

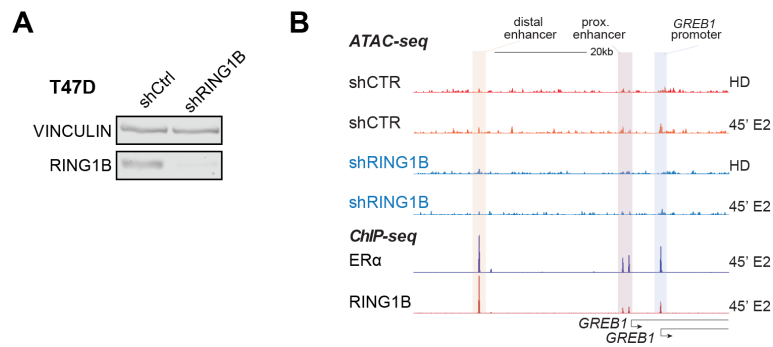
The Polycomb protein RING1B enables estrogen-mediated gene expression by promoting enhancer-promoter interaction and R-loop formation

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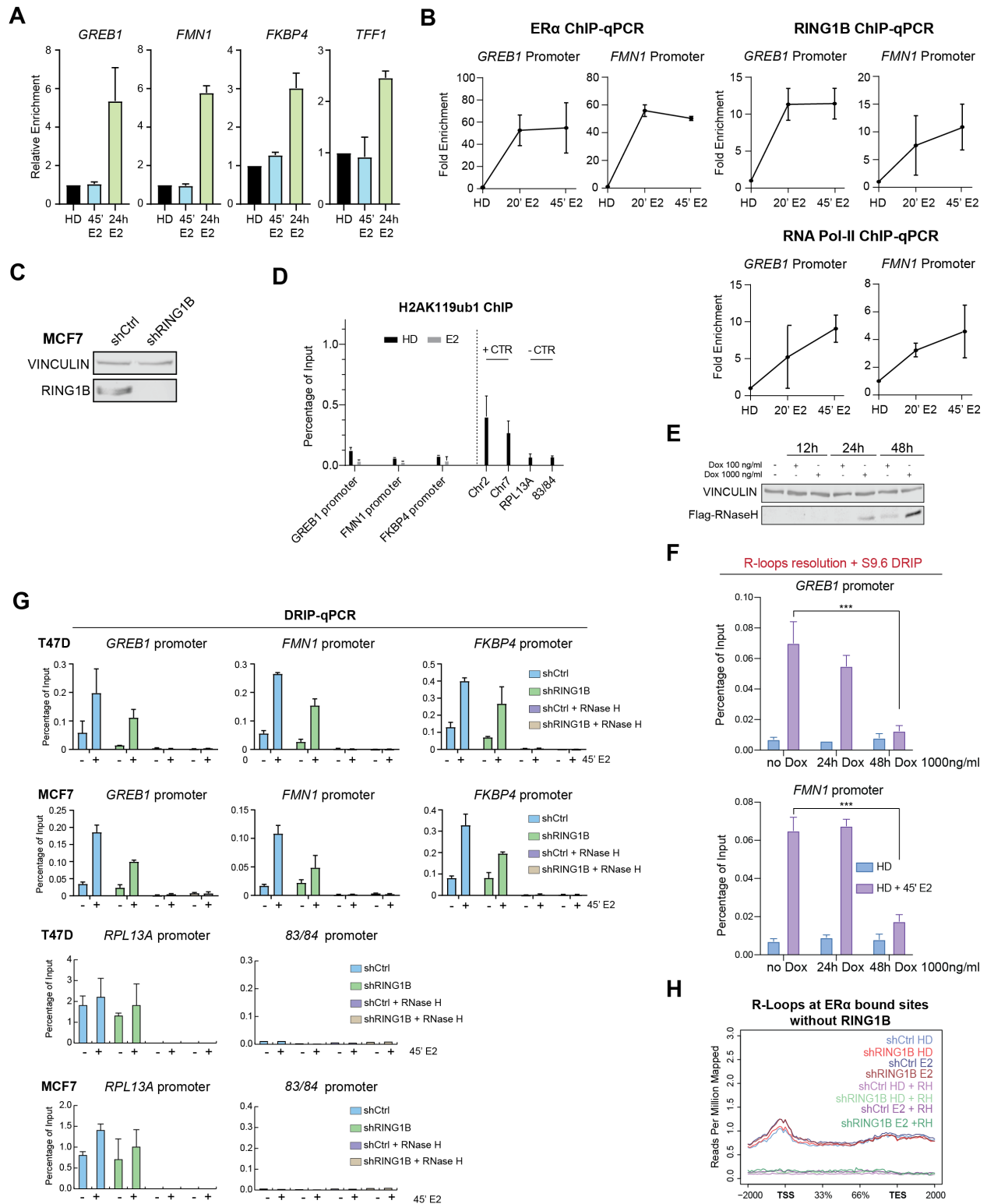
Supplementary Figures

