

**Title: Oxidative stress-induced impairment of trophoblast function causes preeclampsia through the unfolded protein response pathway**

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**SUPPLEMENTARY DETAILS**

**Methods**

**Protocol for early villi explant culture:** Tissue explants from early villi (n=5) were cultured. Small sections weighing 10 mg was cut and plated in Collagen-I coated single well of a 12 well plate (Corning). The tissue sections were washed thoroughly using 1X phosphate buffered saline (PBS) followed by addition of RPMI-1640 medium (HyClone) containing 10% FBS (Invitrogen) and 1% penicillin-streptomycin (Invitrogen). The well plate was then kept at 37°C in a humidified chamber with 21% O<sub>2</sub> and 5% CO<sub>2</sub>.

For experiments, sections from each early villi was divided into four groups- (i) control, (ii) sections treated with H<sub>2</sub>O<sub>2</sub> for 24 h, (iii) sections treated with N-acetyl cysteine (NAC) for 24 h and (iv) sections pre-treated with NAC for 2 h followed by H<sub>2</sub>O<sub>2</sub> treatment for another 24h.

**Legends:**

**Fig 1.** Early villi (10 weeks), term placenta (38 weeks) and Preeclampsia placenta (36 weeks) were collected from the patients and measured for dimensions. PE placenta (Fig. 1c) was smaller in size compared to term placenta (Fig 1b). Early villi (Fig 1a) tissue was well branched.

**Fig 2a.** Patient samples were screened by checking the level of the biomarker of Preeclampsia. sFlt-1 concentration was measured in the blood serum of the normal pregnant women (NP) and Preeclamptic women (PE) collected two days prior to delivery. **Fig 2(b, c).** Concentration of sFlt-1 was also measured in the conditioned media of the trophoblast cells (BeWo & HTR8/SVneo) treated with H<sub>2</sub>O<sub>2</sub>. All data are shown as Mean ± Standard deviation. Results are representative of at least twelve independent experiments. \* p < 0.05; \*\* p < 0.01; \*\*\* p < 0.001 (Student's t-test).

**Fig 3a-b.** Images of trophoblast cells HTR8/SVneo & BeWo respectively were taken after the cells were treated with different dose of H<sub>2</sub>O<sub>2</sub> viz. 20 μM, 40 μM, 80 μM, 100 μM, 200 μM. The images were taken at 100X Magnification. Each experiment was repeated three times.

**Fig 4.** Flow-cytometric analysis was done to determine the sub lethal dose of H<sub>2</sub>O<sub>2</sub> in the trophoblast cells. A tabular representation of the data has been given. All data are shown as Mean ± Standard deviation. Results are representative of at least three independent experiments. \* p < 0.05; \*\* p < 0.01; \*\*\* p < 0.001 (Student's t-test).

**Fig 5.** Effect of H<sub>2</sub>O<sub>2</sub> on the viability of BeWo and HTR8/SVneo cells. **(a-b)** WST-1 and Annexin-FITC-PI staining were used to assess and quantify the viability of cells. Cells were treated with different doses of H<sub>2</sub>O<sub>2</sub> to induce oxidative stress in the cells. WST-1 assay was performed with BeWo and HTR8/SVneo cells to check the sub-lethal dose. **(c-d)** Cell death was measured by flow cytometry using Annexin V-FITC and PI double staining. BeWo and HTR8/SVneo cells were exposed to the chosen concentrations of H<sub>2</sub>O<sub>2</sub>. In each density plot quadrant Q1: shows necrotic cells (Annexin<sup>-</sup> PI<sup>+</sup>); Q2: late apoptotic cells (Annexin<sup>+</sup> PI<sup>+</sup>); Q3: shows the viable cells (Annexin<sup>-</sup> PI<sup>-</sup>) and Q4: early apoptotic cells (Annexin<sup>+</sup> PI<sup>-</sup>) Results are representative of at least three independent experiments..

**Fig 6.** Protein expression of Caspase 3 and PARP was assessed using western blotting. The data was analysed using ImageJ. All data are shown as Mean ± Standard deviation. Results are representative of at least three independent experiments. \* p < 0.05; \*\* p < 0.01; \*\*\* p < 0.001 (Student's t-test).

**Fig 7.** The early villi explants plated on Collagen-I coated well plate have been shown. Four different wells have been considered four different treatment groups.

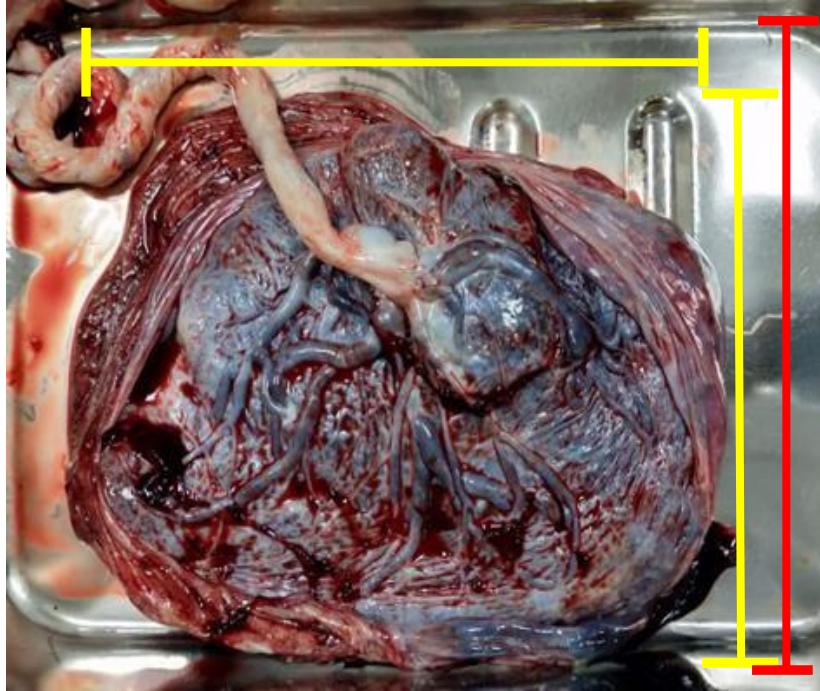
Suppl.Table 1. Details of primers used in this study.

<b>S.No</b>	<b>Genes</b>	<b>Forward Primer (5' – 3')</b>	<b>Reverse Primer (5' – 3')</b>	<b>Amplicon Size (bp)</b>
1	SYN-1	CTGTTGGACTTACTTCACCCAAA	GGTACGGAGGGTTTCATGTAGT	169
2	SYN-2	AGCCCCTATTTGTGTTATGGC	GGAATTGGTTGTGGGTGTATGT	134
3	MFSD2A	CCATTGATGAGGAGAGGCGG	CCTTCTGTGGCCTTCTGCAT	162
4	SLC1A5	TCATGTGGTACGCCCTGT	GCGGGCAAAGAGTAAACCCA	186
5	DYSF	AAGAACAGCGTGAACCCTGTA	CCTCTCGGAGTGGGACCTT	157
6	β-hCG	CTACTGCCCCACCATGACCC	GCAGAGTGCACATTGACAGC	172
7	MMP-2	ATGACAGCTGCACCACTGAG	ATTTGTTGCCCAGGAAAGTG	174
8	MMP-9	CTCGAACTTTGACAGCGACA	GCCATTCACGTCGTCCTTAT	187
9	TIMP-1	CATTGCTGGAAAACCTGCAGGA	GCAGTTTGCAGGGGATGGAT	167
10	TIMP-2	GGCTGCGAGTGCAAGATCAC	TCGAGAAACTCCTGCTTGGG	197
11	UPA	CCCAGGAAATGGGACAGGG	ACAGTTCGCCTGTTCGTATCT	199
12	PAI	CATCCTGGAACCTGCCCTACC	AGGGAGAACTTGGGCAGAAC	174
13	PLAC8	GGAACAAGCGTCGCAATGAG	AAAGTACGCATGGCTCTCCTT	152
14	GAPDH	ACGGATTTGGTCGTATTGGG	CGCTCCTGGAAGATGGTGAT	214

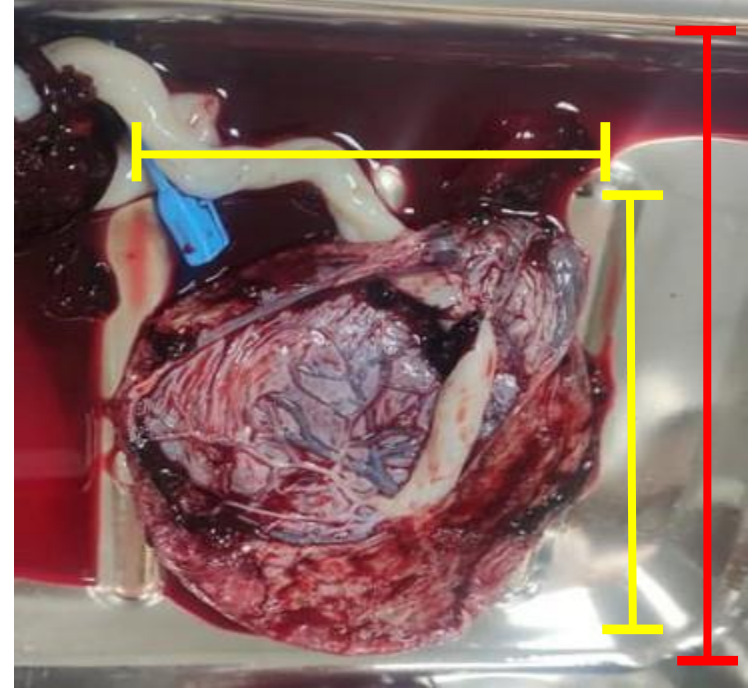
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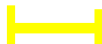


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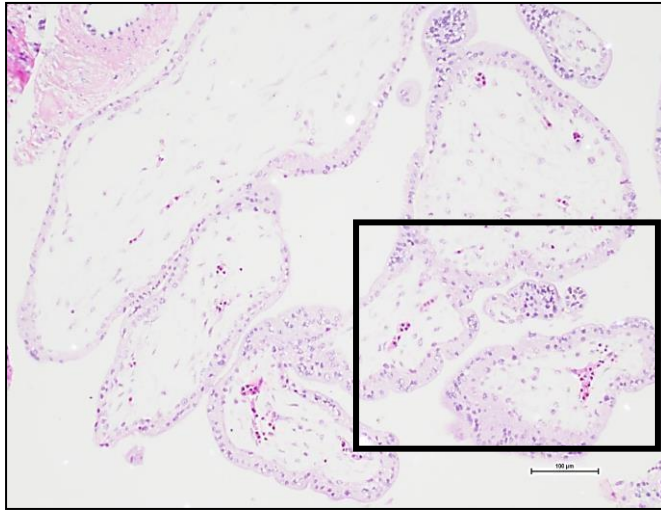
c.



	Dimensions of the container
	Diameter of the placenta

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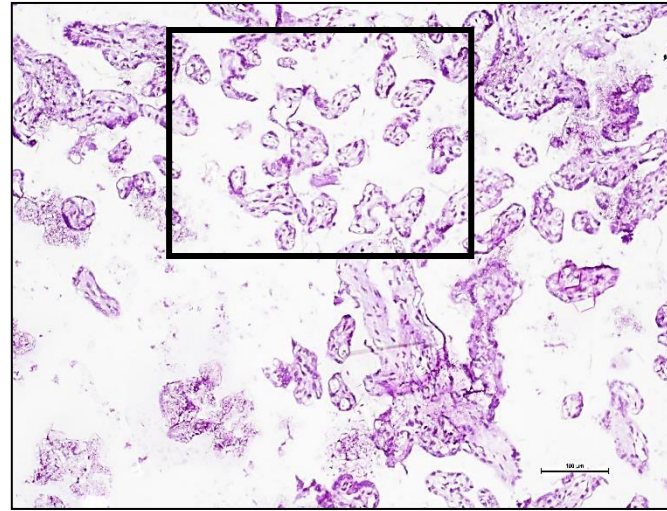
**EP**



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**b.**

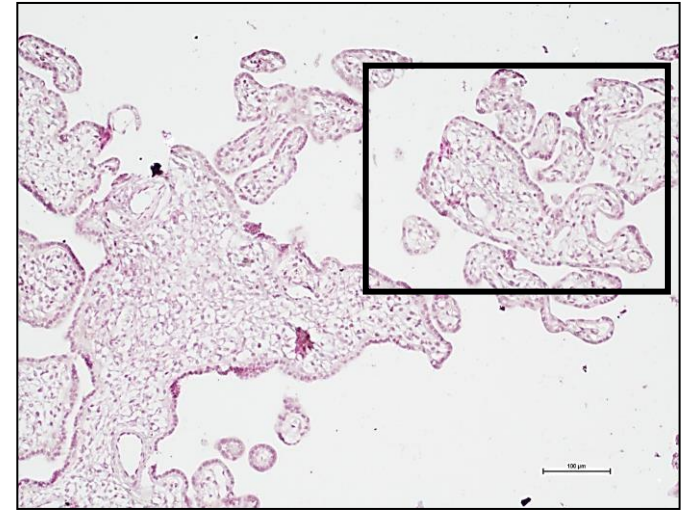
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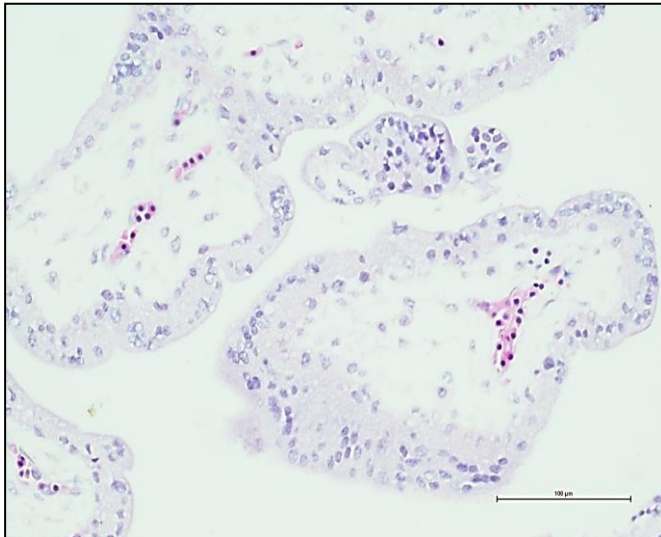
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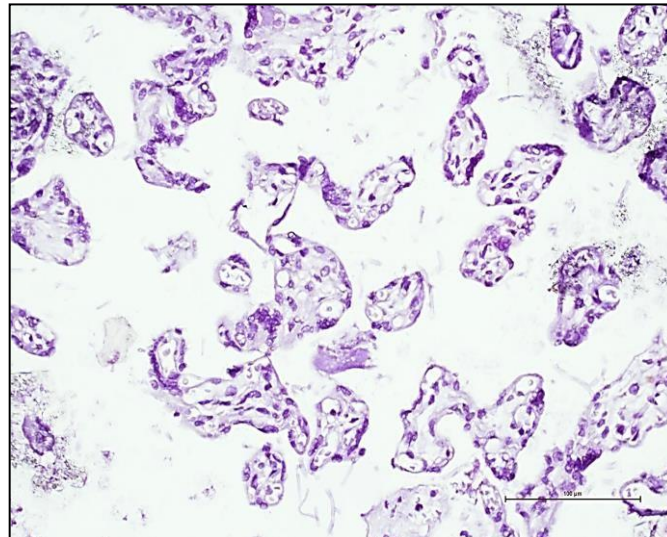
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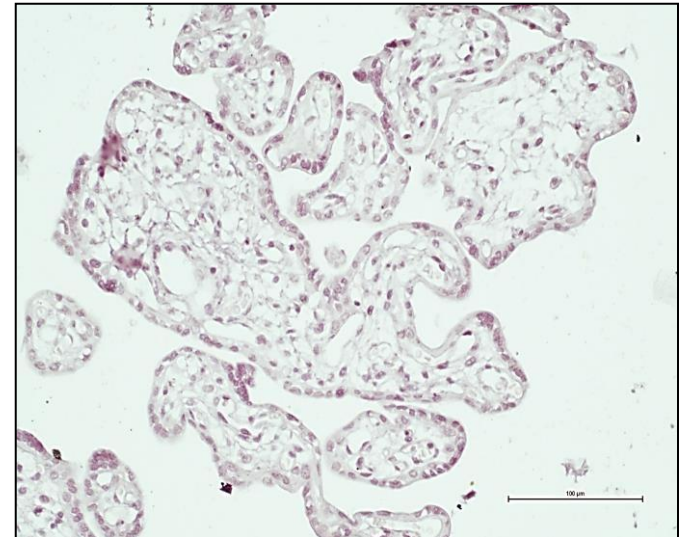
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**20X**

