| 1 | Supplementary Information |
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| 3 | ClusterMap for multi-scale clustering analysis of spatial gene expression |
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| 5 | Yichun He ^{1,2,8} , Xin Tang ^{1,2,8} , Jiahao Huang ² , Jingyi Ren ^{2,3} , Haowen Zhou ² , Kevin Chen ⁴ , |
| 6 | Albert Liu ^{2,3} , Hailing Shi ^{2,3} , Zuwan Lin ^{4,2} , Qiang Li ¹ , Abhishek Aditham ^{2,5} , Johain |
| 7 | Ounadjela ^{2,6} , Emanuelle I Grody ^{2,6} , Jian Shu ^{2,6,7} , Jia Liu ^{1*} & Xiao Wang ^{2,3*} |
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| 9 | ¹ John A. Paulson School of Engineering and Applied Sciences, Harvard University, Cambridge, |
| 10 | MA, USA. |
| 11 | ² Broad Institute of MIT and Harvard, Cambridge, MA, USA. |
| 12 | ³ Department of Chemistry, Massachusetts Institute of Technology, Cambridge, MA, USA. |
| 13 | ⁴ Department of Chemistry and Chemical Biology, Harvard University, Cambridge, MA, USA. |
| 14 | ⁵ Department of Biological Engineering, Massachusetts Institute of Technology, Cambridge, MA, |
| 15 | USA. |
| 16 | ⁶ Whitehead Institute for Biomedical Research, Cambridge, MA, USA. |
| 17 | ⁷ Cutaneous Biology Research Center, Massachusetts General Hospital, Harvard Medical School, |
| 18 | Boston, MA, USA. |
| 19 | ⁸ These authors contributed equally: Yichun He, Xin Tang. |
| 20 | *e-mail: wangxiao@broadinstitute.org; jia_liu@seas.harvard.edu. |



23 Performance comparison of ClusterMap and predefined distribution-based method in

simulated data. Different colors represent different segmentation results. Note that the gene

- identity of each spot is randomly assigned from 1 to 5 as pseudo gene type. Left: ground truth;
- 26 Middle: ClusterMap results; Right: results using predefined distribution-based method²⁰.





- 29 Supplementary Figure 2
- 30 Comparison of RNA sampling approaches between ClusterMap using absolute physical
- 31 distance with other methods²⁰ using k-nearest neighbours (kNN) in simulated data. Three
- 32 examples of RNAs with various local density demonstrating that ClusterMap preserves the local
- 33 physical density information while kNN does not consider the physical density of RNAs.





Illustration of sub-cellular, cellular, and tissue region analyses. a, Subcellular analysis process 37 of the panel IV in Figure 1d by ClusterMap. A three-channel (magenta: *Malat1*; cyan: *ActB*; blue: 38 DAPI) composite image shows raw fluorescent signals. After preprocessing mRNA molecules 39 with specific genes located, ClusterMap first performs cellular resolution and identifies individual 40 41 cells. Then a mesh graph that models the relationships among mRNA spots in the cell is generated to compute the NGC coordinates and K-means clustering separate spots into two regions using 42 joint physical and NGC coordinates. 100 times K-means clustering was performed with different 43 seeds and showed the consistent same results. Finally, a convex hull is constructed from the 44 nucleus spots, denoting the nucleus boundary. The pattern of ClusterMap-constructed nucleus 45 boundary is compared with the DAPI staining. Scale bar: 20µm. b, Examples of the cell 46 identification in ClusterMap procedures in Fig. 2a. Upper: DAPI staining showing the cell nuclei. 47 Middle: mRNA spots. Lower: Clustering results. c, The accuracy of cell identification results from 48 eight STARmap⁶ datasets compared with corresponding expert-annotated labels. BZ5, BZ9, BZ14, 49 BZ19: four STARmap⁶ 166-gene sets in mouse medial prefrontal cortex (mPFC); BD2, BD6: two 50 STARmap⁶ 160-gene sets in mouse V1. BY1, BY3: two STARmap 1020-gene sets in mouse V1. 51 52 The horizontal line is at 80% accuracy. **d**, ClusterMap constructs the tissue regions after cell-typing. First, the neighborhood cell-type composition (NCC) of each cell is computed by considering a 53 sliding window over the cell-type map. Then both the NCC and physical locations of cells are 54 55 combined for K-means clustering. Cells with highly correlated neighboring cell-type composition and close spatial distances are merged into a single tissue region signature. 56



