

Fig. S1. Major immune populations in mouse dura mater. (A) Flow cytometry analysis of blood, BM and dura immune cells. Left panels, representative flow cytometry plots of NK cells (CD45⁺NK1.1⁺), B cells (CD45⁺CD19⁺), CD4 T cells (CD45⁺NK1.1⁻CD19⁻CD11b⁻CD3⁺CD4⁺), CD8 T cells (CD45⁺NK1.1⁻CD19⁻CD11b⁻CD3⁺CD8⁺), (CD45⁺NK1.1⁻CD19⁻CD11b⁺CD3⁻Ly6G⁺Ly6C^{int}), Ly6C⁺ Granulocytes monocytes (CD45+NK1.1-CD19-CD11b⁺CD3⁻Ly6G⁻Ly6C⁺), Ly6C⁻ monocytes (CD45⁺NK1.1⁻CD19⁻CD11b⁺CD3⁻Ly6G⁻Ly6C⁻CD64⁻) and Macrophages (CD45⁺NK1.1⁻CD19⁻CD11b⁺CD3⁻Lv6C⁻CD64⁺Cx3CR1⁺). Right panel, frequency of the immune populations identified in blood, BM and dura, out of total CD45⁺ (n=3 mice; data generated from a single experiment). (B) Flow cytometry staining for B cells in brain and dura. The dot-plot reports the absolute number of B cells (n=5 mice; data generated from two independent experiment). (C) Flow cytometry staining for B1a and B1b cells in dura and peritoneal cavity (PC). The dot-plot reports the percentage of B1a and B1b cells within the CD19⁺CD23⁻ population (n=3 mice; data generated from a single experiment). (D) Representative confocal images of cerebral cortex, choroid plexus, dentate gyrus and spinal cord in CD19-Tomato mice. B cells are not present in the CNS parenchyma (scale bar=50µm). (E) Representative confocal images of B cells in the leptomeninges, showing both extravascular B cells (white arrowhead) and intravascular B cells (yellow arrowhead) (scale bar=50μm).



Fig. S2. Meningeal B cell trafficking through the dura lymphatics. (A) Confocal imaging of dura mater (SS: sagittal sinus; TS: transverse sinus) under Tomato-lectin (blood vessels) and Lyve-1 (lymphatic vessels) staining (low magnification image scale bar=50µm; high magnification image scale bar=50µm). (B) Representative confocal image of dura mater stained for Lyve-1 and B220. B cells were localized both within (white arrowhead) and outside (blue arrowhead) the lymphatic vessels (scale bar=20µm).



Fig. S3. Imaging of immature B cells in dura mater. (A) Representative confocal image of GFP⁺ B cells in the sagittal sinus of *Rag1-GFP* mice (representative of n=5 mice; low magnification image scale bar=100 μ m; high magnification image scale bar=50 μ m). (B) Representative confocal images of dura B cells stained for immature B cell markers (CD93, Ki67, and GL7) and mature B cell marker (CD20) in CD19-tomato mice. White arrowheads indicate CD20⁻ B cells (each column displays a representative image from two different mice; scale bar=20 μ m).



Fig. S4. Phenotypic characterization of meningeal B cells. (A) Flow cytometry analysis of blood, bone marrow (BM), brain and dura B cells. B cells were gated on CD45⁺CD19⁺ and Lineage (CD3, CD11b, F4/80 and Gr-1) negative and were further divided into early (B220^{lo}CD43^{hi}), late (B220^{lo}CD43^{lo}) and mature (B220^{hi}CD43⁻) stages. Histograms show the staining intensity of mature B cell markers and developing B cell markers in the three subsets (data generated from a single experiment). (B) Flow cytometry analysis of IL-7R⁺ (CD127) B cells in BM and brain from C57BL/6 and Rag1^{-/-} mice (mean \pm SEM; n=4 mice per group; Mann Whitney *U* test **P*<0.05; data generated from a single experiment). (C) Flow cytometry analysis of IgD and IgM expression in B cells derived from blood, spleen, BM and dura (representative of n=3 mice; data generated from a single experiment).



Fig. S5. scRNA-seq analysis of dura cells. Top left, UMAP of total 12,351 cells from dura. Cells were colored by cell type and annotated with inferred cluster identities. Top right, frequency of cell types identified in dura. Bottom, expression of lineage specific genes in the corresponding cell types (n=3 mice; data generated from a single experiment).



