

THE LANCET

Supplementary appendix

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Supplement to: Predecki M, Thomson T, Clarke CL, et al. Immunological responses to SARS-CoV-2 vaccines in kidney transplant recipients. *Lancet* 2021; published online Oct 4. [http://dx.doi.org/10.1016/S0140-6736\(21\)02096-1](http://dx.doi.org/10.1016/S0140-6736(21)02096-1).

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Table S1. Clinical characteristics of the main cohort and subgroup of patients who had cellular responses assessed.

Characteristic		Main Cohort N=920 (%)	Subgroup N=106 (%)	p value
Gender	Male	604 (65.7)	71 (67.0)	0.78
	Female	316 (34.3)	35 (33.0)	
Age	Years (Median)	59 (48-67)	53 (42-62)	<0.0001
Ethnicity	Caucasian*	404 (43.9)	43 (40.6)	0.51
	Black	84 (9.1)	14 (13.2)	
	Indoasian	301 (32.7)	31 (29.2)	
	Other	131 (14.2)	18 (17.0)	
Cause of ESKD	Polycystic kidney disease	107 (11.6)	8 (7.5)	0.59
	Glomerulonephritis*	272 (29.6)	34 (32.1)	
	Diabetic nephropathy	171 (18.6)	17 (16.0)	
	Urological	72 (7.8)	8 (7.5)	
	Unknown	208 (22.6)	26 (24.5)	
	Other	90 (9.8)	13 (12.3)	
Vaccinated ≤1 year post transplant	Yes	92 (10.0)	45 (42.5)	<0.0001
	No	828 (90.0)	61 (57.5)	
Time vaccinated post transplant	Years (Median)	6.6 (6.1-7.3)	2.0 (0.4-7.9)	<0.0001
Immunosuppression at diagnosis	CNI monotherapy*	458 (49.8)	44 (41.5)	0.11
	CNI/anti-proliferative	240 (26.1)	30 (28.3)	
	CNI/steroids	56 (6.1)	2 (1.9)	
	CNI/anti-proliferatives/steroids	149 (16.2)	26 (24.5)	

	Anti-proliferatives/steroids	4 (0.4)	-	
	Other	13 (1.4)	4 (3.8)	
Induction immunotherapy	Alemtuzumab*	636 (69.3)	64 (60.4)	0.067
	IL2 receptor blocker	103 (11.2)	30 (28.3)	
	None/Unknown	181 (19.7)	12 (11.3)	
Transplant number	1 st	806 (87.6)	93 (87.7)	0.97
	≥2 nd	114 (12.4)	13 (12.3)	
Diabetes	No	592 (64.3)	75 (70.8)	0.19
	Yes	328 (35.7)	31 (29.2)	
Vaccine type	BNT162b2	490 (53.3)	51 (48.1)	0.32
	ChAdOx1	430 (46.7)	55 (51.9)	
Time between vaccinations	Days (median)	74 (66-77)	63 (63-77)	<0.0001
Time of serological test post-boost	Days (median)	31 (27-35)	31 (29-34)	0.38

ESKD – end stage kidney disease, CNI- calcineurin inhibitor *Comparator

Table S2. Demographic characteristics of HCW comparator cohort

Characteristic		HCW (prior infection) n=25 (%)	HCW (infection-naïve) n=40 (%)
Gender	Male	11 (44.0)	15 (37.5)
	Female	14 (56.0)	25 (62.5)
Age	Years (Median)	32.9 (26.0-47.1)	42.8 (33.4-45.7)
Vaccine type	BNT162b2	18 (72.0)	32 (80.0)
	ChAdOx1	7 (28.0)	8 (20.0)
Time between vaccinations	Days (median)	68 (61-75)	66 (61-69)
Time of serological test post-boost	Days (median)	28 (21-29)	24 (21-29)

Table S3. Clinical characteristics associated with seroconversion following SARS-CoV-2 vaccination in 768 infection-naïve kidney transplant recipients

Characteristic		Failure to seroconvert N=343 (%)	Seroconversion N=425 (%)	p value
Gender	Male	228 (66.5)	280 (65.9)	0.86
	Female	115 (33.5)	145 (34.1)	
Age	Years (Median)	60 (49-67)	59 (48-67)	0.30
Ethnicity	Caucasian	164 (47.8)	200 (47.1)	0.14
	Black	31 (9.0)	26 (6.1)	
	Indoasian	94 (27.4)	143 (33.6)	
	Other	54 (15.7)	56 (13.2)	
Cause of ESKD	Polycystic kidney disease	40 (11.7)	51 (12.0)	0.74
	Glomerulonephritis	106 (30.9)	123 (28.9)	
	Diabetic nephropathy	69 (20.1)	75 (17.6)	
	Urological	27 (7.9)	39 (9.2)	
	Unknown	66 (19.2)	98 (23.1)	
	Other	35 (10.2)	39 (9.2)	
Vaccinated ≤1 year post transplant	Yes	42 (12.2)	17 (4.0)	<0.0001
	No	301 (87.8)	408 (96.0)	
Time vaccinated post transplant	Years (Median)	6.8 (2.4-13.6)	6.7 (3.2-11.9)	0.78
Immunosuppression at diagnosis	CNI monotherapy*	97 (28.3)	284 (66.8)	<0.0001
	CNI/anti-proliferative	123 (35.9)	74 (17.4)	
	CNI/steroids	21 (6.1)	26 (6.1)	
	CNI/anti-proliferatives/steroids	95 (27.7)	36 (8.5)	
	Anti-proliferatives/steroids	3 (0.9)	1 (0.2)	
	Other	4 (1.2)	4 (0.9)	

