

Supplementary Information

Genetic disruption of *Plasmodium falciparum* Merozoite Surface Antigen 180 (PfMSA180) suggests an essential role during parasite egress from erythrocytes

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Figure Legends

Supplementary Figure 1: PfMSA180 is a conserved protein in *Plasmodium* species causing human malaria. PF3D7- *P. falciparum* 3D7, PmUG01- *P. malariae* UG01, PocGH01- *P. ovale curtisi* GH01, PKNH- *P. knowlesi* strain H, PVP01- *P. vivax* P01. Multiple sequence alignment revealed most conserved residues are in the N-terminal region: residues 1-163, central region: residues 417-731 and C-terminal region: residues 1085-1455. The C-terminal region contains most identical residues (~37%). “*” identical residues; “:” conserved substitutions; “.” semi-conserved substitutions.

Supplementary Figure 2: Immobilised Metal Affinity Chromatography (IMAC) purification of PfMSA180 recombinant proteins. PL- Total lysate, FT- flow through from IMAC column, W1- 10mM imidazole, W2- 20mM imidazole; elution profiles using increasing concentrations of imidazole (50 mM - 500 mM), collected each in 2ml fractions.

(a) Construct 1 (C1) was expressed as soluble protein and was purified from a bacterial pellet supernatant obtained after sonication; (b) Construct 2 (C2), (c) Construct 3 (C3) and (d) Construct 4 (C4) were expressed in inclusion bodies and the protein was purified from these inclusion bodies obtained after sonication.

Supplementary Figure 3: Measurement of the antibody responses in rabbits against recombinant PfMSA180 proteins. (a) C1 PfMSA180, (b) C2 PfMSA180, (c) C3 PfMSA180, (d) C4 PfMSA180. Immunogenicity of recombinant PfMSA180 proteins was analyzed by ELISA. The serum samples were serially diluted and end point titers were assessed. Pre-immune sera were used as controls. High titer antibodies were obtained against all recombinant PfMSA180 proteins. The error bars represent the standard error of the mean.

Supplementary Figure 4: Measurement of the antibody responses in mice against recombinant PfMSA180 proteins. (a) C1 PfMSA180, (b) C2 PfMSA180, (c) C3 PfMSA180, (d) C4 PfMSA180. Immunogenicity of recombinant PfMSA180 proteins was analyzed by ELISA. The serum samples were serially diluted and end point titers were assessed. Pre-immune sera were used as controls. High titer antibodies were obtained against all recombinant PfMSA180 proteins. The error bars represent the standard error of the mean.

Supplementary Figure 5: Putative PfSUB1 processing sites in PfMSA180. The high molecular weight parasite protein MSA180 is predicted to possess potential PfSUB-1 proteolytic cleavage sites¹. The three potential PfSUB1 cleavage sites are depicted along with the predicted MSA180 processing fragments.

Supplementary Figure 6: Schematic representation of the generation of inducible *pfmsa180* gene knockout parasites.

- (a) DiCre recombinase-expressing *P. falciparum* parasites II-3 were transfected with pDC2-Cas9-hDHFRyFCU² with gRNA (guide RNA) sequence and a repair DNA plasmid carrying two loxPint sequences (SERA2 intron with a 34 bp lox P site)³ flanking a 94 bp recodonised ORF segment and having 5' homology region of 271 bp and a 3' homology region of 303 bp. The selected gRNA sequence targets Cas9 to a sequence close to the 5' end of the ORF.
- (b) Homologous DNA repair utilises the linearised repair plasmid as homology template resulting in the insertion of the two loxPint sequences flanking the 94 bp recodonised ORF sequence. On addition of rapamycin a functional DiCre-recombinase catalyses the recombination between the two loxP sites resulting in the deletion of 197 bp including the recodonised ORF fragment. This excision changes the reading frame resulting in early termination of translation for PfMSA180.

Supplementary Figure 7: Transgenic *pfmsa180* DNA sequence.

- (a) Part of the DNA sequence of the transgenic *msa180* ORF before excision: The *pfmsa180* ORF with the loxPint (green) and recodonised piece of DNA (Yellow) flanked by two loxPint sites.
- (b) Part of the DNA sequence of the transgenic *msa180* ORF after excision: Rapamycin induced excision leads to the removal of one loxPint and the piece of recodonised DNA.
- (c) Generation of premature STOP codon after excision: After excision the frame shift in DNA sequence results in the truncation of MSA180 at amino acid 81. Codons are depicted here including the premature stop codon TAG (Red).

Supplementary Figure 8: Purified PfMSA180 recombinant protein fragments (C1-4).

- (a) Purified recombinant PfMSA180 products (C1-4) separated by SDS-PAGE were stained with coomassie brilliant blue. Ni-NTA elutes were pooled, dialysed and concentrated. M-Protein marker, BC- Before concentration, AC- After concentration. 1-7 correspond to

collected fractions of elutes of the purified antigen. The dashed rectangles represent the area of cropped gel images shown in Figure 1b.

(b) Immunoblots stained with anti- hexa-histidine tag antibody followed by secondary alkaline phosphatase-conjugated secondary antibody. The dashed rectangles represent the area of cropped immunoblot images shown in Figure 1b.

Supplementary Figure 9: Expression of PfMSA180 in wild-type but not conditional MSA180 knockout parasites.

Full size immunoblots showing a high molecular weight antigen and its processed protein fragments in wild type parasites (WT), whilst bands are absent in the inducible knockout (iKO) parasites (after rapamycin treatment). Immunoblots were probed with MSA180-specific antibodies raised against recombinant MSA180 (a) Construct 1, (b) Construct 2, (c) Construct 3 and (d) Construct 4. (e) Immunoblot probed with anti-CyRPA antibodies were used as a positive loading control. The dashed rectangles represent areas of the immunoblots shown in Figure 2.

Supplementary Figure 10: PCR to confirm integration of the repair plasmid into the *pfmsa180* locus.

(a) Full size agarose gel picture showing diagnostic PCR products. Presence of correctly sized bands in lane I and II confirms integration of repair the plasmid (including a floxed 94 bp recodonised DNA sequence) in one transgenic parasite clone (clone 1) and differentiates it from wild type parasites (II-3 DiCre). Product sizes in lanes III between clone 1 and wild type parasites (II-3 DiCre) indicate the larger size of 980 bp after integration of the loxP-flanked repair DNA fragments, compared to 774 bp in the wild type parasites. The dashed rectangle represents the part of the agarose gel image shown in Figure 5b.

(b) Full size agarose gel picture showing PCR analysis of successful excision of a floxed recodonised *pfmsa180* sequence after the addition of rapamycin. C- Control DMSO treated parasites, R- Rapamycin treated parasites. Reduction of PCR band size in Lane III of ‘R’ compared to ‘C’ confirms the successful deletion of the floxed piece of DNA after rapamycin treatment. For band sizes please see Figure 5. The dashed rectangle represents part of the agarose gel image shown in Figure 5c.

Supplementary Table 1: List of primers used for the cloning of PfMSA180 constructs and for the conditional gene knockout study

Primer	Sequence
MSA180C1FP	CTATAGGGCCTCGTGC <u>CATATGAATGAGAAAAATAGGAAAGCTATT</u>
MSA180C1RP	CTATA <u>GGGTATAGTCTCGAGAGTTGAATTGGGTGACGAA</u>
MSA180C2FP	GTATA <u>CCCTGAACATTGCATATGAACAAAGAACATCTTAATCTTC</u>
MSA180C2RP	GCTTG <u>TGGCTCGAGATTATATATATTTGGATCATC</u>
MSA180C3FP	CATGTATACGAATGCC <u>CATATGAATAATGTACACGATAACAGC</u>
MSA180C3RP	ACGCTTGG <u>CTCGAGATCATTGTTCACTGTTAG</u>
MSA180C4FP	GTATGTAATGTGG <u>CTAGCAATAAGGAAGAGGATATGAATG</u>
MSA180C4RP	CTGACTACGTG <u>CTCGAGATTCTAAAATCTAGTCATC</u>
MSA180NextFP	ATGGTTCATTGTCC <u>CTTTGTAGTG</u>
MSA180NextRP	GGCAAAGGG <u>TTGTTGATAAGGG</u>
MSA180NintFP	GAAGTT <u>CGATAAGTTATAGACGAGTTC</u>
MSA180NintRP	TATCGAA <u>CTTCTTAGAGTCTTAGGG</u>
MSA180_185guideF	<u>ATTGGAAGAATTAAAGTATGCAT</u>
MSA180_185guideR	<u>AAACATGCATACTTAATTCTTC</u>

Supplementary Table 2: Immunoprecipitation by PfMSA180 polyclonal antibodies.

