



## Supplementary Information for

### Therapeutic Inhibition of USP9x-Mediated Notch signaling in Triple-Negative Breast Cancer

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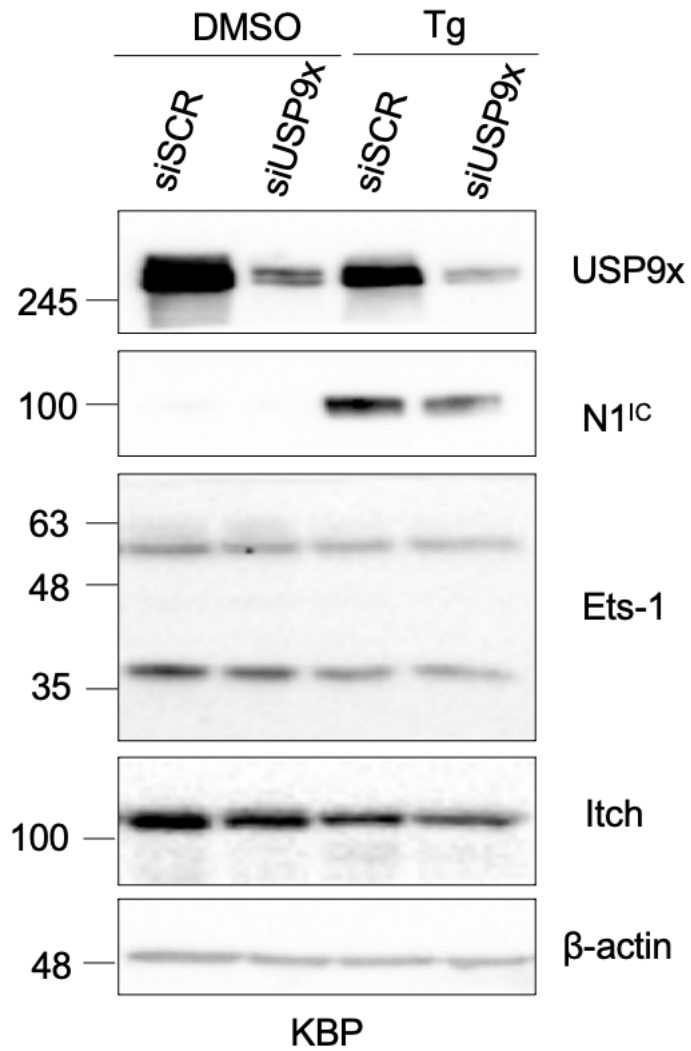
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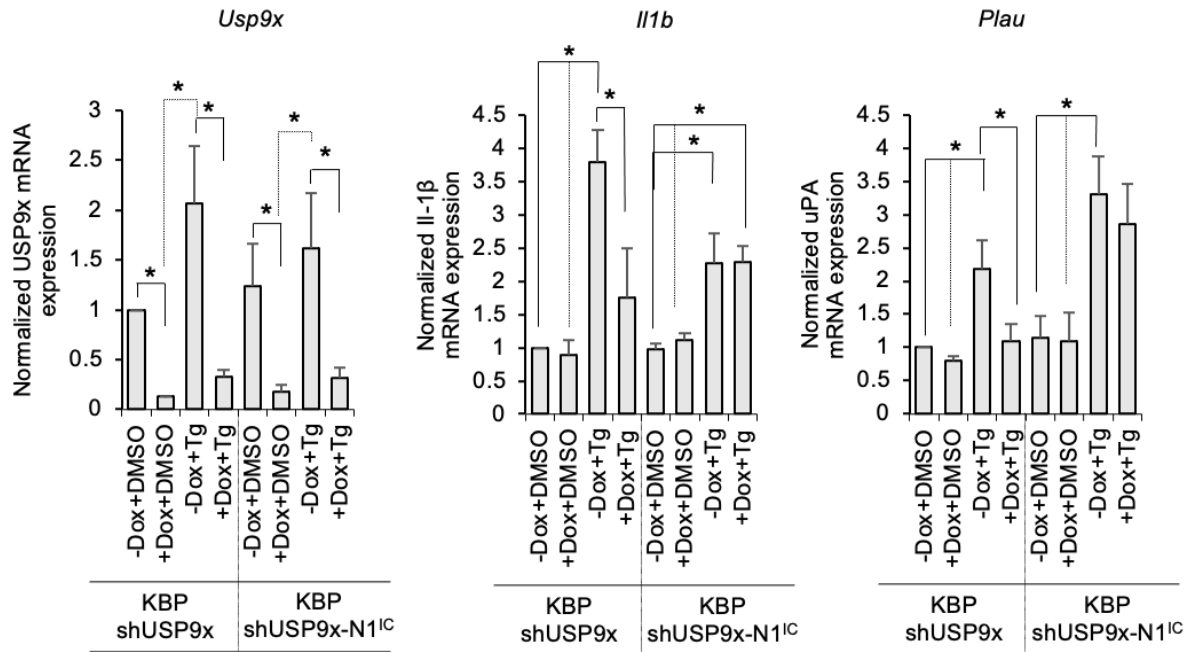
#### **This PDF file includes:**