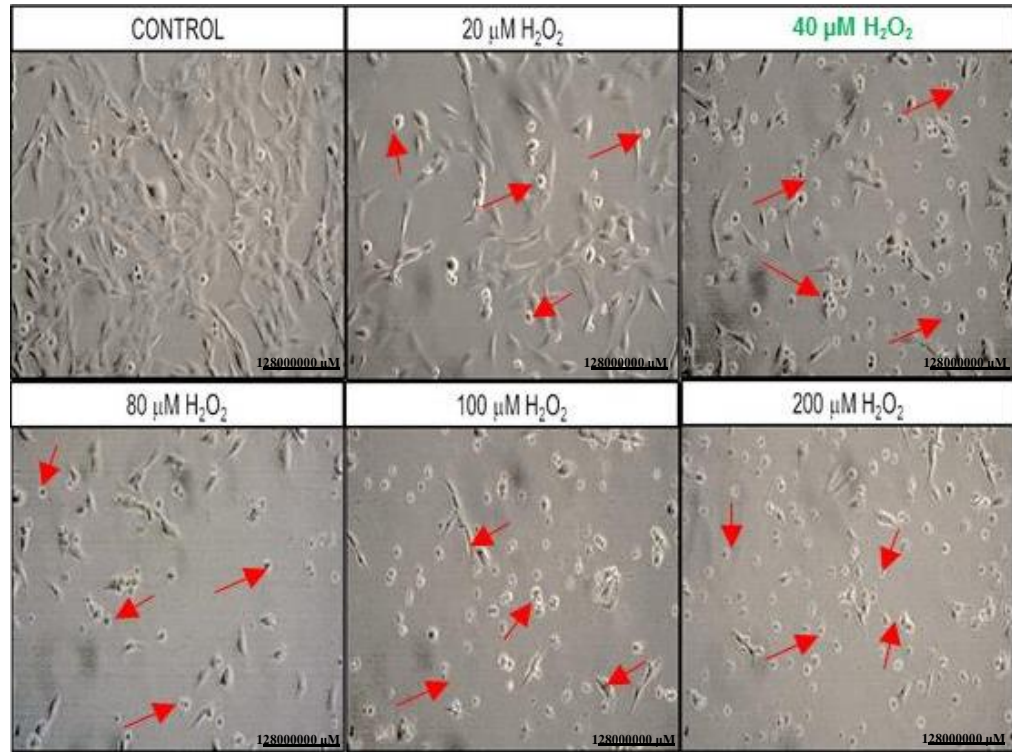


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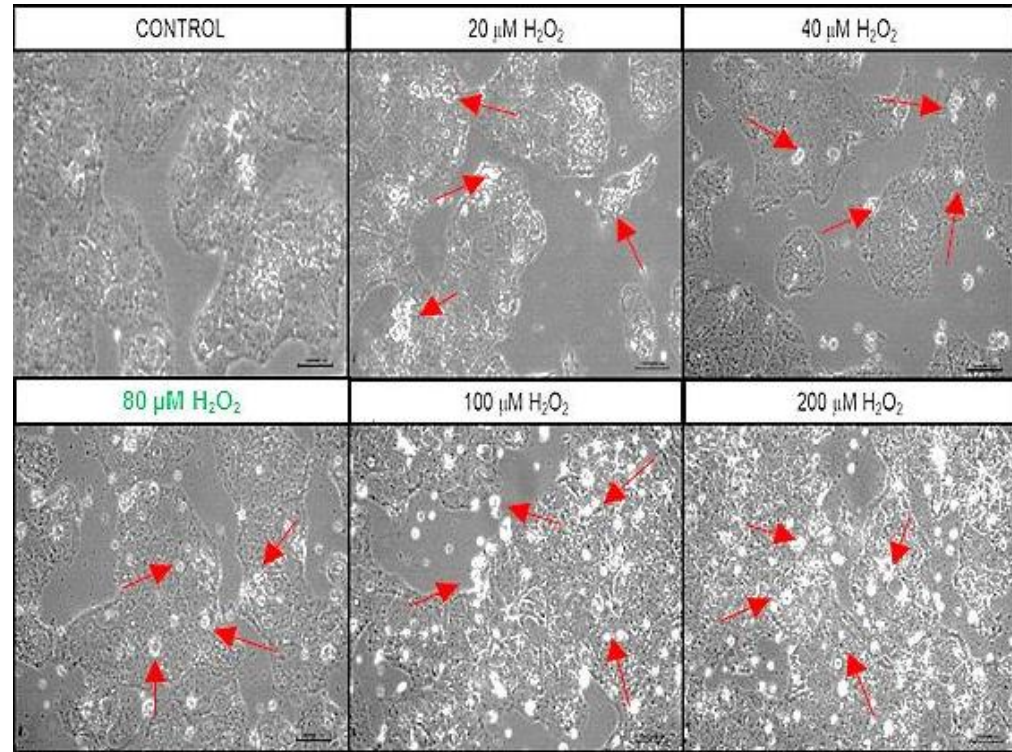
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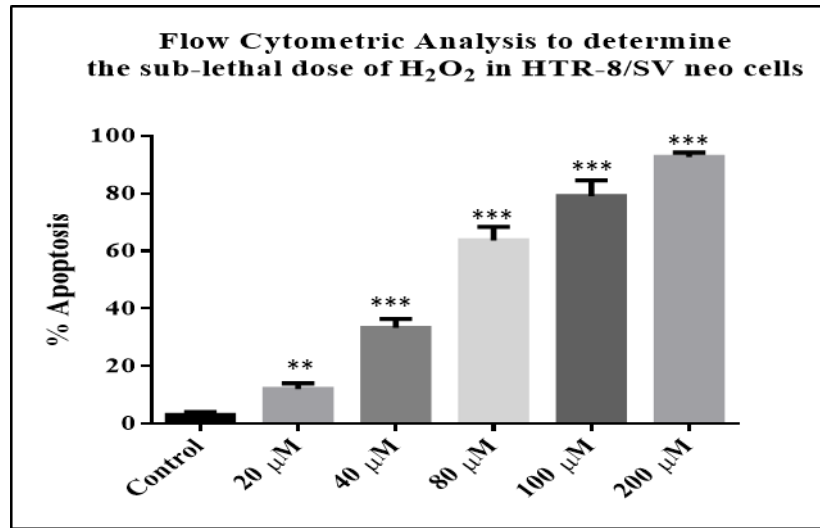
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**b.**



*n=3; Magnification: 10X*

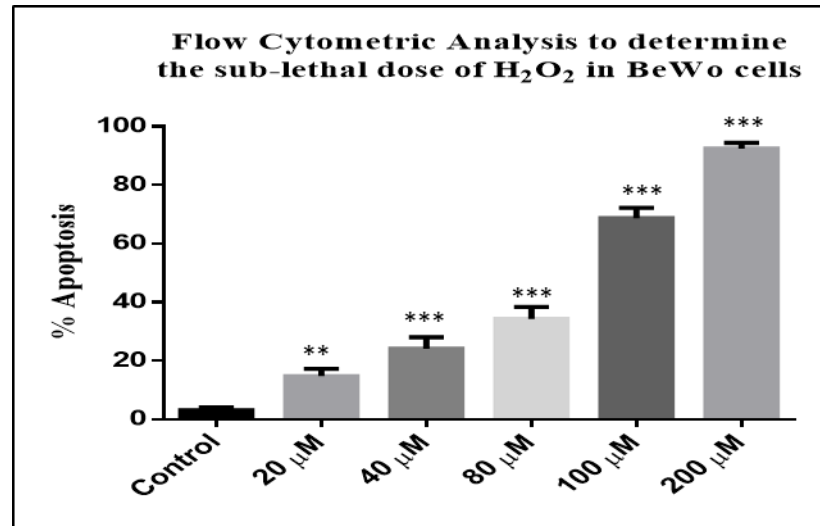
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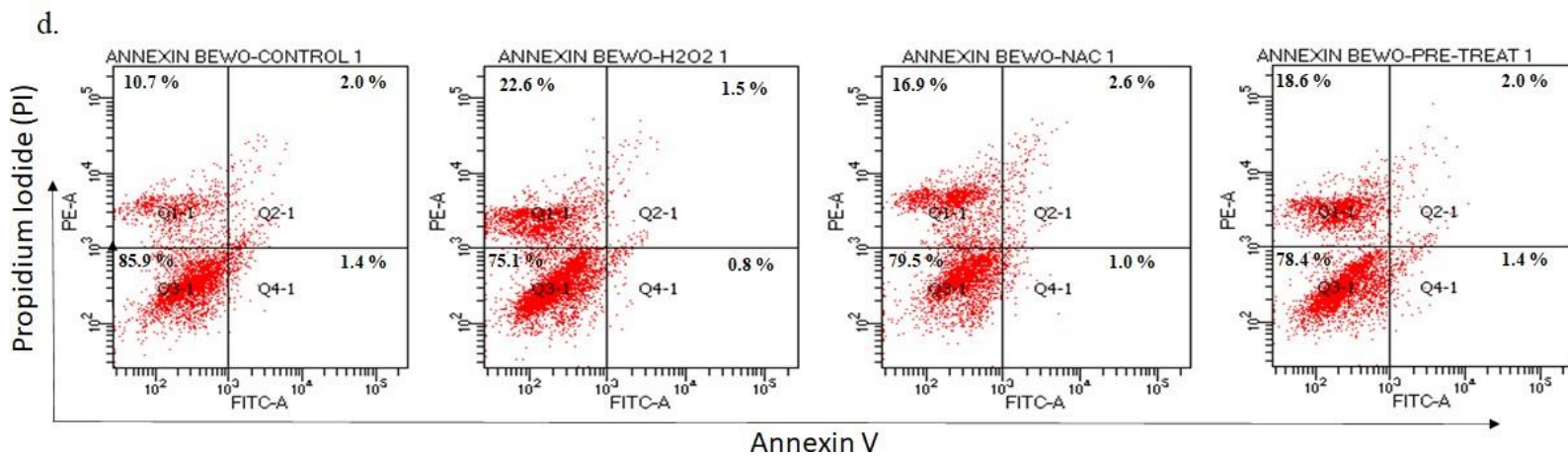
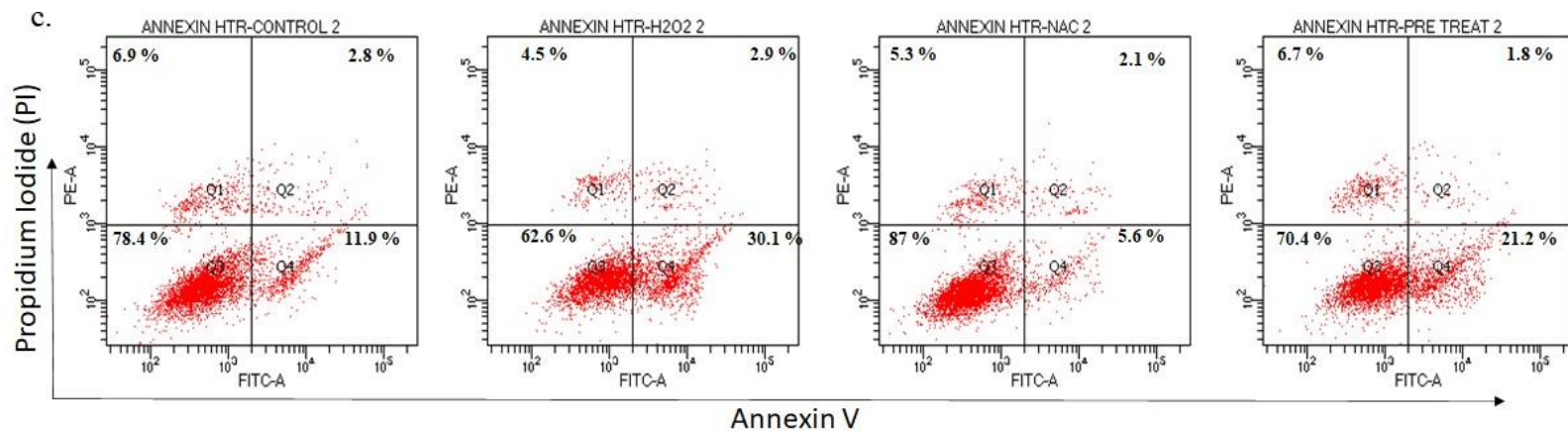
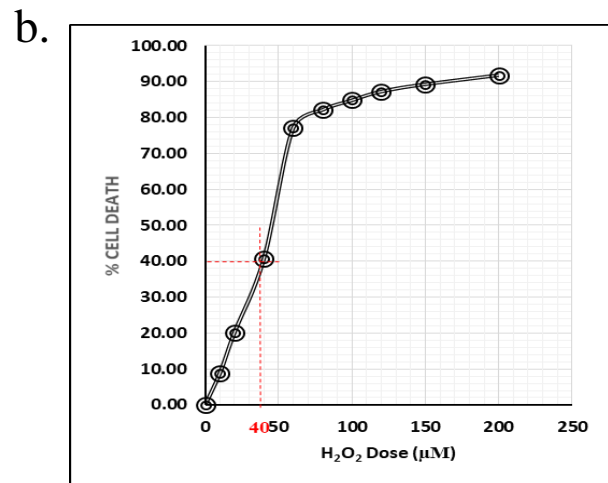
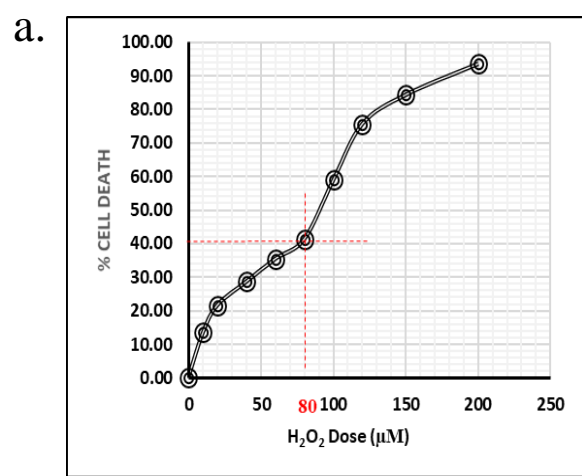
Treatment Groups in HTR-8/ SV neo cells	Percent Apoptosis
Control	2 ± 1
20 μM H <sub>2</sub> O <sub>2</sub>	12 ± 2 **
40 μM H <sub>2</sub> O <sub>2</sub>	34 ± 2 ***
80 μM H <sub>2</sub> O <sub>2</sub>	62 ± 5 ***
100 μM H <sub>2</sub> O <sub>2</sub>	87 ± 7 ***
200 μM H <sub>2</sub> O <sub>2</sub>	92 ± 5 ***

c.

