

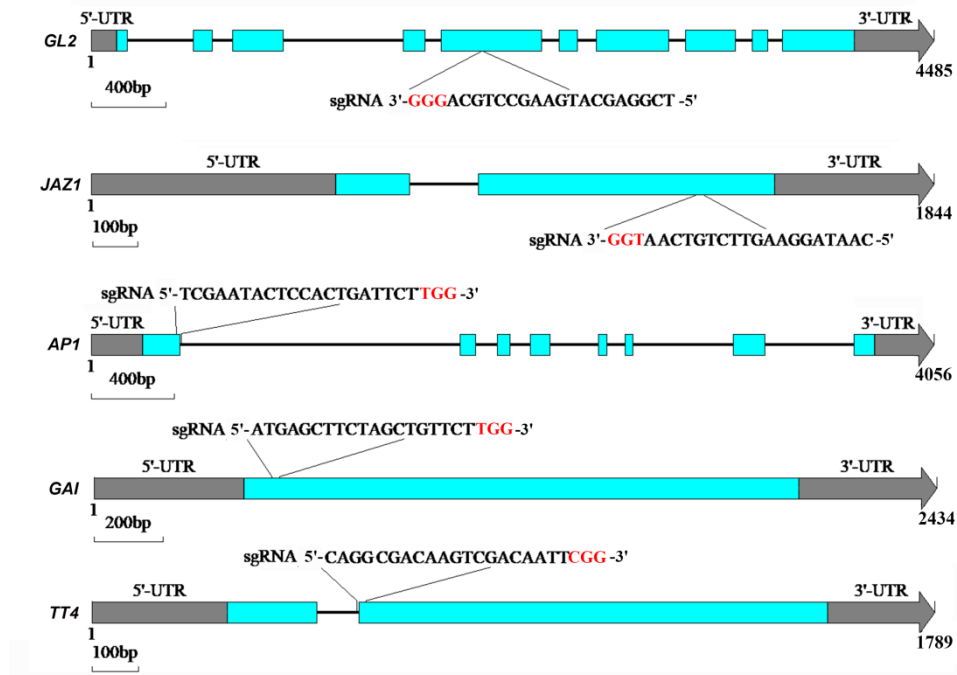
SUPPLEMENTAL MATERIAL

***GLABRA2*-based selection efficiently enriches Cas9-generated non-chimeric *Arabidopsis* mutants in T1 generation**

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Supplemental Figure S1 The target sites of genes for monogenic mutation used in this study. The structures of *GL2*, *AP1*, *TT4*, *JAZ1*, and *GAI* are represented by exons (light blue bars), introns (black lines), and untranslated regions (UTRs) (gray bars). The sgRNA target sites are indicated and the protospacer adjacent motif (PAM) sites are highlighted in red.

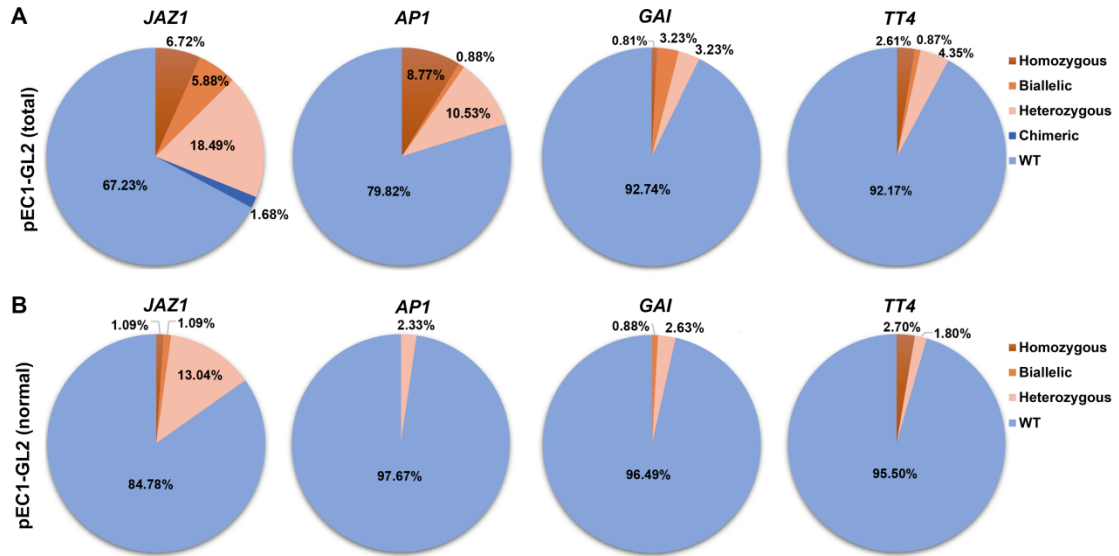
gl2 #1 Allele1: GCAGCTCTTGG-----CTTCATGC (-23 bp)
Allele2: GCAGCTCTTGG-----CTTCATGC (-23 bp)
WT: GCAGCTCTTGGACGCACTCCCTATCCCCTGCAGGCTTCATGC

