

**Supplementary Table 1.**Primers used for detection and sequence determination of *bla* genes

<i>bla</i> gene	Purpose	Primer name	Nucleotide sequence 5'–3'	Reference
<i>bla</i> <sub>CMY</sub>	Detection and sequencing	cmy-F	GACAGCCTCTTCTCCACA	1
		cmy-R	TGGAACGAAGGCTACGTA	1
	Sequencing	CMY2-outF	GTTACAATGTGTGAGAAGCAGTC	2
		CMY2-outR	ATGGGATTTTCCTTGCTGTA	2
		CMY2-R0	CAGTATTTCGTGACCGGA	2
CMY2-F3	CTGGATTACGGTTCCGCA	2		
<i>bla</i> <sub>CTX-M</sub>	Detection and sequencing	CTX-MU1	ATGTGCAGYACCAGTAARGT	3
		CTX-MU2	TGGGTRAARTARGTSACCAGA	3
	Sequencing	CTXM14 upstream F1	AGCAGTCTAAATTCCTCGTG	This study
		CTXM14 upstream R3	GCTGGCAATCAATTTGTTT	This study
		CTX-outF2	GCCAAGGATAATACTAATAGAGG	2
		CTX-outR	GCGGAATGATAGAAAGAGATGAG	2
		CTX-M-14 F653	ACGTGGCTCAAAGCAATAC	4
		CTX-M-14 R1070	CGTGAAGAAGGTGTGCTGA	4
	Detection	Mab/F	GGGGAGCTCATAAAATCTTGAAGAC	5
		Mab/R	GGGGATCCCTTACCAATGCTTAATCA	5
Sequencing	TEM-1R ant	TGATTGCTGGTTACGGTGA	This study	
	TEM-1R upstream	GGTTAGCTCCTTCGGTCCTC	This study	
	TEM1F downstream	GGATGGAGGCGGATAAAGTT	This study	
	TEM-1F Tn3 rec	ACGAAAGGGCCTCGTGATA	This study	
	TEM-1 seq OutF	ATGGTTTCTTAGACGTCAGG	This study	
	TEM-1 seq OutR	CATAAACAAAGCGTCTTGAAC	This study	
<i>bla</i> <sub>SHV</sub>	Detection	SHV-F	AGGATTGACTGCCTTTTGG	6
		SHV-R	ATTTGCTGATTCGCTCG	6

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**Supplementary Table 2.**Epidemiological information about the *bla* gene-carrying *Salmonella* isolates from this study

Serotype	<i>bla</i> gene	Location of samples collected	Facility type	Facility name <sup>a</sup>	Egg production company's name	Sample origin	Isolation date	Strain name
Ohio <sup>b</sup>	<i>bla</i> <sub>CMY-2</sub>	Prefecture A	Layer farm	a5	Company A	Eggshell	January 2012	<b>SEOhiM1960<sup>c</sup></b>
			Pullet-rearing farm	Uncertain A <sup>d</sup>	Company A	Environment	July 2009	SEOhiM1505
			Pullet-rearing farm	a7	Company A	Environment	November 2009	SEOhiM1579
			Pullet-rearing farm	a1	Company A	Environment	January 2010	<b>SEOhiM1593<sup>c</sup></b>
							May 2010	SEOhiM1615
							October 2010	SEOhiM1668-2
							November 2010	SEOhiM1782
							April 2011	SEOhiM1798
							May 2011	SEOhiM1802
							July 2011	SEOhiM1822
				March 2012	SEOhiM1984			
				June 2012	<b>SEOhiM2008<sup>c</sup></b>			
Untypeable with O7:b:-	<i>bla</i> <sub>CMY-2</sub>	Prefecture A	Pullet-rearing farm	a1	Company A	Environment	February 2012	SEOhiM1977
Braenderup	<i>bla</i> <sub>CMY-2</sub>	Prefecture B	Layer farm	Uncertain B <sup>e</sup>	Company B	Environment	October 2011	SEBraM1898
Cerro	<i>bla</i> <sub>CTX-M-14</sub>	Prefecture C	Layer farm	q1	Company Q	Eggshell	August 2012	<b>SECerM2017<sup>c</sup></b>

<sup>a</sup> Each facility belonged to egg production companies A, B, or Q.<sup>b</sup> *Salmonella enterica* subsp. *enterica* serovar Ohio.<sup>c</sup> Detailed information about the four strains appearing in bold type is shown in Table 3.<sup>d</sup> Uncertain A must either be facility a1, a6, or a7 in egg production company A.<sup>e</sup> Uncertain B must be one of the 12 facilities belonging to egg production company B.