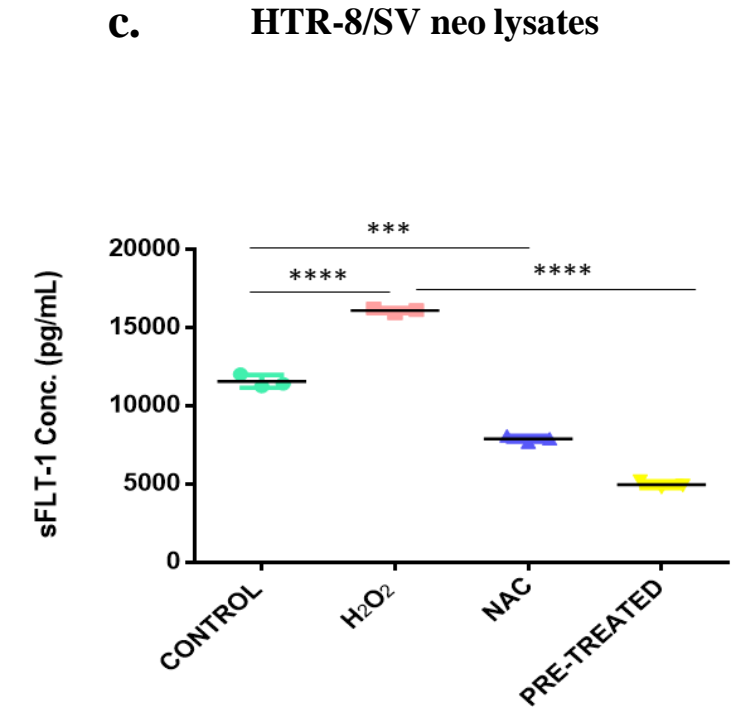
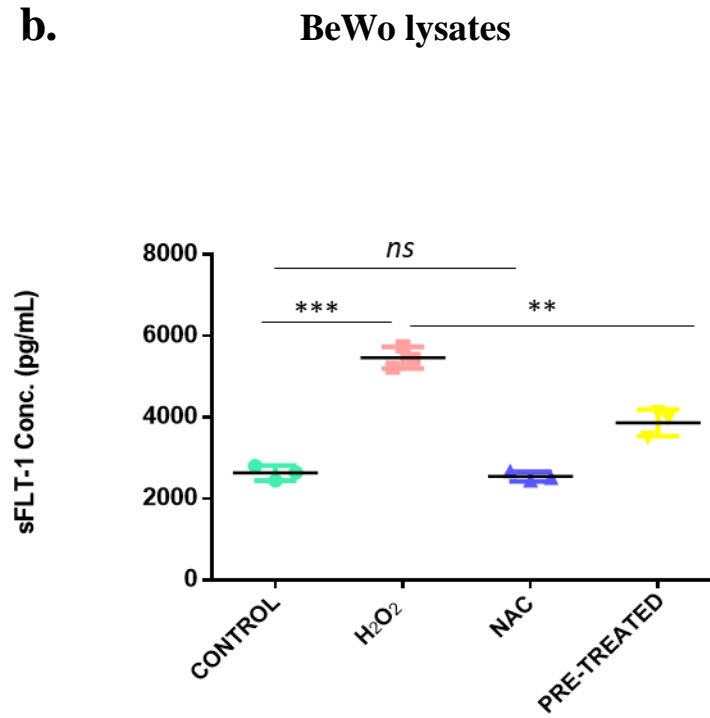
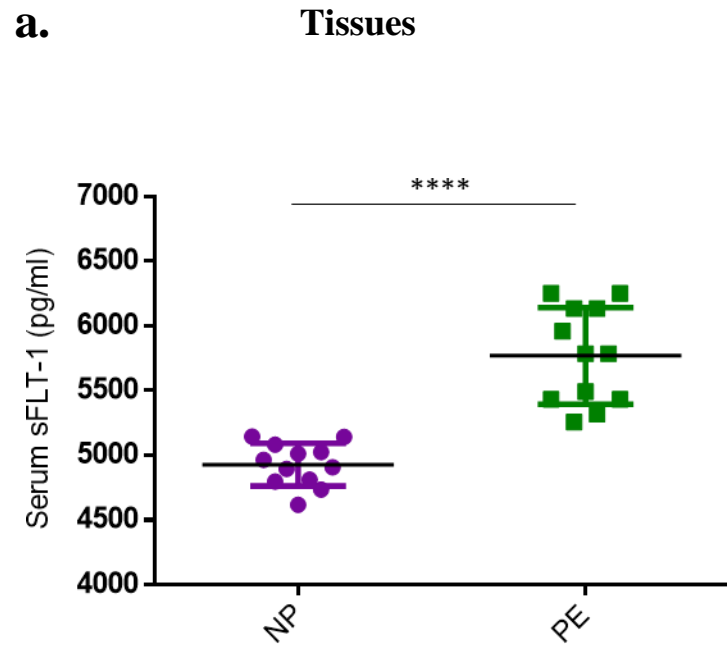


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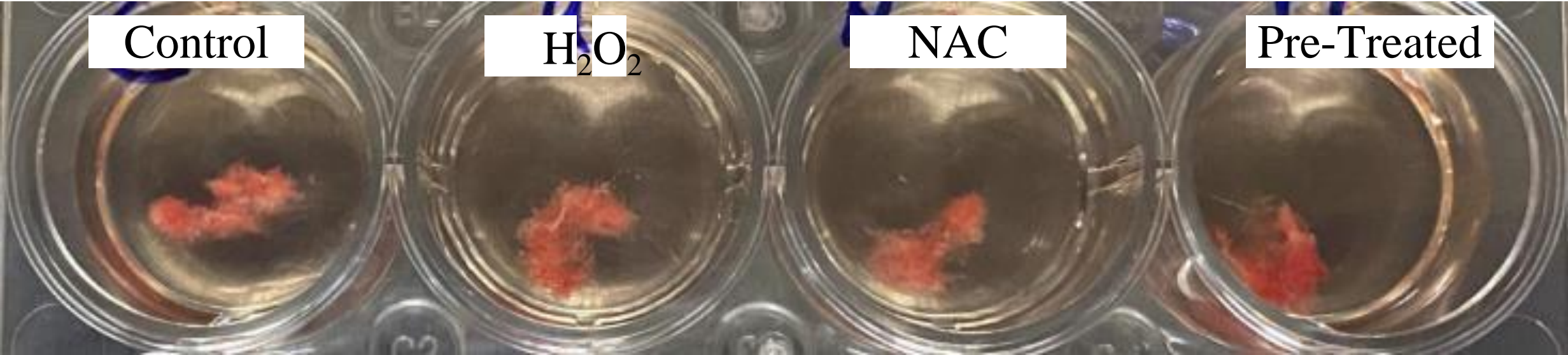
Treatment Groups in BeWo cells	Percent Apoptosis
Control	2 ± 1
20 μM H <sub>2</sub> O <sub>2</sub>	12 ± 2 **
40 μM H <sub>2</sub> O <sub>2</sub>	24 ± 2 ***
80 μM H <sub>2</sub> O <sub>2</sub>	38 ± 5 ***
100 μM H <sub>2</sub> O <sub>2</sub>	69 ± 7 ***
200 μM H <sub>2</sub> O <sub>2</sub>	90 ± 9 ***





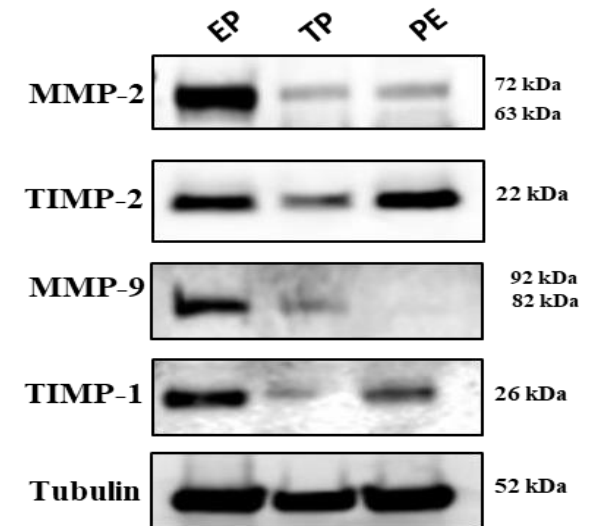
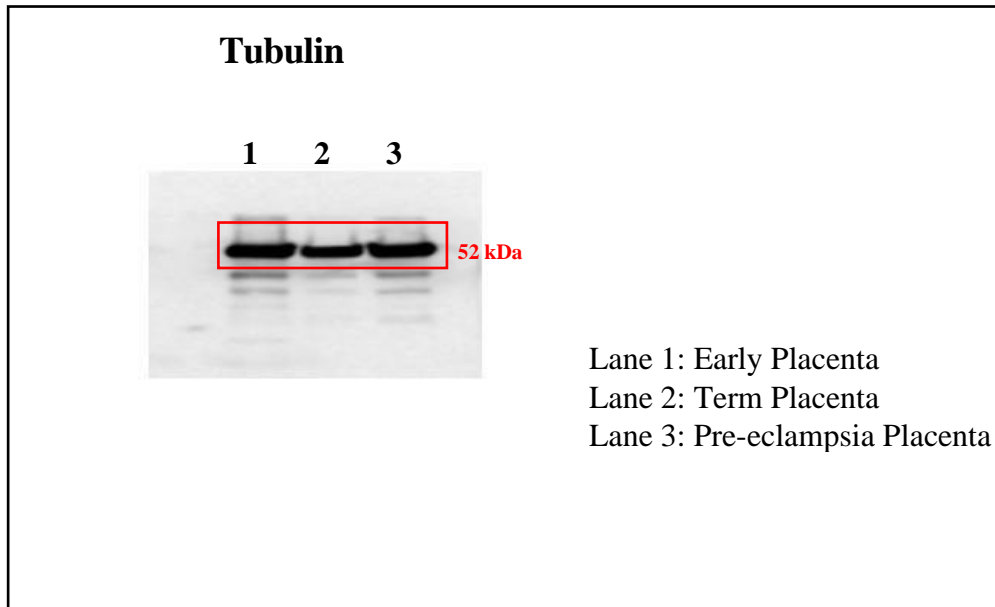
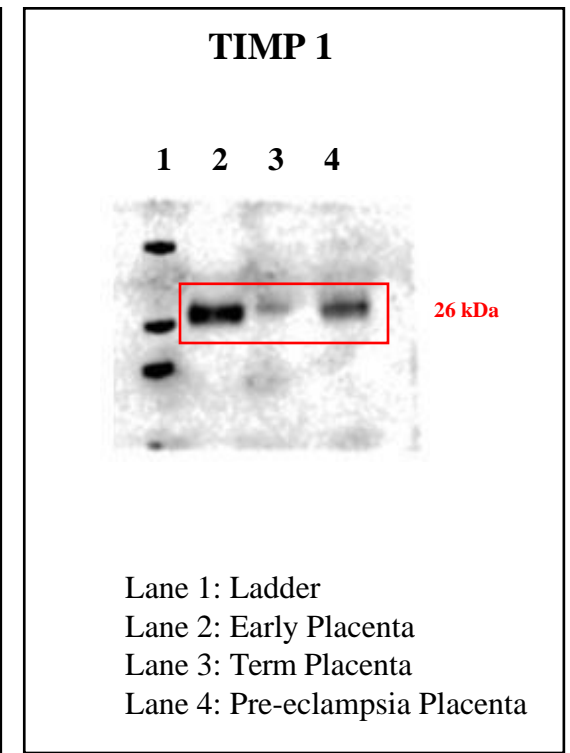
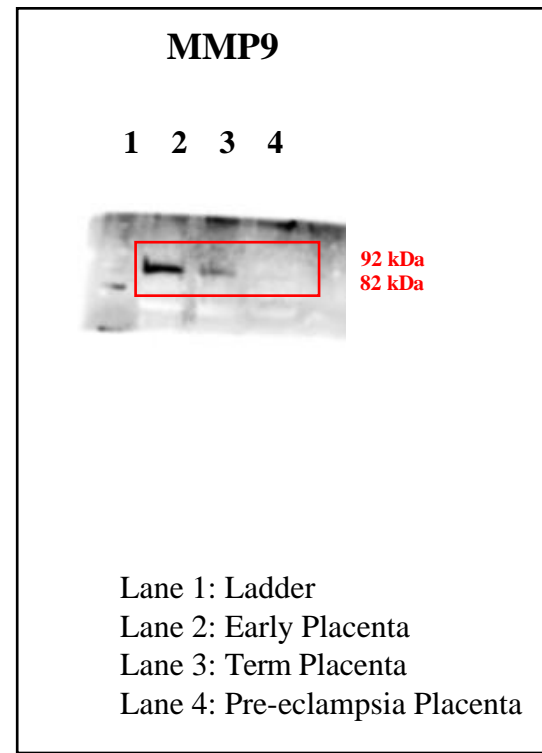
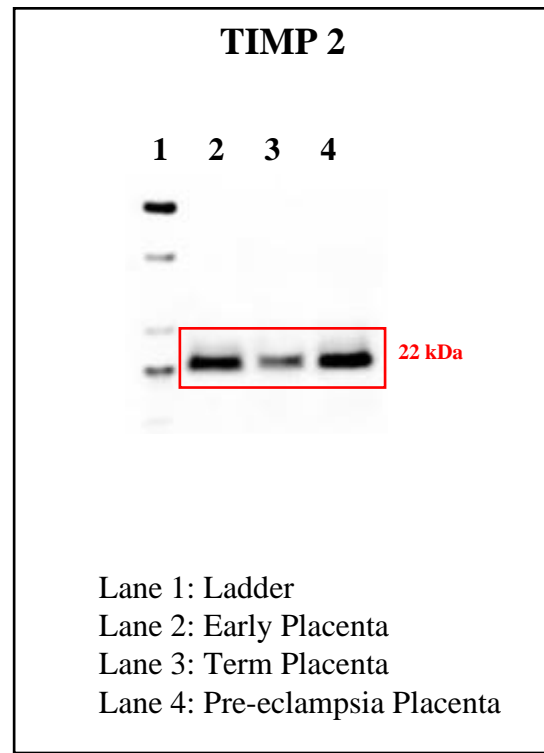
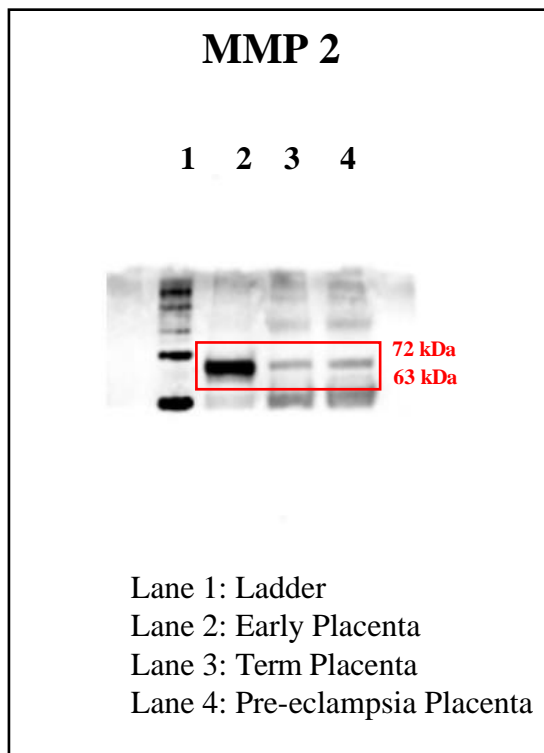