Fig. S6. scRNA-seq analysis of BM cells. Top left, UMAP of total 36,452 cells from BM. Cells were colored by cell type and annotated with inferred cluster identities. Top right, frequency of cell types identified in BM. Bottom, expression of lineage specific genes in the corresponding cell types (n=3 mice; data generated from a single experiment).



Fig. S7. scRNA-seq analysis of blood cells. Top left, UMAP of total 42,370 cells from blood. Cells were colored by cell type and annotated with inferred cluster identities. Top right, frequency of cell types identified in blood. Bottom, expression of lineage specific genes in the corresponding cell types (n=3 mice; data generated from a single experiment).



Fig. S8. Transcriptional signature of the identified B cell clusters. (A) Differential gene expression analysis (cluster vs total; cutoff= $log_2FC>0.5$) and enriched biological processes in each B cell cluster (from C0 to C9). (B) Expression of marker genes differentially expressed during B cell development. (C) BrdU pulse-chase labeling of proliferating B cells in BM and dura. BrdU incorporation was assessed at three time-points (24hrs, 3 days and 5 days post-injection) in early (CD19⁺CD43^{hi}B220^{lo}), late (CD19⁺CD43^{lo}B220^{lo}) and mature (CD19⁺CD43⁻B220^{hi}) B cell. Dynamic distribution of BrdU⁺ B cells among the three subsets is shown in the bottom panel (n=3 mice per time-point; data generated from two independent experiments).



Fig. S9. High-throughput characterization of immune cells from blood, BM, and dura based on CyTOF analysis. Upper panels: major immune populations in blood, bone marrow, and dura displayed by tSNE. Bottom panels: heatmap showing the surface marker repertoire for each individual population (n=3 mice; data generated from a single experiment).



Fig. S10. The hematopoietic niche in the calvaria. (A) Image of the region enriched of BM cells in the mouse skull (red box). (B) Brightfield image under toluidine blue staining of the calvarial bone marrow. (C) FACS analysis of the calvarial CD45⁺ BM cells. Both IL7R⁺ B cells and Lin⁻Sca1⁺Kit⁺ stem cells were found. (D) EM micrographs of the calvarial BM, showing both myeloid cells (bona-fide macrophages, granulocytes, mast cells and erythrocytes) and lymphoid cells (bona-fide lymphocytes). (E) Confocal images of cranial sagittal section from a CD19-Tomato mouse stained with Tomato-lectin. Whereas no B cells are visible in the brain parenchyma, abundant B cells are present within the calvarial BM.



Fig. S11. Meningeal B cells derive from the calvaria. (**A**) Flow cytometry analysis of donor-derived (CD45.2⁺) and host-derived (CD45.1⁺) B cells and CD4 T cells in different compartments of head-reconstituted mice (representative of two mice; data generated from a single experiment). (**B**) Four weeks parabiosis between wild-type C57BL/6 and CD19-Tomato mice. Flow cytometry analysis of the wild type parabiont shows the percentage of B cells chimerism in blood, spleen, brain, and dura. (**C**) Frequency of parabiont-derived B cells in the indicated tissues (mean \pm SEM; n=5 mice; one-way ANOVA and Bonferroni post-hoc test ***P*<0.01, ****P*<0.001; data generated from three independent experiments). (**D**) Adoptive transfer by iv. injection of 20x10⁶ CD19-Tomato splenocytes into wild-type C57BL/6 mouse. Flow cytometry analysis one week post-injection shows the percentage of donor-derived B cells in blood, spleen, brain, and dura. (**E**) Frequency of donor-derived B cells in the indicated tissues (mean \pm SEM; n=3 mice; one-way ANOVA and Bonferroni post-hoc test ***P*<0.01, ****P*<0.001; data representative of two independent experiments).



Fig. S12. B cells trafficking through the skull vascular channel. (A) Left, schematic of sample preparation for skull cryosection and X-ray tomography. Right, representative confocal images of skull vascular channels (scale bar= 50μ m). (B) X-ray tomography images displayed as *z*-series showing a vascular channel opening onto the sinus (white arrowhead). (C) Representative confocal images of IgM⁻ B cells trafficking through the skull vascular channels (scale bar= 20μ m). (D) Orthogonal projection of a B cell within a vascular channel (red arrowhead).



Fig. S13. *Cxcl12* expression in the sagittal sinus. Representative confocal images of dura sagittal sinus from a WT and *a Cxcl12-DsRed* reporter mouse under Lyve-1 staining. High magnification images of Cxcl12-expressing fibroblasts (scale bar=100 μ m) and IgM⁺ B cells are shown on the right (scale bar=10 μ m).