The antibodies were raised to PfMSA180 Construct 3 (C3) and Construct 4 (C4) and a significant number of peptides derived from MSA180 were observed. The Mass Spectrometry analysis was searched against the Plasmodium database on Uniprot⁴.

Table 2a: List of proteins identified by the polyclonal antibodies

Accession	Description	Unique peptides	PSM
Q8IJQ4	Uncharacterized protein PF3D7_1014100	53	190
Q8I0U8	Merozoite surface protein 1 PF3D7_0930300	45	136
C6KTB4	Acetyl-CoA synthetase, putative PF3D7_0627800	32	71
Q8I0V3	60 kDa chaperonin PF3D7_1232100	21	63
Q8IE67	Phosphoribosylpyrophosphate synthetase PF3D7_1325100	15	41
Q8IKF0	Eukaryotic initiation factor 4A PF3D7_1468700	10	35
Q8IKH8	40S ribosomal protein S3 PF3D7_1465900	8	27
Q8IAX5	40S ribosomal protein S16, putative PF3D7_0813900	6	24
O97266	Eukaryotic translation initiation factor 4E PF3D7_0315100	7	22
C6KT18	Histone H2A PF3D7_0617800	3	17
Q8I542	Calcyclin binding protein, PF3D7_1238100	5	10
Q8IIX0	60S acidic ribosomal protein P1, putative PF3D7_1103100	2	7

Table 2b: List of PfMSA180 Peptides detected in immunoprecipitates with polyclonal antibodies

Peptides Detected with C3 Antibody
IKGNSEEFSFDNELPEQTESFPLNKPQDHEAFYNLK
IHNILKDFNINENIMTNK
IKGNSEEFSFDNELPEQTESFPLNKPQDHEAFYNLKK
SAIDKYVHYEYKR
GNSEEFSFDNELPEQTESFPLNKPQDHEAFYNLK
KIANTIYVNVGQSGINGFFNFFDFREK
KHHTNVYEPNDEEKQNEQK
NIYNMNNVHDTAYYHNSR
LTNNFKENDEGLKNENNINNNEDNQNDNMNIVLGK
NLTEFLENTER
NFYNISNENGDNTFNNNNNNMDNK
NFYNISNENGDNTFNNNNNNMDNKKR
MNGKLPIDDPKNIYNMNNVHDTAYYHNSR
NHMMLSNEQFINKNK
NNSETNENISESNSGNPELNNENSYSVK
LSYFNLP SLK
INYIFFNYIPLENYVNNGDALDFR
VTGDSVENINEQTNNNQYPNTEYNTIQR
VGDQFFPTYSNLGKDDHDLEHSAK
ELAEISTSNLFYPKKDIILR
ELAEISTSNLFYPKK
ELAEISTSNLFYPK
DNNYYYYNSDNNNNYNER
DMPSLEDNFYEHLKYPDINTIHIYYNASPVK
ALLQQSNKDTPIHK
YFPTKDMPSLEDNFYEHLKYPDINTIHIYYNASPVK
YKDNNYYYYNSDNNNNYNER
NLTEFLENTERINTFVR
NHMMLSNEQFINK
YPDINTIHIYYNASPVK
YMAENKFNLPMSSSEVENK
NYKNLTEFLENTER
VDVIDEK
TIIIDEIKSK
ENDEGLKNENNINNNEDNQNDNMNIVLGK
ISENLR
LPIDDPKNIYNMNNVHDTAYYHNSR
DMPSLEDNFYEHLKYPDINTIHIYYNASPVKLNEVNDLK
DMQGNNNIKIEQNK
ANQQFFSYK
MNEFDYINNFSASYLLNQLIIFQDKFNYIK

ITSDILYK
FIPINAFITLENK
IFYINSYR
MVNDTWITPYAFVVYSK
HHTNVYEPNDEEKQNEQK
NHMMLSNEQFINKNKYAK
FNLPMSSEVENK
SAIDKYVHYEYK
Peptides Detected with C4 Antibody
IKGNSEEFSFDNELPEQTESFPLNKPQDHEAFYNLK
IHNILKDFNINENIMTNK
SAIDKYVHYEYKR
GNSEEFSFDNELPEQTESFPLNKPQDHEAFYNLK
NIYNMNNVHDTAYYHNSR
NLTEFLENTER
NFYNISNENGDNTFNNNNNNMDNK
NFYNISNENGDNTFNNNNNNMDNKKR
LNEVNDLK
INYIFFNYIPLENYVNNGDALDFR
VTGDSVENINEQTNNNQYPNTEYNTIQR
VGDQFFPTYSNLGKDDHDLEHSAK
ELAEISTSNLFYPKKDIILR
ELAEISTSNLFYPK
DMPSLEDNFYEHLKYPDINTIHIYYNASPVK
YKDNNYYYNNSDNNNYNER
NLTEFLENTERINTFVR
NHMMLSNEQFINK
NYKNLTEFLENTER
VDVIDEK
TIIIDEIKSK
ENDEGLKNENNINNNEDNQNDNMNIVLGK
ISENLR
DFNINENIMTNK
ANQQFFSYK
MNGKLPIDDPK
ITSDILYK
FIPINAFITLENK
IFYINSYR
FNLPMSSEVENK

References:

1. Withers-martinez, C. *et al.* Plasmodium subtilisin-like protease 1 (SUB1): Insights into the active-site structure , specificity and function of a pan-malaria drug target. *Int. J. Parasitol.* **42**, 597–612 (2012).
2. Knuepfer, E., Napiorkowska, M., Ooij, C. Van & Holder, A. A. Generating conditional gene knockouts in Plasmodium – a toolkit to produce stable DiCre recombinase-expressing parasite lines using CRISPR / Cas9. *Sci. Rep.* 1–12 (2017).
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3. Jones, M. L. *et al.* A versatile strategy for rapid conditional genome engineering using loxP sites in a small synthetic intron in Plasmodium falciparum. *Sci. Rep.* 1–9 (2016).
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4. Consortium, T. U. The Universal Protein Resource (UniProt). *36*, 190–195 (2008).

Supplementary Figure 1:

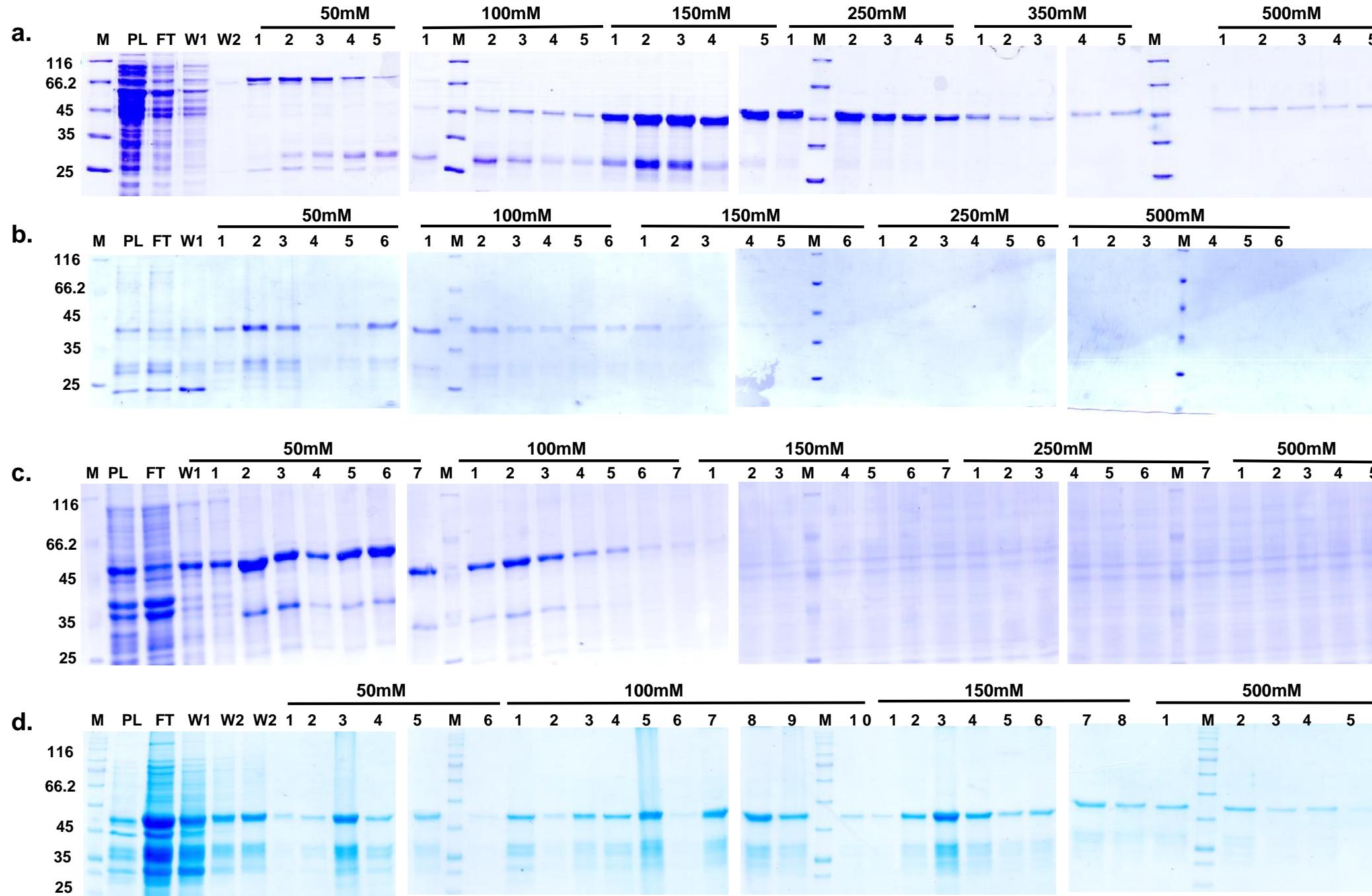
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PoCGH01	MLRIIYFSFFFPLFSLFLISGHDAI-SNVEDKTKKAILLLKNTFIDNEEYKEPNDLN	59
PKNH	MSRITFLFSLSIILFFFLLPGQNAL-TIDDDKNKRATLLALLKNTFIDNKGNKKSDDIKG	59
PVP01	MPRITPLFLLSILLSFFLFGQNAL-TNDDDTNKRATLLALLKNTFIDNTENKKPDDINT	59
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PF3D7	ALEHIQNSELYPKDSKKFDKFIDEFFSYNNIHVNFTDEEKRILHISGVFKEFYVDVDNLN	120
PmUG01	ALENIINNMKLHPTDNDKFDFLDALFKHHNIYVTLDDHKRRIIHISGVLFYVDVDTLT	119
PoCGH01	ALENIINNMNIHPTDNKKFDNFLEELFKHYNVHVTFSMDKDKRVLHLSGVNLNDVYVDVDSL	119
PKNH	ALENIKNMTLQPTDTDKFDKFLDQFLKFQIYVTFSDKDKRVLHLSGVNLNEVYVDVESLS	119
PVP01	ALENIINNMTLHPTDTDKFNKFLDHFLKFHIYVSFSDKDKRVLHLSGVNLNEVYVDVESLS	119
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PmUG01	EEKQKEYFNTFYEKGHTLINLILHSNLIHTKHDKDVKENHKEETKPNHPDSPIPDEEPN	179
PoCGH01	KGNTKEYFNGIHKKALSLINLVLHSNLVHPKYAEINIEKGSQTGNDLQENS----QD	173
PKNH	KENLQKHFDSDLYEKGLNLINLIVHSNLVHPKYDETEMHGVDAAEKDH-----	166
PVP01	EENLQKHFDHSVHEKGLNLINLIVHSNLVHPKYDETAVGMEGEPEQ-----	166
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PF3D7	-----	163
PmUG01	LEKEPIPDDVSKGEVNHQLNYNNVYDSDIEVESNHEVEPNHEVEPNHAVEPNHA	239
PoCGH01	TEG-----EPKGN-----TGEGHEPVYDFKNNQGGEPDNAYGHI--	207
PKNH	VS-----	168
PVP01	TDG-----PTEGP-----GSHG-----KVDPPQEGDPQEEEGPSQE	197
PF3D7	-----	163
PmUG01	VEPNHAVEPNHATEPNHATEPNPENEPIPDDVSKGEVNHQLNYNNVYDSDIEVESNHEVE	299
PoCGH01	-----	207
PKNH	-----	168
PVP01	GHPQE-----	203
PF3D7	-----	163
PmUG01	PNHEVEPNHAVEPNHAVEPNHATEPNHATEPNHATEPNHAVEPNHATEPNPENE	359
PoCGH01	-----QGEEPVHEYEQNQGGEPDNACEHIQGEEPVH---EY---EY-----N	243
PKNH	-----DNYAENIKGPAHYEESQ---EY-----E	188
PVP01	---VDPTQENHPHEKVDPPQEGDPAQESHPHQKDGPAQQDHSQ---EY-----A	246
PF3D7	-----	163
PmUG01	PIPDDVSKGEVNHQLNYNNVYDSDIEVEPNHAAEPNPENEPIPDDVSKGE-VNHQLNYNN	418
PoCGH01	QGE-----EPVHEYEQNQG-----EEP-----VHEYEQNQGEEPVHEYEQNQ	280
PKNH	EMP-----EYADHYHYGGN-----KED-----PDDMDYENGEEYDAQKYQDD	225
PVP01	VTP-----EYADHYHYGGH-----EED-----PEDMDYENGEEYDAQGDPPD	283
PF3D7	-----	163
PmUG01	VYGSDHEDETNHETEFKQEETHNYEAKQED--EHKEHYEHSHTEPEKEVAHSYEAH	476
PoCGH01	G-----EEPVHEYEQNQGEEPVHEYEHNQEGESDN-----A-----YEL-YHGGD	319
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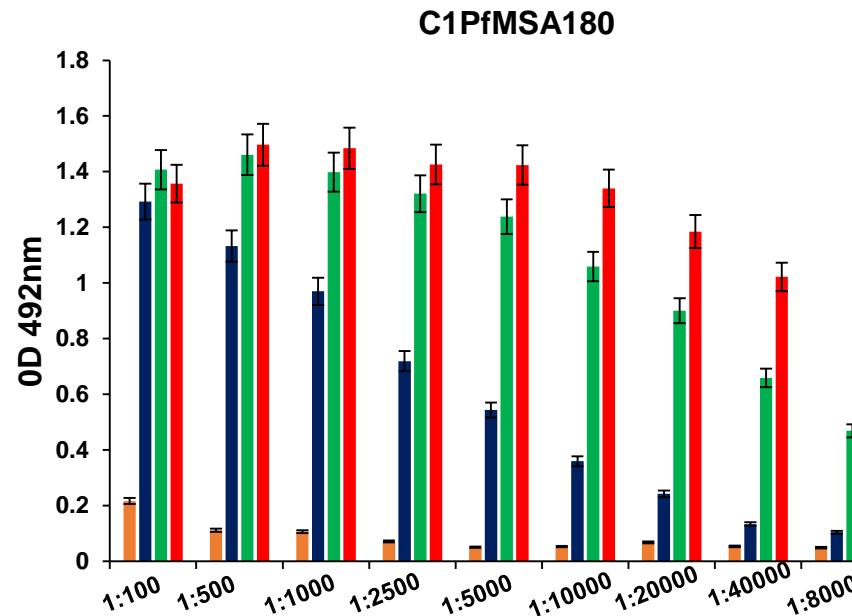
