

Supporting Information

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N2-polarized neutrophils guide BMSC recruitment and initiate bone regeneration: A missing piece of the bone regeneration puzzle

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Figure S1. Intramuscular ectopic implantation of gelatin sponge, gelatin sponge with 1 μ g IL-8, 5 μ g IL-8, 10 μ g IL-8, and 5 μ g LPS. Gelatin sponge with a low IL-8 dose (1 μ g) was insufficient to recruit myeloid cells and induce endochondral ossification, while a high IL-8 dose (10 μ g) led to the rapid degradation of gelatin, indicating predominant catabolism. The gelatin sponge with LPS was completely degraded and pathological fat liquefaction was observed at the implantation site (n = 3 for each group).



Figure S2. Time-course of circulatory neutrophils and monocytes quantified using flow cytometry after treatment with anti-Ly6G or anti-F4/80 neutralizing antibodies. The

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neutralizing antibodies efficiently and specifically depleted neutrophils or monocytes in peripheral blood without affecting other cell type (n = 3 for each group).



Figure S3. Immunohistochemical staining of Col I and Col II in the ectopic tissue after depletion of neutrophils/macrophages (N-/M-). Neutrophil/macrophage depletion significantly hindered chondrogenic and osteogenic differentiation around the implantation site (n = 3 for each group).



Figure S4. Time-course of circulatory myeloid cells in nonlethal irradiated mice quantified using flow cytometry. The number of myeloid cells in peripheral blood decreased by over 90% three days post-irradiation (n = 3 for each group).





Figure S5. Flow cytometry analysis of isolated myeloid cells from unirradiated mice. The positive rates of the purified monocytes, neutrophils, and BMSCs were 85.9%, 97.2%, and 97.7%, respectively, which met the requirements of subsequent experiments.



Figure S6. Wound-healing assay of BMSC migration co-cultured with neutrophils or IL-8-treated neutrophils for 24 h in transwell. IL-8-treated neutrophils significantly enhanced the migration capacity of BMSCs.

	Table	S1.	Primer	sequence	es used	in	RT-I	PCR
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Primer	Sequences
mouse IL-10 Forward	GGTTGCCAAGCCTTATCGGA
mouse IL-10 Reverse	GGGGAGAAATCGATGACAGC
mouse TGFβ1 Forward	CTTCAATACGTCAGACATTCGGG
mouse TGFβ1 Reverse	GTAACGCCAGGAATTGTTGCTA
mouse TNF Forward	GGCAGGTCTACTTTGGAGTC
mouse TNF Reverse	TCGAGGCTCCAGTGAATTCG
mouse IFNγ Forward	AACGCTACACACTGCATCTTG
mouse IFNγ Reverse	GACTTCAAAGAGTCTGAG