Suppl. Fig. 7



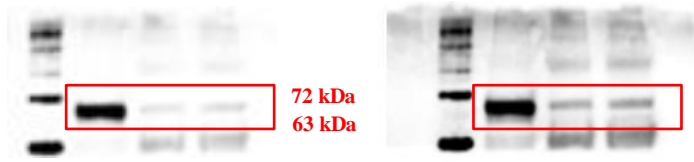
Original Blots





**Figure 2h.**

### MMP-9

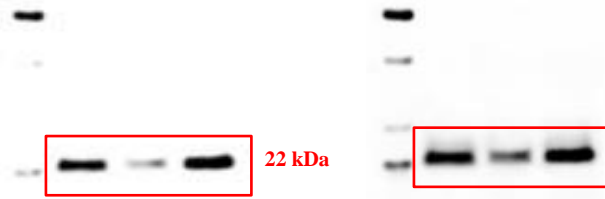


Exposure 1

Exposure 2

(Incorporated image)

### TIMP-2

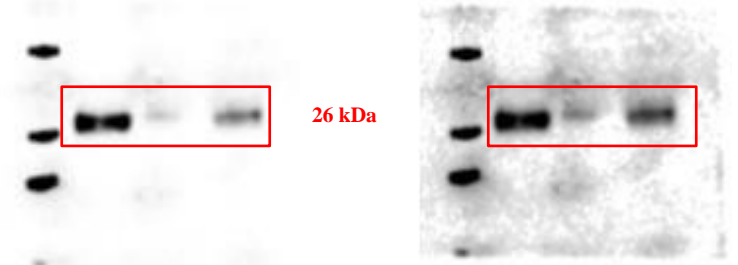


Exposure 1

Exposure 2

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### TIMP-1



Exposure 1

Exposure 2

(Incorporated image)

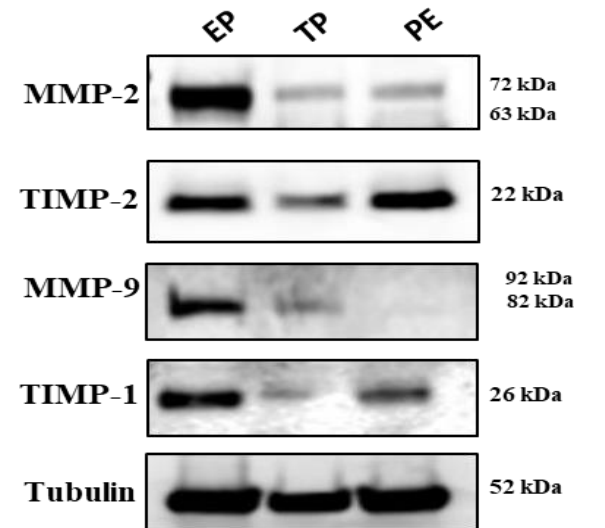
### MMP-2



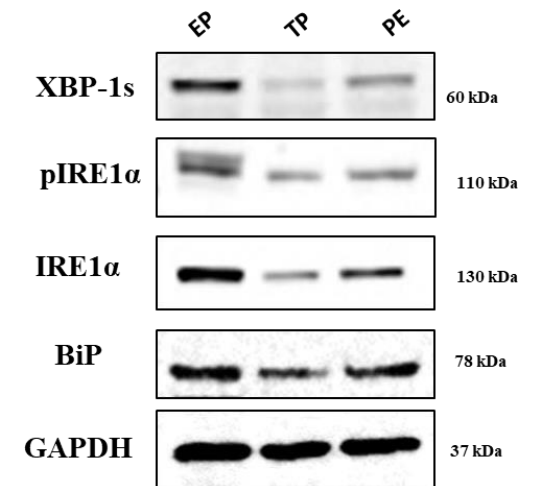
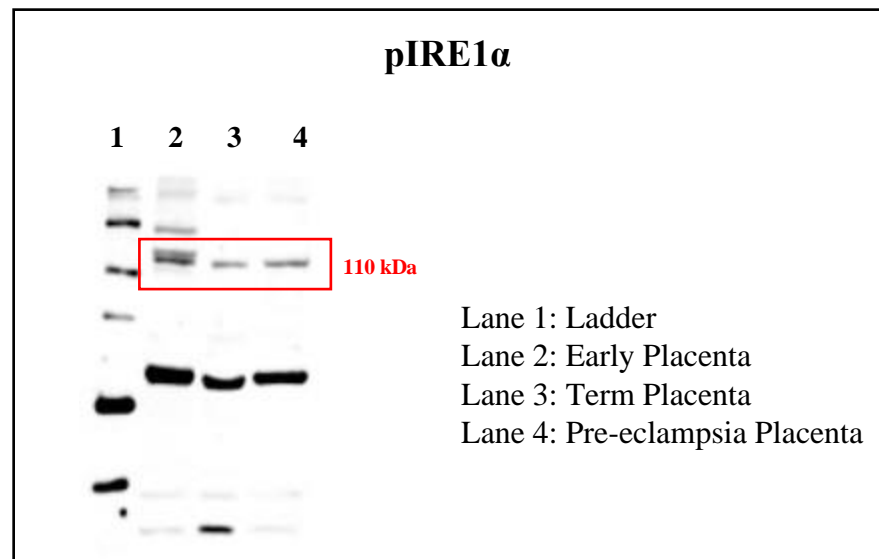
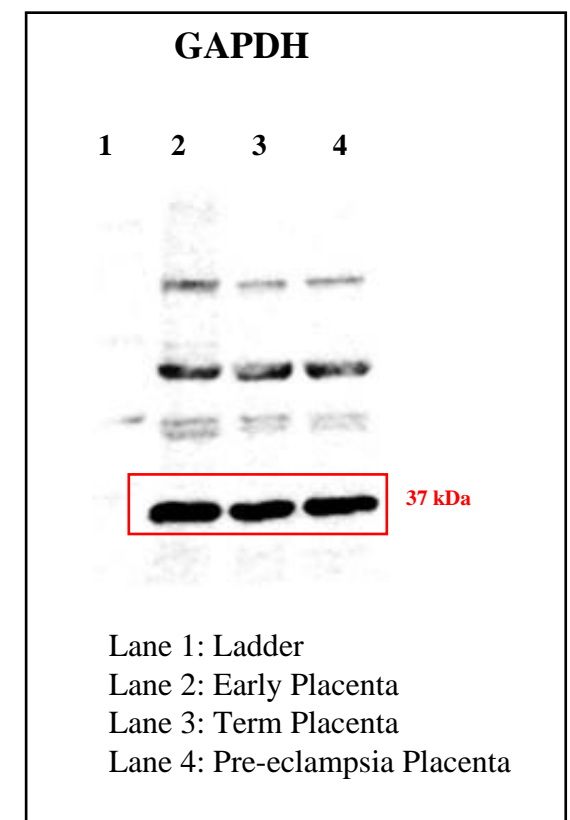
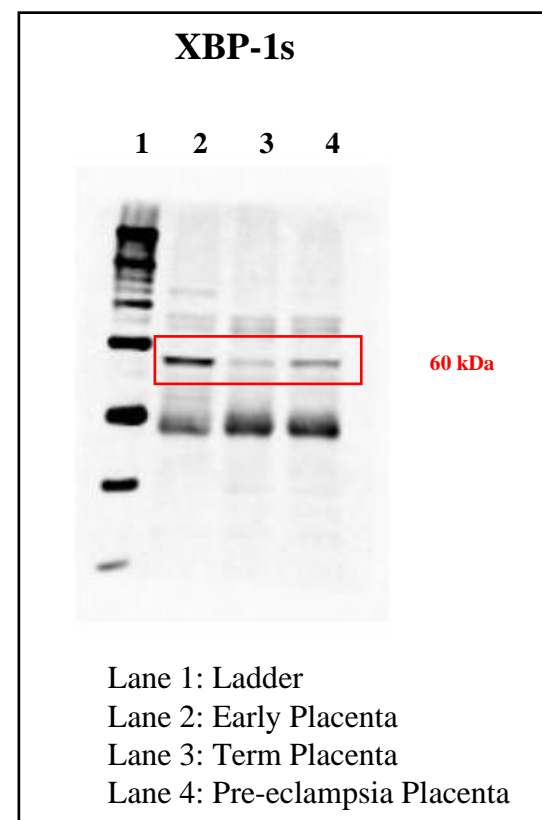
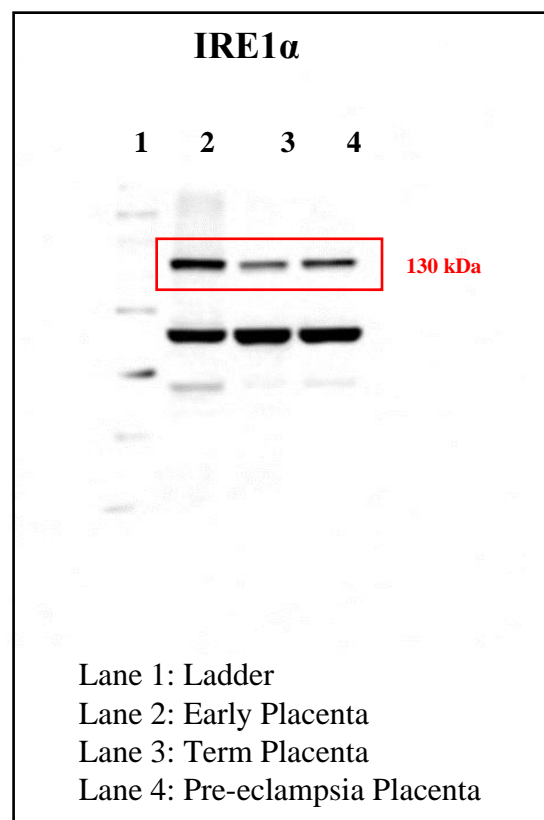
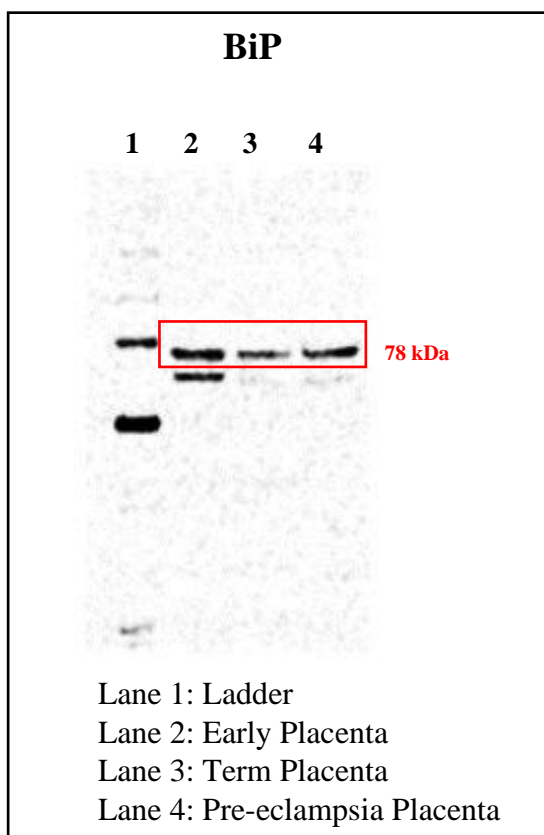
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Exposure 2

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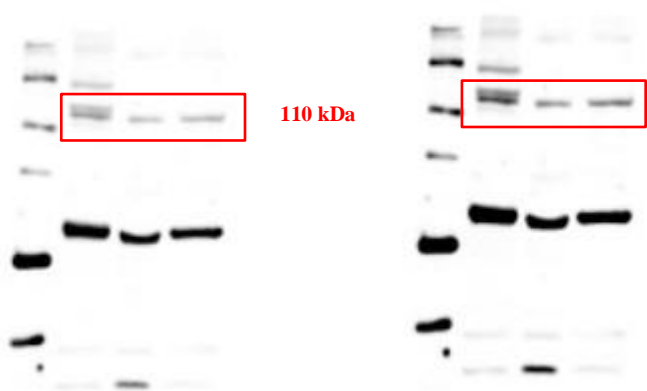