Identification of cell types in mouse primary cortex (V1) and medial prefrontal cortex 60 (mPFC). a,b, UMAP and heatmap visualization of all excitatory, inhibitory and non-neuronal cell 61 types in STARmap⁶ mouse V1 1020-gene (two replicates: BY1 and BY3). The heatmap represents 62 z-scored expression matrix of all cell types, showing clustering of five most differentially 63 expressed genes per cell type. Genes shown are selected based on a false discovery rate (FDR)-64 adjusted *p*-value threshold of 0.05 (Benjamini-Hochberg correction) and a minimum log₁₀ fold 65 change of 0.4, using a two-sided, unpaired t-test, for genes that are expressed in cells within each 66 cluster versus cells in any other cluster. c, e, UMAP visualization of all excitatory, inhibitory and 67 non-neuronal cell types in STARmap⁶ 160-gene datasets in mouse V1 (two replicates: BD2, BD6, 68 (c)), and STARmap⁶ 166-gene datasets in mPFC (four replicates, BZ5, BZ9, BZ14, BZ19, (e)). d, 69 f, Spatial organization map of cell types in BD2 and BD6 (d), and in BZ5, BZ9, BZ14 and BZ19 70 **(f)**. 71



Cell-type correlation matrices comparison of ClusterMap-based and manually-segmented 74 cell types. a,b, Comparison on STARmap mouse V1 1020-gene datasets. Heatmaps of Pearson 75 correlation (a) and -log(p-value) (b) for null hypothesis testing. The p value is based on a t statistic 76 which has *n*-2 degrees of freedom and 95% confidence interval. The single-cell gene expression 77 profiles from ClusterMap with manual annotation are compared. c,d, Comparison on STARmap 78 mouse V1 160-gene datasets. Heatmaps of correlation (c) and -log(p-value) (d) comparing the 79 single-cell gene expression profiles from ClusterMap with manual annotation. e,f, Comparison on 80 STARmap mouse mPFC 166-gene datasets. Heatmaps of correlation (e) and -log(p-value) (f) 81 comparing the single-cell gene expression profiles from ClusterMap with manual annotation. 82 Horizontal: ClusterMap; vertical: manual annotation. 83



Analyses of the STARmap⁶ mouse placental dataset. a, ClusterMap generates the cell
segmentation map of the STARmap⁶ mouse placenta 903-gene dataset, including 7,224 cells. Scale
bar: 100 µm. b, Statistics of ClusterMap identified placental cells as shown in (a). Left: Histogram
of detected reads (DNA amplicons) per cell. Middle: Histogram of genes per cell. Right:
Correlation plot between genes per cell and reads per cell. c, Heatmap visualization of 12 cell types.
Names are in the right panel of (d). d, UMAP from label transfer results with scRNA-seq,
compared with UMAP of the Louvain clustering²² in ClusterMap.



Sub-clustering within one cell type using cell niche compositions in STARmap mouse 95 placenta 903-gene dataset. a, Schematic indicating how cells in one cell type are sub-clustered 96 based on either gene expression (Louvain clustering²²) or the cell niche compositions (K-means 97 clustering¹⁹). **b**, UMAP of gene expression sub-clustering (top) or cell niche composition sub-98 clustering (bottom) in Maternal Decidua-1 (MD-1). c, Spatial subtype maps using gene expression 99 (top) or cell niche composition (bottom) in MD-1. d, Heatmap of sub-clustering using gene 100 expression (top) or cell niche composition sub-clustering (bottom) in MD-1. Gene markers in the 101 top heatmaps of gene expression sub-clustering are 0: GPNMB, 1: CXCL14. Row names in the 102 bottom heatmaps of cell niche composition sub-clustering are cell types in numbers annotated in 103 Figure 3. 104

108 **ClusterMap analyses across different experimental methods. a**, The cell segmentation map of 109 whole osmFISH mouse somatosensory cortex (SSp) datasets. Scale bar: 100 μm. **b,c**, UMAP and 110 heatmap visualization of 31 cell types in osmFISH datasets. **d**, The 2D cell segmentation map of 111 whole MERFISH mouse preoptic area (POA) datasets. Scale bar: 200 μm. **e,f**, UMAP and heatmap 112 visualization of 9 cell types in MERFISH datasets. The number of cells increased from 6,471 to 113 8,538 for osmFISH, from 2,620 to 2,924 for pciSeq, from 6,977 to 10,320 for MERFISH. The 114 number of reads increased from 1,248,106 to 1,690,328 for osmFISH, from 31,246 to 31,750 for

115 pciSeq, from 1,927,913 to 3,065,171 for MERFISH.

ClusterMap analyses of ISS data. a, Cell segmentation map shows the cell segmentation results by ClusterMap. Colors are randomly assigned to each cell mask. b, Cell type map shows the cell type calling results. Colors are assigned according to their corresponding cell type categories. c, Tissue region map shows laminar structure of hippocampus. Scale bar: 200 µm. d, Side-by-side comparison of cell type compositions in each tissue region from ClusterMap and pciSeq of the ISS data.

