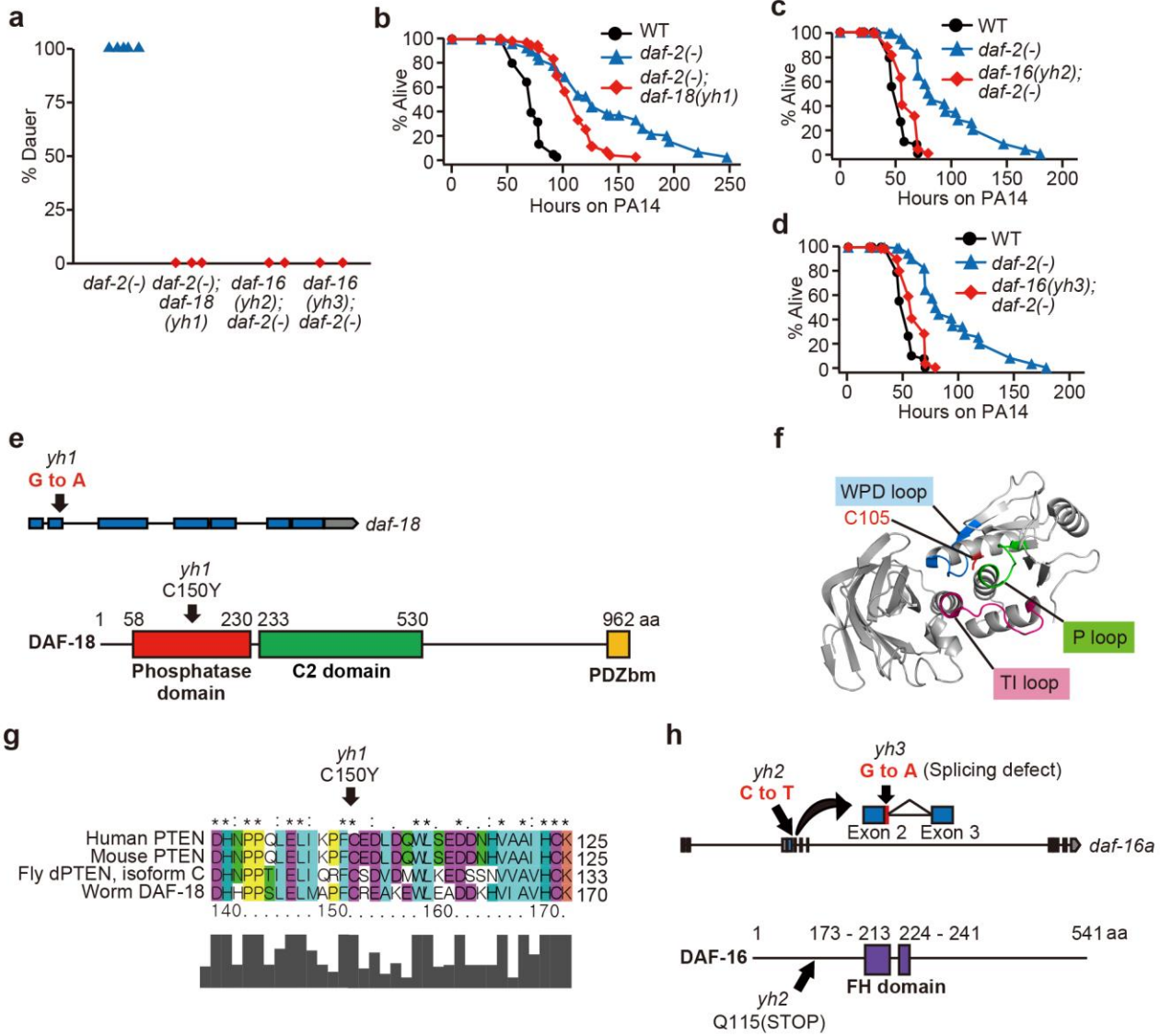


1 Supporting Information

2 Supplementary Figures

Supplementary Figure 1



3 Supplementary Fig. 1: Identification of mutations that suppress dauer phenotypes

4 with small effects on immunity in *daf-2(e1370)* mutants. (a) Percentage of 25°C

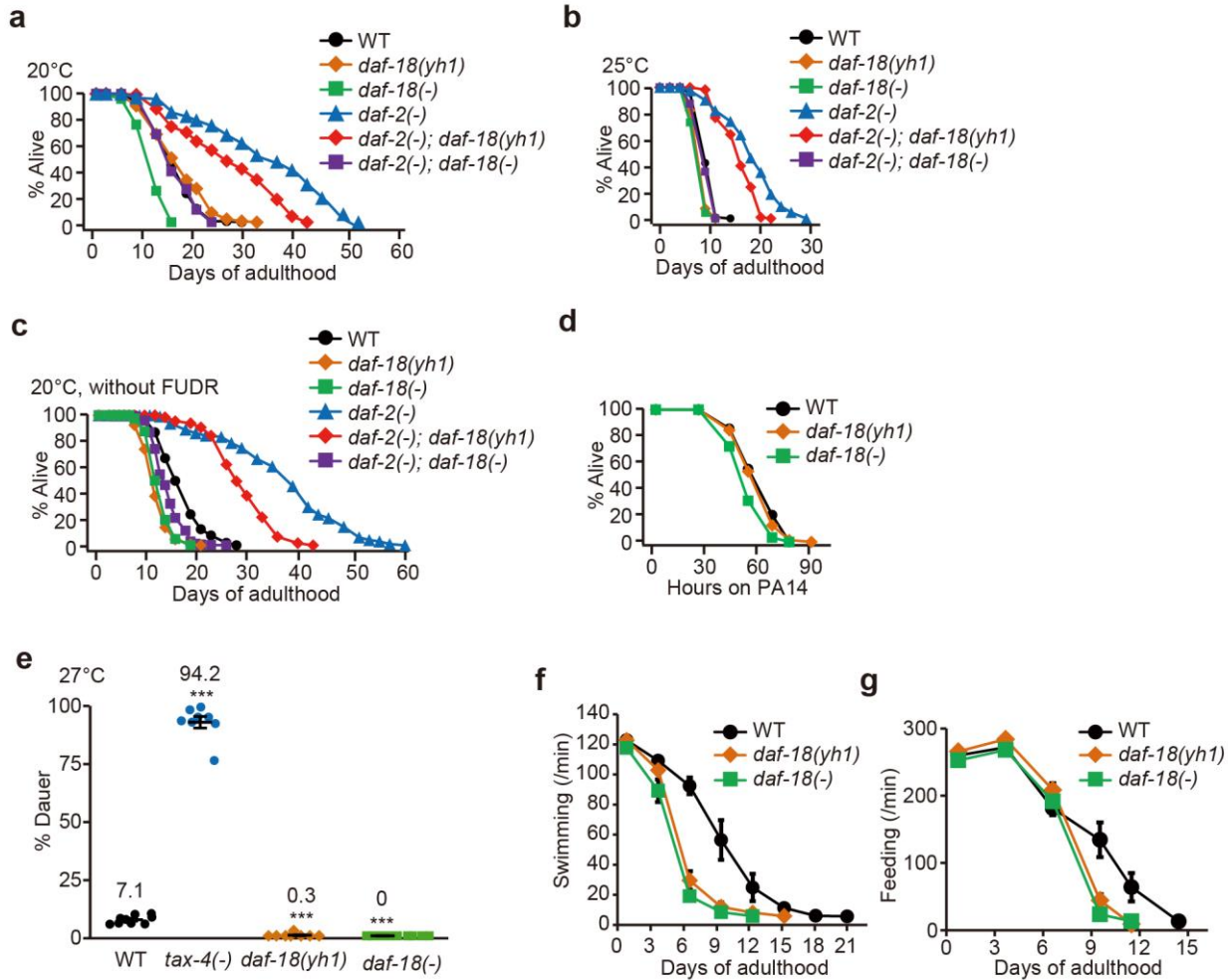
5 dauer formation of *daf-2(e1370); daf-18(yh1)* [*daf-2(-); daf-18(yh1)*], *daf-16(yh2); daf-*

6 *2(-)*, and *daf-16(yh3); daf-2(-)* animals, which were identified from our mutagenesis