Figure S1



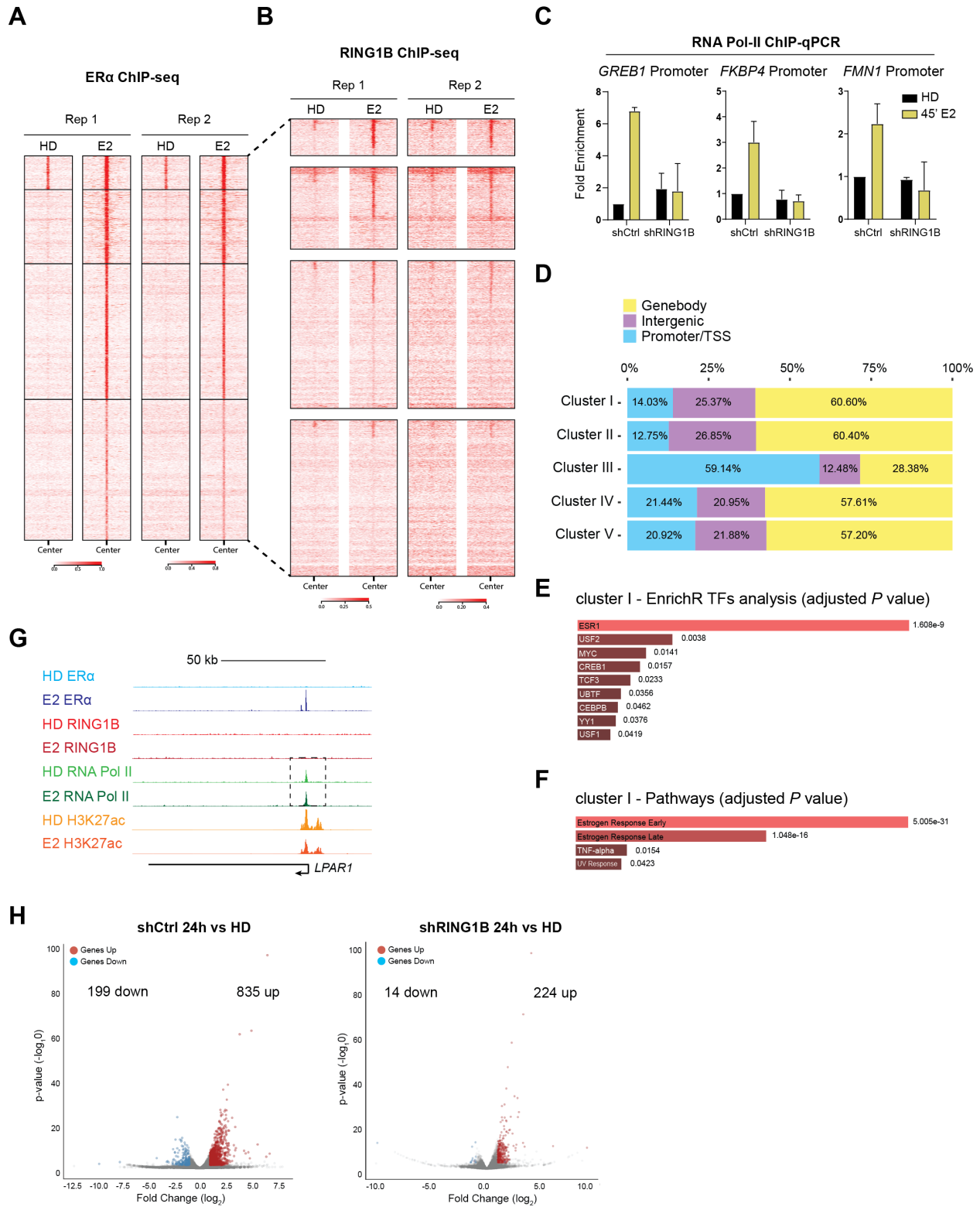
Supplementary Figure S1. (A) Western blot of RING1B and VINCULIN (loading control) in control and RING1B-depleted T47D cells. **(B)** ATAC-seq signal in control and RING1B-depleted T47D cells before and after 45min of E2. The three H3K27ac-enriched sites bound by both RING1B and ER α are highlighted and labeled as distal enhancer, proximal enhancer, and *GREB1* promoter.

