## SUPPLEMENTARY INFORMATION

## Mapping protein interactions in the active TOM-TIM23 supercomplex

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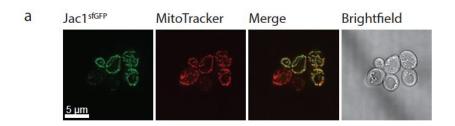
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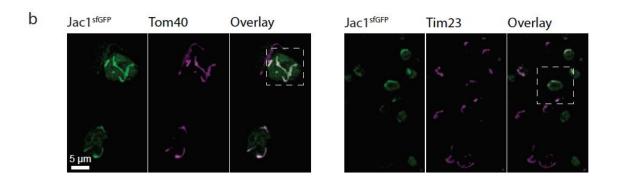
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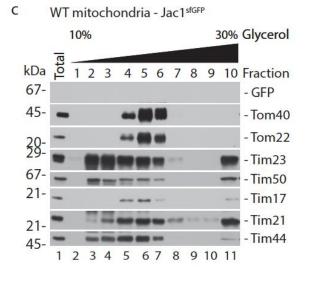
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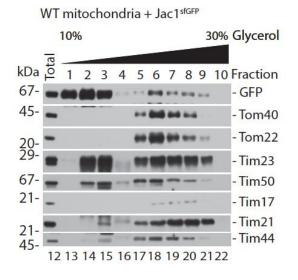
Supplementary Figures 1 to 4

Supplementary Tables 1 to 4

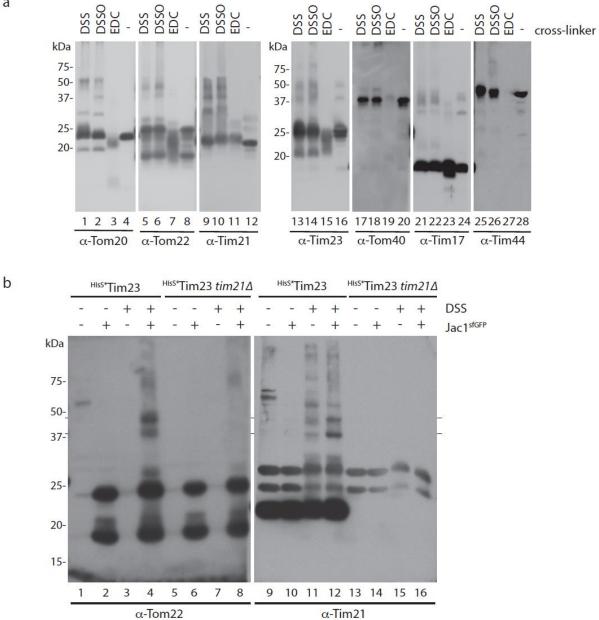






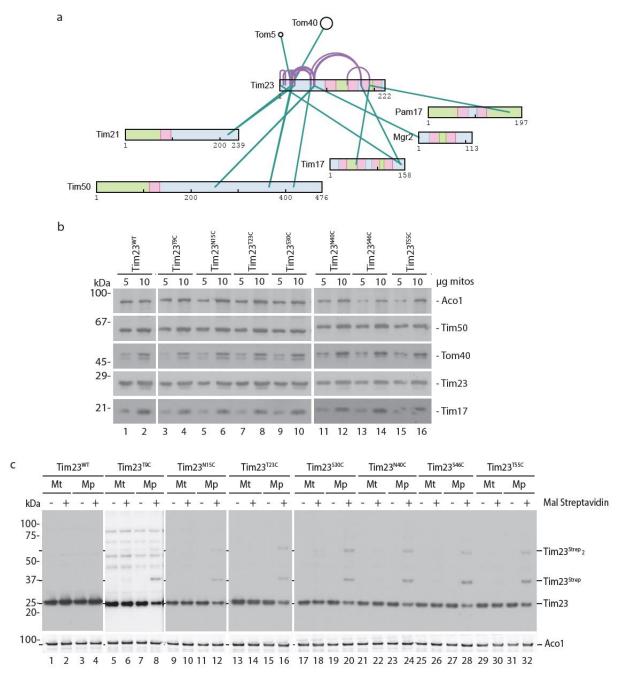


Supplementary Fig. 1. Supercomplex-forming Jac1<sup>sfGFP</sup> localizes to mitochondria. a Yeast cells were transformed with Jac1sfGFP (green)-encoding plasmid. After induction of expression, cells were co-stained with MitoTracker<sup>TM</sup> Orange CMTMRos (red). Merged green and red fluorescence images are shown, indicating co-localization (yellow). (Scale bar =  $5 \mu m$ ). Representative images from three independent experiments are depicted. b Two-colour confocal microscopy of veast cells expressing Jac1<sup>sfGFP</sup>. Cells were labelled with antibodies against GFP (green) and Tom40 (magenta) or Tim23 (magenta). (Scale bar, 5 µm). Representative images from three biological replicates are depicted. c Solubilized wild type mitochondria were fractionated on 10-30% glycerol gradients following import of Jac1<sup>stGFP</sup>. Fractions were collected from top, TCA precipitated, and analyzed by SDS-PAGE followed by western blotting. Total: 2% of mitochondria loaded on the gradient. Representative experiment from a biologic triplicate is shown.

