

1 SUPPLEMENTAL DATA

2 **Sectm1a Deficiency Aggravates Inflammation-Triggered Cardiac Dysfunction**  
3 **through Disruption of LXRA Signaling in Macrophages**  
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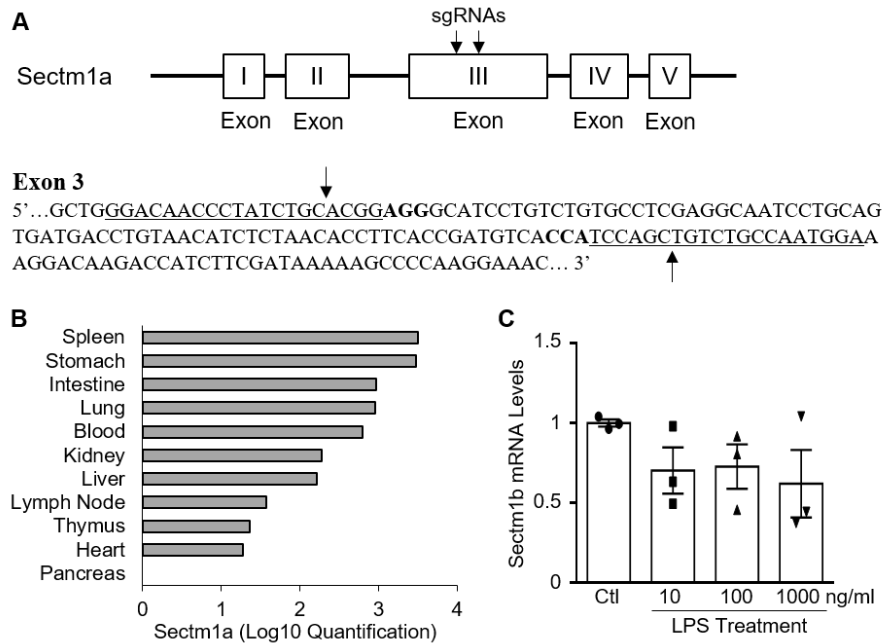
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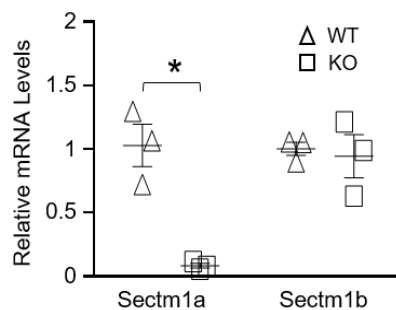


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32 **Fig. S1** Generation of knockout mouse model of Sectm1a. (A) Two gRNAs targeting to Exon3 were  
 33 selected to inject with Cas9mRNA into one-cell embryos. Because of the Cas9 activity, sequence between  
 34 the gRNA targeting sites were deleted. The PAM of each gRNA targeting site is highlighted in bold. The  
 35 spacer sequence is underlined. The cutting site of Cas9 is indicated by arrow. (B) Expression profile of  
 36 Sectm1a in various organs of WT mice were determined using qRT-PCR (n=3). (C) WT-BMDMs were  
 37 treated with indicated doses of LPS for 24 h, Sectm1b expression levels were measured with qRT-PCR  
 38 (n=3). (\*,  $p < 0.05$ , data are presented as Mean  $\pm$  SEM, Student's t test).