Induction immunotherapy	Alemtuzumab* IL2 receptor blocker None/Unknown/Other	194 (56.6) 60 (17.5) 89 (25.9)	333 (78.4) 23 (5.4) 69 (16.2)	<0.0001
Transplant number	1 st ≥2 nd	292 (85.1) 51 (14.9)	380 (89.4) 45 (10.6)	0.07
Diabetes	No Yes	210 (61.2) 133 (38.8)	294 (69.2) 131 (30.8)	0.021
Vaccine type	BNT162b2 ChAdOx1	141 (41.1) 202 (58.9)	269 (63.3) 156 (36.7)	<0.0001
Time between vaccinations	Days (median)	74 (66-78)	75 (68-77)	0.76
Time of serological test post-boost	Days (median)	31 (27-35)	31 (27-35)	0.49

ESKD – end stage kidney disease, CNI-Calcineurin inhibitors, *indicates comparator group

Table S4. Comparison of clinical characteristics of infection-naïve patients receiving ChAdOx1 versus BNT162b2 vaccines

Characteristic		ChAdOx1 N=358 (%)	BNT162b2 N=410 (%)	p value
Gender	Female	117 (32.7)	143 (34.9)	0.52
	Male	241 (67.3)	267 (65.1)	
Age	Years (Median)	59 (49-67)	60 (48-67)	0.45
Ethnicity	Caucasian	175 (48.9)	189 (46.1)	0.046
	Black	35 (9.8)	22 (5.4)	
	Indoasian	98 (27.4)	139 (33.9)	
	Other	50 (14.0)	60 (14.6)	
Cause of ESKD	Polycystic kidney disease	47 (13.1)	44 (10.7)	0.62
	Glomerulonephritis	113 (31.6)	116 (28.3)	
	Diabetic nephropathy	66 (18.4)	78 (19.0)	
	Urological	26 (7.3)	40 (9.8)	
	Unknown	74 (20.7)	90 (22.0)	
	Other	32 (8.9)	42 (10.2)	
Vaccinated ≤1 year post transplant	Yes	39 (10.9)	20 (4.9)	0.0025
	No	319 (89.1)	390 (95.1)	
Immunosuppression at diagnosis	CNI monotherapy*	166 (46.4)	215 (56.4)	0.33
	CNI/anti-proliferative	94 (26.3)	103 (25.1)	
	CNI/steroids	25 (7.0)	22 (5.4)	
	CNI/anti-proliferatives/steroids	69 (19.3)	62 (15.1)	
	Anti-proliferatives/steroids	2 (0.6)	2 (0.5)	
	Other	2 (0.6)	6 (1.5)	
Induction immunotherapy	Alemtuzumab*	235(65.6)	292 (71.2)	0.078
	IL2 receptor blocker	48 (13.4)	35 (8.5)	

	None/Unknown/Other	75 (20.9)	83 (20.2)	
Transplant number	1 st ≥2 nd	311 (86.9) 47 (13.1)	361 (88.0) 49 (12.0)	0.62
Diabetes	No Yes	231 (64.5) 127 (35.5)	273 (66.6) 137 (33.4)	0.55
Seroconversion	Yes No	156 (43.6) 202 (56.4)	269 (65.6) 141 (34.4)	<0.0001
Time between vaccinations	Days (median)	74 (66-78)	74.5 (68-77)	0.95
Time of serological test post-boost	Days (median)	31 (26-34)	31 (28-35)	0.065

ESKD – end stage kidney disease, CNI=calcineurin inhibitors, *indicates comparator group

Table S5. Clinical characteristics associated with seroconversion in 358 infection-naïve patients receiving ChAdOx1

Variable	Reference Group	Denominator	Response Rate (%)	Univariable		Multivariable	
				OR (95% CI)	p value	OR (95% CI)	p value
Age	≥60	178	69 (38.8)	0.64 (0.44-1.03)	0.068	0.53 (0.33-0.84)	0.008
Gender	Male	241	104 (43.2)	0.95 (0.61-1.48)	0.82		
Ethnicity	Caucasian	175	76 (43.4)	0.99 (0.65-1.50)	0.96		
Cause ESRD	Glomerulonephritis	113	51 (45.1)	1.10 (0.70-1.72)	0.69		
Time of vaccine post-transplant	<1 year	39	7 (17.9)	0.25 (0.11-0.58)	0.0013	0.23 (0.09-0.56)	0.0012
Number of transplants	≥2	47	19 (40.4)	0.86 (0.46-1.61)	0.64		
Transplant induction agent	Alemtuzumab	235	124 (52.8)	3.18 (1.97-5.12)	<0.0001	-	
Maintenance Immunosuppression	CNI monotherapy	166	104 (62.7)	4.52 (2.89-7.06)	<0.0001	4.12 (2.44-6.95)	<0.0001
Diabetes	Yes	127	51 (40.2)	0.8 (0.52-1.25)	0.33		

CNI=calcineurin inhibitors

Table S6. Clinical characteristics associated with seroconversion in 410 infection-naïve patients receiving BNT162b2

Variable	Reference Group	Denominator	Response Rate (%)	Univariable		Multivariable	
				OR (95% CI)	p value	OR (95% CI)	p value
Age	≥60	208	134 (64.4)	0.90 (0.60-1.35)	0.61		
Gender	Male	267	176 (65.9)	1.04 (0.68-1.59)	0.86		
Ethnicity	Caucasian	189	124 (65.6)	0.99 (0.66-1.50)	0.99		
Cause ESRD	Glomerulonephritis	116	72 (62.1)	0.81 (0.52-1.26)	0.34		
Time of vaccine post-transplant	<1 year	20	10 (50.0)	0.51 (0.21-1.25)	0.14	-	
Number of transplants	≥2	49	26 (53.1)	0.55 (0.30-1.0)	0.05	-	
Transplant induction agent	Alemtuzumab	292	209 (71.6)	2.43 (1.57-3.79)	0.0001	-	
Maintenance Immunosuppression	CNI monotherapy	215	180 (83.7)	6.13 (3.87-9.69)	<0.0001	6.34 (3.71-11.06)	<0.0001
Diabetes	Yes	137	80 (58.4)	0.62 (0.41-0.96)	0.03	0.51 (0.32-0.83)	0.006

