

APPENDIX for “Multistability maintains redox homeostasis in human cells”

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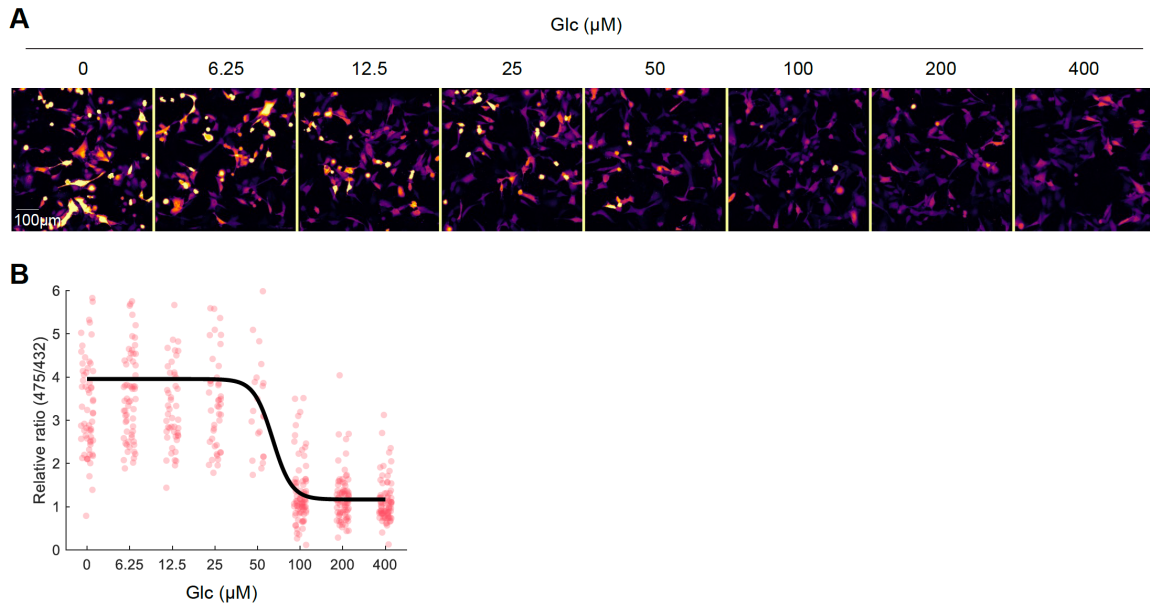
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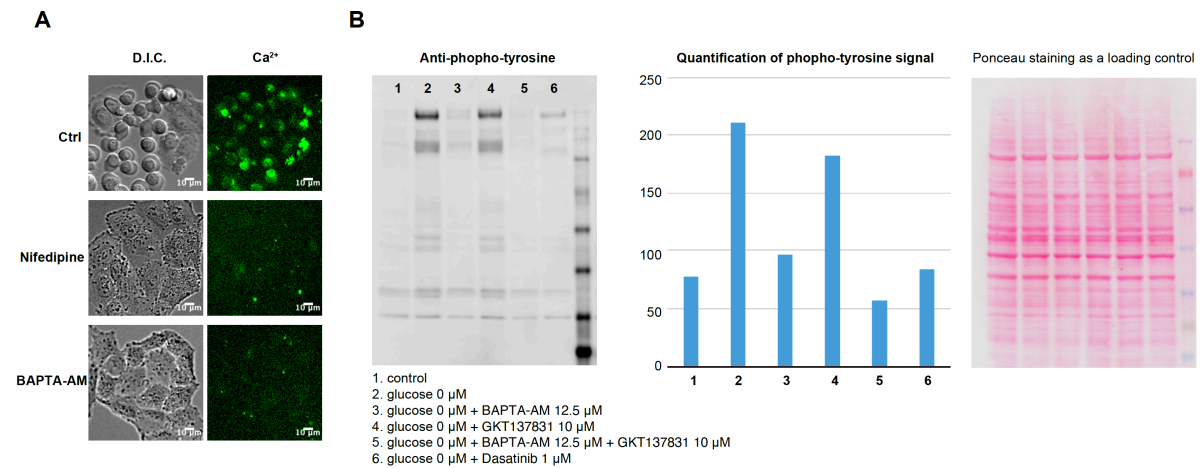
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Appendix Figure S1. Ultrasensitivity of the ROS-response curve in U87-MG cells

(A) Representative HyPer7 images of U87-MG-HyPer7 cells 10 h after glucose deprivation (from 400 μM to 0 μM). (B) A dose-response curve of ROS 10 h after glucose deprivation. For each glucose concentration, the ROS levels of 80 cells were quantified. To deconvolute the effects of non-responsive cells, they were not considered in the analyses of the lower five glucose concentrations (0 μM , 6.3 μM , 12.5 μM , 25 μM and 50 μM). A Hill exponent (n) was fitted to be 6.5, with a 95% confidence interval of 2-11.



Appendix Figure S2. Validation of calcium and NOX inhibitors.

(A) Representative differential interference contrast (D.I.C., left panel) and fluorescent images of calcium indicator Fluo-4 AM (right panel) of LN18 cells 3 h after glucose deprivation. Cells were pre-treated with Nifedipine (10 μ M) and BAPTA-AM (12.5 μ M) 1 h before glucose deprivation to inhibit calcium signaling. (B) Activity of tyrosine kinase signaling was probed with western blot analysis using an anti-phospho-tyrosine antibody (4G10, Cell Signaling Technology) under normal [glucose] (band 1), no glucose (band 2), and no glucose with addition of BAPTA-AM alone (band 3), GKT137831 alone (band 4) and both (band 5). A multi-targeted tyrosine kinase inhibitor, Dasatinib (1 μ M, band 6), was used as a positive control for suppression of tyrosine kinase signaling. The middle and right panels show the quantifications of phospho-tyrosine and the protein loading control respectively.

