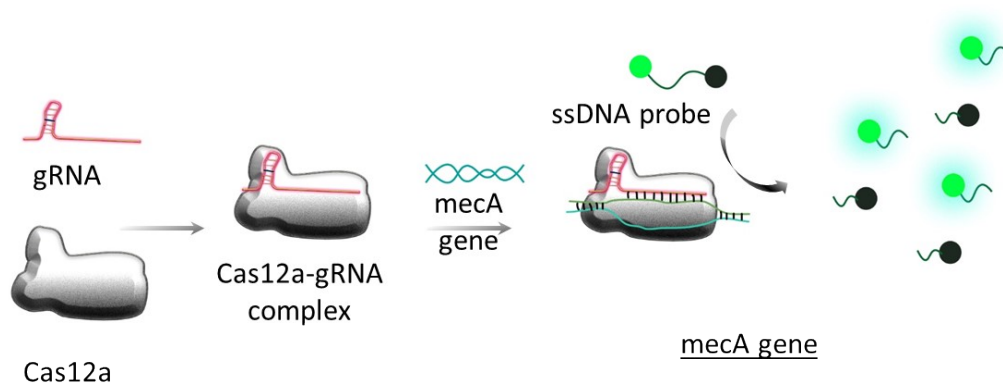


Electronic Supplementary Information

An Amplification-Free Ultra-Sensitive Electrochemical CRISPR/Cas Biosensor for Drug-Resistant Bacteria Detection

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Scheme S1 Schematic of the fluorometric assay for the CRISPR/Cas biosensor.

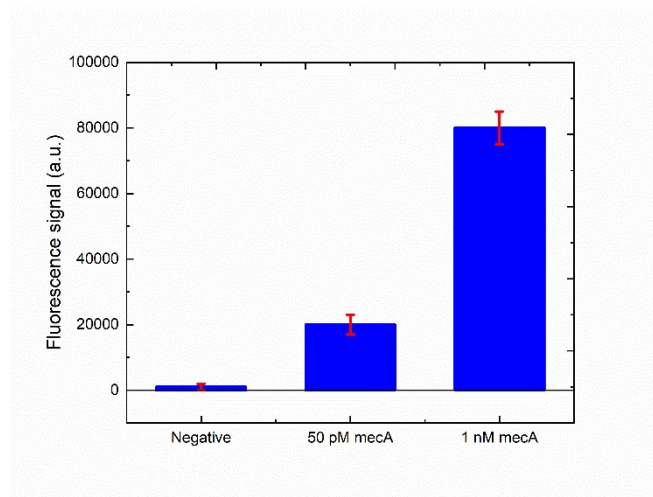


Figure S1. Fluorometric signal of 1 nM and 50 pM *mecA* compared to the negative control (i.e. without the *mecA* gene target).