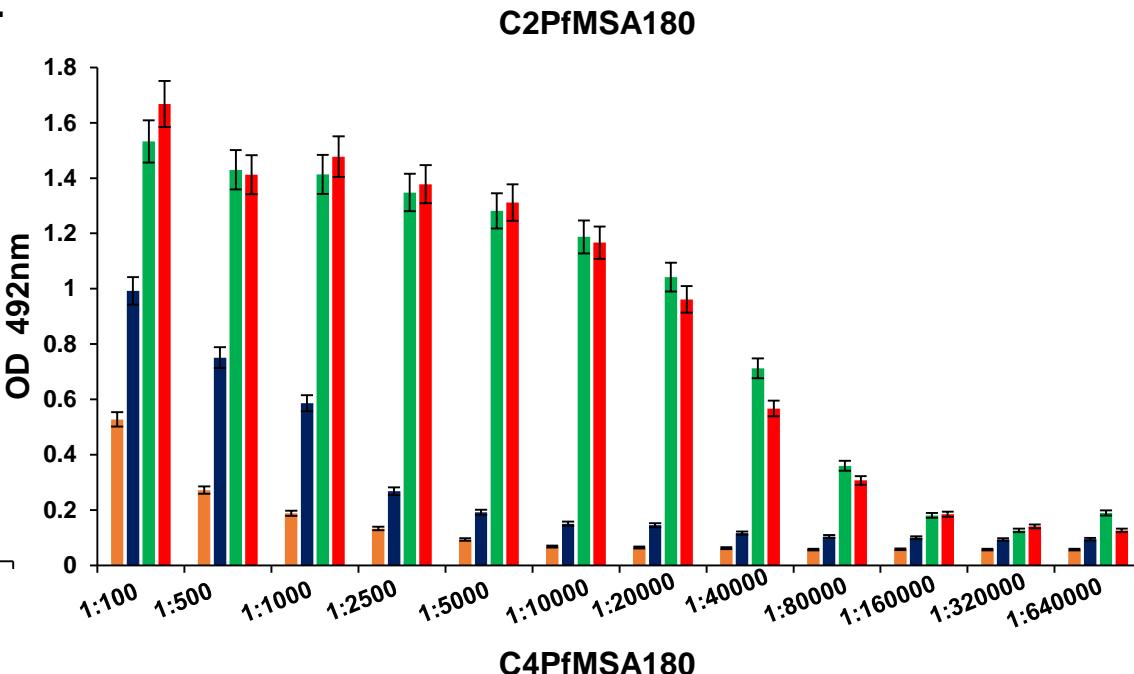
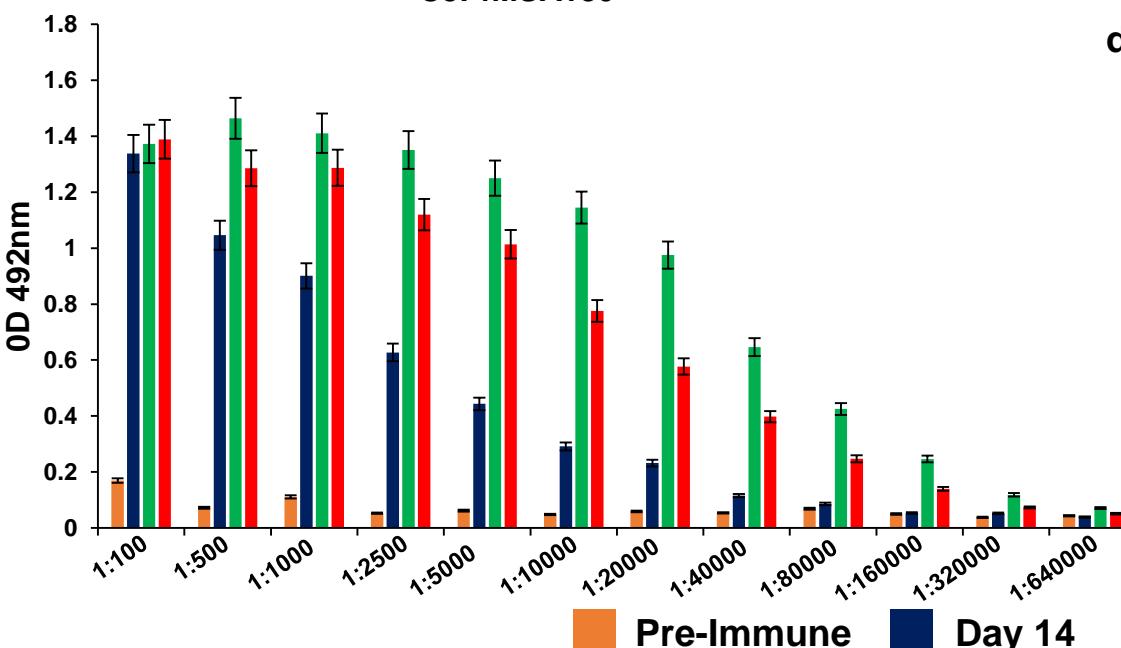
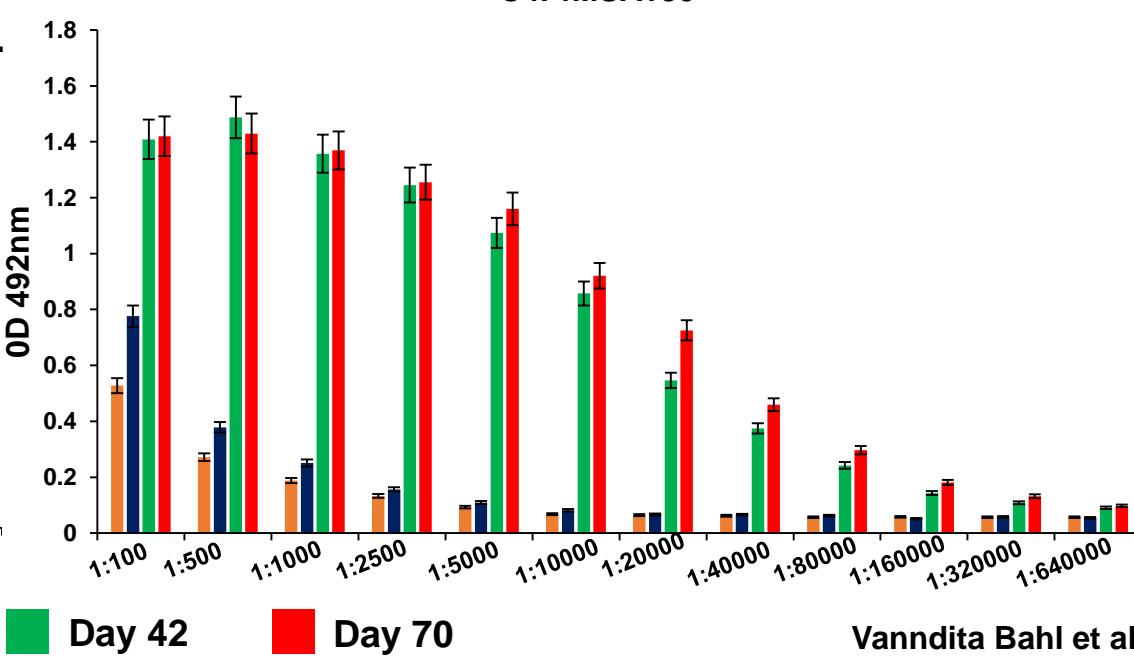
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PoCGH01	I-----TEYYDNEGH---NEENYKNKNIKYVGKKLNLLAMKNIKLNNNSTGEFKV	368
PKNH	N-----DHNDHDDHNDHNDHDEEEKQKIKYVGKKLNLLAMKNIKLNNNSTGEFKV	325
PVP01	-----EHDEHDEHGEHDEPYHEEDKKKIKYVGKKLNLLAMKNIKLNNNSTGEFKV	342
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PmUG01	NFYTNVNYINQPPSNS-GSNHQEKYEYTEINQESKKKKKS-----D	578
PoCGH01	NFYTNVNYIS-PYSINPFPSQDMHKDNKVYEEGRNGKEEKYDEYSGGVSDGDK--DDS	425
PKNH	NFYTNYMNYINTPYVEPPFPFHLDIYEYAEVYSGDKIYPKD-----	366
PVP01	NFYTNVNYINTPYGAPLLPFHKDSYEYAEVYSGDKLHPKK-----	383
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PF3D7	HTEEDNVVV-----SATKGNQKEETEK-KENHENNAINPKYMN-----	291
PmUG01	KMDNSYFVKEVPTVVGDEG-----EGDDIVNTNDIKQVNENLYNTNKAYDHMYNK	630
PoCGH01	SGSHGGYYAGK-----EEGETEDGYNKRNAIRNIYEKMNNQSE-EEHEYEE	471
PKNH	HKDDNIYYGGENELIPQHVKGEMQKEALQEGSYHGYKSTMKGMYENIKRSG-KK-----	420
PVP01	HGDEQMYYVGEKELIPVHGKGDMQK---EGPYDVYKGAMKGIYENIKKAA-KK-----	433
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PF3D7	-----YETYYKKIFNAIFEQIDKLNLFEIKKNKNSETNEN-----	328
PmUG01	K-----IEGMEEKANKYYPYDDSS-----EYA-----DNARYLKKGN---	662
PoCGH01	NAGEEQGKEGNKEWERDWYKDENSE-----G-----ERCKRDEHFV-----	507
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PoCGH01	-----EEETEYAD-SYFDGEDDV-NRKYTEEQTEYSOKDDVEGMQLGNK	550
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PmUG01	KPGKSEAKASHFPYTYYNPYYMYSLNSTGSPKYNNSKYNNGYTNGHNNEYEYSNKYN	762
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PKNH	APAEKEKKGIDFSYTYHNPYMFKLGSNMPGKKAQP-SKGAPAKGGL-----	538
PVP01	APSEKGKKGIDFAYTYYNPYMFKLGSNMPGKKAQP-SKGAPAKGGL-----	578
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PF3D7	-----NN--EDNQNDNMN-----	406
PmUG01	DKYNDKYNKYSNKYSNKYNNEYNNEYNNEYNNEYNKYNNEYNKYNNEYNKYN	822
PoCGH01	-----QEKLVGE-----EKWK-----	607
PKNH	-----GGKDDKEEEEEEVDD-----	553
PVP01	-----GGKGHDEEEEVADE-----EEEEEE-----	599
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PmUG01	NEYNNKYNKYNKYSDKYSDRYKKYNNNNYNNYNKKQNKKSLYDFMLQKKALEKE-HML	881
PoCGH01	-----D-----	608
PKNH	-----	553
PVP01	-----EAEEVADAE-----DVADEDAEEVADED-ADV	626