Appendix Table S1 - Variable Concentration Ranges

Molecule	Location	Organism	Concentration	Unit	Ref	Assumption made to convert unit	Converted quantity	Converted unit	Note
SLC7A11	Cell	Human	200 ppm		PaxDB (https://pax-db.org/protein/1846679); See table 002	10000 ppm ~ 500 uM (See Table EV2)	10 uM		Upper bound
GSH	Cell	Human	10 mM		Bionumbers #113865; Forman et al. Mol. Aspects Med. 2009	NA	10000 uM		Upper bound
GSH	Cell	Human	1 mM		Bionumbers #113865; Forman et al. Mol. Aspects Med. 2009	NA	1000 uM		Lower bound
GSH/GSSG	Cell	Human	100 NA		Zilka et al. Oncol. Lett. 2012	NA	100 NA		Upper bound
GSH/GSSG	Cell	Human	10 NA		Zilka et al. Oncol. Lett. 2012	NA	10 NA		Lower bound
GSSG	Cell	Human	1 mM		Calculated from above	NA	1000 uM		Upper bound
GSSG	Cell	Human	0.01 mM		Calculated from above	NA	10 uM		Lower bound
Glu	Cell	Human	20 mM		Bionumbers #100790	NA	20000 uM		NA
Glu	Cell	Human	10 mM		Featherstone. ACS Chem. Neurosci. 2010	NA	10000 uM		Upper bound
Glu	Cell	Human	5 mM		Featherstone. ACS Chem. Neurosci. 2010	NA	5000 uM		Lower bound
Cys2	Cell	Human	2 nmol / mg protein		Jamalpoor et al. Biomed. Chromatogr. 2017, Fig. 3	Cellular protein concentration 300 mg	600 uM		Cystinotic cell
Cys2	Cell	Human	0.05 nmol / mg protein		Jamalpoor et al. Biomed. Chromatogr. 2017, Fig. 3	Cellular protein concentration 300 mg	15 uM		Healthy cell
Cys	Cell	Human	1.5 nmol / mg protein		Sato et al. J. Biol. Chem. 2005, Fig. 7	Cellular protein concentration 300 mg	450 uM		NA
Cys	Cell	Human	4 nmol / mg protein		Stipanuk et al. J. Nutr. 2006, Fig. 4	Cellular protein concentration 300 mg	1200 uM		Lower bound
Cys	Cell	Human	6 nmol / mg protein		Stipanuk et al. J. Nutr. 2006, Fig. 4	Cellular protein concentration 300 mg	1800 uM		Upper bound
NADPH	Cell	Human	3 uM		Zou et al. Nat. Protoc. 2018, Table 2	NA	3 uM		NA
NADPH/NADP+	Cell	Human	300 NA		Zou et al. Nat. Protoc. 2018, Table 2	NA	300 NA		Upper bound
NADPH/NADP+	Cell	Human	15 NA		Zou et al. Nat. Protoc. 2018, Table 2	NA	15 NA		Lower bound
NADP+	Cell	Human	200 nM		Zou et al. Nat. Protoc. 2018, Table 2	NA	0.2 uM		Upper bound
NADP+	Cell	Human	10 nM		Zou et al. Nat. Protoc. 2018, Table 2	NA	0.01 uM		Lower bound
ROS	Cell	Human	0.01 uM		Giorgio et al. Nat. Rev. Mol. Cell Biol. 2007, Box 1	NA	0.01 uM		Proliferative
ROS	Cell	Human	0.1 uM		Giorgio et al. Nat. Rev. Mol. Cell Biol. 2007, Box 1	NA	0.1 uM		Proliferative
ROS	Cell	Human	1 uM		Giorgio et al. Nat. Rev. Mol. Cell Biol. 2007, Box 1	NA	1 uM		Growth arrest
ROS	Cell	Human	100 uM		Antunes and Cadenas. Free Radic. Biol. Med. 2001, Fig. 2	NA	100 uM		Cell death
ROS	Cell	Human	1 uM		Huang et al. ACS Synth. Biol. 2016, Fig. 4 and 5	NA	1 uM		Cell death
ROS	Cell	Human	50 nM		Sies. Redox Biol. 2017;	NA	0.5 uM		Cell death
ROS	Cell	Human	0.01 uM		Sies and Jones. Nat. Rev. Mol. Cell Biol. 2020	NA	0.01 uM		Proliferative
ROS	Cell	Human	0.1 uM		Sies. Redox Biol. 2017;	NA	0.1 uM		Stress;
ROS	Cell	Human	0.1 uM		Sies and Jones. Nat. Rev. Mol. Cell Biol. 2020	NA	0.1 uM		Stress;
ROS	Cell	Human	1 uM		Sies. Redox Biol. 2017;	NA	1 uM		Growth arrest; Cell death
ROS	Cell	Human	1 uM		Sies and Jones. Nat. Rev. Mol. Cell Biol. 2020	NA	1 uM		Growth arrest; Cell death
RTK	Cell	Human	100 nM		Take generic number from Cell Biology by the Numbers (http://book.bionumbers.org/what-are-the-absolute-numbers-of-signaling-proteins/)	NA	0.1 uM		NA
PPTase	Cell	Human	100 nM		Take generic number from Cell Biology by the Numbers (http://book.bionumbers.org/what-are-the-absolute-numbers-of-signaling-proteins/)	NA	0.1 uM		NA
Ca2+	Cell	Human	100 nM		Bionumbers #107490;	NA	0.1 uM		NA
Glc	RPMI-1640	NA	11.11 mM		Clapham. Cell 2007	NA	0.1 uM		NA
Gln	RPMI-1640	NA	2 mM		General RPMI-1640 recipe	NA	11110 uM		NA
Cys2	RPMI-1640	NA	0.2 mM		General RPMI-1640 recipe	NA	2000 uM		NA
Cys2	RPMI-1640	NA	0.2 mM		General RPMI-1640 recipe	NA	200 uM		NA