7 screen, and *daf-2(-)* animals that were used as a control ($n \geq 267$ for each condition, the
8 assays were performed at least twice). **(b-d)** Survival curves of *daf-2(-); daf-18(yh1)* **(b)**,
9 *daf-16(yh2); daf-2(-)* **(c)**, and *daf-16(yh3); daf-2(-)* **(d)** mutants on PA14 compared with
10 *daf-2(-)* mutant and wild-type (WT) worms ($n = 180$ for each condition). **(e-h)** Molecular
11 natures of *yh1*, *yh2*, and *yh3* alleles identified from our mutagenesis screen (See
12 Supplementary Table 1 for details). **(e)** The locus of the *yh1* allele in *daf-18* and the
13 corresponding change in *C. elegans* DAF-18/PTEN protein. *yh1* mutation changes G to
14 A at the 2nd exon of *daf-18*, which causes the cysteine to tyrosine change at the location
15 of 150th amino acid in DAF-18/PTEN. Amino acid (aa) numbers are indicated at the top
16 of the protein domains. Phosphatase domain (58 aa – 230 aa), C2 domain (233 aa –
17 530 aa), and PDZ-binding motif (PDZbm) are indicated as boxes. **(f)** Three-dimensional
18 structure of human PTEN¹. C105 resides in a region between two catalytic loops, WPD
19 loop and a highly-conserved core P loop. **(g)** Alignment of amino acid sequences of
20 human PTEN, mouse PTEN, *Drosophila* (fly) dPTEN isoform C, and *C. elegans* (worm)
21 DAF-18. A gray histogram represents the conservation of the sequences. **(h)** The loci of
22 *yh2* and *yh3* alleles in *daf-16* isoform a [*daf-16a*] and the corresponding changes in *C.*
23 *elegans* DAF-16/FOXO protein. *yh2* causes a nonsense mutation by a C to T transition
24 at the 2nd exon of *daf-16a*, resulting in premature termination at the 115th glutamine. *yh3*
25 causes a G to A transition at the 5' splicing site of intron 2, resulting in a predicted pre-
26 mRNA splicing defect, leading to the acquisition of a premature stop codon. The
27 structural analysis showed that *daf-16(yh2)* and *daf-16(yh3)* mutations may prevent the
28 translation of the forkhead (FH) domain (173 aa - 213 aa and 224 aa - 241 aa) of DAF-
29 16/FOXO due to generation of premature stop codons. See Supplementary Dataset 2

30 and 3 for additional repeats and statistical analysis for the survival and the dauer assay
31 data shown in this figure. See also Source Data for data points used for the derivation of
32 data.

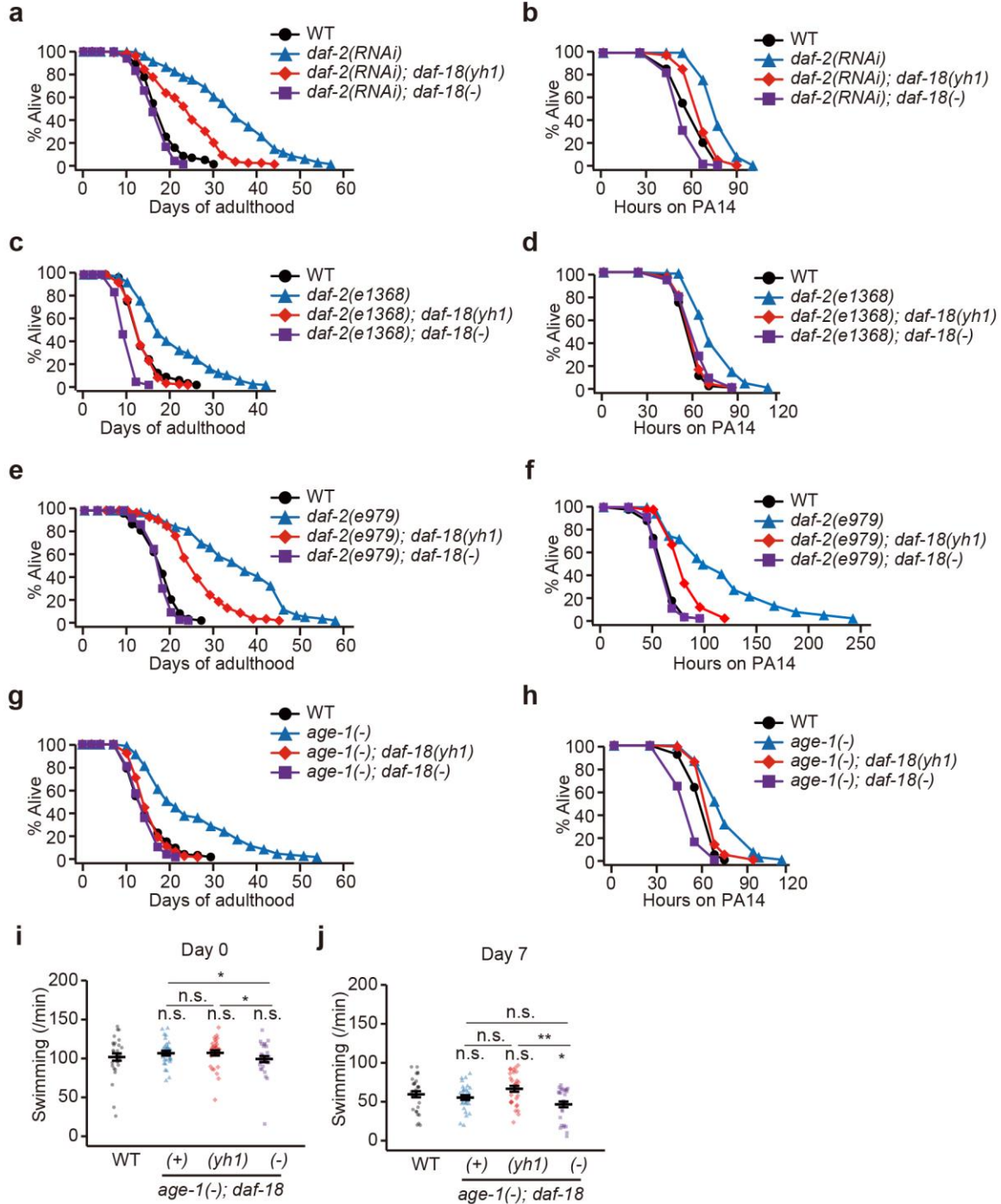
Supplementary Figure 2



33 **Supplementary Fig. 2: *daf-18(yh1)* decreases pathogen resistance, lifespan, and**
 34 **health span in *daf-2(-)* and wild-type animals. (a-c)** The effects of *daf-18(yh1)* and
 35 *daf-18(nr2037)* [*daf-18(-)*] on the lifespan of wild-type (WT) and *daf-2(e1370)* [*daf-2(-)*]
 36 animals at 20°C (**a**), at 25°C (**b**), and at 20°C without FUDR treatment (**c**) (n ≥ 225 for
 37 each condition). *daf-18(yh1)* substantially decreased the lifespan of WT animals with an
 38 extent similar to that caused by *daf-18(-)* at 20°C (four out of six trials). The lifespan
 39 curves of WT, *daf-2(-)*, *daf-2(-); daf-18(yh1)*, and *daf-2(-); daf-18(-)* shown in panel **a** are
 40 the same experimental sets shown in Fig. 1i, and those in **c** are the same experimental
 41 sets shown in Supplementary Fig. 10b and Fig. 6b. (**d-g**) The effects of *daf-18(yh1)* and

42 *daf-18(-)* on pathogen resistance (n = 180 for each condition) (**d**), dauer formation at
43 27°C (n ≥ 790 for each condition, from at least eight trials) (**e**), swimming span (n = 10
44 for each condition, from one trial) (**f**), and feeding span (n = 10 for each condition, from
45 one trial) (**g**). The survival of *daf-18(yh1)* animals under PA14 infection was similar to
46 that of WT worms, whereas *daf-18(-)* worms displayed reduced survival (**d**). Both *daf-*
47 *18(yh1)* (0.3%) and *daf-18(-)* (0%) mutations substantially suppressed dauer
48 phenotypes at 27°C; WT animals displayed 7.1% of dauers at 27°C, and *tax-4(p678)*
49 [*tax-4(-)*] animals, which display a constitutive dauer phenotype at 27°C², was used as a
50 positive control (**e**). In addition, *daf-18(yh1)* and *daf-18(-)* mutations similarly reduced
51 the swimming (motility) and feeding spans (**f,g**). Error bars indicate the standard error of
52 mean (s.e.m., ****p* < 0.001, two-tailed Student's *t*-test relative to WT). See
53 Supplementary Dataset 2, 3, and 4 for additional repeats and statistical analysis for the
54 survival, dauer, and health span assay data shown in this figure. See also Source Data
55 for data points used for the derivation of data.