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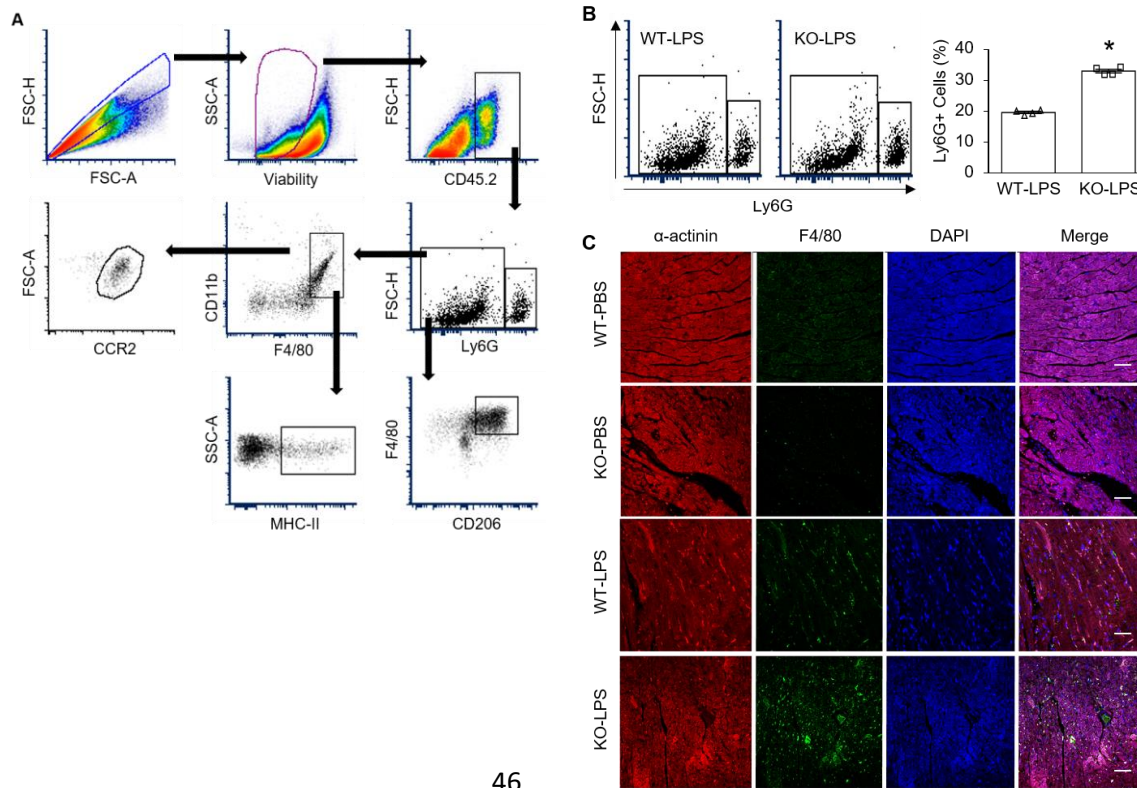
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42 **Fig. S2** q-PCR results of spleen samples confirmed Sectm1a was deleted, whereas Sectm1b gene  
 43 expression was not affected (n=3). (\*,  $p < 0.05$ , data are presented as Mean  $\pm$  SEM, Student's t test).

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47 **Fig. S3** Sectm1a KO mice show stronger cardiac inflammation. (A) Gating strategy for Flow Cytometry  
 48 analysis of cardiac macrophages. (B) Representative plots and quantification of neutrophils (Ly6G+) in  
 49 the heart of WT and Sectm1a KO mice 12 h after LPS treatment (n=4). (C) 12 h after LPS injection, heart  
 50 samples from WT and Sectm1a KO mice were harvested and stained with markers for cardiomyocytes ( $\alpha$ -  
 51 actinin) and macrophage (F4/80), DNA was stained with DAPI (blue). (Scale bar, 50  $\mu$ m; \*,  $p < 0.05$ , data  
 52 are presented as Mean  $\pm$  SEM, Student's t test).

