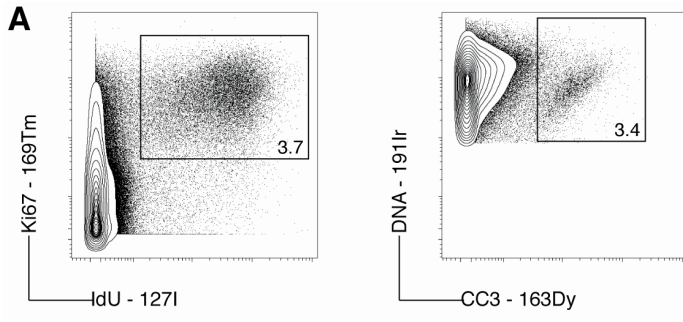
(D) Stacked bar graph displaying relative abundance of KPC PDA MNB FlowSOM clusters grouped into major MNB cell sub-types.

(E) UMAP projection of single CD45^{pos} CD3^{pos} cells (T cells) from 18 of the 19 tumors analyzed for the mesenchymal stroma with color-coded FlowSOM clusters (1-20). Each sample contributes an equal number of cells to the dataset. Total of 5×10^5 cells displayed.

(F) UMAP projection from (E) displaying overlaid relative signal intensity of example phenotypic markers.

(G) Heatmap of marker median mass intensities (MMIs) displayed as z-scores. Each FlowSOM T cell cluster is grouped by unsupervised hierarchical clustering based on marker MMIs. Cell-type annotations based on canonical phenotypic markers are listed.

(H) Stacked bar graph displaying relative abundance of KPC PDA T cell FlowSOM clusters grouped into major T cell sub-types.

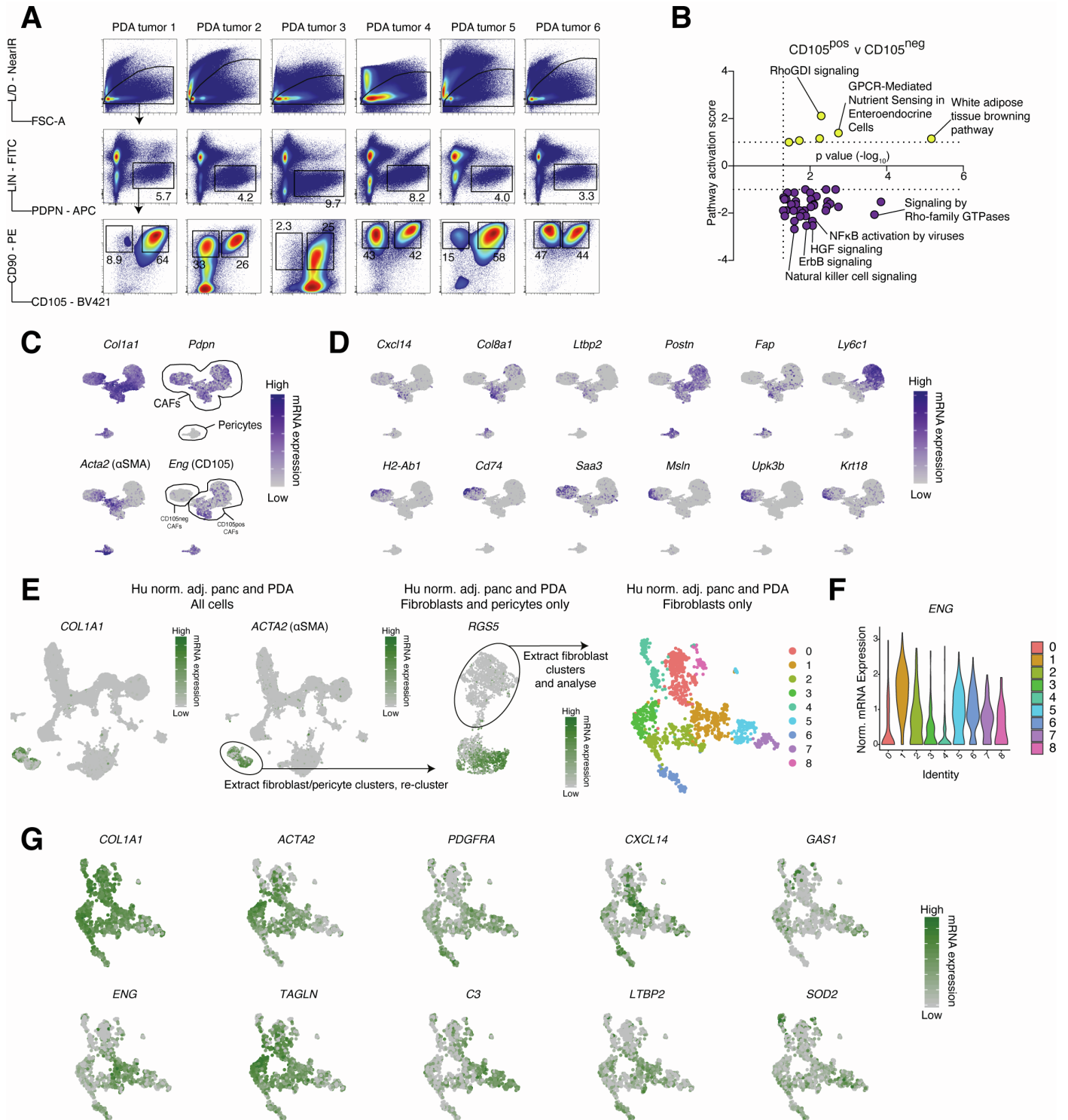


Supplemental Figure 3, related to Figure 2. Co-regulated CAF and immune subsets within the PDA tumor microenvironment

(A) Example mass cytometry (MC) data with Ki67 and IdU (left) and DNA and cleaved caspase-3 (CC3) (right). Cells in S-phase, at the time the tumor was collected, can be clearly identified as Ki67^{pos} IdU^{pos} and apoptotic cells as CC3^{pos}.

