#### **Supporting information**

# Haloperidol Metabolite II Valproate Ester (S)-(-)-MRJF22: Preliminary Studies as Potential Multifunctional Agent Against Uveal Melanoma

Carla Barbaraci,<sup>†,#,§</sup> Giovanni Giurdanella,<sup>‡,§</sup> Claudia Giovanna Leotta,<sup>#</sup> Anna Longo,<sup>‡</sup> Emanuele Amata,<sup>†</sup> Maria Dichiara,<sup>†</sup> Lorella Pasquinucci,<sup>†</sup> Rita Turnaturi,<sup>†</sup> Orazio Prezzavento,<sup>†</sup> Ivana Cacciatore,<sup>¢</sup> Elisa Zuccarello,<sup>Σ</sup> Gabriella Lupo,<sup>‡</sup> Giovanni Mario Pitari,<sup>\*,#</sup> Carmelina Daniela Anfuso,<sup>\*,‡</sup> Agostino Marrazzo<sup>\*,†</sup>

<sup>†</sup>Department of Drug and Health Sciences, University of Catania, Viale A. Doria 6, 95125 Catania, Italy

<sup>‡</sup>Department of Biomedical and Biotechnological Sciences, School of Medicine, University of Catania, Via S. Sofia 97, 95123 Catania, Italy

<sup>#</sup>Vera Salus Ricerca S.r.l, Via Sigmund Freud 62/B, 96100 Siracusa, Italy

<sup>*o*</sup>Department of Pharmacy, "G. D'Annunzio" University of Chieti-Pescara, Via dei Vestini 31, 66100 Chieti Scalo, Italy

<sup>2</sup>Taub Institute for Research on Alzheimer's Disease and the Aging Brain, Columbia University, New York, New York 10032, United States.

#### **Corresponding Author**

- \*Agostino Marrazzo: phone, (+39) 095 7834250; email, marrazzo@unict.it
- \* Carmelina Daniela Anfuso: phone: (+39) 095 4781170; email: daniela.anfuso@unict.it
- \* Giovanni Mario Pitari: phone, (+39) 0931 1987360; email, giovanni.pitari@verasalusricerca.it

Table of contents		
Chemistry	S3	
Figure S1	S5	
Figure S2	S6	
Figure S3	S7	
Figure S4	S8	
Figure S5	S9	
NMR spectra	S10	
HR-MS spectra	S12	

#### Chemistry

General procedure for the synthesis of compounds (–)-3 and (+)-3. A solution of 4-chloro-1-(4-fluorophenyl)butan-1-one (2, 1 mmol) in dry THF was added dropwise to a solution of (+) or (–)-diisopinocanphenylchloroborane (DIP-Cl) in THF dry at –25 °C. The mixture was stirred at this temperature for 16 h under N<sub>2</sub>. The solvent was removed under vacuum and the residue was taken up into diethyl ether. To this solution was carefully added diethanolamine (DEA) and let stirred under N<sub>2</sub> overnight. The white salt formed was separated by filtration through Celite<sup>®</sup> and the filtrate was concentred. The reaction mixture was quenched with a solution of HCl 1M and the aqueous phase extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 x 50 mL). The organic phase was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and evaporated. The crude product was purified by flash chromatography (1:9 EtOAc/Cy) to obtain the desired products.

(*R*)-(+)-4-chloro-1-(4-fluorophenyl)butan-1-ol [(+)-3]. Compound (+)-3 was prepared according to the general procedure using 2 (0.400 g, 2.0 mmol) and (+)-DIP-Cl (2.570 g, 8.0 mmol). Yield: 0.385 g (95%), white solid.  $[\alpha]_D^{20}$ = +37° (*c*1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.42–7.28 (m, 2H), 7.07–6.99 (m, 2H), 4.70 (t, *J* = 6.0 Hz, 1H), 3.58–3.52 (m, 2H), 2.57–2.30 (m, 2H), 2.12–1.63 (m, 3H). <sup>13</sup>C NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  164.7 (*J*<sub>CF</sub> = 325.0 Hz), 141.2, 131.1 (*J*<sub>CF</sub> = 8.5 Hz), 115.7 (*J*<sub>CF</sub> = 35.0 Hz), 72.1, 44.2, 37.2, 24.3. Anal. Calcd for C<sub>10</sub>H<sub>12</sub>ClFO: C, 59.27; H, 5.97; N, 17.49. Found: C, 59.50; H, 6.07; N, 17.58.

(*S*)-(-)-4-chloro-1-(4-fluorophenyl)butan-1-ol [(-)-3]. Compound (-)-3 was prepared according to the general procedure using 2 (0.500 g, 2.0 mmol) and (-)-DIP-Cl (0.600 g, 12.0 mmol). Yield: 0.340 g (84%), white solid.  $[\alpha]_D^{20} = -41^\circ$  (*c*1.2, CHCl<sub>3</sub>). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.42–7.28 (m, 2H), 7.07–6.99 (m, 2H), 4.70 (t, J = 6.0 Hz, 1H), 3.58–3.52 (m, 2H), 2.57–2.30 (m, 2H), 2.12–1.63 (m, 3H). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  164.7 ( $J_{CF} = 325.0$  Hz), 141.2, 131.1 ( $J_{CF} = 8.5$  Hz), 115.7 ( $J_{CF} = 35.0$  Hz), 72.1, 44.2, 37.2, 24.3. Anal. Calcd for C<sub>10</sub>H<sub>12</sub>ClFO: C, 59.27; H, 5.97; N, 17.49. Found: C, 59.45; H, 6.07; N, 17.79.