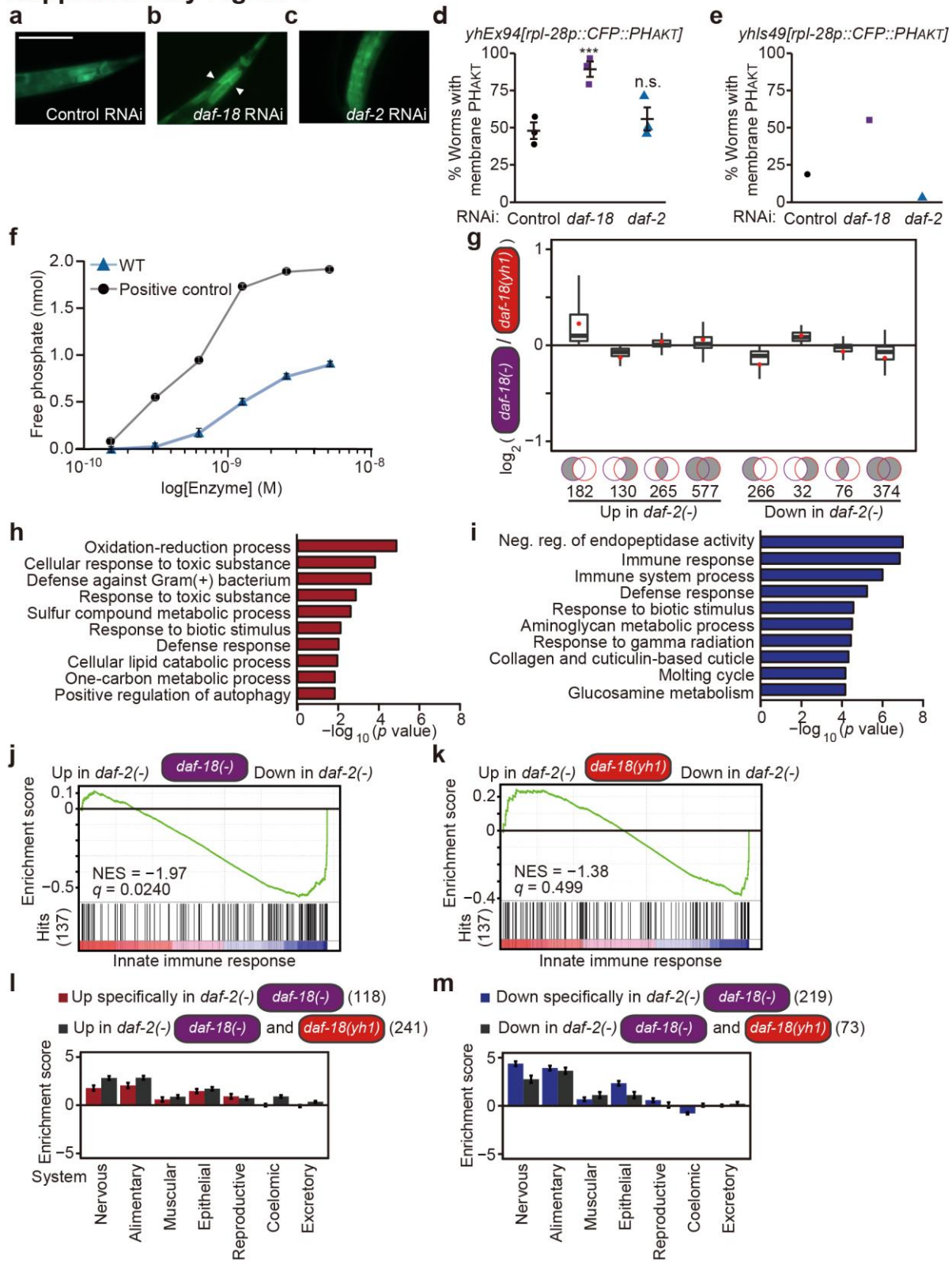
Supplementary Figure 3



56 **Supplementary Fig. 3: *daf-18(yh1)* retains lifespan and pathogen resistance in**
 57 **worms with reduced IIS in multiple conditions. (a-h) The effects of *daf-18(yh1)* and**
 58 ***daf-18(nr2037)* [*daf-18(-)*] on lifespan at 20°C (n ≥ 270 for each condition; n = 156 for**

59 *daf-2(e979)* animals) and pathogen resistance (n = 180 for each condition; n = 35 for
60 *daf-2(e979)* animals) of *daf-2(RNAi)* (a,b), *daf-2(e1368)* (c,d), *daf-2(e979)* (e,f), and
61 *age-1(hx546)* [*age-1(-)*] (g,h) animals. (i,j) Swimming rate (n = 30 for each condition,
62 body bends per minute in liquid measured from three independent trials) of day 0 (i) and
63 day 7 adult (j) wild-type (WT), *age-1(-); daf-18(+)*, *age-1(-); daf-18(yh1)*, and *age-1(-);*
64 *daf-18(-)* animals. Error bars indicate the standard error of mean (s.e.m., **p* < 0.05, ***p*
65 < 0.01, n.s.: not significant, two-tailed Student's *t*-test relative to WT unless otherwise
66 noted). See Supplementary Dataset 2 and 4 for additional repeats and statistical
67 analysis for the survival assay and health span assay data shown in this figure. See
68 also Source Data for data points used for the derivation of data.