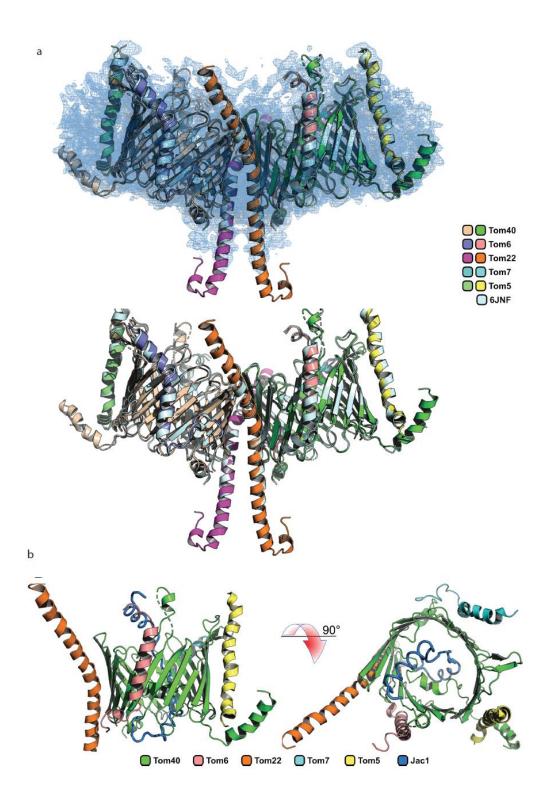


Supplementary Fig. 2. Chemical cross-linking of supercomplex subunits. a Supercomplex isolation was carried out from <sup>HisS\*</sup>Tim23 mitochondria subjected to Jac1<sup>sfGFP</sup> import. Isolation was performed using SUMO\* protease. Isolated complex was treated with DMSO, 2 mM DSS, 2 mM DSSO and 10 mM EDC. Samples were analyzed by SDS-PAGE and western blotting. DSS: disuccinimidyl suberate, DSSO: disuccinimidyl sulfoxide, EDC: 1-Ethyl-3-(3-dimethylaminopropyl)carbodiimide. Three independent experiments were carried out. b TIM23 complex was isolated from HisS\*Tim23 or HisS\*Tim23 *tim21∆* mitochondria in the absence or presence of Jac1<sup>sfGFP</sup>. Eluted samples were treated with DMSO or 2 mM DSS. Samples were analyzed by SDS-PAGE and western blotting. Three replicates were carried out.

a



**Supplementary Fig. 3. Tim23 N-terminus localizes to the IMS. a** Overview of intraand inter- protein cross-links obtained for Tim23. Colour code indicates domain location. blue: IMS, pink: IMM and green: matrix. **b** Purified mitochondria from wild type cells and cells expressing Tim23 cysteine mutants were subjected to SDS-PAGE and immunoblotting. Representative image from two independent experiments. **c** Cysteine modification assay for Tim23 was carried out in mitochondria and mitoplasts from wild type and Tim23 cysteine mutants in the presence or absence of maleimide streptavidin. Samples were subjected to SDS-PAGE and western blotting. Representative image from three replicates.



**Supplementary Fig. 4. Extended structure of the TOM complex.** a *de novo* extension of intermembrane space (IMS) domain of Tom22 (magenta and orange) based on available TOM complex structures (PDB id: 6JNF, EMDB ID: 9851). Rosetta tools were utilized for extending the structure. Superposition of the published TOM complex atomic model (PDB id: 6JNF) supports the extended (modelled) TOM complex. **b** Single TOM complex subunit with front and top view. Colour code indicates individual subunits.

**Supplementary Table 1:** List of identical inter-protein cross-links obtained by three methods: 1) TIM23 complex isolation followed by cross-linking, 2) whole mitochondria *in organello* cross-linking, and 3) *in organello* cross-linking followed by TIM23 complex isolation, in resting (- Jac1<sup>sfGFP</sup>) or active (+ Jac1<sup>sfGFP</sup>) state.