**Figure 2h.**



**Figure 4a.**

**pIRE1 $\alpha$**



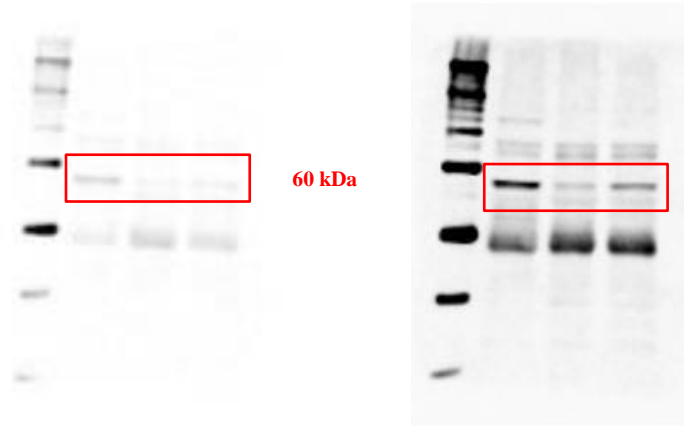
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Exposure 1

Exposure 2

(Incorporated image)

**XBP-1s**



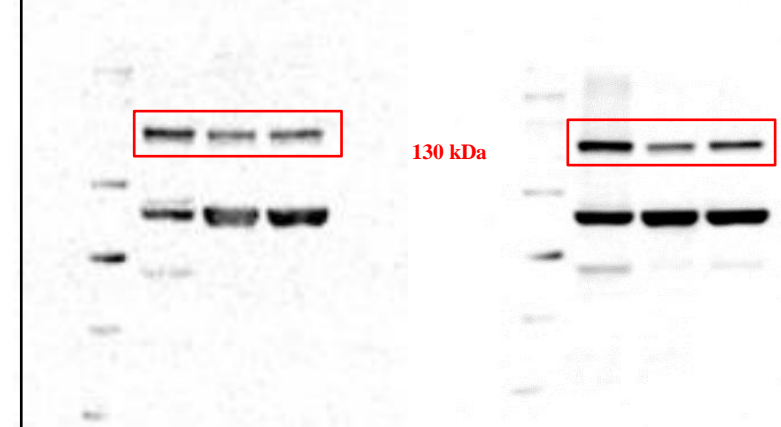
60 kDa

Exposure 1

Exposure 2

(Incorporated image)

**IRE1 $\alpha$**



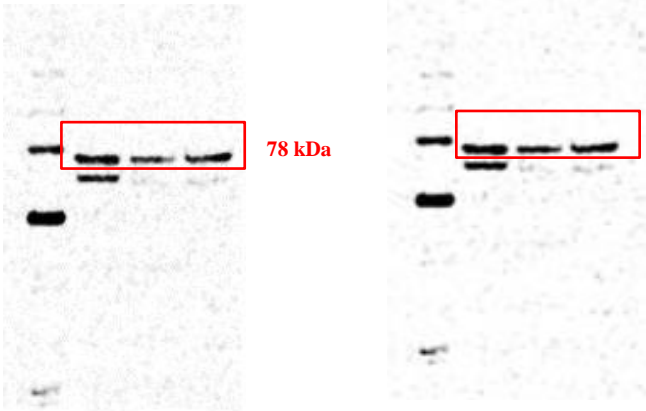
130 kDa

Exposure 1

Exposure 2

(Incorporated image)

**BiP**



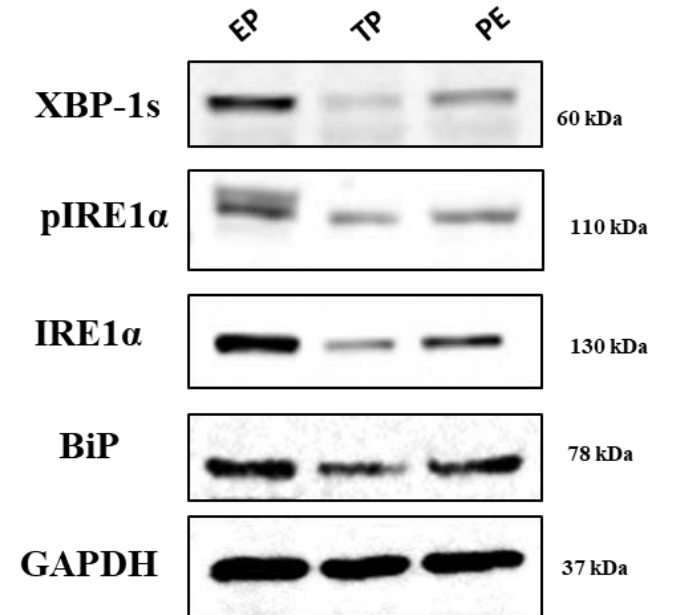
78 kDa

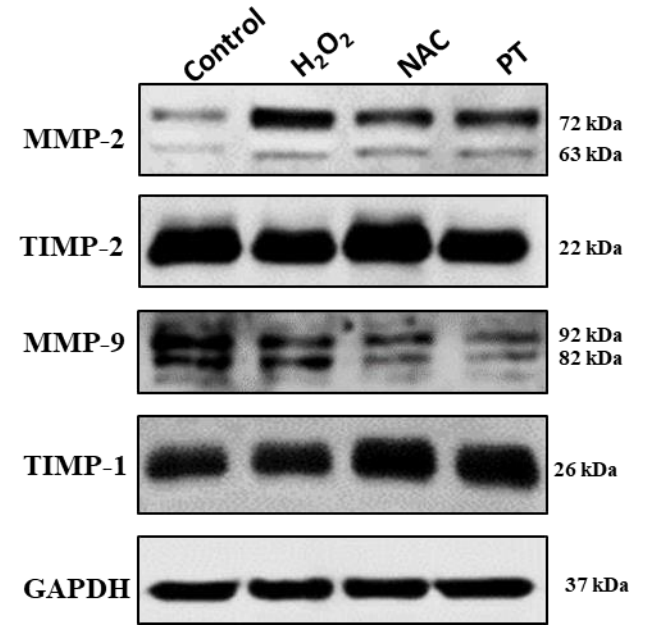
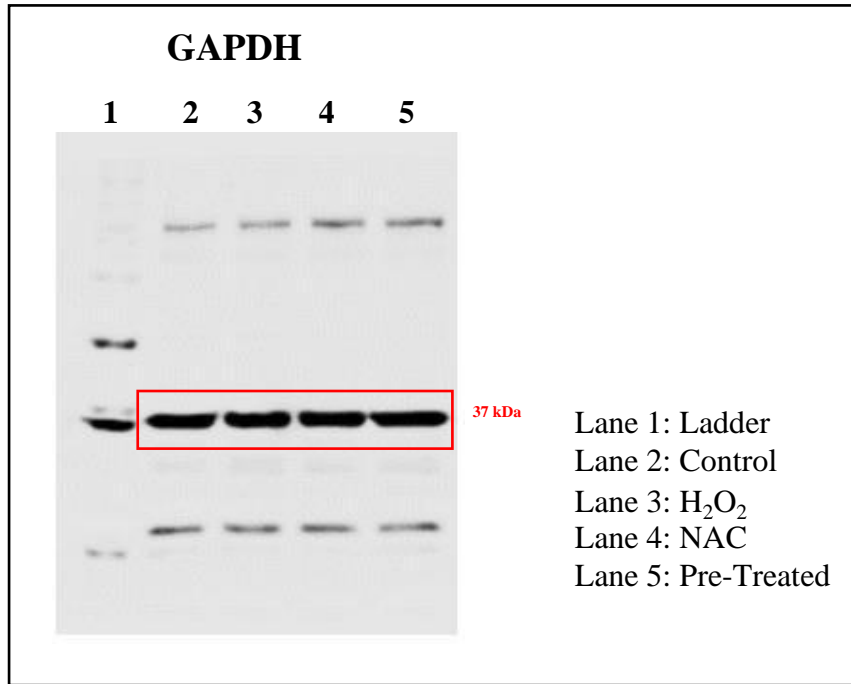
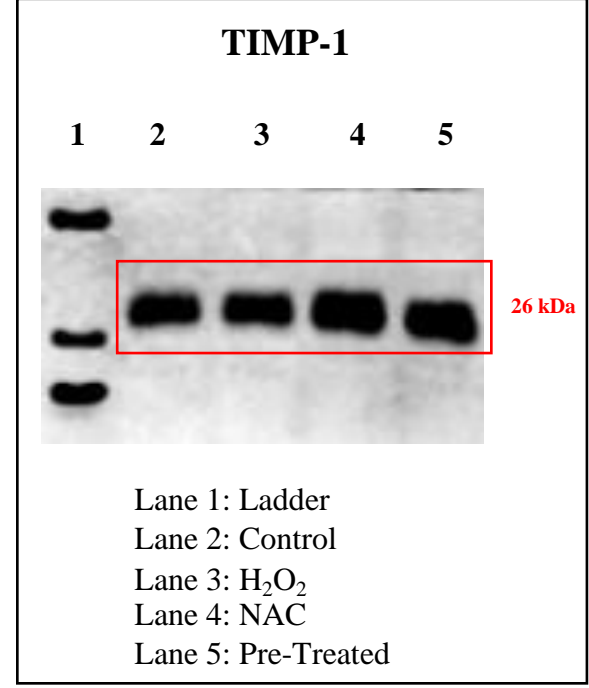
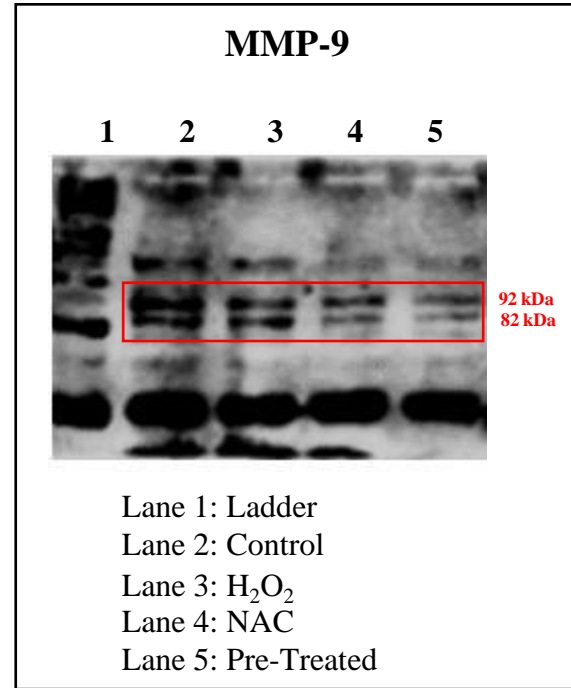
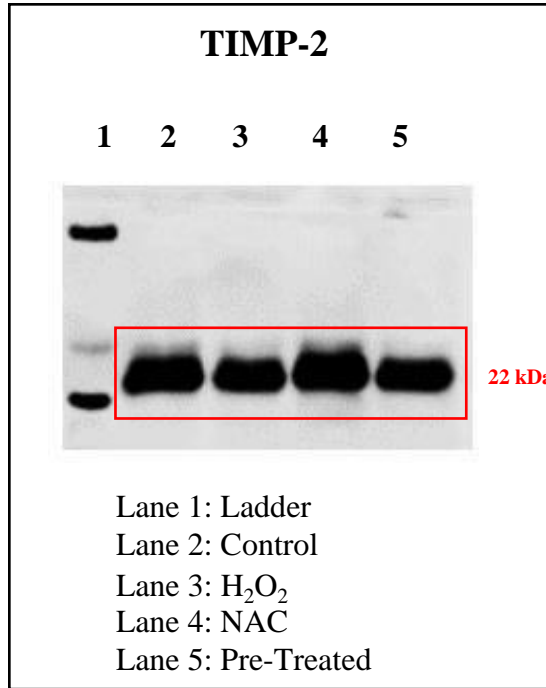
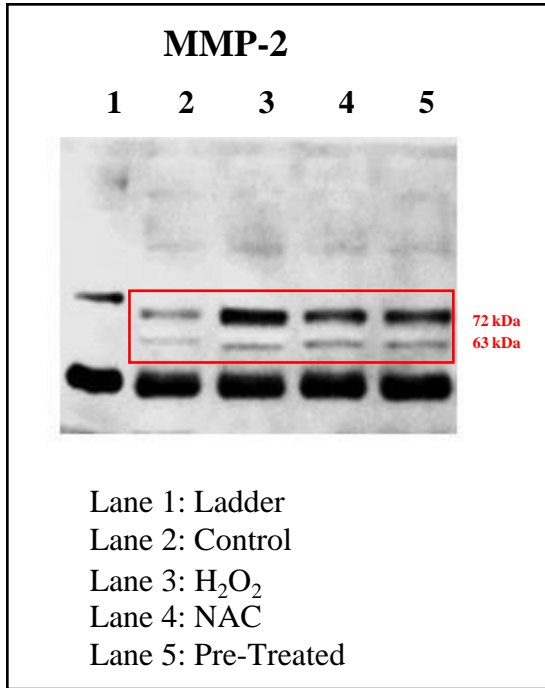
Exposure 1

Exposure 2

(Incorporated image)

**Figure 4a.**

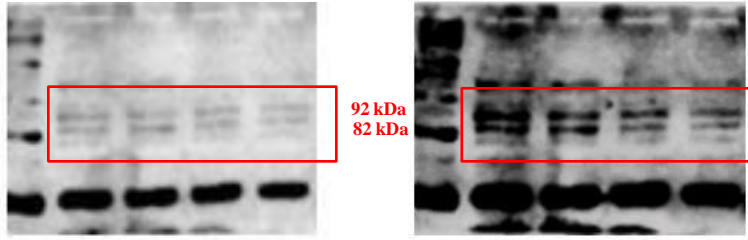




**Figure 7b.**



### MMP-9

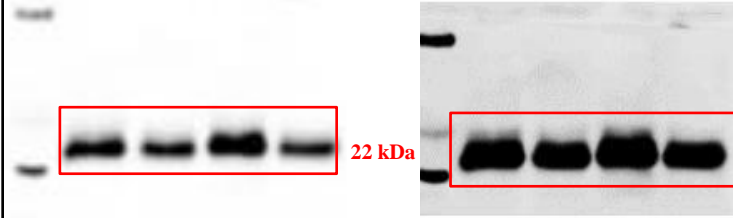


Exposure 1

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(Incorporated image)