Appendix Table S2 - Estimation of SLCTA11 using Actin
Molecule **Location Organism** **Concentration Unit** **Ref** **Assumption made to convert unit** **Converted quantity** **Converted unit** **Note**

Total actin	Cell	Mammals	500 uM	Lodish et al. Freeman. 2000, Molecular Cell Biology, 4th Edition, Section 18.1	NA	500 uM	NA
Total actin	Cell	Acanthamoeba	200 uM	Pollard et al. Annu. Rev. Biophys. Biomol. Struct. 2000, Table 1	NA	200 uM	NA
Total actin	Cell	Dictyostellium	250 uM	Pollard et al. Annu. Rev. Biophys. Biomol. Struct. 2000, Table 1	NA	250 uM	NA
Total actin	Cell	Mammals (Neutrophils)	400 uM	Pollard et al. Annu. Rev. Biophys. Biomol. Struct. 2000, Table 1	NA	400 uM	NA
Total actin	Cell	Mammals (Platelets)	550 uM	Pollard et al. Annu. Rev. Biophys. Biomol. Struct. 2000, Table 1	NA	550 uM	NA
Polymerized actin	Cell	Eukaryotes	1000 uM	Bionumbers #109980	NA	1000 uM	NA
Total actin	Cell	Unspecified	100 uM	Bionumbers #109293	NA	100 uM	NA
Total G-actin	Cell	Unspecified	100 uM	Bionumbers #112131	NA	100 uM	NA
Total actin	Cell	Dictyostellium	250 uM	Bionumbers #109565	NA	250 uM	NA
ACTA1	Cell	Human	150 ppm	PaxDB (https://pax-db.org/protein/1854428)			normalized average
ACTB	Cell	Human	5000 ppm	PaxDB (https://pax-db.org/protein/1853523)			normalized average
ACTC1	Cell	Human	800 ppm	PaxDB (https://pax-db.org/protein/1847228)			normalized average
ACTG1	Cell	Human	5000 ppm	PaxDB (https://pax-db.org/protein/1851354)			normalized average
Total Actin	Cell	Human	10000 ppm	Summation over PaxDB entries	10000 ppm	~ 500 uM	normalized average
SLCTA11	Cell	Human	1.5 ppm	PaxDB (https://pax-db.org/protein/1846679)	10000 ppm	~ 500 uM	normalized average
SLCTA11	Cell	Human	200 ppm	PaxDB (https://pax-db.org/protein/1846679)	10000 ppm	~ 500 uM	Upper bound
Total protein	Cell	Human	10×10^9	Cell biology by the numbers; http://book.bionumbers.org/how-many-proteins-are-in-a-cell/			Cell volume ~ 3 pL; Avogadro's number ~ 6×10^{23}
			molecules				50 mM

Appendix Table S3 - Nutrient Uptake Km

Nutrient	Transporter	Measure	Value	Unit	Ref	Assumption made to convert unit	Converted quantity	Converted unit	Note
Glc	SLC2A1	Km	1.5 mM		Lodish et al. Freeman. 2000. Molecular Cell Biology. 4th Edition, Section 15.3	NA	1500 μ M	NA	NA
Gln	SLC1A5	Km	0.1 mM		Pingitore et al. Biochim. Biophys. Acta 2013, Fig. 9A	NA	100 μ M	NA	NA
Gln	SLC1A5	Km	30 μ M		Fuchs and Bode. Semin. Cancer Biol. 2005	NA	30 μ M	NA	Lower estimate
Gln	SLC1A5	Km	90 μ M		Fuchs and Bode. Semin. Cancer Biol. 2005	NA	90 μ M	NA	Higher estimate
Gln	SLC7A5	Km	1.6 mM		Fuchs and Bode. Semin. Cancer Biol. 2005	NA	1600 μ M	NA	NA
Gln	SLC38A2	Km	2.3 mM		Menchini and Chaudhry. Neuropharmacol. 2019	NA	2300 μ M	NA	NA
Cys2	SLC7A11	Km	0.05 mM		Thomas et al. PLOS ONE 2015	NA	50 μ M	NA	NA
Glu	SLC7A11	Km	7.5 mM		Thomas et al. PLOS ONE 2015	NA	7500 μ M	NA	Export Km for intracellular Glu