General procedure for the synthesis of compounds (+)-4 and (–)-4. Both compounds were synthesized as reported in literature.<sup>33</sup> To a solution of 4-(4-chlorophenyl)hydroxypiperidine (4 mmol) and KHCO<sub>3</sub> (4 mmol) in dry DMF (12 mL) was added dropwise a solution (–)-3 or (+)-3 (1 mmol). The reaction mixture was heated to 80 °C for 24 h. The solvent was removed under vacuum and the residue was dissolved in CHCl<sub>3</sub> and washed with NaHCO<sub>3</sub> (3 x 25 mL). The organic layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and evaporated. The crude product was purified by flash chromatography (1:9 MeOH/CH<sub>2</sub>Cl<sub>2</sub>) to obtain the desired products.

(*R*)-(+)-4-(4-chlorophenyl)-1-[4(4-fluorophenyl)-4-hidroxybutyl]piperidin-4-ol [(*R*)-(+)-HP-mII, (+)-4]. According to general procedure, compound (+)-4 was prepared by reacting 4-(4-chlorophenyl)hydroxypiperidine (1.140 g, 5.0 mmol) and compound (+)-3 (0.362 g, 1.79 mmol). Yield 0.189 g (28%), white solid. Mp: 145-146.5 °C; of,  $[\alpha]_D^{20}$ = +66° (*c*1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.47–7.27 (m, 6H), 7.05–6.95(m, 2H), 4.65–4.62 (m, 1H), 3.05–2.99 (m, 1H), 2.85–2,79 (m, 1H), 2.67–2,42 (m, 4H), 2.28–2.12 (m, 2H), 2.01–1.79 (m, 8H). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  164.7 (*J*<sub>CF</sub> = 325.5 Hz), 145.7, 138.3, 130.4, 130.4 (*J*<sub>CF</sub> = 8.5 Hz), 120.9, 115.8 (*J*<sub>CF</sub> = 35.5 Hz), 73.0, 71.1, 55.7, 48.1, 39.6, 39.4, 24.8. Anal. Calcd for C<sub>21</sub>H<sub>25</sub>ClFNO<sub>2</sub>: C, 65.19; H, 6.77; N, 3.62. Found: C, 65.37; H, 7.02; N, 3.20.

(*S*)-(-)-4-(4-chlorophenyl)-1-[4(4-fluorophenyl)-4-hidroxybutyl]piperidin-4-ol [(*S*)-(-)-HP-mII, (-)-4]. According to general procedure, compound (-)-4 was prepared by reacting 4-(4-chlorophenyl)hydroxypiperidine (0.865 g, 4.0 mmol) and compound (-)-3 (0.206 g, 1.0 mmol). Yield 0.291 g (77%), white solid. Mp: 145–146 °C.  $[\alpha]_D^{20}$ =-65° (*c*1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.47–6.83 (m, 8H), 4.65–4.62 (m, 1H), 3.05–2.99 (m, 1H), 2.85–2,79 (m, 1H), 2.67–2,42 (m, 4H), 2.31–2.23 (m, 2H), 2.10–1.50 (m, 8H). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  164.67 (*J*<sub>CF</sub> = 325.5 Hz), 145.7, 138.3, 130.4, 130.4 (*J*<sub>CF</sub> = 8.5 Hz), 120.9, 115.8 (*J*<sub>CF</sub> = 35.5 Hz), 73.0, 71.1, 55.7, 48.1, 39.6, 39.4, 24.8. Anal. Calcd for C<sub>21</sub>H<sub>25</sub>ClFNO<sub>2</sub>: C, 65.19; H, 6.77; N, 3.62. Found: C, 64.97; H, 6.47; N, 3.99.



Compound	Retention time	Area%
( <i>R</i> )-(+)-MRJF22	9.7	45.7
(S)-(-)-MRJF22	10.7	54.3



Compound	Retention time	Area%	ee%
( <i>R</i> )-(+)-MRJF22	9.7	95.3	92
(S)-(-)-MRJF22	10.7	4	



Compound	Retention time	Area%	ee%
( <i>R</i> )-(+)-MRJF22	9.7	2.3	95.4
( <i>S</i> )-(-)-MRJF22	10.7	97.7	

Figure S1. HPLC chromatograms of (±)-MRJF22, (*R*)-(+)-MRJF22, and (*S*)-(-)-MRJF22.

Effects of (-)-1 and (+)-1 on HREC viability at three different time points.



