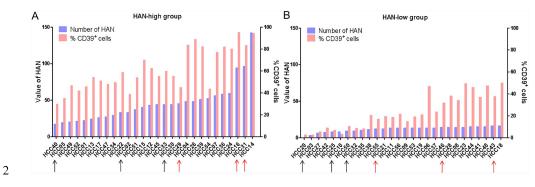
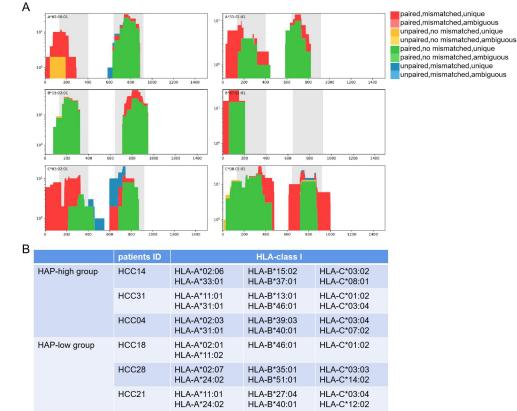
Supplemental Figures and legends



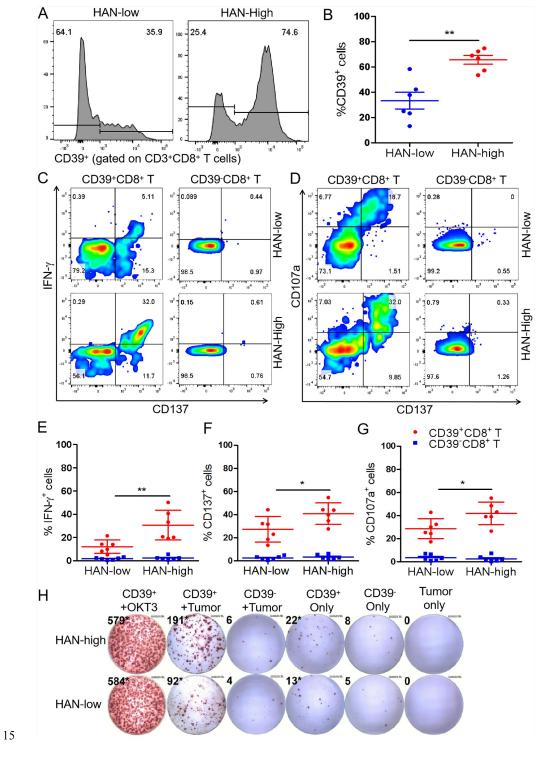
3 Supplemental Figure 1

- 4 Summary of the frequency of CD39⁺CD8⁺ TILs and value of HAN for all the 56 HCC
- 5 patients. (A-B) Red bars represented the frequency of CD39⁺CD8⁺ TILs and blue bars
- 6 represented the value of HAN. Arrows indicated the six representative patients
- 7 selected for TILs and autologous tumors organoids killing assay. Red arrows indicated
- 8 three candidate patients selected for specific high affinity peptide identification assay.



10 Supplemental Figure 2

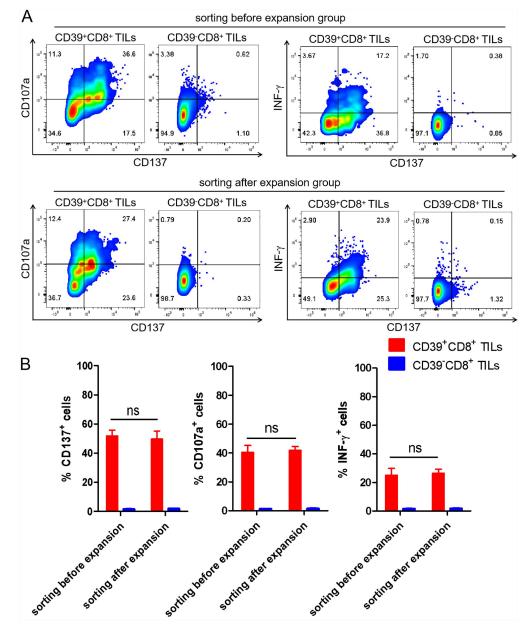
- 11 HLA alleles Analysis by OptiType(v1.3.3) were shown for six candidate patients. (A)
- 12 The coverage plot of the HLA alleles for representative patient HCC14 analyzed by
- 13 OptiType were shown. Listed HLA alleles were with high expression abundance.
- 14 (B) Summary HLA alleles of all the six candidate patients were shown in this table.