**Supplementary Table 3.**Comparison of the *bla*<sub>CMY-2</sub>-harboring-IncA/C<sub>2</sub> plasmids isolated from *Salmonella enterica* in this study with IncA/C<sub>2</sub> plasmids from Enterobacteriaceae<sup>a</sup>

Plasmid name	Plasmid replicon type	Antimicrobial resistance genes			Number of IS26 elements	Number of <i>ISEcp1</i> elements within the region flanking the <i>bla</i> gene <sup>b</sup>	Size (bp)	Isolation year	Country	Source	Accession number	Bacteria	Reference
		<i>bla</i> <sub>CMY-2</sub>	Common antimicrobial resistance genes	Other antimicrobial resistance genes									
pSEOhiM1593a	IncA/C <sub>2</sub>	<i>bla</i> <sub>CMY-2</sub>	<i>aph(3'')-Ib</i> , <i>aph(6)-Id</i> , <i>floR</i> , <i>sul2</i> , <i>tet(A)</i>	ND <sup>c</sup>	3	1	92,251	2010	Japan	Pullet-rearing farm environment	AP024348	<i>Salmonella enterica</i> serovar Ohio	This study
pSEOhiM1960a	IncA/C <sub>2</sub>	<i>bla</i> <sub>CMY-2</sub>	<i>aph(3'')-Ib</i> , <i>aph(6)-Id</i> , <i>floR</i> , <i>sul2</i> , <i>tet(A)</i>	ND	3	1	92,095	2012	Japan	Egg shells	AP024350	<i>Salmonella enterica</i> serovar Ohio	This study
pCVM21550	IncA/C <sub>2</sub>	<i>bla</i> <sub>CMY-2</sub>	<i>aph(3'')-Ib</i> , <i>aph(6)-Id</i> , <i>floR</i> , <i>sul2</i> , <i>tet(A)</i>	ND	3	1	120,340	Uncertain	USA	Swine	CP009564	<i>Salmonella enterica</i> serovar Newport	[1]
p2016K-0796	IncA/C <sub>2</sub>	<i>bla</i> <sub>CMY-2</sub>	<i>aph(3'')-Ib</i> , <i>aph(6)-Id</i> , <i>floR</i> , <i>sul2</i> , <i>tet(A)</i>	<i>aph(3')-Ia</i> , <i>sul1</i> , <i>tet(B)</i> , <i>tet(O)</i>	5	1	179,850	Uncertain (2015–2017)	USA	Human	MH760469	<i>Salmonella enterica</i> serovar Heidelberg	[6]
pAR060302	IncA/C <sub>2</sub>	<i>bla</i> <sub>CMY-2</sub>	<i>aph(3'')-Ib</i> , <i>aph(6)-Id</i> , <i>floR</i> , <i>sul2</i> , <i>tet(A)</i>	<i>aac(3)-VIa</i> , <i>ant(3'')-Ia</i> , <i>sul1</i>	3	1	166,530	2002	USA	Cow	HQ023864	<i>Escherichia coli</i>	[3]
pYDC637	IncA/C <sub>2</sub>	<i>bla</i> <sub>CMY-2</sub>	<i>aph(3'')-Ib</i> , <i>aph(6)-Id</i> , <i>floR</i> , <i>sul2</i> , <i>tet(A)</i>	<i>aadA5</i> , <i>ant(3'')-Ia</i> , <i>aph(3')-Ia</i> , <i>dfrA17</i> , <i>rmtE</i> , <i>sul1</i>	3	1	199,469	2013	USA	Human	KP056256	<i>Escherichia coli</i>	[4]
pKP-Gr642	IncA/C <sub>2</sub>	Not detected but <i>bla</i> <sub>CMY-4</sub>	<i>aph(3'')-Ib</i> , <i>aph(6)-Id</i> , <i>floR</i> , <i>sul2</i> , <i>tet(A)</i>	<i>aac(6)-II</i> , <i>aadA1</i> , <i>bla</i> <sub>NIM-19</sub> , <i>dfrA1</i>	2	1	162,787	2011	Greece	Human	KR559888	<i>Klebsiella pneumoniae</i>	[5]
p205880-Ct1/2	IncA/C <sub>2</sub>	<i>bla</i> <sub>CMY-2</sub>	<i>aph(3'')-Ib</i> , <i>aph(6)-Id</i> , <i>floR</i> , <i>sul2</i> , <i>tet(A)</i>	<i>aac(6)-II</i> , <i>sul1</i>	3	1	153,373	2012	China	Human	MF344573	<i>Klebsiella pneumoniae</i>	[2]
pSN254b	IncA/C <sub>2</sub>	<i>bla</i> <sub>CMY-2</sub>	<i>aph(3'')-Ib</i> , <i>aph(6)-Id</i> , <i>floR</i> , <i>sul2</i> , <i>tet(A)</i>	<i>aadA7</i> , <i>sul1</i>	2	1	152,216	Uncertain	Canada	Uncertain	KJ909290	<i>Aeromonas salmonicida</i>	[7]

<sup>a</sup> Plasmids with >99% query coverage against pSEOhiM1593a nucleotide sequences were identified by BLASTN searches in this list.<sup>b</sup> Number of *ISEcp1* elements within 5,000 bp of the *bla* gene.<sup>c</sup> ND: no antimicrobial resistance gene was detected.