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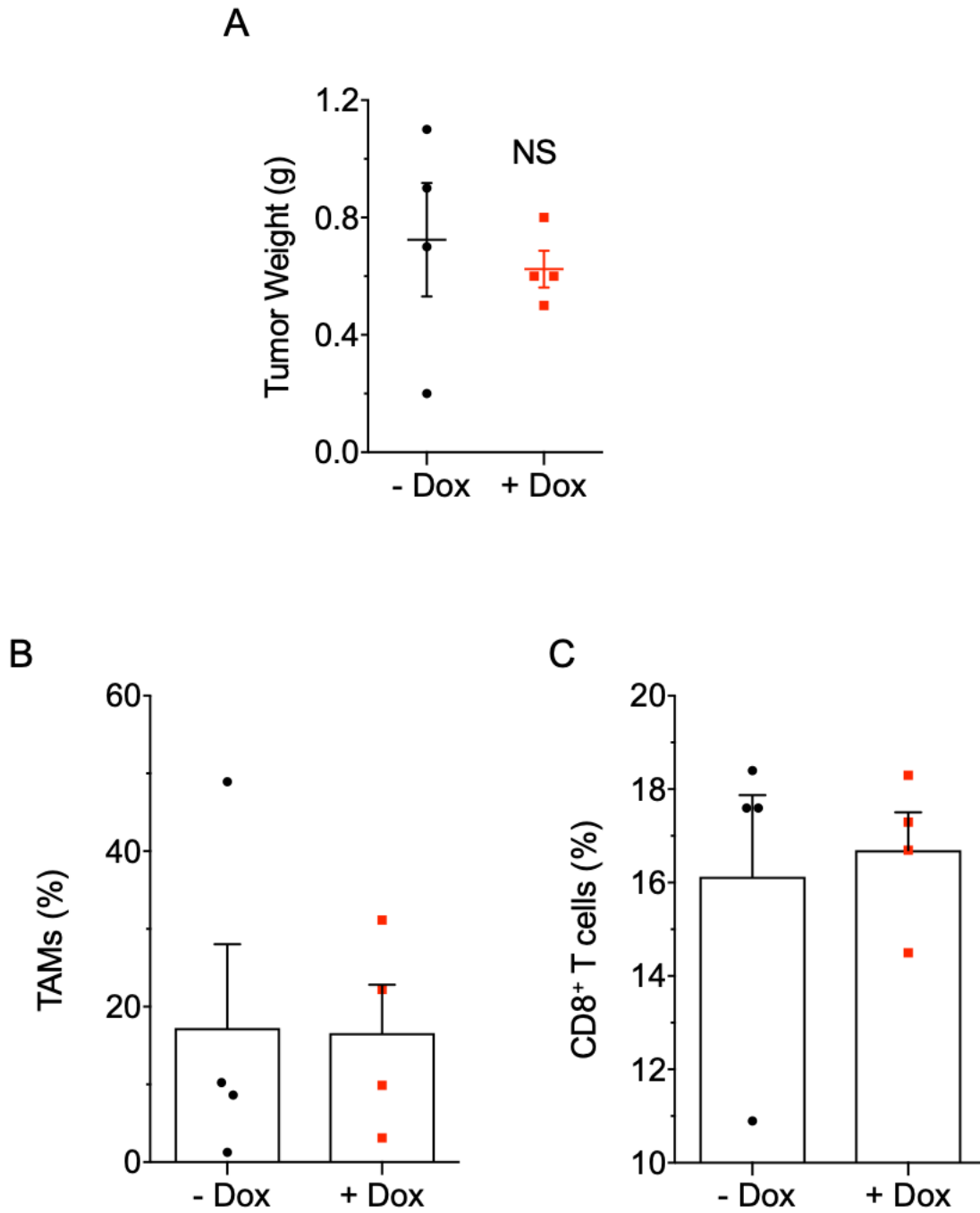
Tables S1 to S2