Fig. S14. Meningeal B cells during aging. (A) Flow cytometry analysis of dura B cells in young and aged mice (mean \pm SEM; n=13 mice; Unpaired Student's *t* test ****P*<0.001; data generated from three independent experiments). (**B**) Confocal imaging of B cells in the sagittal sinus of young and aged mice. (**C**) Frequency of B cells clones ranked by size. Each line represents an individual mouse. (**D**) Fraction of B cell clones shared between blood and dura in young and aged mice. (**E**) Flow cytometry intracellular staining of ApoE in dura ABC population (data generated from a single experiment). (**F**) Principal component analysis of V-region diversity in dura ABCs and dura mature B cells from young and aged mice. (**G**) Frequency of somatic mutations in V-region from dura mature B cells and dura ABCs expressing *Ighm* (violin plot, Mann Whitney *U* test ****P*<0.001).



Fig. S15. IgM plasma cells accumulate in the aged dura. (A) Plasma cells cluster in dura enriched for signature genes (*IgJ* and *Xbp1*). (B) Representative images of dura plasma cells (IgG⁺MHC-II⁻B220⁻) (scale bar=10µm). (C) Differential expression levels of *Igha* and *Ighm* in dura plasma cells from young and aged mice. (D) Dura plasma cells were imaged and quantified in young and aged mice. The cartoon indicates the approximate position of three regions of interest ($y,x,z = 1000\mu m \times 350\mu m \times 10\mu m$) used for imaging. Plasma cells number per mm of sagittal sinus was determined by averaging the absolute count in the three regions of interest in each mouse (scale bar=50µm). (E) Number of total plasma cells per mm of sagittal sinus in young and aged mice (mean ± SEM; n=8 mice; unpaired Student's *t* test ****P*<0.001; data generated from two independent experiments). (F) Number of IgA, IgG and IgM plasma cells per mm of sagittal sinus in young and aged mice (mean ± SEM; n=4-8 mice per group; two-ways ANOVA and Bonferroni post-hoc test ***P*<0.01, ****P*<0.001; data generated from two independent experiments).



	Fig. S16. Dura B cells are equipped with functional BCRs, and MOG-specific B cells are reduced in dura
	compared to tibial BM. (A) Flow cytometry analysis of B cells from Nur77 ^{GFP} mice in multiple compartments: dura,
	tibial BM, skull BM, blood, spleen, and cervical lymph nodes (cLN). B cells were gated as: IgM/IgD double negative
	(DN), IgM ⁺ , IgM ⁺ IgD ¹⁰ (transitional), and IgD ⁺ . Frequency of these population and percentage of GFP ⁺ cells was
5	determined in all compartments. (B) Percentage of different B cell populations from all analyzed tissues (mean ±
	SEM; n=7-11 mice per group; two-ways ANOVA and Bonferroni post-hoc test ***P<0.001; data generated from two
	independent experiments). (C) Percentage of GFP ⁺ B cell in different populations from all analyzed tissues (mean ±
	SEM; n=6-9 mice per group; two-ways ANOVA and Bonferroni post-hoc test **P<0.01, ***P<0.001; data generated
	from two independent experiments). (D) Flow cytometry analysis of B cells from <i>IGH</i> ^{MOG} mice in dura and tibial BM.
10	B cells were gated as: MHC-II ⁺ IgM ⁺ and MHC-II ⁻ IgM ⁻ . Percentage of MOG-binding B cells was determined each
	population. Wild-type mice served as negative control. (E) Quantification of MOG-specific B cells in dura and tibial
	BM MHC-II ⁺ IgM ⁺ B cells (mean \pm SEM; n=7 mice; unpaired Student's t test **P<0.01; data generated from two
	independent experiments). (F) Quantification of MOG-specific B cells in dura and tibial BM MHC-II-IgM ⁻ B cells
	(mean \pm SEM; n=7 mice; unpaired Student's <i>t</i> test; data generated from two independent experiments).
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Movie S1

Two photon in vivo imaging of meningeal B cells.

Movie S2

20 X-ray tomography of the intact mouse skull.

Table S1

Transcriptional signature for each B cell cluster identified by scRNA-seq.

25 **Table S2**

Surface markers expression for each B cell cluster identified by CyTOF.

Table S3

Transcriptional signature of dura ABCs identified by scRNA-seq.

Table S4

Complete antibody list used for flow cytometry, immunofluorescence and CyTOF staining.

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