Appendix Table S4 - Flux Ranges

Process	Value	Unit	Ref	Assumption made to convert unit	Converted quantity	Converted unit	Note
Cys2 uptake	0.2	nmol / mg protein / min	Thomas et al. PLOS ONE 2015, Fig. 6	Cellular protein concentration 300 mg / mL	60	uM / min	Lower measurement
Cys2 uptake	0.8	nmol / mg protein / min	Thomas et al. PLOS ONE 2015, Fig. 6	Cellular protein concentration 300 mg / mL	240	uM / min	Higher measurement
Cys2 uptake	0.6	nmol / mg protein / min	Ye et al. J. Neurosci. 1999, Fig. 5D	Cellular protein concentration 300 mg / mL	180	uM / min	NA
Cys2 uptake	7	fmol / cell / hr	Hostos et al. Dev. Cell 2016, Fig. 1	Cell volume 3 pL	40	uM / min	NA
Gln uptake	3.5	pmol / ug protein / min	Parker et al. Metab. Eng. 2017, Fig. 2	Cellular protein concentration 300 ug / uL	1000	uM / min	NA
Gln uptake	100	fmol / cell / hr	Hostos et al. Dev. Cell 2016, Fig. 1	Cell volume 3 pL	1700	uM / min	NA
Gln uptake	300	fmol / cell / hr	Jain et al. Science 2012, Database S1	Cell volume 3 pL	5000	uM / min	Max from 60 cell lines
Gln uptake	70	fmol / cell / hr	Jain et al. Science 2012, Database S1	Cell volume 3 pL	1200	uM / min	Median from 60 cell lines
Gln uptake	10	fmol / cell / hr	Jain et al. Science 2012, Database S1	Cell volume 3 pL	170	uM / min	Min from 60 cell lines
Glu export	2	pmol / ug protein / min	Parker et al. Metab. Eng. 2017, Fig. 2	Cellular protein concentration 300 ug / uL	600	uM / min	NA
Glu export	20	fmol / cell / hr	Hostos et al. Dev. Cell 2016, Fig. 1	Cell volume 3 pL	100	uM / min	NA
Glu export	100	fmol / cell / hr	Jain et al. Science 2012, Database S1	Cell volume 3 pL	500	uM / min	Max from 60 cell lines
Glu export	10	fmol / cell / hr	Jain et al. Science 2012, Database S1	Cell volume 3 pL	50	uM / min	Median from 60 cell lines
Glu export	-3	fmol / cell / hr	Jain et al. Science 2012, Database S1	Cell volume 3 pL	-15	uM / min	Min from 60 cell lines;
Glu anaplerosis	1	pmol / ug protein / min	Parker et al. Metab. Eng. 2017, Fig. 2	Cellular protein concentration 300 ug / uL	300	uM / min	Net uptake instead of export
Glu to other metabolites	0.5	pmol / ug protein / min	Parker et al. Metab. Eng. 2017, Fig. 2	Cellular protein concentration 300 ug / uL	150	uM / min	Flux unaccounted for in the diagram
Gln to other metabolites / total labeling	0.1	NA	Hostos et al. Dev. Cell 2016, Fig. 1	Gln uptake flux 1700 uM / min (Hostos et al. Dev. Cell 2016)	170	uM / min	Gln channeled to soluble, polar fraction (metabolites)
Glc uptake	900	fmol / cell / hr	Jain et al. Science 2012, Database S1	Cell volume 3 pL	15000	uM / min	Max from 60 cell lines
Glc uptake	300	fmol / cell / hr	Jain et al. Science 2012, Database S1	Cell volume 3 pL	5000	uM / min	Median from 60 cell lines
Glc uptake	40	fmol / cell / hr	Jain et al. Science 2012, Database S1	Cell volume 3 pL	700	uM / min	Min from 60 cell lines
Glc uptake	300	fmol / cell / hr	Hostos et al. Dev. Cell 2016, Fig. 1	Cell volume 3 pL	5000	uM / min	NA
Glc uptake	40	pmol / ug protein / min	Parker et al. Metab. Eng. 2017, Fig. 2	Cellular protein concentration 300 ug / uL	12000	uM / min	NA
Glc uptake	0.5	umol / uL packed cell / hr	Tanner et al. Cell Syst. 2018, Fig. 1	70 ug / uL packed cells (Fan et al. Nature 2014, Fig. S9); Cellular protein concentration 300 ug / uL	36000	uM / min	Basal rate
Glc uptake	1	umol / uL packed cell / hr	Tanner et al. Cell Syst. 2018, Fig. 1	70 ug / uL packed cells (Fan et al. Nature 2014, Fig. S9); Cellular protein concentration 300 ug / uL	72000	uM / min	Ras-stimulated rate
Glc uptake	1.25	pmol / cell / hr	Liu et al. Nat. Cell Biol. 2020, Fig. S1	Cell volume 3 pL	7000	uM / min	NA
NADPH synthesis by NADK	12	pmol / 10*6 cell / hr	Liu et al. Cell Metab. 2018, Fig. 1 (in text)	Cell volume 3 pL	0.07	uM / min	Very slow compared to regeneration; Assume to be total NADPH/+ to be constant
OXPPP flux	2	nmol / uL packed cell / hr	Fan et al. Nature 2014, Fig. 1	70 ug / uL packed cells (Fan et al. Nature 2014, Fig. S9); Cellular protein concentration 300 ug / uL	150	uM / min	NA
OXPPP flux / glycolysis flux	0.04	NA	Liu et al. Nat. Cell Biol. 2020, Fig. 1d	Glc uptake flux 7000 uM / min (Liu et al. Nat. Cell Biol. 2020)	280	uM / min	NA
OXPPP flux / glycolysis flux	0.02	NA	Bionumbers #112684	NA	NA	NA	NA
NADPH regeneration by OXPPP	2	NADPH / Glc	Generic; 1 from G6PD, 1 from PGD	NA	NA	NA	NA
NADPH regeneration total	10	nmol / uL packed cell / hr	Fan et al. Nature 2014, Fig. 1	70 ug / uL packed cells (Fan et al. Nature 2014, Fig. S9); Cellular protein concentration 300 ug / uL	720	uM / min	NA
Fraction NADPH consumed by anabolism	0.9	NA	Fan et al. Nature 2014, Fig. 4	NA	NA	NA	Upper bound
Fraction NADPH consumed by anabolism	0.7	NA	Fan et al. Nature 2014, Fig. 4	NA	NA	NA	Lower bound

