Supplementary material

Role of pyruvate in maintaining cell viability and energy production under high-glucose conditions

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Supporting Information: Tables (4) and Figures (5)

[Table S1] Metabolome analysis; the levels of metabolites involved in glycolysis, TCA cycle, pentose phosphate pathway and purine synthetic pathway in IMS32 cells at 1 h in the [Glc 5 mM/Pyr (+)], [Glc 5 mM/Pyr (-)], [Glc 15 mM/Pyr (+)] and [Glc 15 mM/Pyr (-)]. ND: not detected.

	Concentration (pmol/10 ⁶ cells)						
Compound	[Glc 5 mM/Pyr (+)]	[Glc 5 mM/Pyr (-)]	[Glc 15 mM/Pyr (+)]	[Glc 15 mM/Pyr (-)]			
Glucose 6-phosphate	53	41	48	4			
Fructose 6-phosphate	19	ND	3.6	2			
Fructose 1,6-bisphosphate	1,015	583	1,457	2,11			
Glyceraldehyde 3-phosphate	ND	19	ND	26			
Dihydroxyacetone phosphate	ND	109	47	79			
3-Phosphoglyceric acid	54	62	68	7			
2-Phosphoglyceric acid	2.1	3.4	3.1	3			
Phosphoenolpyruvate	ND	ND	ND	Ν			
Pyruvic acid	1,191	ND	3,151	50			
Lactatic acid	10,962	14,594	26,997	18,89			
Acetyl CoA	16	18	4.0	6			
Citric acid	3,481	2,594	3,696	2,27			
cis-Aconitic acid	35	9.7	24	2			
Isocitric acid	ND	ND	ND	Ν			
2-Oxoglutaric acid	873	249	1,020	24			
Succinic acid	656	608	666	55			
Fumaric acid	349	ND	529	Ν			
Malic acid	2,545	545	4,163	72			
6-Phosphogluconate	6.6	ND	ND	Ν			
Ribulose 5-phosphate	0.2	13	2.1	2			
Ribose 5-phosphate	5.0	5.6	4.4				
Xylulose 5-phosphate	ND	ND	ND	Ν			
Sedoheptulose 7-phosphate	ND	ND	ND	Ν			
Erythrose 4-phosphate	ND	161	ND	N			
Ribose 1-phosphate	ND	ND	ND	N			
ADP-ribose	34	35	22				
PRPP	9.5	16	16	1			
IMP	51	36	36	ć			
Inosine	ND	ND	ND	Ν			
XMP	2.9	3.3	ND	1			
Adenylosuccinic acid	9.2	8.5	13	9			
AMP	523	518	441	29			
ADP	1,841	2,013	2,320	1,65			
ATP	10,636	11,361	12,873	10,51			
NAD^+	1,729	1,839	1,936	1,21			
NADH	110	134	183	17			
$NADP^+$	144	126	130	10			
NADPH	82	87	121	10			

[Table S2] DNA microarray analysis; the relative expression of genes encoding glycolytic enzymes in IMS32 cells at 1 h in the [Glc 5 mM/Pyr (+)], [Glc 5 mM/Pyr (-)], [Glc 15 mM/Pyr (+)] and [Glc 15 mM/Pyr (-)].