Cross- linker	Inter-protein cross-links	Approach 1 Isolation followed by		Approach 2 in organello cross-		Approach 3 <i>in organello</i> cross-linking	
IIIKEI		cross-linking		linking		followed by isolation	
		- Jac1 <sup>sfGFP</sup>	+ Jac1 <sup>sfGFP</sup>	- Jac1 <sup>sfGFP</sup>	+ Jac1 <sup>sfGFP</sup>	- Jac1 <sup>sfGFP</sup>	+ Jac1 <sup>sfGFP</sup>
EDC	Tom70 <sup>103</sup> - Tom20 <sup>183</sup>	-	+	-	+	NA	NA
EDC	Tom5 <sup>1</sup> - Tom20 <sup>90</sup>	-	+	-	+	NA	NA
EDC	$Tom 5^1$ - $Tom 20^{182}$	-	+	+	-	NA	NA
EDC	$Tom 5^1$ - $Tom 20^{183}$	-	+	+	-	NA	NA
DSS	Tim23 <sup>190</sup> - Tim17 <sup>55</sup>	+	-	-	-	+	-
DSS	Hsp70 <sup>445</sup> - Mge1 <sup>104</sup>	+	+	+	+	+	+
DSS	Hsp70 <sup>83</sup> - Mge1 <sup>119</sup>	-	+	-	+	+	+
DSS	Hsp70 <sup>444</sup> - Tim44 <sup>228</sup>	-	+	-	-	-	+
DSS	Hsp70 <sup>514</sup> - Tim44 <sup>215</sup>	+	-	-	-	+	-
DSS	Hsp70 <sup>574</sup> - Tim44 <sup>198</sup>	+	+	-	-	+	+
DSS	Pam16 <sup>34</sup> - Pam18 <sup>101</sup>	+	+	+	-	+	+
DSS	Pam16 <sup>89</sup> - Pam18 <sup>135</sup>	-	+	-	-	-	+

-: cross-link not detected

+: cross-link detected

NA: not analyzed

Supplementary Table 2: List of yeast strains generated in this study.

Strain	Genotype			
<sup>HisS*</sup> Tim23 <i>tim21</i> ∆	HisS*Tim23 <i>tim21::TRP</i>			
BY4741-pRG13	BY4741 + [pRG13-Jac1 <sup>sfGFP</sup> (LEU2)]			
Tim21 <sup>FLAG</sup>	YPH499 <i>tim21::</i> TIM21 <sup>FLAG</sup> (HIS3)			
Tim21 <sup>FLAG</sup> +Tim21 <sup>WT</sup>	YPH499 <i>tim21::</i> TIM21 <sup>FLAG</sup> (HIS3) + [pFL39-TIM21 (TRP)]			
YPH499-pRG26	YPH499 + [pRG26 (TRP)]			
YPH499-pRG27 (Tim23 <sup>ALFA</sup> )	YPH499 + [pRG27-pTIM23 <sup>ALFA</sup> (TRP)]			
YPH499-pRG28 (Tim17 <sup>ALFA</sup> )	YPH499 + [pRG28-pTIM17 <sup>ALFA</sup> (TRP)]			
YPH499-pRG30 (Tim50 <sup>ALFA</sup> )	YPH499 + [pRG30-pTIM50 <sup>ALFA</sup> (TRP)]			
YPH499-pRG33 (Tim44 <sup>ALFA</sup> )	YPH499 + [pRG33-pTIM44 <sup>ALFA</sup> (TRP)]			
YPH499-pRG35 (Pam18 <sup>ALFA</sup> )	YPH499 + [pRG35-pPAM18 <sup>ALFA</sup> (TRP)]			
YPH499-pRG37 (Pam16 <sup>ALFA</sup> )	YPH499 + [pRG37-pPAM16 <sup>ALFA</sup> (TRP)]			
YPH499-R673 (Tim23 <sup>WT</sup> )	MB29 <i>tim23</i> ∆ + [pRS413-pORFt TIM23 (HIS3)]			
YPH499-pRG39 (Tim23 <sup>T9C</sup> )	MB29 <i>tim23</i> ∆ + [pRS413-pORFt TIM23 T9C (HIS3)]			
YPH499-pRG40 (Tim23 <sup>N15C</sup> )	MB29 <i>tim23</i> ∆ + [pRS413-pORFt TIM23 N15C (HIS3)]			
YPH499-pRG41 (Tim23 <sup>T23C</sup> )	MB29 <i>tim23</i> ∆ + [pRS413-pORFt TIM23 T23C (HIS3)]			
YPH499-pRG42 (Tim23 <sup>S30C</sup> )	MB29 <i>tim23</i> ∆ + [pRS413-pORFt TIM23 S30C (HIS3)]			
YPH499-pRG44 (Tim23 <sup>N40C</sup> )	MB29 <i>tim23</i> ∆ + [pRS413-pORFt TIM23 N40C (HIS3)]			
YPH499-pŔG45 (Tim23 <sup>S46C</sup> )	MB29 <i>tim23</i> ∆ + [pRS413-pORFt TIM23 S46C (HIS3)]			
YPH499-pRG46 (Tim23 <sup>T55C</sup> )	MB29 <i>tim23</i> ∆ + [pRS413-pORFt TIM23 T55C (HIS3)]			