Supplementary Figure 4

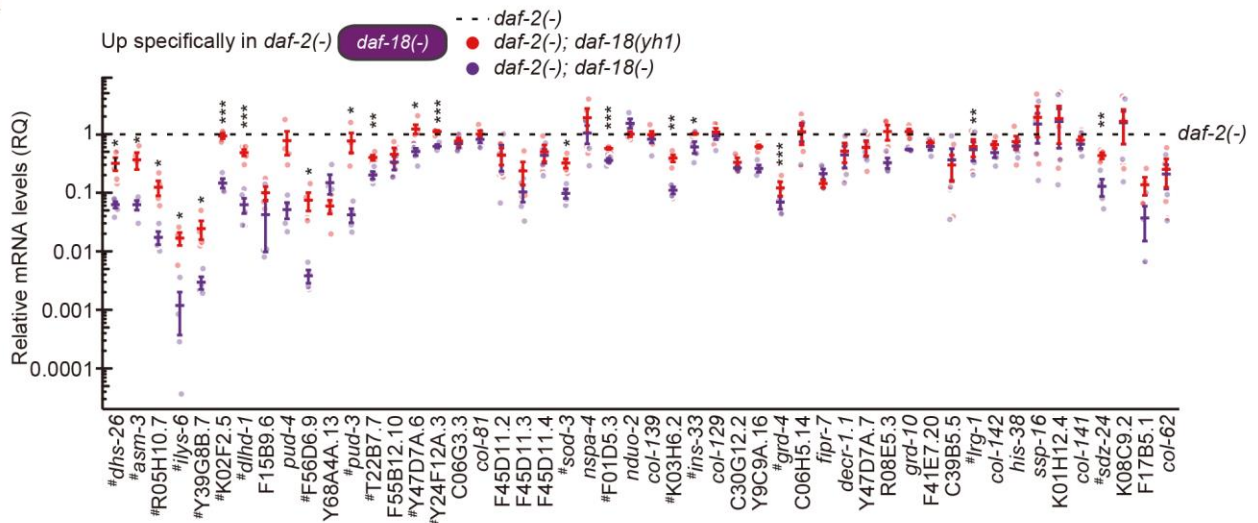


70 **on phosphatase activity and various downstream targets in reduced IIS**
71 **conditions. (a-c)** Subcellular localization of mouse PH_{AKT} for monitoring PIP₃ levels in
72 *C. elegans*. Representative images of *rpl-28p::CFP::PH_{AKT}*-expressing animals treated
73 with control RNAi **(a)**, *daf-18* RNAi **(b)**, or *daf-2* RNAi **(c)**. *rpl-28p* is a ubiquitous *rpl-28*
74 (ribosomal protein large subunit 28) gene promoter. Worms expressing *CFP::PH_{AKT}* as
75 an extrachromosomal array, *yhEx94[rpl-28p::CFP::PH_{AKT}, odr-1p::RFP]*. Scale bar: 50
76 μ m. Arrowhead: membrane CFP::PH_{AKT}. **(d)** Quantification of panels **a-c** ($n > 10$ for
77 each condition, from three independent trials). Error bars indicate the standard error of
78 mean (s.e.m., *** $p < 0.001$, n.s.: not significant, two-tailed Student's *t*-test relative to
79 WT). **(e)** Quantification of fluorescent protein levels of integrated CFP::PH_{AKT} worms,
80 *yhls49[rpl-28p::CFP::PH_{AKT}; odr-1p::RFP]* ($n > 10$ for each condition, from one trial). **(f)**
81 Dose-dependent changes in the protein phosphatase activity of human recombinant
82 PTEN (WT) and protein tyrosine phosphatase 1B (PTP1B, positive control). The
83 indicated amounts of recombinant proteins were tested for the protein phosphatase
84 assay ($N = 3$). Note that the same dose-dependent protein phosphatase activity curve of
85 WT is shown in Fig. 3h as well. **(g)** Overall magnitudes of gene expression changes
86 caused by *daf-18(yh1)* were smaller than those by *daf-18(nr2037) [daf-18(-)]* in *daf-*
87 *2(e1370) [daf-2(-)]* mutants. Venn diagrams with gray area display overlaps between
88 differentially expressed genes. Box plots represent five-number summaries of
89 expression changes of genes that are included in the gray areas in the Venn diagrams.
90 Red dots in the box plots display mean expression changes. **(h,i)** Overrepresented
91 gene ontology (GO) terms of the genes that were upregulated **(h)** and downregulated **(i)**
92 in *daf-2(-)* mutants compared specifically with *daf-2(-); daf-18(-)* animals but not with

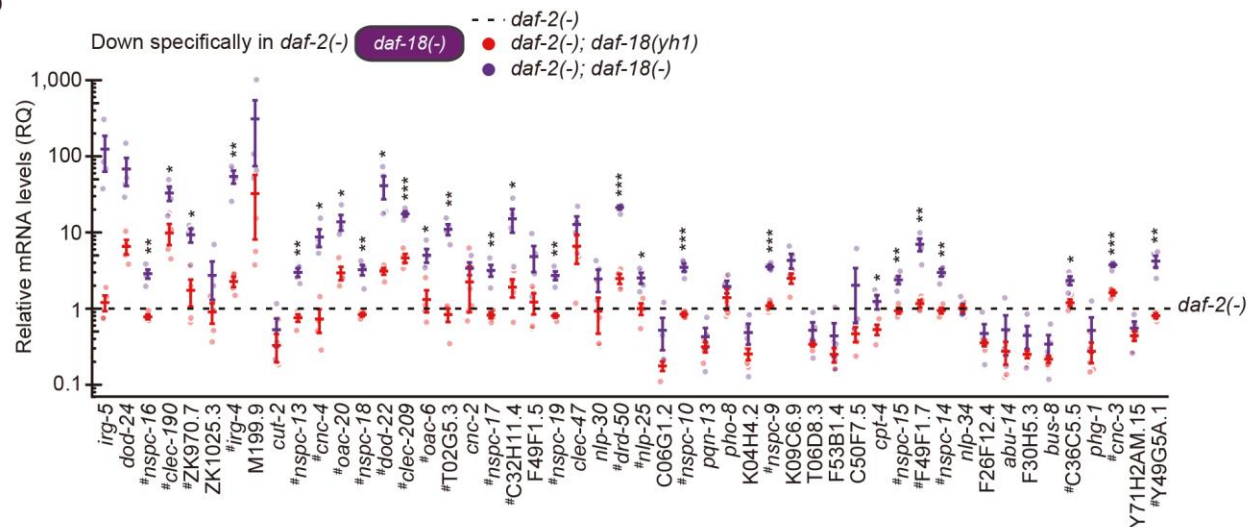
93 *daf-2(-); daf-18(yh1)* worms. *p* values were calculated by using hypergeometric test.
94 **(j,k)** Innate immune response genes (curated as a GO term) were significantly induced
95 by *daf-18(-)* **(j)**, but not by *daf-18(yh1)* **(k)**, in the *daf-2(-)* mutant background. NES:
96 normalized enrichment score. *q* values were obtained by calculating the false discovery
97 rate corresponding to each NES. **(l)** Tissue enrichment analysis of genes upregulated in
98 *daf-2(-)* animals compared specifically with *daf-2(-); daf-18(-)* animals but not with *daf-*
99 *2(-); daf-18(yh1)* animals, or generally compared with both *daf-2(-); daf-18(-)* and *daf-*
100 *2(-); daf-18(yh1)* animals. **(m)** Tissue enrichment analysis of genes downregulated in
101 *daf-2(-)* animals compared specifically with *daf-2(-); daf-18(-)* animals but not with *daf-*
102 *2(-); daf-18(yh1)* animals, or generally compared with both *daf-2(-); daf-18(-)* and *daf-*
103 *2(-); daf-18(yh1)* animals. See Source Data for data points used for the derivation of
104 data.

Supplementary Figure 5

a



b

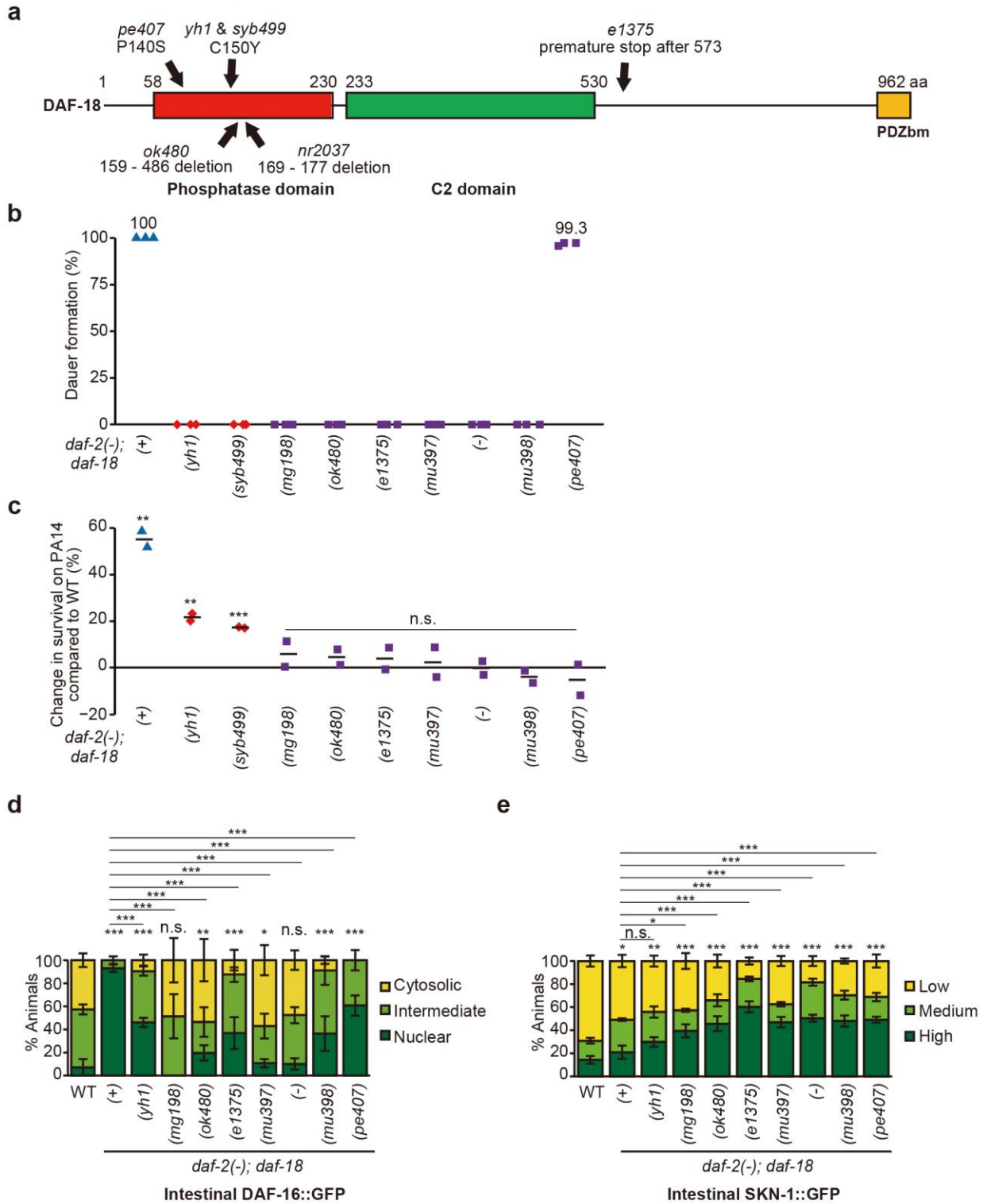


105 **Supplementary Fig. 5: Confirmation of genes whose expression was greatly**
 106 **affected by *daf-18(-)* compared with that by *daf-18(yh1)* in *daf-2(-)* mutants.**

