

Supplementary Information, Furman et al., Handcuffing intrinsically disordered regions in Mlh1-Pms1 disrupts mismatch repair

Supplementary Figure S1. Functional domains of yeast Mlh1 and Pms1.

(A) Cartoon representations of the Mlh1 and Pms1 subunits, with the N-terminal ATP binding and C-terminal endonuclease/MLH interaction domains separated by IDRs. Amino acid locations of the domains in yeast Mlh1 and Pms1 are shown. IDR deletions that do not disrupt or only weakly disrupt Mlh1 or Pms1 functions (black lines) or confer *mlh1* or *pms1* null phenotypes (red lines), are shown (Plys *et al*, 2012). The green triangles indicate insertion sites for FRB and FKBP into the Mlh1 and Pms1 IDRs. FRB and FKBP were inserted immediately after the indicated amino acid position of Mlh1 and Pms1. (B) SDS-PAGE analysis (8% gel stained with Coomassie blue R250) of wild-type, Complex #2 (mlh1-FKBP₄₆₄, pms1-FRB₆₂₇), Complex #3 (mlh1-FRB₄₆₄, pms1-FKBP₄₆₀), Complex #4 (mlh1-FKBP₄₆₄, pms1-FRB₄₆₀), and Complex #5 (mlh1-FRB₃₅₅-pms1-FKBP₄₆₀). Complex #1 (mlh1-FRB₃₅₅, FKBP₄₆₄, PMS1) is shown in the right panel with a molecular weight standard and wild-type included for reference. Note the presence of higher molecular weight Mlh1-Pms1 complexes seen in this panel as described previously (Hall et al. 2001).

Supplementary Figure S2. Analysis of Complex #2 (mlh1-FKBP₄₆₄, pms1-FRB₆₂₇) in DNA binding, ATP binding, and endonuclease assays.

(A) MST analysis of Complex #2 in the presence and absence of 49 bp homoduplex DNA (20 nM), ATP (1 mM), and rapamycin (1 μ M). Three independent experiments were performed (error bars indicate the mean \pm standard deviation) using at least two independently purified batches of each protein. F_{norm} was calculated by dividing F_{hot} (average fluorescence value in the heated state) by F_{cold} (average fluorescence value measured in the cold state before the infrared laser is turned on) and plotted as parts per thousand (%). See the Materials and Methods for details. (B) ATP hydrolysis activity of Complex #2; (0.40 μ M) was determined in the presence

and absence of PCNA (0.250 μ M), 49-bp homoduplex DNA (0.75 μ M), and rapamycin (1 μ M). Error bars indicate \pm one standard deviation for three replicates. (C) Endonuclease activity of Complex #2 (50 nM) was determined on a closed circular DNA substrate (cc) in the presence (+) or absence (-) of MnSO₄, ATP, rapamycin, and yeast PCNA/RFC (Materials and Methods). MnSO₄, ATP, rapamycin, RFC and PCNA were included at 5 mM, 0.5 mM, 1 μ M, 125 nM and 250 nM, respectively. n= nicked product. (D) Endonuclease activities of Complex #2 were determined at the indicated concentrations. Assays were performed in the presence of MnSO₄, ATP, RFC, PCNA. Rapamycin was included as indicated. Error-bars indicate the standard deviation of three replicates (Supplementary Figure S3). In panels B, C, and D, the analysis of Mlh1-Pms1 presented in Fig 3 is shown again for comparison purposes.

Supplementary Figure S3. Gel images of protein titration endonuclease activity assays for wild-type, Complex #2 and Complex #5. The assays shown in this figure were used to make the graphs presented in Figure 3D and Supplementary Figure S2D. Endonuclease assays (Materials and Methods) were performed on closed circular plasmid DNA in the presence of MnSO₄, ATP, RFC, PCNA without (A) or with (B) rapamycin. cc = closed circular, n= nicked, l = linear.

Supplementary Figure S4. Gel images of protein titration endonuclease activity assays for wild-type and Complex #1. (A) Additional replicates of the assay shown in Figure 4C. Note that Assays 1 and 2 were performed with pUC18 DNA substrate and 3 and 4 with pUC19 DNA substrate. The "1/2" notation refers to conditions in which Mlh1-Pms1 and Complex #1 were present at 25 nM. (B) Individual titrations used to make the graph presented in Figure 4D. Endonuclease assays (Materials and Methods) were performed on closed circular plasmid DNA in the presence of Mlh1-Pms1, Complex #1, RFC, MnSO₄, ATP at the concentrations presented in Figure 4C, and increasing concentrations of PCNA (12.5, 25, 50, 100, 200 nM) in the absence of rapamycin.

Supplementary Table S1. Strains used in this study.

Strains used in this study were derived from the S288c background.

Supplementary Table S2. Plasmids used in this study.

Full plasmid descriptions can be found in the Materials and Methods. *mlh1-FRB_{XXX}*, *mlh1-FKBP_{XXX}*, *pms1-FRB_{XXX}*, and *pms1-FKBP_{XXX}* refer to the amino acids (XXX) in Mlh1 or Pms1 immediately after which the FRB or FKBP domains were inserted (Figure 1; Materials and Methods).

Supplementary Table S3. XL-MS lysine crosslinks for Mlh1-Pms1 (A), Complex 5 (B) and summary of intermolecular crosslinks for Mlh1-Pms1 and Complex #5 (C)

Lower case k indicates site of cross-link

Uniprot: <https://www.uniprot.org>

Score: MaXLinker score (Yugandhar et al. 2020)

#CSMs: Number of cross-link spectrum matches

XL MH+: Deconvoluted mass of the cross-link. It is a standard practice to present the deconvoluted mass along with the mass of an additional H+

Z: Charge of the cross-link

XL m/z: Cross-link mass divided by its charge

Supplementary Table S4. Phenotypic analysis of *mlh1* and *pms1* alleles that map within or near Mlh1-Pms1 crosslinking sites as determined by XL-MS

+ indicates phenotype similar to wild-type, - indicates similar to MMR null, +/- intermediate phenotype.