Supplementary Table 3: List of plasmids used in this study.

Plasmids	Description	
GST-nGFP-Strep (R286)	GST-Thrombin-GFP Nanobody-Strep in pGEX-4T3 (Amp)	
pRG9	14HIS-SUMO-Jac1 <sup>sfGFP Q7R</sup> in pSUMO (Kan)	
Tim44 <sup>C-term</sup> (R662)	Tim44 residues 244-431 in pPROEX HTc (Amp)	
Tim44 (1014)	Tim44 full length (without presequence) in pQE60 (Amp)	
pRG13	Jac1 <sup>sfGFP Q7R</sup> in p425Gal1 (Amp, LEU2)	
pRG26	ALFA-ALDH terminator in pRS414 (Amp, TRP1)	
pRG27	Tim23 in pRG26 (Amp, TRP1)	
pRG28	Tim17 in pRG26 (Amp, TRP1)	
pRG30	Tim50 in pRG26 (Amp, TRP1)	
pRG33	Tim44 in pRG26 (Amp, TRP1)	
pRG35	Pam18 in pRG26 (Amp, TRP1)	
pRG37	Pam16 in pRG26 (Amp, TRP1)	
pFL39 (R84)	empty vector for yeast expression (Amp, TRP1)	
pGB 9607-7 (R85)	Tim21 in pFL39 for expression in yeast (Amp, TRP1)	
R673	pORFt Tim23 in pRS413 (Amp, HIS3)	
pRG39	pORFt Tim23 T9C in pRS413 (Amp, HIS3)	
pRG40	pORFt Tim23 N15C in pRS413 (Amp, HIS3)	
pRG41	pORFt Tim23 T23C in pRS413 (Amp, HIS3)	
pRG42	pORFt Tim23 S30C in pRS413 (Amp, HIS3)	
pRG44	pORFt Tim23 N40C in pRS413 (Amp, HIS3)	
pRG45	pORFt Tim23 S46C in pRS413 (Amp, HIS3)	
pRG46	pORFt Tim23 T55C in pRS413 (Amp, HIS3)	

Supplementary Table 4: Antibodies used in this study.

Antibody	Source	Identifier	Dilution	
GFP (western	Roche	11814460001	1:1000	
blot)				
GFP (microscopy)	Thermo Fisher Scientific	A-11120		
Anti-ALFA HRP-	NanoTag Biotechnologies	N1502-HRP	1:500	
conjugated				
FLAG	Sigma	F3165	1:500	
Peroxidase	Jackson ImmunoResearch Lab	016-030-084	1:500	
Streptavidin (SA-				
HRP)				
Goat anti Rabbit	Jackson ImmunoResearch Lab	111-035-144	1:10,000	
lgG (H+L) HRPO				
Tim23	Rabbit polyclonal, self-made	3846	1:500	
Tim23 <sup>IMS</sup>	Rabbit polyclonal, self-made	133	1:750	
Tim17	Rabbit polyclonal, self-made	4968	1:200	
Tim50	Rabbit polyclonal, self-made	3314	1:400	
Tim21	Rabbit polyclonal, self-made	3111	1:400	
Tim44	Rabbit polyclonal, self-made	3869	1:250	
Hsp70	Rabbit polyclonal, self-made	4945	1:50,000	
Pam16	Rabbit polyclonal, self-made	3872	1:200	
Pam18	Rabbit polyclonal, self-made	3955	1:200	
Pam17	Rabbit polyclonal, self-made	5017	1:400	
Tom70	Rabbit polyclonal, self-made	3530	1:400	
Tom40	Rabbit polyclonal, self-made	4901	1:200	
Tom22	Rabbit polyclonal, self-made	3533	1:400	
Tom20	Rabbit polyclonal, self-made	3534	1:400	
Tom5	Rabbit polyclonal, self-made	162	1:500	
Aco1	Rabbit polyclonal, self-made	5004	1:500	