107 Relative mRNA levels of selected genes whose expression was highly affected by *daf-*
 108 *18(nr2037)* [*daf-18(-)*] compared with that by *daf-18(yh1)* in *daf-2(e1370)* [*daf-2(-)*] using
 109 quantitative RT-PCR (N ≥ 4). (a) Among the top-ranked 50 genes that were upregulated
 110 in *daf-2(-)* animals compared with *daf-2(-); daf-18(-)* (fold change > 2, Benjamini and
 111 Hochberg (BH)-adjusted *p* value < 0.05), but marginally with *daf-2(-); daf-18(yh1)* worms

112 (fold change < 2), 19 genes (marked as #) displayed the same tendency as RNA-seq
113 data by using qRT-PCR. **(b)** Among the 50 genes that were robustly downregulated in
114 *daf-2(-)* animals compared with *daf-2(-); daf-18(-)* worms (fold change > 2, BH-adjusted
115 *p* value < 0.05) but slightly with *daf-2(-); daf-18(yh1)* animals (fold change < 2), 25
116 genes (marked as #) displayed the same tendency as RNA-seq data by using qRT-PCR.
117 #: *daf-2(-)* vs. *daf-2(-); daf-18(-)* and *daf-2(-); daf-18(yh1)* vs. *daf-2(-); daf-18(-)*, *p* < 0.05.
118 RQ: relative quantity. Black dotted lines indicate the RQ value of *daf-2(-)* animals set as
119 1. Error bars indicate the standard error of mean (s.e.m., **p* < 0.05, ***p* < 0.01, ****p* <
120 0.001, two-tailed Student's *t*-test). See Supplementary Dataset 5 for the details of
121 primer sequences. See also Source Data for data points used for the derivation of data.

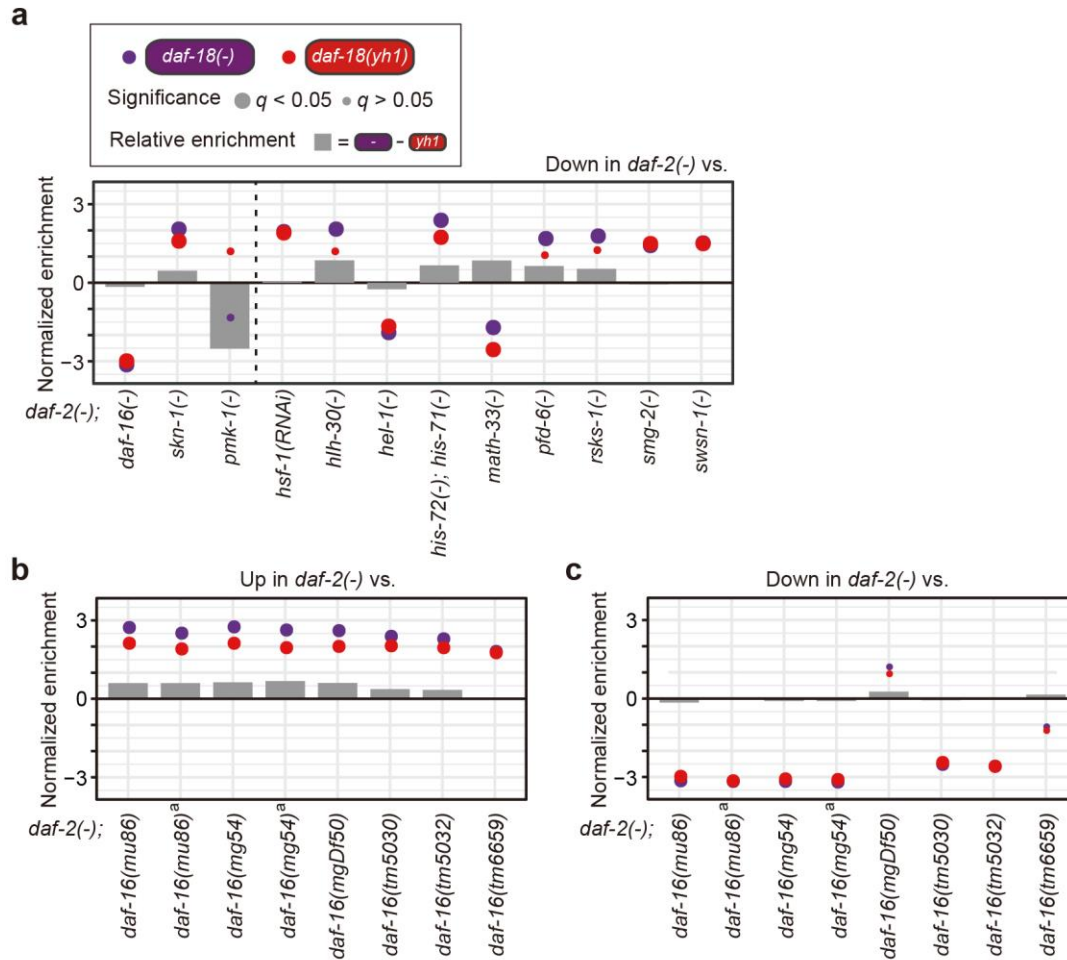
Supplementary Figure 6



123 **formation, immunity, and the subcellular localization of DAF-16/FOXO and SKN-**
124 **1/NRF2 in *daf-2(-)* mutants. (a)** Locations of *daf-18* hypomorphs that were used for the
125 experiments in the current work³⁻¹⁰. *pe407* was isolated from the mutagenesis screen as
126 a suppressor that restores the plasticity of salt chemotaxis in *casy-1(tm718)* mutant
127 backgrounds¹⁰. *pe407* causes P140S change in the phosphatase domain of DAF-
128 18/PTEN. *yh1* (this study) and *syb499* (CRISPR knock-in version of *yh1*) results in
129 C150Y change, at the phosphatase domain of DAF-18. Amino acid (aa) numbers are
130 indicated at the top of the protein domains. Phosphatase domain, C2 domain, and PDZ-
131 binding motif (PDZbm) are indicated as boxes. *nr2037* (169 aa – 177 aa deletion)^{3,4} and
132 *ok480* (159 aa – 486 aa deletion)⁷ are deletion alleles of *daf-18*, and *e1375* has a 30 bp
133 insertion that introduces a premature stop codon at 573 aa^{5,8}. **(b,c)** The effects of
134 various *daf-18* mutant alleles on dauer formation at 25°C (n ≥ 287 for each condition,
135 from three to five independent trials; the assays were performed 96 hrs after eggs were
136 placed) **(b)** and the survival of worms on the pathogen (PA14) using a big-lawn assay (n
137 = 180 for each condition, from two independent trials) **(c)** in the *daf-2(-)* background.
138 Wild-type (WT), *daf-2(e1370)* [*daf-2(-)*], *daf-2(-); daf-18(yh1)*, *daf-2(-); daf-18(syb499)*,
139 *daf-2(-); daf-18(mg198)*, *daf-2(-); daf-18(ok480)*, *daf-2(-); daf-18(e1375)*, *daf-2(-); daf-*
140 *18(mu397)*, *daf-2(-); daf-18(nr2037)* [*daf-18(-)*], *daf-2(-); daf-18(mu398)*, and *daf-2(-);*
141 *daf-18(pe407)* animals were examined. Consistent with previous reports^{5,8,11}, *daf-2(-);*
142 *daf-18(e1375)* animals developed slowly but eventually became adults at the time dauer
143 formation was assayed. Horizontal lines in panel **c** represent mean values (***p* < 0.01,
144 ****p* < 0.001, n.s.: not significant, two-tailed Student's *t*-test relative to WT). **(d,e)** The
145 effects of various *daf-18* mutant alleles on the subcellular localization of DAF-16::GFP