(B-C) Correlation matrix with Spearman correlation coefficients of all pairwise comparisons of mesenchymal stromal and immune cell subset frequencies (B) and proliferation rates (C). All pairwise apoptosis rate correlations are included in Table S2.

Samples were compared using Spearman correlation adjusted for multiple testing using Benjamini-Hochberg correction (J,K). * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.



Supplemental Figure 4, related to Figure 3. CD105 expression discriminates two distinct CAF populations in murine and human PDA

(A) FACS gating used to isolate EpCAM^{neg} CD31^{neg} CD45^{neg} CD90^{pos} PDPN^{pos} - CD105^{pos} (n=6) and - CD105^{neg} (n=6) PDA CAFs. L/D: Live/Dead, LIN: EpCAM CD31 CD45. Scatter gating for singlets is not shown.

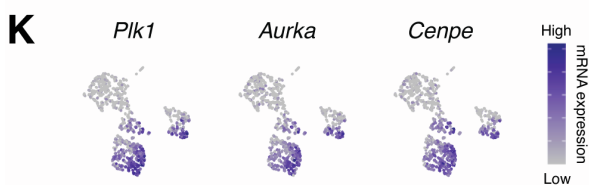
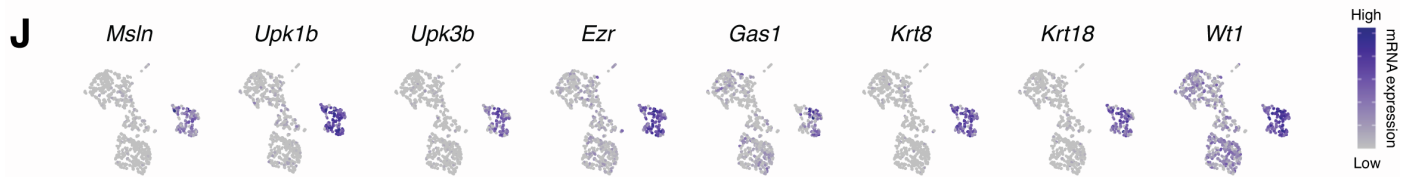
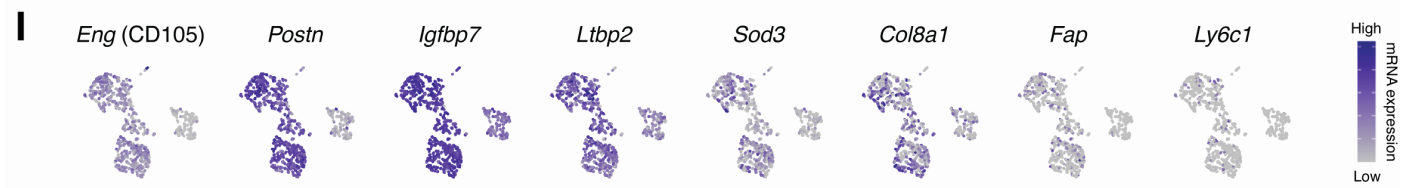
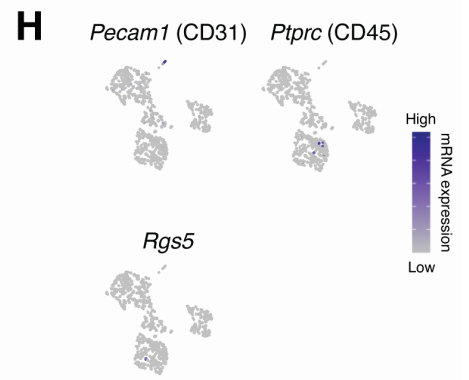
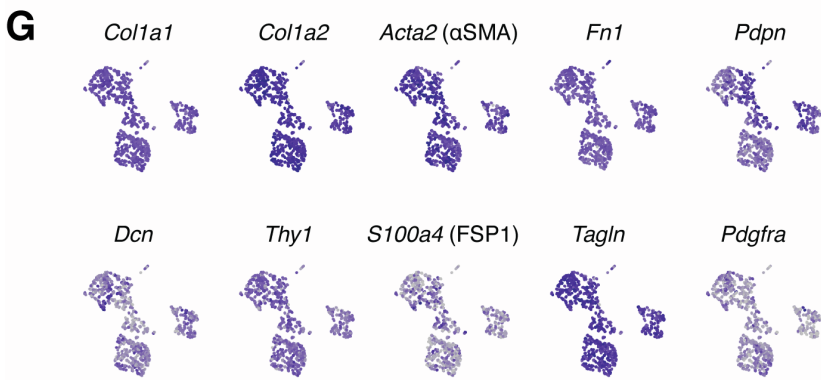
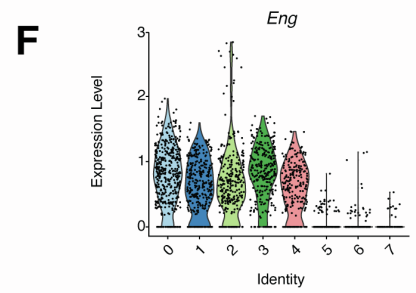
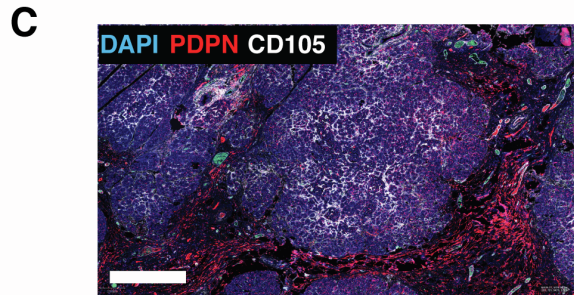
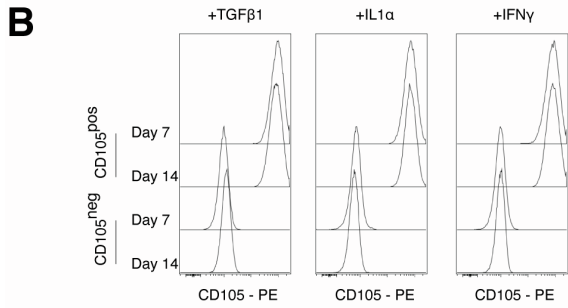
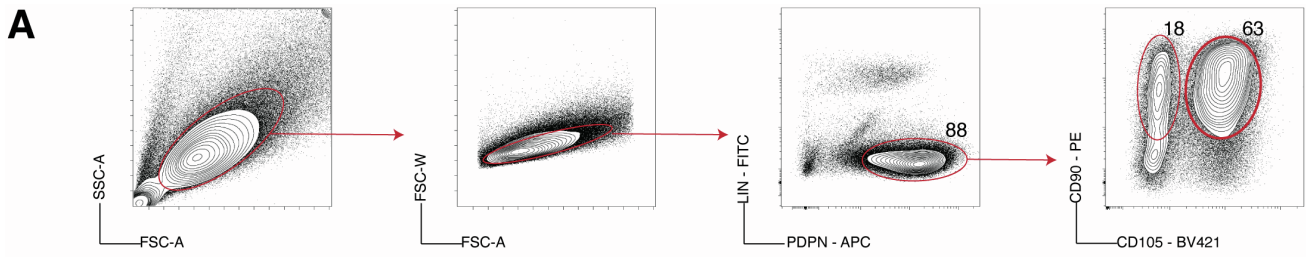
(B) Ingenuity Pathway Analysis (IPA) of isolated PDA CAF gene expression, displaying pathway activation score. Displaying plots for positively enriched (enriched in CD105^{pos}, yellow) (B) and negatively enriched (enriched in CD105^{neg}, purple) (C).