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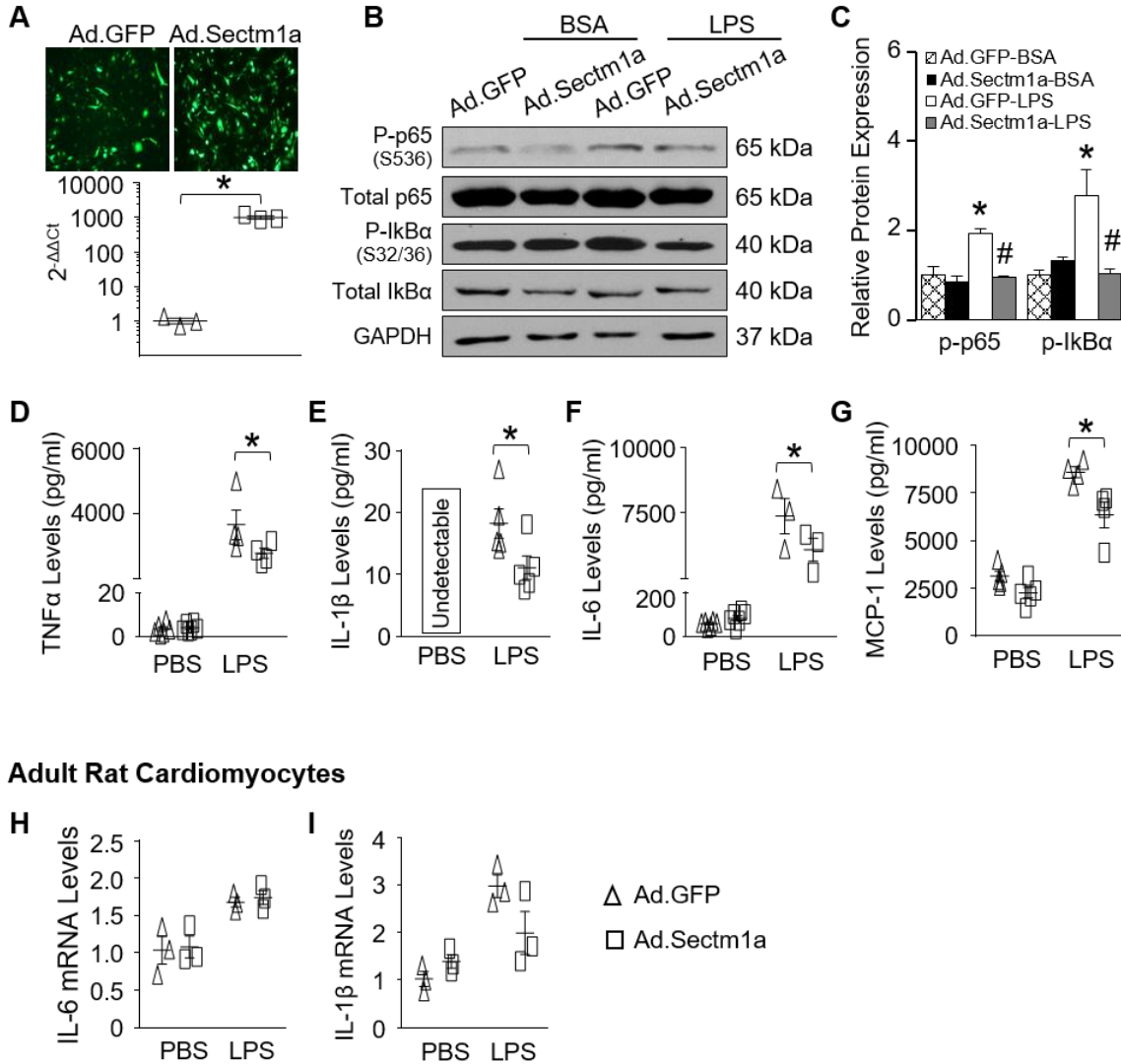
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## Bone Marrow-Derived Macrophages



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59 **Fig. S4** Overexpression of Sectm1a reduces cytokine production in BMDMs, but not in cardiomyocytes.