## References

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**Supplementary Table 4.**  
*Salmonella* serotypes from eggshells and egg production environments

O-group	Serotypes	Number of isolates
O:4 (B)	Agona <sup>a</sup>	9
	Typhimurium	3
O:7 (C <sub>1</sub> )	Braenderup <sup>b</sup>	36
	Infantis	22
	Ohio <sup>b</sup>	22
	Mbandaka	10
	Thompson	8
	Montevideo	6
	Livingston	4
	Singapore	4
	Othmarschen	3
	Bareilly	2
	Tennessee	2
	Untypeable with O7: b: - <sup>b</sup>	1
O:8 (C <sub>2</sub> -C <sub>3</sub> )	Corvallis	15
	Newport	5
	Kentucky	4
	Manhattan	2
O:9 (D <sub>1</sub> )	Enteritidis	5
O:3,10 (E <sub>1</sub> )	Orion	3
O:13 (G)	Untypeable with O13: y: -	2
O:18 (K)	Cerro <sup>c</sup>	20
O:35 (O)	Alachua	21
O:40 (R)	Johannesburg	3
	Others (including 12 serotypes) <sup>d</sup>	12
	Total	224

<sup>a</sup> *Salmonella enterica* subsp. *enterica* serovar Agona.

<sup>b</sup> Including AmpC  $\beta$ -lactamase producers.

<sup>c</sup> Including an extended-spectrum  $\beta$ -lactamase producer.

<sup>d</sup> Including Chincol, Gaminara, Havana, Idikan, Oranienburg, Potsdam, Putten, Rissen, Schwarzengrund, Senftenberg, Virchow, and O16: l, v: -.

**Supplementary Table 5.**

Antimicrobial resistance rates of *Salmonella* isolates<sup>a</sup> originating from Japanese layer breeding chains and egg processing facilities.

Antimicrobial	Number of resistant isolates	Resistance rate <sup>b</sup> (%)
Streptomycin	30	13.4
Tetracycline	26	11.6
Ampicillin	20	8.9
Chloramphenicol	16	7.1
Cefpodoxime	15	6.7
Nalidixic acid	6	2.7
Ciprofloxacin	4	1.8
Kanamycin	1	0.4
Azithromycin	0	0
Meropenem	0	0

<sup>a</sup> Of the 224 *Salmonella* isolates tested, 40 showed resistance to at least one antimicrobial agent.

<sup>b</sup> The resistance rate for each antimicrobial agent was calculated as the percentage of resistant isolates among 224 isolates in this study.

**Supplementary Table 6.**Determination of minimum inhibitory concentration (MIC) for *Salmonella enterica* using Etest<sup>a</sup>

Strains	Serovars	Location of <i>bla</i> genes	<i>bla</i> genes	MIC ( $\mu$ g/ml) of each antimicrobial <sup>b</sup>					
				CTRX	CAZ	CTX	CPDX	ABPC / SBT	ABPC
SEOhM1593	<i>S.</i> Ohio <sup>c</sup>	Plasmid	<i>bla</i> <sub>CMY-2</sub>	96	48	>32	> 256	24	> 256
SEOhM1960	<i>S.</i> Ohio	Chromosome and plasmid	<i>bla</i> <sub>CMY-2</sub>	96	48	>32	> 256	48	> 256
SEOhM2008	<i>S.</i> Ohio	Chromosome	<i>bla</i> <sub>CMY-2</sub>	16	12	12	> 256	16	> 256
SECerM2017	<i>S.</i> Cerro	Chromosome	<i>bla</i> <sub>CTX-M-14</sub>	16	1	12	64	6	> 256

<sup>a</sup> Etest (bioMérieux, Marcy-l'Étoile, France) was used for MIC determination using agar plates.<sup>b</sup> ABPC: ampicillin, ABPC / SBT: ampicillin-sulbactam, CAZ: ceftazidime, CPDX: cefpodoxime, CTRX: ceftriaxone, CTX: cefotaxime.<sup>c</sup> *S. enterica* subsp. *enterica* serovar Ohio.