

185 Appendix

186 Supplementary methods

187 Dataset

188 We analyse 123,867 variant-specific RT-PCR tests performed in France on the same number of individuals
189 between Jan 26 and Mar 19, 2021. The main assay used was IDTM SARS-CoV-2/UK/SA Variant Triplex
190 (ID SOLUTION) but for 4159 tests (3,4%) performed before Feb 3, 2021 we used the VirSNiP SARS-CoV-
191 2 Spike del+501 (TIB MOLBIOL) assay. The sampling varied between French regions and we excluded
192 from the analysis regions with less than 100 tests. 936 tests were also removed because the sampling region
193 was missing.

194 These tests have probes with 3 targets: a control one in the virus N gene, the Δ 69-70 deletion, and the
195 N501Y mutation. For Alpha variants, both the deletion and the mutation are present. For Beta or Gamma
196 variants, only the N501Y is detected. For VirSNiP assay, it is based on 501 and 69/70 fragments amplified
197 and analyzed with a melting curve using mutation-specific probes, as described earlier ([Haim-Boukobza
198 et al., 2021](#)). As indicated in [Haim-Boukobza et al. \(2021\)](#), the test specificity was confirmed internally
199 using next-generation sequencing.

200 The main cofactors in the analysis were the assay used, the patient age, the sampling date, the sampling
201 region, and the sampling facility (hospitals or city screening).

202 For 119,708 ID SOLUTION tests, we also analyse the cycle threshold value (Ct) of the virus control
203 gene of the assay. Ct values greater than 30 were ignored because they may provide unreliable results
204 regarding the variant-specific probes LoD (Limit of Detection). Indeed, the latter are located in the S
205 gene, which tends to exhibit higher Ct values than the N gene ([Alizon et al., 2021](#)).

206 Linear model for the Ct analysis

207 We performed a type I error for the analysis-of-variance. Our response variable was the Ct value. The
208 main covariate of interest was the strain and it could take 4 values (Alpha, Beta or Gamma, wild type,
209 or other). The other covariates were the age, the sampling facility (hospital or city), the sampling date,
210 and the geographical region. We also considered an interaction between sampling region and date. We
211 used a type-I analysis of variance (ANOVA) and added the strain covariate last. The motivation for this
212 is that with the sequential assumption of the summing of the squares (type I method), the order in which

213 the covariates are tested matters, and, in the case of an uneven sampling, the last one in the list is less
214 likely to be significant. Therefore, our assumption decreases the risk of erroneously attributing observed
215 variance to a variant effect.

216 We used a F-test to determine whether the addition of the strain effect statistically improved the
217 explanation of the data.

218 Generalised linear model to correct for variant sampling bias

219 As indicated in (Haim-Boukobza et al., 2021), for a given variant category (Alpha or Beta/Gamma) we first
220 perform a generalised linear model with a binomial error distribution where the variable of interest is the
221 binary variant variable (with values ‘variant’ or ‘wild type’) and the explanatory variables are the sampling
222 date, the sampling region, and the individual age. We also include an interaction between sampling region
223 and date. We then use the residuals of this model to infer the transmission advantage of the variant.

224 Logistic growth fitting

225 We used the fitted values of a GLM model applied to the data after removing samples from hospitals
226 (the sampling location effect was also obviously removed from the model) to perform the inference of a
227 two-parameter logistic growth kinetic curve: $f(t) = (1 + e^{-\rho(t-\tau)})^{-1}$, where $f(t)$ is the frequency of the
228 variants in the new infections at time t , ρ is the relative growth rate of the variants and τ is the time
229 at which f reaches 1/2. This method is indeed more appropriate to deal with temporal auto-correlation
230 biases in proportion time series (Davies et al., 2021; Volz et al., 2021).

231 The parameter estimation was performed using the `drc` package in R both at the national and the
232 regional level (for regions with at least 1,000 samples). The confidence intervals of the fitted curves rely
233 on those of the estimated date of reaching half proportion of new infections (τ).