CNI=calcineurin inhibitors

Table S7. Clinical characteristics associated with seroconversion in 768 infection-naïve patients receiving a SARS-CoV-2 vaccine

Variable	Reference Group	Denominator	Response Rate (%)	Univariable		Multivariable	
				OR (95% CI)	p value	OR (95% CI)	p value
Age	≥60	386	203 (52.6)	1.25 (0.94-1.66)	0.12	0.71 (0.51-0.99)	0.043
Gender	Male	508	280 (55.1)	0.97 (0.72-1.32)	0.97		
Ethnicity	Caucasian	364	200 (54.9)	0.97 (0.73-1.29)	0.84		
Cause ESRD	Glomerulonephritis	229	123 (53.7)	0.91 (0.67-1.24)	0.55		
Time of vaccine post-transplant	<1 year	59	17 (28.8)	0.30 (0.17-0.53)	<0.0001	0.30 (0.16-0.57)	0.0002
Number of transplants	≥2	96	45 (46.9)	0.68 (0.44-1.04)	0.08	-	
Transplant induction agent	Alemtuzumab	527	333 (63.2)	2.78 (2.03-3.82)	<0.0001	1.22 (0.82-1.80)	0.32
Maintenance Immunosuppression	CNI monotherapy	381	284 (74.5)	5.11 (3.75-6.96)	<0.0001	5.20 (3.57-7.57)	<0.0001
Diabetes	Yes	264	131 (49.6)	0.70 (0.52-0.94)	0.02	0.65 (0.46-0.91)	0.014
Vaccine	BNT162b2	410	269 (65.6)	2.47 (1.85-3.31)	<0.0001	2.46 (1.78-3.41)	<0.0001

CNI - calcineurin inhibitors

Table S8. Characteristics of infection-naïve patients undergoing assessment of serological and cellular responses by vaccine type

Characteristic		BNT162b2 N=40 (%)	ChAdOx1 N=39 (%)	p value
Gender	Male	30 (75.0)	25 (64.1)	0.30
	Female	10 (25.0)	14 (35.9)	
Age	Years (Median)	57 (46-64)	50 (39-56)	0.016
Ethnicity	Caucasian	18 (45.0)	19 (48.7)	0.30
	Black	2 (5.0)	6 (15.4)	
	Indoasian	14 (35.0)	8 (20.5)	
	Other	6 (15.0)	6 (15.4)	
Cause of ESKD	Glomerulonephritis*	10 (25.0)	16 (41.0)	0.026
	Other	30 (75.0)	23 (59.0)	
Vaccinated ≤1 year post transplant	Yes	3 (7.5)	25 (64.1)	<0.0001
	No	37 (92.5)	14 (35.9)	
Immunosuppression at diagnosis	CNI monotherapy*	21 (52.5)	10 (25.6)	0.015
	Other	19 (47.5)	29 (74.4)	
Induction immunotherapy	Alemtuzumab	29 (72.5)	16 (41.0)	0.0005
	Other	11 (27.5)	23 (59.0)	
Transplant number	1 st	36 (90.0)	33 (84.6)	0.47
	≥2 nd	4 (10.0)	6 (15.4)	
Diabetes	No	23 (57.5)	32 (82.1)	0.018
	Yes	17 (42.5)	7 (17.9)	

ESKD – end stage kidney disease, CNI - calcineurin inhibitors

Table S9. Analysis of clinical characteristics associated with detectable T-cell responses in infection-naïve patients after the first-year post-transplant

Nine of 79 (11.4%) infection-naïve patients had detectable T-cell responses post-vaccination, none of the 28 patients vaccinated within the first-year post-transplant and 9/51 (17.6%) of patients vaccinated after the first post-operative year, $p=0.019$. Analysis was performed of independent variables associated with T-cell responses in infection-naïve patients after the first-year post-transplant.

Variable	Reference Group	Univariable	
		OR (95% CI)	p value
Age	≥60 years	0.23 (0.03-1.98)	0.18
Gender	Male	0.50 (0.11-2.19)	0.36
Cause of ESKD	Glomerulonephritis	1.41 (0.30-6.62)	0.66
Number of transplants	≥2	0.93 (0.09-9.03)	0.95
Transplant induction agent	Alemtuzumab	1.75 (0.32-9.55)	0.52
Maintenance Immunosuppression	CNI monotherapy	0.17 (0.19-3.78)	0.17
Vaccine	BNT1262b2	1.40 (0.25-7.72)	0.70
Diabetes	Yes	1.3 (0.30-5.57)	0.72

Table S10. Characteristics associated with lack of detectable immunological (serological and T-cell) response in 79 infection-naïve transplant patients

Variable	Reference Group	Denominator	Event rate (%)	Univariable		Multivariable	
				OR (95% CI)	p value	OR (95% CI)	p value
Age	≥60	23	10 (43.5)	0.95 (0.36-2.54)	0.92		
Gender	Male	55	25 (45.5)	1.17 (0.44-3.08)	0.76		
Ethnicity	Caucasian	37	16 (43.2)	0.92 (0.38-2.25)	0.86		
Cause ESKD	Glomerulonephritis	26	8 (30.8)	0.43 (0.16-1.15)	0.09	-	
Time of vaccine post-transplant	<1 year	28	5 (17.9)	0.15 (0.5-0.46)	0.0009	0.19 (0.04-0.96)	0.045
Number of transplants	≥2	10	4 (40.0)	0.82 (0.21-3.16)	0.77		
Transplant induction agent	Alemtuzumab	45	27 (60.0)	4.88 (1.87-13.79)	0.0017	1.89 (0.49-7.35)	0.30
Maintenance Immunosuppression	CNI monotherapy	31	25 (80.6)	15.83 (5.11-49.06)	<0.0001	13.3 (3.68-48.14)	<0.0001
Diabetes	Yes	24	11 (45.8)	1.09 (0.42-2.87)	0.86		
Vaccine	BNT162b2	40	25 (62.5)	4.83 (1.85-12.65)	0.0013	1.63 (0.42-6.55)	0.46

CNI - calcineurin inhibitors

Figure S1. Correlation between anti-S concentrations and clinical characteristics in infection-naïve patients.