(C-D) Reanalysis of single cell transcriptomic data from Elyada et al. The pre-defined fibroblast enriched fraction was used and only collagen expressing cells analyzed. Data is displayed as UMAP projections with overlaid expression levels of example genes, including canonical fibroblast genes (*Col1a1*, *Pdpn*, *Acta2*) and Eng (the gene that encodes CD105) (C), genes with increased expression in CD105^{pos} PDA CAFs (*Cxcl14*, *Col8a1*, *Ltbp2*, *Postn*, *Fap* and *Ly6c1*) and gene with increased expression in CD105^{neg} PDA CAFs (*H2Ab1*, *Cd74*, *Saa3*, *Msln*, *Up3kb* and *Krt18*) (D).

(E) Single cell transcriptomic analysis of human (Hu) PDA tumor (n=14) and normal adjacent (norm adj) (n=2) (Steele et al., 2020). Displayed as UMAP projections with the expression levels of listed genes overlaid. The method for extracting fibroblast cells is shown using example genes to navigate appropriate clusters. Fibroblast clusters are visualized by color-coding (right).

(F) Violin plots displaying *ENG* expression levels (standard Seurat normalized) of the fibroblast clusters from (E). Plots display maximum and minimum data range with width representing probability density.

(G) UMAP projection of fibroblasts from (E), with overlaid expression of *COL1A1* and *ENG*, myCAF-associated genes (*ACTA2* and *TAGLN*), iCAF-associated genes (*PDGFRA* and *C3*), example CD105^{pos} fibroblast signature genes (*CXCL14* and *LTBP2*) and example CD105^{neg} fibroblast signature genes (*GAS1* and *SOD2*).