60 (A) Representative images of BMDMs infected with adenovirus encoding Sectm1a (or GFP as control),

61 and qRT-PCR result validating that overexpression of Sectm1a was successful (n=3). (B) Western

62 blotting and quantification of phosphorylated p65 and IkBα in BMDMs infected with adenovirus,

63 followed by LPS treatment (n=3 dishes of BMDMs for isolation of proteins). (D-G) Concentration of

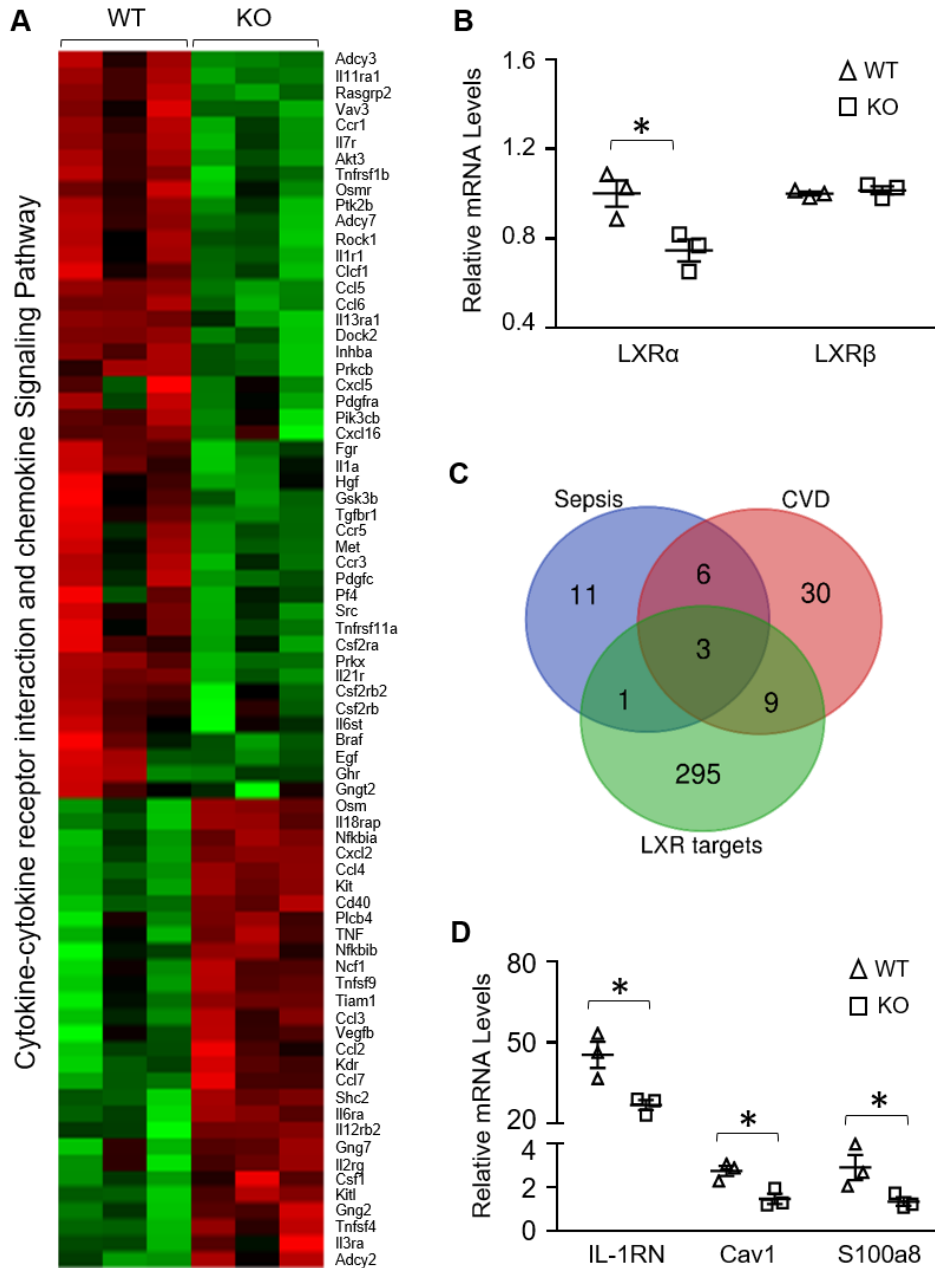
64 cytokines: TNFα (D), IL-1β (E), IL-6 (F), and MCP-1 (G) from cell culture supernatant were determined

