Supplementary information

Structural basis for UFM1 transfer from UBA5 to UFC1

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Structure	UFC1-UBA5 (389-404)	UBA5 (347-404)-UFC`1	UFC1 Y110A&F121A
Beamline	ESRF ID23	BESSY II BL14.1	XtalLab Pro (Rigaku) Pilatus 200K detector
			(Dectris)
Wavelength (Å)	0.9720	0.9763	1.54
Space group	P212121	P64	P 41212
Unit Cell a, b, c (Å), α, β, γ (°)	47.1, 67.2, 133.8, 90, 90, 90	84.6, 84.6, 60.8, 90, 90, 120	46.16 46.16 143.76 90 90 90
Resolution range (Å) ^a	33.71 – 2.40 ^b (2.47-2.40) ^b	46.78-2.65 (2.78-2.65)	33.24 - 2.20 (2.28 - 2.20)
Total reflections ^a	297,196 (14,591) ^b	149,370 (20,269)	10,416 (917)
Unique reflections ^a	14,084 (704) ^b	7,314 (953)	8,488 (804)
Completeness sperical (%) ^a	81.3 (49.4) ^b	100.0 (100.0)	99.82 (99.14)
Multiplicity ^a	21.1 (20.7)	20.4 (21.4)	1.2 (1.1)
R _{meas} (%) ^{a,c}	13.6 (142.9) ^b	11.2 (600.2)	15.3 (71.9)
/<0(1)> ^a	16.9 (2.5) ^b	13.1 (0.6)	13.3 (2.2)
CC _{1/2} ^{a,d}	0.999 (0.868) ^b	0.990 (0.292)	0.99 (0.47)
Wilson B-factor ^e (Å ²)	52.1 ^b	127.1	25.44
Refineme	ent statistics		
Rwork	0.219	0.219	0.205
R _{free}	0.288	0.264	0.256
No. of protein monomers in a.u	2	1	1
Number	of atoms		
Macromolecules	3,000	1,488	1,316
Solvent	52	-	59
Number of protein residues	354	183	164
RMS bond lengths (Å)	0.007	0.004	0.013
RMS bond angles (°)	1.59	1.45	1.82
Ramachandran favored (%) ^f	95.7	86.7	97.5
Ramachandran allowed (%)	4	13.3	2.5
Ramachandran outliers (%) ^f	0.3	0.0	0.0
Clashscore ^f	8.9	21.2	10.23
Average B-factor protein (Å ²)	60.1	157.1	28.77
Average B-factor solvent (Å ²)	40.6	-	33.38

Supplementary Table 1: Data collection and Refinement statistics of crystal structures

^aValues for the highest resolution shell are given in parentheses

^b Values are given for data subjected to anisotropic ellipsoidal truncation using the STARANISO server¹

^c $R_{meas} = \Sigma_h [m/(m - 1)]^{1/2} \Sigma_i |I_{h,i} - \langle I_h \rangle | / \Sigma_h \Sigma_i I_{h,i}$ ^d CC_{1/2} is defined in ² ^e Wilson B-factor was estimated by SFCHECK³. ^f The Ramachandran statistics and clashscore statistics were calculated using MOLPROBITY.

Supplementary Table 2: Thermodynamic parameters of binding experiments measured by isothermal titration calorimetry

Experiment	[Syr] (M)	[Cell] (M)	N1 (sites)	N1 Error	N1 Dependency	KD1 (M)	KD1 Error (M)	KD1 Dependency	N2 (sites)	N2 Error	N2 Dependency	KD2 (M)	KD2 Error (M)	KD2 Dependency
UBA5 (Δ349-389) into UFC1	1.50E- 03	1.50E- 04	1.07	0.131	0.995	5.50E- 06	8.69E- 07	0.887	6.80E- 03	0.139	1	8.00E- 07	7.55E- 07	0.996
UBA5 (333- 404) into UFC1	1.60E- 03	1.60E- 04	1.26	4.6E- 02	0.989	2.54E- 06	8.67E- 08	0.659	1.4E- 02	4.6E- 02	0.996	5.64E- 08	3.98E- 08	0.601
UBA5 (Y372A) into UFC1	8.7E- 04	9E-05	6.8E- 02	8.9E- 02	0.997	1.22E- 07	1.02E- 07	0.969	1.11	8.3E- 02	0.992	1.94E- 06	1.57E- 07	0.805
UBA5 1- 404 M401R into UFC1	9.00E- 04	1.00E- 04	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
UBA5 1- 404 L397R into UFC1	9.00E- 04	1.00E- 04	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A