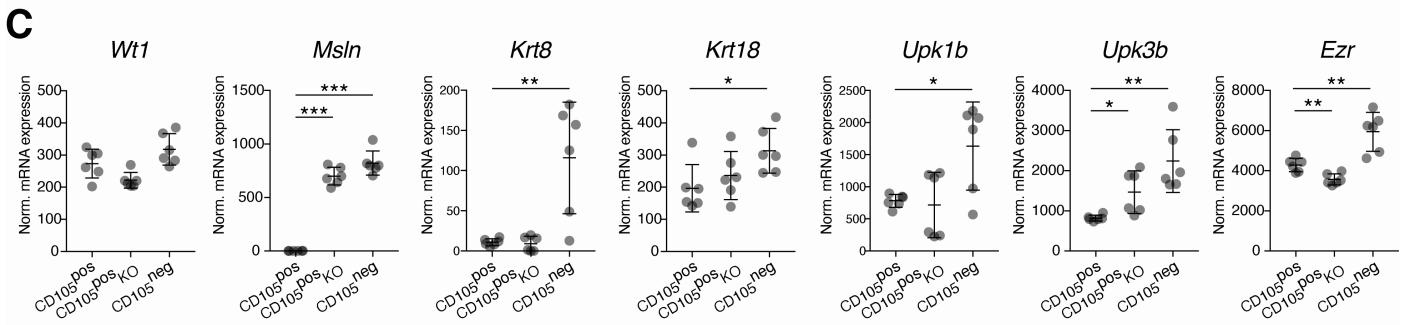
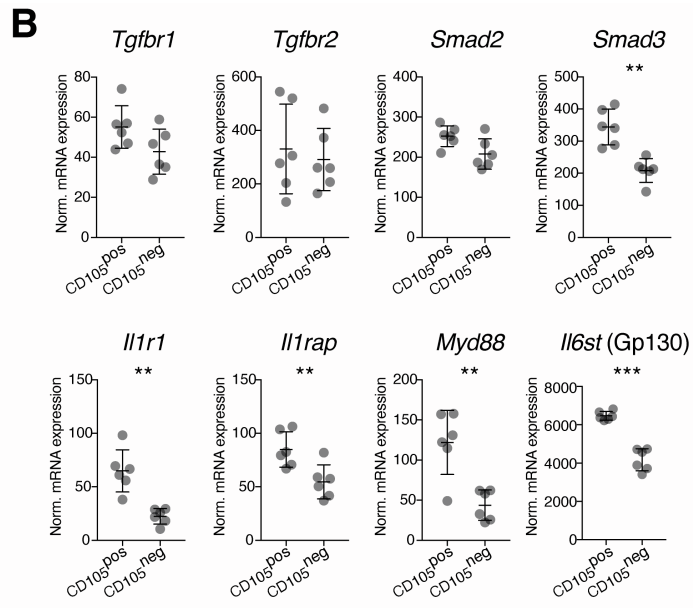
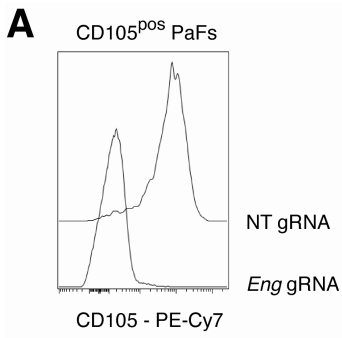
Supplemental Figure 5, related to Figure 4. Phenotypic plasticity of mesenchymal marker expression

(A) Representative FACS gating strategy to isolate CD105^{pos} and CD105^{neg} pancreatic fibroblasts (PaFs) after 7 days of *in vitro* expansion from normal healthy pancreas. L/D: Live/Dead, LIN: EpCAM CD31 CD45. Plots are representative of n=8 experiments.

(B) Flow cytometry analysis of CD105 in populations of purified CD105^{pos} and CD105^{neg} PaFs after 7 and 14 days in culture with TGFβ1, IL1α or IFNγ. Plots are representative of n=4 experiments.

(C) Representative immunohistochemistry (IHC) analysis of normal but inflamed human pancreas tissue, adjacent to PDA tumors. Stained with DAPI (blue) and with antibodies targeting podoplanin (PDPN) (red) and CD105 (white). Scale bar = 750 μm.

(D-J) Single cell transcriptomic analysis of *in vitro* expanded fibroblasts from healthy mouse pancreas (n=3), after 7 days of culture (same time point for the FACS isolation described in (A)). Displayed as UMAP projections (D, E, G-K). Comparison of the n=3 samples demonstrates concordant results (D). Clusters are defined in (E) and *Eng* gene expression in each cluster displayed (F). Expression levels of example genes are overlaid, including canonical fibroblast genes (G), canonical genes of endothelial, immune and perivascular cells (H), *Eng* and other CD105^{pos} fibroblast signature genes (I), CD105^{neg} signature and mesothelial-associated genes (J) and cell cycle-associated genes (K).