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PmUG01 NNDSGNNNDNIKLSISKNENITKHSRNSIDKYLFYEYKKISNNILQYYDDLNSKIPQFKDN 1722
PocGH01 NK-A--NSKYKLKISQNVHITKYSRNAIDKYIYYEYKKISNNIIQYYEELNPKIHEYSD 1477
PKNH PSKD--LNSYKLKIAQNEFITKFSRSAIDKYMYYEYRKISNNIVQHHVELSPKLGENLAE 1415
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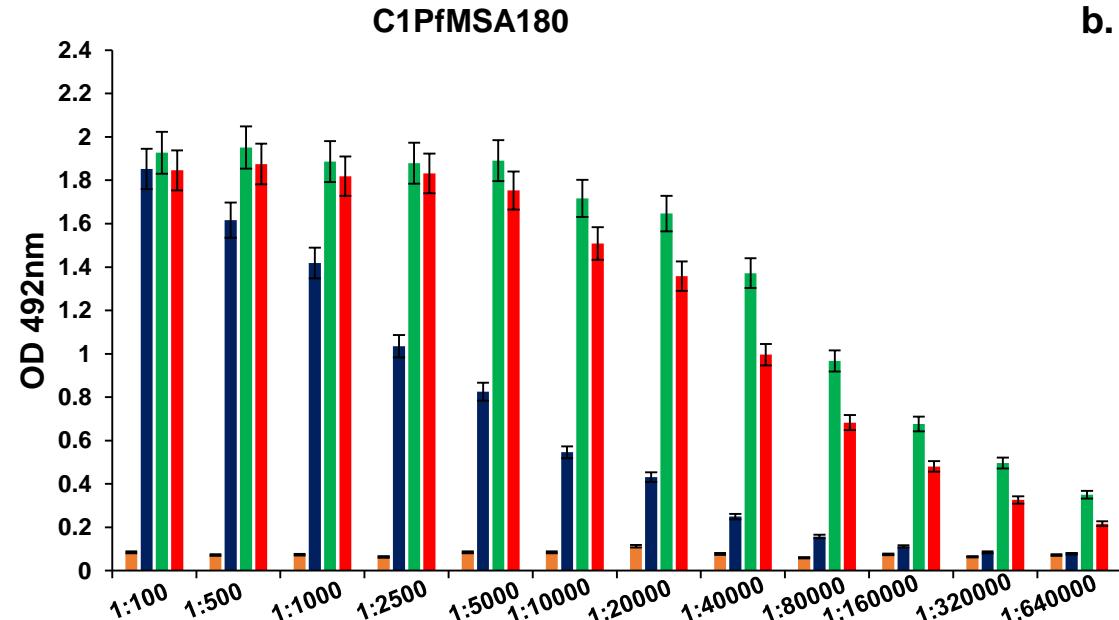
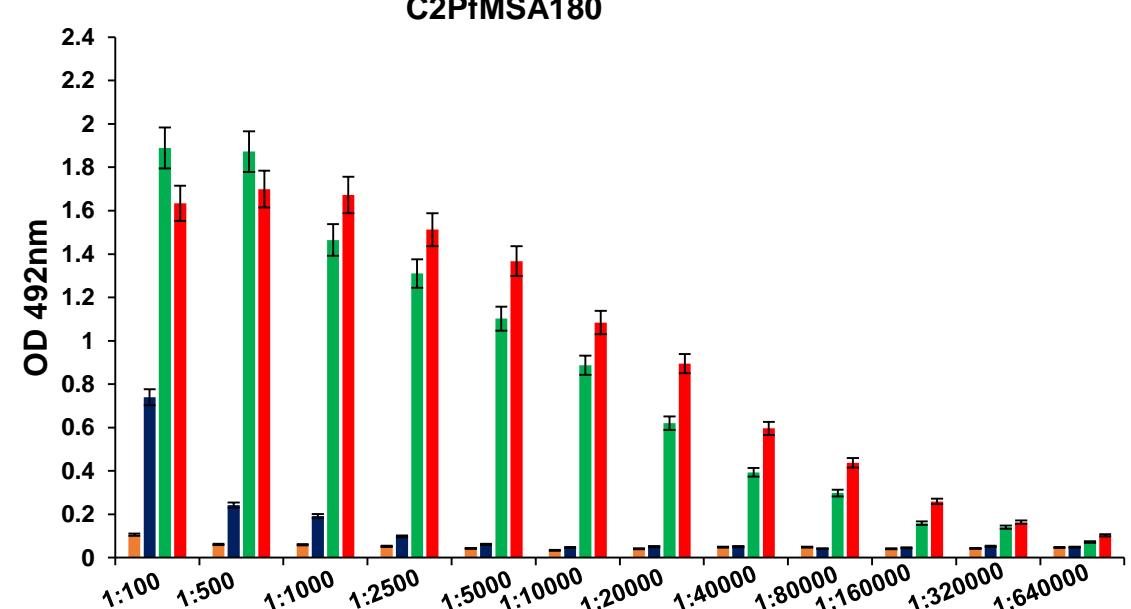
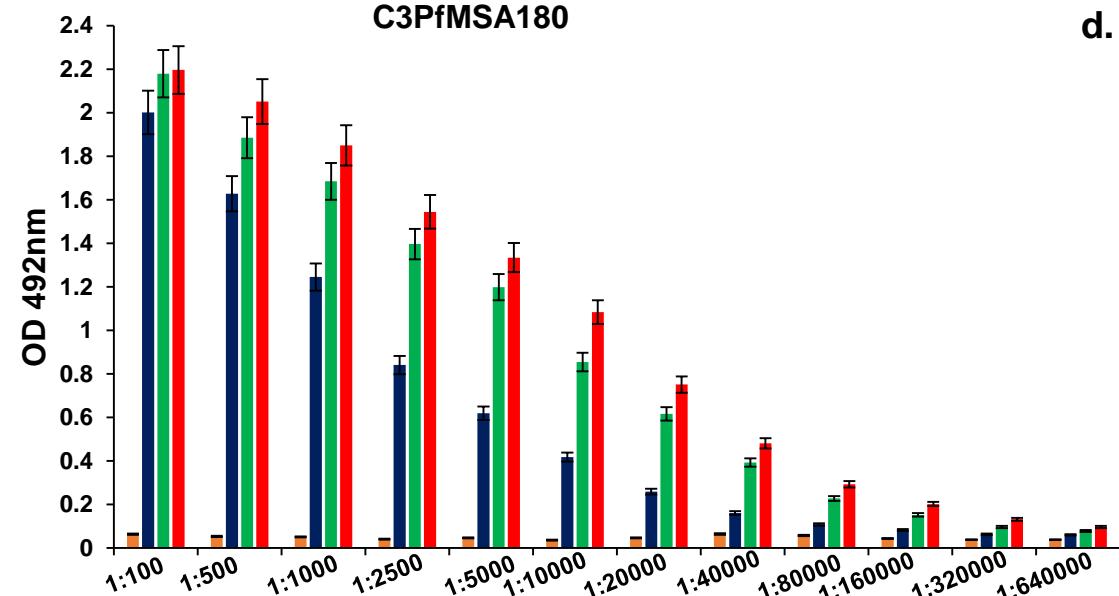
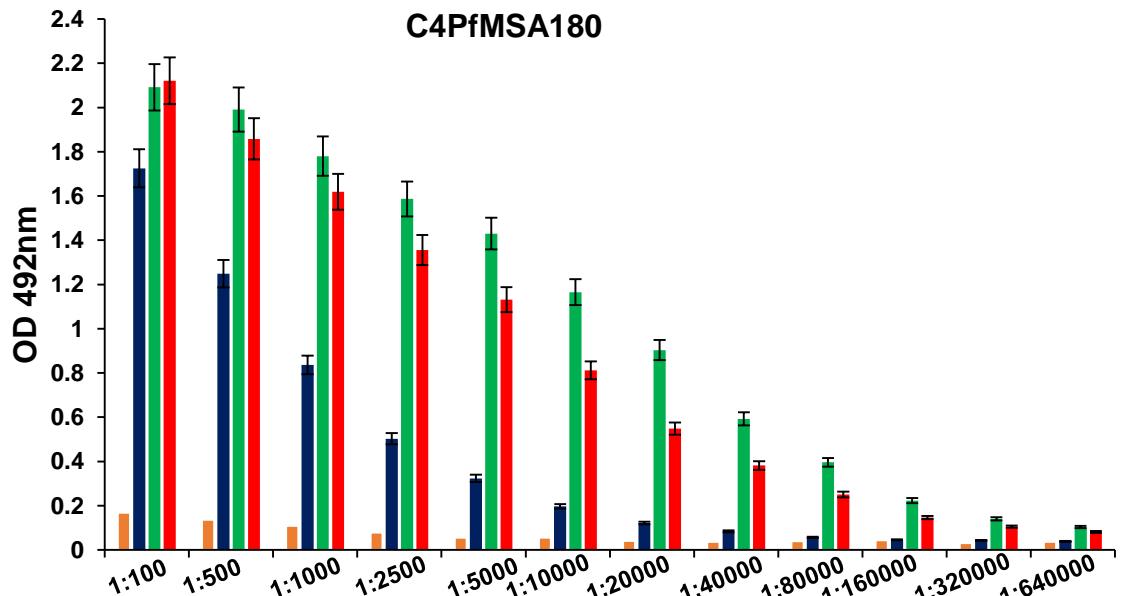
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PKNH FIILDEGVNYLFFNYAPNENHINEEA----- 1501
PVP01 FIILDEGVNYLFFNYVPNENHINYAA----- 1596
*: * : : * : * : * : * : * : * : * : * : * : * : * : * : * : * : * : * :



