

Supplemental Figures







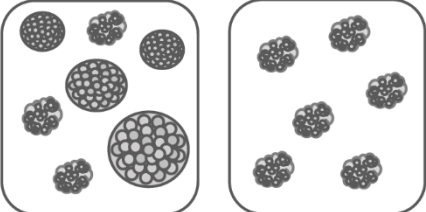
High-throughput imaging assay for drug screening of 3D prostate cancer organoids

Correspondence:

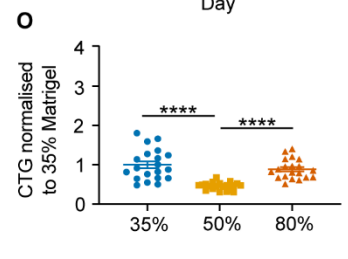
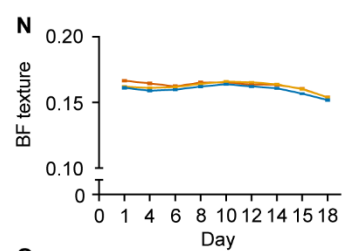
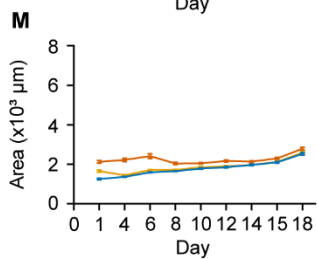
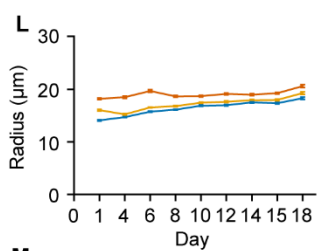
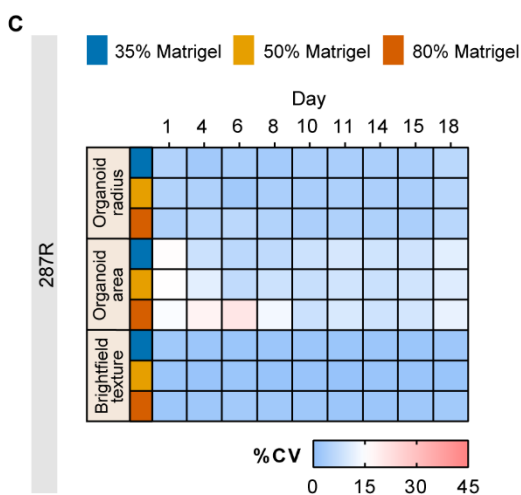
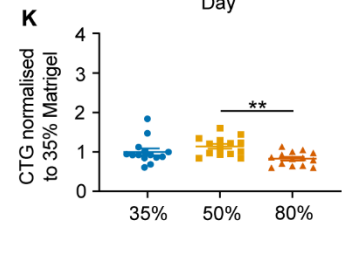
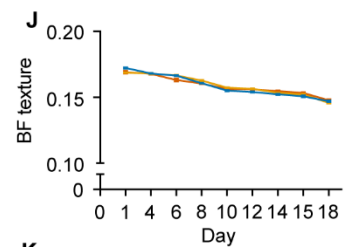
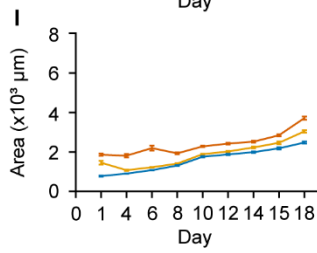
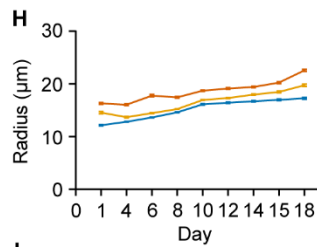
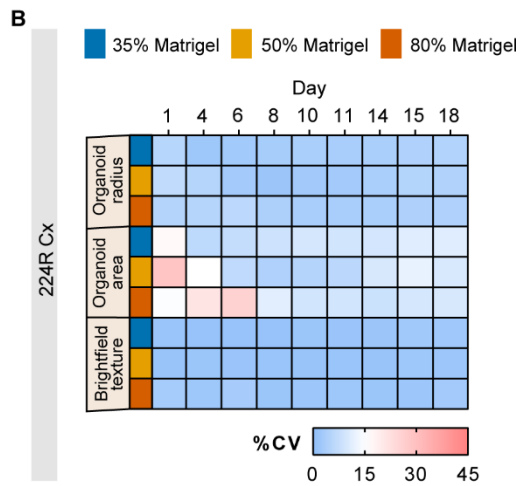
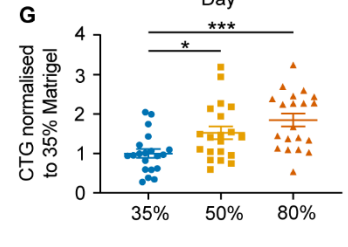