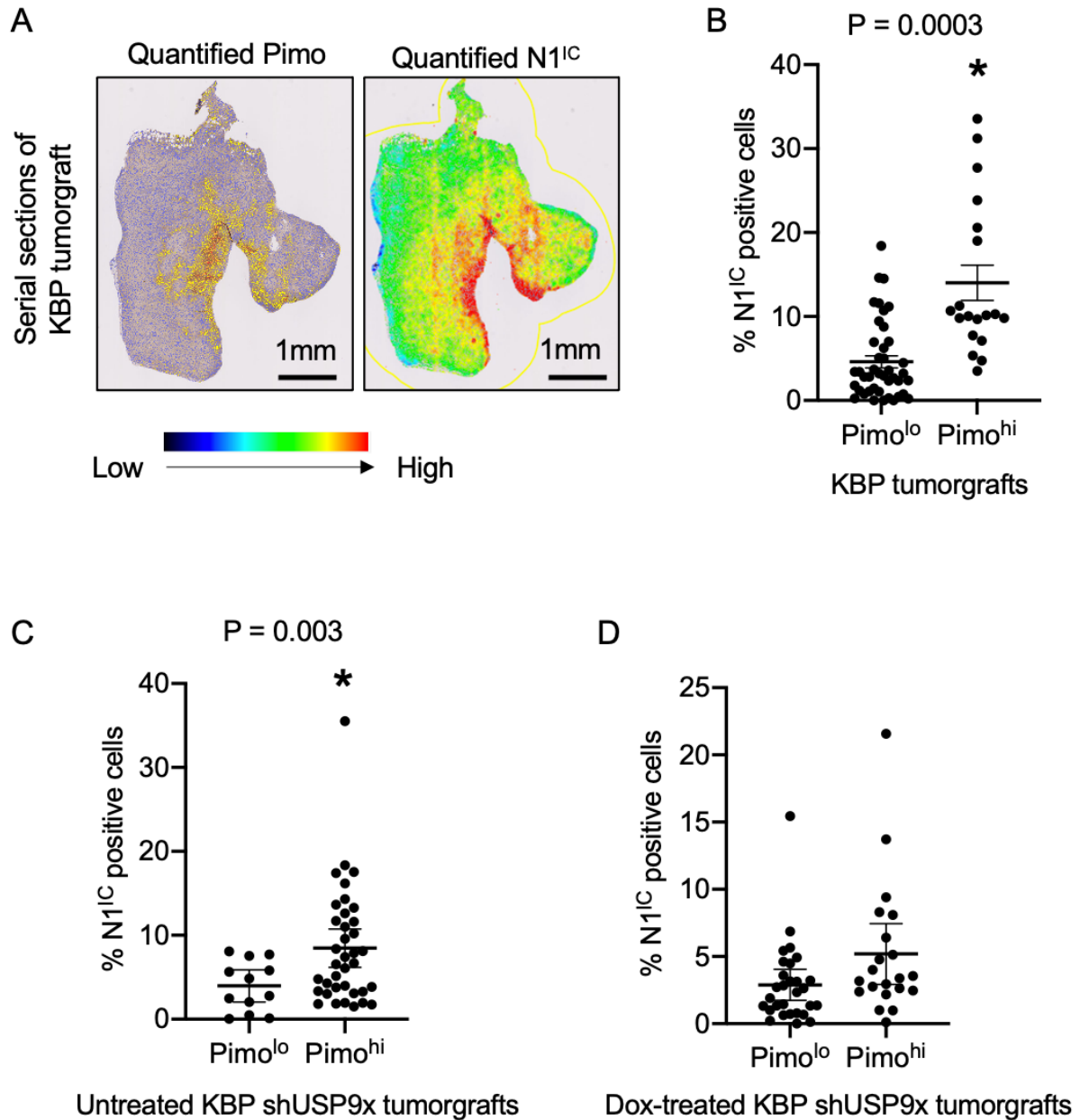
**Suppl Fig 1. USP9x knockdown does not affect the expression of Ets-1 or Itch in KBP cells.** Immunoblot of USP9x, N1<sup>IC</sup>, Ets-1 and Itch in KBP cells transfected with either siSCR or USP9x siRNA and treated with either DMSO or 1 μM Tg. β-actin is included as a loading control. Mw markers are shown in kilodaltons.