REFERENCES

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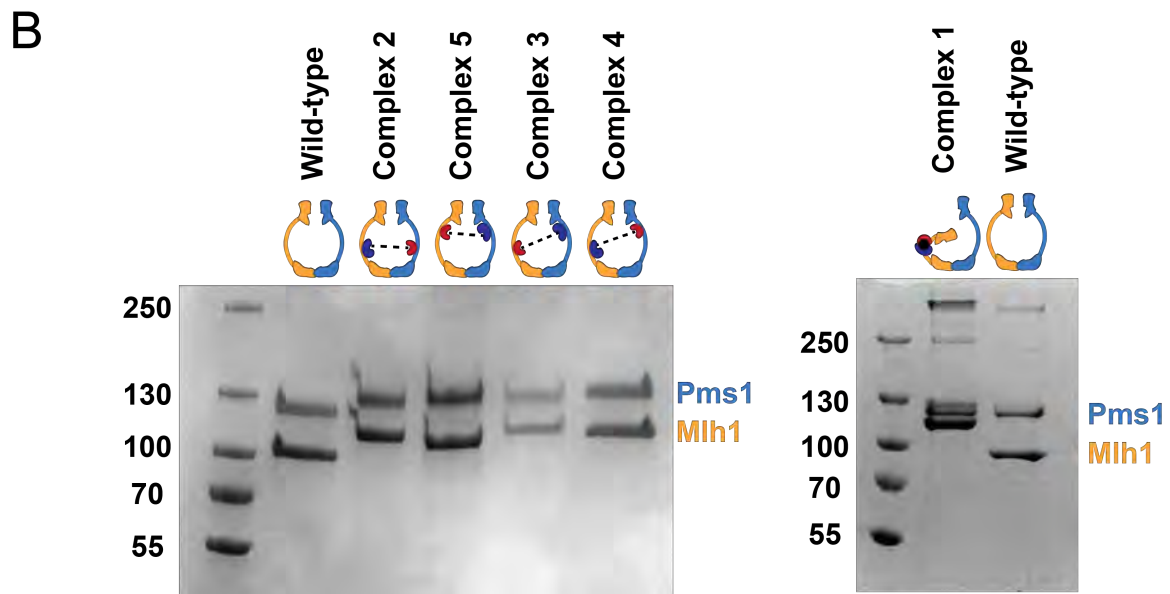
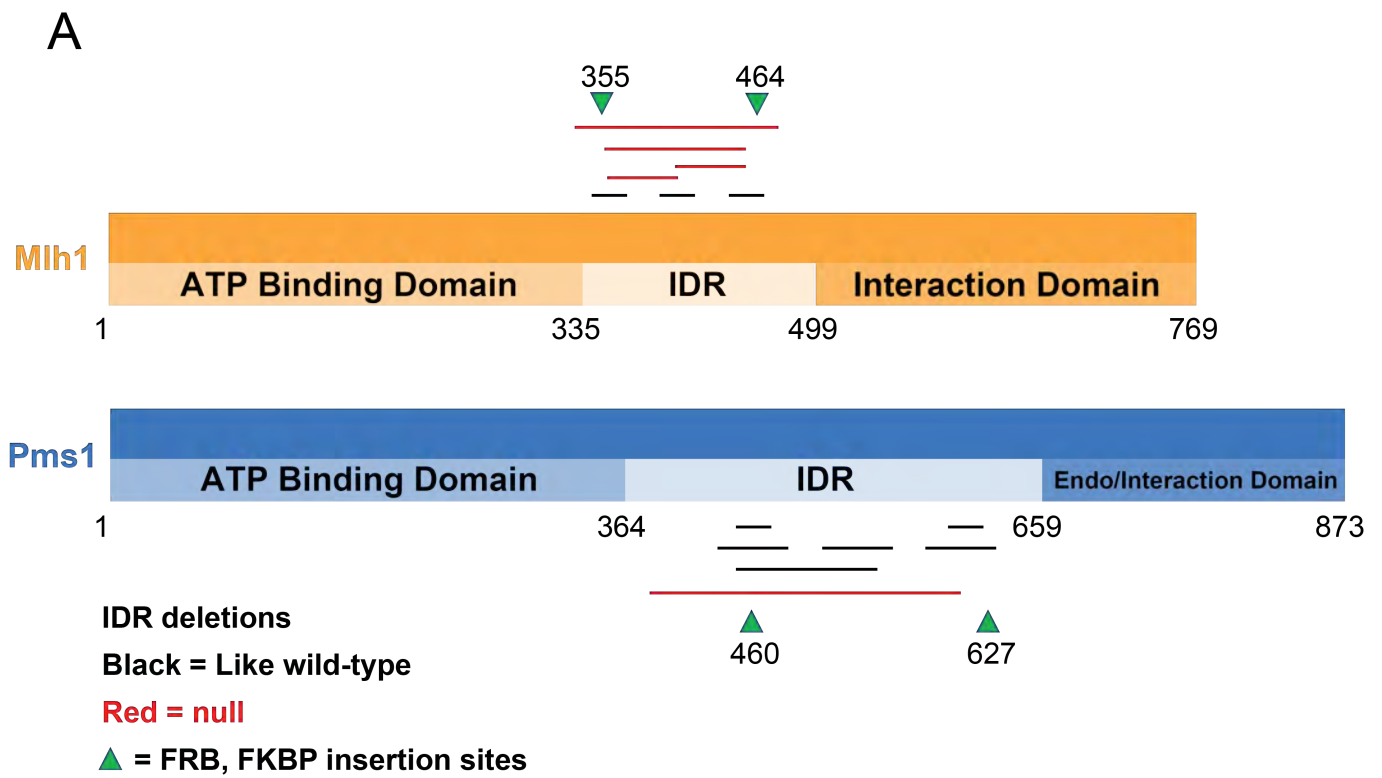
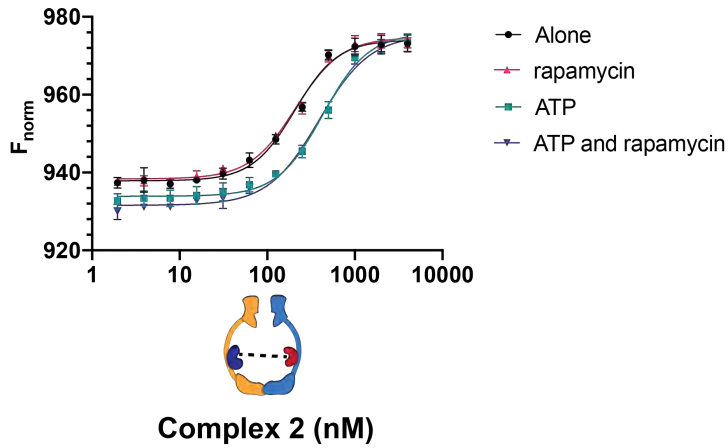
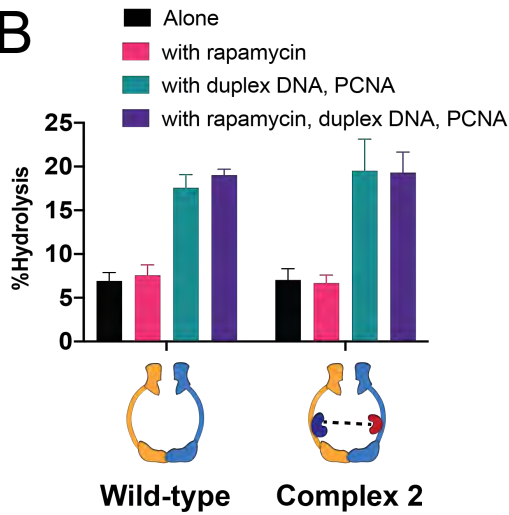