gl2 #6 Allele1: CGCACTCCCTATCCCCTGC-----TCCGACGATCA (-11 bp)
Allele2: CGCACTCCCTATCCCCTGC**A**AGGCTTCATGCTCCGACGATCA (+1 bp)
WT: CGCACTCCCTATCCCCTGC-AGGCTTCATGCTCCGACGATCA

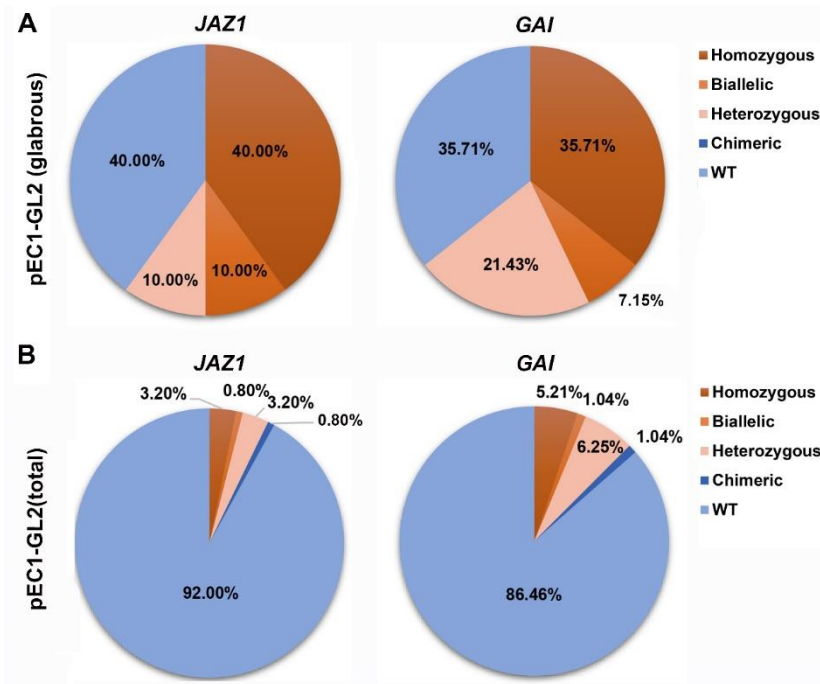
gl2 #11 Allele1: CGCACTCCCTATCCCCTGCA-GCTTCATGCTCCGACGATCAAG (-1 bp)
Allele2: CGCACTCCCTATCCCCTGCA-----TGCTCCGACGATCAAG (-7 bp)
WT: CGCACTCCCTATCCCCTGCAGGCTTCATGCTCCGACGATCAAG

gl2 #21 Allele1: CGCACTCCCTATCCCCTGC-----TGCTCCGACGATCAAG (-8 bp)
Allele2: CGCACTCCCTATCCCCTGC-----TGCTCCGACGATCAAG (-8 bp)
WT: CGCACTCCCTATCCCCTGCAGGCTTCATGCTCCGACGATCAAG