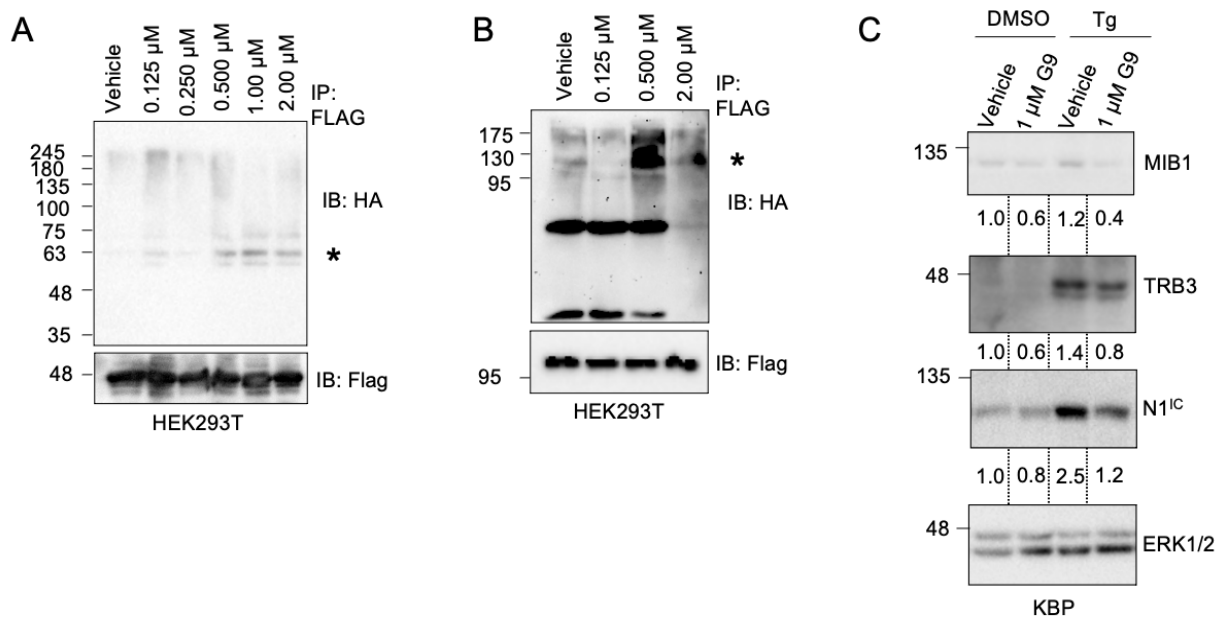
**Suppl Fig 2. Ectopic N1<sup>IC</sup> expression rescues the effect of USP9x knockdown on the expression of Notch target genes.** qPCR analysis of *Usp9x* and Notch target genes (*Il1b* and *Plau*) in uninfected or retrovirus infected (expressing N1<sup>IC</sup>) KBP shUSP9x and treated with either DMSO or 1  $\mu$ M Tg in the absence or presence of doxycycline. Expression levels were normalized to GAPDH. Experiments were performed in triplicate; error bars represent SEM. The asterisks (\*) denotes  $P < 0.05$  relative to the respective control.  $P$  values are determined by one-way ANOVA and corrected for multiple comparisons using Tukey's test.



**Suppl Fig 3. Doxycycline treatment has no effect on KBP allograft growth. (a)** Weight of KBP allografts fed a regular or doxycycline-containing diet (n = 4 per group). **(b and c)** Quantification (by flow cytometry) of TAMs (Gr-1<sup>-</sup> F4/80<sup>+</sup> CD11b<sup>+</sup>) **(b)** and CD8<sup>+</sup> T **(c)** infiltrates in KBP tumors expressed as % of CD45<sup>+</sup> cells. Error bars represent SEM. NS, non-significant. *P*-value is determined by the two-tailed Student's *t*-test.