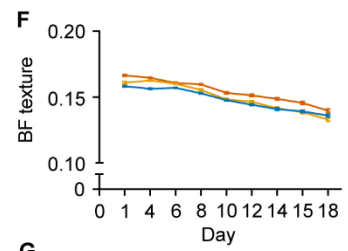
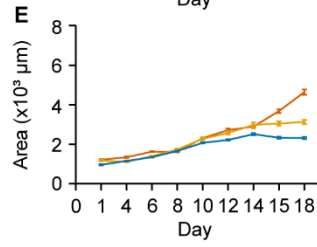
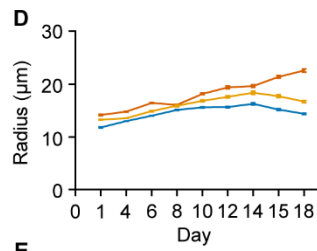
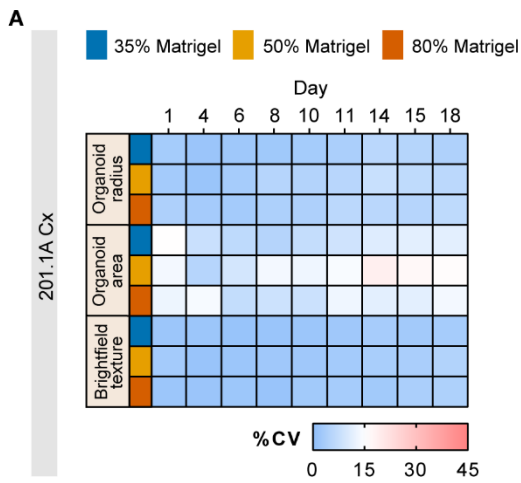
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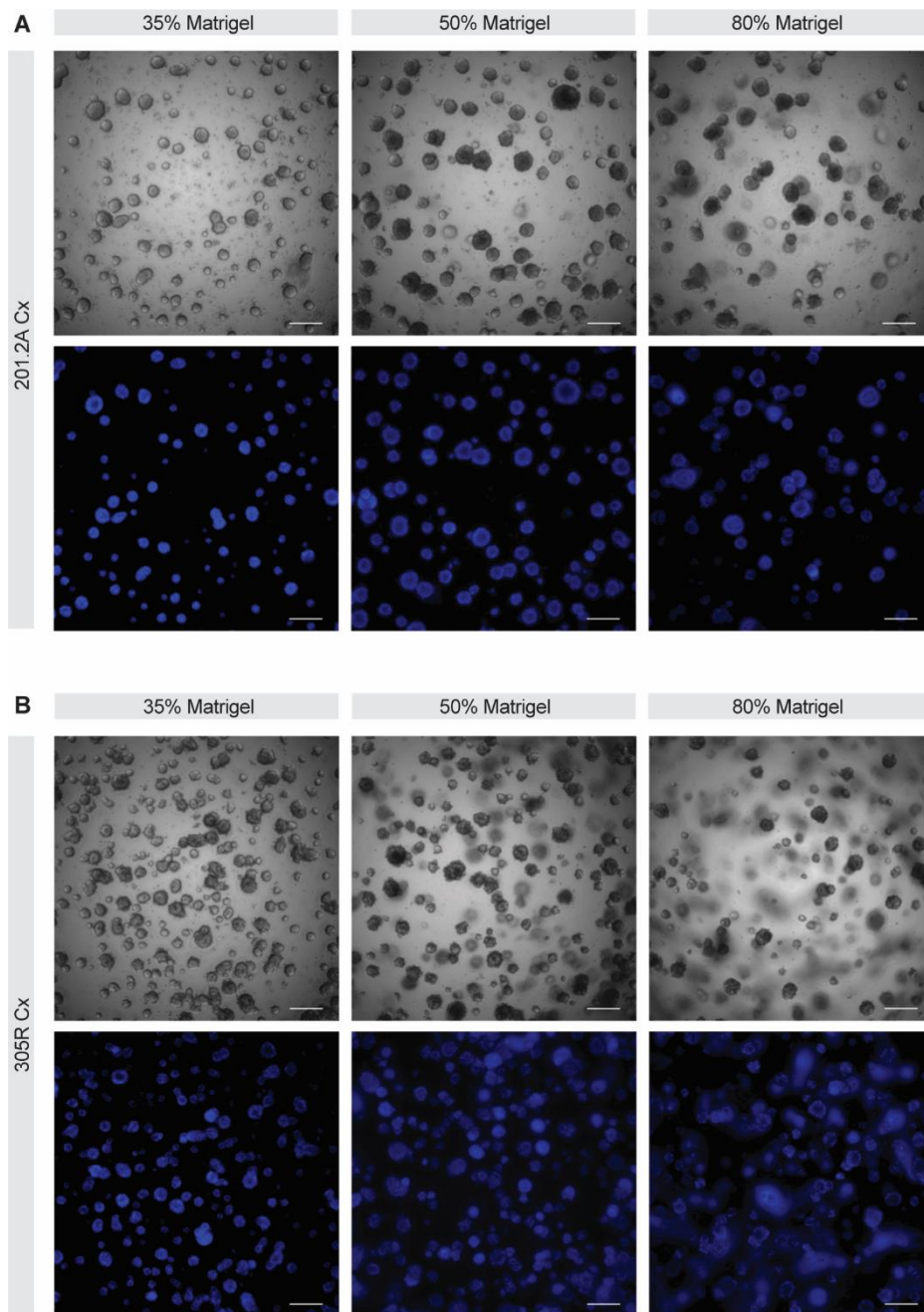
Supplementary Figure 1: Representative plate layout of a high throughput screening assay. (A) Plate layout of organoids grown in different concentrations of Matrigel treated with DMSO and dose responses of talazoparib or carboplatin. (B) Plate layout of a 3D compound screen including DMSO, six positive controls and 42 library compounds.