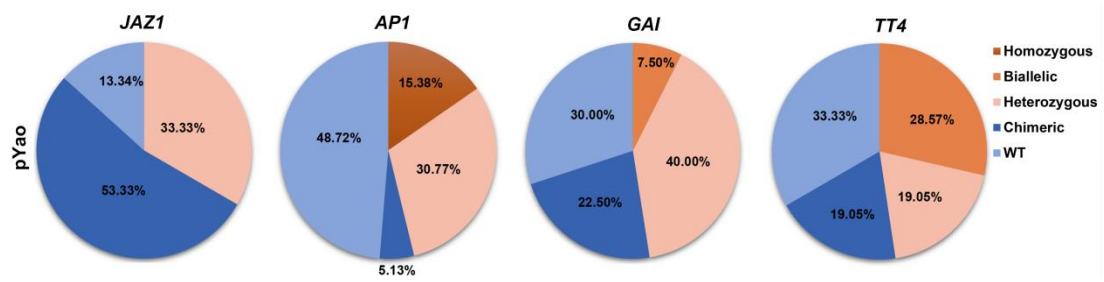
Supplemental Figure S2 DNA sequences of the *gl2* mutants. The reference sequence of WT is shown on the bottom. The indels in mutant alleles are depicted by a red dash (deletion) or red letter (insertion).



Supplemental Figure S3 Analyses of the frequencies of mutations for all transgenic T1 plants (total) and those with normal trichomes (normal) group obtained using the pEC1-GL2 vector. A, Percentages of WT and different types of mutation at four target sites among all T1 plants obtained using pEC1-GL2 vector. B, Percentages of WT and different types of mutation at four target sites among T1 plants with normal trichomes obtained using the pEC1-GL2 vector.



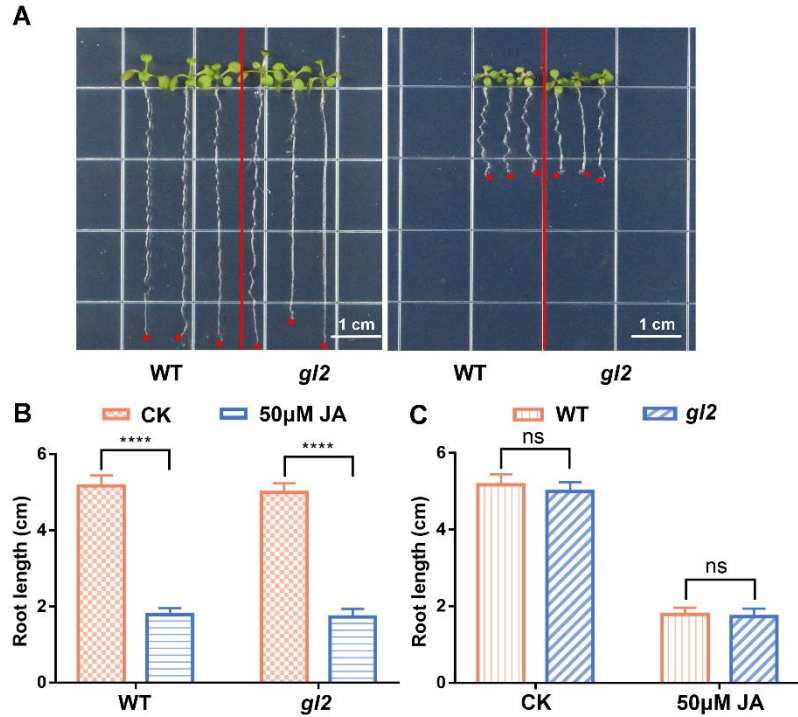
Supplemental Figure S4 Analyses of the frequencies of *JAZ1* and *GAI* mutations for all transgenic T1 plants (total) and glabrous (glabrous) group obtained using the pEC1-GL2 vector. A, Percentages of WT and different types of mutation at *JAZ1* and *GAI* target sites among glabrous T1 plants obtained using pEC1-GL2 vector. B, Percentages of WT and different types of mutation at *JAZ1* and *GAI* target sites among all T1 plants obtained using the pEC1-GL2 vector.