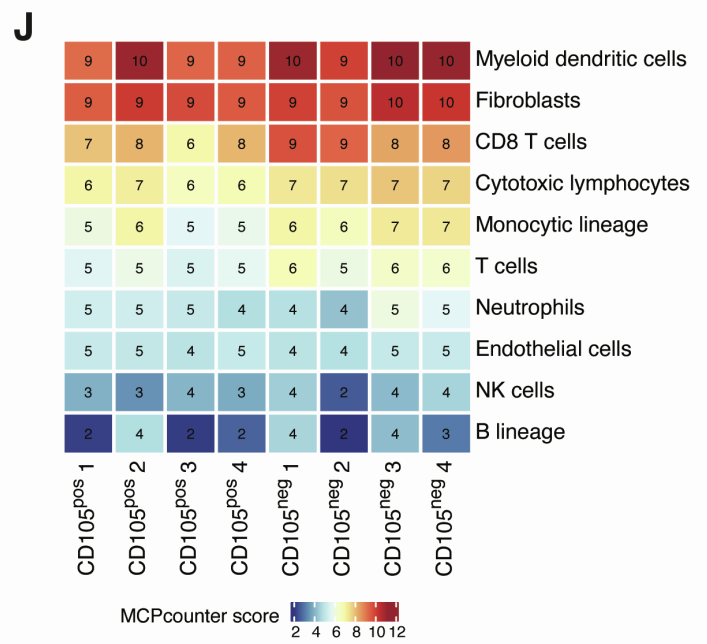
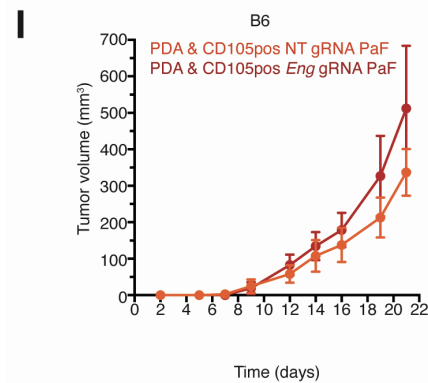
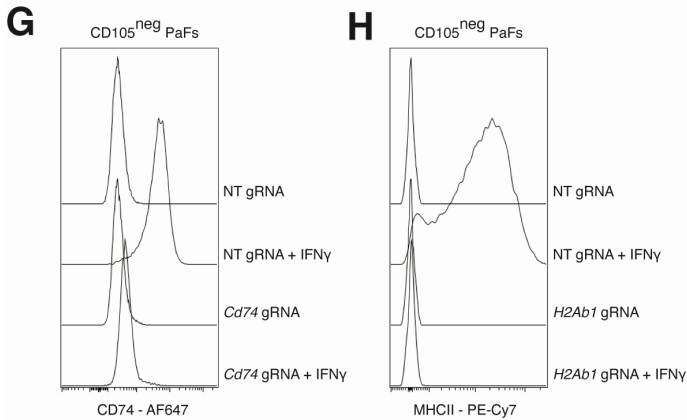
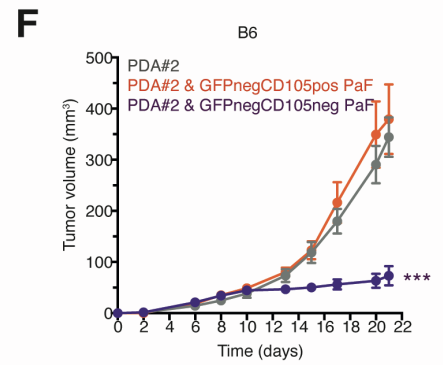
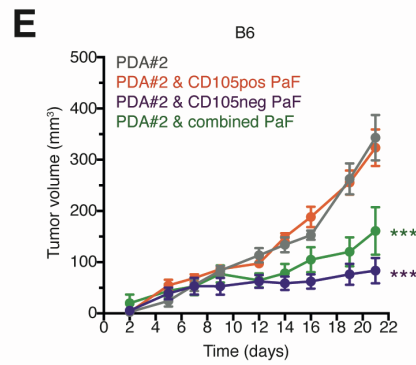
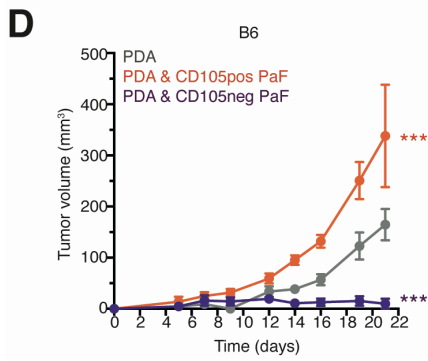
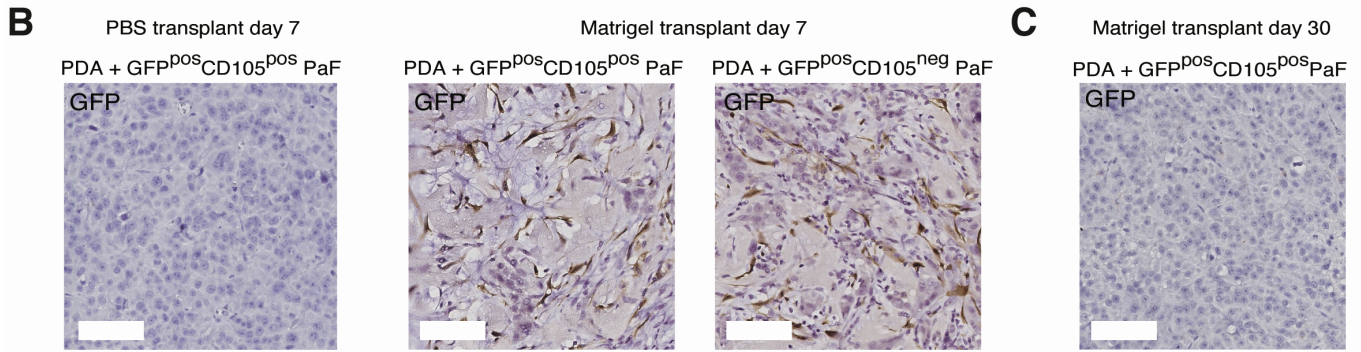
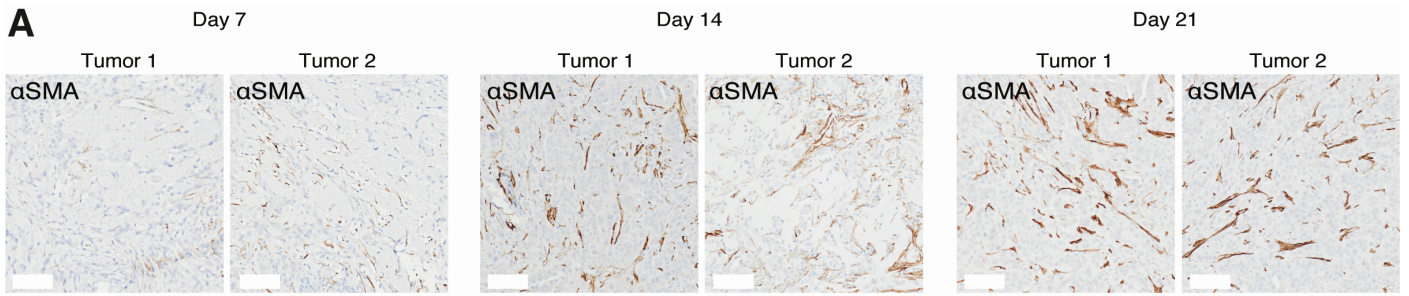
Supplemental Figure 6, related to Figure 5. Differential signaling engagement of CD105^{pos} and CD105^{neg} PaFs