- a. Kidney transplant patients who were receiving calcineurin inhibitor (CNI) monotherapy had significantly higher anti-S concentrations, 75 (7.1-646) BAU/ml, compared with patients receiving CNI in combination with anti-proliferative agents (mycophenolate mofetil or azathioprine), with or without corticosteroids, 7.1 (7.1-28) BAU/ml, $p < 0.0001$
- b. Comparing patients receiving combination therapy alone, there was no difference between those who had received Alemtuzumab induction, 7.1 (7.1-67) BAU/ml compared with IL-2 receptor antibodies, 7.1 (7.1-7.12) BAU/ml, $p = 0.06$. Comparing patients who received Alemtuzumab as induction alone, those who are maintained on CNI monotherapy, 73 (7.1-656) BAU/ml, had significantly higher anti-S than those on combination therapy, $p < 0.0001$.
- c. There was no difference in anti-S between females, 17 (7.1-264) BAU/ml compared with males, 11 (7.1-223) BAU/ml, $p = 0.76$
- d. There was no correlation between cause of end stage kidney disease (ESKD) and anti-S concentrations. Patients with ESKD secondary to diabetes had a median anti-S concentration of 9 (7.1-174) BAU/ml, polycystic kidney disease (APKD), 18 (7.1-415) BAU/ml, Glomerulonephritis (GN), 9 (7.1-238), Urological causes of ESKD, 14 (7.1-205) BAU/ml, and unknown causes of ESKD, 15 (7.1-221) BAU/ml, $p = 0.83$
- e. There was no correlation between ethnicity and anti-S concentrations. Patients from Indoasian, Black and White backgrounds having a median anti-S concentration of 19 (7.1-272), 7.1 (7.1-51) and 13 (7.1-270) respectively, $p = 0.13$.
- f. Patients who were within their first-year post-transplant when vaccinated had significantly lower anti-S, 7.1 (7.1-21) BAU/ml compared with those patients who were vaccinated after the 1st year, 17 (7.1-271) BAU/ml, $p < 0.0001$.
- g. A significant inverse correlation was seen between anti-S response and age in patients who received BNT162b2, $r = -0.14$, $p = 0.0038$ (i), but not in patients who received ChAdOx1, $r = -0.08$, $p = 0.13$ (ii).

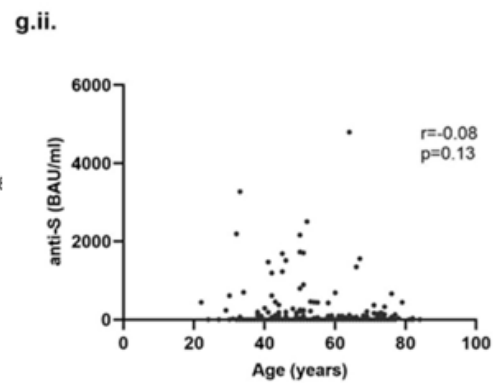
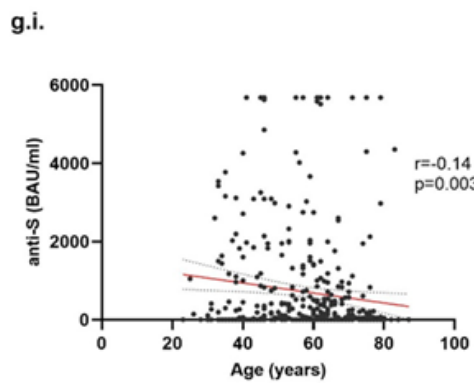
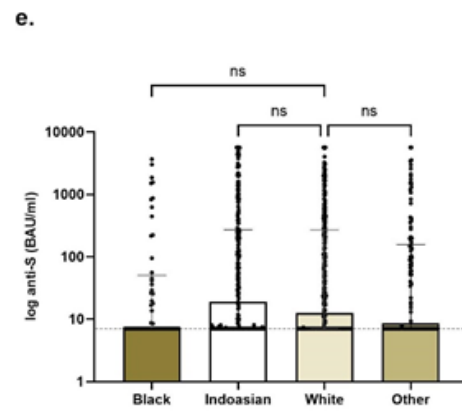
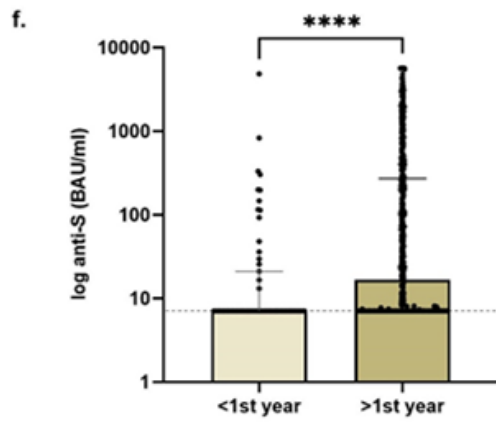
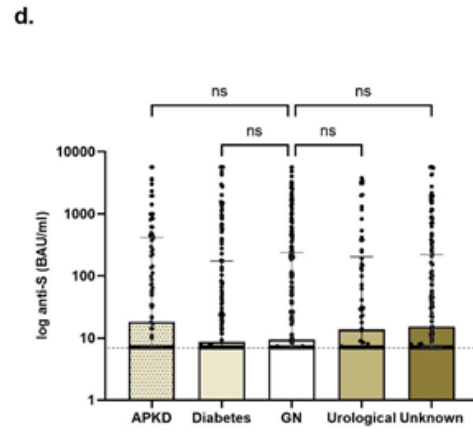
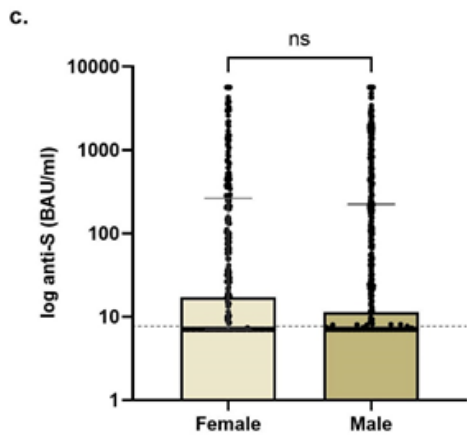
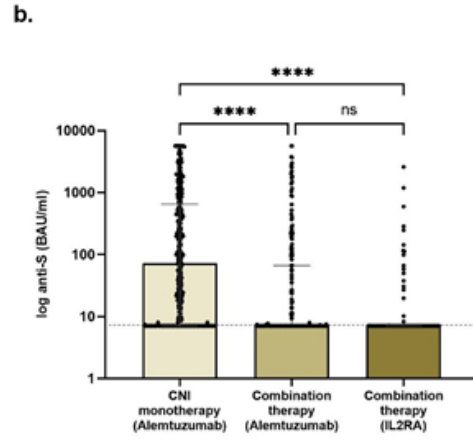
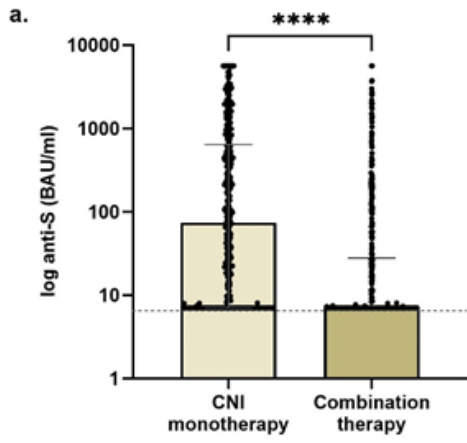