146 and SKN-1::GFP in *daf-2(-)* mutants. **(d)** Quantification of the subcellular localization of
147 DAF-16::GFP in the intestines of indicated strains. Cytosolic: predominant cytosolic
148 localization, intermediate: partial nuclear localization, nuclear: predominant nuclear
149 localization ($n \geq 28$ for each condition, from four to seven independent trials). **(e)**
150 Quantification of the subcellular localization of SKN-1::GFP in the intestinal cells of
151 indicated strains. Low: very dim GFP in the nuclei, medium: $< 50\%$ of the nuclei with
152 SKN-1::GFP, high: $> 50\%$ of the nuclei with SKN-1::GFP ($n \geq 144$ for each condition,
153 from four to eight independent trials). The quantification data of DAF-16::GFP
154 subcellular localization in WT, *daf-2(-)*, *daf-2(-); daf-18(yh1)*, and *daf-2(-); daf-18(-)*
155 animals shown in panel **d** are the same experimental sets shown in Fig. 5g, and those
156 in panel **e** are the same experimental sets shown in Fig. 5i. Error bars represent the
157 standard error of mean (s.e.m., $*p < 0.05$, $**p < 0.01$, $***p < 0.001$, n.s.: not significant,
158 Chi-squared test). See Supplementary Dataset 2 and 3 for additional repeats and
159 statistical analysis for the survival and dauer assay data shown in this figure. See also
160 Source Data for data points used for the derivation of data.

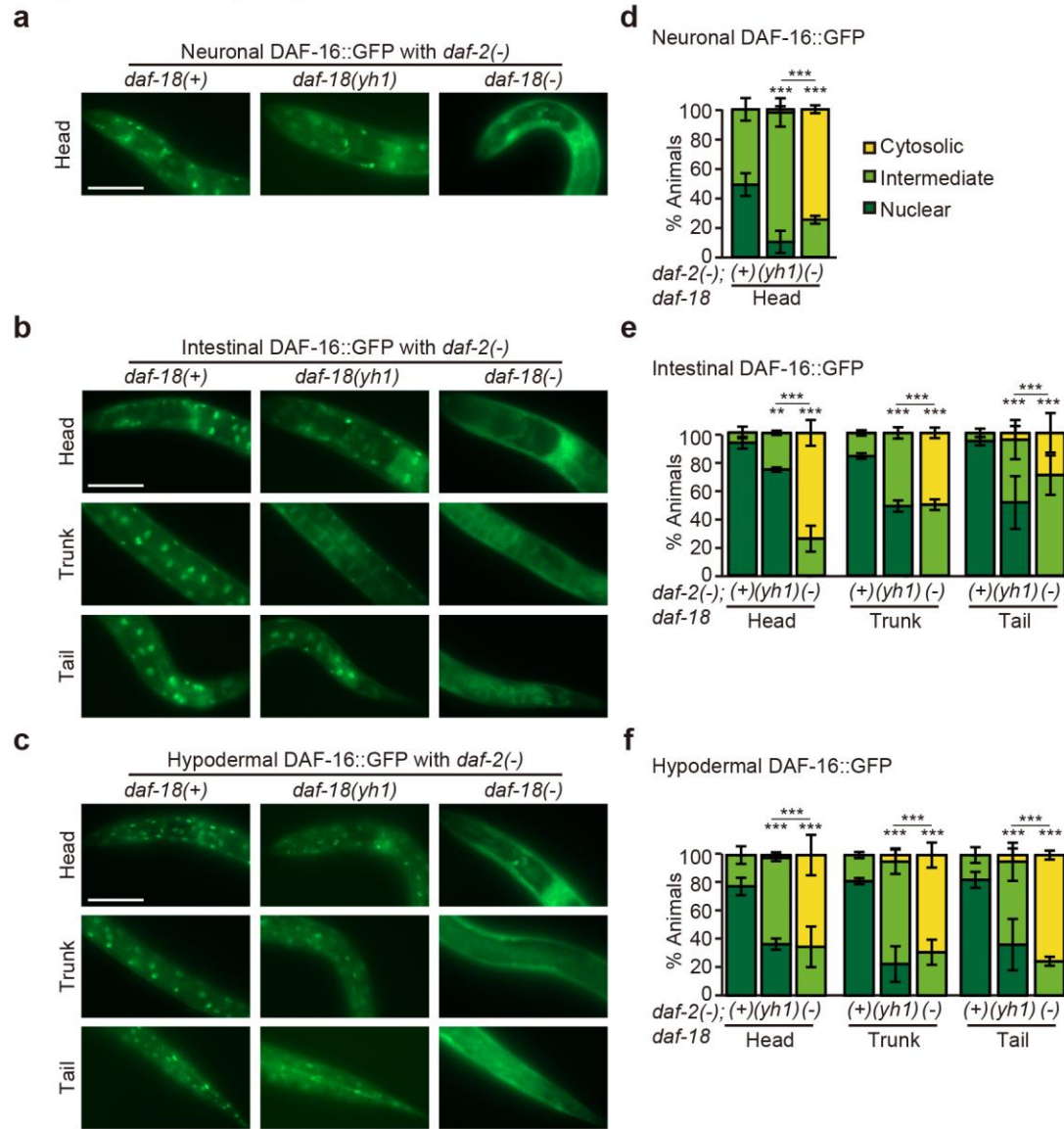
Supplementary Figure 7



161 **Supplementary Fig. 7: Differential effects of *daf-18(yh1)* and *daf-18(-)* on the**
 162 **expression of various target genes in worms with reduced insulin/IGF-1 signaling.**
 163 (a-c) Normalized enrichment of gene expression changes in *daf-2(e1370)* [*daf-2(-)*]
 164 mutants compared to *daf-2(-); daf-18(yh1)* or *daf-2(-); daf-18(nr2037)* [*daf-18(-)*]
 165 mutants. (a) Genes downregulated in *daf-2(-)* animals compared to various lifespan
 166 mutants in the *daf-2(-)* background were used for the analysis. Shown are DAF-
 167 16/FOXO¹², SKN-1/NRF2¹³, PMK-1/p38 MAP kinase¹⁴, HSF-1/heat shock factor 1¹⁵,
 168 HLH-30/TFEB¹⁶, HEL-1/DEAD-box RNA helicase¹⁷, histone H3.3¹⁸, MATH-
 169 33/deubiquitylating enzyme (at 25°C)¹⁹, PFD-6/prefoldin⁶²⁰, RSKS-1/S6K²¹, SMG-

170 2/UPF1²², and SWSN-1/BAF155/170²³ target genes. **(b-c)** Genes upregulated **(b)** and
171 downregulated **(c)** in *daf-2(-)* animals compared to various alleles of *daf-16* mutants in
172 the *daf-2(-)* background were used for the analysis. Shown are data using *daf-*
173 *16(mu86)*¹², *daf-16(mu86)*^a, *daf-16(mg54)*¹², *daf-16(mg54)*^a, *daf-16(mgDf50)*²⁴, *daf-*
174 *16(tm5030)*¹², *daf-16(tm5032)*¹², and *daf-16(tm6659)*¹². Relative enrichment indicates
175 the difference of gene expression changes caused by *daf-18(yh1)* or *daf-18(-)*. *q* values
176 were obtained by calculating the false discovery rate corresponding to each normalized
177 enrichment. ^a: GSE111338 in GEO. See Supplementary Table 2 for details.