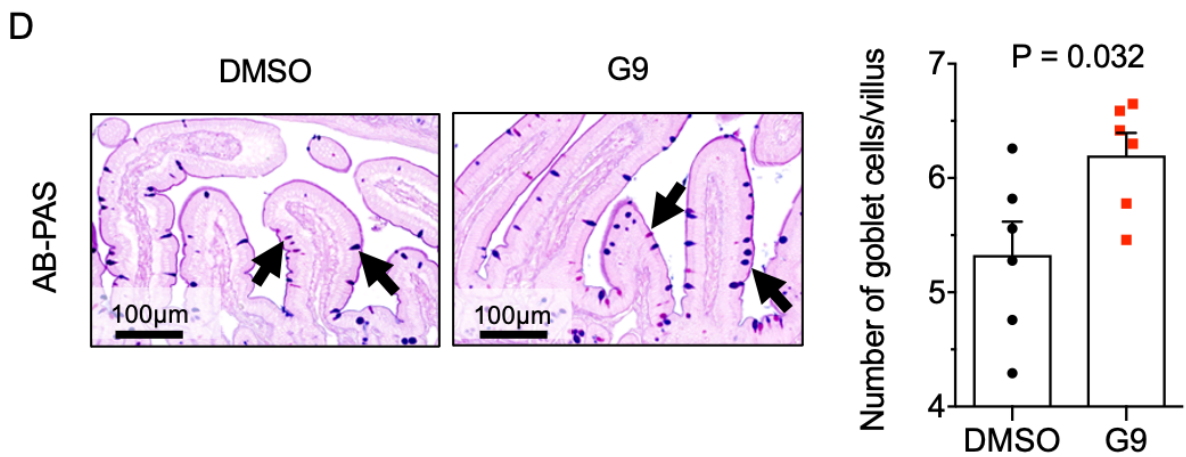
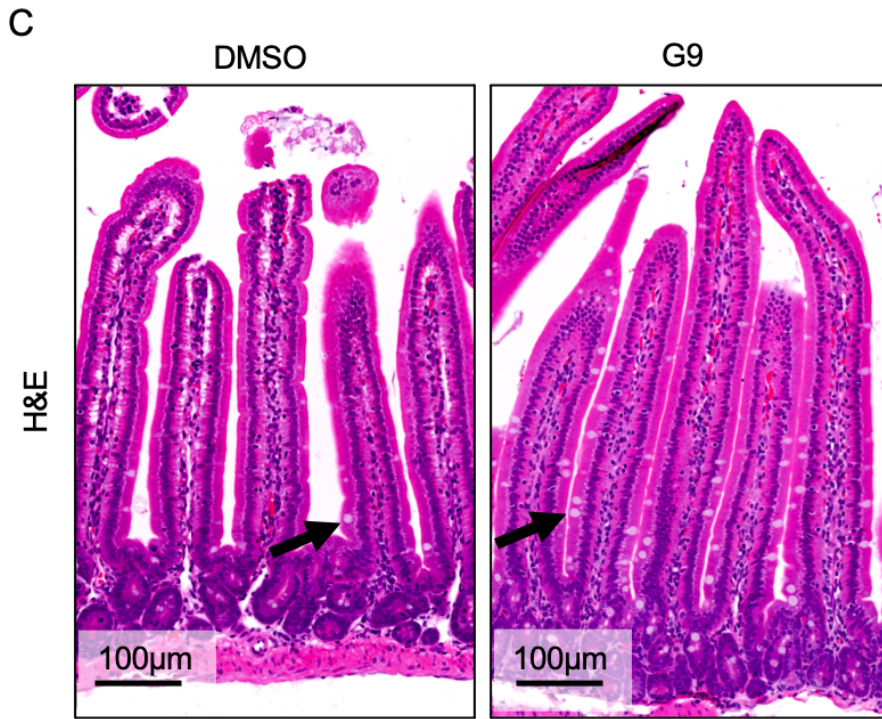
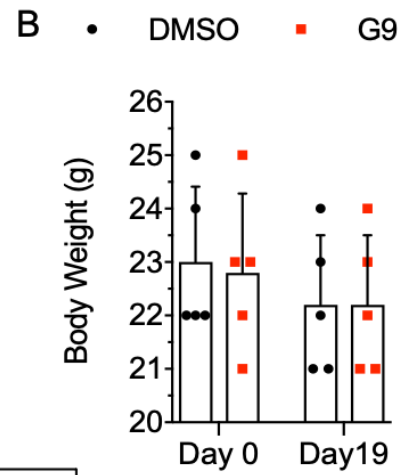
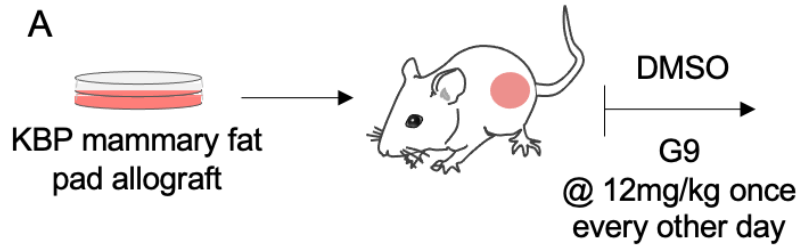