(A) Flow cytometry validation of knockout of cell surface CD105 in CD105^{pos} pancreatic fibroblasts (PaFs) by CRISPR-Cas-9 gene editing.

(B) mRNA expression of genes associated with TGF β signal transduction (*Tgfrb1*, *Tgfrb2*, *Smad2* and *Smad3*), IL1 signal transduction (*Il1r1*, *Il1rap* and *Myd88*) and *Il6st* (Gp130) in CD105^{pos} (n=6) and CD105^{neg} (n=6) PaFs. Gene expression measured by RNA-seq and quantified as DEseq2 median ratio normalized expression values. Displaying expression values as mean \pm standard deviation (SD).

(C) mRNA expression of genes associated with mesothelial identity in CD105^{pos}, CD105^{pos} KO and CD105^{neg} PaFs, Gene expression measured by RNA-seq and quantified as DEseq2 median ratio normalized expression values. Displaying expression values as mean \pm standard deviation (SD).

Samples are compared using unpaired t-test (B-C). ns. non-significant, *p<0.05, **p<0.01, ***p<0.001.



Supplemental Figure 7, related to Figure 6. CD105^{neg} fibroblasts restrict tumor growth *in vivo*

(A) Immunohistochemistry (IHC) of subcutaneous tumors from mono-transplanted KPC PDA tumor cells in Matrigel at indicated time points. Stained with an α SMA targeting antibody and Haematoxylin. Representative of n=4-5 tumors. Scale bar = 80 μ m.

(B) IHC of subcutaneous tumors from co-transplanted KPC PDA tumor cells and GFP^{pos} pancreatic fibroblasts (PaFs) into syngeneic B6 mice. Stained with a GFP targeting antibody and Haematoxylin. No GFP^{pos} fibroblasts are observed in the growing tumors after 7 days when the cells are injected in PBS (left) but are highly abundant in growing tumors in which cells are injected in Matrigel (right). Scale bar = 80 μ m.

(C) No GFP^{pos} fibroblasts are retained 30 days after KPC PDA tumor cells and GFP^{pos} PaFs are co-transplanted. This likely represents a limitation of this model for studies of long-term cancer cell and fibroblast interactions *in vivo*.

(D-F) Tumor growth of subcutaneous injection of 10⁵ PDA tumor cells or co-transplantation with 10⁵ CD105^{pos} or CD105^{neg} pancreatic fibroblasts (PaFs) in syngeneic B6 mice. Shown are independent experiment from Figure 6B (D), with alternative KPC PDA cell line from a fully backcrossed B6 mouse (E) and with PaFs with no GFP expression (F). n=5 mice per condition.

