

Supplementary Materials for

Cosmc deficiency causes spontaneous autoimmunity by breaking B cell tolerance

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Figs. S1 to S6

Supplementary Figure Legends

Fig. S1 BC-CosmcKO mice demonstrated autoimmune disease-like pathological features.

(A) Representative images of ocular manifestations of female (11-20 months old) and male mice (13-18 months old) from both BC-*Cosmc*KO and WT littermate controls shown. (**B**) Examples of ocular histology of male BC-*Cosmc*KO and WT mice by hematoxylin and eosin (H&E) staining. Scale bar represents 500 μ m. (**C**) Summary of abnormal ocular manifestations in both male and female mice. (**D**) Wet weight of spleens of young female (left) at 2-4 months old, and aged male (right) at 13-18 months old. Each symbol (black square and open circle for WT and BC-*Cosmc*KO mice, respectively) represents an individual mouse, graphed as mean \pm 1 standard error of the mean (SEM). Unpaired two-tailed Student's *t*-tests were performed to determine statistical significance with *** = p<0.001, ns = significant. (**E**) Steatosis of male mice from both BC-*Cosmc*KO and WT littermate controls at 13 to 18 months old. Liver sections were stained with H&E. Scale bar represents 100 μ m. Representative images from both WT and BC-*Cosmc*KO mice were shown. Photo Credit: Junwei Zeng, BIDMC.

Fig. S2 Disrupted B cell tolerance in BC-CosmcKO mice.

(A) Autoreactive antibodies of BC-CosmcKO mice at 2 months old. Serum samples (1:90 dilution) from both BC-CosmcKO and WT littermate controls were assayed by HEp-2 cell staining. Bound mouse IgM (red) detected by goat anti-mouse IgM conjugated with Alexa fluor-568, and bound IgG (green) detected by goat anti-mouse IgG conjugated with Alexa fluor-488. The WT-1 shown is the only one (1 out of 17) demonstrating minimal HEp-2 reactivity. Images were acquired at 63x magnification. Scale bar represents 20 µm. (B) Summary of assayed samples from BC-CosmcKO mice. (C) Immunoglobulin deposition in kidneys of female BC-CosmcKO mice. Kidney images acquired at 20x magnification. Representative images from both WT (n=5) and BC-CosmcKO male mice (n=6) shown. Scale bar represents 50 μ m. (**D**, E) Urine was collected from aged BC-CosmcKO and WT mice of both genders for analysis by Chemstrips (D), and Bicinchoninic Acid (BCA) protein assay (E). WT (n=7) and BC-CosmcKO mice (n=9) for male, WT (n=7) and BC-CosmcKO mice (n=7) for female in **D**. WT (n=4) and BC-CosmcKO mice (n=8) for male, WT (n=6) and BC-CosmcKO mice (n=7) for female in E. The female urinary protein data (by BCA) are representative of two independent experiments with similar results, at least 6 animals in each group. (F, G) Histopathology scores of kidneys of both aged mice: WT (n=6) and BC-CosmcKO mice (n=9) for male in F, and WT (n=8) and BC-CosmcKO mice (n=12) for female in G. Representative images of H&E staining of kidneys of aged female shown and boxed area shown at x40 magnification in G (right). Scale

bars represent 100 μ m (left) and 20 μ m (right) respectively. The glomerular index is the score of the category (a), detailed in the Methods. (H) Blue native agarose polyacrylamide gel electrophoresis (BN-APAGE) analysis for IgM and IgG of both aged WT and BC-*Cosmc*KO mice sera. Four and three biological replicates of female and male respectively are shown. For **D-G**, each symbol (black square and open circle for WT and BC-*Cosmc*KO mice, respectively) represents an individual mouse, graphed as mean ± 1 standard error of the mean (SEM). Serum or tissues were collected from age-matched aged (11-20 months old) and young (2-4 months old) mice of indicated gender. Unpaired two-tailed Students *t*-tests were performed to determine statistical significance with ns stands for non-significant, * = p<0.05, ** = p<0.01.

Fig. S3 Immune effectors in BC-CosmcKO mice.