Imaging feature	Schematic representation
Area	
MeanRadius	
Eccentricity	
FormFactor	
MeanIntensity	
Texture	
Texture_Variance	

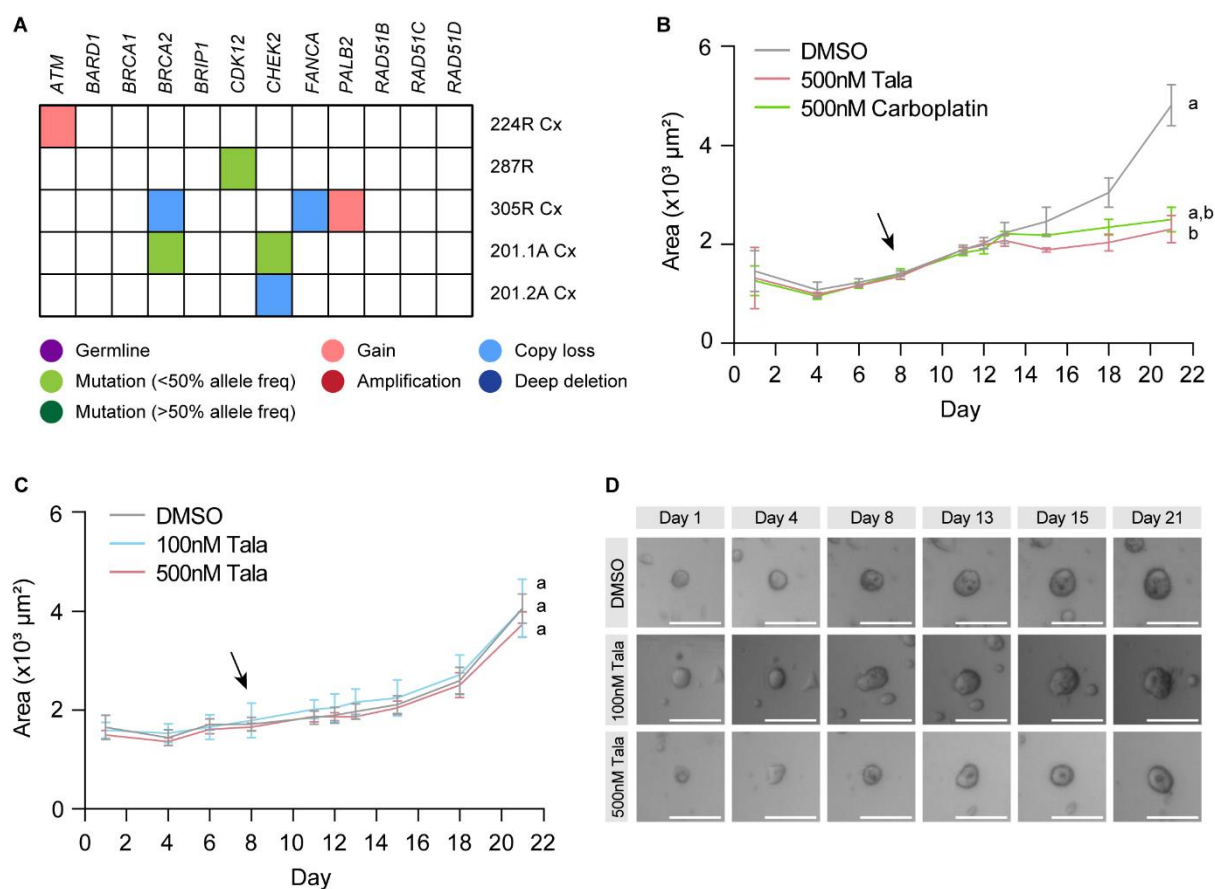
Supplementary Figure 2: Schematic representation of imaging features used for organoid analyses. Imaging features include area, mean radius, eccentricity, form factor, mean intensity, texture and texture variance. A detailed description is also provided in Table 1.