Figure S1

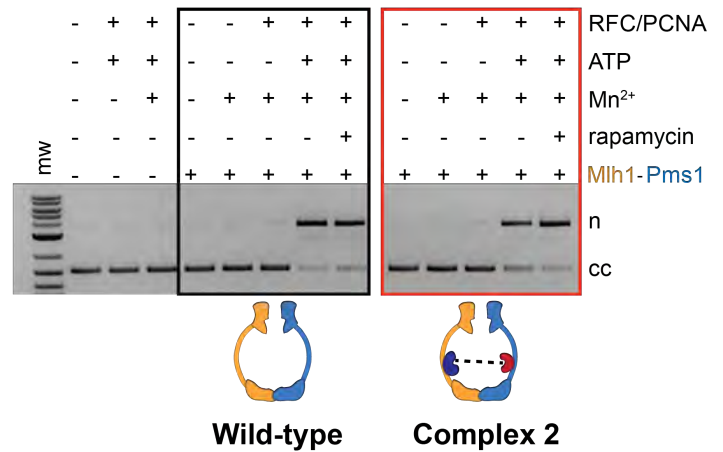
A



B



C



D

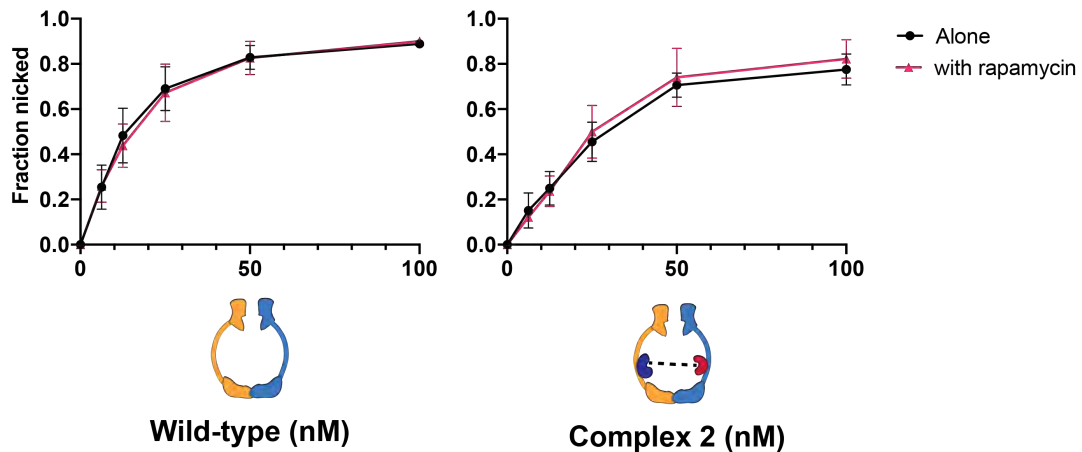
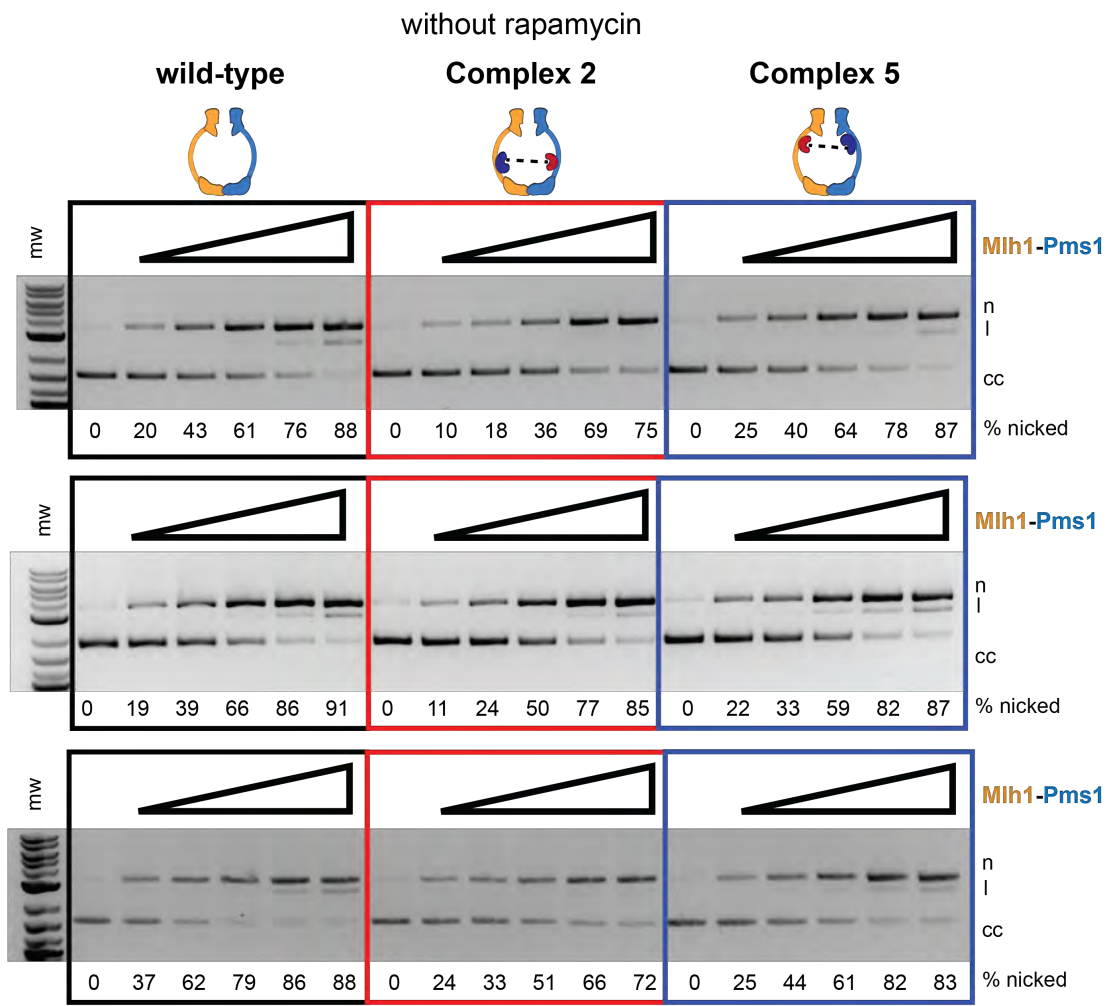