Figure S2. Correlation between anti-S concentrations and diagnostic characteristics in patients with prior infection.

- a. Anti-S concentrations in patients who were diagnosed by PCR compared with serology were 324 (111-844) and 758 (172-1985) BAU/ml respectively, $p=0.59$, in patients receiving ChAdOx1; and 1659 (546-3684) and 2752 (641-5680) BAU/ml respectively, $p=0.76$, in patients receiving BNT162b2.
- b. Anti-S concentrations in patients who were anti-NP positive compared with negative at the time of testing, were 503 (37-2094) and 701 (153-1694) BAU/ml respectively, $p=0.91$, in patients receiving ChAdOx1; and 2512 (499-5680) and 2350 (780-5022) BAU/ml respectively, $p=0.99$, in patients receiving BNT162b2

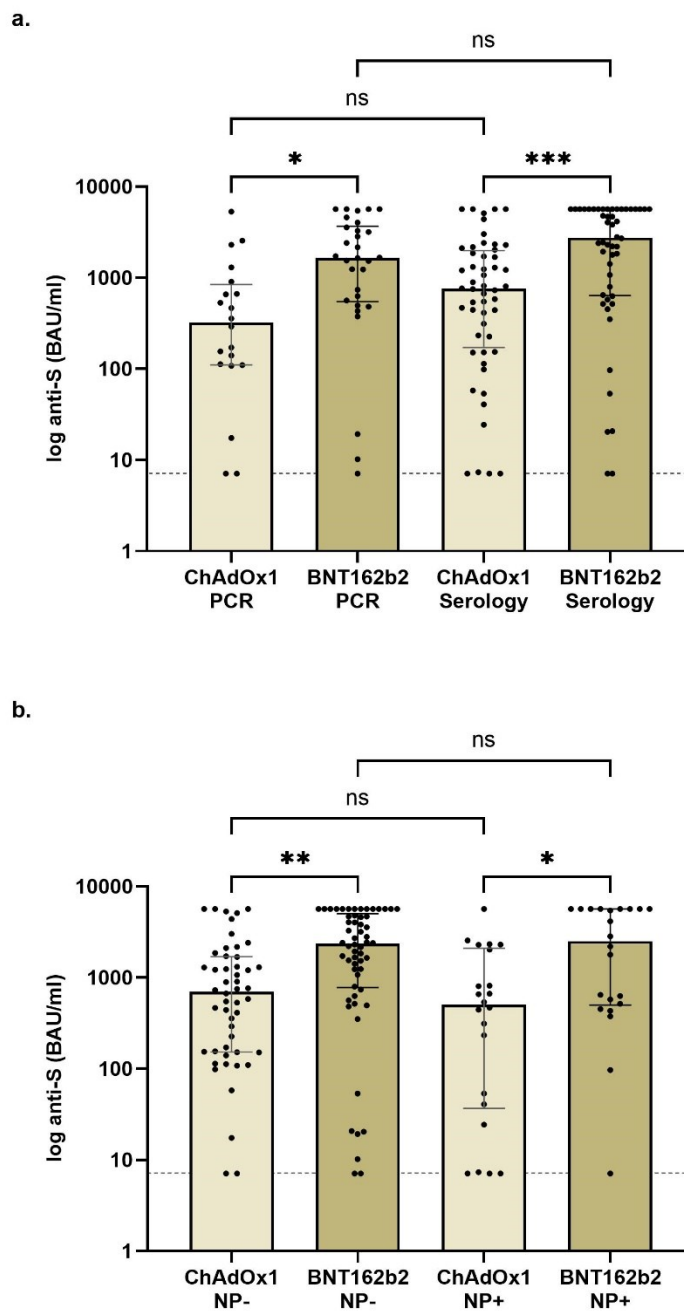
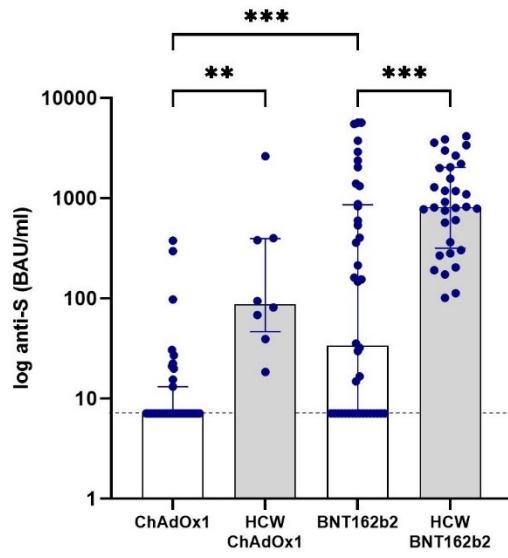