Blood samples were collected from indicated groups for A-D. Determination of TNF α (A) and IFN γ (B) levels in sera were conducted by ELISA, according to manufacturer's instructions. Dashed lines represent the minimal detection dose, 1.88 pg/ml for TNF α , and 2 pg/ml for IFN γ . Detection of full length C3 complement and its breakdown fragments of young (C) and aged (D) mice was analyzed by Western blot. Asterisks in D indicates presence of aberrant complement C3 and breakdown subunits in the aged female BC-CosmcKO mice. Each symbol (black square and open circle for WT and BC-*Cosmc*KO mice, respectively) represents an individual mouse, graphed as mean \pm 1 standard error of the mean (SEM). Sera were collected from age-matched aged (11-20 months old) and young (2-4 months old) mice of indicated gender. ns = not significant. Unpaired two-tailed Student's *t*-tests were performed to determine statistical significance with * = p<0.05, ** = p<0.01.

Fig. S4 Enhanced basal activation of Cosmc-deficient B cells.

(A) Expression levels of CD9 and CD1d on follicular B cells (B220⁺ or CD19⁺ CD23⁺ CD21¹⁰ FOB, upper row) and marginal zone B cells (B220⁺ or CD19⁺ CD23¹⁰ CD21^{hi} MZB, bottom row) by flow cytometry. Expression levels of CD86, CD80, and MHC II on (**B**) FOB cells and (**C**) MZ B cells. Splenocytes are prepared from both BC-*Cosmc*KO and WT littermate controls. The results are representative of three independent experiments with at least 3 animals in each group. Each symbol (black square and open circle for WT and BC-*Cosmc*KO mice, respectively) represents an individual mouse, graphed as mean ± 1 standard error of the mean (SEM). Cells were prepared from young (2-4 months old) male mice. Unpaired two-tailed Student's *t*-tests were performed to determine statistical significance with * = p<0.05, ** = p<0.01, *** = p<0.001.

Fig. S5 Cosmc deficiency reduces PNA binding on germinal center B cells.

Binding analysis of biotinylated PNA, MAL-II, SNA, and Con A, followed by streptavidin-Alexa fluor 488, on neuraminidase or PBS treated WT and BC-*Cosmc*KO splenic B cell subsets (A): follicular B cells (B220⁺ or CD19⁺CD23⁺CD21^{lo} FOB, top two rows) marginal zone B cells (B220⁺ or CD19⁺CD23^{lo}CD21^{hi} MZB, bottom two rows). (B) Representative histogram plots of PNA binding of splenic germinal center B cells (B220⁺ or CD19⁺GL7⁻ Fas⁺) and non-germinal center B cells (B220⁺ or CD19⁺GL7⁻ Fas⁻). Right, mean fluorescence intensity (MFI) of PNA binding on indicated B cell population is shown for each WT and BC-*Cosmc*KO mice. (C) Representative FACS plots of the germinal center B cell population in MLN simultaneously stained with CD19, GL7, Fas, and CD38, and analyzed by two gating strategies (CD19⁺GL7⁺ Fas⁺ left, and CD19⁺CD38^{lo} Fas⁺ middle). Right, percentage of GC B cells of total B cells shown for WT and BC-*Cosmc*KO mice of indicated tissue. Each symbol (black square and open circle for

WT and BC-*Cosmc*KO, respectively) represents an individual mouse, graphed as mean ± 1 standard error of the mean (SEM). Cells were prepared from young (2-4 months old) male mice in **A** and **B**, and from young male and female mice in **C**. Unpaired two-tailed Student's *t*-tests were performed to determine statistical significance with ** = p<0.01, *** = p<0.001. ns = not significant.

Fig. S6 Antigen specific Immune responses in BC-CosmcKO mice.

NP-specific immunoglobulin responses to immunization with (A) NP-LPS, (B) NP-Ficoll. (A) Both WT (n=9) and BC-*Cosmc*KO mice (n=11) were intraperitoneally immunized with NP-LPS. (B) Both WT (n=9) and BC-*Cosmc*KO mice (n=9) were intraperitoneally immunized with NP-Ficoll. ELISA determined the titers of NP-specific IgM (A and B) of sera collected from immunized mice at indicated time points. Data were from two experiments with similar results. (C) Phospho-flow cytometry analysis of WT and BC-*Cosmc*KO B cells stimulated with anti-IgM antibody for indicated periods of time. Representative histogram plots of phosphorylated SYK and total cellular proteins (pan-tyrosine) shown. Each symbol (black square and open circle for WT and BC-*Cosmc*KO, respectively) represents an individual mouse, graphed as mean ± 1 standard error of the mean (SEM). Young (2-4 months old) male mice were used in A-C. Unpaired two-tailed Student's *t*-tests were performed to determine statistical significance with ** = p<0.01.

Supplementary Figures