Figure S2

A



B

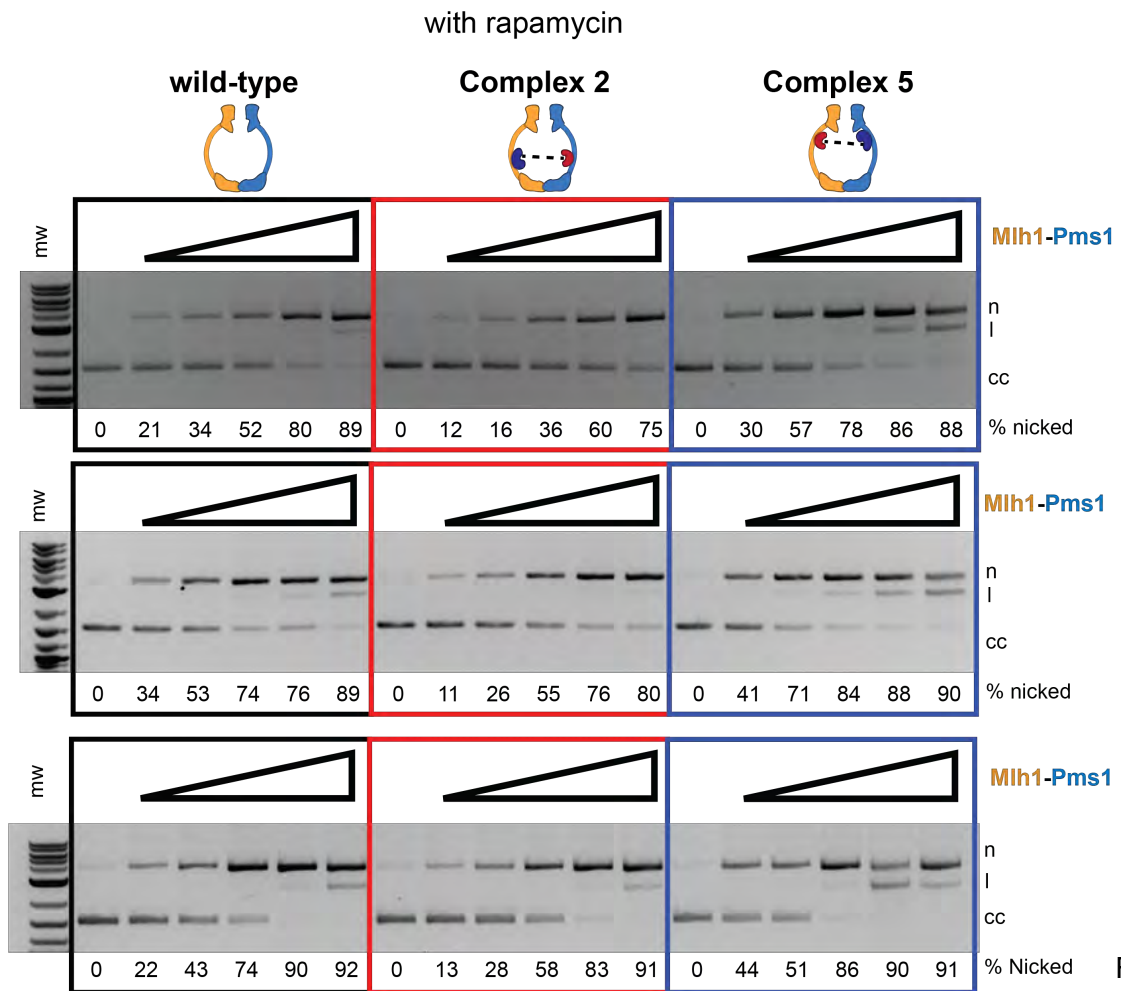
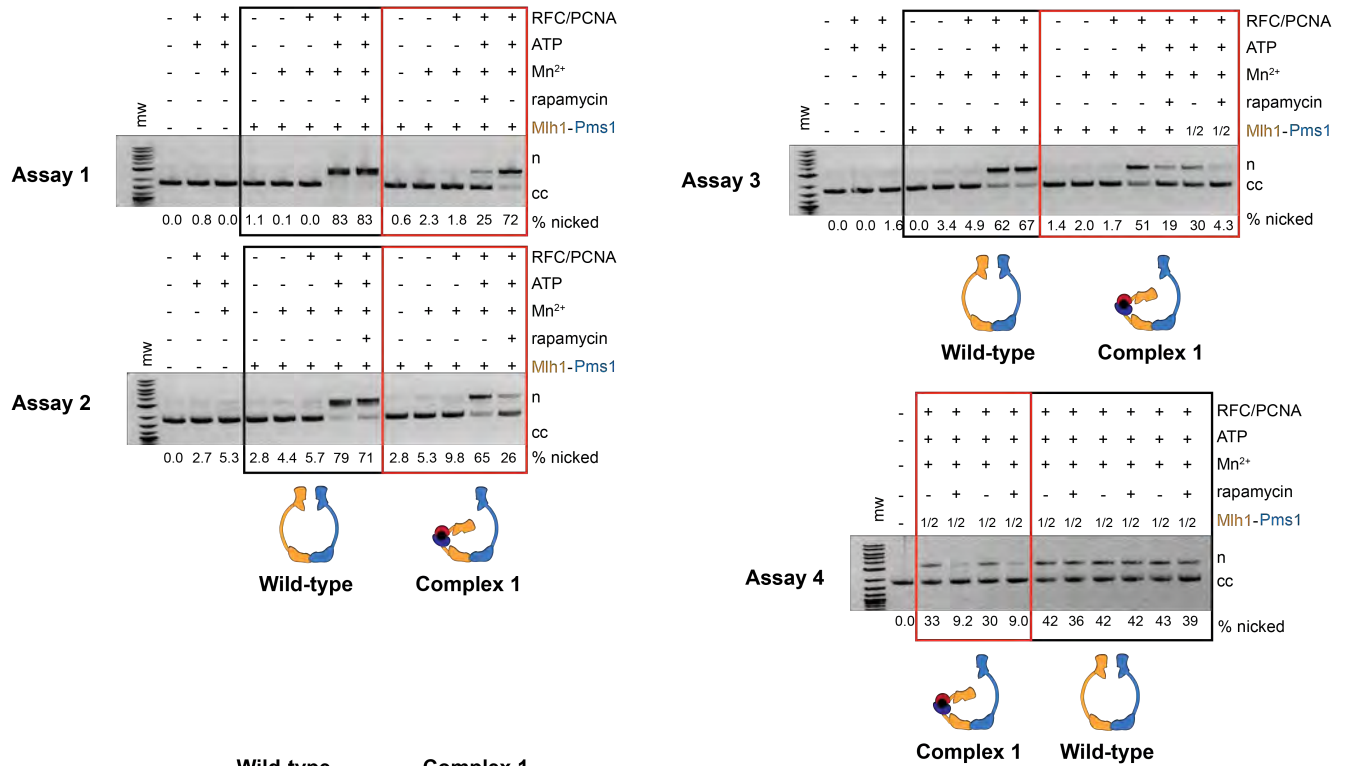