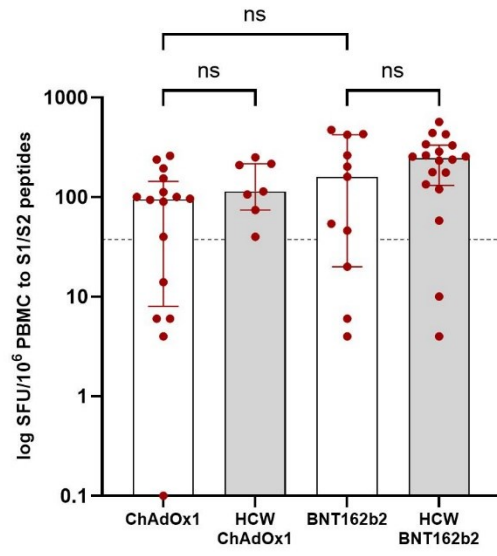
Figure S3. Immunological responses to SARS-CoV-2 vaccination in 106 kidney transplant recipients who underwent paired serological and T cell assessments

- a. Anti-S concentrations in infection-naïve patients who received BNT162b2 were higher compared with those patients who had received ChAdOx1, with anti-S concentrations of 34 (7.1-861) BAU/ml and 7.1 (7.1-13) BAU/ml respectively, $p=0.0005$. These anti-S concentrations in patients were significantly lower than infection naïve HCW who received the corresponding vaccine, with HCW receiving BNT162b2 and ChAdOx1 having a median anti-S of 815 (318-2033), $p=0.0003$, and 88 (47-395) BAU/ml, $p=0.01$, respectively. Black dotted line represents 7.1 BAU/ml, the cut off for a positive result.
- b. There was no difference in median SFU/ 10^6 PBMCs in previously exposed patients who received BNT162b2 compared with ChAdOX1, with 160 (20-422) SFU/ 10^6 PBMCs and 95 (8-144) SFU/ 10^6 PBMCs respectively, $p=0.42$. There were no differences between T-cell responses in previously exposed patients and HCW receiving the corresponding vaccine, with HCW receiving BNT162b2 and ChAdOx1 having a median 246 (131-332) SFU/ 10^6 PBMCs, $p=0.63$ and 114 (74-216) SFU/ 10^6 PBMCs, $p=0.85$ respectively. Data points of 0 SFU/ 10^6 PBMCs are represented as 0.1 for visualisation on a log scale. Black dotted line indicates threshold for a positive ELISpot, 40 SFU/ 10^6 PBMCs which was calculated from unvaccinated, infection naïve HCW.
- c. There was no difference in anti-S concentrations in patients with prior infection who received BNT162b2, 1238 (497-2829) BAU/ml, compared with ChAdOx1, 366 (119-1273) BAU/ml, $p=0.07$. There were also no differences between patients and HCW with prior exposure who received BNT162b2, median anti-S 2189 (1236-3303) BAU/ml, $p=0.72$, or HCW who received ChAdOx1, median anti-S 753 (574-867) BAU/ml, $p=0.91$. Black dotted line represents 7.1 BAU/ml, the cut off for a positive result.

a.



b.



c.