Supplementary Table 3: predicted pKa and buried surface area (BSA) of the active site cysteine in E2 proteins

Protein	PDB code	Catalytic Cysteine	BSA	рКа
UFC1	2Z6P	116	4%	8.51
UBC9	1U9A	93	36%	11.28
UBCH5C	1X23	85	26%	11.33
UBC12	1Y8X	111	31%	10.91
UBCH7	4Q5H	86	18%	9.57
UBCH5B	3TGD	85	23%	11.13
UBC13	1JBB	87	16%	9.86
UBCH5A	2C4P	85	37%	10.79
UBCH10	4YII	114	17%	9.13
UBE2K	1YLA	92	24%	10.27
UBCH6	5LBN	131	21%	10.25
RAD6B	2YBF	88	23%	9.25
UBE2F	3FN1	116	37%	9.49
UBC7	2CYX	89	49%	9.47
UBE2H	2Z5D	106	19%	9.02
UBE2S	5BNB	95	30%	9.28
UBE2T	1YH2	86	31%	9.77
UBE2U	1YRV	89	31%	10.44
E2E2	1Y6L	85	22%	9.56
UBCH8	1WZV	85	18%	9.63
UBC1	2AAK	88	15%	9.47
RAD6A	6CYO	88	22%	9.86
CDC34	3RZ3	93	31%	10.01
ATG3	2DYT	234	35%	10.29

Supplementary Table 4: Cavity volume in selected E2 proteins

Protein	Volume (Å ³)	Residues defining the cavity
UFC1-2Z6P	377	T93, A94, P95, E96, I97, Y110, K114, I115, C116,
		L117, T118,
		D119, F121, K122, L137, G141, L142
UFC1 Y110A and	126	P95, E96, I97, I115, I117, K122, L137
F121A		
UBC13-1JBB	163.1	P67, K68, V69, R85, I86, L91, K92
UBC12-1Y8X	110.4	P91, K92, V93, K94, V110, L115, R116
UBC9-1U9A	No proper cavity;	V92, C93, L94, L97, E98
	16.6	
UBCH5B-3TGD	No cavity	
UBCH57-4Q5H	66.1	K67, I68, Q84, V85
UBCH5C-1X23	No cavity	
E2E2-1Y6L	No cavity	
UBCH8-1WZV	No cavity	
UBC1-2AAK	No cavity	
RAD6A-6CYO	No proper cavity:	P68, T69, L92, Q93
	5.0	
UBCH5A-2C4P	No cavity	
UBCH10-4YII	No cavity	
UBE2K-1YLA	No cavity	
UBCH6-5LBN	No cavity	
RAD6B-2YBF	163.8	K66, P67, P68, T69, V70, R71, S86, I87, L92, Q93
UBE2F-3FN1	89.1	P95, P96, K97, V98, E114, I115, L120
UBC7-2CYX	96.3	P69, K70, M71, Y83, R87, V88, C89, L93, H94
UBE2H-2Z5D	98.2	R73, D75, S86, I87, G88, M90, S102, T104
UBE2S-5BNB	198.1	L64, P74, P75, K76, G77, E93, I94, C95, V96, L99,
		K100, V112, I116
UBE2T-1YH2	No cavity	
UBE2U-1YRV	82.8	P68, V69, V70, Q87, P88, L93, D94
CDC34-3RZ3	No proper cavity:	V92, I94, L97
	6.8	



Supplementary Figure 1: The composition of the UFC1-UBS asymmetric unit. The asymmetric unit (black rectangle) comprises two molecules of UFC1 (orange and light orange) and two molecules of UBS (red and magenta). The UBS molecules shown in the asymmetric unit are fused to UFC1 molecules that arrive from other asymmetric units. The first two residues of UBS (D389 and S390) and the last two residues of UFC1 (N166 and Q167) were not modeled due to the absence of electron density. The first and last seen residues of UBS are shown in bold while the last seen residues of UFC1 is shown in regular font. The first and last seen residues of UBS and UFC1, respectively, are connected by dotted line and the distance shown could be spanned by the missing residues.