Supplementary Figure 3: Comparison of organoid growth from diverse phenotypes of prostate cancer in different concentrations of Matrigel. (A-C) Heatmaps of percent coefficient of variation for **(A)** 201.1A-Cx, **(B)** 224R-Cx, and **(C)** 287R organoids across 18 days for organoid radius, area, and brightfield texture in 35% (blue), 50% (yellow) and 80% Matrigel (orange). The growth of organoids from 201.1A-Cx **(D-G)**, 224R-Cx **(H-K)**, and 287R **(L-O)** in different concentrations of Matrigel was measured over 18 days ($n = 15$ wells per tumor). Graphs show changes in organoid radius **(D, H, L)**, area **(E, I, M)**, and brightfield texture **(F, J, N)**. Metabolic activity **(G, K, O)** was measured with CTG and normalized to 35% Matrigel. All data are mean \pm SEM; * $P < 0.05$; ** $P < 0.01$, *** $P < 0.001$, **** $P < 0.0001$; one-way ANOVA with post hoc Tukey's test. Abbreviations: CV – coefficient of variation, CTG – CellTiter-Glo.



Supplementary Figure 4: Maximum projections of brightfield and fluorescence microscopy of organoids grown in different concentrations of Matrigel. Representative images of (A) 201.2-Cx and (B) 305R-Cx organoids grown for 21 days in 35%, 50% or 80% Matrigel. Images are maximum projections from three Z heights for brightfield microscopy and fluorescence microscopy of Hoechst staining. Scale bars = 500 μ m.



Supplementary Figure 5: Alterations in DNA damage repair genes and change

in area over time in organoids after treatment. (A) Plot summarising genomic

alterations in DNA damage repair genes based on targeted DNA sequencing of PDX

tissue. There are no homozygous mutations or deep deletions. **(B)** Graph showing the