ClusterMap analyses in the 3D datasets. a, Statistics of ClusterMap identified cells in the 3D 126 STARmap⁶ cardiac organoid²⁷ 8-gene dataset. Left: Histogram of detected reads (DNA amplicons) 127 per cell. Middle: Histogram of genes per cell. Right: Correlation plot between genes per cell and 128 reads per cell. **b**, **c**, UMAP and heatmap visualization of three cell types in the STARmap⁶ cardiac 129 organoid 8-gene dataset. The number of cells in each cell type is as follows: cardiomyocytes, 929; 130 induced pluripotent stem cells (iPSCs), 489; mesenchymal stem cells (MSCs), 101. d, 3D four-131 channel composite raw fluorescent image of the first sequencing round shows spatial arrangement 132 of mRNA molecules in the STARmap⁶ mouse V1 28-gene dataset. Width 184 µm, height 194 µm, 133 depth 100 µm. e, Statistics of ClusterMap identified cells in (d). Left: Histogram of detected reads 134 (DNA amplicons) per cell. Middle: Histogram of genes per cell. Right: Correlation plot between 135 genes per cell and reads per cell. f, g, UMAP and heatmap visualization of three cell types of (d). 136

Performance comparison of ClusterMap using physical density, gene distance, and joint 139 140 information. a, Examples of cell segmentation using only physical density information (left), gene 141 (NGC) distance information (middle), and joint information (right). **b**,**c**, Bar plots demonstrating the percentage of over-/under- segmented cells in ground truth cells (b) and overall accuracy (c) 142 143 in using physical distances information, gene (NGC) distance information, and joint information. d,e, Bar plots demonstrating the percentage of over-/under- segmented cells in ground truth cells 144 (left) and overall accuracy (right) in using physical distances information, random (d) or identical 145 (e) gene (NGC) distance information, and joint information. f, Raw DAPI image of the targeted 146 mouse placenta tissue. Scale bar: 20µm. g,h, Line plots showing the number of cells and overall 147 accuracy to the radius. i-m, Two examples of the hippocampus regions in STARmap mouse V1 148 1020-gene datasets showing raw spatial transcriptomics data (i,l), ClusterMap results without 149 DAPI (**j**,**m**), and ClusterMap results with DAPI (**k**,**n**). Scale bar: 20 µm. **o**,**p**, Bar plots showing 150 the percentage of over-/under- segmented cells (o) and overall accuracy (p) from ClusterMap 151 without and with DAPI. 152

Comparison on STARmap mouse placenta data Predefined distribution-based method

Comparison on pciSeq hippocampus data

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Performance comparison of ClusterMap and other method across different types of in situ 155 transcriptomic data. a, Example of a region in the STARmap⁶ mouse V1 1020-gene dataset with 156 157 DAPI signals (gray) showing ground truth cell nuclei. Red contours show cell boundaries identified by predefined distribution-based method²⁰ (left) and ClusterMap (right), respectively. 158 **b.c**, As in (a) but using the STARmap⁶ mouse placenta 903-gene dataset and published pciSeq⁴ 159 dataset. d, Bar plots demonstrating the remaining RNA numbers, cell numbers and segmentation 160 accuracy for each dataset. In each bar plot, results from predefined distribution-based method²⁰, 161 ClusterMap, and published reports were shown in blue, red, and yellow, respectively. 162

164 Supplementary Table

- 165 **Supplementary Table 1.** Summary of the name, *in situ* sequencing protocol, number of genes,
- number of cells, number of reads, number of cell types, corresponding figures and note of 7
- 167 datasets.

| Dataset | Experimental Method | # Genes | # Cells | # Reads | # Cell types | Figures | Note |
|---|------------------------|------------|------------|-----------|-----------------|--|--------------------------------|
| STARmap mouse V1 1020-gene | STARmap | 1,020 | 1,599 | 863,426 | 16 | Fig. 1c, Fig. 2, Supplementary Figs. 3,4,5 | Source: Ref. 6. 2D analysis |
| STARmap mouse placenta 903- gene | STARmap | 903 | 7,224 | 5,090,930 | 12 | Fig. 3, Fig. 4, Supplementary Figs. 6,7 | New data. 2D analysis |
| MERFISH mouse POA | MERFISH | 140 | 10,320 | 3,065,171 | 9 | Fig. 5, Supplementary Fig. 8 | Source: Ref. 3. 3D analysis |
| pciSeq mouse hippocampus | ISS | 98 | 2,924 | 31,750 | 23 | Fig. 5, Supplementary Fig. 9 | Source: Ref. 4. 2D analysis |
| osmFISH mouse SSp | osmFISH | 33 | 8,538 | 1,690,328 | 31 | Fig. 5, Supplementary Fig. 8 | Source: Ref. 5. 2D analysis |
| STARmap cardiac organoid 8- gene | STARmap | 8 | 1,519 | 47,594 | 3 | Fig. 6, Supplementary Fig. 10 | New data, 3D analysis |
| STARmap mouse V1 28- gene | STARmap | 28 | 24,590 | 753,396 | 11 | Fig. 6, Supplementary Fig. 10 | Source: Ref. 6. 3D analysis |