65 by ELISA (n=3-5). (H-I) after infecting cardiomyocytes with adenovirus followed by LPS treatment, gene

66 expression of IL-6 and IL-1β in adult rat cardiomyocytes were measured by qRT-PCR (n=3). (\*,  $p < 0.05$ ,

67 data are presented as Mean ± SEM, 2-way ANOVA).

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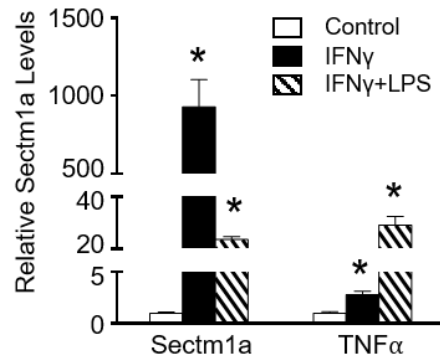


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70 **Fig. S5** Gene expression profile analysis reveals inflammatory phenotype of BMDMs due to disrupted  
 71 LXR signaling. (A) Heatmap comparison of genes involved in cytokine-cytokine receptor interaction and  
 72 chemokine signaling pathway. (B) Expression of LXRα and LXRβ as determined by RNA-seq analyses;  
 73 (C) Venn diagram showing overlapped genes involved in sepsis, cardiovascular disease, and LXR-related  
 74 signaling from our RNA-seq analyses. (D) Three genes overlapped among the 3 pathways mentioned in  
 75 the Venn diagram (C) (n=3 of each genotype). (\*,  $p < 0.05$ , data are presented as Mean ± SEM, 2-way  
 76 ANOVA).

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80 **Fig. S6** WT BMDMs were treated with IFN $\gamma$  alone (10 ng/ml), or together with LPS (10 ng/ml) for 4 h,  
 81 gene expression of Sectm1a and TNF $\alpha$  were determined by qRT-PCR (n=3). (\*,  $p < 0.05$ , data are  
 82 presented as Mean  $\pm$  SEM, Student's t test).

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**Table S1. Echocardiographic measurements in WT and Sectm1a-KO mice after 12 h of LPS injection**

	WT-PBS	KO-PBS	WT-LPS	KO-LPS
Diameter;s	2.72 $\pm$ 0.09	2.41 $\pm$ 0.4	2.43 $\pm$ 0.15	2.92 $\pm$ 0.14
Diameter;d	3.93 $\pm$ 0.09	3.98 $\pm$ 0.14	3.09 $\pm$ 0.16 *	3.37 $\pm$ 0.16
Volume;s	27.90 $\pm$ 2.17	30.67 $\pm$ 2.39	21.46 $\pm$ 3.19	33.46 $\pm$ 3.99
Volume;d	67.68 $\pm$ 3.48	69.91 $\pm$ 5.65	38.43 $\pm$ 4.55 *	47.42 $\pm$ 5.19
Stroke Volume	39.78 $\pm$ 1.53	39.24 $\pm$ 3.51	16.97 $\pm$ 1.85 *	13.96 $\pm$ 1.47
Ejection Fraction	59.25 $\pm$ 1.38	55.86 $\pm$ 1.3	45.10 $\pm$ 2.7 *	29.63 $\pm$ 1.56 #
Fractional Shortening	31.02 $\pm$ 0.92	28.74 $\pm$ 0.89	21.67 $\pm$ 1.48 *	13.44 $\pm$ 0.77 #
Cardiac Output	22.06 $\pm$ 0.72	22.08 $\pm$ 2.22	5.14 $\pm$ 0.95 *	4.99 $\pm$ 0.59