Figure S2



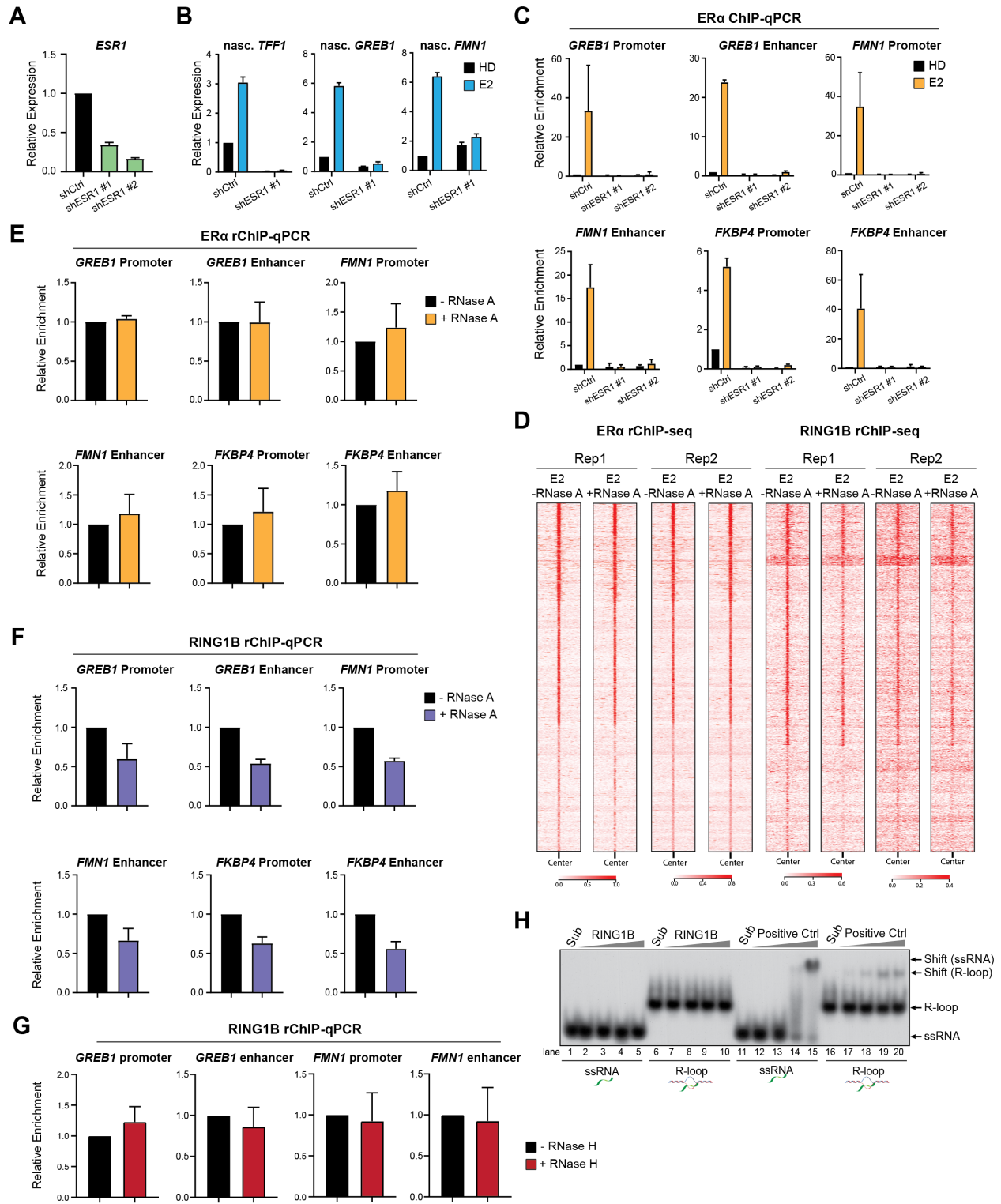
Supplementary Figure S2. (A) RT-qPCR of *GREB1*, *FMN1*, *FKBP4*, and *TFF1* in T47D parental cells before and after 45min and 24h of E2 stimulation. Error bars represent the standard deviation of two independent replicates. **(B)** ER α , RING1B, and RNA Pol-II ChIP-qPCR in T47D parental cells before and after 20min and 45min of E2 at RING1B and ER α co-regulated sites within the *GREB1* and *FMN1* promoters. Error bars represent the standard deviation of two independent replicates. **(C)** Western blot of RING1B and VINCULIN (loading control) in control and RING1B-depleted MCF7 cells. **(D)** H2AK119ub1 ChIP-qPCR before and after 45min of E2 administration at promoters co-occupied by RING1B and ER α . Two regions in chromosomes 2 and 7 were used as a positive control (4) and *RPL13A* and *83/84* were used as negative controls. **(E)** Flag WB in dox-inducible Flag-RNase H T47D cells treated with 100ng/ml or 1000ng/ml of dox for 12h, 24h and 48h. VINCULIN was used as a loading control. **(F)** DRIP-qPCR of RING1B/ER α co-targets before and after 45min of E2 induction in the presence of no dox or dox (1000ng/ml) for 24h and 48h. *** p-value < 0.001, ** p-value < 0.005, two-tailed t-test. **(G)** DRIP-qPCR in T47D and MCF7 shCTR and shRING1B cells before and after 45min of E2 addition at ER α and RING1B co-bound sites within the promoters of ER α target genes. No signal was detected in samples treated with RNase H, demonstrating that the detected enrichment is specific for R-loops. *RPL13A* and the *83/84* locus were used as positive and negative controls, respectively. Error bars represent the standard deviation of two biological replicates. **(H)** Average DRIP-seq signal at ER α target genes not occupied by RING1B.