16 Supplemental Figure 3

17 CD39 and effector molecules expressed on TILs from surgically removed HCC fresh

18	tumor samples. (A-B) The frequency of CD39 ⁺ CD8 ⁺ T cells in each group was
19	detected by flow cytometry. (C-G)The expression of IFN- γ , CD107a and CD137 on
20	CD39 ^{+/-} CD8 ⁺ T cells was detected by flow cytometry. The dots represent different
21	patients. Data are presented as mean \pm SEM (n=6), *P< 0.05, **P < 0.01.(H) CD39 ^{+/-}
22	CD8 ⁺ TILs were respectively sorted from surgically removed fresh tumor sample and
23	co-cultured with or without (TILs only) autologous tumor cells, then the IFN- γ
24	secretion was investigated by ELISPOT after 24 hours. OKT3 was used as positive
25	control.





27 Supplemental Figure 4

The IFN-γ secretion, CD137 and CD107a expression on CD39^{+/-} TILs had no bias in expansion before sorting group and expansion after sorting group. (A-B) TILs were isolated from surgically removed HCC fresh tumor sample and divided into two groups. The one was the sorting before expansion group which TILs were sorted immediately and then CD39^{+/-} TILs were expanded respectively for 14 days. Another

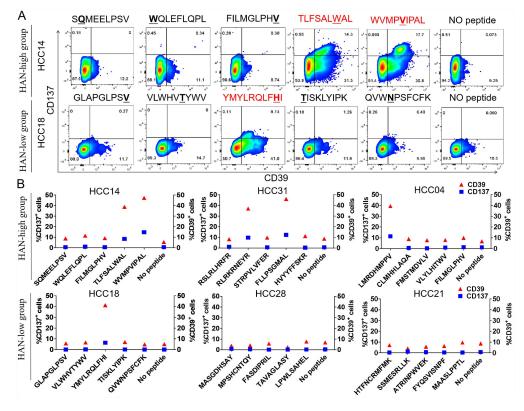
- 33 one was the sorting after expansion group which TILs were expanded first for 14 days
- and then sorted by the marker of CD39. At day 14, TILs of each group was
- 35 co-cultured with tumor cells, and then the function of CD39^{+/-} subsets was assessed by
- flow cytometry with intracellular staining of IFN-γ, CD107a and CD137. (B)
- 37 Summary of the frequency of IFN-γ, CD107a and CD137 positive was shown.

Α									
Patient ID	Positive	Peptides					Negativ		
U	control	1	2	3	4	5	contro (NC)		
HCC14	ОКТ3	S <u>Q</u> MEEL PSV	WQLEFL QPL	FILMGL PH <u>V</u>	TLFSAL <u>W</u> AL	WVMP <u>V</u> I PAL	No peptide		
HCC31	ОКТ3	RSLRLH RF <u>R</u>	RLRK <u>R</u> N EYR	ST <u>R</u> PVL VFER	FLLPSG MAL	HVYYFF <u>S</u> KR	No peptide		
HCC04	ОКТ3	LMRDH	CLM <u>H</u> HL AQA	FMSTMD	VLYLHI T <u>W</u> V	FILMGLP H <u>V</u>	No peptide		
HCC18	ОКТ3	GLAPGL PS <u>V</u>	VLWHV <u>T</u> YWV	YMYLRQ LF <u>H</u> I	<u>T</u> ISKLYI PK	QVW <u>N</u> PS FCFK	No peptid		
HCC28	ОКТ3	MASGD <u>H</u> SAY	MPS <u>H</u> C NTQY	FA <u>S</u> DIPR IL	TAVAGL <u>A</u> SY	LP <u>W</u> LSA HEL	No peptid		
HCC21	ОКТ3	HTFNCR MFMK	SSMES <u>R</u> LLK	A <u>T</u> RINP WVEK	FY <u>Q</u> SVI SNPF	MAAS <u>L</u> P PTL	No peptid		
6	OKT3	1	2	3	4	5	NC		
HCC14		0			115*	139*	U C		
HAP-high group C04 HCC31	83*0	2	15*		143*		• 0		
₽ V		60* 0							
ICC18	62 [*] 0	0		20*					
HAP-low group 1 HCC28 H	69*0	0					0		
HA C21	97*0	0	C C	Longy B			0		