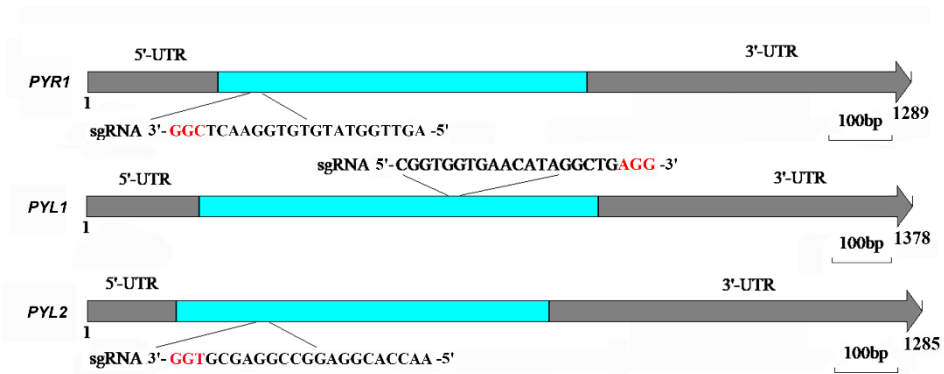
Supplemental Figure S5 Analyses of the frequencies of mutation in T1 plants obtained using the pYao vector. Percentages of WT and different types of mutation are shown for four target sites.



Supplemental Figure S6 Mutation of *GL2* does not affect flower development or seed color. A, Representative flowers from WT and the *gl2* mutant. B, Seeds of WT and the *gl2* mutant.



Supplemental Figure S7 Mutation of *GL2* does not affect plant response to JA. A, Representative T2 seedlings of WT and the *gl2* mutant grown on 1/2 MS plates with or without 50 μM JA. B, Analyses of primary root length to compare the JA response of WT and the *gl2* mutant. C, Analyses of primary root length to compare the growth of WT and the *gl2* mutant on 1/2 MS plates with or without 50 μM JA. Data are shown as mean ± SD (n = 15). ns: no significant difference; ****: P<0.0001 determined by Student's *t*-test.



Supplemental Figure S9 The target sites of genes for polygenic mutation used in this study. The structures of *PYR1*, *PYL1*, and *PYL2* are represented by exons (light blue bars), and untranslated regions (UTRs) (gray bars). The sgRNA target sites are indicated and the protospacer adjacent motif (PAM) sites are highlighted in red.

■ Homozygous
 ■ Biallelic
 ■ Heterozygous
 ■ Chimeric
 ■ WT

Line \ Target	#1	#2	#3	#4	#5	#6	#7	#8	#9	#10	#11	#12	#13	#14	#15	#16	#17	#18	#19	#20
<i>PYR1</i>	Chimeric	Heterozygous	Chimeric	Biallelic	Chimeric	Heterozygous	Chimeric	Chimeric	Biallelic	Chimeric	Biallelic	Chimeric	Chimeric	Chimeric	Chimeric	Heterozygous	Biallelic	Chimeric	Chimeric	Chimeric
<i>PYL1</i>	Heterozygous	Heterozygous	Biallelic	Biallelic	Chimeric	Chimeric	Homozygous	Chimeric	Heterozygous	Chimeric	Chimeric	Chimeric	Chimeric	Heterozygous	Chimeric	Chimeric	Biallelic	Chimeric	Chimeric	Chimeric
<i>PYL2</i>	Chimeric	Chimeric	Chimeric	Heterozygous	Chimeric	Chimeric	Heterozygous	Chimeric	Chimeric	Chimeric	Chimeric	Chimeric	Chimeric	Chimeric	Chimeric	Chimeric	Heterozygous	Chimeric	Chimeric	Chimeric

Supplemental Figure S10 Mutation types at each target sites of multiplex edited T1 plants with normal trichomes. The types of mutation at *PYR1*, *PYL1* and *PYL2* loci in individual T1 plants with normal trichomes were analyzed.

Supplemental Table S1 Numbers of all transgenic T1 plants and glabrous T1 plants

for each target gene obtained using the pEC-GL2 vector.

Target	T1 (total)	T1 (glabrous)	glabrous /total
<i>JAZ1</i>	119	27	22.69%
<i>API</i>	114	28	24.56%
<i>GAI</i>	124	10	8.06%
<i>TT4</i>	115	4	3.48%

Supplemental Table S2 Mutation frequencies of four target genes using different CRISPR-Cas9 systems.