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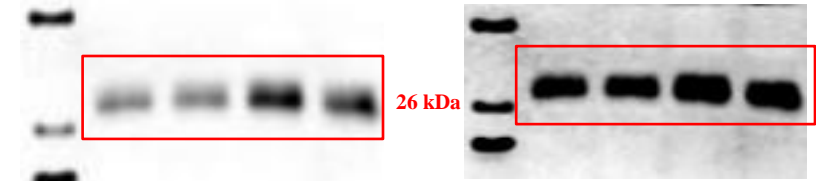


Exposure 1

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### TIMP-1

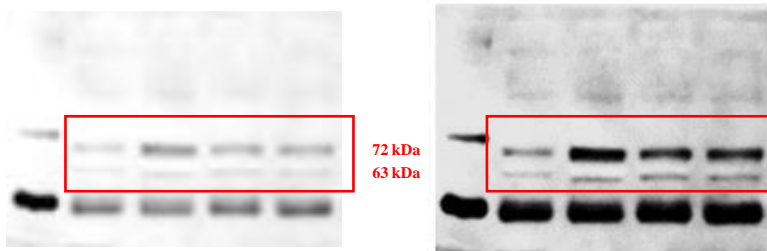


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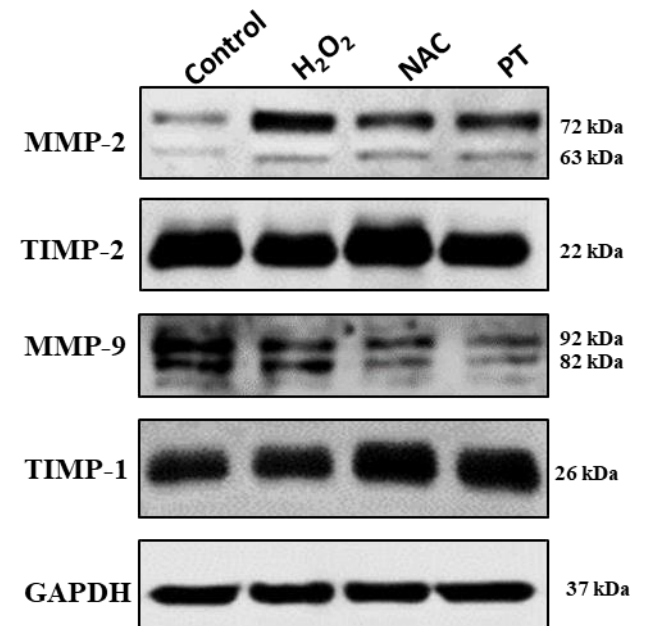
### MMP-2



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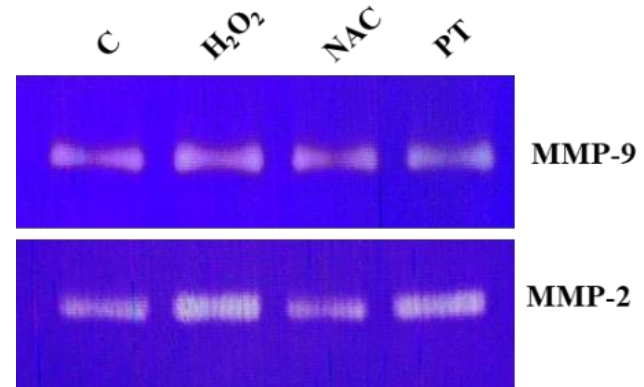
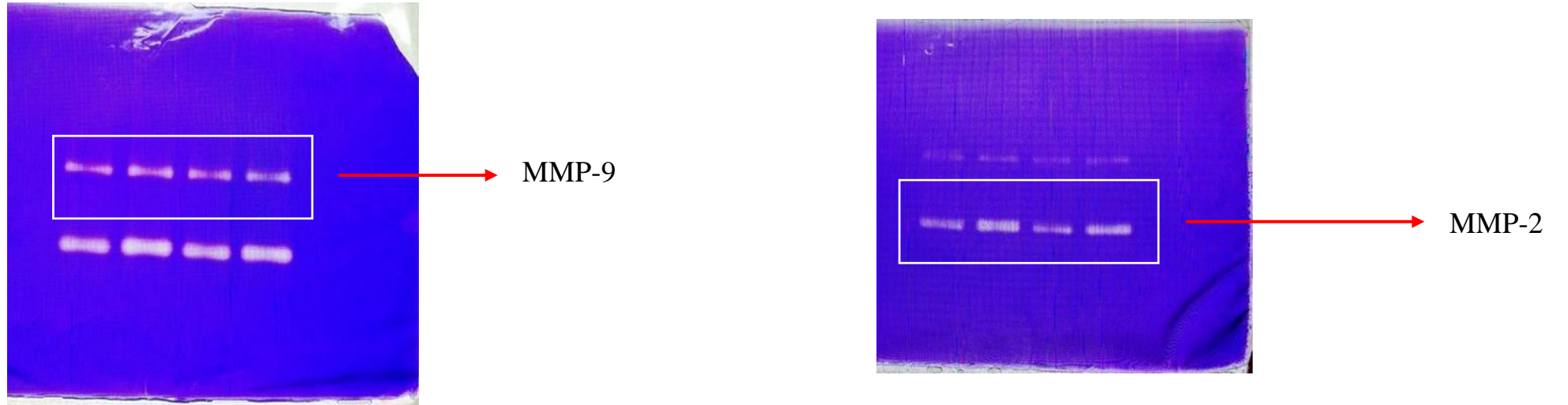
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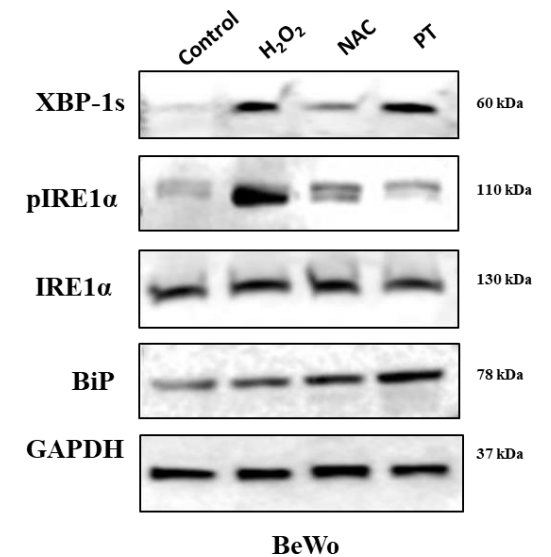
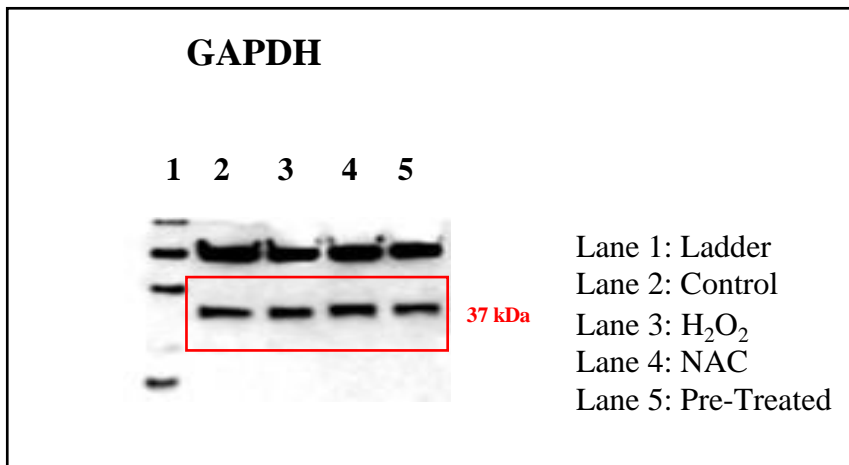
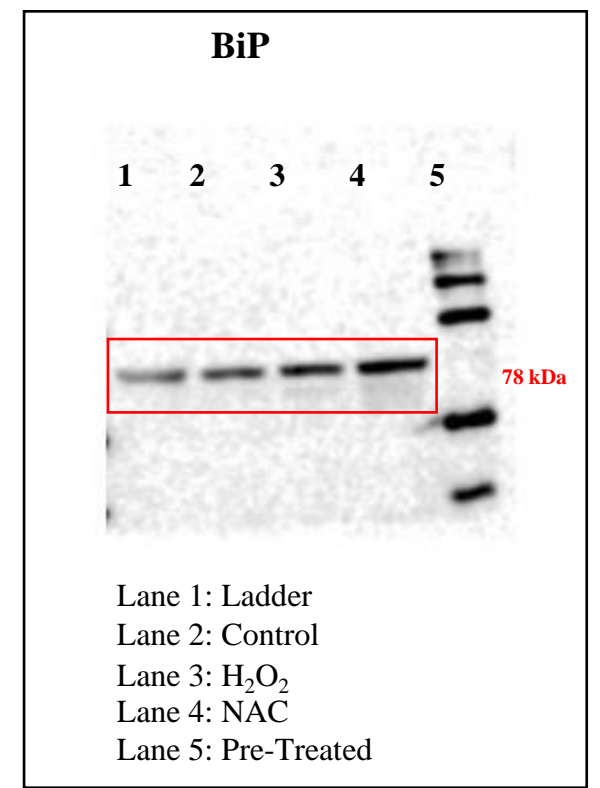
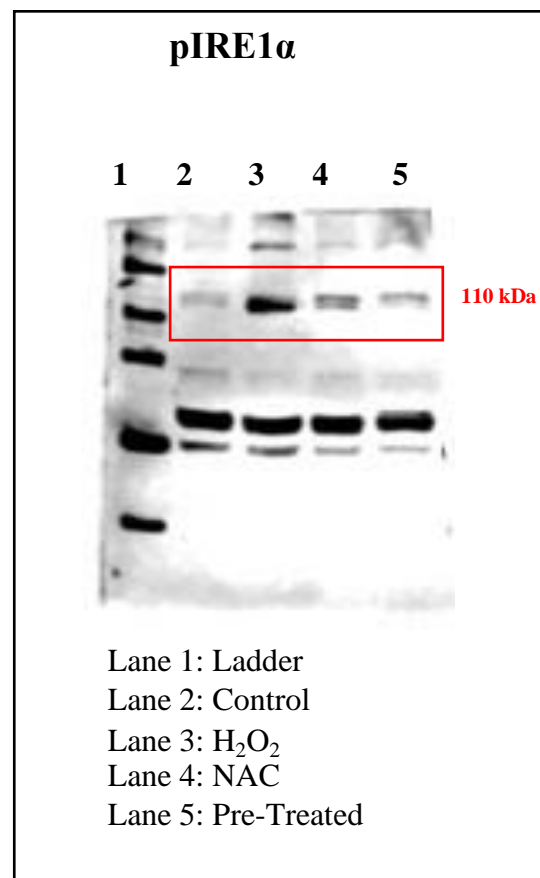
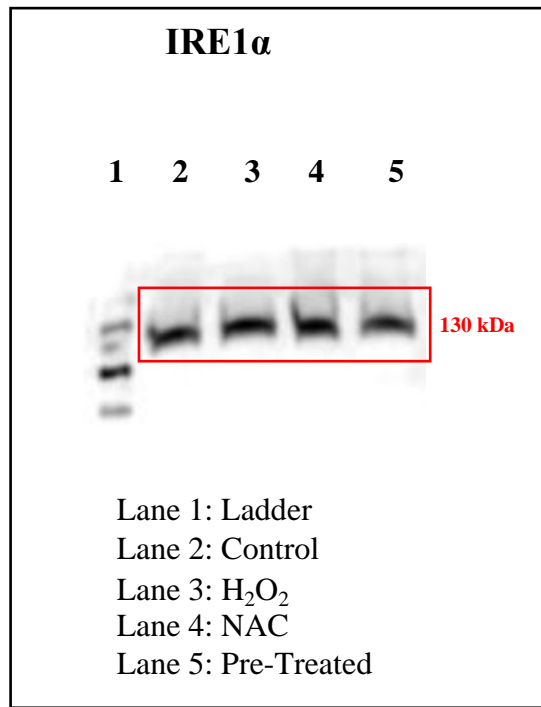
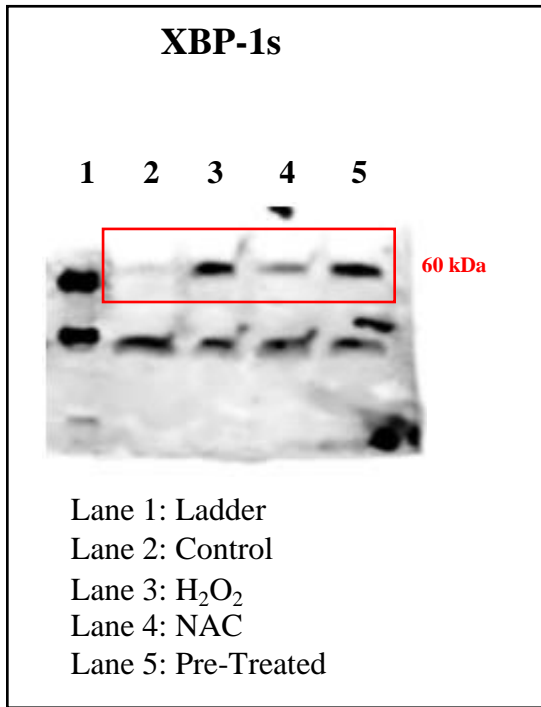
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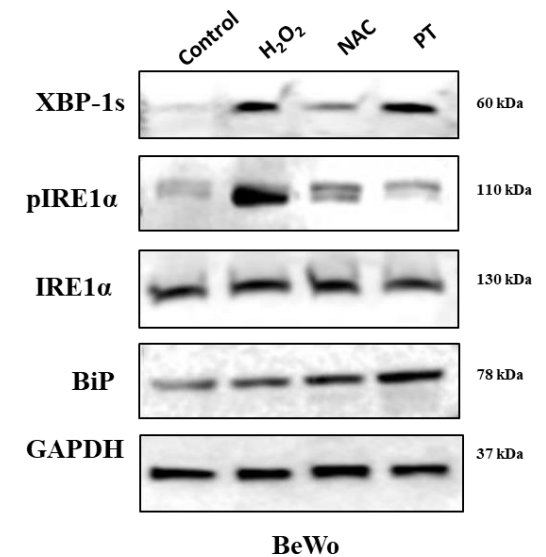
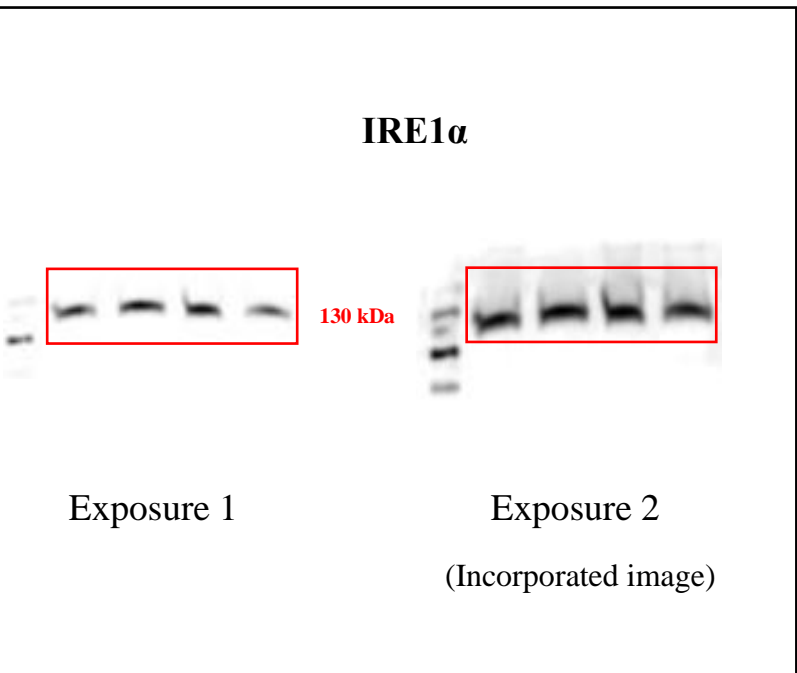
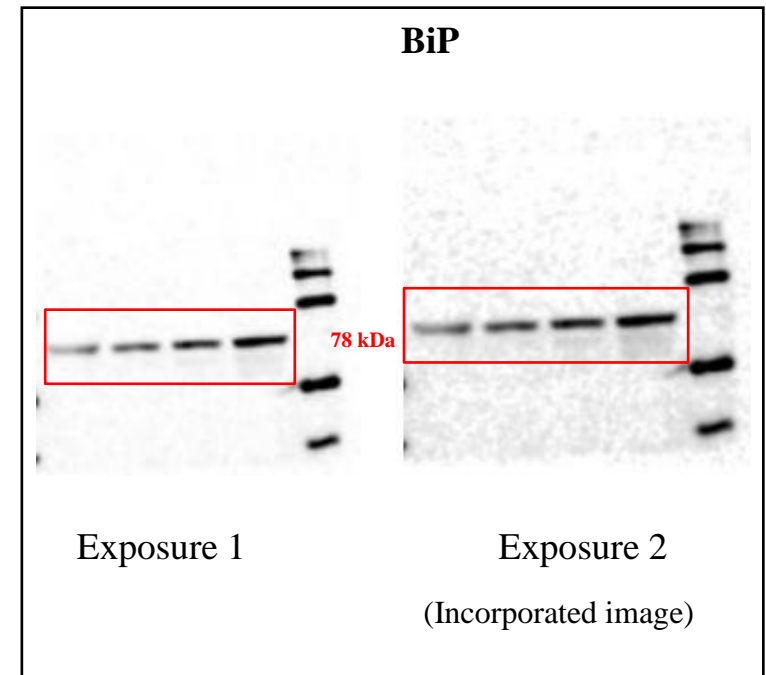
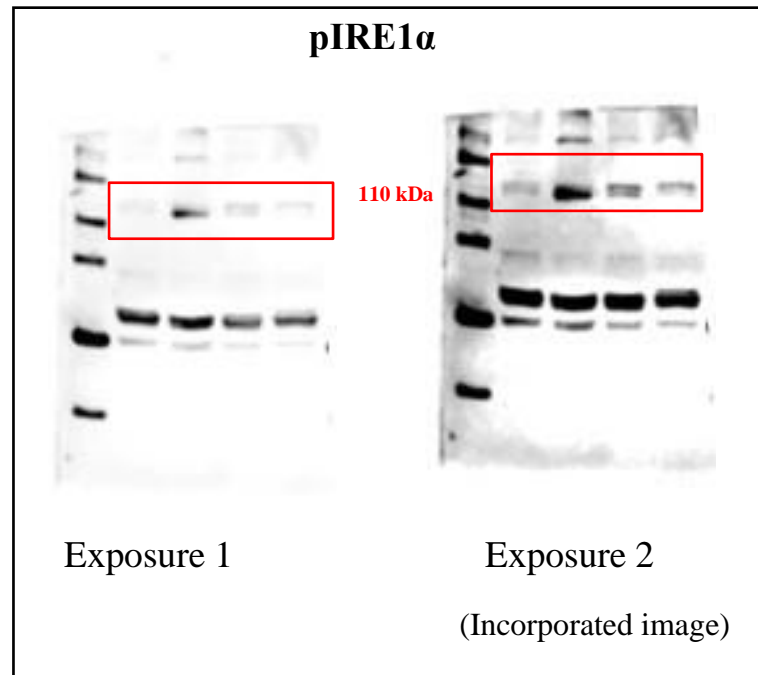
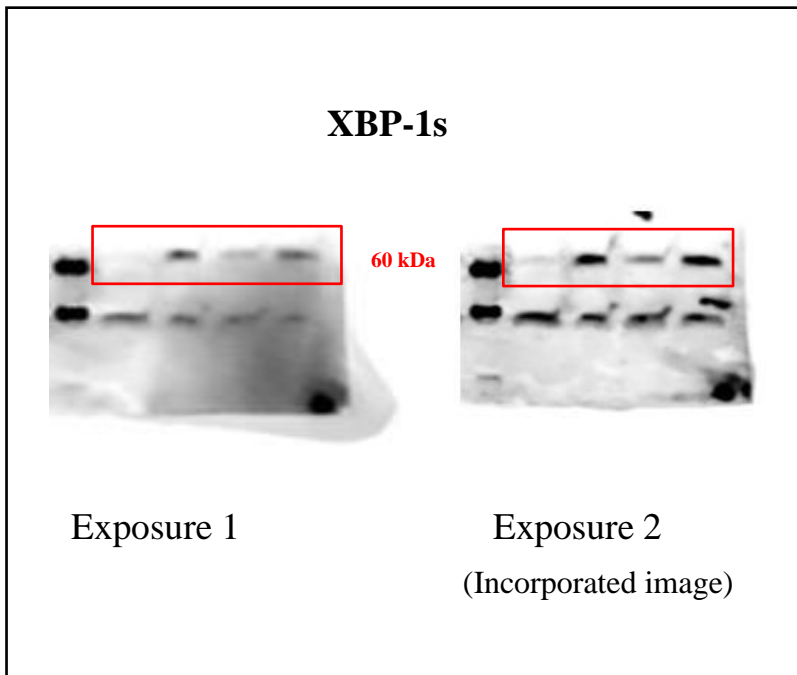
**Figure 7b.**