234 The unitless estimated transmission advantage is expressed in terms of multiplicative gain in repro-
235 duction number with respect to that of the wild type, such that $\mathcal{R}_{\text{variant}} = (1 + \text{ETA}) \times \mathcal{R}_{\text{wild type}}$. Its
236 calculation was made by solving the Euler-Lotka equation ($\mathcal{R}_{\text{variant}} \int_0^\infty e^{-\rho t} w(t) dt = 1$) assuming a serial
237 interval w following a Weibull distribution with a mean and SD of 4.8 and 2.3 days (Nishiura et al., 2020)
238 and a constant $\mathcal{R}_{\text{wild type}}$ equal to 1. The confidence interval rely on those of the estimated relative growth
239 rate.

240 The estimate of the frequency of variant on Mar 12, 2021, was done by first estimating the proportion p_x
241 of a given variant x compared to the wild type (while ignoring the other variant y) and second performing

242 the same analysis to look at the proportion p_x^T of wild type and x compared to the whole population (x
243 plus y plus wild type). The frequency of variant X was then obtained as $p_x \times p_x^T$.

244 Supplementary figures

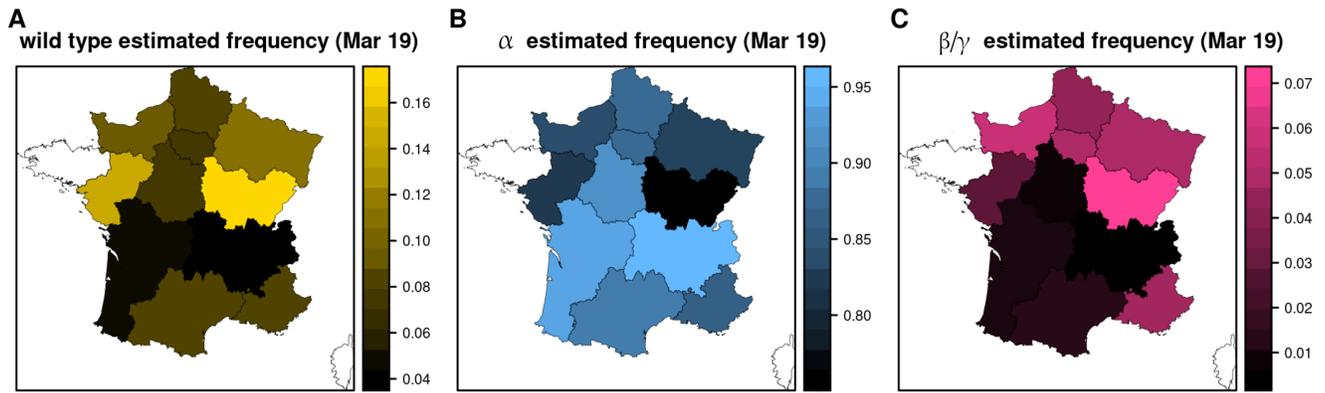


Figure S1: Estimated proportion of new infections caused by A) wild type, B) Alpha variant, and C) Beta or Gamma variants on Mar 12, 2021, in French regions. Regions with insufficient sampling are in white.

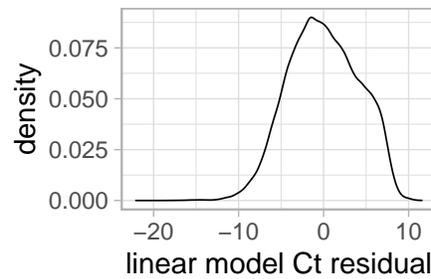


Figure S2: Distribution of the residual values of the multiple regression linear model.