D:			not			detecte
Gene	Protein	5 mM glucose pyruvate+	5 mM glucose pyruvate-	15 mM glucose pyruvate+	15 mM glucose pyruvate-	15 mM glucose pyruvate /15 mM glucose pyruvate
Hk1	Hexokinase 1	55.72	53.28	56.08	52.79	0.9
Hk2	Hexokinase 2	1.08	1.05	1.06	0.95	0.9
Hk3	Hexokinase 3	0.01	0.02	0.01	0.01	0.8
G6pd2	Glucose-6-phosphate dehydrogenase 2	0.10	0.09	0.10	0.12	1.2
Pfkfb1	6-phosphofructo-2- kinase/fructose-2,6-	0.59	0.58	0.58	0.45	
Pfkfb2	bisphosphatase 1 6-phosphofructo-2- kinase/fructose-2,6-	0.15	0.14	0.13	0.13	0.7
Pfkfb3	bisphosphatase 2 6-phosphofructo-2-	0.98	1.09	0.94	1.11	0.9
Pfkfb4	kinase/fructose-2,6- bisphosphatase 3 6-phosphofructo-2-	1.87	1.97	2.11	1.91	1.
191901	kinase/fructose-2,6- bisphosphatase 4	1.07	1.57	2.11	1.91	0.
Aldoa	Aldolase A	96.94	97.41	106.91	112.99	1.
Aldob	Aldolase B	ND	ND	ND	ND	Ν
Aldoc	Aldolase C	0.38	0.40	0.42	0.58	1.
Tpil	Triosephosphate Isomerase 1	29.91	31.75	32.05	34.35	1.
Gapdh	Glyceraldehyde 3-phosphate dehydrogenase	102.74	110.52	109.72	106.62	0.
Pgkl	Phosphoglycerate kinase 1	43.40	42.99	42.56	46.64	1.
Pgk2	Phosphoglycerate kinase 2	0.02	0.02	0.02	0.02	1.
Pgaml	Phosphoglycerate mutase 1	57.52	58.11	59.24	60.45	1.
Pgam2	Phosphoglycerate mutase 2	0.05	0.04	0.04	0.05	1.
Pgam5	Phosphoglycerate mutase 5	8.77	8.89	9.06	10.53	1.
Enol	Enolase 1	61.53	62.80	62.73	62.85	1.
Enolb	Enolase 1b	58.90	59.31	62.26	60.43	0.
Eno2	Enolase 2	1.66	1.70	1.49	1.70	1.
Eno3	Enolase 3	3.05	3.34	3.31	3.63	1.
Eno4	Enolase 4	0.03	0.02	0.03	0.03	1.
Pklr	Pyruvate kinase isotype L/R	0.03	0.02	0.02	0.02	1.
Pkm	Pyruvate kinase	31.59	32.83	34.37	34.67	1.

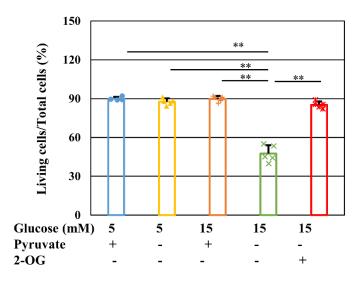
[Table S3] DNA microarray analysis; the relative expression of genes encoding enzymes of collateral glucose-utilising pathways in IMS32 cells at 1 h in the [Glc 5 mM/Pyr (+)], [Glc 5 mM/Pyr (-)], [Glc 15 mM/Pyr (+)] and [Glc 15 mM/Pyr (-)].

Gene P	Protein	5 mM glucose pyruvate+	5 mM glucose pyruvate-	15 mM glucose pyruvate+	15 mM glucose pyruvate-	15 mM glucose pyruvate- /15 mM glucose pyruvate+
Tkt	Transketolase	32.49	33.64	34.86	32.96	0.95
Gfpt1	Glutamine-fructose-6- phosphate aminotransferase 1	2.91	2.80	2.73	2.48	0.91
Gfpt2	Glutamine-fructose-6- phosphate aminotransferase 2	0.06	0.07	0.07	0.08	1.09
Akr1b3	Aldose reductase	24.42	24.82	25.73	27.06	1.05
Sord	Sorbitol dehydrogenase	4.84	5.25	5.28	5.77	1.09

[Table S4] DNA microarray analysis; the relative expression of genes encoding enzymes of TCA cycle in IMS32 cells at 1 h in the [Glc 5 mM/Pyr (+)], [Glc 5 mM/Pyr (-)], [Glc 15 mM/Pyr (+)] and [Glc 15 mM/Pyr (-)].