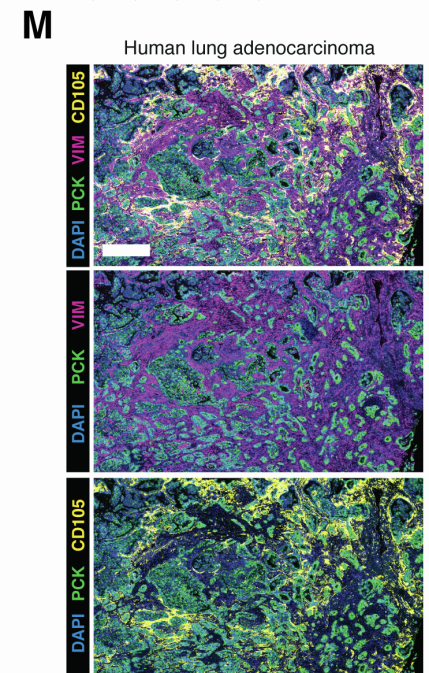
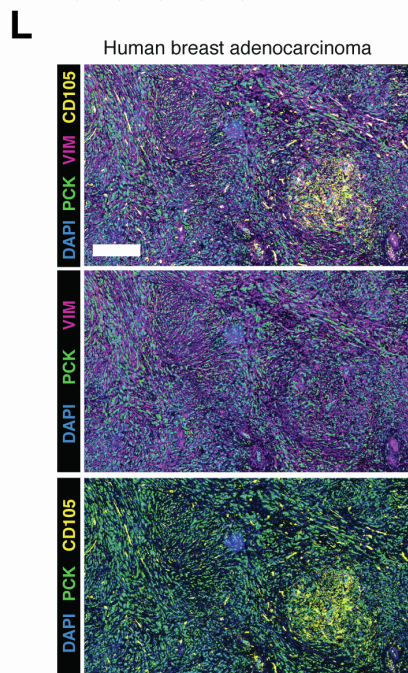
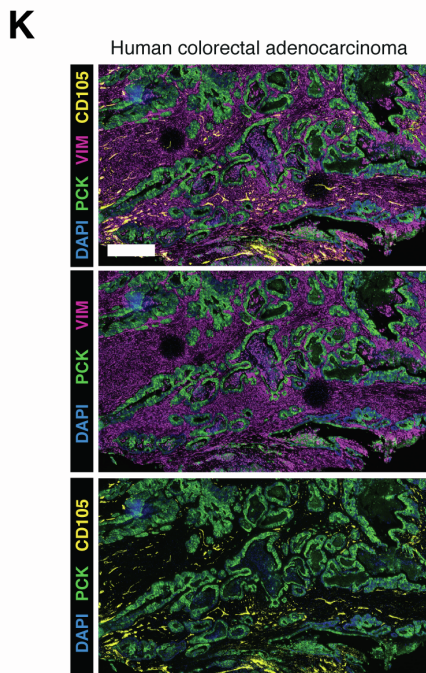
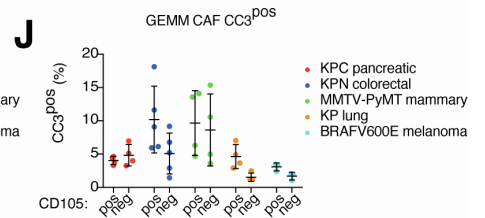
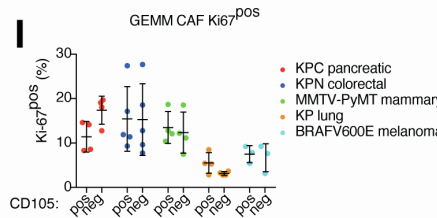
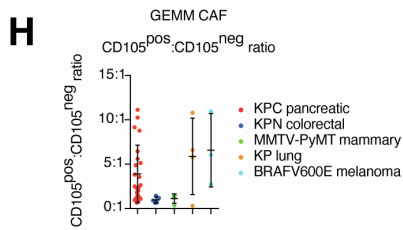
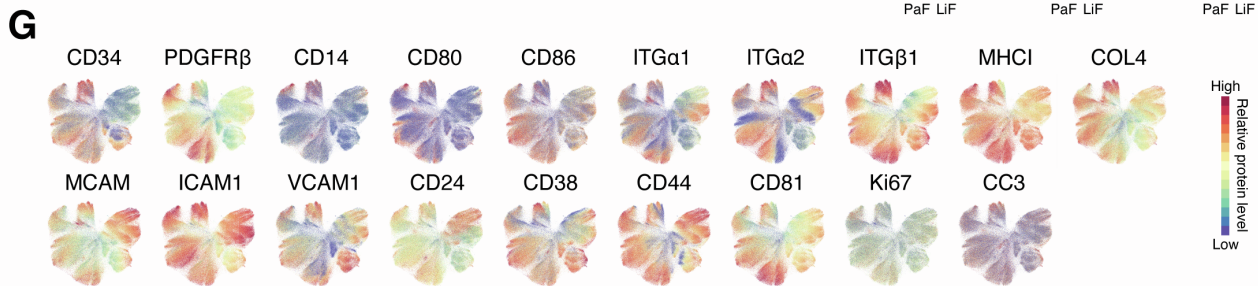
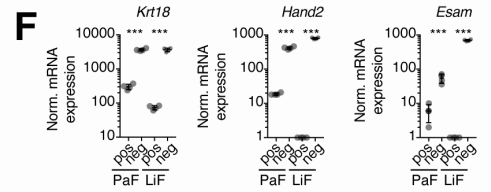
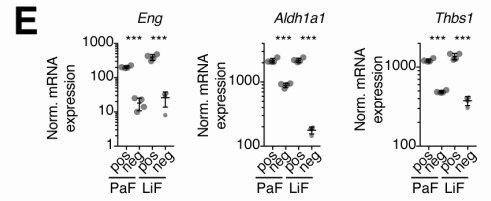
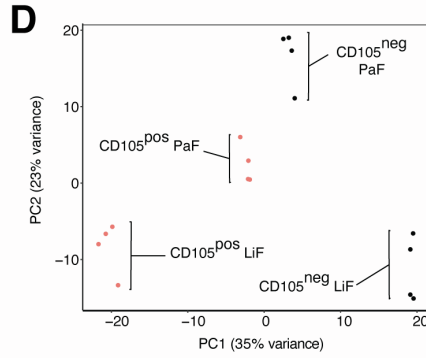
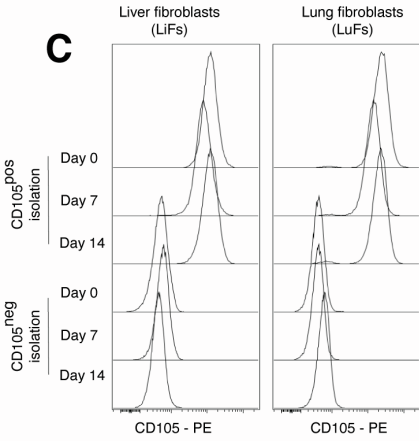
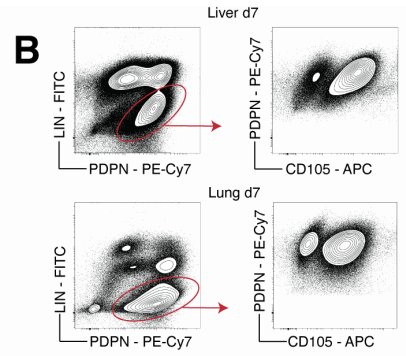
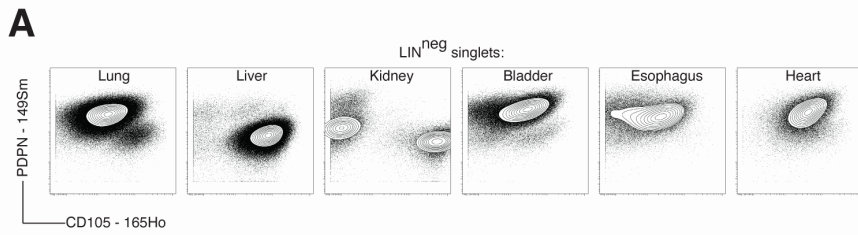
(G, H) Flow cytometry validation of CD74 (I) and MHCII (J) knock out in CD105^{neg} PaFs by CRISPR-Cas-9 gene editing.

