

## C-H functionalisation tolerant to polar groups could transform fragment-based drug discovery (FBDD)

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### Assignment of key polar fragment functionalities required for binding to protein

Using the 131 examples of FBDD campaigns detailed in the five Mini-perspectives: Fragment-to-Lead Medicinal Chemistry Publications (2015-2019),<sup>1-5</sup> we initially examined the X-ray or NMR structural information of both the hit and lead (where available) to define the types of fragment polar functional groups making direct interactions with proteinogenic amino acid groups. Water mediated and/or interactions that were not maintained by the lead compound were discounted, as instead we chose to only focus on key hydrogen-bonding interactions required for fragment-protein binding (for further discussion see main text of manuscript).

Pleasingly, the majority (96/131; 73%) of the hit-to-lead papers analysed in this published dataset, had X-ray or NMR structural information detailing the binding of both the fragment hit and lead (or close analogues thereof) to the protein of interest. In some cases, however, structural information for either the hit, lead or both was missing but a putative binding mode was suggested through computational modelling (23/131; 18%), we have highlighted these cases accordingly (Footnote 1, Table S1). For a small number of examples (10/131; 8%), no structural information was available for either the hit, lead or both, therefore determination of the key fragment polar functionalities interacting with the protein was not possible (entries listed in Footnote 2, Table S2). Furthermore, in some cases there was a shift in binding mode for the lead compared to the fragment (13/131; 10%) or the core structure of the fragment was changed enough that it was perceived to be a scaffold hop (6/131; 5%). We have highlighted these examples in our analysis and only assigned fragment polar functionalities interacting with the protein that were maintained by the lead.

### Defining growth vectors

We recognise that defining nominal growth vectors is somewhat subjective, so we created a set of guidelines to try to ensure consistency (Supplementary Information, Figure S1).

- Nominal growth vectors are highlighted as red bonds, when it is not synthetically sensible to highlight the observed change as nominal growth, a synthetically viable bond is instead highlighted in cyan, (e.g. Figure S2, 2015-17)
- A growth vector is defined as being where a new group has been added to the fragment, even if this group is small e.g. ArC–H → ArC–Me (Figure S1, 2015-2)
- If a pre-existing group is modified only slightly (e.g. homologation/ dehomologation) and does not engage any additional protein interactions, this is not counted as a growth vector e.g. nPr → Et (Figure S1, 2015-6)
- If a ring or heterocycle has been changed or expanded, without changing the pharmacophore, this is not defined e.g. pyridine → pyrazole (Figure S1, 2015-7), 6- → 7-membered ring expansion (Figure S1, 2015-4)
- Groups removed from a fragment are not highlighted e.g. ArC–Cl → ArC–H (Figure S1, 2015-2)
- In some cases, a fragment atom was changed to enable a growth vector, this has been highlighted e.g. pyridyl-N → phenyl-CH (Figure S1, 2015-2)
- If a heteroatom has been added to the initial fragment scaffold, this is highlighted in red even if this is not a growth vector (Figure S1, 2019-1), we have done this to highlight the breadth of different heterocycles encountered in FBDD
- The type of bond being formed when growing from the fragment is defined irrespective of the starting fragment atom e.g. the C(sp<sup>2</sup>)–N segment includes cases where a nitrogen is added to a fragment-C(sp<sup>2</sup>) atom and where a C(sp<sup>2</sup>) atom (e.g. arene or alkene) is added to a nitrogen atom located on the fragment

For the majority of the cases in Table S1, defining nominal growth vectors under the constraints listed above was relatively straightforward, however, some cases were more challenging and Figure S2 details a number of select examples to illustrate the range of situations encountered during this analysis. For example, in entry 2015-1 (Figure S2), the fragment hit is entirely encompassed by the lead and one ArC–H has been elaborated with a C–C coupling,

this case is clear-cut. Conversely, entry 2015-17 (Figure S2) shows an example where the approximate designation of growth clearly conflicted with what was synthetically viable. Here, nominal growth is observed to be double alkylation of the amide N–H (shown with red arrows), however amide bond formation is synthetically straightforward and would permit a greater scope of analogues accessible in SAR exploration. In instances like this, the synthetically viable, rather than the strictly nominal, growth vector has been defined (Table S1 & Figure S2, cyan bonds).

In our analysis, we also found examples requiring both the designation of a strictly nominal (red bond) and a more synthetically viable growth vector (cyan bond). This is highlighted in the case of 2017-14 (Figure S2), where ArC–F  $\rightarrow$  to ArC–OAr growth is nominal (red bond), however, the nominal growth vector of the sulfonamide is observed to be from the CH of the methyl group. Considering the robustness of sulfonamide chemistry and the challenge of methyl C–H activation, we have defined the synthetically viable bond between the aniline and the sulfur as being the growth for this case (Figure S2, cyan bond).