Gene		5 mM glucose pyruvate+	5 mM glucose pyruvate-	-	15 mM glucose pyruvate-	15 mM glucose pyruvate- /15 mM glucose pyruvate+
Cs		1	54.28			
Acol	Aconitase 1	0.07	0.08	0.06	0.07	
Aco2	Aconitase 2	12.19	12.30	12.85	12.19	
Idh1	Isocitrate dehydrogenase (NADP(+)) 1	0.33	0.38	0.37	0.42	1.14
Idh2	Isocitrate dehydrogenase (NADP(+)) 2	29.15	29.20	31.37	31.34	1.00
Idh3a	Isocitrate Dehydrogenase (NAD(+)) 3 Catalytic Subunit Alpha		8.60	8.81	9.63	
Idh3b	Isocitrate Dehydrogenase (NAD(+)) 3 Non-Catalytic Subunit Beta		4.29	4.53	4.74	
Idh3g		19.99	19.00	19.79	22.13	
Oadh	Oxoglutarate dehydrogenase	9.65	10.01	9.72	8.85	1.12
Sucla2	· · ·	4.34				0.91
Suclg1	0	6.49	6.28	6.77	7.18	1.07
Suclg2	Succinate-CoA Ligase GDP-Forming Subunit Beta		0.63	0.68	0.67	
Sdha	Succinate Dehydrogenase Complex Flavoprotein Subunit A		8.99	9.35	9.48	
Sdhb	Succinate Dehydrogenase Complex Iron Sulfur Subunit B		34.23	37.53	38.31	
Sdhc		9.48	9.58	9.84	10.61	1.02
Sdhd	-		16.31	18.79	20.84	
Fh1	•	12.58	12.35	13.68	13.47	1.11 0.98
Mdh1	2	17.24				017 0
Mdh1b	, ,	0.13				1.05
Mdh2		77.00				1.00

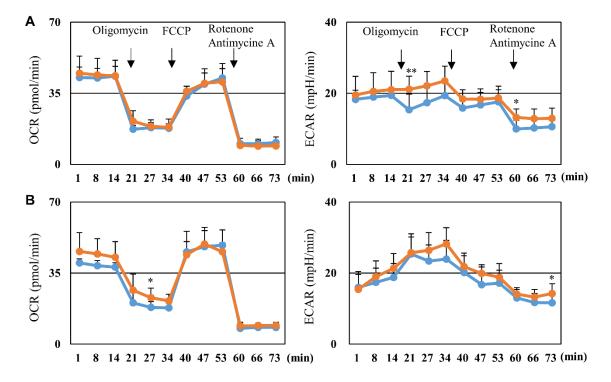
Fig. S1



[Figure S1] 2-OG prevents DRG neuronal death under high-glucose pyruvate-starved conditions

Cell viability at 6 h in the [Glc 5 mM/Pyr (+)] (blue), [Glc 5 mM/Pyr (-)] (yellow), [Glc 15 mM/Pyr (+)] (brown), [Glc 15 mM/Pyr (-)] (green), and [Glc 15 mM/Pyr (-)/2-OG (+)] (red) groups was determined by trypan blue staining. Data represent mean + SD from 4-5 values from independent culture (individual values are depicted as circles, triangles, pluses, crosses and asterisks). Statistical analysis of the data was performed by one-way ANOVA followed by post hoc comparisons with Tukey HSD test. ** P<0.01.

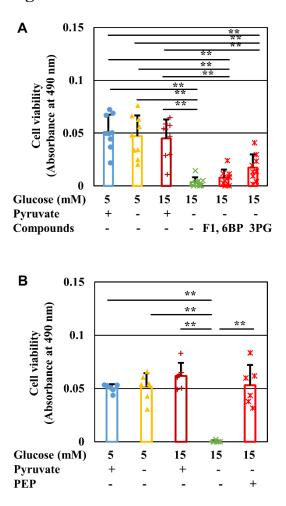




[Figure S2] The effects of 2-OG supplementation on OCR and ECAR under normal- and highglucose conditions in the presence of pyruvate