Supplementary Figure 2: Stereographic representations of the UBS helical region with a composite OMIT electron density map contoured at 1 σ . The map coefficients were calculated using Phenix⁴ and the figure was prepared using CCP4mg software⁵. A. In the UFC1-UBS structure, the UBS region and its partner UFC1 region are shown as stick models with carbon atoms in orange and cyan, respectively. B. In the UBA5(347-404)-UFC1 structure the UBS region and the binding region of the crystallographically related UFC1 are shown as stick models with carbon atoms in orange and cyan, respectively.

А

UBA5 L397R

UBA5 M401R



Supplementary Figure 3: Mutations in UBS prevent binding to UFC1. ITC experiment showing no binding affinity between UFC1 and indicated UBA5 mutations. Top graph represents raw data of heat flow versus time. The area under the peaks of the upper panel was integrated and plotted as kcal per mole of UFC1 as a function of binding stoichiometry in the bottom panel. Graph represents the enthalpy change versus the molar ratio of UFC1 to UBA5 mutants. Both graphs show no binding of UFC1 to the above mutants.



Supplementary Figure 4: Mutations in the indicated charged residues of UFC1 or UBS hardly affect UFM1 transfer. SDS-PAGE analysis showing the charging of UFC1 WT or mutant with WT or the indicated UBA5 mutants. The gels are representative of two independent experiments. Source data are provided as a source data file.



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В
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UFC1





Supplementary Figure 5: The surface for UBS binding is unique to UFC1. A. A comparison between electrostatic potential surfaces of indicated E2 enzymes. Yellow dotted line corresponds to the border of UFC1 surface that binds the UBS and the corresponding surface in other E2 proteins. B. Clashes of UBS (blue) with UBC9 (red) surface (indicated with black box) but not with UFC1(orange) surface.



Supplementary Figure 6: Size exclusion chromatography with multi-angle light scattering analysis of UBA5 (347-404) fragment. Protein was loaded on an analytical SEC column (Superdex 75 10/300 GL) equilibrated with a buffer containing 20 mM Tris (pH 7.5), 50 mM NaCl. Molecular mass within the chromatographic peak was calculated using ASTRA software, version 7 (Wyatt Technologies). The measured molecular weight of 6.6 kDa fits the expected molecular weight of this fragment (6.4 kDa). Source data are provided as a source data file.



Supplementary Figure 7: Superpositions of UFC1 with indicated E2 enzymes. The active site Cys is in yellow sphere. UFC1 is colored as in Fig.1C.