Appendix Table S5 - Fluxes Used To Estimate Parameters

Process	Flux value used	Flux unit	Model	Note	Interpretation	Parameter estimated	Parameter unit
NADPH regeneration by Glc metabolism	500	uM/min	$k_{\text{NadphRedGlc}} \cdot \text{glc} / ((k_{\text{glcIn}} + \text{glc}) \cdot (\text{nadplot} - \text{nadph}) \cdot (1 + \text{switchNadphRed}))$	Assume total flux = 1000 uM/min; Assume switchNadphRed = 0 for estimation process	Not added to Glc for NADPH	$k_{\text{NadphRedGlc}}$	/min
NADPH regeneration by Glu metabolism	500	uM/min	$k_{\text{NadphRedGlu}} \cdot \text{glu} \cdot (\text{nadplot} - \text{nadph}) \cdot (1 - \text{switchNadphRed})$	Assume total flux = 1000 uM/min; Assume switchNadphRed = 0 for estimation process	Not added to Glc for NADPH	$k_{\text{NadphRedGlu}}$	/min
Cys2 uptake	30	uM/min	$k_{\text{Cys2Im}} \cdot \text{cys2o} / (k_{\text{Cys2In}} + \text{cys2o}) \cdot \text{xt} \cdot \text{glu}$	Assume $\text{xt} = 0.1$ uM; Assume Cys2 uptake flux = 30 uM/min; Both lower estimates; Assume 1:1 exchange; Same as Glu export flux	Not added to Cys2 for Cys	k_{Cys2Im} ; k_{GluEx}	/uM/min
Glu export	30	uM/min	$k_{\text{GluEx}} \cdot \text{cys2o} / (k_{\text{Cys2In}} + \text{cys2o}) \cdot \text{xt} \cdot \text{glu}$	Assume $\text{xt} = 0.1$ uM; Assume Glu export flux = 30 uM/min; Both lower estimates; Assume 1:1 exchange; Same as Glu export flux	Not added to Cys2 for Cys	k_{GluEx} ; k_{Cys2Im}	/uM/min
Cys2 reduction by NADPH	30	uM/min	$k_{\text{Cys2Red}} \cdot \text{cys2} \cdot \text{nadph}$	Assume all uptaken Cys2 gets reduced to Cys; Assume NADPH provides all reducing power needed; Same as NADPH consumed by Cys2		k_{Cys2Red} ; $k_{\text{NadphOxCys2}}$	/uM/min
NADPH consumed by Cys2	30	uM/min	$k_{\text{NadphOxCys2}} \cdot \text{cys2} \cdot \text{nadph}$	Assume all uptaken Cys2 gets reduced to Cys; Assume NADPH provides all reducing power needed;			
NADPH consumed by anabolism	800	uM/min	$k_{\text{NadphOxAnab}} \cdot \text{nadph}$	Same as Cys2 reduction by NADPH Assume anabolism consumes 80% NADPH		$k_{\text{NadphOxCys2}}$; k_{Cys2Red}	/uM/min
Gln uptake and Glu production	300	uM/min	$k_{\text{GluPro}} \cdot \text{glu} / (k_{\text{glnIn}} + \text{glu})$	Assume all Gln uptaken hydrolyzed to Glu; Assume Gln uptake = 300 uM/min; A lower estimate	Not added to Gln	k_{GluPro}	uM/min
GSH synthesis	30	uM/min	$k_{\text{GshPro}} \cdot \text{cys} \cdot \text{glu}$	Assume 10% of Glu produced goes to GSH; An upper bound estimate		k_{GshPro}	/uM/min

Appendix Table S6 - Variable Steady States to Estimate Parameters

Molecule	State	Value	Unit	Note
ROS	Full nutrient	0.01	uM	
GSH	Full nutrient	1000	uM	
GSSG	Full nutrient	100	uM	
NADPH	Full nutrient	1	uM	
Cys2	Full nutrient	1	uM	Lower estimate;Consistent with lower SLC7A11 used
Cys	Full nutrient	100	uM	Lower estimate;Consistent with lower SLC7A11 used
Glu	Full nutrient	10000	uM	
Ca2+	Full nutrient	0.1	uM	
RTK-phospho / RTK-total	Full nutrient	0.1	NA	Ref. Graham et al. Mol. Syst. Biol. 2012
RTK-phospho / RTK-total	Glc deprivation	1	NA	Ref. Graham et al. Mol. Syst. Biol. 2012
PPTase-reduced / PPTase-total	Full nutrient	1	NA	Ref. Graham et al. Mol. Syst. Biol. 2012
PPTase-reduced / PPTase-total	Glc deprivation	0.5	NA	Ref. Graham et al. Mol. Syst. Biol. 2012

Appendix Table S7 - Predetermined or Arbitrary Parameters

Parameter	Type	Value	Range	Unit	Note
xct	Constant	0.1	0 to 10	uM	Model low SLC7A11 cell
n1	Hill exponent	5	1 to n	NA	Ultrasensitivity in auto-inhibition
fRosGlc	Artificial handle	0.1	0 to 1	NA	Arbitrary
fRosGlu	Artificial handle	0.4	0 to 1	NA	Arbitrary
switchNadphRed	Artificial handle	0	-1 to 1	NA	Model non-addicted cell
foldRos	Artificial handle	1	0 to n	NA	Base line