Fig. S3

A



B

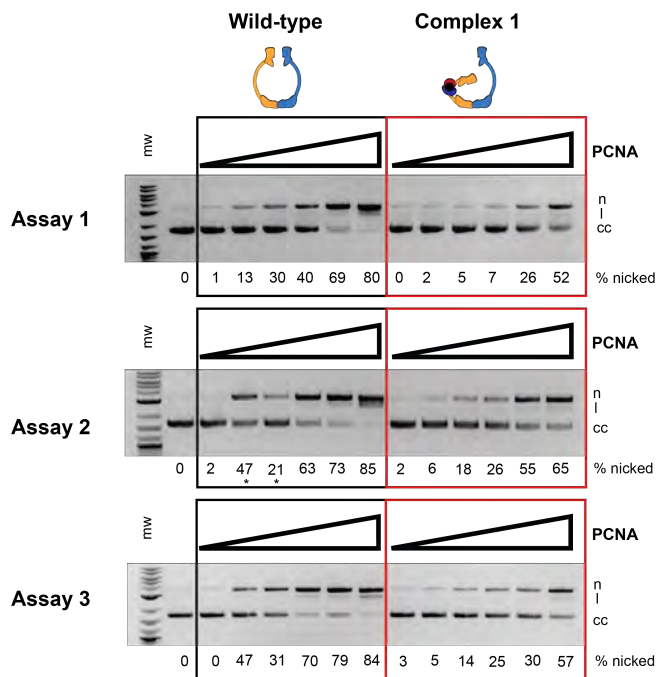


Figure S4

Table S1. Strains used in this study

EAY1269	<i>MATa, ura3-52, leu2Δ1, trp1Δ63, lys2::insE-A14</i>
EAY4209-4211	<i>MATa, ura3-52, leu2Δ1, trp1Δ63, lys2::insE-A14, tor1-1-S1972A</i>
EAY4212-4215, 4449	<i>MATa, ura3-52, leu2Δ1, trp1Δ63, lys2::insE-A14, tor1-1-S1972A, fpr1Δ::KANMX</i>
EAY4450	<i>MATa, ura3-52, leu2Δ1, trp1Δ63, lys2::insE-A14, tor1-1-S1972A, fpr1Δ::NATMX</i>
EAY4488-4490	<i>MATa, ura3-52, leu2Δ1, trp1Δ63, lys2::insE-A14, tor1-1-S1972A, fpr1Δ::NATMX, mlh1Δ::KANMX</i>
JJY70	<i>MATa, fpr1Δ::NAT, tor1-1, leu2-3,112, ura3-52, his3-Δ200, trp1-Δ901, suc2-Δ9 lys2-801; GAL</i>
BJ2168	<i>MATa, ura3-52, leu2-3, 112, trp1-289, prb1-1122, prc1-407, pep4-3</i>
<i>mlh1</i> integrations	
EAY4470-4471	<i>MATa, ura3-52, leu2Δ1, trp1Δ63, lys2::insE-A14, tor1-1-S1972A, fpr1Δ::NATMX, mlh1-FKBP₄₆₄::KANMX</i>
EAY4472-4473	<i>MATa, ura3-52, leu2Δ1, trp1Δ63, lys2::insE-A14, tor1-1-S1972A, fpr1Δ::NATMX, mlh1-FRB₃₅₅::KANMX</i>
EAY4474-4475,4491	<i>MATa, ura3-52, leu2Δ1, trp1Δ63, lys2::insE-A14, tor1-1-S1972A, fpr1Δ::NATMX, mlh1-FRB₄₆₄::KANMX</i>
EAY4598-4600	<i>MATa, ura3-52, leu2Δ1, trp1Δ63, lys2::insE-A14, tor1-1-S1972A, fpr1Δ::NATMX, mlh1-FRB₃₅₅,FKBP₄₆₄::KANMX</i>
<i>pms1</i> integrations	
EAY4492-4494, 4504	<i>MATa, ura3-52, leu2Δ1, trp1Δ63, lys2::insE-A14, tor1-1-S1972A, fpr1Δ::NATMX, pms1-FRB₄₆₀::LEU2</i>
EAY4495-4496, 4505	<i>MATa, ura3-52, leu2Δ1, trp1Δ63, lys2::insE-A14, tor1-1-S1972A, fpr1Δ::NATMX, pms1-FKBP₄₆₀::LEU2</i>
EAY4497-4498, 4506	<i>MATa, ura3-52, leu2Δ1, trp1Δ63, lys2::insE-A14, tor1-1-S1972A, fpr1Δ::NATMX, pms1-FRB₆₂₇::LEU2</i>
EAY4499-4500, 4507	<i>MATa, ura3-52, leu2Δ1, trp1Δ63, lys2::insE-A14, tor1-1-S1972A, fpr1Δ::NATMX, pms1-FKBP₆₂₇::LEU2</i>
<i>mlh1, pms1</i> integrations	
EAY4451-4452, 4486	<i>MATa, ura3-52, leu2Δ1, trp1Δ63, lys2::insE-A14, tor1-1-S1972A, fpr1Δ::NATMX, mlh1-FKBP₄₆₄::KANMX, pms1-FRB₆₂₇::LEU2</i>
EAY4453-4454, 4508	<i>MATa, ura3-52, leu2Δ1, trp1Δ63, lys2::insE-A14, tor1-1-S1972A, fpr1Δ::NATMX, mlh1-FRB₄₆₄::KANMX, pms1-FKBP₄₆₀::LEU2</i>
EAY4455-4456, 4509	<i>MATa, ura3-52, leu2Δ1, trp1Δ63, lys2::insE-A14, tor1-1-S1972A, fpr1Δ::NATMX, mlh1-FKBP₄₆₄::KANMX, pms1-FRB₄₆₀::LEU2</i>
EAY4457-4458, 4487	<i>MATa, ura3-52, leu2Δ1, trp1Δ63, lys2::insE-A14, tor1-1-S1972A, fpr1Δ::NATMX, mlh1-FRB₃₅₅::KANMX, pms1-FKBP₄₆₀::LEU2</i>
EAY4501-4503	<i>MATa, ura3-52, leu2Δ1, trp1Δ63, lys2::insE-A14, tor1-1-S1972A, fpr1Δ::NATMX, mlh1-FRB₃₅₅::KANMX, pms1-FKBP₆₂₇::LEU2</i>