	α2	α3	α4	
3TGD_UBCH5B	22222222222		معمعه بععمه	1000000.00
3TGD UBCH5B	98 TISKVLLSI.CSLLCD	N PDDPLVPEIARIYKT	DREKYNRI	AREWTQ. KYAM
1J7D UBC13	100 QIRTVLLSI.QALLSAN	NPDDPLANDVAEQWKTN	VEAQAI	ARAWTR.LYAMNNI
1U9A_UBC9	108 TIKQILLGI.QELLNE	NIQDPAQAEAYTIYCQN	IRVEYEKR	VRAQAK.KFAPS
1X23_UBCH5C	98 TISKVLLSI.CSLLCD	NPDDPLVPEIARIYKT	ORDKYNRI	SREWTQ.KYAM
1Y6L_UBE2E2	155 TISKVLLSI.CSLLTDO	NPADPLVGSIATQYMTN	IRAEHD	[ARQWTK.RYAT
4Q5E_UBCH7	100 KTDQVIQSL.IALVNDI	QPEHPLRADLAEEYSKE	ORKKFS	AEEFTK.KYGEKRPVD
4YII_UBCH	127 DVRTILLSI.QSLLGE	NIDSPLNTHAAELWKNF	PTAFKKYLQETYSKQ	VTSQEP
1Y8X_UBC12	124 TINSIIYGL.QYLFLE	NPEDPLNKEAAEVLQNN	IRRLFEQN	VQRSMRGGYIGSTYF.ERCLK
1WZW_UBCH8	100 KTCQVLEAL.NVLVNR	NIREPLRMDLADLLTQN	VPELFR	AEEFTL.RFGVDRPS
6CYO_RAD6A	101 DVSSILTSI.QSLLDE	NPNSPANSQAAQLYQEN	IKREYEKR	VSAIVE.QSWRDC
1FZY_UBC1	101 TLKSALISL.QALLQSI	EPNDPQDAEVAQHYLRE	DRESFNKT	AALWTR.LYAS
1YLA_UBE2K	105 TLRTVLLSL.QALLAAA	AEPDDPQDAVVANQYKQN	NPEMFKQT	ARLWAH.VYAGAPVSSPEYTKKIENLCAMGFDRNAVIVALSSKSWDVETATELLLSN
2YBF_RAD6B	101 DVSSILTSI.QSLLDE	NPNSPANSQAAQLYQEN	NKRE	VSAIVE.QSWNDS
2C4P_UBCH5A	98 TVSKVLLSI.CSLLCDI	NPDDPLVPDIAQIYKSC	OKEKYNRH	[AREWTQ.KYAM
5LBN_UBCH6	144 TISKVLLSI.CSLLTDO	CNPADPLVGSIATQYMTN	NRAEHDRM	LARQWTK.RYAT
3FN1 UBE2F	135 TLKDVVWGL.NSLFTDI	L.NFDDPLNIEAAEHHLRC	OKEDFRNK	VDDYIKRYAR
2CYX_UBE2G2	115 SVEKILLSV.VSMLAEI	NDESGANVDASKMWRD	DREQFYKI	AKQIVQ.KSLGL
2Z5D_UBE2H	100 DLTNIFESFLPQLLAY	NPIDPLNGDAAAMYLHF	RPEEYKQK	IKEYIQ.KYATEEALKEQEEG
2F4W_UBE2J2	110 SVSTILTGL.LSFMVEH	GPTLGSI.ETSDFTKRQLAVQ	OSLAFNLKDKVFCELFPEV	VEEIKQ.KQKAQDELSSRPQTLP
5BNB_UBE2S	108 GIRHVLLTI.KMLLIHI	NPESALNEEAGRLLLEN	NYEEYAAR	ARLLTE.IHG
1YH2_UBE2T	103 NIATVLTSI.QLLMSEI	NPDDPLMADISSEFKYN	KPAFL	ARQWTE.KHARQKQKADEEE.MLDNLP
1YRV_UBE2U	104 TLSSILLAL.QVMLSNI	VLENPVNLEAARILVKC	DESLYRTI	LRLFNR.P
2Z6P UFC1	133 GL.AHLMALGLGPV	LAVE PDLIQKGVIQHKEKCNQ		
-	a2.			
	000000000000000000000000000000000000000	200000000000000		

Supplementary Figure 8: sequence alignment of E2 enzymes. The secondary structural elements covering the region of helix 2 to the C-terminus are shown for the indicated E2s (top). The corresponding secondary element for that region in UFC1 is shown underneath the sequence alignment.



Supplementary Figure 9: Overlay of ${}^{1}H{-}{}^{15}N$ HSQC spectra of 0.2 mM UFC1 alone(black), and in the presence of equimolar concentration of UBA5³⁴⁷⁻⁴⁰⁴ (red, upper panel) or five-fold excess of UBA5³⁴⁷⁻³⁹² (pink, bottom panel).

Supplementary References

- 1 Tickle, I. Staraniso: Use of a Webgl-Based 3-D Interactive Graphical Display to Represent and Visualise Data Quality Metrics for Anisotropic Macromolecular Diffraction Data. *Acta Crystallogr A* **75**, E162-E162, doi:10.1107/S205327331909394x (2019).
- 2 Karplus, P. A. & Diederichs, K. Linking Crystallographic Model and Data Quality. *Science* **336**, 1030-1033, doi:10.1126/science.1218231 (2012).
- 3 Vaguine, A. A., Richelle, J. & Wodak, S. J. SFCHECK: a unified set of procedures for evaluating the quality of macromolecular structure-factor data and their agreement with the atomic model. *Acta Crystallogr D* **55**, 191-205, doi:Doi 10.1107/S0907444998006684 (1999).
- 4 Afonine, P. V. *et al.* Towards automated crystallographic structure refinement with phenix.refine. *Acta crystallographica. Section D, Biological crystallography* **68**, 352-367, doi:10.1107/S0907444912001308 (2012).
- 5 Potterton, L. *et al.* Developments in the CCP4 molecular-graphics project. *Acta crystallographica.* Section D, Biological crystallography **60**, 2288-2294, doi:10.1107/S0907444904023716 (2004).