**Suppl Fig 4. Co-localization of hypoxia and Notch activation is USP9x-dependent.** (a) Representative quantified IHC images of the hypoxia marker pimonidazole (Pimo) and N1<sup>IC</sup> in serial sections of KBP allografts. The color bar indicates the expression level of Pimo and N1<sup>IC</sup>. Scale bars: 1mm. (b-d) Percentage of N1<sup>IC</sup>+ cells in Pimo<sup>lo</sup> (< 20% pimo<sup>+</sup> cells) vs Pimo<sup>hi</sup> (> 20% cells Pimo<sup>+</sup>) regions of KBP (b), untreated KBP shUSP9x (c) and doxycycline-treated KBP shUSP9x (d) allografts (14-18 x 20,000 μm<sup>2</sup> regions were surveyed in n = 4 tumors per group). Error bars represent SEM. \* = P < 0.05. P indicates the significance value determined by the two-tailed Student's *t*-test with Welch's correction for unequal variances.



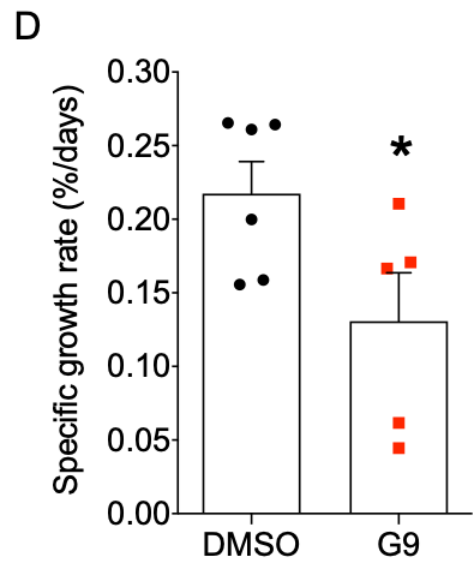
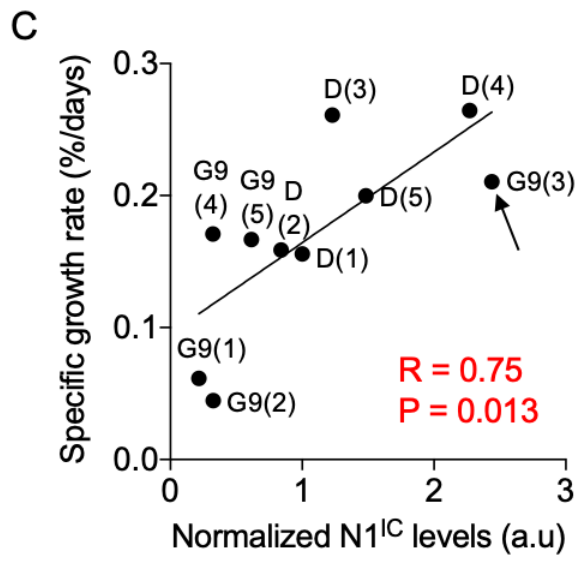
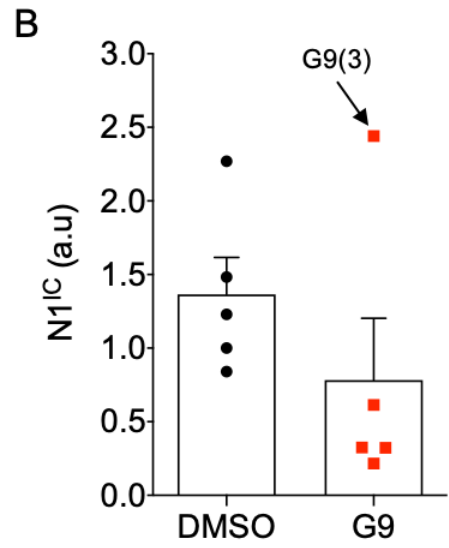
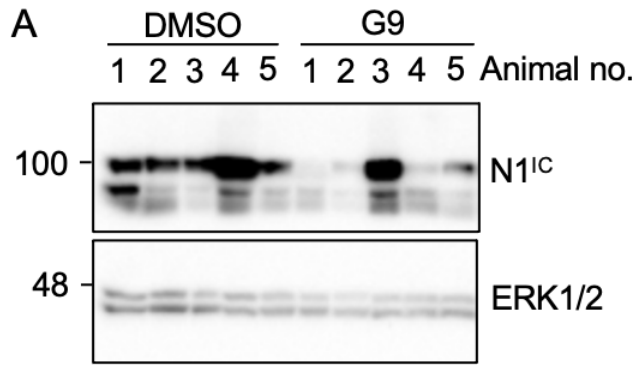
**Suppl Fig 5. G9 inhibits stress-induced Notch activation in murine KBP cells.**