mean area of 224R-Cx organoids grown for 21 days and treated with DMSO, 500 nM

talazoparib, or 500 nM carboplatin from day 8. Arrow indicates the first day of

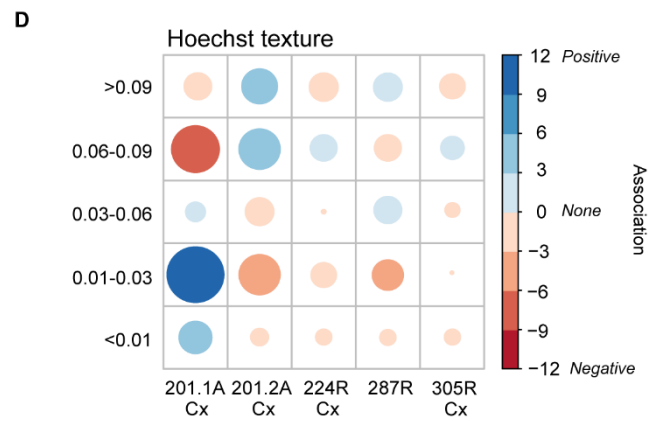
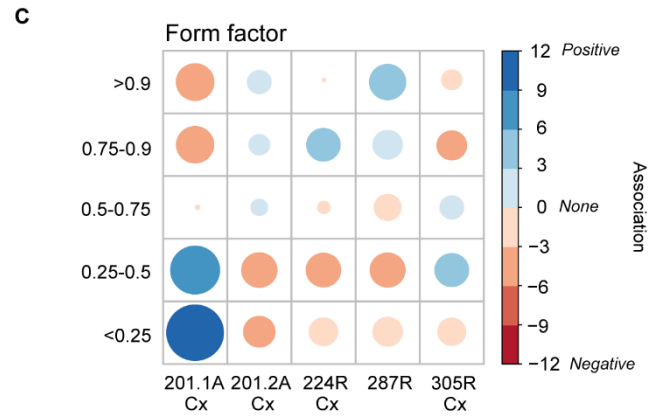
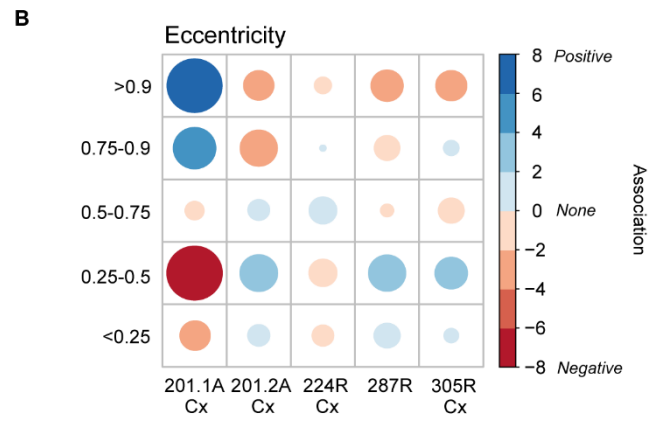
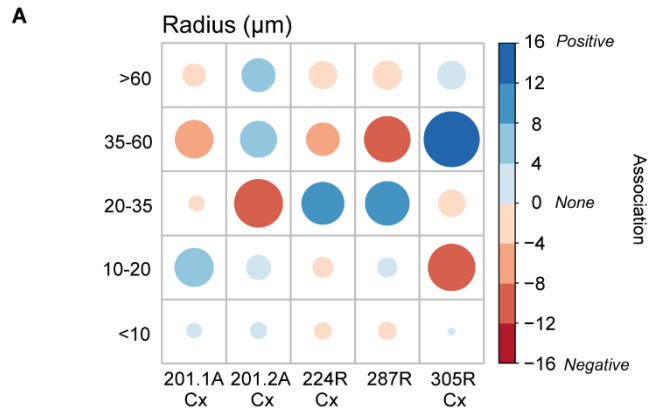
treatment. **(C)** Graph showing the mean area of 287R organoids grown for 21 days

and treated with DMSO, 100 or 500nM talazoparib from day 8. Data are mean \pm SEM.

^a $P < 0.001$, statistical analysis of change in organoid area across time within each

treatment group; ^b $P < 0.001$, statistical analysis of organoid area within each treatment group compared to vehicle control; two-way ANOVA with Dunnett's post hoc test. **(D)**

Representative images of 287R single organoids over time that were treated with DMSO, 100 or 500 nM talazoparib from day 8. Scale bars = 250 μm . Abbreviations: CTG – CellTiter-Glo, Tala – talazoparib.



Supplementary Figure 6: Schematic of Chi-squared analysis results for association between organoid models per bin. The graphical matrices (A, B, C, D) represent Chi-squared tests of the association between organoid models (column) in each bin (row). Positive (blue) and negative (red) residuals specify the association between each organoid per bin. A positive association indicates a higher proportion of organoids in a bin, whereas a negative association indicates a lower proportion of organoids in a bin. The size of residuals (circles) is proportional to the magnitude of the association, positive or negative.