Figure S3



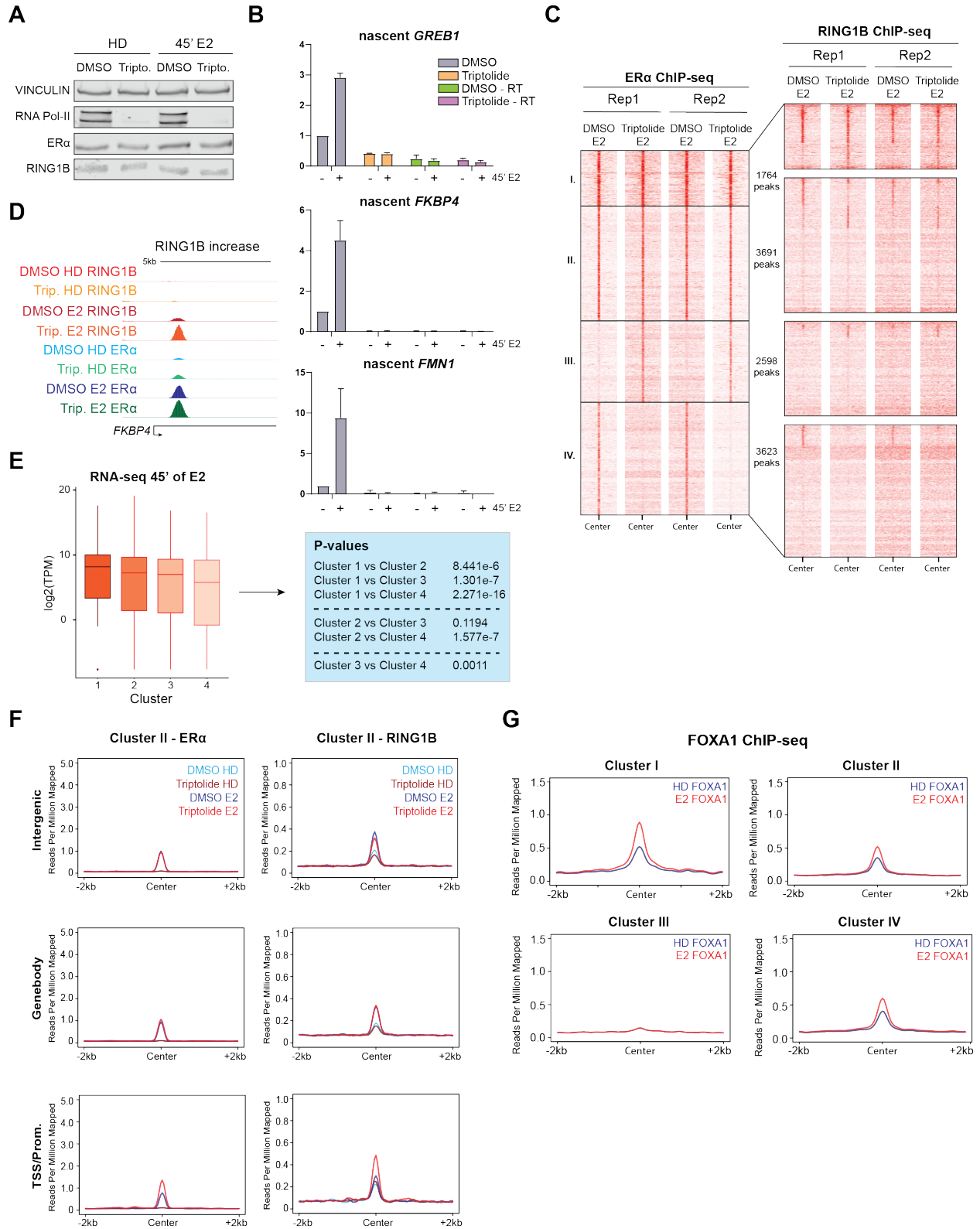
Supplementary Figure S3. (A-B) Heatmap of each ER α (A) or RING1B (B) ChIP-seq replicate before and after 45min of E2 organized in order of the 4 clusters from Figure 3A. **(C)** RNA Pol-II ChIP-qPCR in shCTR and shRING1B T47D cells before and after 45min of E2 at RING1B and ER α co-regulated sites. Error bars represent the standard deviation of two independent replicates. **(D)** Genomic annotation of RNA Pol II peaks within each cluster from Figure 3C. **(E-F)** EnrichR TF analysis (E) and EnrichR pathway analysis (F) of genes assigned to cluster I RNA Pol II peaks. **(G)** ER α , RING1B, RNA Pol II, and H3K27ac signal before and after 45min of E2 at *LPAR1*, which is regulated only by ER α . **(H)** Volcano plots (adjusted *P* value) of deregulated genes in T47D-shCTR and shRING1B cells after 24 hours of E2.