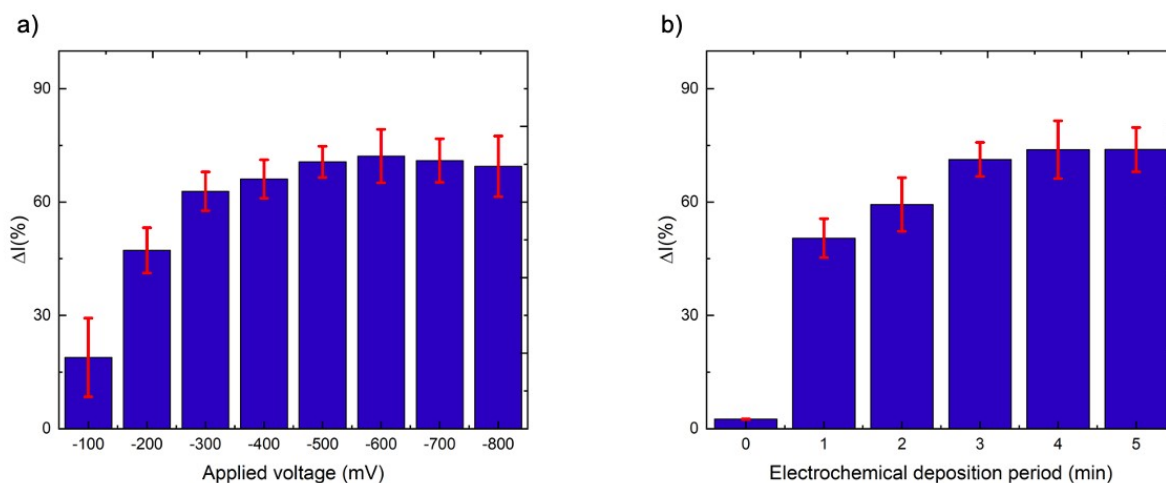


Figure S2 Electrochemical reduction optimization for E-Si-CRISPR as function of (a) applied voltage (-100 to -800 mV), and (b) electrodeposition time (0 to 5 minutes).

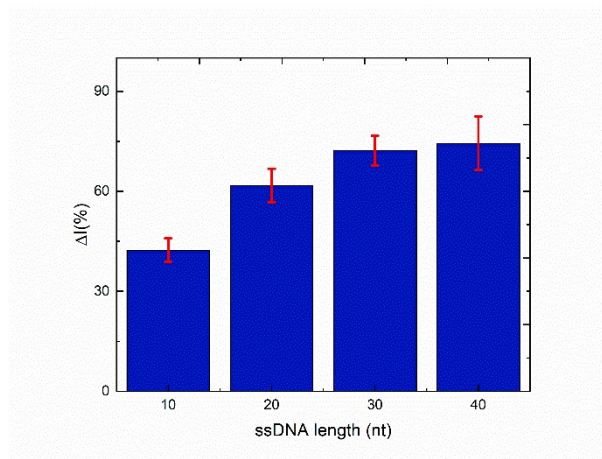


Figure S3 Thiolated ssDNA length optimization on the electrode surface for E-Si-CRISPR, studied from 10 to 40 nt.

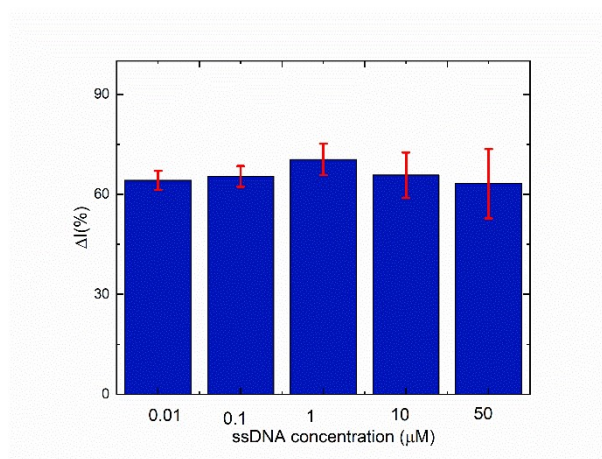


Figure S4 Thiolated ssDNA concentration optimization on electrode surface for E-Si-CRISPR, studied from 0.01 to 50 μM. The ssDNA was modified and the surface passivated by MCH later.

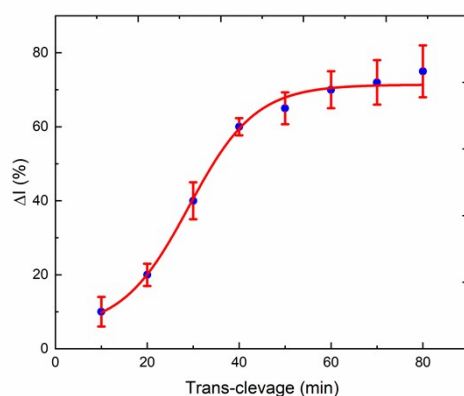


Figure S5 Trans-cleavage period optimization of the lysed MRSA for E-Si-CRISPR, studied from 10 to 80 minutes.