We have also encountered more complex examples when defining growth vectors in this dataset, such as 2019-16 (Figure S2). In this example, though the change of an aromatic ethyl to a phenyl can be defined as a simple nominal growth vector, designating the other vectors proved more difficult due to inverted stereochemistry between the fragment and the lead, in addition to the change in linking atom within the fragment scaffold. In this case, we have defined the ArC–N  $\rightarrow$  ArC–O as a synthetically viable growth vector but have also highlighted the methyl  $\rightarrow$  benzyl switch at the stereogenic centre as this comprises both the nominal growth and a change in stereochemistry from the initial fragment (Figure S2).

Table Entry	Fragment	Lead	Guidelines for defining vectors and growth (specific examples highlighted in lead)
2015-2			<p><math>C_{Ar}-Cl \rightarrow C_{Ar}-H</math> not defined ●</p> <p>Groups removed from the original fragment during elaboration are not highlighted</p> <p><math>N \rightarrow C</math> (pyridine to benzene) defined ●</p> <p>Atom changed in fragment to enable synthetic growth</p> <p><math>C_{Ar}-H \rightarrow C_{Ar}-Me</math> defined ●</p> <p>Growth highlighted even if the group added is small</p>
2015-6			<p><math>C_{Ar}-H \rightarrow C_{Ar}-Me</math> not defined ●</p> <p>Modification of pre-existing group on fragment does not engage additional protein interactions ∴ not defined as a growth vector</p>
2015-4			<p><math>6 \rightarrow 7</math> membered cyclic amine not defined ●</p> <p>Ring expanded to improve solvent interactions and not required to enable growth or synthesis</p>
2015-7			<p>Pyridine <math>\rightarrow</math> Pyrazole not defined ●</p> <p>Heterocycle changed but pharmacophore is maintained and no additional protein interactions are acquired</p>
2019-1			<p><math>C \rightarrow N</math> (benzene to pyridine) highlighted ●</p> <p>Heteroatom added during elaboration to pick up additional protein interactions</p>
<p><b>Key:</b></p> <p> Fragment functionality required for protein binding</p> <p> Growth vector (origin and direction of growth during fragment to lead elaboration)</p> <p> Group highlighted to show where growth was defined</p> <p> Group highlighted to explain why growth was not defined</p> <p> Bond formed during growth vector elaboration (nominal / observed growth)</p>			

**Figure S1** Illustrates the guidelines we used to define nominal growth vectors. Fragment and corresponding lead showing the fragment polar binding groups (blue circles) and the nominal fragment growth vectors (red arrows). The new binding groups added onto the lead during fragment elaboration represent hypothetical synthetic bonds (red or cyan bonds). Guidelines for defining growth vectors are summarised in the final column.

Table Entry	Fragment	Lead	Growth Vectors	Bond Designation
2015-1			1 growth vector (C <sub>Ar</sub> -H)	Assigned growth (red): C <sub>Ar</sub> -H → C <sub>Ar</sub> -C <sub>Ar</sub>
2015-17			2 growth vectors (2 x NH)	Assigned growth (cyan): amide bond†
2017-14			2 growth vectors (C <sub>Ar</sub> -F & C <sub>Alkyl</sub> -H)	Assigned growth (red): C <sub>Ar</sub> -F → C <sub>Ar</sub> -OAr Assigned growth (cyan): sulfonamide bond†
2019-16			3 growth vectors (2 x C <sub>Ar</sub> -H & *C <sub>Alkyl</sub> -H) <i>n.b. includes change in stereochemistry</i>	Assigned growth (red): *C <sub>Alkyl</sub> -H → *C <sub>Alkyl</sub> -C <sub>Alkyl</sub> Assigned growth (red): C <sub>Ar</sub> -H → C <sub>Ar</sub> -C <sub>Ar</sub> Assigned growth (cyan): aryl ether†

**Key:**

Fragment functionality required for protein binding



Growth vector (origin and direction of growth during fragment to lead elaboration)

Bond formed during growth vector elaboration (nominal / observed growth)

Synthetically viable bond defined (in the absence of pertinent nominal growth)

† Defined bond is more synthetically viable to enable scope

**Figure S2** Shows specific examples of nominal and or synthetically viable growth. For each entry, the polar binding groups on the fragment are highlighted (blue circles) in addition to the nominal fragment growth vectors elaborated in the lead to increase binding affinity (red arrows). The new binding groups added onto the lead during fragment elaboration represent hypothetical synthetic bonds (red or cyan bonds).

## Astex Overlay Page Help <https://astx.com/interactive/F2L-2021/>

### Overview

The overlay pages provide a curated view of a series of protein-ligand structures. The structures can be explored and displayed through the hierarchical menus in the right hand panel.

Structures have some basic top-level controls: checkboxes and colour pickers to control the protein, ligand, waters and simple molecular surfaces.