(I) Tumor growth of subcutaneous injection of 10⁵ PDA tumor cells co-transplanted with 10⁵ CD105^{pos} PaFs with Eng expression disrupted by CRISPR-Cas-9 gene editing. CD105^{pos} PaFs transfected with non-targeting (NT) gRNAs are used as control. n=5 mice per condition.

(J) MCPcounter analysis of bulk gene expression profiles of subcutaneous KPC PDA and CD105^{pos} PaFs or CD105^{neg} PaF co-transplanted tumors at day 10.

Data is displayed as mean tumor volumes \pm standard error of the mean (SEM) (D-F and I).

Conditions are compared using 2-way ANOVA (B-G). *p<0.05, **p<0.01, ***p<0.001.



Supplemental Figure 8, related to Figure 7. CD105^{pos} and CD105^{neg} fibroblasts are identified in normal and tumor-bearing tissues

(A) Mass cytometry (MC) analysis of *in vitro* expanded, low passage, primary fibroblast isolations from mouse organs not displayed in Figure 7A. Cells expressing canonical markers of epithelial, immune, endothelial and perivascular cells are excluded from the analysis and live, singlets are displayed in plots showing PDPN and CD105 levels. LIN: EpCAM CD45 CD31

(B) Flow cytometry analysis of EpCAM^{neg} CD45^{neg} CD31^{neg} (LIN^{neg}) and PDPN^{pos} liver (top) and lung (bottom) fibroblasts from day 7 primary cell isolations, an earlier time-point than the analysis of these isolations by MC in Figure 7A and Figure S7A.

(C) Flow cytometry analysis of purified and CD105^{pos} and CD105^{neg} liver (LiF) and lung (LuF) fibroblasts after 1 and 2 weeks of *in vitro* culture. Plots are representative of n=4 experiments.

(D) Principle component (PC) analysis of differentially expressed genes between *in vitro* expanded CD105^{pos} and CD105^{neg} LiFs and pancreatic fibroblasts (PaFs).

(E-F) mRNA expression of genes associated with CD105 expression in liver and pancreatic fibroblasts. Gene expression measured by RNA-seq and quantified as DEseq2 median ratio normalized expression values. Displaying expression values as mean ± standard deviation (SD).

(G) UMAP projection from Figure 7B, displaying relative intensity of additional phenotypic markers not included in Figure 7F.

(H) MC analysis of the ratio of CD105^{pos}:CD105^{neg} CAFs in tumors from genetically engineered mouse models (GEMMs) of pancreatic (KPC) (n=23), colorectal (KPN) (n=5), mammary (MMTV-PyMT) (n=4), lung (KP) (n=4) and melanoma (BRAF^{v600E}) (n=3) cancer. Data from the n=19 PDA tumor samples analyzed in Figure 1 were combined to increase the accuracy of the ratio measured for the pancreatic GEMM tumors. Displayed as mean ± standard deviation (SD).

(I-J) MC analysis of proliferation (I) and apoptosis (J) rates of CD105^{pos} and CD105^{neg} GEMM CAFs. n=5 tumors for each genotype (paired CD105^{pos} and CD105^{neg} fractions). Displayed as mean ± SD.

(K-M) Immunohistochemistry (IHC) analysis of additional representative human colorectal (K), breast (L) and lung adenocarcinoma (M) tumor samples stained for pan-cytokeratin (PCK) (green), vimentin (VIM) (purple), CD105 (yellow) and DAPI (blue). Scale bar = 500 μm.