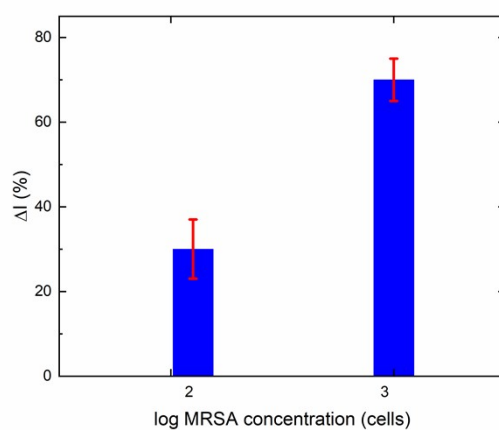


Figure S6 Electrochemical signal of spiked MRSA cells in human serum with concentration of ca. 10^2 and 10^3 cells per detection using optimized condition as used in Figure 5b.

Table S1. PCR primers for *mecA* amplification, and the oligonucleotides for E-Si-CRISPR

Name	Oligonucleotide Sequence
LF	5' AGATTGGGATCATAGCGTCAT 3'
LB	5' TTGAGGGTGGATAGCAGTACC 3'
up <i>mecA</i>	5'-TCT TCA TGT TGG AGC TTT TTA TCG <u>TAA A</u> -3'
down <i>mecA</i>	5'- <u>TTT ACG</u> ATA AAA AGC TCC AAC ATG AAG A-3'
mmPAM	5' TCTTCATGTTGGAGCTTTTTATCG <u>TATA</u> -3'
mmT1	5' TCTTCATGTTGGAGCTTTTTATCT <u>TAAA</u> -3'
mmA5	5' TCTTCATGTTGGAGCTTTTAATCG <u>TAAA</u> -3'
mmG10	5' TCTTCATGTTGGAGTTTTATCG <u>TAAA</u> -3'
10nt	5' AAAAAAAAAA-3'
20nt	5' AAAAAAAAAATTAAAAAAAAA-3'
30nt	5' AAAAAAAAAAAAAAAAAATTTAAAAAAAAAAAAAAAA-3'
40nt	5' AAAAAAAAAAAAAAAAAAAAAAAAAATTAAAAAAAAAAAAAAAAA-3'
FAM Probe	FAM-TTAATT-BHQ1
gRNA	5'-uaa uuuu cau cua agu gua gaucga uaa aaa gcu cca aca ug-3'

Table S2. A comparison of reported electrochemical techniques for CRISPR/Cas biosensors.

Year	Enzyme	Signal Amplification	Target	LoD	Electrochemical method	Assay Time (h)	Ref
2019	Cas12a	-	HPV-16	50 pM	SWV	2	1
2019	Cas13a	HRP/AntiFAM	miRNA-19b	10 pM	CA	4	2
2021	Cas13a	Catalytic hairpin assembly	miRNA-21	2.6 fM	DPV	2	3
2020	Cas12a	Catalytic hairpin assembly	HPV-16 and -18	30 pM	DPV	1.5	4
2021	Cas12a	Rolling circle amplification	ATP	0.46 pM	SWV	5	5
2021	Cas13a	Catalytic hairpin DNA circuit	NSCLC-related RNAs	0.5 fM	SWV	1	6
2021	Cas12a	Recombinase-assisted amplification	Listeria monocytogenes	0.68 aM	SWV	1.5	7
2021	Cas12a	-	MRSA	3.5 fM	SWV	1.5	This work

SWV: square wave voltammetry, *CA*: chronoamperometry, *DPV*: differential pulse voltammetry

References

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3. <https://doi.org/10.1016/j.talanta.2020.121878>
4. <https://doi.org/10.1021/acssensors.9b02461>
5. <https://doi.org/10.1021/acs.analchem.1c00805>
6. <https://doi.org/10.1016/j.bios.2021.113027>
7. <https://doi.org/10.1016/j.bios.2021.113073>