Strains used in this study were derived from the S288c background.

Table S2. Plasmids used in this study

Plasmid	Relevant genotype	Vector type	Source
pRS415	empty vector	<i>ARS-CEN, LEU2</i>	Christianson et al., 1992
pRS416	empty vector	<i>ARS-CEN, URA3</i>	Christianson et al., 1992
pEAA213	<i>MLH1::KANMX</i>	<i>ARS-CEN, LEU2</i>	Alani lab
pEAA238	<i>PMS1</i>	<i>ARS-CEN, HIS3</i>	Alani lab
pEAA248	<i>MLH1</i>	<i>ARS-CEN, URA3</i>	Alani lab
pEAA67	<i>MLH1, PMS1, MSH2</i>	<i>ARS-CEN, URA3</i>	Alani lab
pEAA672	<i>mlh1-FRB₃₅₅::KANMX</i>	<i>ARS-CEN, LEU2</i>	Alani lab; this study
pEAA674	<i>mlh1-FKBP₄₆₄::KANMX</i>	<i>ARS-CEN, LEU2</i>	Alani lab; this study
pEAA675	<i>mlh1-FRB₄₆₄::KANMX</i>	<i>ARS-CEN, LEU2</i>	Alani lab; this study
pEAA713	<i>mlh1-FRB₃₅₅,FKBP₄₆₄::KANMX</i>	<i>ARS-CEN, LEU2</i>	Alani lab; this study
pEAA676	<i>pms1-FRB₄₆₀</i>	<i>ARS-CEN, HIS3</i>	Alani lab; this study
pEAA677	<i>pms1-FKBP₄₆₀</i>	<i>ARS-CEN, HIS3</i>	Alani lab; this study
pEAA678	<i>pms1-FRB₆₂₇</i>	<i>ARS-CEN, HIS3</i>	Alani lab; this study
pEAA671	<i>pms1-FKBP₆₂₇</i>	<i>ARS-CEN, HIS3</i>	Alani lab; this study
pEAE448	<i>GAL1-mlh1-FRB₃₅₅(FLAG₄₉₉)-VMA1-CBD</i>	<i>2μ, TRP1</i>	Alani lab; this study
pEAE447	<i>GAL1-mlh1-FKBP₄₆₄(FLAG₄₉₉)-VMA1-CBD</i>	<i>2μ, TRP1</i>	Alani lab; this study
pEAE446	<i>GAL1-mlh1-FRB₄₆₄(FLAG₄₉₉)-VMA1-CBD</i>	<i>2μ, TRP1</i>	Alani lab; this study
pEAE460	<i>GAL1- mlh1-FRB₃₅₅,FKBP₄₆₄-VMA1-CBD</i>	<i>2μ, TRP1</i>	Alani lab; this study
pEAE435	<i>GAL10-pms1-FRB₄₆₀</i>	<i>2μ, LEU2</i>	Alani lab; this study
pEAE433	<i>GAL10-pms1-FKBP₄₆₀</i>	<i>2μ, LEU2</i>	Alani lab; this study
pEAE431	<i>GAL10-pms1-FRB₆₂₇</i>	<i>2μ, LEU2</i>	Alani lab; this study
pEAI453	<i>pms1-FRB₄₆₀::LEU2</i>	<i>ARS-CEN, LEU2, HIS3</i>	Alani lab; this study
pEAI454	<i>pms1-FKBP₄₆₀::LEU2</i>	<i>ARS-CEN, LEU2, HIS3</i>	Alani lab; this study
pEAI455	<i>pms1-FRB₆₂₇::LEU2</i>	<i>ARS-CEN, LEU2, HIS3</i>	Alani lab; this study
pEAI468	<i>pms1-FKBP₆₂₇::LEU2</i>	<i>ARS-CEN, LEU2, HIS3</i>	Alani lab; this study
pEAI160	<i>mlh1Δ::KANMX</i>	<i>Integration</i>	Alani lab
pEAE269	<i>GAL1-MLH1(FLAG₄₉₉)-VMA1-CBD</i>	<i>2μ, TRP1</i>	Alani lab
pMH8	<i>GAL10-PMS1</i>	<i>2μ, LEU2</i>	Hall and Kunkel, 2001

Full plasmid descriptions can be found in the Materials and Methods. *mlh1-FRB_{xxx}*, *mlh1-FKBP_{xxx}*, *pms1-FRB_{xxx}*, and *pms1-FKBP_{xxx}* refer to the amino acids (XXX) in Mlh1 or Pms1 immediately after which the FRB or FKBP domains were inserted (Figure 1; Materials and Methods).