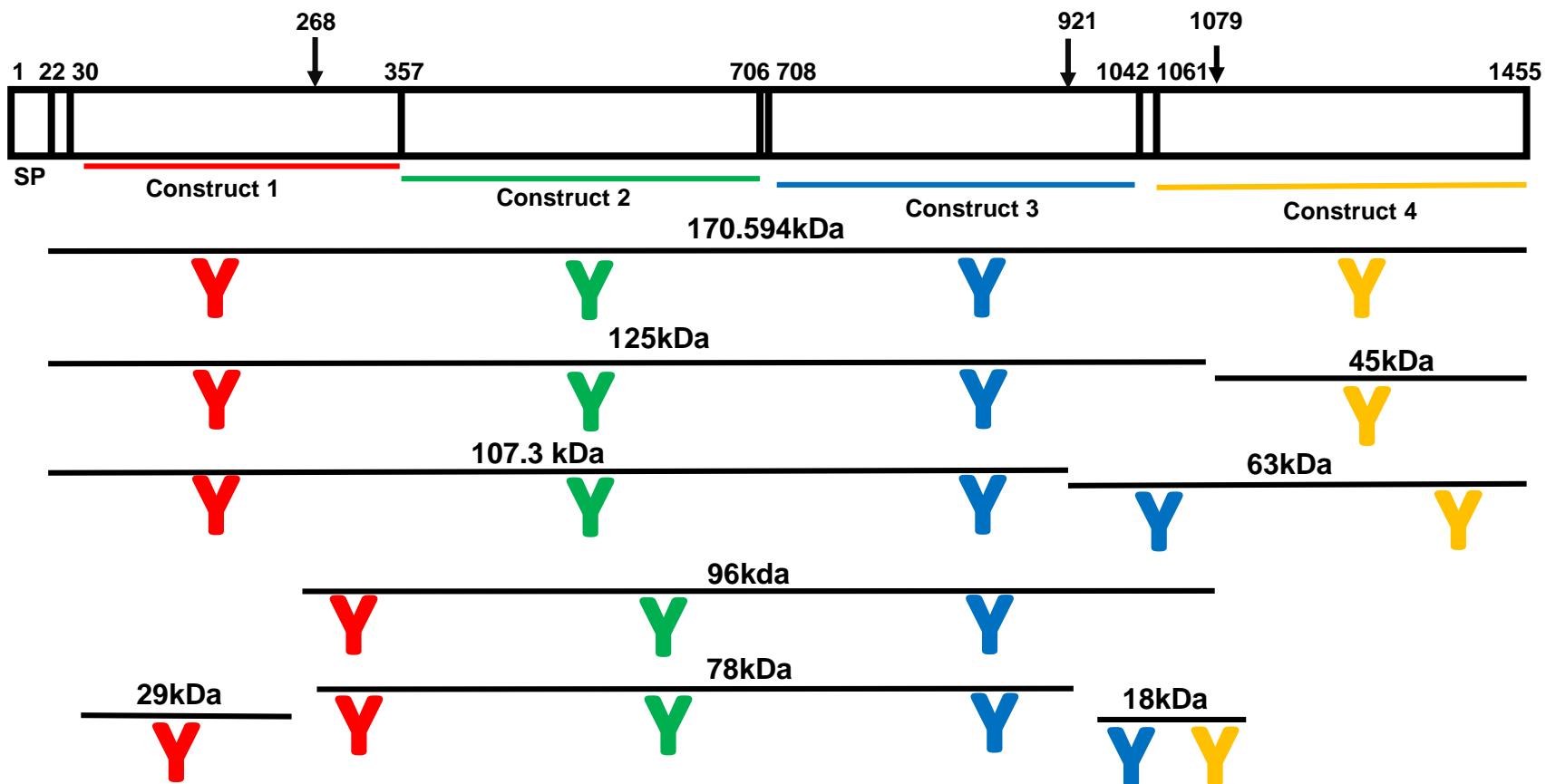
a.**b.****c.****d.**

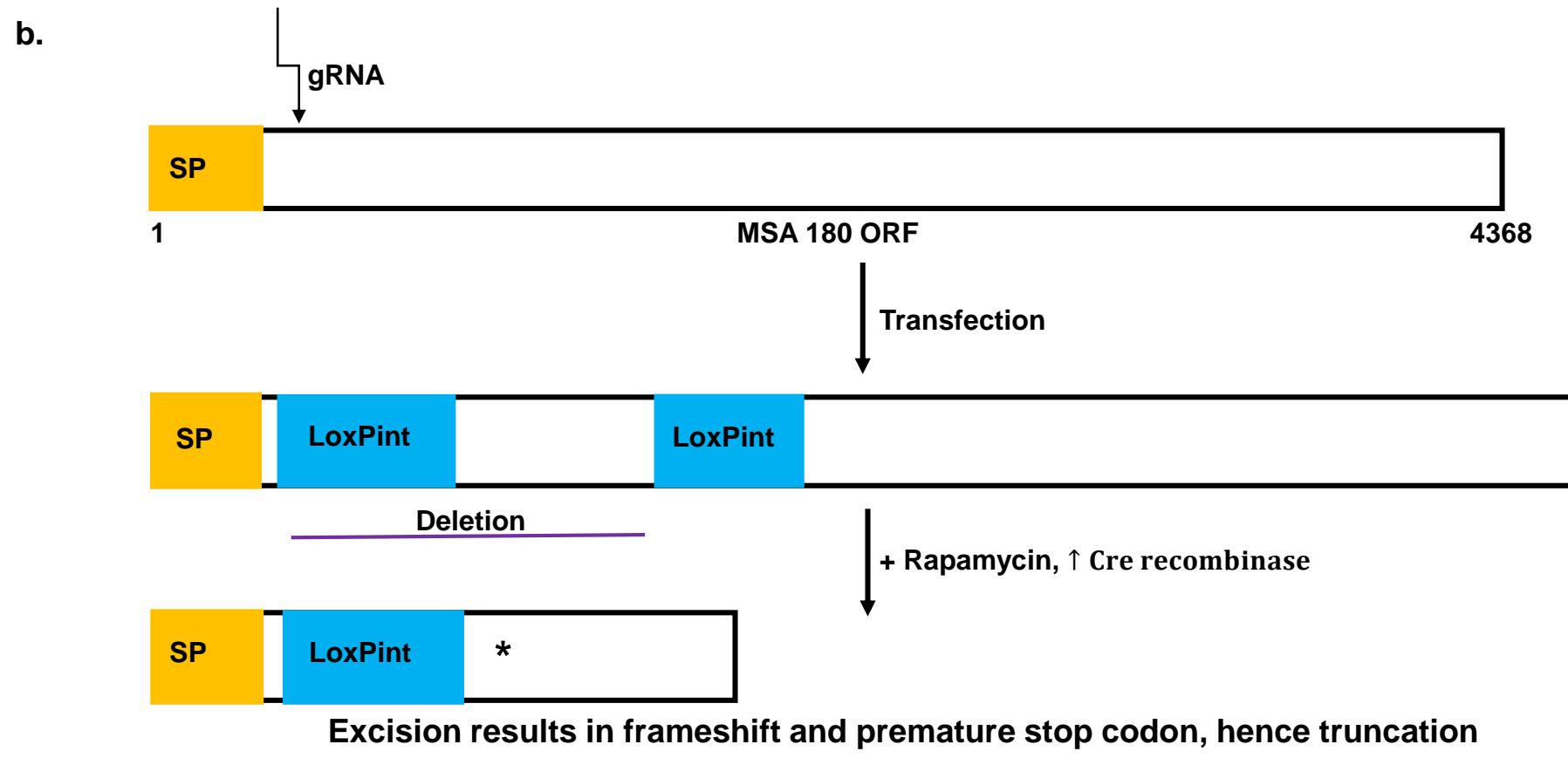
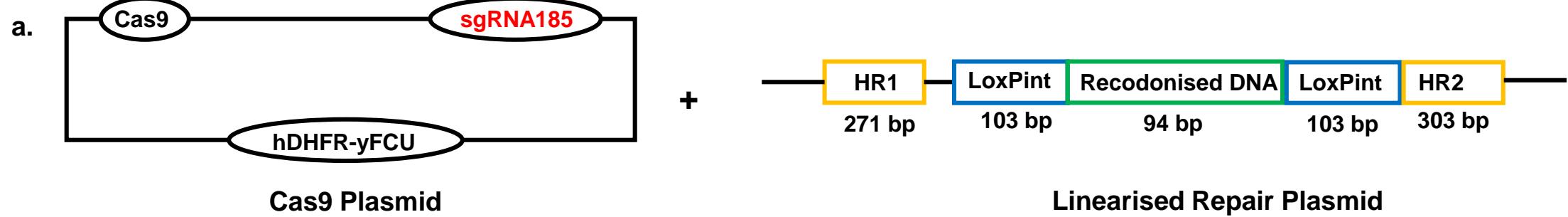
Pre-Immune Day 14