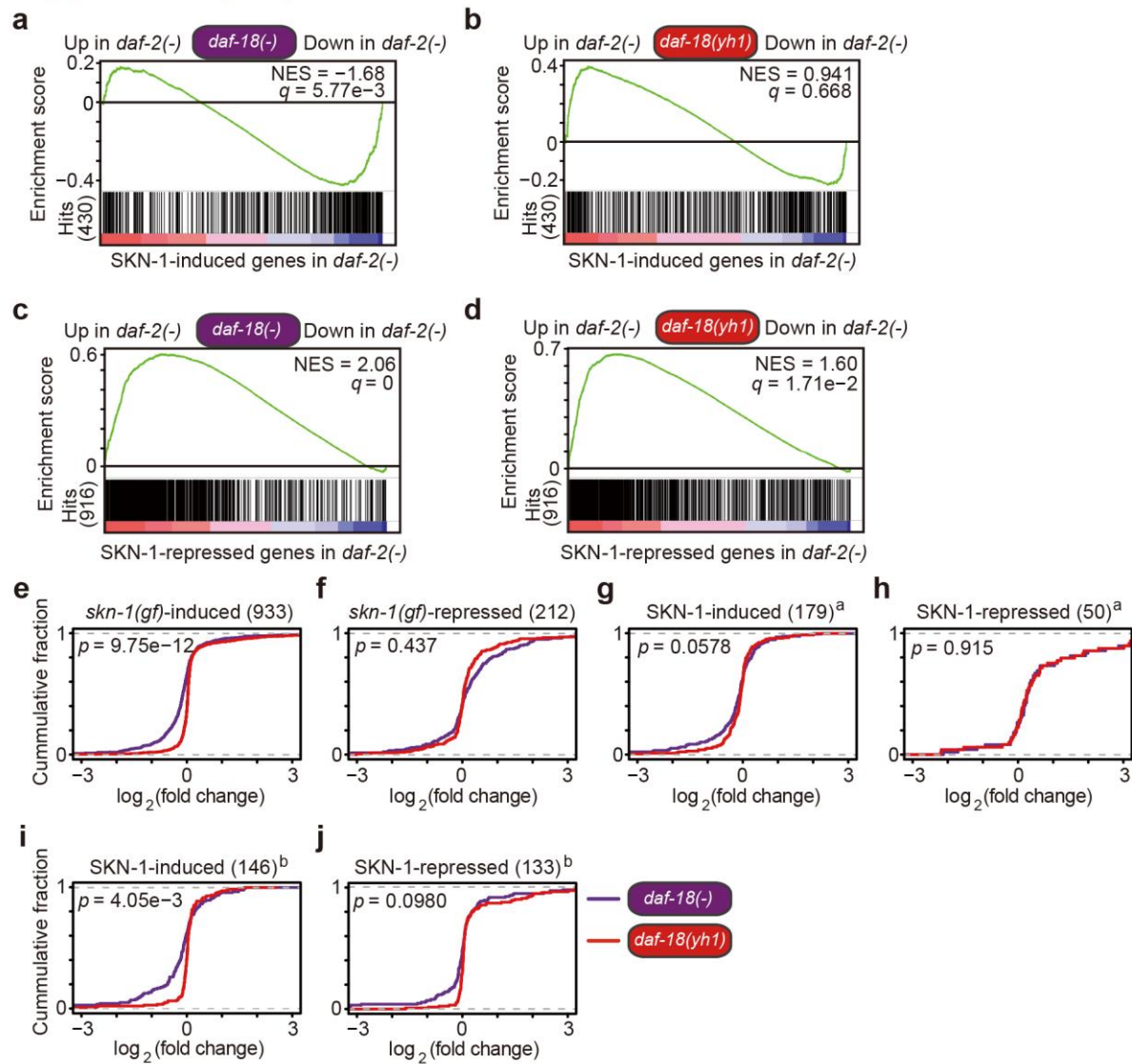
Supplementary Figure 8



178 **Supplementary Fig. 8: *daf-18(yh1)* partially retains the nuclear localization of DAF-**
 179 **16::GFP in various tissues of *daf-2(-)* mutants. (a-c)** Representative images of the
 180 neuronal (a), intestinal (b), and hypodermal (c) DAF-16::GFP localization in head, trunk,
 181 and tail regions of *daf-2(e1370)* [*daf-2(-)*], *daf-2(-); daf-18(yh1)*, and *daf-2(-); daf-*
 182 *18(nr2037)* [*daf-18(-)*] animals. Scale bar: 50 μ m. (d-f) Quantification of data shown on
 183 panels a-c. The subcellular localization of DAF-16::GFP was scored as follows. Nuclear:

184 predominant nuclear localization, intermediate: partial nuclear localization, cytosolic:
185 predominant cytosolic localization ($n \geq 31$ for each condition, from three independent
186 trials). Error bars represent the standard error of mean (s.e.m., $**p < 0.01$, $***p < 0.001$,
187 Chi-squared test relative to WT unless otherwise noted). See also Source Data for data
188 points used for the derivation of data.

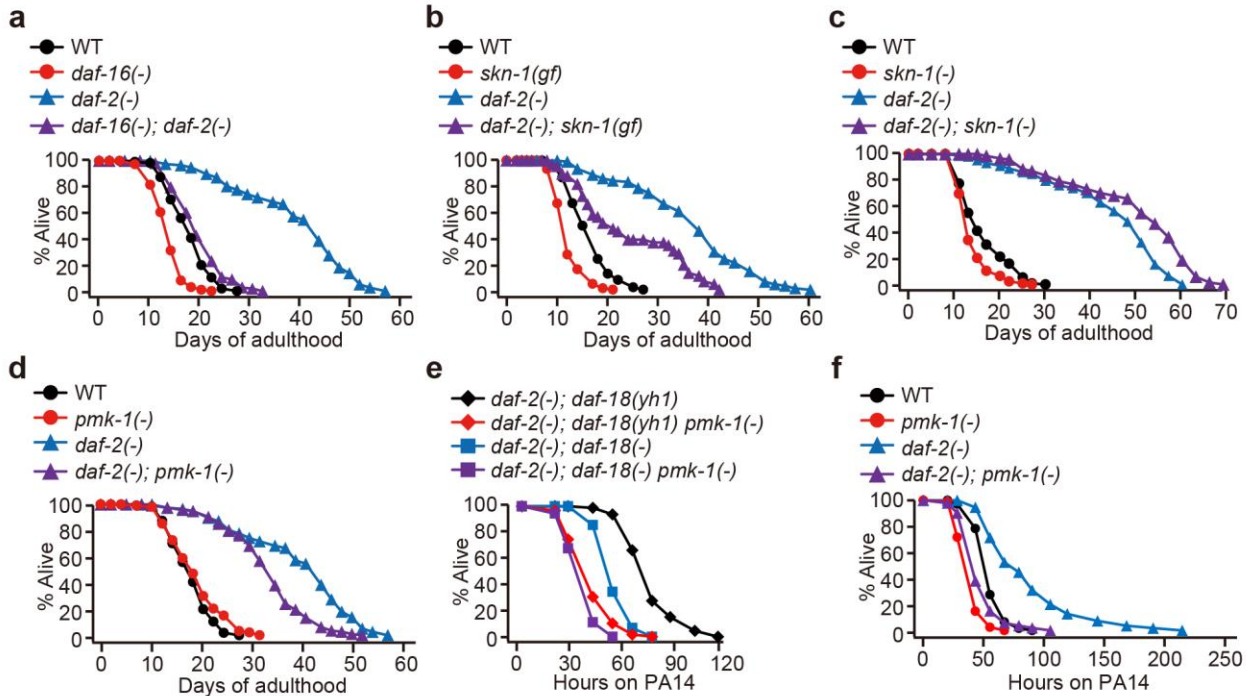
Supplementary Figure 9



189 **Supplementary Fig. 9: DAF-18/PTEN downregulates SKN-1/NRF2 in animals with**
 190 **reduced insulin/IGF-1 signaling. (a-d)** The extent of gene expression changes
 191 conferred by *daf-18(yh1)* and *daf-18(nr2037)* [*daf-18(-)*] in *daf-2(e1370)* [*daf-2(-)*]
 192 animals. Genes whose expression was upregulated (**a,b**) and downregulated (**c,d**) in
 193 *daf-2(e1370)* [*daf-2(-)*] mutants compared to *daf-2(-)*; *skn-1(zu67)* mutants are shown.
 194 Data using *daf-2(e1368)* and *daf-2(-)* mutants¹³ were pooled. NES: normalized
 195 enrichment score. q values were obtained by calculating the false discovery rate

196 corresponding to each NES. **(e-j)** Cumulative fraction of genes in an ascending order of
197 the extent of gene expression changes conferred by *daf-18(yh1)* and *daf-18(-)* in wild-
198 type animals. **(e,f)** Shown are genes whose expression was upregulated **(e)** and
199 downregulated **(f)** in wild-type worms compared to *skn-1(lax188)* [*skn-1(gf)*] mutants²⁵.
200 **(g-j)** Genes whose expression was upregulated **(g,i)** and downregulated **(h,j)** in wild-
201 type worms compared to the worms treated with *skn-1* RNAi are shown ^{a: 26; b: 27}. *p*
202 values were calculated by using two-tailed paired permutation test.