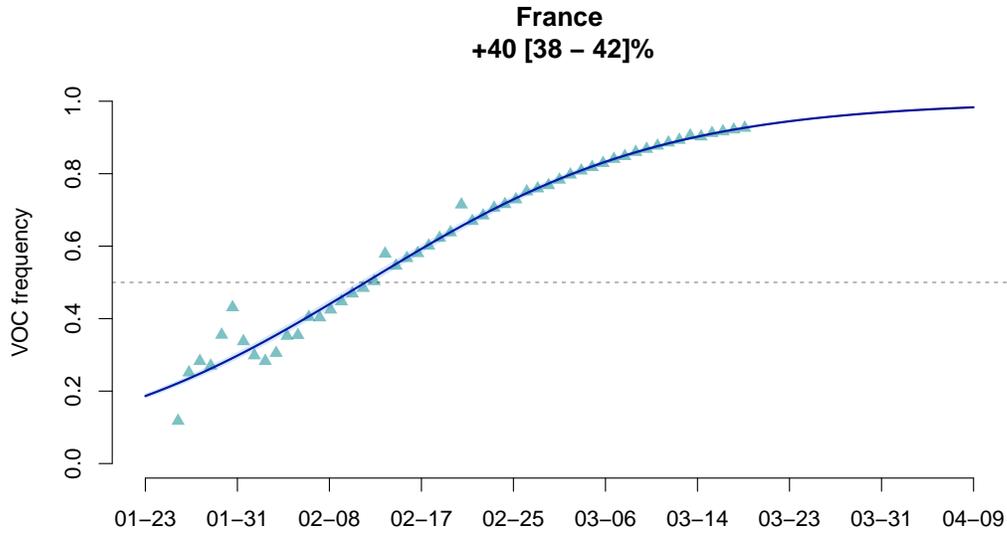


Figure S3: **Estimating the transmission advantage of the Alpha variant over the wild type strain.** The dots indicate the GLM-fitted values values and the line is the output of the logistic growth model estimation. The top figures indicate the estimated transmission advantage of the Alpha variant (with respect to the wild type reproduction number) and its 95%-confidence interval. The x-axis shows the date (month-day format).

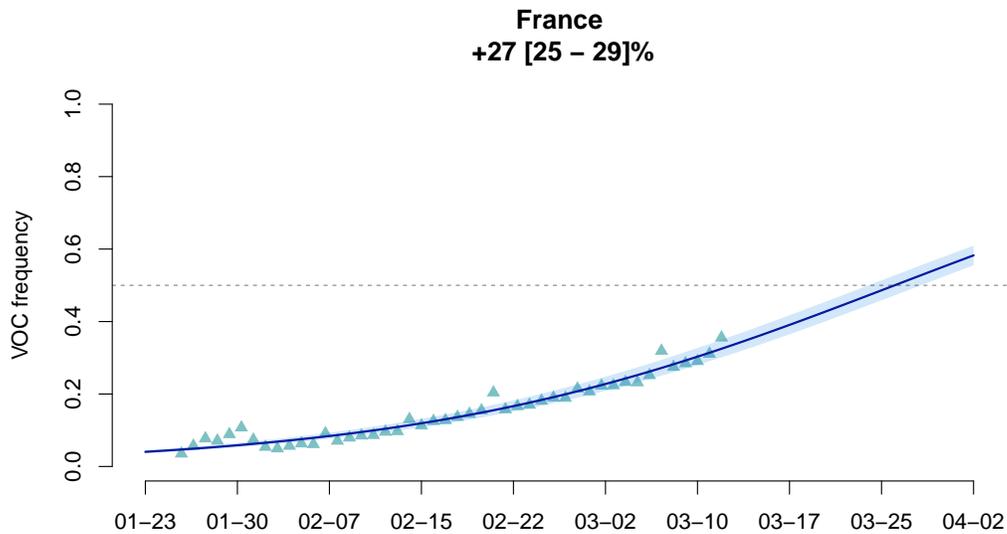


Figure S4: **Estimating the transmission advantage of the Beta or Gamma variants over the wild type strain.** See Figure S3 for details.

245 **Additional bibliography**

- 246 Nishiura H, Linton NM, Akhmetzhanov AR. Serial interval of novel coronavirus (COVID-19) infections.
247 International Journal of Infectious Diseases. 2020 Apr;93:284-286.
248 Available from: [https://www.ijidonline.com/article/S1201-9712\(20\)30119-3/abstract](https://www.ijidonline.com/article/S1201-9712(20)30119-3/abstract).