Day 42 Day 70

a.**b.****c.****d.**

Pre-Immune Day 14 Day 42 Day 70





a. SEQUENCE BEFORE EXCISION

ATGAATCGAATATTTATTTGTTTACTATTTGGTTATCTCTGTATCTGGTAAAATGTTAATAATAAAAAACTGTAATGAGAAAAATAGGAAAGCTATTTAT
TAGCTTATTAAAAAATTCAATTAGTAGATAATAAGGATTATAACAATAGTGAAGAATTAAAGTATGCTCTTGAGCACATACA **gtaaataaaaaataatacaATAACTTCGTATAG**
CATACATTATACGAAGTTAtatatatgtatatatatatttatattttatattcttttag GAACAGTGAGCTACCCCTAAAGACTCTAAGAAGTCGATAAGTTATAGACGAGTTCTC
TTACTACAACATACACGTTAACCTCACAGAC gtaaataaaaaataatacaATAACTTCGTATAGCATACATTATACGAAGTTAtatatatgtatatatatatttatatttttag
GAAGAAAAAAGAATATTACATATATCAGGTGTCTCAAAGAATTTATGTAGATGTAGATAATTAAATAAGATGAAATGAAAGAATATTTAAGAAAAATTATGAAAAAGG

b. SEQUENCE AFTER EXCISION

ATGAATCGAATATTTATTTGTTTACTATTTGGTTATCTCTGTATCTGGTAAAATGTTAATAATAAAAAACTGTAATGAGAAAAATAGGAAAGCTATTTAT
TAGCTTATTAAAAAATTCAATTAGTAGATAATAAGGATTATAACAATAGTGAAGAATTAAAGTATGCTCTTGAGCACATACA **gtaaataaaaaataatacaATAACTTCGTATAGC**
ATACATTATACGAAGTTAtatatatgtatatatatatttatattttatattcttttag GAAGAAAAAAGAATATTACATATATCAGGTGTCTCAAAGAATTTATGTAGATGTAGATAATT
AAATAAAGATGAAATGAAAGAATATTTAAGAAAAATTATGAAAAAGG

c. PREMATURE STOP CODON AFTER EXCISION

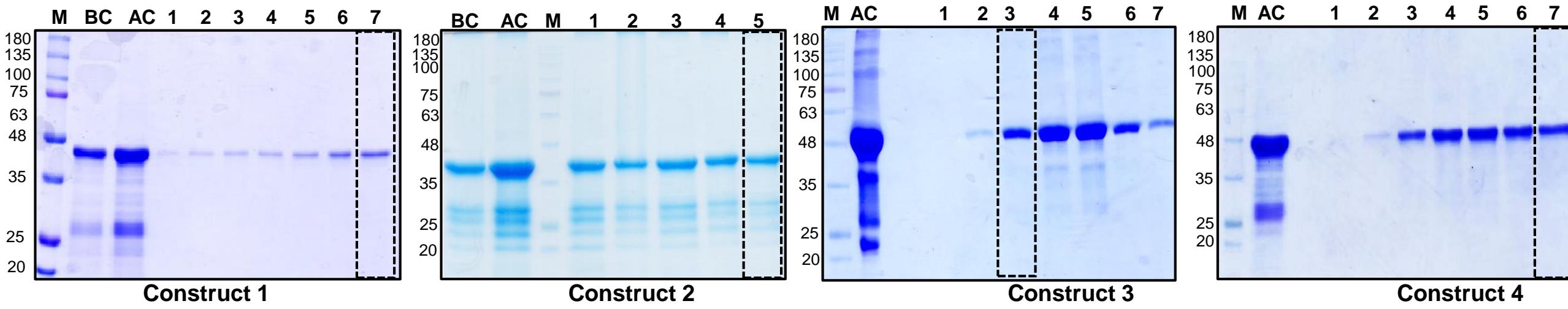
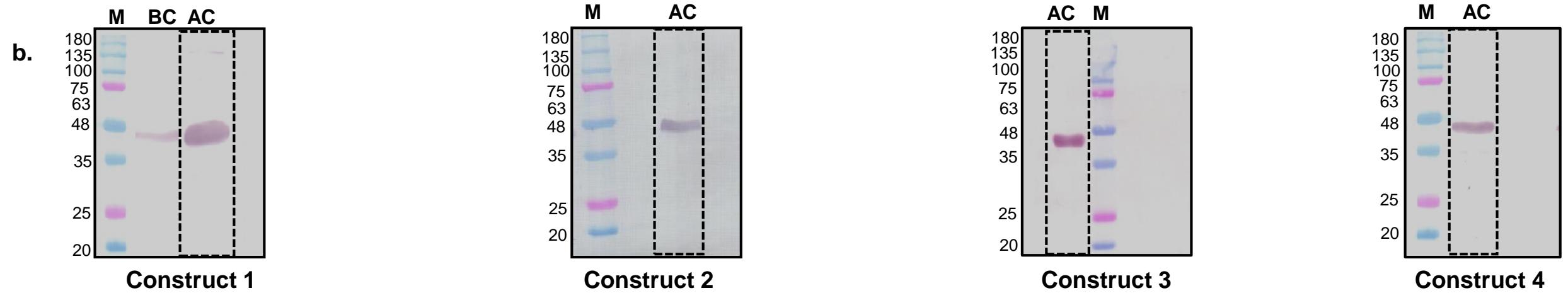
ATG AAT CGA ATA TTT TAT TTT TGT TTG TTT ACT ATT TTG TTT TGG TTA TCT CTT GTA TCT GGT GAA AAT GTT AAT AAT AAA AAC TGT AAT GAG AAA AAT AGG
AAA GCT ATT TTA TTA GCT TTA AAA AAT TCA TTA GTA GAT AAT AAG GAT TAT AAC AAT AGT GAA GAA TTA AAG TAT GCT CTT GAG CAC ATA CAG AAG
AAA AAA GAA TAT TAC ATA TAT CAG GTG TCT TCA AAG AAT TTT ATG **TAG** ATG TAG ATA ATT TAA ATA AAG ATG AAA TGA AAG AAT ATT TTA AGA AAA ATT ATG
AAA AAG G