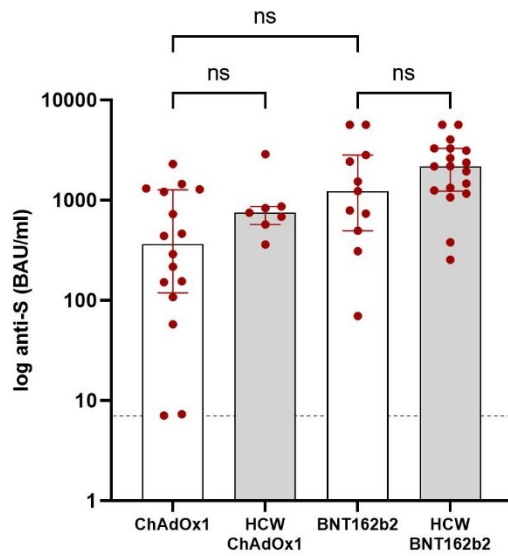
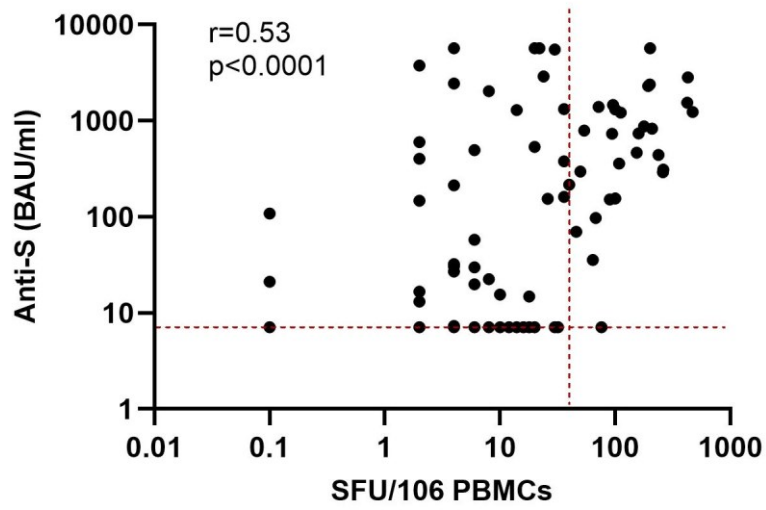


Figure S4. Correlation between anti-S (BAU/ml) and T-cell responses (SFU/10⁶ PMBCs)



1 **Supplemental Methods**

3 ***Study population and objectives***

4 We undertook a cohort study of 920 kidney transplant patients who had received 2 doses of either
5 BNT162b2 or ChAdOx1. The aim of the study was to evaluate immune responses to 2 dose vaccination
6 with either BNT162b2 or ChAdOx1. Patients were included if they were able to provide consent for
7 research and were participating in the vaccination programme. Participants were vaccinated as part of
8 their usual care, with two doses of either 0.5mL ChAdOx1-S (Oxford AstraZeneca) or 30ug BNT162b2
9 (Pfizer-BioNTech) at the specified dosing interval.

10 All patients were recruited from within Imperial College Renal and Transplant Centre into one of two
11 prospective studies. The first is the OCTAVE study, an Observational Cohort Study of T-cells,
12 Antibodies and Vaccine Efficacy in SARS-CoV-2 in people with chronic diseases and/or secondary
13 immunodeficiency, which is part of the UK COVID-19 Immunity National Core Study Programme.
14 The OCTAVE study was approved by the Health Research Authority, Research Ethics Committee
15 (Reference:21/HRA/0489). The second study is a prospective longitudinal study ‘The effect of COVID-
16 19 on Renal and Immunosuppressed patients’, sponsored by Imperial College London. This study was
17 approved by the Health Research Authority, Research Ethics Committee (Reference: 20/WA/0123).

18 A subgroup of the first 106 (11.5%) patients recruited underwent more in-depth immunological analysis
19 of serological and cellular responses to SARS-CoV2 vaccination. A group of 65 healthcare workers
20 (HCW), with a median age of 38 (30-46) years, were used as a comparator. Fifty and 15 HCW received
21 the BNT162b2 and ChAdOx1 vaccines, respectively. The median interval between vaccinations in the
22 HCW was 68 (61-70) days, with a median time to sampling post-vaccination of 28 (21-28) days.

23 Anonymised data were collected on age, gender, ethnicity, primary renal disease, diabetes, induction
24 and maintenance immunosuppression, date(s) and number of renal transplants. Under the terms of our
25 ethical approval for healthy volunteers data were collected only on age, gender and ethnicity.

27 ***Serological testing***

28 Serum was tested for antibodies to both the nucleocapsid protein (anti-NP) and spike protein (anti-S).
29 Anti-NP was tested using the Abbott Architect SARS-CoV-2 IgG 2 step chemiluminescent
30 immunoassay (CMIA) according to manufacturer’s instructions. This is a non-quantitative assay and
31 samples were interpreted as positive or negative with a threshold index value of 1.4. The presence of
32 anti-NP was used as a marker of natural infection. For vaccine responses, spike protein antibodies (anti-
33 S IgG) were assessed using the Abbott Architect SARS-CoV-2 IgG Quant II CMIA. Anti-S antibody
34 titres are quantitative with a threshold value of 7.1 BAU/ml for positivity, and an upper level of
35 detection of 5680 BAU/ml.

37 ***T cell ELISpot***

38 SARS-CoV-2 specific T-cell responses were detected using the T-SPOT® Discovery SARS-CoV-2
39 (Oxford Immunotec) according to the manufacturer's instructions. In brief, peripheral blood
40 mononuclear cells (PBMCs) were isolated from whole blood samples with the addition of T-Cell
41 Xtend™ (Oxford Immunotec) if samples were taken more than 12 hours prior to processing. All samples
42 were processed within 24 hours of venepuncture. 250,000 PBMCs were plated into individual wells of
43 a T-SPOT® Discovery SARS-CoV-2 plate. The assay measures immune responses to SARS-CoV-2
44 spike protein peptide pools (S1 protein and S2 protein), in addition to positive PHA
45 (phytohemagglutinin) and negative controls. Cells were incubated and interferon- γ secreting T cells
46 were detected. Spot forming units (SFU) were detected using an automated plate reader (Autoimmun
47 Diagnostika). Spot count in the negative well was subtracted from spot count for spike protein wells
48 and this was used to calculate spike protein specific responses per 10^6 PBMC. Assays with a negative
49 PHA response were deemed to be failed assays and excluded from analysis (n=1 for HCW and n=2 for
50 transplant recipients). Infection-naïve, unvaccinated participants were used to identify a threshold for a
51 positive response using mean +3 standard deviation SFU/ 10^6 PBMC, as previously described. This
52 resulted in a cut-off for positivity of 40 SFU/ 10^6 PBMC (1).