Supplementary Figure 10



203 **Supplementary Fig. 10: Effects of *daf-16* RNAi, *skn-1(gf)*, *skn-1(-)*, and *pmk-1***
 204 **RNAi on lifespan and pathogen resistance. (a-c)** Effects of *daf-16* RNAi [*daf-16(-)*] (n
 205 = 120 for each condition) (a), *skn-1(lax188)* [*skn-1(gf)*] (n ≥ 295 for each condition) (b),
 206 *skn-1(zj15)* [*skn-1(-)*] (n ≥ 270 for each condition) (c), and *pmk-1* RNAi [*pmk-1(-)*] (n =
 207 120 for each condition) (d) on the lifespan of wild-type (WT) and *daf-2(e1370)* [*daf-2(-)*]
 208 worms. Please note that we used *skn-1(zj15)*, a point mutation that causes mis-splicing
 209 and reduces the mRNA levels of *skn-1*, because a strong *skn-1* mutant allele, *skn-*
 210 *1(mg570)*, causes sickness and short lifespan²⁸; indeed, we found that *mg570*
 211 substantially increased vulval rupture phenotypes in worms: 6% in WT and 22% in *daf-*
 212 *2(-); daf-18(nr2037)* [*daf-18(-)*] backgrounds. (e,f) The effects of *pmk-1* RNAi (n ≥ 108
 213 for each condition) on the survival of *daf-2(-); daf-18(yh1)* and *daf-2(-); daf-18(-)* (e), and
 214 WT and *daf-2(-)* worms (f) against PA14 infection. Although *daf-2(-); daf-18(-)* animals
 215 displayed increased PMK-1 activity compared with *daf-2(-)* or *daf-2(-); daf-18(yh1)*

216 mutants shown in Fig. 5, reduced DAF-16/FOXO activity in *daf-2(-); daf-18(-)* animals
217 may have decreased the survival of the worms on PA14. These results are consistent
218 with a previous report showing that genetic inhibition of DAF-16/FOXO, which acts
219 downstream of DAF-18/PTEN, increases PMK-1 target gene expression in *daf-2*
220 mutants¹⁴. In contrast to lifespan results shown in Fig. 6d, we found that PMK-1 was
221 required for the enhanced pathogen resistance of *daf-2(-)* and *daf-2(-); daf-18(yh1)*
222 animals and for the normal survival of WT upon PA14 infection. Our data indicate that
223 reducing the activity of PMK-1 can extend lifespan in *daf-2(-); daf-18(-)* animals, while
224 decreasing immunity in these animals. These data are consistent with a previous report
225 showing that lifespan and immunity can be regulated in opposite directions by one
226 genetic factor²⁹. See Supplementary Dataset 2 for statistical analysis and additional
227 repeats for the data shown in this figure. See also Source Data for data points used for
228 the derivation of data.

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314 **Supplementary Tables**

315 **Supplementary Table 1. Molecular natures of mutant alleles identified from our**
 316 **mutagenesis screen.**

Allele	yh1	yh2	yh3
Gene	<i>daf-18</i>	<i>daf-16</i>	<i>daf-16</i>
Base change	G to A	C to T	G to A
Function	C150Y	Q115(STOP)	Splice donor
Isoforms affected	All	<i>a/c/d/f/h/i/k/l/m</i>	<i>a/c/d/f/h/i/k/l/m</i>
Domain affected	Phosphatase domain	Forkhead domain	Forkhead domain

317

318 **Supplementary Table 2. References for multiple target genes in *daf-2* mutants that**
 319 **were used for analysis in Fig. 5 and Supplementary Fig. 7.**

Genes	Strains	Figure	Reference	
DAF-16- induced/repressed	<i>daf-2(e1370)</i> vs. <i>daf-16(mu86)</i> ; <i>daf-2(e1370)</i>	Fig. 5a and Supplementary Fig. 7a,b	Chen <i>et al.</i> , 2015	
	<i>daf-2(e1370)</i> vs. <i>daf-16(mg54)</i> ; <i>daf-2(e1370)</i> <i>daf-2(e1370)</i> vs. <i>daf-16(tm6659)</i> ; <i>daf-2(e1370)</i> <i>daf-2(e1370)</i> vs. <i>daf-16(tm5032)</i> ; <i>daf-2(e1370)</i> <i>daf-2(e1370)</i> vs. <i>daf-16(tm5030)</i> ; <i>daf-2(e1370)</i>	Supplementary Fig. 7b,c		
	<i>daf-2(e1370)</i> vs. <i>daf-16(mu86)</i> ; <i>daf-2(e1370)</i> <i>daf-2(e1370)</i> vs. <i>daf-16(mg54)</i> ; <i>daf-2(e1370)</i>	Supplementary Fig. 7b,c		GSE111338 in GEO
	<i>daf-2(e1370)</i> vs. <i>daf-16(mgDf50)</i> ; <i>daf-2(e1370)</i>	Supplementary Fig. 7b,c		Kumar <i>et al.</i> , 2015
SKN-1- induced/repressed	<i>daf-2(e1370)</i> vs. <i>daf-2(e1370)</i> ; <i>skn-1(zu67)</i> <i>daf-2(e1368)</i> vs. <i>daf-2(e1368)</i> ; <i>skn-1(zu67)</i>	Fig. 5a and Supplementary Fig. 7a	Ewald <i>et al.</i> , 2015	
PMK-1- induced/repressed	<i>daf-2(e1368)</i> vs. <i>daf-2(e1368)</i> ; <i>pmk-1(km25)</i>	Fig. 5a and Supplementary Fig. 7a	Troemel <i>et al.</i> , 2006	

HSF-1-induced/repressed	<i>daf-2(e1370)</i> vs. <i>hsf-1(RNAi)</i> ; <i>daf-2(e1370)</i>	Fig. 5a and Supplementary Fig. 7a	Lee <i>et al.</i> , 2021
HLH-30-induced/repressed	<i>daf-2(e1370)</i> vs. <i>daf-2(e1370)</i> ; <i>hlh-30(tm1978)</i>	Fig. 5a and Supplementary Fig. 7a	Lin <i>et al.</i> , 2018
HEL-1-induced/repressed	<i>daf-2(e1370)</i> vs. <i>hel-1(gk148684)</i> ; <i>daf-2(e1370)</i>	Fig. 5a and Supplementary Fig. 7a	Seo <i>et al.</i> , 2015
Histone H3.3-induced/repressed	<i>daf-2(e1370)</i> vs. <i>daf-2(e1370)</i> ; <i>his-72(tm2066)</i> ; <i>his-71(ok2289)</i>	Fig. 5a and Supplementary Fig. 7a	Piazzesi <i>et al.</i> , 2016
MATH-33-induced/repressed	<i>daf-2(e1370)</i> vs. <i>daf-2(e1370)</i> ; <i>math-33(tm3561)</i> at 25°C (restrictive temperature)	Fig. 5a and Supplementary Fig. 7a	Heimbucher <i>et al.</i> , 2015
PFD-6-induced/repressed	<i>daf-2(e1370)</i> vs. <i>pf6-6(gk493446)</i> ; <i>daf-2(e1370)</i>	Fig. 5a and Supplementary Fig. 7a	Son <i>et al.</i> , 2018
RSKS-1-induced/repressed	<i>daf-2(e1370)</i> vs. <i>daf-2(e1370)</i> ; <i>rsk-1(ok1255)</i>	Fig. 5a and Supplementary Fig. 7a	Chen <i>et al.</i> , 2013
SMG-2-induced/repressed	<i>daf-2(e1370)</i> vs. <i>smg-2(qd101)</i> ; <i>daf-2(e1370)</i>	Fig. 5a and Supplementary Fig. 7a	Son <i>et al.</i> , 2017
SWSN-1-induced/repressed	<i>daf-2(e1370)</i> vs. <i>daf-2(e1370)</i> ; <i>swn-1(os22)</i>	Fig. 5a and Supplementary Fig. 7a	Riedel <i>et al.</i> , 2013