	<i>JAZ1</i>	<i>API</i>	<i>GAI</i>	<i>TT4</i>
pYao	86.66%	51.28%	70.00%	66.67%
pEC1	35.91%	10.00%	14.64%	22.50%
pEC1-GL2	32.77%	20.18%	7.26%	7.83%
pEC1-GL2 (glabrous)	92.59%	75.00%	50.00%	100.00%

Supplemental Table S3 Primers used in this study

Primer name	Primer sequence
GL2-sgRNA-F	ATTGTCGGAGCATGAAGCCTGCA
GL2-sgRNA-R	AAACTGCAGGCTTCATGCTCCGA
AP1-sgRNA194-F	ATTGTCGAATACTCCACTGATTCT
AP1-sgRNA194-R	AAACAGAATCAGTGGAGTATTCGA
JAZ1-sgRNA1-F	ATTGCAATAGGAAGTCTGTCAA
JAZ1-sgRNA1-R	AAACTTGACAGAACTTCCTATTG
TT4-sgRNA296-F	ATTGCAGGCGACAAGTCGACAATT
TT4-sgRNA296-R	AAACAATTGTCGACTGTGCGCCTG
GAI-sgRNA1-F	ATTGATGAGCTTCTAGCTGTCT
GAI-sgRNA1-R	AAACAGAACAGCTAGAAGCTCAT
GL2-2Tar-F	TAGAGTCGAAGTAGTGATTGTGAGACCTGAATTCAGGTCTCAGTTTTAGAGCTAGAAATAG C
GL2-2Tar-R	GCTATTTCTAGCTCTAAAACCTGCAGGCTTCATGCTCCGACAATCTCTTAGTCGACTCTA
YAOp-F	CAATACCATGGTTATACTAGTGATGGGAAATTCATGAAAAC
YAOp-R	GGTCTTGTAATCCATCTAGATTCTTCTCTCTCACTCCCTC
4DT-1F	TAGAGTCGAAGTAGTGATTGAGTTGGTATGTGTGGAAGTGTGTTAGAGCTAGAAATAG
4DT-1R	CAGCCTATGTTACCACCGAATCTCTTAGTCGACTCTA
4DT-2F	GGTGGTGAACATAGGCTGGTTTTAGAGCTAGAAATAGC
4DT-2R	GCTATTTCTAGCTCTAAAACGGTACCCGCTCCGGCCTCCGTGGTTCAATCACTACTTCGTCTC TA
4DT-3F	ACCACGGAGGCCGGAGCGGTTTTAGAGCTAGAAATAGC
4DT-3R	GCTATTTCTAGCTCTAAAACCTGCAGGCTTCATGCTCCGACAATCACTACTTCGACTCTA
GL2-IF	GTTCCAAAACCGCCGCA
GL2-IR	TAAACACAATCCCCGCATCTC
AP1-IF	TCTCCATAAATAAAGATCCCGAGAC
AP1-IR	CTGGCTAATCAAAAAATCAAAAA
JAZ1-IF	TTGACTATATTCTACGCCGGGC
JAZ1-IR	GTAACAAAATGAAAACAAAACACACCT
TT4-IF	ATAATGGTGATGGCTGGTGCT
TT4-IR	TCAAGGTGGGTGTCAGAGGG
GAI-IF	CCCCATCACGAGCCTTTTTAT
GAI-IR	CGTCGCTGTAGTGGTTCCAC
PYR1-IF	CATGTTACCTTCATCTTCCT
PYR1-IR	ATCATCTCCGAGTTACCTTCC
PYL1-IF	CCAACTCTCCCAATCAATCGCC
PYL1-IR	CCCCAAATTCATCATTACCTAACC
PYL2-IF	ACAAACCCCAACCTCAA
PYL2-IR	TCGACGAACTCAAGCCGCTC

Supplemental Data Set S1 Sequences for plasmids used in this paper.

pEC1-GL2 plasmid:

GTTTACCCGCCAATATATCCTGTCAAACACTGATAGTTTAACTGAAGGCGGGAAACGA
CAATCTGATCCAAGCTCAAGCTGCTCTAGCATTCCGCCATTCAGGCTGCGCAACTGTTGG
GAAGGGCGATCGGTGCGGGCCTCTTCGCTATTACGCCAGCTGGCGAAAGGGGGATGTG
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pYao plasmid:

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