Expanding a structure displays further controls for different display styles and controls to turn on electron density maps (where available). The maps are often clipped to the immediate vicinity of the ligand to minimize file sizes.

### Mouse Controls

- **Rotate** - *Left* button hold and move
- **Zoom** - *Shift+Left* button hold and move, **OR** *Right* button hold and drag (up/down)
- **Translate** - *Ctrl+Left* button hold and move
- **Adjust clipping planes** - *Scroll* mousewheel (**OR** "-" and "+" keys)
- **Pick** - *Left* click on an atom (see measurements below)
- **Centre** - *Middle* click on an atom or bond

### Keyboard Shortcuts

The following keyboard shortcuts are available when the NGL Viewer has focus (i.e. after you click on the viewer area).

#### General

- **(c)**entre – recentre on the last picked atom
- **(r)**eset – zooms to view all *loaded* structures
- **Sp(i)n** – toggle spin mode
- **Roc(k)** – toggle rock mode
- **(\_-)** – decrease depth-of-field (move clipping planes together) **OR** mouse scrollwheel up
- **(\_+)** – increase depth of field (move clipping planes apart) **OR** mouse scrollwheel down

#### Measurements

Pick to select atoms, then:

- **(d)**istance – operates on last two picked atoms
- **(a)**ngle – operates on last three picked atoms
- **(t)**orsion – operates on last four picked atoms

**(Shift-d/a/t)** clears distances, angles, torsions respectively

## References

1. C. N. Johnson, D. A. Erlanson, C. W. Murray and D. C. Rees, *Journal of Medicinal Chemistry*, 2017, **60**, 89-99.
2. C. N. Johnson, D. A. Erlanson, W. Jahnke, P. N. Mortenson and D. C. Rees, *Journal of Medicinal Chemistry*, 2018, **61**, 1774-1784.
3. P. N. Mortenson, D. A. Erlanson, I. J. P. de Esch, W. Jahnke and C. N. Johnson, *Journal of Medicinal Chemistry*, 2019, **62**, 3857-3872.
4. D. A. Erlanson, I. J. P. de Esch, W. Jahnke, C. N. Johnson and P. N. Mortenson, *Journal of Medicinal Chemistry*, 2020, **63**, 4430-4444.
5. W. Jahnke, D. A. Erlanson, I. J. P. de Esch, C. N. Johnson, P. N. Mortenson, Y. Ochi and T. Urushima, *Journal of Medicinal Chemistry*, 2020, **63**, 15494-15507.





























Total entries		131		191														230											230																					
Year	Entry	Fragment Hit - PDB Code (where available)	Lead - PDB Code (where available)	Institution	Target	Binding pose changed?	Scaffold Hop?	Fragment functionalities interacting with proteins														Nominal growing vectors											Bond formation																	
								# fragment / protein interactions	Arom CH	Aliph CH	Arom N	Arom NH	Anilin eNH	Aliph NH	Amid eNH	CO	Acid COOH	Arom OH	Aliph OH	Arom Hal	Other polar funct.	# nominal growing vectors	Arom CH	Aliph CH	Arom N	Arom NH	Aniline NH	Aliph NH	Amide NH	CO	Acid COOH	Arom OH	Aliph OH	Arom Hal	Other polar funct.	C(sp2)-C(sp2)	C(sp2)-C(sp3)	C(sp3)-C(sp3)	C(sp2)-C(alkyne)	C(sp2)-C(nitrile)	Csp2-N	Csp3-N	amide	Csp2-O	Csp3-O	C-Hal	sulfonamide			
2018	5			Technische Univ Braunschweig, RWTH Aachen, ManFos	DYRK1A	N	N	1																																1										
2018	6	6G82 	6G8N 	Astec	ERK1/2	N	N	2			1																																							
2018	7	6CJE* 	6CK6 	eFFector Therapeutics et al	MNK1/2	N	N	1			1																																							
2018	8			A*Star et al	MNK1/2	N	N	2	1		1																																							
2018	9			A*STAR	PKC iota	N	N	2			1	1																																						
2018	10		5N9R* 	Almac, Queen's Univ Belfast	USP7		N	3	1		1																																							
2018	11	5O4V 	5MU6 	Imperial	Human N-myristoyltransferase	N	N	1						1																																				
2018	12	5OU2 	5OU3 	Univ Cambridge, Univ Cape Town, Univ Melbourne	IMPDH	N	N	1					1																																					
2018	13	6F20 	6FK* 	Sprint Bioscience et al	MTH1	N	N	2			1	1																																						
2018	14			Abbvie	NAMPT	N	N																																											
2018	15	5XUJ 	5XUJ* 	Astellas	PDE10A	N	N	1						1																																				

Table S1 An assessment of 131 Fragment-to-Lead campaigns detailing i) polar fragment functionality interacting with proteins, ii) the nature of the atom growth originated from during fragment-to-lead elaboration and iii) the observed bonds formed during this process.