Appendix Table S8 - Estimate Parameter Values	Flux value used	Flux unit	Model	Known parameters and estimated steady state Parameter estimated	Estimated value	Parameter unit
Estimation step	Process					
1	NADPH regeneration by Glc metabolism	500	$\mu\text{M}/\text{min}$	$k\text{NadphRedGlc} \cdot \text{glc} / (\text{KglcIn} + \text{glc}) \cdot (\text{nadpIn} - \text{nadph}) \cdot (1 + \text{switchNadphRed})$	$k\text{NadphRedGlc}$	5750 / $\mu\text{M}/\text{min}$
				$\text{glc} = 10000 \text{ }\mu\text{M};$ $\text{KglcIn} = 1500 \text{ }\mu\text{M};$ $\text{nadpIn} = 1 \text{ }\mu\text{M};$ $\text{nadp} = 0.1 \text{ }\mu\text{M};$ $\text{switchNadphRed} = 0;$		
2	NADPH regeneration by Glu metabolism	500	$\mu\text{M}/\text{min}$	$k\text{NadphRedGlu} \cdot \text{glu} \cdot (\text{nadpIn} - \text{nadph}) \cdot (1 - \text{switchNadphRed})$	$k\text{NadphRedGlu}$	0.5 / $\mu\text{M}/\text{min}$
3	NADPH consumed by anabolism	800	$\mu\text{M}/\text{min}$	$k\text{NadphOxAnab} \cdot \text{nadph}$	$k\text{NadphOxAnab}$	800 / $\mu\text{M}/\text{min}$
				$\text{glu} = 10000 \text{ }\mu\text{M};$ $\text{nadpIn} = 1 \text{ }\mu\text{M};$ $\text{nadph} = 0.9 \text{ }\mu\text{M};$ $\text{switchNadphRed} = 0;$ $\text{nadp} = 1 \text{ }\mu\text{M};$		
4	Cys2 uptake	30	$\mu\text{M}/\text{min}$	$k\text{Cys2In} \cdot \text{cys2o} / (\text{Kcys2In} + \text{cys2o}) \cdot \text{xt} \cdot \text{glu}$	$k\text{Cys2In}; k\text{GluEx}$	0.0375 / $\mu\text{M}/\text{min}$
				$\text{cys2o} = 0.2 \text{ }\mu\text{M};$ $\text{Kcys2In} = 0.05 \text{ }\mu\text{M};$ $\text{xt} = 0.1 \text{ }\mu\text{M};$ $\text{glu} = 10000 \text{ }\mu\text{M};$		
4	Glu export	30	$\mu\text{M}/\text{min}$	$k\text{GluEx} \cdot \text{cys2o} / (\text{Kcys2In} + \text{cys2o}) \cdot \text{xt} \cdot \text{glu}$	$k\text{GluEx}; k\text{Cys2In}$	0.0375 / $\mu\text{M}/\text{min}$
				$\text{cys2o} = 0.2 \text{ }\mu\text{M};$ $\text{Kcys2In} = 0.05 \text{ }\mu\text{M};$ $\text{xt} = 0.1 \text{ }\mu\text{M};$ $\text{glu} = 10000 \text{ }\mu\text{M};$		
5	Cys2 reduction by NADPH	30	$\mu\text{M}/\text{min}$	$k\text{Cys2Red} \cdot \text{cys2} \cdot \text{nadph}$	$k\text{Cys2Red}; k\text{NadphOxCys2}$	30 / $\mu\text{M}/\text{min}$
				$\text{cys2} = 1 \text{ }\mu\text{M};$ $\text{nadph} = 1 \text{ }\mu\text{M};$		
5	NADPH consumed by Cys2	30	$\mu\text{M}/\text{min}$	$k\text{NadphOxCys2} \cdot \text{cys2} \cdot \text{nadph}$	$k\text{NadphOxCys2}; k\text{Cys2Red}$	30 / $\mu\text{M}/\text{min}$
				$\text{cys2} = 1 \text{ }\mu\text{M};$ $\text{nadph} = 1 \text{ }\mu\text{M};$		
5	Cys production from Cys2 and NADPH	60	$\mu\text{M}/\text{min}$	$2 \cdot k\text{NadphOxCys2} \cdot \text{cys2} \cdot \text{nadph}$	$k\text{NadphOxCys2}; k\text{Cys2Red}$	30 / $\mu\text{M}/\text{min}$
				$\text{cys2} = 1 \text{ }\mu\text{M};$ $\text{nadph} = 1 \text{ }\mu\text{M};$		
6	NADPH consumed by GSSG	170	$\mu\text{M}/\text{min}$	$k\text{NadphOxGssg} \cdot \text{gssg} \cdot \text{nadph}$	$k\text{NadphOxGssg}$	1.7 / $\mu\text{M}/\text{min}$
				$\text{gssg} = 100 \text{ }\mu\text{M};$ $\text{nadph} = 1 \text{ }\mu\text{M};$		
6	GSSG reduced by NADPH	170	$\mu\text{M}/\text{min}$	$k\text{GshRed} \cdot \text{nadph} \cdot \text{gssg}$	$k\text{GshRed}$	1.7 / $\mu\text{M}/\text{min}$
				$\text{gssg} = 100 \text{ }\mu\text{M};$ $\text{nadph} = 1 \text{ }\mu\text{M};$		
6	GSH regenerated by NADPH	340	$\mu\text{M}/\text{min}$	$2 \cdot k\text{GshRed} \cdot \text{nadph} \cdot \text{gssg}$	$k\text{GshRed}$	1.7 / $\mu\text{M}/\text{min}$
				$\text{gssg} = 100 \text{ }\mu\text{M};$ $\text{nadph} = 1 \text{ }\mu\text{M};$		
7	Gln uptake and Glu production	300	$\mu\text{M}/\text{min}$	$k\text{GluPro} \cdot \text{gln} / (\text{KglIn} + \text{gln})$	$k\text{GluPro}$	315 / $\mu\text{M}/\text{min}$