53

54 ***Definition of prior infection***

55 Prior infection was defined serologically or via past PCR positive confirmed infection. The detection
56 of anti-NP on current or historic samples, and the presence of anti-S at baseline (pre-vaccine) or historic
57 samples, was required for the definition of prior infection by serological methods. Serological screening
58 of transplant patients had commenced in our centre in June 2020, and these data were used to aid
59 identification of patients with prior SARS-CoV-2 exposure prior to vaccination. As part of this
60 protocol, patients were initially screened for anti-NP, and those with a subthreshold anti-NP index value
61 (0.25-1.4), underwent confirmatory testing for natural infection by assessing for receptor binding
62 domain (anti-RBD) antibodies. This was performed using an in-house double binding antigen ELISA
63 (Imperial Hybrid DABA; Imperial College London, London, UK), which detects total RBD antibodies
64 (2-4). The HCW had serological analysis at baseline, which was used to define prior infection alone.

65

66 ***Statistical Analysis***

67 Statistical analysis was conducted using Prism 9.0 (GraphPad Software Inc., San Diego, California).
68 Unless otherwise stated, all data are reported as median with interquartile range (IQR). The aim of the
69 study was to investigate the seroconversion rates in transplant patients receiving either BNT162b2 or
70 ChAdOx, and identify clinical variables associated with **reduced** immunogenicity. In addition, T-cell
71 responses were investigated in a subcohort of patients.

72

73 The Chi-squared test was used for proportional assessments; seroconversion and ELISpot positivity in
74 patients receiving BNT162b2 and ChAdOx1 in both infection-naïve and infection-experienced patients.

75 The Mann-Whitney and Kruskal-Wallis tests were used to assess the difference between 2 or >2 groups,
76 with Dunn's post-hoc test to compare individual groups. A comparison of anti-S concentrations and T-
77 cell responses, in both infection naïve and infection-experienced patients receiving BNT162b2 and
78 ChAdOx1 was undertaken.

79
80 To investigate clinical features associated with seroconversion following vaccination, proportional
81 assessments of clinical characteristics in patients seroconverting compared with those failing to
82 seroconvert was performed. A comparative analysis of clinical features associated with seroconversion
83 by vaccine type was undertaken. Multivariable analysis was carried out using multiple logistic
84 regression using variables which had a p value of <0.15 on univariable analysis.

85

86 ***Acknowledgements and Funding***

87 The OCTAVE trial, which is part of the COVID-19 Immunity National Core Study Programme, was
88 sponsored by the University of Birmingham and funded by a grant from UK Research and Innovation
89 (UKRI) administered by the Medical Research Council (grant reference number MC_PC_20031). It has
90 been designated an Urgent Public Health (UPH) study by the National Institute of Health Research.

91 This research is also supported by the National Institute for Health Research (NIHR) Biomedical
92 Research Centre based at Imperial College Healthcare NHS Trust and Imperial College London. The
93 authors would like to thank the West London Kidney Patient Association, all the patients and staff at
94 ICHNT (The Imperial COVID vaccine group and dialysis staff, and staff within the North West London
95 Pathology laboratories). The authors are also grateful for support from Hari and Rachna Murgai, Milan
96 and Rishi Khosla OBE, The Nan Diamond Fund, Sidharth and Indira Burman, and the Auchy Charitable
97 Foundation. MP is supported by an NIHR clinical lectureship. Work in DCT's lab is supported by a
98 Wellcome Trust Clinical Career Development Fellowship

99

100 ***Role of the funding source***

101 The funding body had no role in data collection, analysis, interpretation, writing of the manuscript or
102 the decision to submit for publication

103

104 ***Data sharing***

105 Individual participant data that underlie the results reported in this article, after de-identification will
106 be shared upon reasonable request to the corresponding author

107

108 ***Contributor statement***

109 MP- Investigation, data curation, data analysis, writing - original draft, review, and editing,
110 conceptualisation, accessed and verified data

111 TT- Investigation, data curation, writing-review and editing

112 CC- Investigation, data analysis, writing-review and editing, conceptualisation, accessed and verified
113 data
114 PMa, SG, RDC, HE, PMo, SMcI, DM, AC, GP- Investigation, writing-review and editing
115 LL- Writing-review and editing, supervision
116 DT- Writing-review and editing, supervision
117 SPM- Writing-review and editing, supervision, conceptualisation
118 PK- Investigation, writing - original draft, review, and editing, conceptualisation
119 MW- Investigation, data curation, data analysis, writing - original draft, review, and editing,
120 conceptualisation, accessed and verified data
121

122 The corresponding authors had full access to all the data and the final decision to submit for
123 publication.
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154

155 1. Predecki M, Clarke C, Brown J, Cox A, Gleeson S, Guckian M, et al. Effect of previous SARS-
156 CoV-2 infection on humoral and T-cell responses to single-dose BNT162b2 vaccine. *Lancet* (London,
157 England). 2021;397(10280):1178-81.

158 2. Rosadas C, Randell P, Khan M, McClure MO, Tedder RS. Testing for responses to the wrong
159 SARS-CoV-2 antigen? *Lancet*. 2020;396(10252):e23.

160 3. Predecki M, Clarke C, Gleeson S, Greathead L, Santos E, McLean A, et al. Detection of SARS-
161 CoV-2 Antibodies in Kidney Transplant Recipients. *J Am Soc Nephrol*. 2020;31(12):2753-6.

162 4. Clarke C, Predecki M, Dhutia A, Ali MA, Sajjad H, Shivakumar O, et al. High Prevalence of
163 Asymptomatic COVID-19 Infection in Hemodialysis Patients Detected Using Serologic Screening. *J Am*
164 *Soc Nephrol*. 2020;31(9):1969-75.

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