Figure S4



Supplementary Figure S4. (A) RT-qPCR of *ESR1* in T47D shCTR and two shESR1 cell lines, showing ~66% ER α depletion in shESR1-1 and ~85% depletion in shESR1 #1. Error bars represent the standard deviation of two independent replicates. **(B)** RT-qPCR of nascent *TFF1*, *GREB1*, and *FMN1* transcripts in T47D shCTR and shESR1-1 cells before and after 45min of E2 stimulation. Error bars represent the standard deviation of two independent replicates. **(C)** ER α ChIP-qPCR in control and ER α -depleted T47D cells before and after 45min of E2 stimulation at RING1B and ER α co-targets. Error bars represent the standard deviation of two independent replicates. **(D)** Heatmaps of each rChIP-seq replicate of ER α (left) and RING1B (right) after 45min of E2 stimulation. **(E-F)** rChIP-qPCR of ER α (E) and RING1B (F) after 45min of E2 stimulation with and without RNase A treatment. Error bars represent the standard deviation of two independent replicates. **(G)** rChIP-qPCR of RING1B after 45min of E2 stimulation with and without prior RNase H treatment of chromatin. Error bars represent the standard deviation of two independent replicates. **(H)** Electrophoretic mobility shift assay (EMSA) using ssRNA and R-loops demonstrate a shift in ssRNA and R-loop migration when incubated with a positive control but not when incubated with RING1B.

Figure S5



Supplementary Figure S5. (A-B) Western blot (A) of RNA Pol II, ER α , RING1B, and VINCULIN (loading control) and RT-qPCR (B) in T47D cells treated with vehicle (DMSO) or triptolide, before and after 45min of E2. Error bars represent the standard deviation from two biological replicates. **(C)** Heatmap of both replicates of ER α (left) and RING1B (right) ChIP-seq in cells treated with vehicle (DMSO) or triptolide upon 45 min of E2 organized in order of the ER α clusters from Figure 4F. **(D)** RING1B and ER α ChIP-seq before and after 45min of E2 in cells treated with DMSO or triptolide demonstrating increased ER α and RING1B recruitment at the *FKBP4* promoter following triptolide treatment. **(E)** Boxplot depicting TPM values of genes assigned to each cluster from Figure 4F (left) with comparisons between the 4 clusters (right). **(F)** Average profiles showing the levels of ER α and RING1B chromatin binding at cluster 2 ER α sites, classified by genomic annotation, before and after 45 min of E2 in cells treated with DMSO or triptolide. **(G)** FOXA1 ChIP-seq signal at each of the four ER α clusters before and after 45min of E2 treatment.