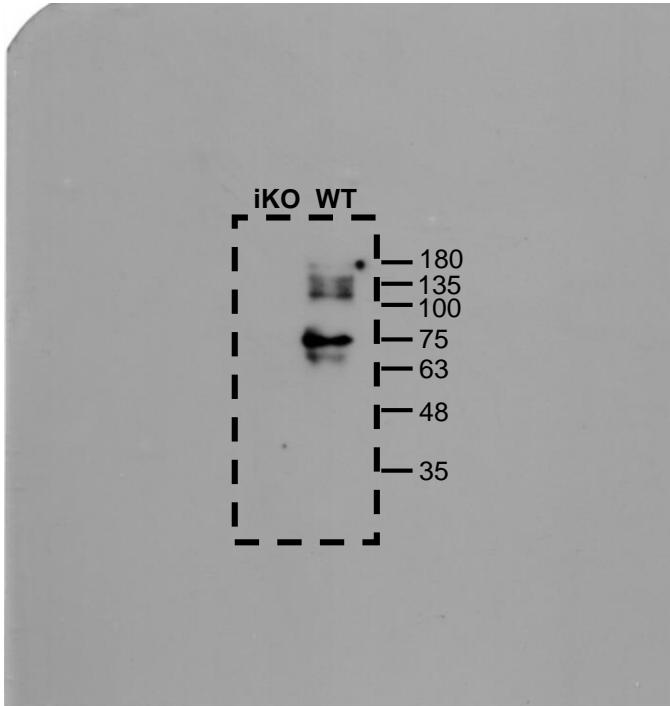
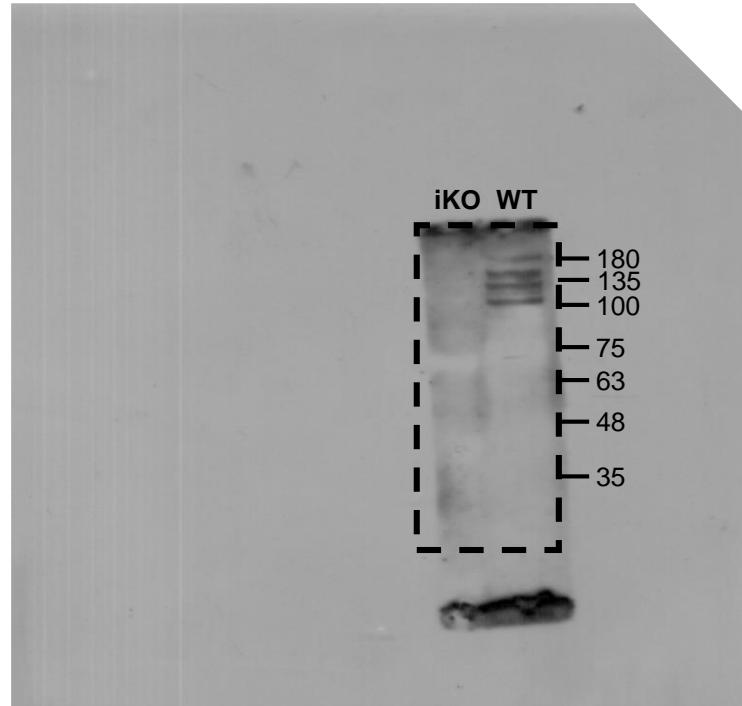
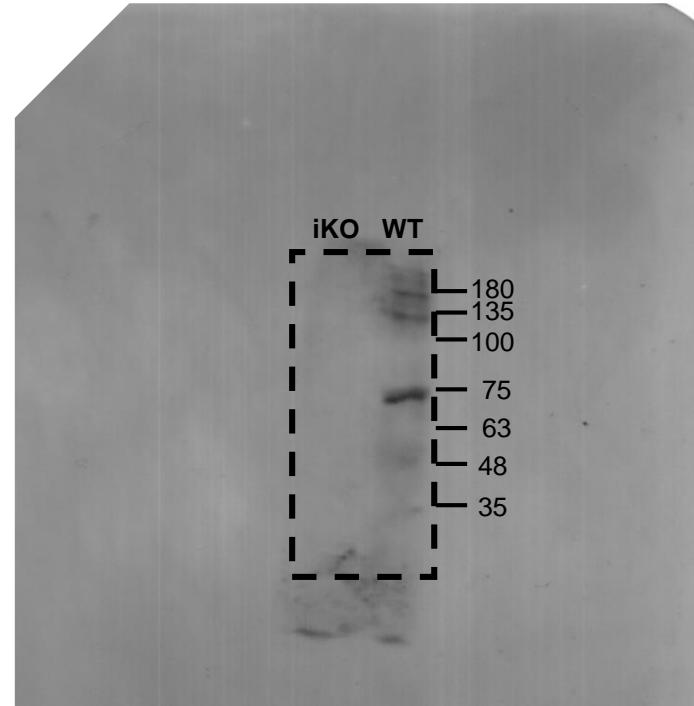
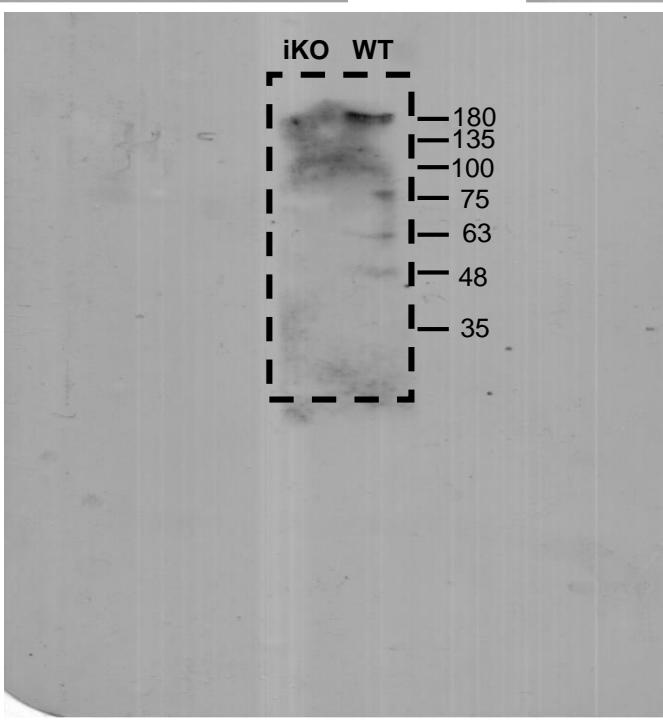
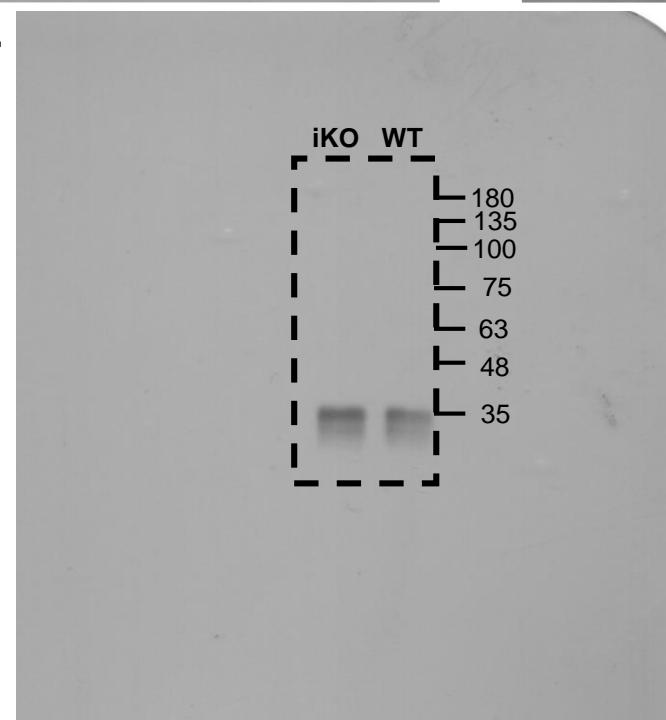


LoxPint sequence (ATAACT-----TTAT: LoxP Sequence)

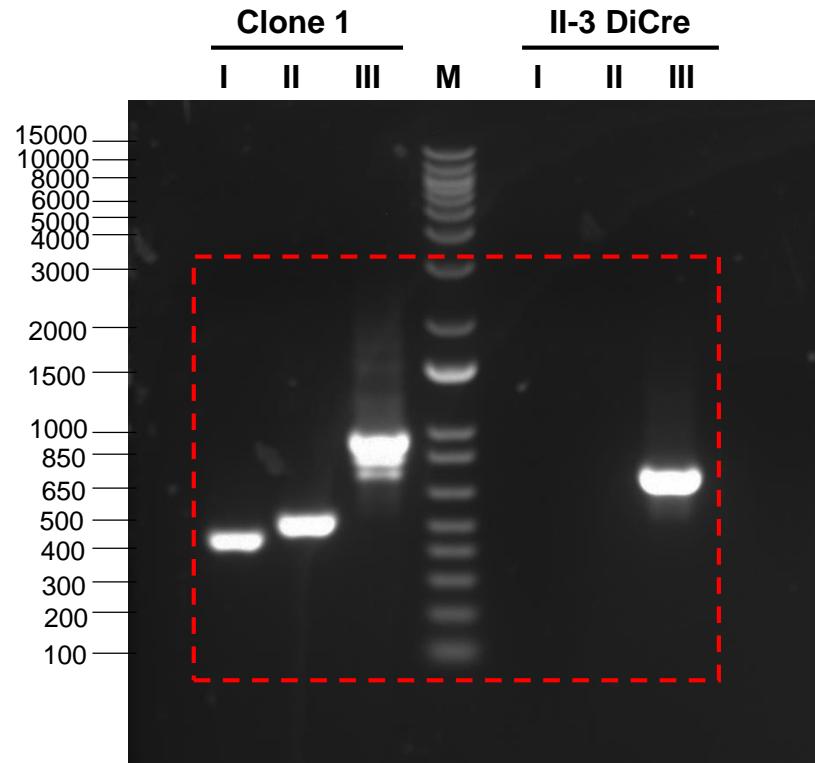


MSA180 Recodonised DNA

a.**b.**

a.**b.****c.****d.****e.**

a.



b.