				$\text{gln} = 2000 \text{ }\mu\text{M};$ $\text{KglIn} = 100 \text{ }\mu\text{M};$		
8	GSH synthesis	30	$\mu\text{M}/\text{min}$	$k\text{GshPro} \cdot \text{cys} \cdot \text{glu}$	$k\text{GshPro}$	0.00005 / $\mu\text{M}/\text{min}$
				$\text{glu} = 10000 \text{ }\mu\text{M};$ $\text{cys} = 100 \text{ }\mu\text{M};$		
9	Cys degradation through other processes	30	$\mu\text{M}/\text{min}$	$k\text{CysDeg} \cdot \text{cys}$	$k\text{CysDeg}$	0.3 / min
				$\text{cys} = 100 \text{ }\mu\text{M};$		
10	Glu degradation through other processes	240	$\mu\text{M}/\text{min}$	$k\text{GluDeg} \cdot \text{glu}$	$k\text{GluDeg}$	0.024 / min
				$\text{glu} = 10000 \text{ }\mu\text{M};$		
11	GSH and GSSG degradation	30	$\mu\text{M}/\text{min}$	$k\text{GshDeg} \cdot \text{gsh} + k\text{GssgDeg} \cdot \text{gssg}$	$k\text{GshDeg}; k\text{GssgDeg}$	0.027 / min
				$\text{ros} = 0.01 \text{ }\mu\text{M};$ $\text{gsh} = 1000 \text{ }\mu\text{M};$ $\text{gssg} = 100 \text{ }\mu\text{M};$		
12	GSH oxidation by ROS	340	$\mu\text{M}/\text{min}$	$2 \cdot k\text{GshOx} \cdot \text{ros} \cdot \text{gsh}^2$	$k\text{GshOx}; k\text{RosDeg}$	0.017 / $\mu\text{M}/\text{min}$
				$\text{ros} = 0.01 \text{ }\mu\text{M};$ $\text{gsh} = 1000 \text{ }\mu\text{M};$		
12	GSSG production from GSH oxidation	170	$\mu\text{M}/\text{min}$	$k\text{GshOx} \cdot \text{ros} \cdot \text{gsh}^2$	$k\text{GshOx}; k\text{RosDeg}$	0.017 / $\mu\text{M}/\text{min}$
				$\text{ros} = 0.01 \text{ }\mu\text{M};$ $\text{gsh} = 1000 \text{ }\mu\text{M};$		
12	ROS removed by GSH	170	$\mu\text{M}/\text{min}$	$k\text{RosDeg} \cdot \text{ros} \cdot \text{gsh}^2$	$k\text{RosDeg}$	0.017 / $\mu\text{M}/\text{min}$
				$\text{ros} = 0.01 \text{ }\mu\text{M};$ $\text{gsh} = 1000 \text{ }\mu\text{M};$		
13	ROS produced by lipid metabolism	170	$\mu\text{M}/\text{min}$	$\text{foldRos} \cdot k\text{RosProBase} \cdot (1 - \text{fRosGlc} - \text{fRosGlu}) \cdot \text{ca} \cdot \text{rtkPhos} \cdot k\text{RosInhRos}^{\text{n1}} / (k\text{RosInhRos}^{\text{n1}} + \text{ros}^{\text{n1}})$	$k\text{RosProBase}$	170000 / $\mu\text{M}/\text{min}$
				$\text{foldRos} = 1;$ $\text{fRosGlc} = 0;$ $\text{fRosGlu} = 0;$ $\text{glc} = 10000 \text{ }\mu\text{M};$ $\text{KglcIn} = 1500 \text{ }\mu\text{M};$ $\text{ca} = 0.1 \text{ }\mu\text{M};$ $\text{rtkPhos} = 0.01 \text{ }\mu\text{M};$ $k\text{RosInhRos} = 0.5;$ $\text{ros}^{\text{n1}} = 5;$		
14	ROS produced by Glc metabolism	170	$\mu\text{M}/\text{min}$	$\text{foldRos} \cdot k\text{RosProGlc} \cdot \text{glc} / (\text{KglcIn} + \text{glc}) \cdot \text{fRosGlc} \cdot \text{ca} \cdot \text{rtkPhos} \cdot k\text{RosInhRos}^{\text{n1}} / (k\text{RosInhRos}^{\text{n1}} + \text{ros}^{\text{n1}})$	$k\text{RosProGlc}$	195500 / $\mu\text{M}/\text{min}$
				$\text{foldRos} = 1;$ $\text{fRosGlc} = 1;$ $\text{fRosGlu} = 0;$ $\text{glc} = 10000 \text{ }\mu\text{M};$ $\text{KglcIn} = 1500 \text{ }\mu\text{M};$ $\text{ca} = 0.1 \text{ }\mu\text{M};$ $\text{rtkPhos} = 0.01 \text{ }\mu\text{M};$ $k\text{RosInhRos} = 0.5;$ $\text{ros}^{\text{n1}} = 5;$		
15	ROS produced by Glu metabolism	170	$\mu\text{M}/\text{min}$	$\text{foldRos} \cdot k\text{RosProGlu} \cdot \text{glu} \cdot \text{fRosGlu} \cdot \text{ca} \cdot \text{rtkPhos} \cdot k\text{RosInhRos}^{\text{n1}} / (k\text{RosInhRos}^{\text{n1}} + \text{ros}^{\text{n1}})$	$k\text{RosProGlu}$	17 / $\mu\text{M}^2 / \text{min}$
				$\text{foldRos} = 1;$ $\text{fRosGlc} = 0;$ $\text{fRosGlu} = 1;$ $\text{glu} = 10000 \text{ }\mu\text{M};$ $\text{ca} = 0.1 \text{ }\mu\text{M};$ $\text{rtkPhos} = 0.01 \text{ }\mu\text{M};$ $k\text{RosInhRos} = 0.5;$ $\text{ros}^{\text{n1}} = 5;$		