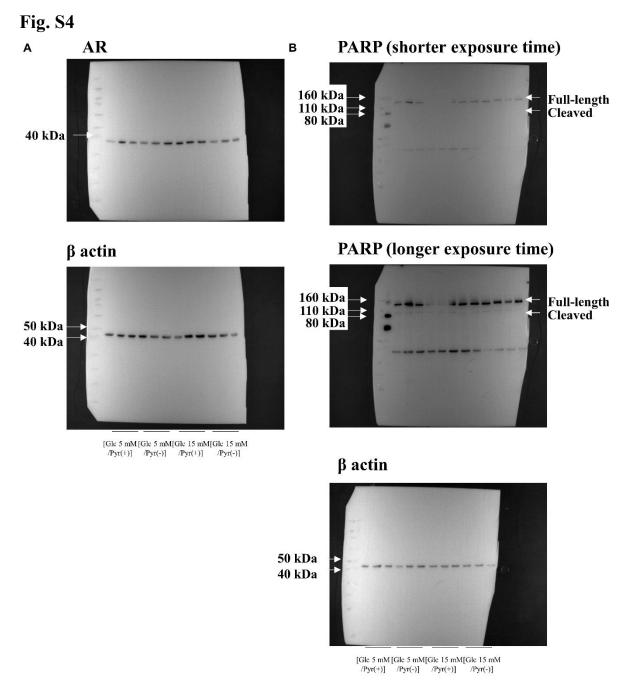
Time courses of OCR (left) and ECAR (right) in the [Glc 5 mM/Pyr (+)/2-OG(-)] (A, blue), [Glc 5 mM/Pyr(+)/2-OG(+)] (A, orange), [Glc 100 mM/Pyr (+)/2-OG(-)] (B, blue), and [Glc 100 mM/Pyr (+)/2-OG(+)](B, orange) groups measured by Extracellular Flux Analyzer. Data represent mean + SD from 9-10 values from two independent cultures. Statistical analysis of the data was performed Mann-Whitney test. * P<0.05, ** P<0.01.

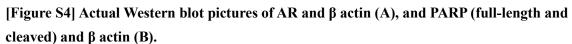
Fig. S3



[Figure S3] 3PG and PEP, but not F1,6BP, prevents IMS32 cell death under high-glucose pyruvate-starved conditions

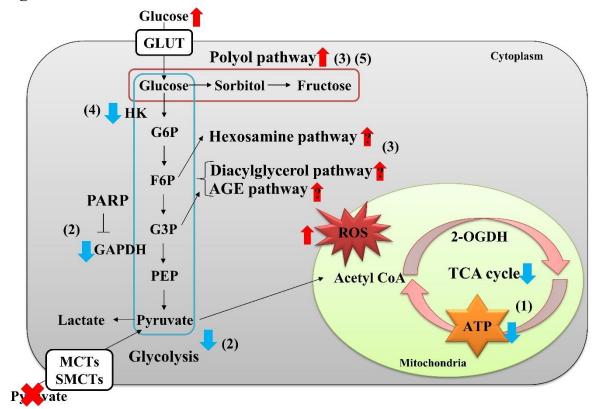
Cell viability at 24 h in the [Glc 5 mM/Pyr (+)] (blue), [Glc 5 mM/Pyr (-)] (yellow), [Glc 15 mM/Pyr (+)] (brown), [Glc 15 mM/Pyr (-)] (green), and [Glc 15 mM/Pyr (-)] supplemented with (A) F1,6BP, 3-phosphogryceric acid (3-PG) or (B) phosphoenolpyruvate (PEP) (red) groups was determined by MTS assay. Data represent mean + SD from six (A) and nine (B) values from two or three independent cultures (individual values are depicted as circles, triangles, pluses, crosses and asterisks). ** P<0.01.

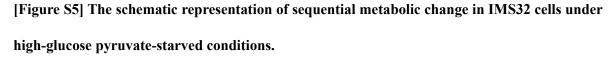




Full length of the membranes presented in Fig. 6C (A) and Fig. 8A (B). AR and β actin (A), and PARP in shorter and longer exposure time and β actin (B) were sequentially visualized in same membranes.







Rapid IMS32 cell death under high-glucose pyruvate-starved conditions can be caused by the sequential metabolic changes as follows; 1) reduced TCA cycle intermediates and mitochondrial ATP production, 2) impaired GAPDH activity and inhibition of glycolytic flux, 3) augmentation of the polyol and other collateral glycolysis pathways, 4) decreased hexokinase (HK) activity and glucose utilization in the glycolytic pathway, and 5) further enhancement of the polyol pathway flux. The schematic image was created by H. Y.