Immunoblot of HA and Flag after anti-Flag immunoprecipitations from HEK293T cells transfected with plasmids expressing HA-Ub and either TRB3-Flag (**a**) or MIB1-Flag (**b**) and treated with either vehicle or varying concentrations of G9 in the presence of proteasome inhibitor, MG132. \* marks the predicted size of monoubiquitinated TRB3-Flag-HA-Ub (**a**), or MIB1-Flag-HA-Ub (**b**). (**c**) Immunoblot of MIB1, TRB3, and N1<sup>IC</sup> in KBP cells treated with either vehicle or 1 μM G9 and treated with either DMSO or 1 μM Tg. Expression of ERK1/2 is included as a loading control. Expression of MIB1, TRB3 and N1<sup>IC</sup> are determined by densitometry and quantified relative to ERK1/2. *Mw* markers are shown in kilodaltons.



**Suppl Fig 6. The effect of G9 on the gastrointestinal epithelium. (a)** Schematic diagram of the experimental design. **(b)** Bodyweight of animals treated with DMSO or G9 (n = 5 mice per group). **(c)** Representative images of H&E-stained sections of murine intestinal villi. **(d)** Left panel: representative images of AB-PAS-stained sections of murine intestinal villi. Right panel: quantification of intestinal goblet cells (i.e secretory cells) from AB-PAS-stained sections (n = 6 mice). Arrows in **(c)** and **(d)** indicate goblet cells. Error bars represent SEM. Scale bars: 100 $\mu$ m. *P* values are determined by the two-tailed Student's *t*-test.





**Suppl Fig 7. G9 suppresses Notch-dependent KBP tumor growth.** (a) Immunoblot of N1<sup>IC</sup> in lysates of spontaneous KBP tumors treated with DMSO or G9. The expression of ERK1/2 is included as a loading control. *Mw* markers are shown in kilodaltons. (b) Expression of N1<sup>IC</sup> determined by densitometry relative to ERK1/2. The arrow indicates N1<sup>IC</sup> expression in G9-treated tumor #3. (c) Correlation analysis of specific growth rate (SGR; calculated using the formula  $\ln(V_2/V_1)/\Delta t$ , where  $V_1$  and  $V_2$  are the measured tumor volumes at the beginning and end of treatment respectively, and  $\Delta t$  is the treatment duration) with tumoral N1<sup>IC</sup> levels. Each data point is labelled with the sample ID (treatment type followed by sample number). R indicates the degree of correlation and P indicates the significance value determined through the Pearson correlation test. (d) SGR of KBP tumors treated with DMSO or G9 (DMSO, n = 6 tumors; G9, n = 5 tumors). Error bars represent SEM. \* = P < 0.05. P values are determined by the two-tailed Student's *t*-test for (b) and (d).