\* ,  $p < 0.05$  when comparing WT-PBS to WT-LPS

# ,  $p < 0.05$  when comparing WT-LPS to KO-LPS

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**Table S2. Echocardiographic measurements in WT and Sectm1a-KO mice with GW3965 and LPS injection**

	WT-DMSO	KO-DMSO	WT-GW3965	KO-GW3965
Diameter;s	3.03 ± 0.26	3.07 ± 0.12	2.43 ± 0.18	3.14 ± 0.19 #
Diameter;d	3.72 ± 0.27	3.55 ± 0.1	3.29 ± 0.17	3.74 ± 0.2
Volume;s	37.51 ± 6.91	37.54 ± 3.3	22.11 ± 4.06	40.19 ± 6.03 #
Volume;d	60.92 ± 9.48	52.77 ± 3.6	45.19 ± 5.83	60.89 ± 8.04
Stroke Volume	23.406875	15.23 ± 0.79	23.07 ± 1.98	20.69 ± 2.05
Ejection Fraction	39.94 ± 2.89	29.28 ± 1.94	53.22 ± 2.7 *	34.68 ± 1.28 #
Fractional Shortening	19.06 ± 1.48	13.35 ± 0.95	26.75 ± 1.58 *	16.2 ± 0.62 #
Cardiac Output	9.51 ± 1.53	5.65 ± 0.61	8.86 ± 1.12	8.82 ± 1.19

\* ,  $p < 0.05$  when comparing WT-DMSO to WT-GW3965

# ,  $p < 0.05$  when comparing WT-GW3965 to KO-GW3965

**Table 3. Echocardiographic measurements in WT and Sectm1a-KO mice with 20 weeks of HFD feeding**

	WT-HFD	KO-HFD
Diameter;s	2.86 ± 0.11	3.29 ± 0.06 *
Diameter;d	4.19 ± 0.1	4.44 ± 0.07
Volume;s	31.77 ± 2.71	43.98 ± 1.97 *
Volume;d	78.97 ± 4.19	90.17 ± 3.38
Stroke Volume	47.2 ± 2.58	46.19 ± 2.41
Ejection Fraction	60.25 ± 2.23	51.1 ± 1.59 *
Fractional Shortening	32.04 ± 1.53	26.01 ± 1.01 *
Cardiac Output	22.65 ± 1.63	19.42 ± 1.26 *

\* ,  $p < 0.05$  when comparing WT-HFD to KO-HFD

**Table S4. Primers used in this study**

<i>qRT-PCR primers</i>		
<b>Gene</b>	<b>Forward</b>	<b>Reverse</b>
Sectm1a	5'-CAGTGATGACCTGTAACATCTC-3'	5'-CAAGTATATCCCTGTGTGGTTCG-3'
Sectm1b	5'-GAGAAGCAGGTAAGAAGCTGGAG-3'	5'-CAGTTCACACCGAAGAACCC-3'
<i>Genotyping primers</i>		
Sectm1a	5'-CATTCTCTCCATACAGGCTGG-3'	5'-CTTGAAGCTGGAGCTCCAC-3'

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**Table S5. Antibodies used for Flow Cytometry**

<b>Name</b>		<b>Company</b>	<b>Cat. #</b>
LIVE/DEAD Stain		Thermo Fisher	L34962
CD45.2	Alexa Flour 488	Biologend	109816
Ly6G	BV421	Biologend	127628
CD11b	APC-eFlour 780	eBioScience	47-0112-80
Ly6C	APC	eBioScience	17-5932-82
F4/80	BV510	Biologend	123135
MHC-II	PerCP-eFluor 710	eBioScience	46-5321-82
CD206	Alexa Flour 700	Bio-Rad	MCA2235A700
CCR2	PE-Cy7	Biologend	150612
CD38	PE-Cyanine 7	eBioScience	25-0381-82
CD301	PE	Biologend	145704

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