#### Supplementary Data for

#### USP7 Facilitates SMAD3 Autoregulation to Repress Cancer Progression in p53-deficient Lung Cancer

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#### Supplemental Figures



# Figure S1. Comparable levels of MDM2 are expressed in wildtype and *USP*7 KO H1299 cells.

Immunoblots of MDM2 in the cell lysates from wildtype (WT) and the indicated USP7 KO H1299 cells.  $\beta$ -actin is the loading control.



## Figure S2. Genome-wide profiling of USP7-responsive enhancers in H1299 cells.

Venn diagrams showing the comparison of H3K27ac ChIP-seq peaks identified in wildtype (left panel, two biological experiments) or in *USP7* KO (right panel, two clonal lines: HKO-E2 and HKO-E3) H1299 lines. Peak overlapping analysis was performed using the *mergePeaks* of HOMER software with "-d given" parameter. The common regions of each genomic background were further analyzed for identifying the USP7-responsive enhancers in Fig. 2A and listed in dataset S1.





# Figure S3. USP7 inactivation significantly downregulates the expression of SMAD3.

RT-qPCR assays showing the relative expression levels of *SMAD3* in wildtype (WT) or *USP7* KO (two independent clones: HKO\_E2 and HKO\_E3) H1299 cell lines treated with 1% FBS. Means  $\pm$  SD from three biological experiments are shown. Student's t-test, \*\*\* p <0.001.



Figure S4. USP7 inactivation has no effects on the SMAD3 protein stability. (Upper) Representative immunoblots showing the stability of SMAD3 in wildtype or *USP7* KO H1299 lines. Cells were treated with cycloheximide (CHX, 20  $\mu$ g/mL) for indicated time (h) and cell lysates were subjected to immunoblotting assays with anti-SMAD3 antibody.  $\beta$ -actin as an internal control. Asterisks denote non-specific signals. Calculated half-lives (t<sub>1/2</sub>) of SMAD3 in each cell line are indicated.

(Bottom) Decay curves for SMAD3 protein in the indicated cell lines. The SMAD3 levels were quantitated by the ImageJ software and the level in the control cells (without CHX treatment) was set to 1. Mean ± SD from 3 independent experiments.

### Supplemental Table S1. Oligos used in this study

### RT-qPCR

SMAD2	F: GCTGGCCTGATCTTCACAGT
	R: CCAGAGGCGGAAGTTCTGTT
SMAD3	F: GCTGTCTACCAGTTGACCCG
	R: AGGACCTTGTCAAGCCACTG
SMAD4	F: GGACTGCACCATACACACCT
	R: AATGGGAGGCTGGAATGCAA

### ChIP-qPCR

SMAD3_EN1	F: AACTGCTCCAGAAACTCTCAA
	R: CACATGAAGCCCAAACCTGTG
SMAD3_EN4	F: TCTCTCTATCGCCAACGTGA
	R: TCCTGGCAGGCCTTTCCTTA
SMAD3_EN9	F: GTTGCTTTCGCCTAACTGGC
	R: AGCAAAGGGATCCACAGACG
SMAD3_EN10	F: GGAAGCAGAGTGGTATTCAGCA
	R: TAGGCAACATGGGGAAAATGGA

### For reporter construction

SMAD3_EN1	F: CACATGCTATCTTCACAGTGTGATCGA
	R: GACAGAAAAAGAAAATAATGTTGACTTCAGTTTGCA
SMAD3_EN2	F: GAATCCTGGTTTTCCAAGTGTTTAGAGG
	R: GATCAGGAGGCCTCCAGCAG
SMAD3_EN3	F: TGTGTGCTTGCTCTGAAGATTCCA
	R: TTGCCTCTGTGCTGCCAAG
SMAD3_EN4	F: TTGCCTCTGTGCTGCCAAG
	R: AAAATGATTGCTTCCTGAGGTCTGGATG
SMAD3_EN5	F: GGTCTCCCCTTAAATGTCATCTAAGAGAG
	R: CGCGGGAGGTGGTGG
SMAD3_EN6	F: CTGTTCCCCCAGACCCTG
	R: ATAGCAAGACCTCTTCTCAACAGAAAAATACAAAAA
SMAD3_EN7	F: AAATCAAGGAACATTGCCCCATCTCC
	R: CTCGGTAAGCACCAGCACATCT

SMAD3_EN8	F: CTTGGCCTGTTGGTGGTGG
	R: GAAAGAATCCAACAACTCAGATATGCAAAATTTTACC
SMAD3_EN9	F: CAGTAATTCTGCAGCCTCCCTCAC
	R: CCAGTCCCAGCTGAGATTCAGA
SMAD3_EN10	F: TCCATAGATCTGACTCTGGAAACACCG
	R: CCTGCCTGTGATTTCACAAGTGT

Fig. 1B









## Fig. 5C



### Fig. 6A



Ab: USP7





## Fig. 6B



## Fig. 6C



## Fig. 7B

Ab: USP7



## Fig. 7G





## Figure S1







### Figure S3



Ab: β-actin



Ab: SMAD3