**Suppl Table 1. Sequences and Primers**

<b>siRNA</b>	<b>Sequence</b>	
<b>siscr</b>	UGGUUUACAUGUCGACUAA UGGUUUACAUGUUGUGUGA UGGUUUACAUGUUUUCUGA UGGUUUACAUGUUUCCUA	
<b>siTRB3</b>	GCAGAUACCCAUUCCACGA GAGAGAGGAUCACGGCCGA CUGGAAGAUGCCUGCGUGA AGGUGGGGCUGUACGGCUA	
<b>siUSP9x</b>	CAAAGGAGAUUUACUAGAA AGAAAUCGCUGGUUAAA ACACGAUGCUIUAGAAUUU GUACGACGAUGUUAUCUCA	
<b>shRNA</b>	<b>Sense</b>	<b>Anti-sense</b>
<b>shUSP9x (5'-3')</b>	GATGAGGAACCTGCATTTTC	GAAATGCAGGTTCCCTCATC
<b>qRT-PCR</b>	<b>Forward Primer</b>	<b>Reverse Primer</b>
<b>Mouse <i>Usp9x</i></b>	AAGGAGCTACTGGCTTTTTCAGA	GATGCGGGAAAGATGAAGTC
<b>Mouse <i>Plau</i></b>	GACAGCCTGGCCTACCATAA	AGTCTGAACCAAACGGAGCA
<b>Mouse <i>Il1b</i></b>	CTCTTGTTGATGTGCTGCTG	GACCTGTTCTTTGAAGTTGACG

**Suppl Table 2. Antibodies**

<b>Immunoblot and IHC</b>	<b>Source</b>	<b>Catalog no.</b>
<b>USP9x</b>	Bethyl Laboratories	A301-351A
<b>TRB3</b>	Santa Cruz Biotechnology	sc-365842
<b>MIB1</b>	Abcam	ab245766
<b>JAG1</b>	CST	2620
<b>Cleaved Notch1 [Val1744]</b>	CST	4147
<b>Notch1 (to detect ectopic N1<sup>C</sup>)</b>	Santa Cruz Biotechnology	sc-6014
<b>p44/42 MAPK (ERK1/2)</b>	CST	9102
<b>HA</b>	Santa Cruz Biotechnology	sc-805
<b>Flag</b>	Santa Cruz Biotechnology	sc-807
<b>β-actin</b>	Santa Cruz Biotechnology	sc-1615
<b>Ets-1</b>	CST	14069
<b>AIP4 / Itch</b>	Santa Cruz Biotechnology	sc-11890
<b>CCL2/MCP-1</b>	Santa Cruz Biotechnology	sc-1785
<b>IL-1β</b>	CST	12242
<b>anti-Mouse IgG, HRP-linked</b>	CST	7076
<b>anti-Rabbit IgG, HRP-linked</b>	CST	7074
<b>anti-goat IgG, HRP-linked</b>	Santa Cruz Biotechnology	sc-2020
<b>Pimonidazole</b>	Hypoxyprobe	PAb2627
<b>SMA</b>	Abcam	ab5964
<b>ELISA</b>	<b>Source</b>	<b>Catalog no.</b>
<b>CCL2</b>	R&D	DY479-05
<b>IL-1β</b>	R&D	DY401-05
<b>Flow cytometry</b>	<b>Source</b>	<b>Catalog no.</b>
<b>CD45</b>	eBioscience	63-0451-82
<b>CD3</b>	Biologend	100234
<b>F4/80</b>	eBioscience	17-4801-82
<b>CD11b</b>	BD biosciences	553310
<b>CD206</b>	Biologend	141719
<b>CD8</b>	eBioscience	45-0081-82
<b>GrmB</b>	Invitrogen	48-8898-82
<b>FoxP3</b>	eBioscience	12-5773-80
<b>Gr-1</b>	eBioscience	45-5931-80
<b>Viability</b>	eBioscience	65-0865-14