Figure. 7n

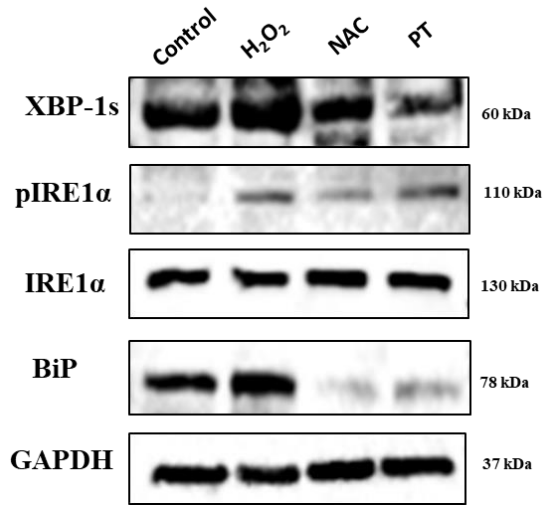
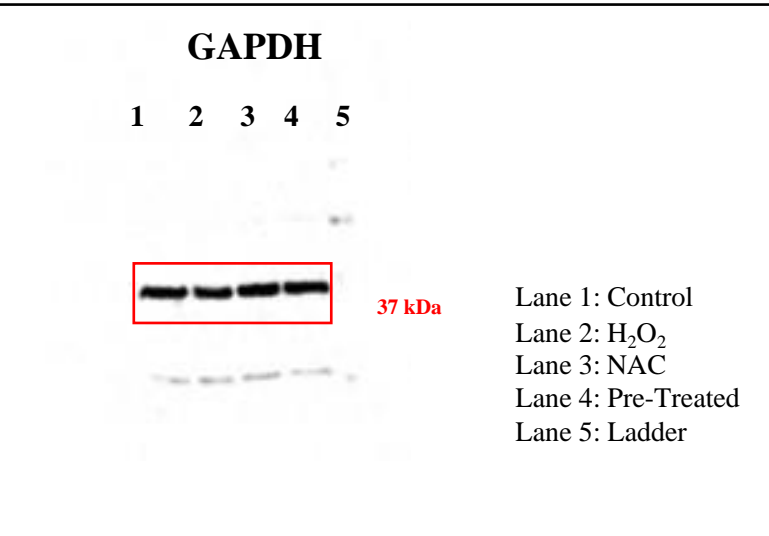
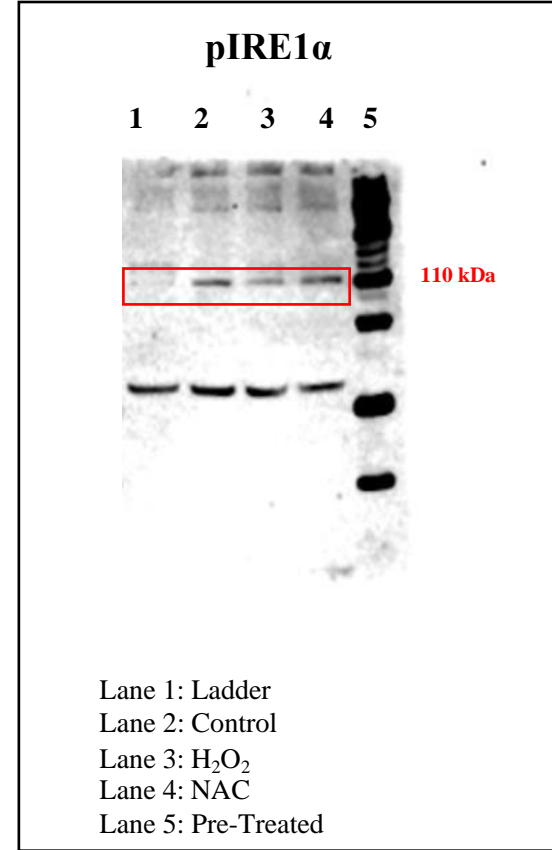
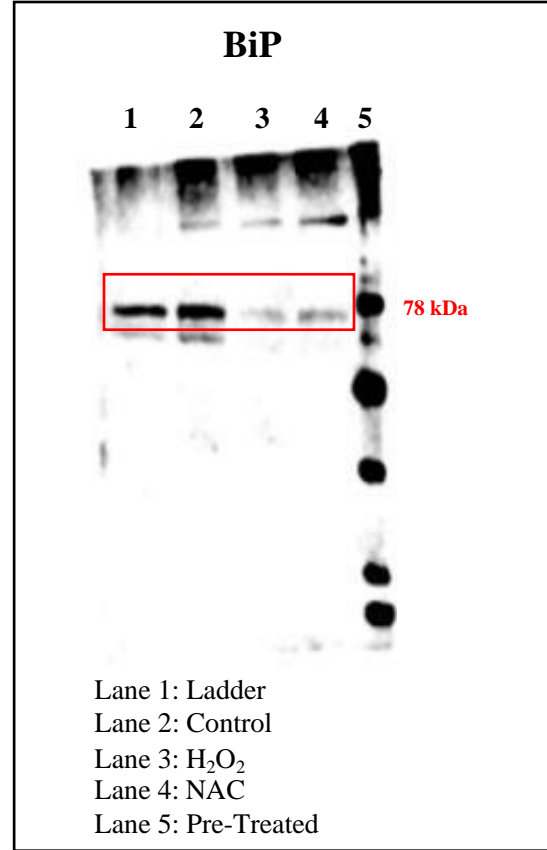
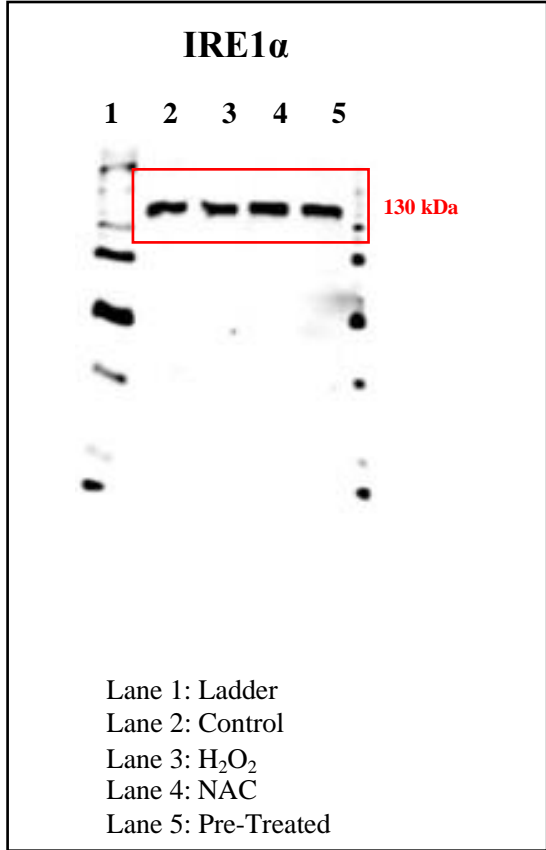
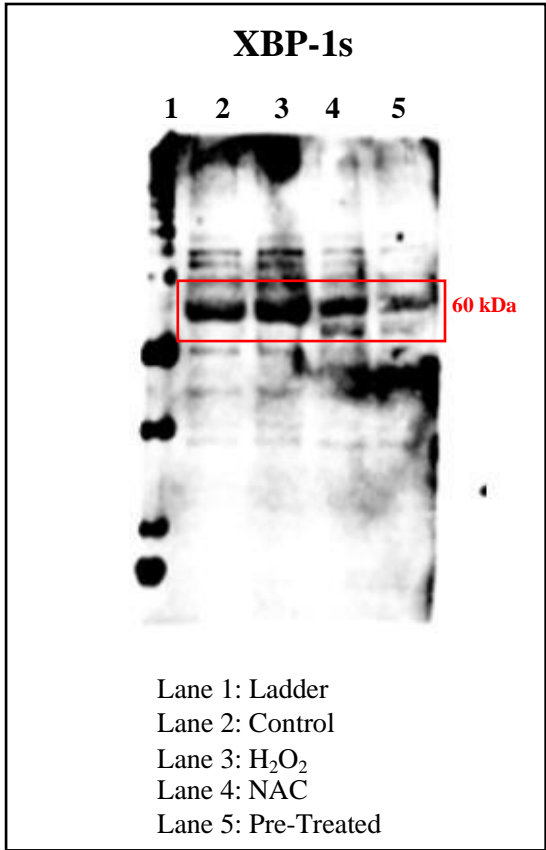