**Figure S2.** MTT values obtained in cells treated with increasing concentration (1.0, 2.5, 5.0, 10.0 and 20.0  $\mu$ M) of (-)-1 (A) and (+)-1 (B) for 24, 48 and 72 h. Data are expressed as a mean  $\pm$  SEM of three independent experiments, each involving three different wells per condition. Statistical analysis was performed using two-way ANOVA, followed by Tukey's test. \**p* <0.05 *vs* control; \**p* <0.05 *vs* 24 h 5  $\mu$ M.





**Figure S3.** A) Representative images of scratched cells treated with VEGF-A (80 ng/mL; b, b' and b''), 5  $\mu$ M of (±)-1 (c, c' and c''), VEGF-A plus 5  $\mu$ M of (±)-1 (d, d' and d''), VEGF-A plus 5  $\mu$ M of (±)-1 plus 2  $\mu$ M of Pentazocine (PTZ, e, e' and e''), VEGF-A plus 5  $\mu$ M of (±)-1 plus 2  $\mu$ M of AC927 (f, f' and f''). Untreated cells were considered as control samples (Ctrl; a, a' and a''). Images show cells at the starting points (0 h after scratch: a, b, c, d, e and f), after 24 h (a', b', c', d', e' and f') and 48 h (a'', b'', c'', d'', e'' and f'') from the starting of the assays. B) Wound closure percentage with (±)-1 was quantified by ImageJ software. Ctrl, vehicle control (DMSO). Values are expressed as a mean ± SEM of three independent experiments, each involving three different wells per condition. Statistical analysis was performed using one-way ANOVA, followed by Tukey's test. \**p* <0.05 *vs* Ctrl; #*p* <0.05 *vs* VEGF-A; §*p* <0.05 *vs* the same conditions without agonist or antagonist.





**Figure S4.** A) Representative images of scratched cells treated with VEGF-A (80 ng/mL; b, b' and b''), 5  $\mu$ M of (+)-1 (c, c' and c''), VEGF-A plus 5  $\mu$ M of (+)-1 (d, d' and d''), VEGF-A plus 5  $\mu$ M (+)-1 plus 2  $\mu$ M of PTZ (e, e' and e''), VEGF-A plus 5  $\mu$ M of (+)-1 plus 2  $\mu$ M of AC927 (f, f' and f''). Untreated cells were considered as control samples (Ctrl; a, a' and a''). Images show cells at the starting points (0 h after scratch: a, b, c, d, e and f), after 24 h (a', b', c', d', e' and f') and 48 h (a'', b'', c'', d'', e'' and f'') from the starting of the assays. B) Wound closure percentage with (+)-1 was quantified by ImageJ software. Ctrl, vehicle control (DMSO). Values are expressed as a mean ± SEM of three independent experiments, each involving three different wells per condition. Statistical analysis was performed using one-way ANOVA, followed by Tukey's test. \**p* <0.05 *vs* Ctrl; #*p* <0.05 *vs* VEGF-A; \**p* <0.05 *vs* the same conditions without agonist or antagonist.





**Figure S5.** A) Representative images of scratched cells treated with VEGF-A (80 ng/mL; b, b' and b''), 5  $\mu$ M of (–)-1 (c, c' and c''), VEGF-A plus 5  $\mu$ M of (–)-1 (d, d' and d''), VEGF-A plus 5  $\mu$ M (–)-1 plus 2  $\mu$ M of PTZ (e, e' and e''), VEGF-A plus 5  $\mu$ M of (–)-1 plus 2  $\mu$ M of AC927 (f, f' and f''). Untreated cells were considered as control samples (Ctrl; a, a' and a''). Images show cells at the starting points (0 h after scratch: a, b, c, d, e and f), after 24 h (a', b', c', d', e' and f') and 48 h (a'', b'', c'', d'', e'' and f'') from the starting of the assays. B) Wound closure percentage with (–)-1 was quantified by ImageJ software. Ctrl, vehicle control (DMSO). Values are expressed as a mean ± SEM of three independent experiments, each involving three different wells per condition. Statistical analysis was performed using one-way ANOVA, followed by Tukey's test. \*p < 0.05 vs Ctrl; #p < 0.05 vs VEGF-A; §p < 0.05 vs the same conditions without agonist or antagonist.

### NMR spectra



<sup>1</sup>H (500 MHz, DMSO-*d*<sub>6</sub>) of (*R*)-(+)-MRJF22(oxalate), [(+)-1].



<sup>13</sup>C NMR (125 MHz, DMSO-*d*<sub>6</sub>) of (*R*)-(+)-MRJF22(oxalate), [(+)-1].



<sup>1</sup>H (500 MHz, DMSO-*d*<sub>6</sub>) of (*S*)-(-)-MRJF22(oxalate), [(-)-1].



<sup>13</sup>C NMR (125 MHz, DMSO-*d*<sub>6</sub>) of (*S*)-(-)-MRJF22(oxalate), [(-)-1].

### **HR-MS** spectra





#### HR-MS SPECTRA of (±)-MRJF22.



### HR-MS SPECTRA of (*R*)-(+)-MRJF22 [(+)-1].



## HR-MS SPECTRA of (S)-(+)-MRJF22 [(-)-1].