40.74	2	1608.863	4	402.9717	QKEFSK	Pms1-P14242	N-Term-159	klPSIK	Pms1-P14242	Linker-468
40	1	2965.459	4	742.1207	DANTINDNDLKDQPKKK	Mlh1-P38920	Linker-471	QkLGDYK	Mlh1-P38920	Linker-475
40	1	2993.34	4	749.0908	mcSQSEQQAQKR	Pms1-P14242	Linker-377	kNISSVFGAGGmR	Pms1-P14242	N-Term-213
40	1	2868.362	4	717.8463	SISKDNVYR	Pms1-P14242	Linker-591	kFEDEILEVNLSTK	Pms1-P14242	Linker-605
40	1	2081.095	4	521.0297	QKEFSK	Pms1-P14242	N-Term-159	IkALDASVVNK	Mlh1-P38920	N-Term-6
39.98	1	3058.582	5	612.5226	VPKER	Mlh1-P38920	C-Term-504	LGDYkVPSIADDEKNALPISK	Mlh1-P38920	Linker-480
39.96	1	1891.99	4	473.7535	NSkDR	Pms1-P14242	N-Term-278	VNVNLTSlkK	Mlh1-P38920	C-Term-515
39.91	1	2930.352	4	733.3438	NGkQMSSlISK	Pms1-P14242	Linker-628	NkDELEDFEQGEK	Pms1-P14242	Linker-649
39.87	1	2008.037	4	502.7651	VPKER	Mlh1-P38920	C-Term-504	VcNLNFIkSkK	Mlh1-P38920	N-Term-253
39.86	1	1559.759	4	390.6956	kTMTR	Pms1-P14242	C-Term-830	QKEFSK	Pms1-P14242	N-Term-159
39.56	1	2343.127	4	586.5375	kTMTR	Pms1-P14242	C-Term-830	LGDYkVPSIADDEK	Mlh1-P38920	Linker-480
29.82	1	3287.701	4	822.681	IkALDASVVNK	Mlh1-P38920	N-Term-6	LQkFEDLSlQlTYGFR	Mlh1-P38920	N-Term-84
29.78	1	1751.9	4	438.7308	DQPKKK	Mlh1-P38920	Linker-471	QkLGDYK	Mlh1-P38920	Linker-475
29.52	1	2934.349	4	734.3432	kTmTR	Pms1-P14242	C-Term-830	VDTSDASLSEDEkAQFINR	Mlh1-P38920	C-Term-717
19.78	1	1584.828	4	396.9629	kSlSK	Pms1-P14242	Linker-589	NkFEISK	Pms1-P14242	C-Term-621

Definitions

lower case k indicates site of cross-link

Uniprot: <https://www.uniprot.org>

Score: MaxLinker score; see Yugandhar et al. (2020) Mol. Cell Proteomics 19, 554-568 (2020).

#CSMs: Number of cross-link spectrum matches

XL MH+: Deconvoluted mass of the cross-link. It is a standard practice to present the deconvoluted mass along with the mass of an additional H+

Z: Charge of the cross-link

XL m/z: Cross-link mass divided by its charge

59.95	1	1785.842	4	447.2163	VkEDR	Mh1-P38920	N-Term-117	SISKDNYR	Pms1-P14242	Linker-591
59.95	1	2200.016	4	550.7598	McSQSEQQAQQR	Pms1-P14242	Linker-377	KSISK	Pms1-P14242	Linker-589
59.93	1	2286.03	4	572.2634	McSQSEQQAQQR	Pms1-P14242	Linker-377	TFPKR	Pms1-P14242	FKBP-17
59.93	1	3225.534	4	807.1393	MLGKYTDOPDFLDDLYK	Pms1-P14242	N-Term-246	IAKfQDVAK	Pms1-P14242	N-Term-83
59.92	1	1559.756	4	390.6948	KTMTR	Pms1-P14242	C-Term-830	QkEFSK	Pms1-P14242	N-Term-159
59.9	1	1967.995	4	492.7546	KSISK	Pms1-P14242	Linker-589	LEGSNSSTPTKK	Pms1-P14242	Linker-467
59.88	1	1635.83	4	409.7134	cSkIR	Pms1-P14242	C-Term-807	TPLKNSR	Pms1-P14242	Linker-583
59.86	1	1661.828	4	416.2128	KSISK	Pms1-P14242	Linker-589	YmKSGNVK	Mh1-P38920	FRB-70
59.85	1	3214.525	5	643.7113	YmKSGNVK	Mh1-P38920	FRB-70	NVKGMEVLEPLHAMMER	Mh1-P38920	FRB-25
59.82	1	1755.897	4	439.7301	VkEDR	Mh1-P38920	N-Term-117	FTTSkLQK	Mh1-P38920	N-Term-81
59.8	1	1942.012	4	486.2588	KSISK	Pms1-P14242	Linker-589	VcNLNFISKK	Mh1-P38920	N-Term-253
59.77	1	4007.996	4	1002.755	RQENKLVLR	Mh1-P38920	Linker-398	IDASQAKITSFLSSSQQFNFEQSSTK	Mh1-P38920	Linker-408
59.77	1	2425.204	4	607.057	IAKfQDVAK	Pms1-P14242	N-Term-83	LEGSNSSTPTKK	Pms1-P14242	Linker-467
59.64	1	1877.041	4	470.0161	KSISK	Pms1-P14242	Linker-589	IkALDASVVNK	Mh1-P38920	N-Term-6
40	1	2209.136	4	553.04	TPLKNSR	Pms1-P14242	Linker-583	FmLGQEVIR	Pms1-P14242	FKBP-52
39.99	1	3385.775	4	847.1997	TVFNKSVASNLITFHISK	Mh1-P38920	N-Term-219	VcNLNFISKK	Mh1-P38920	N-Term-253
39.99	1	2343.124	4	586.5369	KTMTR	Pms1-P14242	C-Term-830	LGDYKVPsIAADDEK	Mh1-P38920	Linker-480
39.97	1	1679.855	4	420.7197	KTMTR	Pms1-P14242	C-Term-830	QENKLVLR	Mh1-P38920	Linker-398
39.96	1	2335.156	4	584.545	VPKER	Mh1-P38920	C-Term-504	LGDYKVPsIAADDEK	Mh1-P38920	Linker-480
39.96	1	1608.865	4	402.9722	QkEFSK	Pms1-P14242	N-Term-159	KLPSIK	Pms1-P14242	Linker-468
39.95	1	2935.378	5	587.8818	NSKDR	Pms1-P14242	N-Term-278	RPVEYSTLLkccNEVYK	Mh1-P14242	N-Term-297
39.54	1	1773.925	4	444.2372	KYmKSGNVK	Mh1-P38920	FRB-70	KSISK	Pms1-P14242	Linker-589
30.6	2	2965.451	4	742.1187	DANTINDNDLKDQPKKK	Mh1-P38920	Linker-471	QkLGDYK	Mh1-P38920	Linker-475
30.53	2	2016.115	4	504.7847	DQPKKK	Mh1-P38920	Linker-471	VNVLNLTskk	Mh1-P38920	C-Term-515
29.77	1	1551.783	4	388.7015	VPKER	Mh1-P38920	C-Term-504	QkEFSK	Pms1-P14242	N-Term-159
29.71	1	2613.505	4	654.132	TPLKNSR	Pms1-P14242	Linker-583	SLPLLLkGYIPSLVK	Mh1-P38920	C-Term-657
29.6	1	2900.422	4	725.8613	RQENKLVLR	Mh1-P38920	Linker-398	DANTINDNDLKDQPK	Mh1-P38920	Linker-467
29.6	1	2041.99	4	511.2533	LEGSNSSTPTKK	Pms1-P14242	Linker-467	KTMTR	Pms1-P14242	C-Term-830
29.52	1	1943.065	4	486.5222	IkALDASVVNK	Mh1-P38920	N-Term-6	VPKER	Mh1-P38920	C-Term-504
29.38	1	1845.964	4	462.2468	RQENKLVLR	Mh1-P38920	Linker-398	VkEDR	Mh1-P38920	N-Term-117
19.87	1	1528.812	4	382.959	VPKER	Mh1-P38920	C-Term-504	DQPKKK	Mh1-P38920	Linker-471
19.76	1	1761.966	4	441.2474	KSISK	Pms1-P14242	Linker-589	RQENKLVLR	Mh1-P38920	Linker-398
19.75	1	1981.026	4	496.0124	KTMTR	Pms1-P14242	C-Term-830	SSIMIGkPLNK	Pms1-P14242	C-Term-825