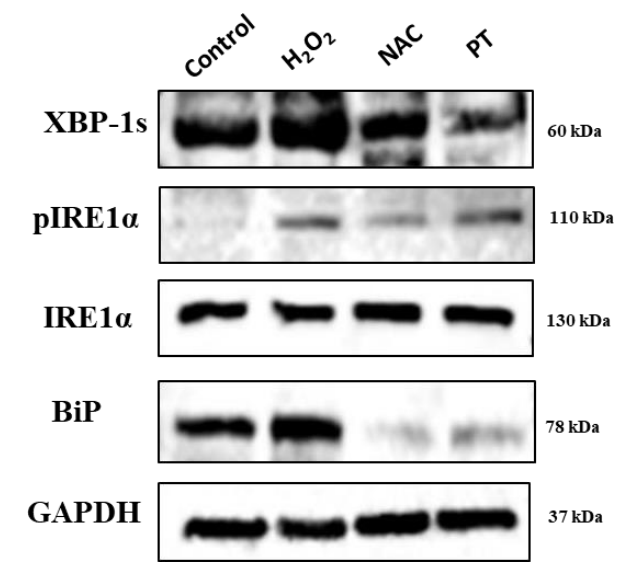
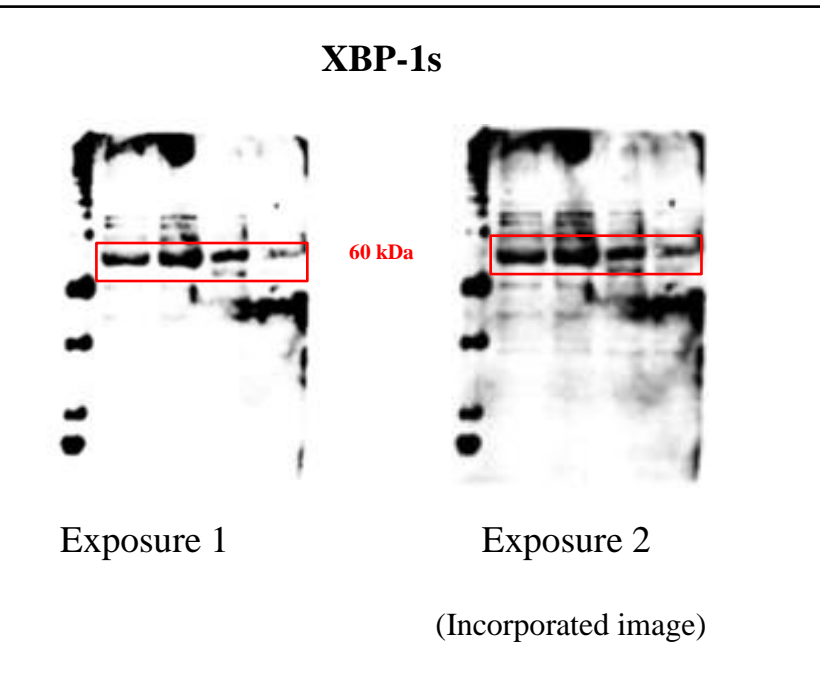
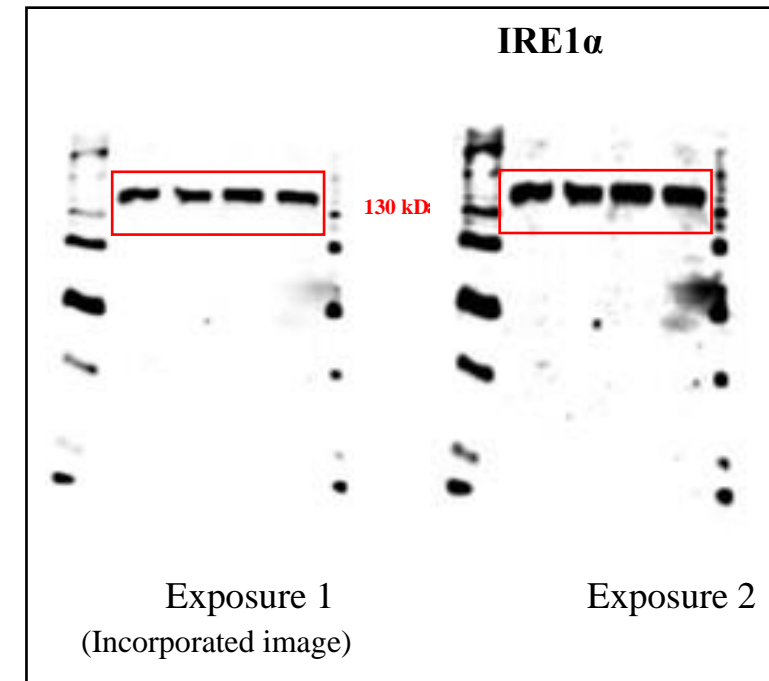
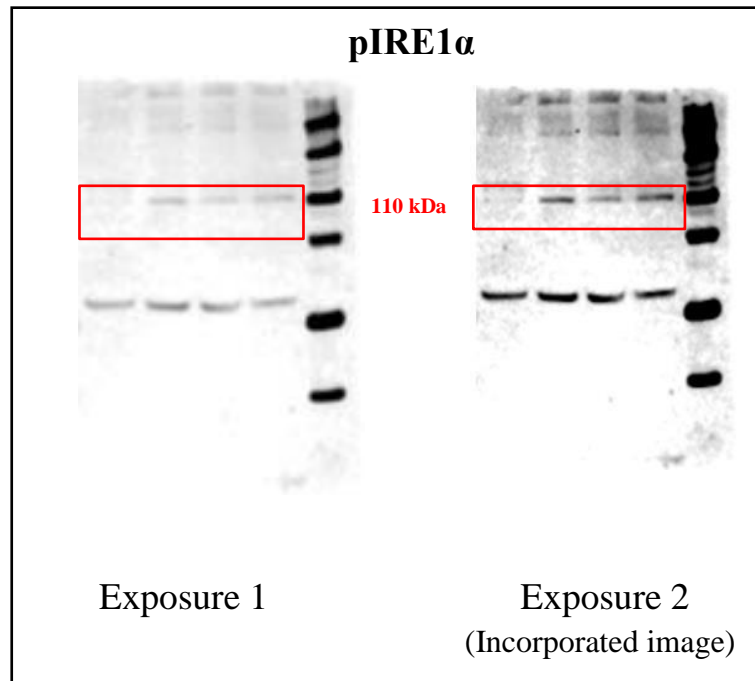
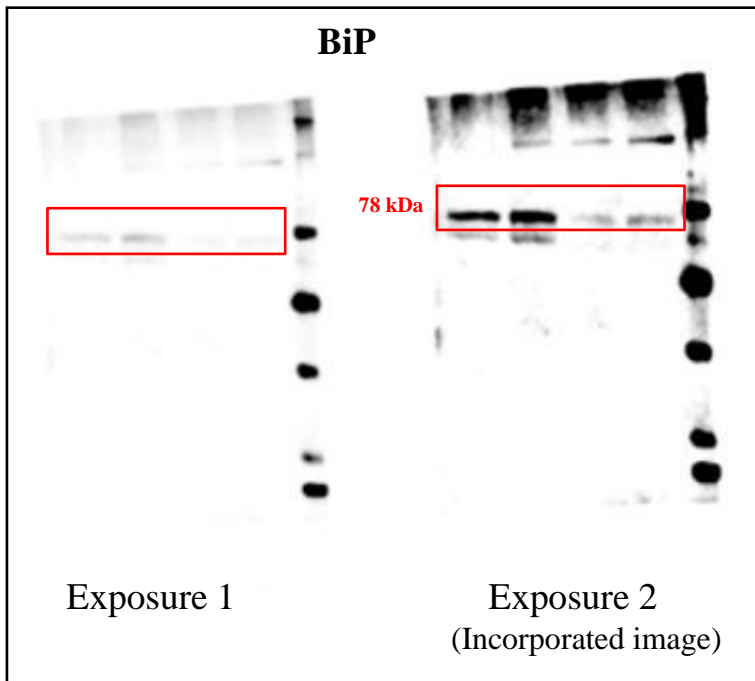
**Figure 9a.**



**Figure 9a.**

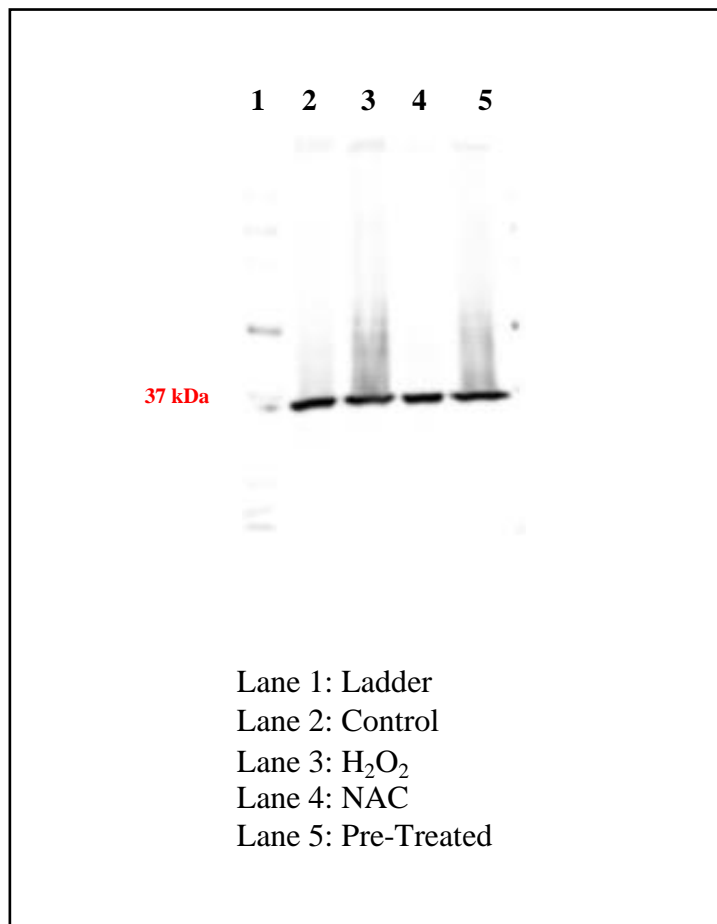


**Figure 9g.**

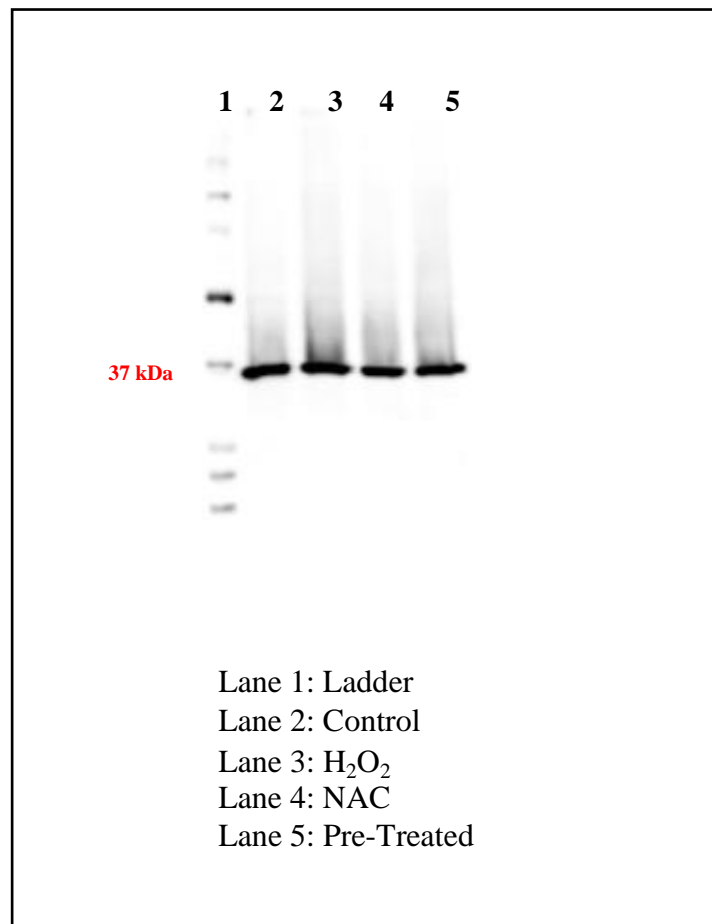


**Figure 9g.**

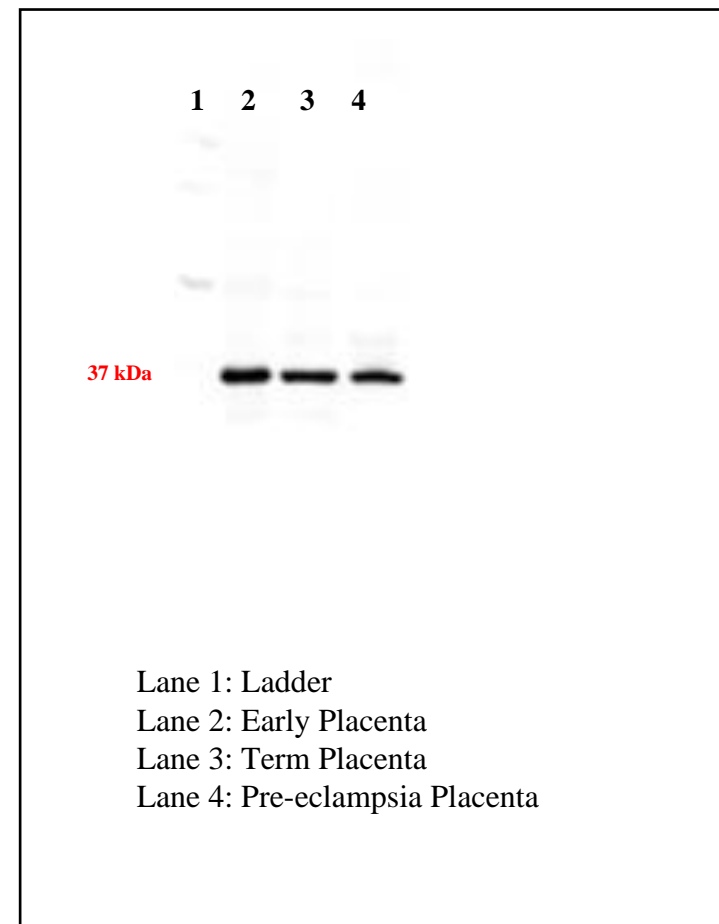
### GAPDH for pIRE1 $\alpha$



**BeWo**



**HTR-8/SV neo**



**Tissues**