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39 Supplemental Figure 5

- 40 Identification of personalized neoantigens for 6 patients. (A-B) Autologous PBMCs
- 41 were stimulated with top five candidate peptides for 10 days, after which autologous
- 42 tumor organoid from candidate patient was added to activate T-cell specific antigen
- 43 response 24h before IFN-γ detection by ELISPOT assays. OKT3 was used as positive
- 44 control, and no-peptide stimulation was tested as negative control.

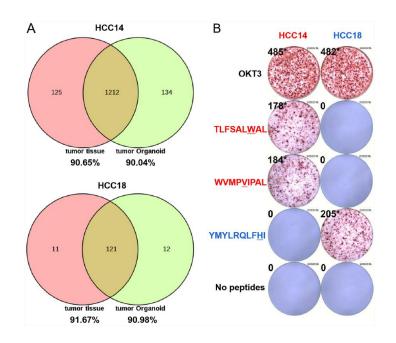


45

46 Supplemental Figure 6

Expression of CD137 and CD39 of tumor-reactive CD39⁺CD8⁺ T cells by stimulation
with candidate HANs from PBMCs. (A) Flow cytometry plots gated on CD8⁺ T cells
to analyze T cell effector sensitivity against tumor organoids by CD137 and CD39
up-regulation. (B) Summary of the frequency of CD39⁺ or CD137⁺ T cells was shown

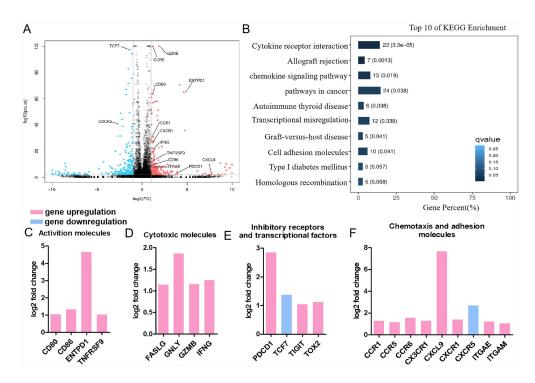
51 for all the six representative patients.



52

53 Supplemental Figure 7

Allogeneic positive peptides couldn't induce IFN-y release for autologous tumor 54 55 organoid killing. (A)Venn diagrams illustrate the number of somatic non-synonymous mutations present in each tumor tissue and their derivative HCC organoids. The 56 57 similarity of mutation from tumor tissue and organoid was greater than 90%. (B) Autologous PBMCs from candidate patients were stimulated with autologous peptides 58 59 (marked by the same color) and allogeneic peptides (marked by the different color) for 10 days, after which autologous tumor organoids from candidate patient was 60 added to activate T-cell specific antigen response 24h before IFN- γ detection by 61 ELISPOT assays. OKT3 was used as positive control, and no-peptide stimulation was 62 tested as negative control. 63



65 Supplemental Figure 8

RNA-seq data of CD39^{+/-}CD8⁺ T cells stimulated by HAN peptide. PBMCs were 66 stimulated by autologous HANs for 10 days and then co-cultured with autologous 67 tumor organoids for 24h. After which CD39^{+/-}CD8⁺ T cells were sorted by FACS for 68 following RNA-seq assay. (A) Volcano graph revealed the difference of genes 69 70 expression analyzed by RNA-seq between CD39^{+/-} CD8⁺ T cells after stimulation by HAN peptide (log2-transformed). The dashed line identified the differently expressed 71 72 genes when using a P value <0.05. (B) KEGG enrichment demonstrated the 73 significant enrichment of top 10 pathways using a P/Q value. (C-F) Bar plots 74 indicated the RNA-seq expression of log2 fold change of major up-regulated and down-regulated genes in CD39⁺CD8⁺ T cells compared to CD39⁻CD8⁺ T cells (P 75 <0.05). Pink bars represented up-regulation and blue bars represented 76 77 down-regulation.