Definitions

Score: MaxLinker score; see Yugandhar et al. (2020) Mol. Cell Proteomics 19, 554-568 (2020).

#CSMs: Number of cross-link spectrum matches

XL MH+: Deconvoluted mass of the cross-link. It is a standard practice to present the deconvoluted mass along with the mass of an additional H+

Z: Charge of the cross-link

XL m/z: Cross-link mass divided by its charge

Uniprot: <https://www.uniprot.org>

lower case k indicates site of cross-link

Table S3C. Summary of intermolecular crosslinks for Mlh1-Pms1 and Complex #5.

Mlh1-Pms1 (15 in total, with the amino acid positions crosslinked in each subunit shown)

Mlh1-N-Term-6 + Pms1-N-Term-83
Mlh1-N-Term-6 + Pms1-N-Term-159
Mlh1-Linker-398 + Pms1-Linker-583
Mlh1-Linker-398 + Pms1-Linker-591
Mlh1-Linker-480 + Pms1-C-Term-830
Mlh1-Linker-489 + Pms1-C-Term-830
Mlh1-Linker-489 + Pms1-Linker-583
Mlh1-Linker-489 + Pms1-Linker-591
Mlh1-C-Term-504 + Pms1-Linker-605
Mlh1-C-Term-515 + Pms1-N-Term-278
Mlh1-C-Term-515 + Pms1-Linker-583
Mlh1-C-Term-515 + Pms1-Linker-589
Mlh1-C-Term-515 + Pms1-Linker-605
Mlh1-C-Term-717 + Pms1-C-Term-672
Mlh1-C-Term-717 + Pms1-C-Term-830

Complex #5 (34, with the amino acid positions crosslinked in each subunit shown)

Mlh1-N-Term-6 + Pms1-Linker-583
Mlh1-N-Term-6 + Pms1-Linker-589
Mlh1-N-Term-81 + Pms1-Linker-583
Mlh1-N-Term-117 + Pms1-N-Term-159
Mlh1-N-Term-117 + Pms1-Linker-467
Mlh1-N-Term-117 + Pms1-Linker-591
Mlh1-N-Term-137 + Pms1-Linker-467
Mlh1-N-Term-137 + Pms1-Linker-583
Mlh1-N-Term-253 + Pms1-Linker-589
Mlh1-Linker-398 + Pms1-Linker-583
Mlh1-Linker-398 + Pms1-Linker-589
Mlh1-Linker-398 + Pms1-C-Term-830
Mlh1-Linker-446 + Pms1-Linker-583
Mlh1-Linker-471 + Pms1-C-Term-830
Mlh1-Linker-472 + Pms1-Linker-583
Mlh1-Linker-480 + Pms1-C-Term-830
Mlh1-Linker-489 + Pms1-Linker-583
Mlh1-C-Term-504 + Pms1-N-Term-159
Mlh1-C-Term-504 + Pms1-Linker-377
Mlh1-C-Term-515 + Pms1-Linker-583
Mlh1-C-Term-515 + Pms1-Linker-589
Mlh1-C-Term-520 + Pms1-Linker-583
Mlh1-C-Term-657 + Pms1-Linker-583
Mlh1-C-Term-657 + Pms1-Linker-591
Mlh1-C-Term-717 + Pms1-C-Term-830
Mlh1-FRB-46 + Pms1-Linker-467
Mlh1-FRB-46 + Pms1-Linker-583
Mlh1-FRB-70 + Pms1-N-Term-159
Mlh1-FRB-70 + Pms1-N-Term-278
Mlh1-FRB-70 + Pms1-Linker-377
Mlh1-FRB-70 + Pms1-Linker-467
Mlh1-FRB-70 + Pms1-Linker-583
Mlh1-FRB-70 + Pms1-Linker-589
Mlh1-FRB-70 + Pms1-C-Term-830

Table S4. Phenotypic analysis of *mlh1* and *pms1* alleles that map within or near Mlh1-Pms1 crosslinking sites (Figure 2A) as determined by XL-MS

Crosslinking site (amino acid position) in Mlh1	<i>mlh1</i> allele	DNA mismatch repair phenotype	Reference
6	<i>R4A, K6A</i>	+/-	Argueso <i>et al</i> , 2003
398	<i>E396A, K398A</i>	+, but – nearby	Argueso <i>et al</i> , 2003
480	<i>D478A, K480A</i>	+	Argueso <i>et al</i> , 2003
489	<i>D486A, D487A, E488A, K489A</i>	+	Argueso <i>et al</i> , 2003
504	<i>K504A, E505A, R506A</i>	+/-	Argueso <i>et al</i> , 2003
515	<i>K515A, K516A</i>	+/-	Argueso <i>et al</i> , 2003
717	<i>E714A, D715A, E716A, K717A</i>	+	Argueso <i>et al</i> , 2003
Crosslinking site (amino acid position) in Pms1			
83	<i>G97A</i>	-	Tran & Liskay, 2000
159	ATP binding domain		Arana <i>et al</i> , 2010
278	Maps to DNA binding region		Hall <i>et al</i> , 2003
583-605	<i>IDR deletion (584-634)</i>	+/-	Plys <i>et al</i> , 2012
672	Maps to endonuclease site		Smith <i>et al</i> , 2013
830	<i>C817R, C848S</i> ; maps to endonuclease site	-	Smith <i>et al</i> , 2013

+ indicates phenotype similar to wild-type, - indicates similar to MMR null, +/- intermediate phenotype.