Appendix Table S9 - Parameter Summary

Parameter	Type	Process	Interpretation	Note	Value
kRosDeg	Rate constant	ROS removal; GSH oxidation	ROS removal by GSH	Same as kGshOx	0.017
kGshOx	Rate constant	ROS removal; GSH oxidation	GSH consumed by ROS	Same as kRosDeg	0.017
kGshRed	Rate constant	GSH regeneration; NADPH oxidation	GSH regenerated by NADPH	Same as kNadphOxGssg	1.7
kNadphOxAnab	Rate constant	Anabolism; NADPH oxidation	NADPH consumed in anabolism		800
kNadphOxGssg	Rate constant	GSH regeneration; NADPH oxidation	NADPH consumed by GSSG	Same as kGshRed	1.7
kNadphOxCys2	Rate constant	Cys production; NADPH oxidation	NADPH consumed by Cys2	Same as kCys2Red	30
kCys2Red	Rate constant	Cys production; NADPH oxidation	Cys2 reduced by NADPH	Same as kNadphOxCys2	30
kPptaseRedGsh	Rate constant	RTK signaling	PPTase reduced by GSH		0.1
kPptaseOxRos	Rate constant	RTK signaling	PPTase oxidized by ROS		10
kRtkPhos	Rate constant	RTK signaling	RTK got phosphorylated		0.1
kRtkDephos	Rate constant	RTK signaling	RTK dephosphorylated by PPTase		10
kRosProBase	Rate constant	ROS production	Basal ROS production; ROS production by lipid metabolism; ROS production not (instantly) responsive to Glc or Glu		170000
kRosProGlc	Rate constant	ROS production	ROS production by Glc metabolism; ROS production responsive to Glc		195500
kRosProGlu	Rate constant	ROS production	ROS production by Glu metabolism; ROS production responsive to Glu		17
kNadpRedGlc	Rate constant	NADPH regeneration	NADPH regenerated by Glc metabolism; NADPH regenerated by OXPPP		5750
kNadpRedGlu	Rate constant	NADPH regeneration	NADPH regenerated by Glu metabolism; NADPH regenerated by IDH1 and ME1		0.5
kGluPro	Rate constant	Gln uptake; Glu production	Gln uptake; Glu production from Gln		315
kGluDeg	Rate constant	Glu metabolism	Glu consumed in other processes		0.024
kGluEx	Rate constant	Glu export; Cys2 uptake	Glu exported in exchange of Cys2	Same as kCys2Im	0.0375
kGshPro	Rate constant	GSH synthesis	GSH de novo synthesis		0.00005
kGshDeg	Rate constant	GSH degradation	GSH exported; GSH export		0.027
kGssgDeg	Rate constant	GSSG degradation	GSSG degradation; GSSG export		0.027
kCysDeg	Rate constant	Cys metabolism	Cys consumed in other processes		0.3
kCalmBase	Rate constant	Ca2+ influx	Basal Ca2+ influx		0.1
kCalmRos	Rate constant	Ca2+ influx	ROS-induced Ca2+ release		1
kCaOut	Rate constant	Ca2+ efflux	Ca2+ efflux		1
kCys2Im	Rate constant	Glu export; Cys2 uptake	Cys2 uptake at the expense of Glu	Same as kGluEx	0.0375
xc	Constant	Glu export; Cys2 uptake	SLC7A11 abundance; Mass conservation		3
nadpTot	Mass conservation	NADPH regeneration	Total NADPH+; Mass conservation		1
pptaseTot	Mass conservation	RTK signaling	Total PPTase; Mass conservation		0.1
rtkTot	Mass conservation	RTK signaling	Total RTK; Mass conservation		0.1
KrosInhRos	Km	ROS production	ROS auto-inhibition; Redox inhibition of OXPHOS / ETC enzymes; Oxidative damages to ETC / mitochondria		0.5
n1	Hill exponent	ROS production	Ultrasensitivity in ROS auto-inhibition		5
glc	Constant	ROS production; NADPH regeneration	Glc abundance in media		10000
gln	Constant	ROS production; Glu production	Gln abundance in media		2000
cys2o	Constant	NADPH oxidation	Cys2 abundance in media		200
KglnIn	Km	Gln uptake	Gln concentration of half max uptake rate		100
Kcys2In	Km	Cys2 uptake	Cys2 concentration of half max uptake rate		50
KglcIn	Km	Glc uptake	Glc concentration of half max uptake rate		1500
fRosGlc	Artificial handle	ROS production	Fraction ROS production dependent on Glc metabolism		0.1
fRosGlu	Artificial handle	ROS production	Fraction ROS production dependent on Glu metabolism		0.4
switchNadphRed	Artificial handle	NADPH regeneration	Fraction NADPH regeneration dependent on Glc metabolism		0.7
foldRos	Artificial handle	ROS production	Strength of ROS production		1